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Transient inhibition of ROR- γ t therapeutically limits intestinal inflammation by reducing T_H17 cells and preserving ILC3

David R. Withers^{1,*}, Matthew R. Hepworth^{2,3,4}, Xinxin Wang^{2,3,4}, Emma C. Mackley¹, Emily E. Halford¹, Emma E. Dutton¹, Clare L. Marriott¹, Verena Brucklacher-Waldert⁵, Marc Veldhoen⁵, Judith Kelsen⁶, Robert N. Baldassano⁶, Gregory F. Sonnenberg^{2,3,4,*}

¹MRC Centre for Immune Regulation, Institute of Immunology & Immunotherapy, College of Medical and Dental Sciences, University of Birmingham, UK

²Joan and Sanford I. Weill Department of Medicine, Gastroenterology Division, Weill Cornell Medicine, New York, New York, USA.

³Department of Microbiology & Immunology, Weill Cornell Medicine, New York, New York, USA.

⁴The Jill Robert's Institute for Research in Inflammatory Bowel Disease, Weill Cornell Medicine, New York, New York, USA.

⁵Babraham Institute, Babraham Research Campus, Cambridge, UK

⁶Division of Gastroenterology, Hepatology and Nutrition, Children's Hospital of Philadelphia, Philadelphia, PA, 19104, USA

**The Sonnenberg and Withers laboratories contributed equally to this work.*

*CORRESPONDENCE: Dave Withers, d.withers@bham.ac.uk; and Gregory F. Sonnenberg, gfsonnenberg@med.cornell.edu

Abstract

RAR-related orphan receptor γ t (ROR- γ t) directs differentiation of pro-inflammatory T helper 17 (T_H17) cells and is a potential therapeutic target in chronic autoimmune and inflammatory diseases¹⁻³. However, ROR- γ t-dependent group 3 innate lymphoid cells (ILC3s) provide essential immunity and tissue protection in the intestine⁴⁻¹¹, suggesting that targeting ROR- γ t could also result in impaired host defense to infection or enhanced tissue damage. Here, we demonstrate that transient chemical inhibition of ROR- γ t in mice selectively reduces cytokine production from T_H17 cells but not ILC3s in the context of intestinal infection with *Citrobacter rodentium*, resulting in preserved innate immunity. Transient genetic deletion of ROR- γ t in mature ILC3s also did not impair cytokine responses in the steady state or during infection. Finally, pharmacologic inhibition of ROR- γ t provided therapeutic benefit in mouse models of intestinal inflammation, and reduced the frequencies of T_H17 cells but not ILC3s isolated from primary intestinal samples of individuals with inflammatory bowel disease (IBD). Collectively, these results reveal differential requirements for ROR- γ t in the maintenance of T_H17 cell versus ILC3 responses, and suggest that transient inhibition of ROR- γ t is a safe and effective therapeutic approach during intestinal inflammation.

While therapeutic targeting of the T_H17 cell pathway is effective in several autoimmune diseases¹²⁻¹⁴, in Crohn's disease, blockade of IL-17A is ineffective and in some cases was associated with worsened disease or increased opportunistic infections¹⁵⁻¹⁷. This is consistent with mouse models demonstrating protective functions for IL-17A and the often co-expressed cytokine IL-22 in the intestine¹⁸⁻²³, which is in part mediated by ROR- γ ⁺ ILC3⁴⁻¹¹. Small-molecular weight inhibitors of ROR- γ are an alternative therapeutic approach, with several identified and found to exhibit efficacy in models of autoimmunity²⁴⁻²⁸. However, the effect of targeting ROR- γ in intestinal immune cells remains unexplored.

To investigate the impact of ROR- γ inhibitors in the intestine, C57BL/6 mice were infected with the enteric pathogen *Citrobacter rodentium*, which elicits ROR- γ ⁺ T_H17 cell and ILC3 responses^{6,7,29,30}, and treated with a vehicle control or GSK805, a orally-available inhibitor of ROR- γ -mediated transcription²⁴. Relative to vehicle control, oral GSK805 treatment reduced the frequency and total number of IL-17A- and IL-22-producing T_H17 cells, but not IFN- γ ⁺ T_H1 cells, in the colon lamina propria (cLP) and mesenteric lymph node (mLN; **Fig. 1a, d and e**). In contrast, GSK805 treatment did not alter the percentage or total numbers of IL-17A- or IL-22-producing ILC3 in the colon or mLN (**Fig. 1b, c and f**). IL-22 production from ILC3s and T cells mediates protective immunity to *C. rodentium* at early versus late stages of infection, respectively^{6,7,29}. Consistent with this, GSK805-treated mice had a comparable burden of *C. rodentium* early during infection, but increased levels at later stages as compared to control treated mice (**Supplementary Fig. 1a**). Preserved innate immunity allowed for 75% of mice to survive at day 20 post infection, in contrast to anti-IL-23 monoclonal antibody administration which resulted in 100% mortality (**Fig. 1g**), due to inhibition of ILC3 and T_H17 cell responses^{7,23,30}. Colon length was increased (**Fig. 1h**) and histologic signs of inflammation and tissue damage were reduced (**Fig. 1i**) in GSK805-treated mice relative to controls. In response to a systemic infection with the opportunistic pathogen *Candida albicans*, vehicle control and GSK805-treated mice had comparable IL-17A⁺ ILC3 responses in the spleen, fungal burdens, and a lack of inflammation in the kidney, as opposed to *Il17a*^{-/-} mice which exhibited significantly increased fungal burdens and kidney inflammation (**P* < 0.05, **Supplementary Fig. 1b-d**).

To further define the requirements for ROR- γ in the maintenance of ILC3 responses, we generated mice where *Rorc* can be temporally deleted in mature ILC3. This was accomplished by crossing *Rorc*^{flxed} mice and *Id2*^{creERT2} mice, which targets cre recombinase activity to ILCs following

tamoxifen administration (**Supplementary Fig. 2a**), to generate $\text{Id2}^{\text{i}\Delta\text{ROR-}\gamma\text{t}}$ mice. After 2 weeks of tamoxifen administration, comparable frequencies of total ILCs ($\text{Lineage}^{\text{-}}(\text{Lin}^{\text{-}})\text{CD127}^{\text{+}}$) were observed in the mLN of control and $\text{Id2}^{\text{i}\Delta\text{ROR-}\gamma\text{t}}$ mice, with efficient deletion of ROR- γt protein (**Fig. 2a**). Despite a loss of ROR- γt , analyses of the total ILCs revealed comparable frequencies of cells with an ILC3-like phenotype, including those expressing CCR6, CD4 and MHCII in the mLN of control and $\text{Id2}^{\text{i}\Delta\text{ROR-}\gamma\text{t}}$ mice (**Fig. 2b, c**). Further, microarray analyses of sort purified $\text{CCR6}^{\text{+}}$ ILCs (gated as $\text{Lin}^{\text{-}}\text{CD127}^{\text{+}}\text{CCR6}^{\text{+}}$) revealed a decrease in *Rorc* transcripts from tamoxifen administered $\text{Id2}^{\text{i}\Delta\text{ROR-}\gamma\text{t}}$ mice relative to *Rorc*^{flxed} mice, but sustained expression of other genes associated with ILC3 function (**Fig. 2d**). To determine how long ILC3-like cells could persist in the absence of *Rorc*, 6 week old $\text{Id2}^{\text{i}\Delta\text{ROR-}\gamma\text{t}}$ mice were continuously administered tamoxifen for 12 weeks. At this time point, cells with an ILC3-like phenotype could be observed in the mLN, with modest changes in absolute cell numbers and surface markers (**Supplementary Fig. 3**). In contrast, an ILC3-like population was not maintained in the small intestine, but rather an increase in the numbers of $\text{Lin}^{\text{-}}\text{CD127}^{\text{+}}\text{NKp46}^{\text{+}}$ ILC1-like cells was evident (**Supplementary Fig. 4**). These results suggest that loss of ROR- γt may cause an expansion of ex-ILC3 or ILC1 in the small intestine³¹⁻³³, or a loss of pro-inflammatory ILC3^{34,35}. Consistent with a loss in pro-inflammatory ILC3, a decrease in $\text{IFN-}\gamma^{\text{+}}$ ILCs was observed in the small intestine, while no change was observed in the mLN (**Supplementary Fig. 4**). Despite changes in the intestine, comparable frequencies and numbers of ILCs lacking detectable ROR- γt expression but able to produce both IL-17A and IL-22 were maintained in the mLN following 12 weeks of continuous tamoxifen administration (**Supplementary Fig. 4**).

Consistent with a preservation of ILC3 function, tamoxifen-administered *Rorc*^{flxed} control and $\text{Id2}^{\text{i}\Delta\text{ROR-}\gamma\text{t}}$ mice exhibited comparable IL-17A⁺ ILC3 responses in the spleen, CFU and a lack of inflammation in the kidney following infection with *C. albicans* (in contrast to *Il17a*^{-/-} mice, **Supplementary Fig. 2b, c**). Furthermore, *Rorc*^{flxed} and $\text{Id2}^{\text{i}\Delta\text{ROR-}\gamma\text{t}}$ mice exhibited comparable IL-22⁺ ILC3 responses in the mLN following *C. rodentium* infection (**Fig. 2e**). To compare these results to transient genetic deletion of ROR- γt in $\text{CD4}^{\text{+}}$ T cells, *Rorc*^{flxed} mice were crossed to $\text{CD4}^{\text{creERT2}}$ mice, to generate $\text{CD4}^{\text{i}\Delta\text{ROR-}\gamma\text{t}}$ mice. IL-17A production was reduced in $\text{CD4}^{\text{+}}$ T cells in the mLN of tamoxifen-administered and *Citrobacter rodentium* infected $\text{CD4}^{\text{i}\Delta\text{ROR-}\gamma\text{t}}$ mice relative to littermate controls (**Fig. 2f**), demonstrating differential requirements for ROR- γt in ILC3 versus $\text{T}_{\text{H}}17$ cells. Further, 75% of $\text{Id2}^{\text{i}\Delta\text{ROR-}\gamma\text{t}}$ mice survived at day 20

post infection with *C. rodentium*, in comparison to mice with a genomic deletion in ROR- γ t (*Rorc*^{-/-} mice) and thus impaired ILC3 and T_H17 development, which exhibited 100% mortality (**Fig. 2g**). CD4^{iΔROR- γ t} mice were partially susceptible at later stages post-infection (**Fig 2g**), yet exhibited increased colon length (**Fig. 2h**) and reduced histologic signs of intestinal inflammation (**Fig. 2i**) relative to control and Id2^{iΔROR- γ t} mice, suggesting that while T_H17 cells are important for immunity at late stages of infection, they also are primary mediators of intestinal damage and inflammation.

Previous studies reported a low turnover rate of ILC3³⁶, and pulse-chase experiments with tamoxifen administration in Id2^{iΔROR- γ t} mice revealed differing half-lives of ILC3 in the mLN and intestine (**Supplementary Fig. 5**). Further, in all tissues assessed, the markers of proliferation were higher in T_H17 cells than ILC3 at steady state (**Supplementary Fig. 6**). However, this did not account for the differential requirements of sustained ROR- γ t expression, as treatment of mice with IL-2/anti-IL-2 complexes induced ILC3 proliferation (**Supplementary Fig. 7**) and equally expanded a population of the ROR γ t⁻ ILC3 in the mLN of tamoxifen treated Id2^{iΔROR- γ t} mice that produced comparable levels of IL-17A and IL-22 relative to ROR γ t⁺ ILC3 in control mice (**Supplementary Fig. 8**).

To experimentally address whether chemical inhibition of ROR- γ t can provide therapeutic benefit in the context of intestinal inflammation, *Il10*^{-/-} mice were aged to 10 weeks (a time point at which spontaneous colonic inflammation is observed), and orally administered either vehicle control or GSK805 at a dose 10 mg/kg/day for two weeks. GSK805 administration selectively reduced the frequency of IL-17A⁺ T_H17 cells in the colonic tissues, while no effect on the frequency of IL-17A⁺ ILC3 was observed (**Fig. 3a**). Furthermore, GSK805 treatment reduced elevated amounts of fecal lipocalin-2 (**Fig. 3b**), prevented shortening of the colon (**Fig. 3c**) and reduced histologic signs of inflammation in the colon (**Fig. 3d**) of *Il10*^{-/-} mice relative to controls. In a second model of intestinal inflammation, CBir1 transgenic CD4⁺ T cells, which recognize an immunodominant and commensal bacteria-derived antigen in Crohn's disease³⁷, were transferred to *Rag1*^{-/-} mice. At two weeks post transfer, intestinal inflammation was observed and then mice were orally administered either vehicle control or GSK805 at a dose 10 mg/kg/day for two weeks. GSK805 treatment significantly reduced the frequency of colonic CBir1-specific T_H17 cells, but not IL-17A⁺ ILC3 (***P* < 0.01, **Fig. 3e**). Further, GSK805 treatment reduced fecal lipocalin-2, colonic shortening and histologic signs of inflammation in the colon of *Rag1*^{-/-} mice receiving a CBir1 T cell transfer, relative

to control mice (**Fig. 3f–h**). Together, these results demonstrate that transient inhibition of ROR- γ t provides therapeutic benefit in the context of two mouse models of intestinal inflammation.

To interrogate how transient inhibition of ROR- γ t modulates human T_H17 cell and ILC3 responses, colonic resection tissues from pediatric individuals with Crohn's disease were digested and single cell suspensions were cultured *in vitro* for 12 hours in the presence of DMSO control or GSK805. ILC3s were identified by gating on Lin⁻ cells and examining for CD127⁺ populations (**Fig. 4a**) followed by co-expression of NKp44 and c-kit. Incubation with GSK805 relative to DMSO control did not alter ILC3 frequencies (**Fig. 4b**), production of IL-22 (**Fig. 4c**), or production of the pro-inflammatory cytokine TNF (**Fig. 4d**). In contrast, *in vitro* culture with GSK805 resulted in a decrease in the frequency of CD4⁺ T cells that produce IL-17A (**Fig. 4e**) and IL-22 (**Fig. 4f**) relative to DMSO. The decrease in IL-17A⁺ and IL-22⁺ T_H17 cells following culture in the presence of GSK805 was observed in cell suspensions from 8 out of 10 pediatric individuals with Crohn's disease (**Fig. 4g**), did not influence IFN- γ ⁺ T cells (**Fig. 4h**), and resulted in comparable or increased frequencies of IL-17A⁺ and IL-22⁺ ILC3 (**Fig. 4i**). Similar data were also obtained when another ROR- γ t inhibitor, Digoxin²⁵, was utilized *in vitro* (**Supplementary Fig. 9**). Collectively, these results demonstrate that similar to mice, transient *in vitro* inhibition of ROR- γ t in cells isolated from individuals with IBD selectively limits T_H17 cells, with limited influence on ILC3s.

Subsets of ILCs and CD4⁺ T helper cells exhibit remarkably similar transcriptional profiles and effector cytokine responses, yet can promote differential functions in the context of immunity, inflammation and tissue repair^{1-3,11,38}. Therefore, therapeutically targeting conserved pathways in the context of chronic inflammatory diseases has the potential to influence both arms of the immune system, and in addition to limiting a pathologic response, could inappropriately target beneficial responses that mediate protective immunity or tissue repair. This may account for off-target consequences of biologic therapies in patients with IBD that broadly inhibit cytokine-cytokine receptor interactions, such as the anti-IL-17 clinical trials that resulted in increased infections or worse disease^{15-17,39}. Here, we uncover an unexpected cellular mechanism by which transient inhibition of the transcription factor ROR- γ t allows for selective targeting of adaptive T_H17 cells without influencing protective ILC3 responses in both mice and *in vitro* cultures with human tissues samples. These data provoke a challenge to our understanding of the modular effects of lineage-specifying transcription factors in the maintenance and function of innate versus adaptive

lymphocytes, and identify ROR- γ t inhibition as an attractive future therapeutic approach to treat IBD and other chronic inflammatory diseases.

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Author Contributions

D.R.W., G.F.S. designed and performed experiments, analyzed data, and wrote the paper; M.R.H., X.W., E.C.M., E.E.H., E.E.D., C.L.M., V.B-W., M.V. designed and performed experiments; J.K. and R.N.B. provided critical reagents and invaluable expertise.

Competing financial interests

The authors declare no competing financial interests.

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Figure Legends

Figure 1. Transient chemical inhibition of ROR- γ t selectively reduces T_H17 responses but not ILC3s during infection. (a–i) C57BL/6 mice were infected with *C. rodentium* and treated daily with vehicle control or GSK805 (10 mg/kg) starting on day –1 and continued for the duration of the experiment (a) Frequency of IL-17A-producing T_H17 cells and (b) IL-17A- or (c) IL-22-producing ILC3 at day 10 post infection. (d) Total numbers of IL-17A⁺ and IL-22⁺ T_H17 cells, (e) IFN- γ ⁺ T cells and (f) IL-17A⁺ and IL-22⁺ ILC3 in the colon at day 10 post infection. (g) Survival curve of infected mice. (h) Colon length and (i) Hematoxylin and eosin stained histologic sections of the distal colon at day 10 post infection, representative of $n = 5$. Scale bar, 200 μ m. All data are representative of five individual mice ($n = 5$) and was replicated in two independent experiments. Data shown are mean \pm SEM (error bars). Statistics compare treatment versus control groups using Student's *t* test. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$.

Figure 2. Transient genetic deletion of ROR- γ t impairs T_H17 cells but not ILC3s during homeostasis and intestinal infection. (a–g) Analyses of ILC3s in *Rorc*^{flxed} control and *Id2* ^{Δ ROR- γ t} mice following daily treatment with tamoxifen for two weeks. (a) Gating for ILCs and subsequent analysis for ROR- γ t. Lin[–] CD127⁺ cells from the mLN were analyzed for (b) CCR6 and (c) CD4 protein, as well as sequential gating for MHCII protein. (d) Relative expression of ILC3-associated genes from mLN sort purified CCR6⁺ ILCs was assessed by genome-wide microarrays. (e–h) Mice were infected with *C. rodentium* and treated daily with tamoxifen starting on day –1. (e) Identification of IL-22-producing ILC3 and (f) IL-17-producing T_H17 cells at day 10 post infection. (i) Survival curve of infected mice. (g) Colon length and (h) Hematoxylin and eosin stained histologic sections of the distal colon at day 10 post infection, representative of $n = 5$. Scale bar, 200 μ m. All data are representative of five individual mice ($n = 5$) and was replicated in two independent experiments. Data shown are mean \pm SEM (error bars). Statistics compare treatment versus control groups using Student's *t* test. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$

Figure 3. Transient inhibition of ROR- γ t selectively limits T_H17 cell responses and reduces intestinal inflammation. (a–d) Ten-week-old *Il10*^{–/–} mice were treated with vehicle control or GSK805

(10 mg/kg) for two weeks. (a) Frequency of IL-17A-producing T_H17 cells (red) or ILC3 (blue). Amount of fecal lipocalin-2 (b), colon length (c), and Hematoxylin and eosin stained histologic sections of the distal colon (d). Scale bar, 200 μm, representative of *n* = 5. (e–h) Adoptive transfer of CBir1 TCR transgenic T cells into *Rag1*^{-/-} mice, followed 2-3 weeks later by daily treatment with either vehicle control or GSK805 (10 mg/kg) for 2 weeks. (e) Analyses of IL-17A-producing T_H17 cells (top) or ILC3 (bottom), (f) amount of fecal lipocalin-2, (g) colon length, and (h) Hematoxylin and eosin stained histologic sections of the distal colon. Scale bar, 200 μm, representative of *n* = 5. All data are representative of five individual mice (*n* = 5) and was replicated in two independent experiments. Data shown are mean ± SEM (error bars). Statistics compare treatment versus control groups using Student's *t* test. **P* < 0.05, ***P* < 0.01, ****P* < 0.001

Figure 4. Transient inhibition of ROR-γt selectively reduces T_H17 cells in intestinal tissue from pediatric individuals with Crohn's disease. (a–i) Intestinal resection tissue from pediatric individuals with Crohn's disease was cultured with DMSO control or GSK805 (0.5 μM) for 12 hours. (a) Identification of Lin⁻IL-7Rα⁺ ILCs. (b) Percentage of NKp44⁺c-kit⁺ ILC3s. (c) Production of IL-22 and (d) TNF by gated c-kit⁺ ILC3s. (e) Expression of IL-17A and (f) IL-22 by CD3⁺ T_H17 cells. (g) Fold change in the percentage of IL-17A⁺ or IL-22⁺ T_H17 cells, (h) IFN-γ⁺ T_H1 cells and (i) IL-17A⁺ or IL-22⁺ ILC3 following culture with GSK805 relative to DMSO. All data are representative or shown for *n* = 10 samples from pediatric individuals with Crohn's disease.

Online methods

Mice

C57BL/6, CD4^{creERT2} (ref. 40), Id2^{creERT2} (ref. 41), *Rorc*^{-/-}, *Il10*^{-/-}, ROSA^{mT/mG}, *Rag1*^{-/-}, and *Rorc*^{floxed} (ref. 24) mice were originally obtained from the Jackson Laboratory (Bar Harbor, ME) and bred and maintained at Weill Cornell Medical College, (New York, NY, USA) or the University of Birmingham (Birmingham, UK). CBir1 mice and *Il17a*^{-/-} mice were previously described^{37,42}. Mice used were a mix of male and female animals, aged 6-10 weeks at the start of the experiment. For experiments at the University of Birmingham, animals were used in accordance with Home Office guidelines at the University of Birmingham. For experiments performed at Weill Cornell, all protocols were approved by the Weill Cornell Institutional Animal Care and Use Committee (IACUC), and all experiments were performed according to the guidelines of the Weill Cornell IACUC. To generate Id2^{ΔROR-γt} and CD4^{ΔROR-γt} mice, Id2^{creERT2} and CD4^{creERT2} mice were crossed to *Rorc*^{floxed} mice respectively. Where noted, experimental and control mice were maintained on Tamoxifen-containing diet (TAM400, Harlan Laboratories) for at least 10-14 days prior to analysis or infection, or were gavaged daily with 10 mg/kg of tamoxifen in corn oil (Sigma). GSK805 (C₂₃H₁₈Cl₂F₃NO₄S) was previously identified by a Fluorescence Resonance Energy Transfer (FRET) assay that examined for inhibition of the ROR-γt ligand-binding domain and cofactor peptide SRC1 and compared to other nuclear receptors²⁴. Inhibition of ROR-γ was not tested. Efficacy and potency was previously demonstrated, and RNAseq data revealed comparable transcriptional changes to that observed in *Rorc* gene deletion²⁴. In this study, GSK805 was provided from EMD Milipore (53136) at a purity ≥ 95% by HPLC. Mice received 10 mg/kg/day of GSK805 dissolved in DMSO and provided by oral gavage in corn oil, or DMSO alone in corn oil as a vehicle control. For BrdU incorporation, mice were given 1.5 mg BrdU i.p. and analyzed after 24 hours. Mice were injected i.p. for 3 consecutive days with IL-2/anti-IL2 complexes generated by mixing recombinant protein and antibody for 30 minutes at 37 °C prior to injection. Per injection, each mouse received 1 μg IL-2 (Peprotech) and 5 μg anti-IL-2 monoclonal antibody (clone JES6-1A12, eBioscience), and control mice received PBS. No animals were excluded from the analyses.

Citrobacter rodentium infection

Citrobacter rodentium (formerly *Citrobacter freundii*, biotype 4280) strain DBS100 was prepared by

selecting a single colony and culturing in LB broth overnight as previously described⁷. Mice were inoculated with approximately 1×10^8 CFU in 200-250 μ L broth via oral gavage. Quantification of fecal CFU was determined by homogenization in sterile PBS and serial dilutions on MacConkey's agar.

***Candida albicans* infection**

Candida albicans infections were performed as previously described⁴³. In brief, 10^4 – 10^5 CFU of *Candida albicans* strain SC5314 were administered to mice by i.v. injection and analyzed three days later.

Mouse models of intestinal inflammation

To induce intestinal inflammation, *Il10*^{-/-} mice were aged to 10 weeks in our animal facilities, followed by 2 weeks of GSK805 treatment as described above. The CBir1 T cell transfer model of intestinal inflammation was previously described⁴⁴. In brief, 2×10^6 CD3⁺CD4⁺ T cells were sort purified as from spleens of CBir1 transgenic mice and adoptively transferred to *Rag1*^{-/-} recipients by i.v. injection. This protocol induces substantial colonic inflammation at 3 to 4 weeks⁴⁴, at which point mice were then treated for 2 weeks with GSK805 as described above.

Flow cytometry

Lymph nodes were teased using fine forceps and digested for 25 minutes in RPMI containing with 250 μ g/ml Collagenase-Dispase (Roche) and 25 μ g/ml DNase I. Digested tissue was then crushed through a 70 μ m nylon mesh. In some experiments, LNs were only crushed through a 70 μ m nylon mesh. Antibody staining was performed at 4 °C for 30 minutes. Antibodies raised against the following mouse antigens were used: B220 (clone RA3-6B2, eBioscience), CCR6 (clone 29-2L17, Biolegend), CD3 (clone 145-2C11, eBioscience), CD4 (clone GK1.5, eBioscience), CD45.1 (clone A20, eBioscience), CD45.2 (clone 104, eBioscience), CD5 (clone 53-7.3, eBioscience), CD11b (clone M1/70, eBioscience), CD11c (clone N418, eBioscience), GATA-3 (TWAJ, eBioscience), MHCII (clone AF6-120.1, eBioscience), IL-7R α (clone A7R34, eBioscience), IL-17A (clone TC11-18H10.1, eBioscience), IL-22 (clone 1H8PWSR, eBioscience), Ki-67 (clone SolA15, eBioscience), NKp46 (clone 29A1.4, eBioscience), ROR- γ t (clone B2D, eBioscience), T-bet (clone eBio4B10, eBioscience). All antibodies were used at a 1:200 dilution. The lineage dump channel contained antibodies against B220, CD3, CD5, NK1.1, CD11b and CD11c all in one cocktail, or

divided into two channels. Intracellular staining was done using the FoxP3 fixation and permeabilization kit (eBioscience) according to manufacturer's instructions. Incorporation of BrdU was detected using FITC BrdU Flow Kit (BD) according to manufacturer's instructions. Addition of Spherotech Accucount blank particles was used to calculate absolute cell numbers. Flow cytometry was performed on a Fortessa analyzer using FACSDiva6.2 software (BD), with data subsequently analyzed with FlowJo software (Tree Star). For cytokine production, cells were cultured *ex vivo* in the presence of either mouse recombinant IL-1 β (14-8012), IL-2 (14-8021), IL-6 (14-8061) and IL-23 (14-8231) at a concentration of 20 ng/mL for 2 hours (all obtained from eBioscience). Cells were then stimulated with 50 ng/mL PMA, 750 ng/mL Ionomycin and 10 μ g/mL Brefeldin A (all obtained from Sigma-Aldrich) for 4 hours.

Microarray analysis

For microarray analysis, Lin⁻CCR6⁺CD90.2⁺ cells were sorted from the mLN of *Rorc*^{flxed} or *Id2* ^{Δ ROR- γ t} mice were lysed directly in TRIzol. RNA was isolated, amplified, reverse-transcribed to cDNA and hybridized to an Affymetrix GeneChip (Mouse Gene 1.0ST), or obtained via the GEO database and normalized as previously described (41). All data sets were deposited in the GEO database and are available at ascension code GSE76201.

Histological sections

Distal colons were fixed with 4% PFA and embedded in paraffin, and 5 μ m sections were cut and stained with Hematoxylin and eosin. Whole kidneys were fixed with 4% PFA and embedded in paraffin, and 5 μ m sections were cut, stained with Periodic acid–Schiff and counterstained with Hematoxylin.

Human tissue and flow cytometry

Deidentified intestinal resection tissues from the colon of pediatric individuals with Crohn's disease were obtained from the Children's Hospital of Philadelphia (CHOP) following an CHOP Institutional Review Board (IRB) approved protocol (14-010981) and informed consent of participating individuals. Tissues were processed by first incubating in 1 mM EDTA, 1 mM DTT and 5% FCS (all obtained from Fisher Scientific) for 30 minutes at 37 °C with shaking to remove intestinal epithelial cells. Supernatants were then discarded and the remaining tissues were incubated in 0.5 mg/mL collagenase/dispase and 20

µg/mL DNase I for 20-30 minutes at 37°C with shaking to obtain the lamina propria fraction. Any remaining tissues were also included following mechanical dissociation and filtering through a 70 µm cell strainer. All cells were then viably cryopreserved at 150 °C in 90% FBS and 10% DMSO for future side-by-side analyses. Following thawing, cells were re-stimulated *ex vivo* with human recombinant IL-1β, IL-2, IL-6, IL-7 and IL-23 (20 ng/mL, eBioscience) for 8 hours in the presence of DMSO control, 0.5 µM GSK805 (EMD Milipore) or 5 µM digoxin (Sigma-Aldrich). Cells were then stimulated with 50 ng/mL PMA, 750 ng/mL Ionomycin and 10 µg/mL Brefeldin A (all obtained from Sigma-Aldrich) for 4 hours. Cells were then stained with antibodies for the following markers: anti-CD3 (clone UCHT1, eBioscience), anti-CD4 (clone OKT-4, eBioscience), anti-CD5 (clone UCHT2, eBioscience), anti-CD11b (clone M1/7B, eBioscience), anti-CD11c (clone 3.9, eBioscience), anti-CD14 (clone 61D3, eBioscience), anti-CD19 (clone 2H7, eBioscience), anti-CD127 (clone A019D5, Biolegend), anti-FcεR1 (clone AER-37, eBioscience), anti-CD117/c-kit (clone 104D2, eBioscience), anti-NKp44 (clone Z231, BC Cytometry). For intracellular staining, cells were fixed and permeabilized using a commercially available kit (eBioscience) and staining with anti-IL-17A (clone eBio64DEC17, eBioscience), anti-IL-22 (clone 22URTI, eBioscience) or anti-IFN-γ (clone 4S.B3, eBioscience). All antibodies were used at a 1:200 dilution. Lineage cocktails are defined in Figure 4 as the x-axis: CD19, CD11b, CD11c, and the y-axis: CD3, CD5, CD14, FcεR1.

Statistics

Data were analysed using GraphPad Prism (version 5.01). Student's t test or Mann-Whitney were used to determine significance which was set at $P < 0.05$. We did not use statistical methods to predetermine sample size, there was no randomization designed in the experiments, and the studies were not blinded. Samples sizes were estimated based upon sample availability and previous experimental studies of the intestinal immune system^{8,9,45,46}. No exclusion criteria were utilized in these studies.

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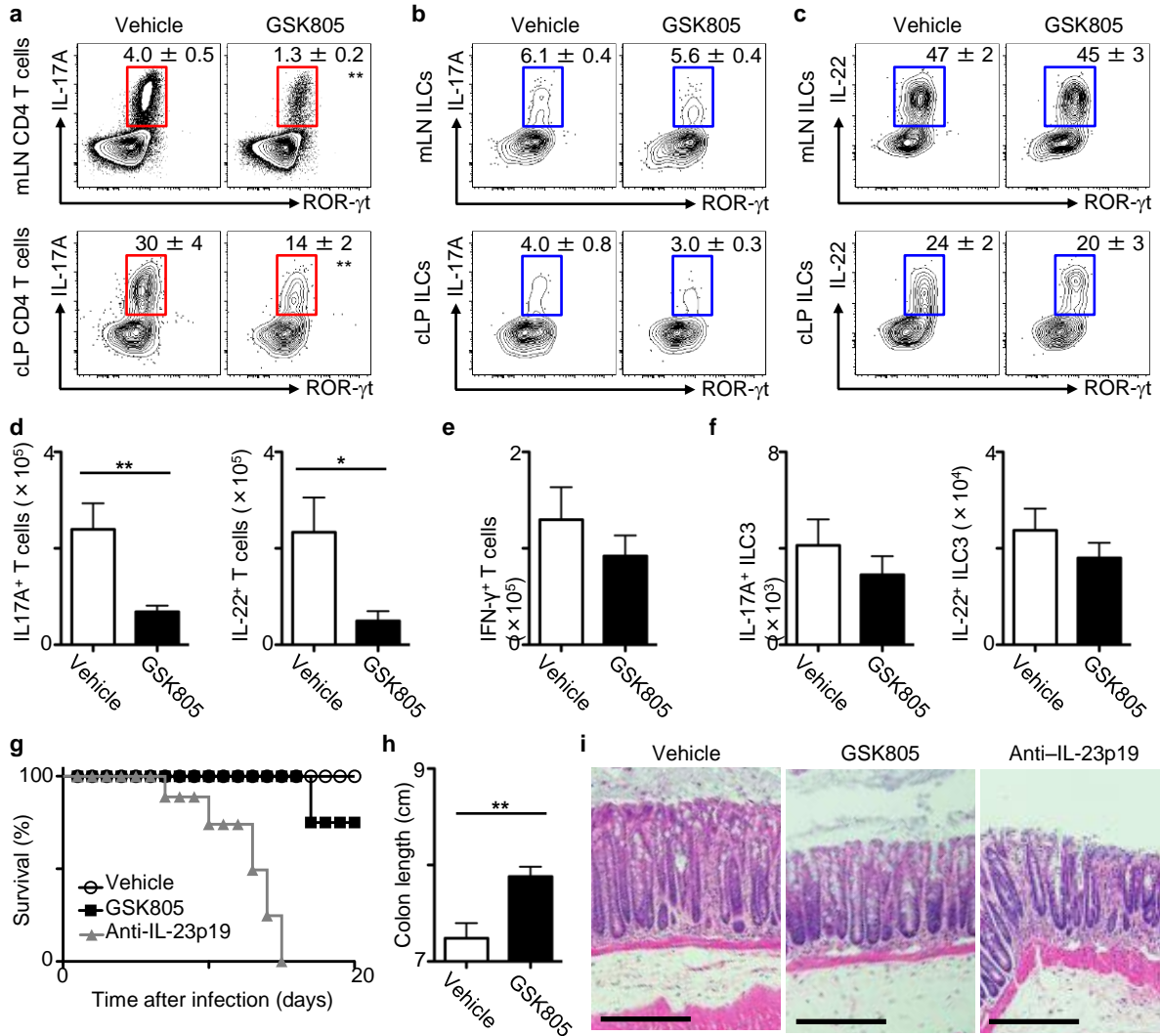


Figure 1

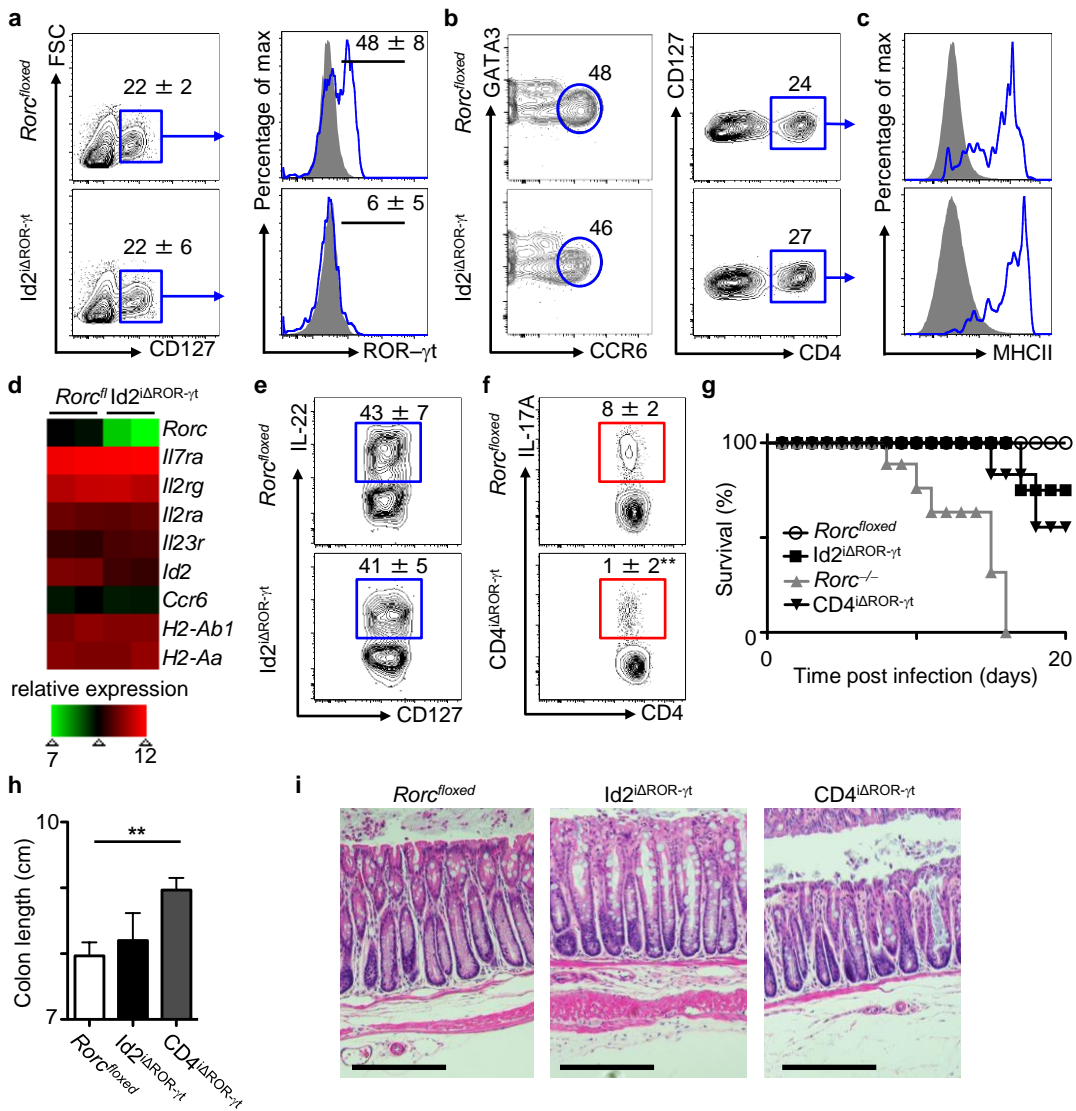


Figure 2

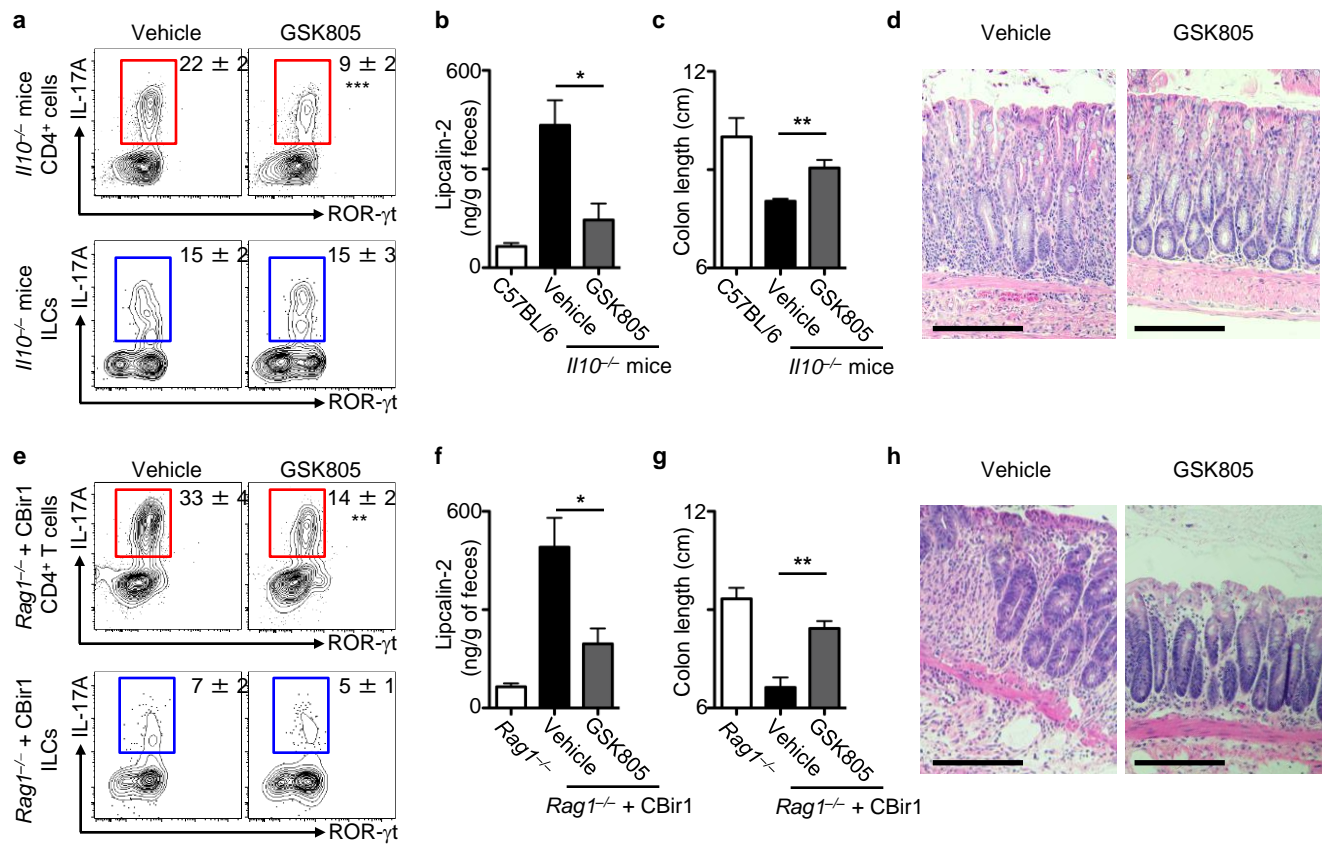


Figure 3

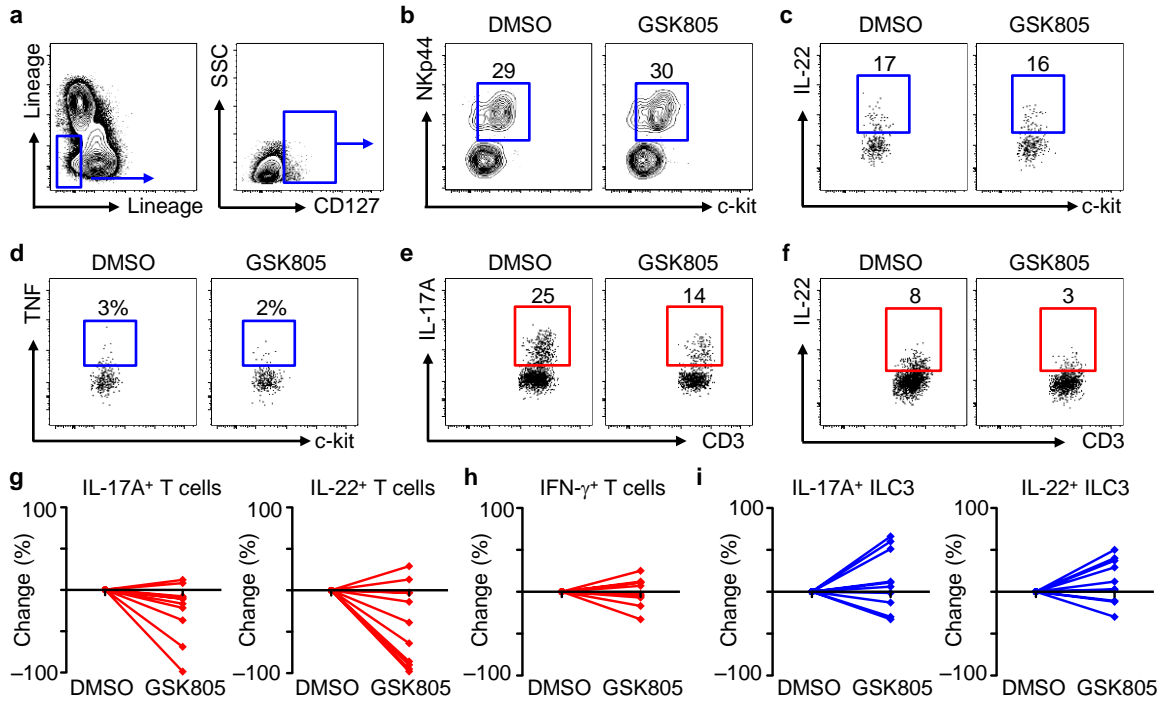
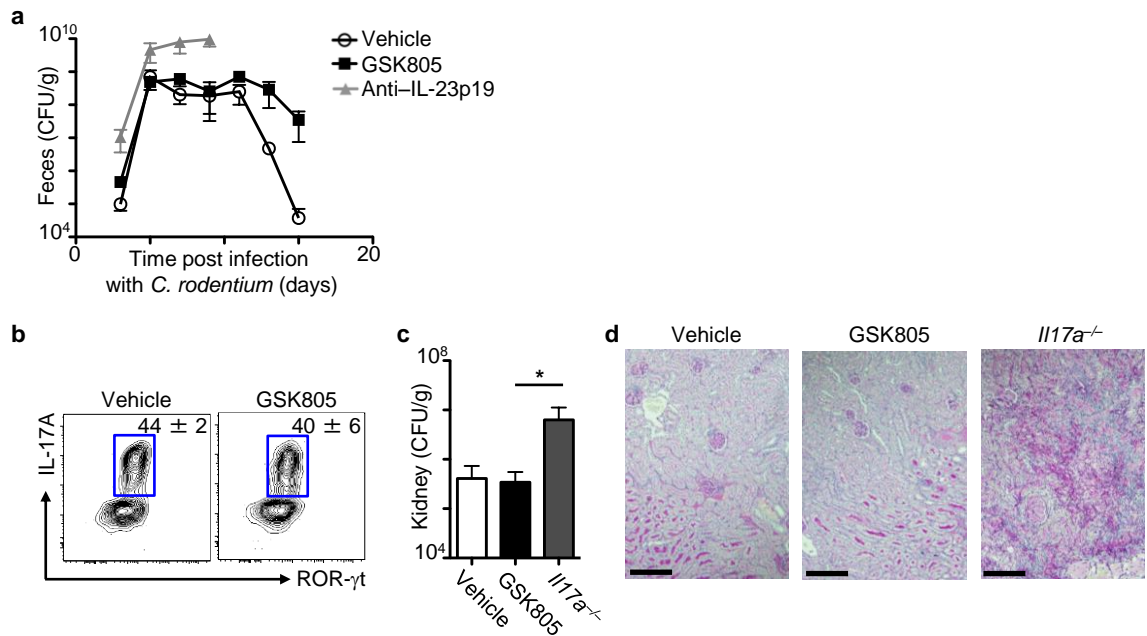


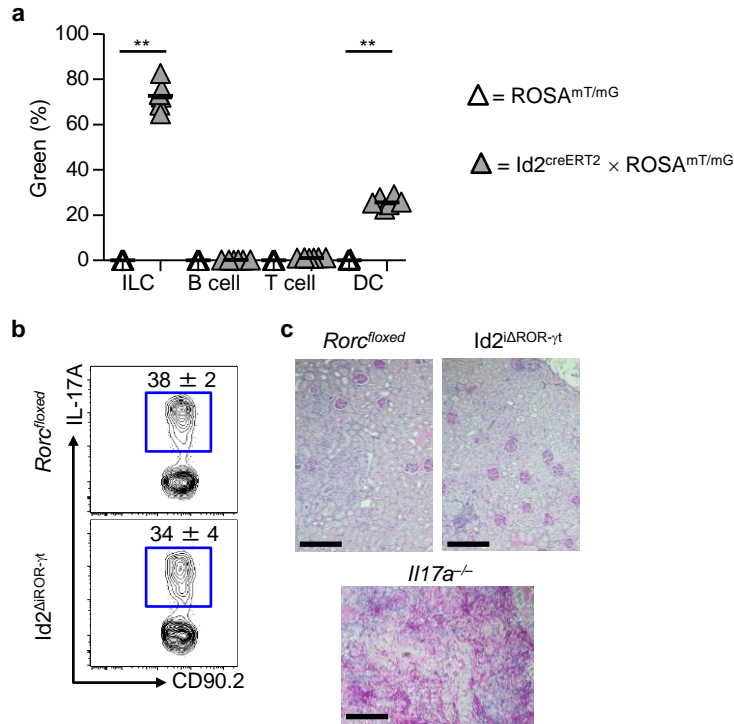
Figure 4

Transient inhibition of ROR- γ t therapeutically limits intestinal inflammation by reducing T_H17 cells and preserving ILC3

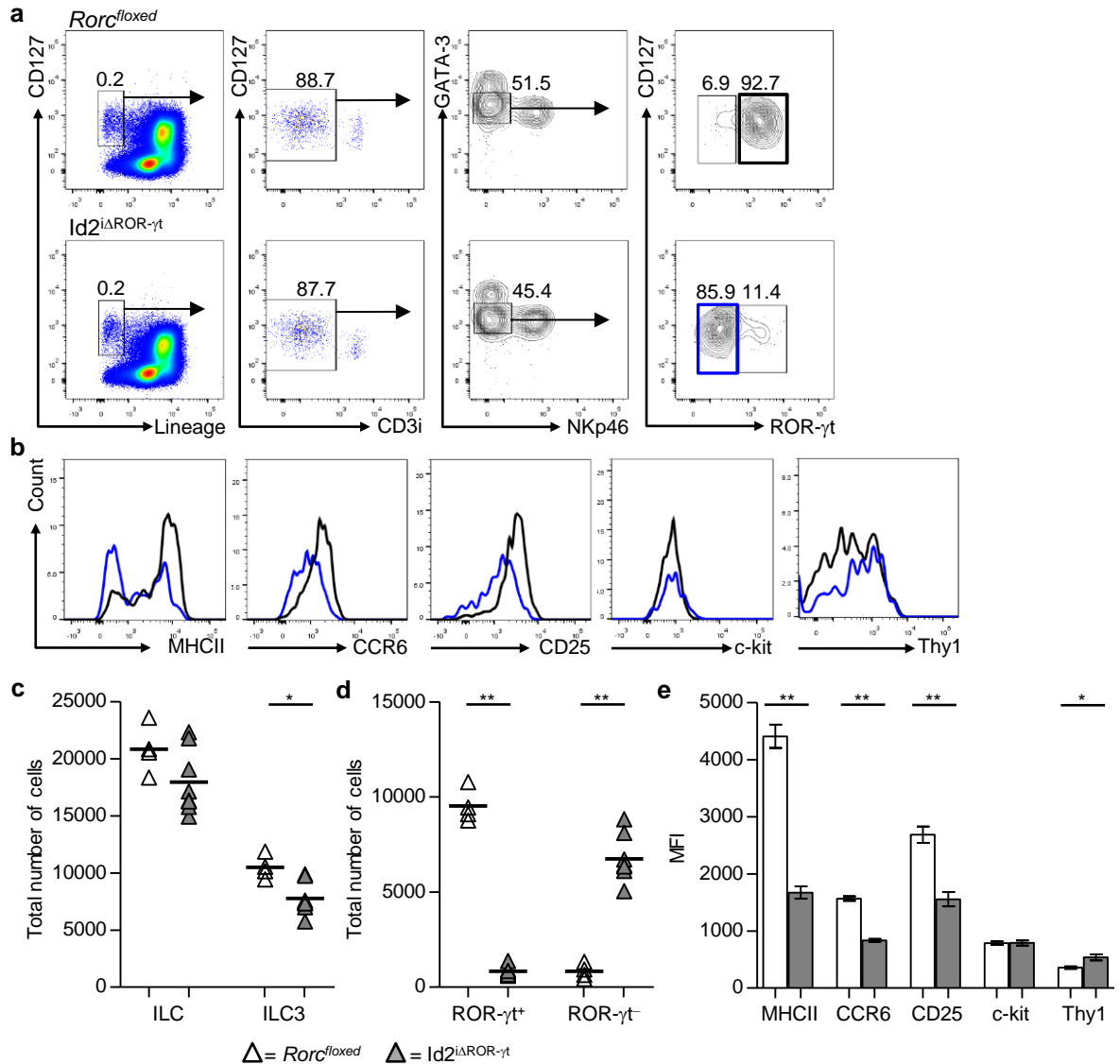
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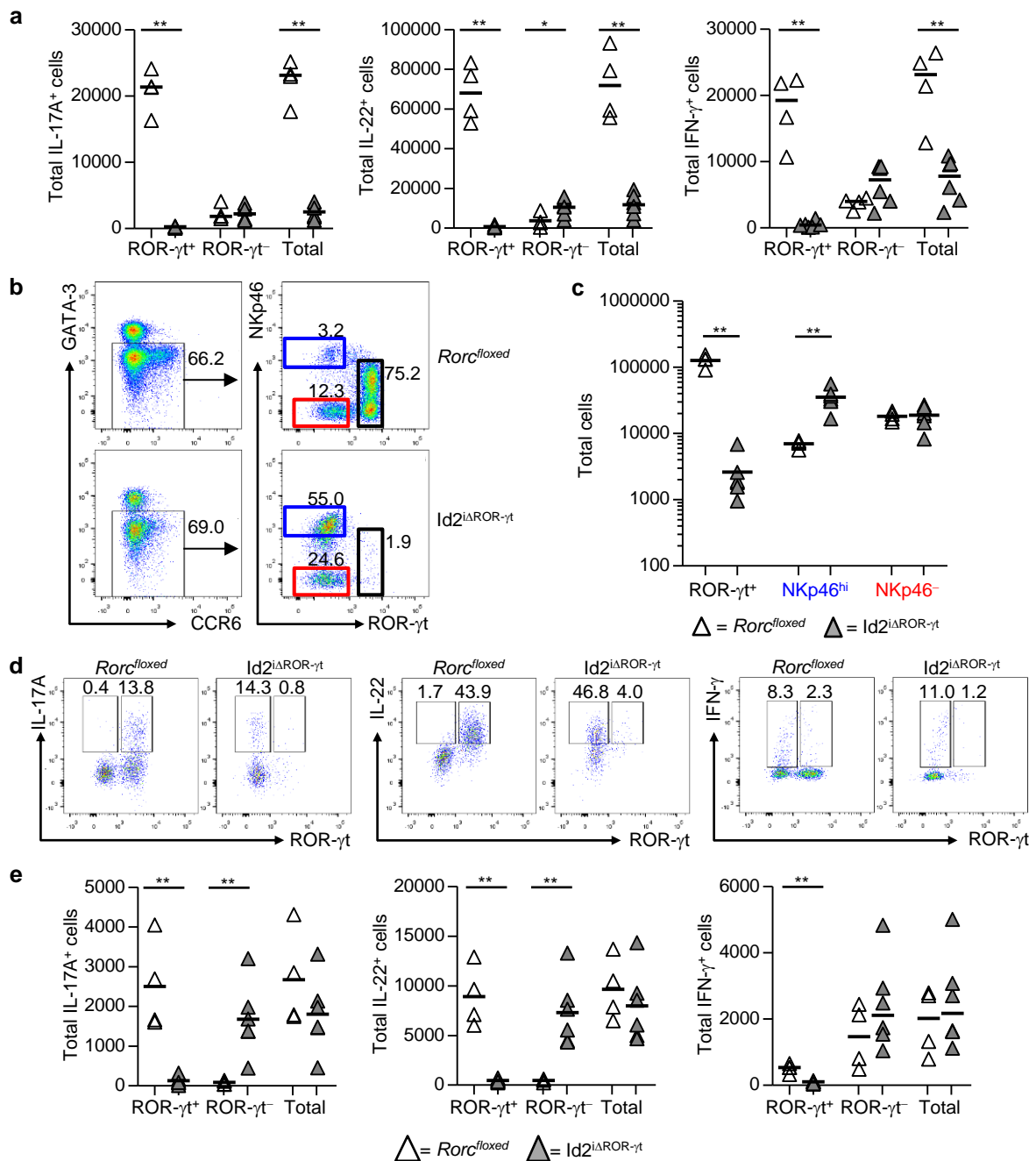
Supplementary Figure 1. Mice treated with GSK805 control early *C. rodentium* and *C. albicans* infections. (a) Kinetic CFU counts per g/feces in mice treated with vehicle control, GSK805 or anti-IL-23p19 monoclonal antibody following *C. rodentium* infection. (b–d) C57BL/6 mice were infected with *C. albicans*, treated daily with vehicle control or GSK805 (10 mg/kg) starting on day -1, and sacrificed on day 3. (b) Frequency of IL-17A producing ILC3. (c) CFU in the kidney and (d) Periodic acid-Schiff stained histologic sections of the kidney. Scale bar, 200 μ m. All data are representative of five individual mice ($n = 5$) and was replicated in two independent experiments. Data shown are mean \pm SEM (error bars). Statistics compare treatment versus control groups using Student's t test. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$.



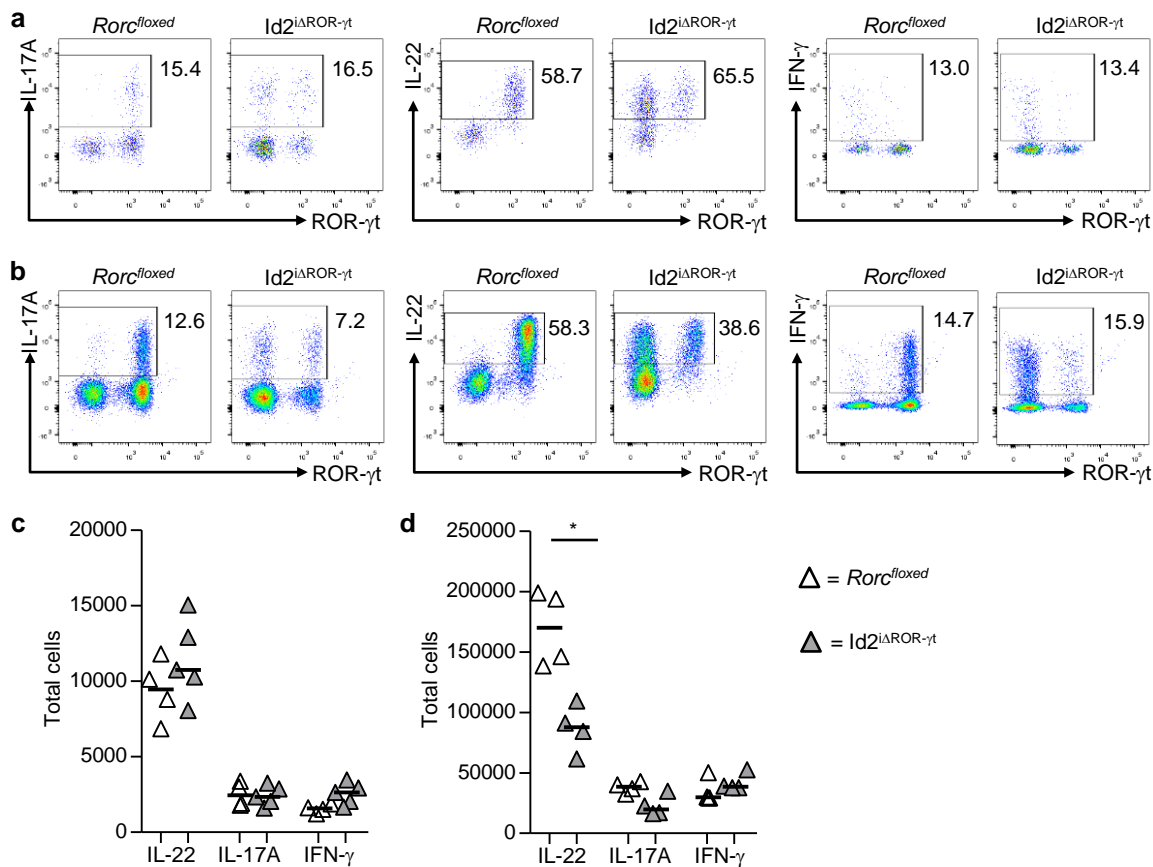
Supplementary Figure 2. Id2^{ΔROR-γt} mice are protected from infection with *C. albicans*. (a) Cre activity in CD127⁺ ILCs was assessed using ROSA^{mT/mG} control and Id2^{creERT2} × ROSA^{mT/mG} mice maintained on tamoxifen containing food for 3 weeks, $n = 5$ ROSA^{mT/mG} control and 6 Id2^{creERT2} × ROSA^{mT/mG} mice. Data pooled from two independent experiments and assessed using Mann-Whitney two tailed non-parametric test. * $P < 0.05$, ** $P < 0.005$; bar represents median. (b,c) Mice were infected with *C. albicans*, treated daily with oral tamoxifen starting on day -1, and sacrificed on day 3. (b) Identification of IL-17A producing ILC3 and (c) Periodic acid-Schiff stained histologic sections of the kidney. Scale bar, 200 μ m.



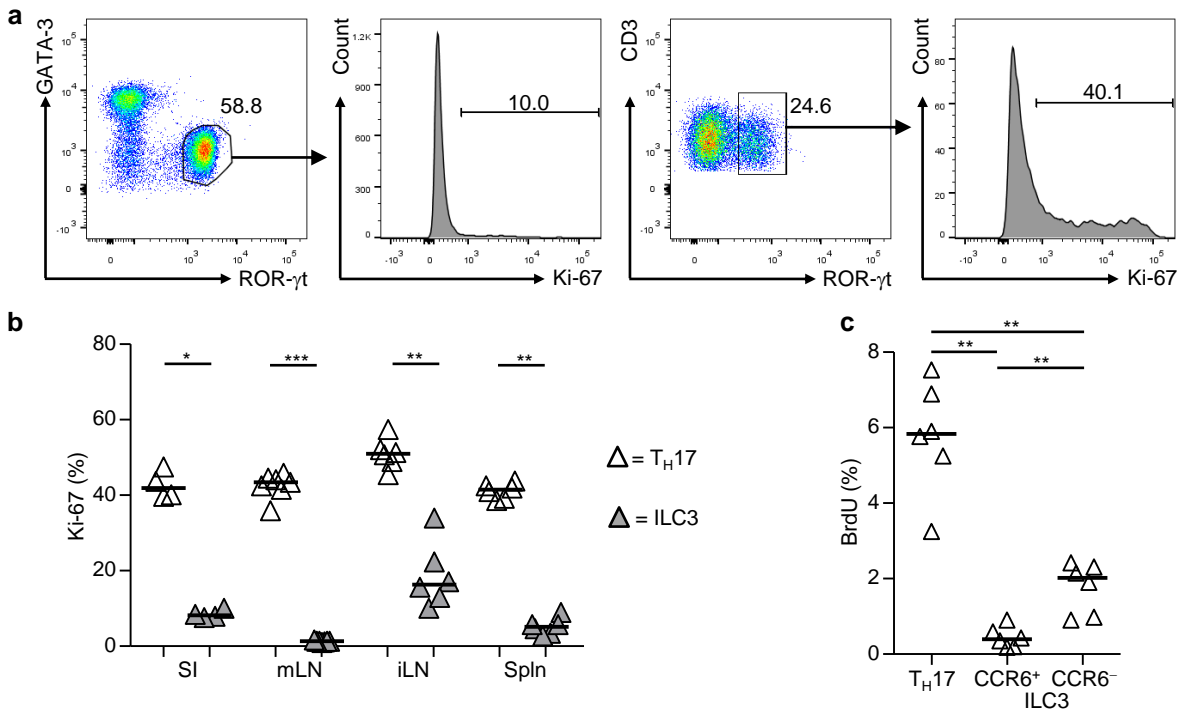
Supplementary Figure 3. Phenotypic analysis of ROR-γt⁺ and ROR-γt⁻ ILC3 populations in the mLN following genetic deletion of *Rorc* for 12 weeks. *Rorc*^{flxed} and *Id2*^{ΔROR-γt} mice were maintained on tamoxifen for 12 weeks and ILCs isolated from the mLN and characterized. (a) Gating strategy to identify ILC3 through exclusion of ILC2 (GATA-3⁺) and ILC1 (NKp46⁺). CD3i, CD3 intracellular. ROR-γt⁺ ILC3 in control *Rorc*^{flxed} mice shown in black, ROR-γt⁻ ILC3 in control *Id2*^{ΔROR-γt} mice shown in blue. (b) Overlays of histograms of MHCII, CCR6, CD25, c-kit and CD90 (Thy1) expression by ROR-γt⁺ ILC3 (black) and ROR-γt⁻ ILC3 (blue) as identified in (a). Total number of ILC and ILC3 (GATA-3⁻NKp46⁻ ILC) (c) and ROR-γt⁺ and ROR-γt⁻ ILC3 (d), bars show median, *n* = 4 (*Rorc*^{flxed}) and 8 (*Id2*^{ΔROR-γt}) mice. (e) Analysis of MFI of MHCII, CCR6, CD25, c-kit and CD90 (Thy1) protein by ROR-γt⁺ (white) and ROR-γt⁻ (grey) ILC3, mean and SEM shown, *n* = 4 (*Rorc*^{flxed}) and 6 (*Id2*^{ΔROR-γt}) mice. Data pooled from two independent experiments and assessed using Mann-Whitney two tailed non-parametric test. **P* < 0.05, ***P* < 0.005; bar represents median or SEM in (e).



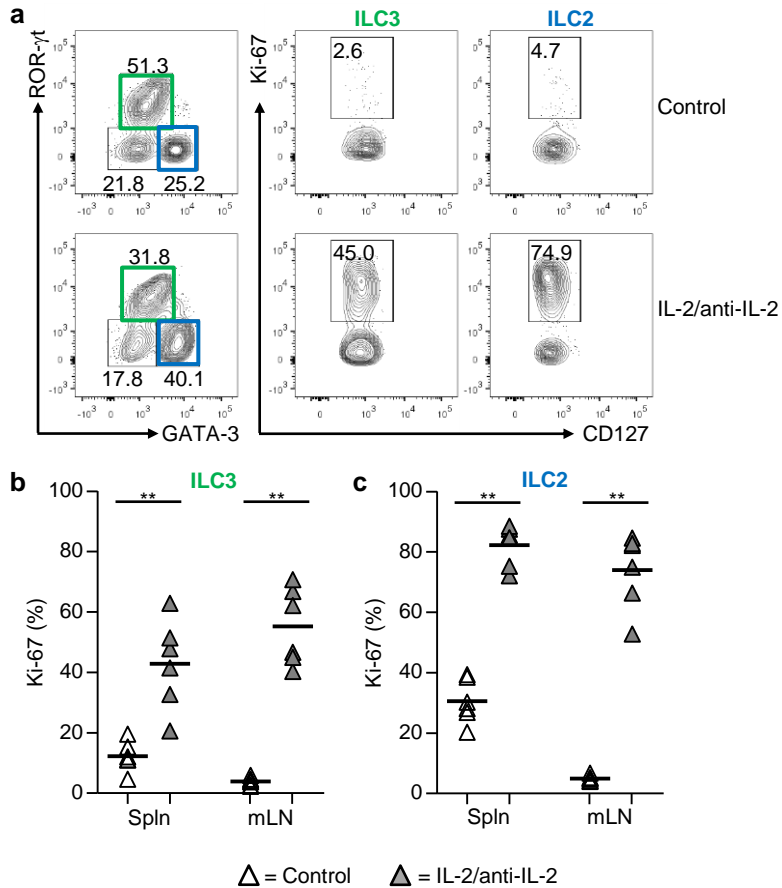
Supplementary Figure 4. Persistence of functional ILC3 in the mLN following genetic deletion of *Rorc* for 12 weeks. *Rorc^{flxed}* and *Id2^{iΔROR-γt}* mice were maintained on tamoxifen containing food for 12 weeks, cells from the mLN and siLP isolated and then stimulated *ex vivo* to assess cytokine expression. **(a)** Absolute numbers of IL-17A⁺, IL-22⁺, and IFN-γ⁺ cells that were ROR-γt⁺, ROR-γt⁻ or total in the small intestine lamina propria (siLP) of *Rorc^{flxed}* (*n* = 4) and *Id2^{iΔROR-γt}* (*n* = 6). **(b)** Gating strategy to assess non-ILC2 (GATA-3⁻) ILC populations in siLP using ROR-γt⁺ and NKp46. ROR-γt⁺ (black) ROR-γt⁻NKp46^{hi} (blue) and ROR-γt⁻NKp46⁻ (red) cells identified and then enumerated **(c)**, bars show median, *n* = 4 (*Rorc^{flxed}*) and *n* = 6 (*Id2^{iΔROR-γt}*) mice. **(d)** Expression of IL-17A, IL-22 and IFN-γ versus ROR-γt in CD45⁺CD127⁺Lin⁻ cells from the mLN, representative plots shown from 2 independent experiments. **(e)** Absolute numbers of IL-17A⁺, IL-22⁺, and IFN-γ⁺ cells that were ROR-γt⁺, ROR-γt⁻ or total in mLN of *Rorc^{flxed}* (*n* = 4) and *Id2^{iΔROR-γt}* (*n* = 6). Data from one of two independent experiments shown. Data assessed using Mann-Whitney two-tailed non-parametric test. **P* < 0.05, ***P* < 0.005.



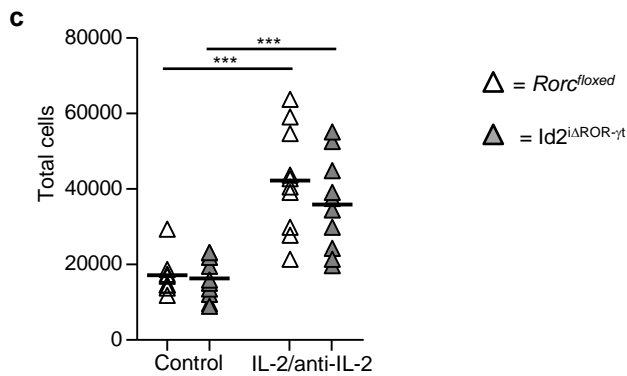
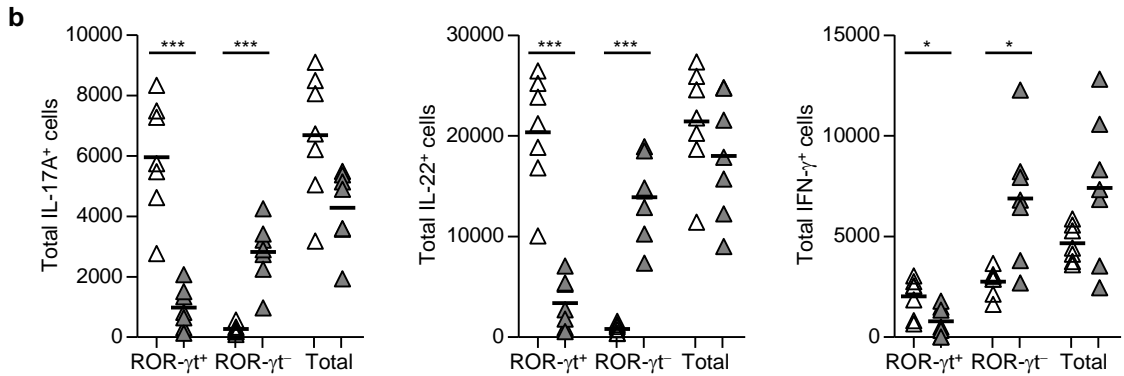
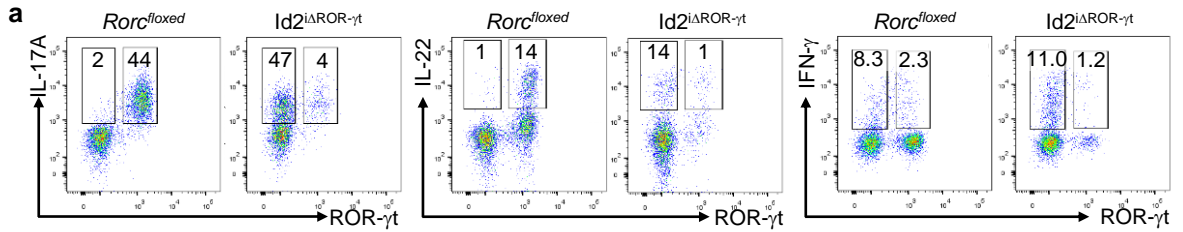
Supplementary Figure 5. ILC3 half-life differs in the mLN and small intestine. *Rorc^{flxed}* and *Id2^{ΔROR-γt}* mice were maintained on food containing tamoxifen for 2 weeks, then normal food for a further 4 weeks. Cells were prepared from mLN and the small intestine lamina propria (siLP) and then stimulated to assess cytokine and ROR-γt expression. IL-17A, IL-22 and IFN-γ protein versus ROR-γt by CD45⁺CD127⁺Lin⁻ cells isolated from (a) mLN and (b) siLP, representative plots shown from 2 independent experiments. Total numbers of IL-17A⁺, IL-22⁺, and IFN-γ⁺ in mLN (c) and siLP (d) of *Rorc^{flxed}* ($n = 4$) and *Id2^{ΔROR-γt}* (mLN $n = 5$, siLP $n = 4$) mice, data pooled from two independent experiments. Data assessed using Mann-Whitney two-tailed non-parametric test. * $P < 0.05$; bar represents median.



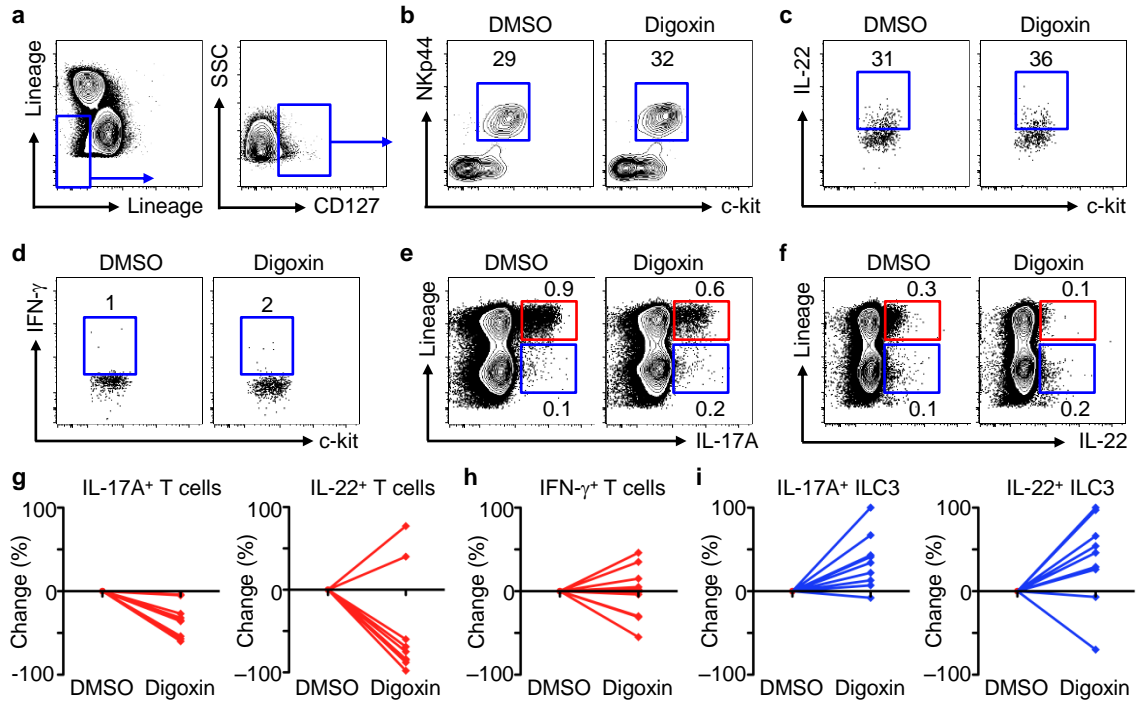
Supplementary Figure 6. T_H17 cells proliferate more than ILC3 in the small intestine. ILC3 and T_H17 populations from different tissues were assessed for evidence of proliferation. **(a)** Expression of Ki-67 by ILC3 (CD45⁺Lin⁻CD127⁺ROR- γ t⁺) cells and T_H1 (CD45⁺CD4⁺CD3⁺ROR- γ t⁺) cells isolated from the small intestine lamina propria (SI) of C57BL/6 mice, data representative of 4 mice. **(b)** Percentage Ki-67⁺ T_H17 and ILC3 in siLP, mLN, inguinal (i) LN and spleen (spln), data pooled from two independent experiments ($n = 4, 7, 6, 6$). **(c)** Percentage of BrdU⁺ T_H17 CCR6⁺ and CCR6⁻ ILC3 in siLP, 18 hours after 1.5 mg BrdU injected i.p. Data pooled from two experiments, $n = 6$ C57BL/6 mice. Data assessed using Mann-Whitney two-tailed non-parametric test. ** $P < 0.05$, * $P < 0.005$; bar represents median.



Supplementary Figure 7. Treatment with IL-2/anti-IL-2 complexes induces ILC2 and ILC3 proliferation. C57BL/6 mice received PBS or IL-2/anti-IL-2 complexes (1 μ g IL-2 and 5 μ g anti-IL-2 monoclonal antibody per injection per mouse) i.p. for three consecutive days and the spleen and mLN were then analyzed on the following day. **(a)** Representative plots of Ki-67 expression by ILC3 and group 2 ILCs (ILC2). Quantitation of ILC3 **(b)** and ILC2 **(c)** expressing Ki-67 in spleen and mLN. Data ($n = 6$) pooled from two independent experiments. Data assessed using Mann-Whitney two-tailed non-parametric test. ** $P < 0.005$; bar represents median.



Supplementary Figure 8. IL-2/anti-IL-2 complexes drive expansion of functional ILC3s in the absence of ROR- γ t expression. *Rorc*^{flxed} and *Id2*^{ΔROR- γ t} mice were maintained on tamoxifen for 10 weeks, given PBS or IL-2/anti-IL-2 complexes i.p. for three consecutive days, and then analyzed 5 days after the final injection. (a) Representative plots of the gating strategy and expression of IL-17A, IL-22 and IFN- γ by ILC from the mLN after stimulation. (b) Quantification of numbers of ROR- γ t⁺, ROR- γ t⁻ and total IL-17A, IL-22 and IFN- γ expressing ILC. Data ($n = 7$) pooled from two independent experiments. Bars show median. (c) Total numbers of ILC in mLN of control and IL-2/anti-IL-2 treated mice. Data pooled ($n = 8-10$) from 3 independent experiments and assessed using Mann-Whitney two-tailed non-parametric test. ** $P < 0.005$.



Supplementary Figure 9. Transient inhibition of ROR- γ t selectively reduces T_H17 cells in intestinal tissues from pediatric individuals with Crohn's diseases. (a–i) Intestinal resection tissue from pediatric IBD patients was cultured with Digoxin or DMSO control for 12 hours and examined for T_H17 cell and ILC3 responses. (a) Identification of Lineage-IL7R α ⁺ ILCs. (b) Percentage of NKp44⁺c-kit⁺ ILC3s. (c) Production of IL-22 and (d) IFN- γ by gated c-kit⁺ ILC3s. (e) Expression of IL-17A and (f) IL-22 by Lineage⁺ T_H17 cells (red) and Lineage⁻ ILC3s (blue). (g) Fold change in the percentage of IL-17A⁺ or IL-22⁺ T_H17 cells, (h) IFN- γ ⁺ T_H1 cells and (i) IL-17A⁺ or IL-22⁺ ILC3. All data are representative or shown for $n = 10$ pediatric individuals with Crohn's disease.