UNIVERSITYOF BIRMINGHAM

Research at Birmingham

The accuracy of cell-free fetal DNA based noninvasive prenatal testing in singleton pregnancies: a systematic review and bivariate meta-analysis

Mackie, Fiona; Hemming, Karla; Allen, Stephanie; Morris, R. Katie; Kilby, Mark; MacKie, Fiona

DOI:

10.1111/1471-0528.14050

Creative Commons: Attribution-NonCommercial-NoDerivs (CC BY-NC-ND)

Document Version Peer reviewed version

Citation for published version (Harvard):

Mackie, F, Hemming, K, Allen, S, Morris, RK, Kilby, M & MacKie, F 2016, 'The accuracy of cell-free fetal DNA based non-invasive prenatal testing in singleton pregnancies: a systematic review and bivariate meta-analysis', BJOG: An International Journal of Obstetrics & Gynaecology. https://doi.org/10.1111/1471-0528.14050

Link to publication on Research at Birmingham portal

Publisher Rights Statement:
Checked for eligibility: 27/04/2016. This is the peer reviewed version of the following article: Mackie FL, Hemming K, Allen S, Morris RK, Kilby MD. The accuracy of cell-free fetal DNA-based non-invasive prenatal testing in singleton pregnancies: a systematic review and bivariate meta-analysis. BJOG 2016; DOI: 10.1111/1471-0528.14050., which has been published in final form at http://onlinelibrary.wiley.com/doi/10.1111/1471-0528.14050/full. This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for Self-Archiving.

General rights

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

- Users may freely distribute the URL that is used to identify this publication.
- · Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
- User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

Download date: 01. Feb. 2019

- 1 The accuracy of cell-free fetal DNA based non-invasive prenatal testing in
- 2 singleton pregnancies: a systematic review and bivariate meta-analysis
- Fiona L Mackie (Clinical Research Fellow)¹, Karla Hemming (Senior Lecturer Public
- 4 Health, Epidemiology and Biostatistics)², Stephanie Allen (Consultant Clinical Scientist
- 5 Genetics)³, R Katie Morris (Senior Lecturer/Honorary Consultant Maternal Fetal
- 6 Medicine)^{1,4}, Mark D Kilby (Professor Fetal Medicine)^{1,4}
- 7 1. Centre for Women's & Children Health and the School of Clinical and Experimental
- 8 Medicine, College of Medical and Dental Sciences, University of Birmingham,
- 9 Birmingham, B15 2TT, UK.
- 10 2. Public Health, Epidemiology and Biostatistics, School of Health and Population
- Sciences, College of Medical and Dental Sciences, University of Birmingham,
- Birmingham, B15 2TT, UK.
- 13 3. West Midlands Regional Genetics Laboratory, Birmingham Women's Hospital NHS
- Foundation Trust, Mindelsohn Way, Edgbaston, Birmingham, B15 2TG,UK.
- 15 4. Fetal Medicine Centre, Birmingham Women's Hospital NHS Foundation Trust,
- Birmingham, B15 2TG, UK.
- 18 Corresponding Author: Dr Fiona Mackie. 3rd Floor Academic Department,
- 19 Birmingham Women's Hospital NHS Foundation Trust, Mindelsohn Way, Edgbaston,
- 20 Birmingham, B15 2TG, UK. fionamackie@doctors.org.uk +44-121-626-4535
- 22 **Running title**: Cell-free fetal DNA based NIPT in singleton pregnancies
- 24 Word Count: **4295**

27

17

21

23

25

26

- 29 Abstract
- 30 Background. Cell-free fetal DNA (cffDNA) non-invasive prenatal testing (NIPT) is
- 31 rapidly expanding and being introduced at varying rates depending on country and
- 32 condition.
- 33 **Objectives.** Determine accuracy of cffDNA-based NIPT for all conditions. Evaluate
- influence of other factors on test performance.
- 35 **Search strategy**. Medline, Embase, CINAHL, Cochrane Library, 1997-April 2015.
- 36 Selection criteria. Cohort studies reporting cffDNA-based NIPT performance in
- 37 singleton pregnancies.
- 38 **Data collection and analysis**. Bivariate or univariate meta-analysis and sub-group
- analysis performed to explore influence of test type and population risk. .
- 40 Main results. 117 studies included which analysed 18 conditions. Bivariate meta-
- 41 analysis demonstrated sensitivities and specificities respectively for: fetal sex
- 42 0.989(95%CI 0.980-0.994) and 0.996(95%CI 0.989-0.998) 11,179 tests; Rhesus D
- 43 0.993(0.982-0.997) and 0.984(0.964-0.993) 10,290 tests; trisomy 21 0.994(0.983-
- 44 0.998) and 0.999(0.999-1.00) 148,344 tests; trisomy 18 0.977(0.952-0.989) and
- 45 0.999(0.998-1.00) 146,940 tests; monosomy X 0.929(0.741-0.984) and 0.999(0.995-
- 46 0.999) 6,712 tests. Trisomy 13 was analysed by univariate meta-analysis with a
- 47 summary sensitivity of 0.906(95%CI 0.823-0.958) and specificity of 1.00(95%CI 0.999-
- 48 0.100) 134,691 tests. False and inconclusive results were poorly reported across all
- 49 conditions. Test type did affect sensitivity and specificity, but there was no evidence
- that population risk did.
- 51 Conclusions. Performance of cffDNA-based NIPT is affected by condition under
- 52 investigation. For fetal sex and Rhesus status NIPT can be considered diagnostic. For
- trisomy 21, 18 and 13, the lower sensitivity, specificity and disease prevalence
- 54 combined with the biological influence of confined placental mosaicism designates it a
- screening test. These factors must be considered when counselling patients and
- assessing the cost of introduction into routine care.

Systematic review registration. PROSPERO CRD42014007174

- **Keywords.** cell-free fetal DNA, non-invasive prenatal testing, diagnostic accuracy
- Tweetable abstract. cffDNA NIPT accuracy high, can be diagnostic for fetal sex and
- Rhesus, but only screening test in aneuploidy

Introduction

63

64

65 maternal plasma and believed to originate from trophoblast. It was first detected by Lo 66 et al. in 1997 (1) and used to note the presence of the Y chromosome to diagnose fetal 67 sex. NIPT can now be used to test for an euploidy, and single gene disorders such as 68 cystic fibrosis, Huntington's disease or thanatophoric dysplasia (2-6). Its advantage is 69 that it is non-invasive, avoiding the 0.5-1% risk of miscarriage associated with 70 amniocentesis/chorionic villus sampling (7) and allows timely therapeutic intervention in 71 conditions such as congenital adrenal hyperplasia (CAH) (8). cffDNA is cleared from 72 plasma (in hours) following delivery ensuring individuality for each pregnancy (9). Non-73 invasive prenatal testing also has health economic implications eliminating the need to 74 give all Rhesus negative women anti-D immunoglobulin prophylaxis. NIPT is being introduced into routine antenatal care across the world at differing 75 76 speeds, largely influenced by technological advances facilitated by the commercial 77 sector. Current guidance in North America and from the International Society for 78 Prenatal Diagnosis advises a positive NIPT for an euploidy to be confirmed by invasive 79 testing (10-12) due to the low risk of a false positive result secondary to confined 80 placental mosaicism (CPM). Inconclusive results occur in up to 8.1% (10), with a repeat 81 sample being successful in up to 80% participants (13). 82 Several systematic reviews and meta-analyses evaluating test accuracy have been 83 published (14-18). However these have several limitations: i) they evaluate individual 84 conditions (e.g. fetal sex, Rhesus status or aneuploidy) thus not allowing comparison; 85 ii) have a high risk of bias as they include case-control studies; iii) utilise inferior 86 statistical techniques for meta-analysis and iv) include studies with a significant risk of 87 verification bias due to all participants not receiving a reference test (e.g. karyotype). 88 The aim of our paper is to produce the most comprehensive systematic review and 89 meta-analysis of NIPT and address these issues: include only cohort studies to reduce 90 bias (19); perform bivariate meta-analysis where possible and thirdly to encompass all

Non-invasive prenatal testing (NIPT) utilises cell-free fetal DNA (cffDNA) present in

indications for antenatal use, so as to enable a more uniformed comparison for the use of NIPT in clinical practice. We also aim to assess aspects of test accuracy that might influence how cffDNA is implemented in the clinical pathway e.g. effect of technique on accuracy and evaluation of false positive, false negative and inconclusive results.

examined.

Methods

This review was performed according to recommended methods (20-23) and an *a priori* designed and registered protocol (PROSPERO CRD42014007174).

Identification of studies

Medline, Web of Science, Embase, CINAHL and the Cochrane Library databases were searched for relevant articles by FLM. Grey literature and reference lists were hand searched. The search terms used were 'noninvasive', 'non-invasive', 'non invasive', 'prenatal diagnosis', 'cell free fetal DNA' and 'cell-free fetal DNA'. The full search strategy is available as online supplementary material (Appendix S1). The date of publication was limited from 1997 to 13 April 2015. There was no limitation on language.

Study selection

Study selection was performed in duplicate (FLM, RKM) involving screening of titles and abstracts, then reviewing full manuscripts of selected articles. Disagreements in selection were resolved by MDK. Articles were included based on the following criteria: *Population:* Women with a singleton pregnancy, any gestation. Populations could include women of varying risk with high-risk women defined as attending for testing due to pre-existing risk factors: a personal or family history of the condition being tested for, high-risk on routine biochemical screening, abnormal ultrasound scan, and/or raised maternal age. Women were considered low-risk if they had none of the above risk factors. *Test:* NIPT based on cffDNA in maternal blood, irrespective of condition being

119 Reference standard: Studies must have compared all the cffDNA results with either: 120 karyotype results or birth outcome (either blood sample or phenotype) as appropriate in 121 all participants. 122 Study design: Cohort studies. 123 Exclusion criteria: pre-implantation testing, fetal cell testing, case-control studies, case 124 series with <5 participants. 125 Data extraction 126 Data were extracted in duplicate on the relevant 2x2 tables comparing the non-invasive 127 test with the reference test used for definitive diagnosis. Data were also extracted on 128 factors which may affect test accuracy: participant characteristics (e.g. obstetric history); and test characteristics (e.g. cut offs used, test technique [e.g. PCR, MPS, 129 130 mass spectrometry]). Information regarding false results and inconclusive results was 131 obtained. When a study used similar laboratory protocols on the same blood samples (e.g. 132 133 different number of replicates performed) only the best results were included. When a 134 study used different laboratory protocols on different blood samples, but the same type 135 of test technique, these samples were grouped together for analysis. If a study sub-136 divided samples based on population characteristics (e.g. high-risk vs. low-risk for a condition, or 1st trimester vs. 2nd trimester vs. 3rd trimester) these were grouped 137 together for the summary statistics, and analysed as a sub-group where appropriate. 138 139 Quality Assessment 140 The quality of the studies was assessed using the QUADAS-2 tool (24). 141 Data synthesis 142 For each study the 2x2 data were used to calculate sensitivity and specificity with 95% 143 confidence intervals. Heterogeneity was explored by assessing the distribution of 144 results in the Forest plots and summary receiver operating characteristic curves 145 (SROC). Summary measures including sensitivities, specificities, diagnostic odds ratio, 146 positive and negative likelihood ratios along with 95% confidence intervals were

calculated using bivariate logistic regression model with an unstructured correlation. This model allows for the correlation between sensitivity and specificity from the same study and for the sensitivities and specificities to have different random effects (25). Meta-analysis was performed when there were more than 5 studies per condition using STATA 13 (StataCorp. 2012, College Station, Texas) (see Appendix S2 for more detail). Sub-group analysis and meta-regression was planned *a priori* to assess effects of study level covariates on test accuracy, namely: population characteristics (level of risk for condition where appropriate i.e. not performed in fetal sex or Rhesus D); test technique (e.g. PCR, MPS) and quality aspects according to QUADAS-2. We used sub-group analyses (as opposed to meta-regression) to assess the influence of all categorical covariates due to model convergence difficulties (26).

Results

The search revealed 4433 studies for inclusion. After reviewing the full article, 117 studies (1, 27-143) were eligible reporting on 18 different conditions, and 472.935 tests (Figure S1). The study characteristics are outlined in Table S1. We were able to produce summary results using the fully unstructured bivariate model for: fetal sex, Rhesus D, trisomy 21, trisomy 18 and monosomy X (Table S2). For trisomy 13, despite a sufficient number of studies (n=15) there was no heterogeneity in specificities across studies so the bivariate model, which takes into account the correlation between the sensitivities and specificities, failed to converge and consequently we fitted a univariate model. Because of this, these results are less methodologically robust. The HSROC curves are presented in Figure S2 and the results from our sub-group analyses in Table S2. There were 5 studies (n=394,130 tests) in which there was differential verification of results, in that some participants had their result confirmed by karyotype and others by phenotype (35, 91, 93, 114, 133). These 5 studies all assessed fetal aneuploidy and utilised NIPT as a screening test in a low-risk population. A sensitivity analysis

175 removing these 5 studies demonstrated no significant effect on the summary results, thus these studies are included in all analyses and Forest plots. 176 The following 12 conditions had insufficient studies for meta-analysis: Rhesus C, 177 178 Rhesus E, 47XXX, 47XXY, 47XYY, trisomy 16, congenital adrenal hyperplasia, deletion-duplication syndromes, sickle cell anaemia, thalassaemia, human platelet 179 antigen 1a, and KEL 1. The Forest plots of these 12 conditions are presented in Figure 180 181 S3. 183 Methodological quality of included studies 184

182

This was assessed according to the Quality Assessment tool for Diagnostic Accuracy Studies (QUADAS-2) (24), the results are demonstrated in Figure S4 and further described in Appendix S3.

187

188

189

190

191

192

193

194

195

196

197

185

186

False results and inconclusive results

Reporting of causes and implications of false positive, false negative and inconclusive results was poor, and varied across all conditions (Table S3). The included studies reported an inconclusive result rate of 0.32-5.3%. This issue was further compounded by a myriad of varying quality control (QC) standards, some studies excluding samples that failed their QC and others implementing no QC steps and therefore reporting some results as false negatives which other studies would have excluded from analysis. Some studies investigated the reasons for their false and inconclusive results and reported these clearly, accounting for all samples. Other studies reported inconclusive results as false negatives or did not report them at all. We describe these results in more detail for each of the conditions investigated.

199

200

198

Results from bivariate meta-analysis

201

202

Fetal Sex

Sixty studies (11,179 tests) evaluated fetal sex and are represented in the Forest plot in Figure 1. Bivariate meta-analysis produced a summary sensitivity of 0.989 (95% CI 0.980 to 0.994) and specificity of 0.996 (95% CI 0.989 to 0.998), a positive likelihood ratio of 255 (95% CI 89 to 729) and negative likelihood ratio of 0.011 (95% CI 0.006 to 0.019). Other summary measures are in Table S2. No significant effect on sensitivity was found with test technique. However there was a difference in specificity with real-time quantitative PCR 0.999 (95%CI 0.991 to 1.00) performing better than conventional PCR 0.939 (95%CI 0.872 to 0.972). For fetal sex, 11/60 studies reported inconclusive results, of these, 5 studies documented an explanation (in order of frequency): assay failure, no reason given, insufficient number of markers present from pre-specified cut-off and low fetal fraction. The commonest reasons given by the authors of the studies for the false results were: no reason given, low fetal fraction (although cffDNA not quantified), low fetal fraction confirmed by authors quantifying cffDNA, possible contamination/DNA degradation/vanishing twin/test failure although not confirmed, and previous male pregnancy, although the latter reason has since been disproven as cell-free fetal DNA is cleared from the maternal circulation hours post-delivery (9).

Rhesus D

Thirty studies (10,290 tests) evaluated fetal Rhesus D status and are represented in Figure 2. Bivariate meta-analysis produced a summary sensitivity of 0.993 (95% CI 0.982 to 0.997) and specificity of 0.984 (95% CI 0.964 to 0.993) a positive likelihood ratio of 61 (95% CI 22 to 167) and negative likelihood ratio of 0.007 (95% CI 0.003 to 0.186). There was a significant difference between test techniques with real-time quantitative PCR sensitivity: 0.997 (95% CI 0.987 to 0.999) demonstrating a higher sensitivity than conventional PCR 0.924 (95%CI 0.832 to 0.968), although it was not possible to assess if there was a difference in those which utilised mass spectrometry (despite sufficient studies, due to convergence issues as detailed in the discussion),

and no difference in specificity was seen (Table S2). For Rhesus D, 13/30 studies reported inconclusive results, of these, 10 studies documented an explanation (in order of frequency): no reason given, RHD gene variant, insufficient number of markers present from pre-specified cut-off, test failure, low fetal fraction. The commonest reasons given for false results were: presumed low fetal fraction (although not quantified by authors), no reason given, presumed RHD gene variant (although not confirmed), confirmed RHD gene variant, test failure, possible contamination/DNA degradation/pipetting error/incorrect neonatal blood testing.

Trisomy 21

Thirty-one studies (148,344 tests) assessed trisomy 21 and are represented in Figure 3A. Bivariate meta-analysis produced a summary sensitivity of 0.994 (95% CI 0.983 to 0.998) and specificity of 0.999 (95% CI 0.999 to 1.00) a positive likelihood ratio of 1720 (95% CI 1111 to 2662) and negative likelihood ratio of 0.006 (95% CI 0.002 to 0.017). Test technique and population risk had no significant effect. For trisomy 21, 14/31 studies reported inconclusive results, of these, 7 studies documented an explanation (in order of frequency): assay failure, confirmed low fetal fraction, no reason given, presumed low fetal fraction/inadequate sequencing depth. The commonest reasons given for false results were: confirmed low fetal fraction, confirmed mosaicism, no reason given, test failure, maternal CNV.

Trisomy 18

Twenty-four studies (146,940 tests) assessed trisomy 18 and are represented in Figure 3B. Bivariate meta-analysis produced a summary sensitivity of 0.977 (95% CI 0.952 to 0.989) and specificity of 0.999 (95% CI 0.998 to 1.00) and a positive likelihood ratio of 1569 (95% CI 810 to 3149) and negative likelihood ratio of 0.023 (95% CI 0.011 to 0.048). Neither test technique or population risk had a significant effect. For trisomy 18, 12/24 studies reported inconclusive results, of these 7 studies documented an

explanation (in order of frequency): low fetal fraction, test failure, no reason given, mosaicism. The commonest reasons given for false results were: confirmed low fetal fraction, confirmed mosaicism, presumed low fetal fraction/human error, maternal CNV, no reason given.

Monosomy X

Eight studies (6712 tests) assessed monosomy X and are represented in Figure 3C. Bivariate meta-analysis produced a summary sensitivity of 0.929 (95% CI 0.741 to 0.984) and specificity of 0.999 (95% CI 0.995 to 0.999) and a positive likelihood ratio of 1337 (95% CI 213 to 8407) and negative likelihood ratio of 0.071 (95% CI 0.017 to 0.292). There was no significant difference with test technique. It was not possible to assess the effect of population risk as there were insufficient low-risk studies. For monosomy X, 5/8 studies reported inconclusive results, of these, 3 studies documented an explanation (in order of frequency): low fetal fraction, presumed human error and no reason given. The commonest reasons given for false results were: mosaicism and no reason given.

The 5 aneuploidy studies which evaluated an unselected obstetric population reported inconclusive results rates of 0.29-5.1% and provided the same reasons for their false and inconclusive results as with the high-risk aneuploidy populations.

Trisomy 13 – univariate meta-analysis

Sixteen studies which equates to 134,691 tests examined trisomy 13, represented in Figure 3D. There was a summary sensitivity of 0.906 (95% CI 0.823 to 0.958) and specificity of 1.00 (95% CI 0.999 to 1.00). The positive likelihood ratio was 453 (95% CI 26 to 7864) and negative likelihood ratio was 0.188 (95% CI 0.080 to 0.44039) with a diagnostic odds ratio of 2788 (95% CI 285 to 27252). For trisomy 13, 6/16 studies reported inconclusive results, of these, 4 studies documented an explanation for

inconclusive results: low fetal fraction, different fragmentation rate, contamination, assay failure and human error. The only reason given for false results was confirmed low fetal fraction.

290

291

292

287

288

289

Results where meta-analysis not possible

The results for these conditions are presented as Forest plots in S3.

293

294

295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

Clinical application for NIPT for Down's syndrome screening

Using published data from the National Down Syndrome Cytogenetic Register (NDSCR) 2012 Annual report we have produced a table detailing the estimated outcomes (livebirth rate, invasive test rate, euploid pregnancy loss rate, undiagnosed aneuploidy livebirth rate) from the current standard Down's Syndrome Screening (DSS) i.e. first trimester combined screening pathway (maternal age, nuchal translucency, beta human chorionic gonadotrophin and pregnancy associated plasma protein A) and from a pathway with NIPT as both contingent (i.e. NIPT offered to women with a positive screen after first trimester combined screening) and first line screening for a population of 100,000 women using crude rates (144) (Table S4). We use the prevalence reported by NDSCR¹ (trisomy 21: 2.2 per 1000 women, trisomy 18: 0.64 per 1000, trisomy 13 0.26 per 1000). This assumes that standards for the first trimester combined screening are "achievable" as described by Fetal Anomaly Screening Programme (FASP) guidance i.e. for trisomy 21 a detection rate of 85% for a screen positive rate of 2% (145). For NIPT the summary measures are those from our meta-analysis. For the contingent screening model the cut-off for high risk is 1:1000 from first trimester combined screening with a detection rate of 96% and false positive rate of 12% (146). This model assumes that all women accept screening when offered as it is not possible to determine yet what the uptake of NIPT would be if offered as a first-line test. It also assumes that all women are required to have an invasive test for karyotyping after a screen positive result from combined or NIPT prior to considering

termination of pregnancy, thus the invasive test rates will be higher than in a real-life population. It assumes a 0.5% pregnancy loss rate from invasive testing (146).

317

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

315

316

These data demonstrate the influence of disease prevalence on test performance. If we compare combined screening with a 1:150 cut-off (i.e. current NHS practice) with NIPT as a first–line test we can reduce the invasive test rate from 2000 to 319 per 100,000 women, the euploid pregnancy loss rate from 9 to 1 per 100,000 and the undiagnosed trisomy 21 live births rate from 32 to 1 per 100,000. If NIPT was used as a contingent screening test for a 1:1000 combined screening cut-off (i.e. as a 2nd test following a positive combined screening result at a 1:1000 cut-off) then these figures are reduced even further compared to combined screening with a 1:150 cut-off: 2000 to 222 per 100,000 women invasive test rate; 9 to 0 euploid pregnancy loss rate, although there is less of a reduction in undiagnosed trisomy 21 live birth rate from 32 to 10. If NIPT was used as a contingent screening test for a 1:150 combined screening cut-off then these figures are: 2000 per 100,000 women invasive test rate; 0 euploid pregnancy loss and 34 undiagnosed trisomy 21 livebirth rate. A two stage contingent screening pathway with a 1:1000 cut-off when compared to NIPT as a first line test affords a reduction in false positive results (12 versus 100 per 100,000 women) that are found at the time of NIPT as the prevalence of disease in the population now undergoing NIPT is much higher. This is at the expense of a 10 fold increase in undiagnosed aneuploidy live births (1 versus 10 per 100,000 women) due to the increased number of false negatives at the first stage of screening that do not undergo NIPT. A cut-off of 1:150 at the first stage for the combined test compared to a 1:150 cut-off for NIPT as a contingent screening test has little effect on the number false negatives (33 versus 34), however the invasive test rate is reduced (2000 versus 188 per 100,000 women).

340

341

342

339

Discussion

Main findings

Our results demonstrate that for fetal sex and Rhesus D status, cffDNA-based NIPT has a high sensitivity and specificity. For aneuploidies: trisomy 21, and in particular trisomy 18 and 13 we have demonstrated improved accuracy from other recent systematic reviews likely due to technological developments. Importantly we found that false results and inconclusive results were poorly reported across all conditions.

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

366

367

368

369

370

347

343

344

345

346

Strengths and limitations

This review was performed according to rigorous methodology with efforts made to reduce bias in participant selection and clinical applicability by excluding case-control studies, performing bivariate meta-analysis and meta-regression analysis and assessing the impact of differential verification (i.e. different reference standards). Bivariate meta-analysis is the recommended approach for the meta- analysis of diagnostic test accuracy studies. This is because a conventional univariate analysis makes assumptions that are known not to be tenable (that the sensitivity and specificity from the same study are independent). However, the bivariate meta-analysis model is a technically difficult model to fit and it is well known that these models might not converge when there are a small number of studies, or when there are zero cells (i.e. sensitivity or specificity close to 100) (26). We observed no indication that other model fits were unstable and so have no reason to be concerned about the statistical validity of the other results. Our review also evaluates more conditions than previously. In addition, our paper has been able to assess the impact of test technique and population risk. We were unable to evaluate the number of samples which failed QC measures as this was reported in varying degrees. When considering the implementation of a new test, information regarding failed tests (147, 148), and inconclusive results is vital. We investigated the reasons for false positive and false negative results within and across studies and attempted to summarise these. This was again hampered by poor reporting with a common reason being low fetal fraction which is difficult to measure accurately and thus has led to variations in approach between

studies. It is especially important to consider this further as low fetal fraction has been shown to be associated with trisomy 18 and triploidies.

A limitation of this work is that it was not possible to account for the many subtle

A limitation of this work is that it was not possible to account for the many subtle differences in laboratory techniques such as comparing the different combinations of genetic markers used for each condition; or the myriad of adjustments made to bioinformatics algorithms as these were so varied. This is where the results from the large studies in screening populations are especially important as there is QC across laboratories and standardisation of techniques (35, 91, 93, 114, 133). In the process of publishing this review, the search was re-run from April 2015 - September 2015 in view of the rapid progression in this area. This yielded 78 new citations, of which 11 additional papers would be eligible for inclusion (3, 149-158), which comprise 10,191 women in total. These studies examine fetal sex (n=436 women), Rhesus D status (n=2965), trisomy 21 (n=6661), trisomy 18 (n=6701), trisomy 13 (n=6495), and monosomy X (n=40), which equate to a small proportion of additional tests, compared to the studies we have already analysed. There is also now one study which investigates than atophoric dysplasia (n=108), although this cannot be included in a meta-analysis as it is the only study to look at this condition thus far. As the search was under a year old when the publication was accepted we have not included these 11 studies in our results. We are confident that if these studies were included they would not impact on our results and conclusions.

391

392

393

394

395

396

397

398

371

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

388

389

390

Interpretation

It is recognised that there are fewer studies in our meta-analyses for trisomy 13 and monosomy X compared to a previous large meta-analysis (14) but this is due to excluding case-control studies and limiting to singletons. This has led to us reporting higher summary sensitivities and specificities than existing analyses, demonstrating how NIPT is advancing, and supporting the belief that NIPT will be used as the first-line screening test in the future. Our clinical application model has highlighted the

_-

importance of low prevalence of disease on the positive predictive value and false positive rate in the case of aneuploidies. Although positive and negative predictive values are useful indicators of test accuracy as they take into account disease prevalence (159), we have not presented these values within this paper due to variation in disease prevalence among included study populations.

404

405

406

407

399

400

401

402

403

Conclusion

This work demonstrates that there is a sufficient body of evidence for the accuracy and reproducibility of cffDNA-based NIPT to allow its introduction into routine clinical practice within the UK, however its role is yet to be decided.

409

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

408

Implications for clinical practice

The findings of this analysis support the use of NIPT as a diagnostic test for fetal sex and Rhesus status due to the nature of these conditions and the populations being tested. For assessment of aneuploidy the test must be considered a "screening test" despite high accuracy due to the low prevalence of disease and influence of biological factors such as CPM. We are aware that the National Screening Committee (NSC) is currently reviewing all the evidence for an euploidy, and is likely to recommend NIPT as a contingency screening test in the UK (Dr Pranav Pandya, Personal Communication, 2015). While for Down's syndrome screening (DSS) this will ensure access to an accurate, non-invasive test and ensure equity for many more women (i.e. test threshold has less of an impact on offering invasive testing and test can be offered throughout gestation not just in a small first trimester window) this must be balanced with consideration of the important ethical repercussions which need addressing (i.e. a test that can assess for multiple conditions and those with a milder phenotype and also test for conditions within the mother e.g. sex-chromosome anomaly or cancers) (160). There are also counselling implications as access to a non-invasive, highly accurate test still needs careful consideration by parents.

--

427

428

429

430

431

432

433

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

Implications for future research

The authors would recommend that the same rigorous assessment of the evidence and accuracy as we have performed be applied in multiple pregnancies once the evidence base is sufficient. The NIHR funded RAPID study which has used NIPT in an NHS setting for women in whom combined testing gave a risk of ≥ 1:1000 will soon be published. This study aims to assess the uptake of NIPT and whether the addition of NIPT to the DSS pathway affects the uptake of DSS and invasive testing; a detailed health economic evaluation using a tool developed in conjunction with the UK NSC; optimal ways to deliver education to women and healthcare professionals; and sensitivity and specificity of NIPT for an euploidy when performed in an NHS regional genetics laboratory. The results from our review indicate the latter (accuracy results from an NHS regional genetics laboratory) will be an important outcome as it will remove the influence of results from the commercial sector and poor reporting. This will allow for improved QC. enable continued assessment on a national basis, and ensure that the cost of NIPT will improve further. Similarly, the conditions for which NIPT will be used are likely to increase: 11 studies which examined single gene mutations and microdeletions could not be included in our meta-analysis due to having fewer than 5 participants; even whilst writing this review larger studies are being reported on these conditions (161). However, an economic evaluation of this first-line screening with NIPT would also need to include maintaining access to a high quality first trimester ultrasound scan including nuchal translucency (NT) assessment, to allow dating, viability, multiple pregnancy, structural anomaly and adnexal assessment, and importantly the assessment of the risk of cardiac anomalies and increased pregnancy loss associated with raised NT.

452

154	Acknowledgements. The articles were translated by FLM and RNM, and Di Carman
455	Lai. Some of these data have been presented at the British Maternal and Fetal
456	Medicine Society Annual Scientific Conference, 2015 (Mackie FL, Morris RK, Hemming
457	K, Allen S, Kilby MD. Cell-free fetal DNA based non-invasive prenatal testing: a
458	systematic review and meta-analysis of diagnostic accuracy. Br J Obstet Gynecol
159	2015;122:Supp 2)
460	
461	Disclosure of interest: We have no disclosures of interests to declare. The ICMJE
162	disclosure forms are available as online supporting information.
463	
164	Contribution to authorship: FLM extracted the data, contributed to the analysis and
465	data interpretation, and drafted the manuscript. RKM assisted extracting the data,
466	contributed to the analysis and data interpretation and amended the manuscript. KH
467	conducted the bivariate meta-analysis and data interpretation and amended the
468	manuscript. SA assisted with data extraction, interpretation of the results and amended
169	the manuscript. MDK conceived, designed and oversaw the work, made final decisions
470	where there were discrepancies and amended the manuscript. MDK is guarantor for
471	the study.
172	
473	Details of ethical approval: not required
174	
175	Funding: FLM is funded by the Richard and Jack Wiseman Trust (Registered charity
476	number: 1036690).
177	
178	References
179 180 181 182	 Lo Y, Corbetta N, Chamberlain P, Rai V, et al. Presence of fetal DNA in maternal plasma and serum. Lancet. 1997;350(9076):485-7. Bréchot P, Mouawia H, Saker A. Diagnostic prénatal non invasif de la mucoviscidose. Arch Pédiatr. 2011;18(1):111-8.

- 483 3. Chitty L, S, Barrett A, McKay F, Lench N, Daley R, Jenkins L. Non-invasive prenatal
- 484 diagnosis of achondroplasia and thanatophoric dysplasia: next-generation sequencing allows
- for a safer, more accurate, and comprehensive approach. Prenat Diagn. 2015;35:656-62
- 486
- 487 4. Bustamante-Aragones A, Trujillo-Tiebas M, Gallego-Merlo J, Rodriguez de Alba M,
- 488 Gonzalez-Gonzalez C, Cantalapiedra D, et al. Prenatal diagnosis of Huntington disease in
- 489 maternal plasma: direct and indirect study. European Journal of Neurology. 2008;15(12):1338-
- 490 44.
- 491 5. González-González M, Garcia-Hoyos M, Trujillo-Tiebas M, Bustamante Aragonés A,
- Rodriguez de Alba M, Alvarez D, et al. Improvement in strategies for the non-invasive prenatal
- diagnosis of Huntington disease. J Assist Reprod Genet. 2008;25(9-10):477-81.
- 494 6. Lench N, Barrett A, Fielding S, McKay F, Hill M, Jenkins L, et al. The clinical
- implementation of non-invasive prenatal diagnosis for single-gene disorders: challenges and progress made. Prenat Diagn. 2013;33(6):555-62.
- 7. Tabor A, Alfirevic Z. Update on procedure-related risks for prenatal diagnosis
- 498 techniques. Prenat Diagn. 2010;27(1):1-7.
- 499 8. Lo Y, Tein M, Lau T, Haines C, Leung T, al. e. Quantitative analysis of fetal DNA in
- maternal plasma and serum: implications for noninvasive prenatal diagnosis. Am J Hum Genet.1998.
- 502 9. Lo Y, Zhang J, Leung T, Lau T, Chang A, Hjelm N. Rapid clearance of fetal DNA from
- 503 maternal plasma. Am J Hum Genet. 1999;64:218-24.
- 504 10. SMFM. Society for Maternal-Fetal Medicine (SMFM) Consult Series #36: Prenatal
- aneuploidy screening using cell free DNA. Am J Obstet Gynecol. 2015;epub ahead of print.
- 506 11. Langlois S, Brock J. SOGC Committee Opinion: Current status in non-invasive prenatal
- detection of Down syndrome, trisomy 18, and trisomy 13 using cell-free fDNA in maternal plasma. J Obstet Gynecol Can. 2013;35:177-81.
- 509 12. ISPD. Position statement from the chromosome abnormality screening committee on
- 510 behalf of the board of the International Society for Prenatal Diagnosis Charlottesville, VA:
- 511 International Society for Prenatal Diagnosis; 2015 [cited 06 May 2015]. Available from:
- 512 http://www.ispdhome.org/public/news/2015/PositionStatementFinal04082015.pdf.
- 513 13. Sonek J, Cuckle H. What will be the role of first-trimester ultrasound if cell-free DNA
- 514 screening for aneuploidy becomes routine. Ultrasound Obstet Gynecol. 2014;44:621-30.
- 515 14. Gil M, Quezada M, Revello R, Akolekar R, Nicolaides K. Analysis of cell-free DNA in
- 516 maternal blood in screening for fetal aneuploidies: updated meta-analysis. Ultrasound Obstet
- 517 Gynecol. 2015;45:249-66.
- 518 15. Devaney S, Palomaki G, Scott J, Bianchi D. Noninvasive fetal sex determination using
- 519 cell-free fetal DNA. JAMA. 2011;306:627-36.
- 520 16. Wright C, Wei Y, Higgins J, Sagoo G. Non-invasive prenatal diagnostic test accuracy for
- fetal sex using cell-free DNA a review and meta-analysis. BMC Res Notes. 2012;5:1-11.
- 522 17. Geifman-Holtzman O, Grotegut C, Gaughan J. Diagnostic accuracy of noninvasive fetal
- 523 Rh genotyping from maternal blood a meta-analysis. Am J Obstet Gynecol. 2006;195:1163-
- 524 75.
- 525 18. Zhu Y, Zheng Y, Li L, Zhou H, Liao X, Guo J, et al. Diagnostic accuracy of non-invasive
- 526 fetal RhD genotyping using cell-free fetal DNA: a meta-analysis. J Maten Fetal Neonatal Med.
- 527 2014;27(18):1839-44
- 528 19. Rutjes A, Reitsma J, Di Nusio M, Smidt N, van Rijn J, Bossuyt P. Evidence of bias and
- variation in diagnostic accuracy studies. CMAJ. 2006;174:469-76.
- 530 20. Cochrane. Cochrane methods working group on systematic reivews of screening and
- diagnostic tests: recommended methods. Cochrane, editor2011.
- 532 21. Deeks J. Systematic reviews in health care: systematic reviews of diagnostic and
- 533 screening tests. BMJ. 2001;323:157-62.

- 534 22. Khan K, Dinnes J, Kleijnen J. Systematic reviews to evaluate diagnostic tests. Eur J
- 535 Obstet Gynecol Reprod Biol. 2001;95:6-11.
- 536 23. Irwig L, Tosteson A, Gatsonis C, Lau J, Colditz G, Chalmers T. Guidelines for meta-
- analyses evaluating diagnostic tests. Ann Intern Med. 1994;120:667-76.
- 538 24. Whiting P, Rutjes A, Westwood M, Mallett S, Deeks J, Reitsma J, et al. QUADAS-2: A
- Revised Tool for the Quality Assessment of Diagnostic Accuracy Studies. Ann Intern Med.
- 540 2011;155(8):529-36.
- 541 25. Reitsma J, Glas A, Rutjes A, Scholten R, Bossuyt P, Zinderman A. Bivariate analysis of
- sensitivity and specificity produces informative summary measures in diagnostic reviews. J Clin
- 543 Epidemiol. 2005;58(982-90).
- 544 26. Takwoingi Y, Guo B, Riley R, Deeks J. Performance of methods for meta-analysis of
- diagnostic test accuracy with few studies or sparse data. Stats Methods Med Res. 2015.
- 546 27. Achargui S, Tijane M, Benchemsi N. Génotypage RHD fœtal par PCR dans le plasma de
- femmes enceintes D négatif. Transfusion Clinique et Biologique. 2011;18(1):13-9.
- 548 28. Aghanoori MR, Vafaei H, Kavoshi H, Mohamadi S, Goodarzi HR. Sex determination
- 549 using free fetal DNA at early gestational ages: a comparison between a modified mini-STR
- genotyping method and real-time PCR. Am J Obstet Gynecol. 2012;207(3):202.e1-e8.
- 551 29. Akolekar R, Farkas DH, VanAgtmael AL, Bombard AT, Nicolaides KH. Fetal sex
- determination using circulating cell-free fetal DNA (ccffDNA) at 11 to 13 weeks of gestation.
- 553 Prenat Diagn. 2010;30(10):918-23.
- 30. Alberti A, Salomon L, Le Lorc'h M, Couloux A, Bussieres L, Goupil S, et al. Non-invasive
- 555 prenatal testing for trisomy 21 based on analysis of cell-free fetal DNA circulating in the
- maternal plasma. Prenat Diagn. 2015;35:471-6.
- 557 31. Al-Yatama M, Mustafa A, Ali S, Abraham S, Khan Z, Khaja N. Detection of Y
- 558 chromosome-specific DNA in the plasma and urine of pregnant women using nested
- polymerase chain reaction. Prenat Diagn. 2001;21(5):399-402.
- 560 32. Al-Yatama M, Mustafa A, Al-Kandari F, Khaja N, Zohra K, Monem R, et al. Polymerase-
- Chain-Reaction-Based Detection of Fetal Rhesus D and Y-Chromosome-Specific DNA in the
- 562 Whole Blood of Pregnant Women during Different Trimesters of Pregnancy. Med Princ Pract.
- 563 2007;16(5):327-32.
- 564 33. Aykut A, Cotulu O, Onay H, Satol S, Ozkinay F. Determination of fetal rhd status by
- maternal plasma DNA analysis. Clin Genet. 2010;78:S108.
- 566 34. Barrett A, McDonnell T, Chan K, Chitty L. Digital PCR Analysis of Maternal Plasma for
- Noninvasive Detection of Sickle Cell Anemia. Clin Chem. 2012;58(6):1026-32.
- 568 35. Bianchi D, Parker R, Wentworth J, Mandankumar R, Saffer C, Das A. DNA sequencing
- versus standard prenatal aneuploidy screening. New Eng J Med. 2014;370:799-808.
- 570 36. Bijok J, Gorzelnik K, Massalska D, Ilnicka A, Pawlowska B, Zimowski J, et al. Non-
- 571 invasive prenatal diagnosis of the most common aneuploidies with cell-free fetal DNA in
- 572 maternal serum preliminary results. Ginekol Pol. 2014;85:208-13.
- 573 37. Bombard AT, Akolekar R, Farkas DH, VanAgtmael AL, Aquino F, Oeth P, et al. Fetal RHD
- 574 genotype detection from circulating cell-free fetal DNA in maternal plasma in non-sensitized
- 575 RhD negative women. Prenat Diagn. 2011;31(8):802-8.
- 576 38. Boon EMJ, Schleeht HB, Martin P, Daniels G, Vossen R, Den Dunnen JT, et al. Y
- 577 chromosome detection by Real Time PCR and pyrophosphorolysis-activated DNA isolated from
- 578 maternal polymerisation using free fetal plasma. Prenatal Diagnosis. 2007;27(10):932-7.
- 579 39. Bustamante-Aragones A, Rodriguez De Alba M, Gonzalez-Gonzalez C, Trujillo-Tiebas
- 580 MJ, Diego-Alvarez D, Vallespin E, et al. Foetal sex determination in maternal blood from the
- 581 seventh week of gestation and its role in diagnosing haemophilia in the foetuses of female
- 582 carriers. Haemophilia. 2008;14(3):593-8.
- 583 40. Chen H, Wang T, He G, Zhu L, Ma T. Gene analysis of free fetal DNA in maternal
- plasma. Journal of Tongji Medical University. 2001;21(4):329-31.

- 585 41. Chen SP, Lau TK, Zhang CL, Xu CM, Xu ZF, Hu P, et al. A method for noninvasive
- 586 detection of fetal large deletions/duplications by low coverage massively parallel sequencing.
- 587 Prenat Diagn. 2013;33(6):584-90.
- 588 42. Chi C, Hyett J, Finning K, Lee C, Kadir R. Non-invasive first trimester determination of
- 589 fetal gender: a new approach for prenatal diagnosis of haemophilia. BJOG. 2006;113(2):239-
- 590 42.
- 591 43. Chitty L, Finning K, Wade A, Soothill P, Martin B, Oxenford K, et al. Diagnostic accuracy
- 592 of routine antenatal determination of fetal RHD status across gestation: population based
- 593 cohort study. BMJ. 2014;349(g5243).
- 594 44. Chiu R, Lau T, Leung T, Chow K, et al. Prenatal exclusion of beta thalassaemia major by
- 595 examination of maternal plasma. The Lancet. 2002;360(9338):998-1000.
- 596 45. Clausen FB, Christiansen M, Steffensen R, Jorgensen S, Nielsen C, Jakobsen MA, et al.
- 597 Report of the first nationally implemented clinical routine screening for fetal RHD in D-
- 598 pregnant women to ascertain the requirement for antenatal RhD prophylaxis. Transfusion.
- 599 2012;52(4):752-8.
- 600 46. Costa JM, Benachi A, Gautier E, Jouannic JM, Ernault P, Dumez Y. First-trimester fetal
- 601 sex determination in maternal serum using real-time PCR. Prenat Diagn. 2001;21(12):1070-4.
- 602 47. Costa JM, Giovangrandi Y, Ernault P, Lohmann L, Nataf V, El Halali N, et al. Fetal RHD
- 603 genotyping in maternal serum during the first trimester of pregnancy. Br J Haematol.
- 604 2002;119(1):255-60.
- 605 48. Cremonesi L, Galbiati S, Foglieni B, Smid M, Gambini D, Ferrari A, et al. Feasibility Study
- for a Microchip-Based Approach for Noninvasive Prenatal Diagnosis of Genetic Diseases. Ann N
- 607 Y Acad Sci. 2004;1022(1):105-12.
- 608 49. Davalieva K, Dimcev P, Efremov GD, Plaseska-Karanfilska D. Non-invasive fetal sex
- determination using real-time PCR. J Matern Fetal Neonatal Med. 2006;19(6):337-42.
- 610 50. Deng ZH, Wu GG, Li Q, Zhang X, Liang YL, Li DC, et al. Noninvasive genotyping of 9 Y-
- chromosome specific STR loci using circulatory fetal DNA in maternal plasma by multiplex PCR.
- 612 Prenat Diagn. 2006;26(4):362-8.
- 613 51. Fan C, Blumenfeld Y, Chitkara U, Hudgins L, Quake S. Noninvasive diagnosis of fetal
- aneuploidy by shotgun sequencing DNA from maternal blood. PNAS. 2008;105(42):16266-71.
- 615 52. Fernandez-Martinez FJ, Galindo A, Garcia-Burguillo A, Vargas-Gallego C, Nogues N,
- 616 Moreno-Garcia M, et al. Noninvasive fetal sex determination in maternal plasma: a prospective
- 617 feasibility study. Genet Med. 2012;14(1):101-6.
- 618 53. Ferres M, Lichten L, Sachs A, Lau K, Bianchi D. Early experience with noninvasive DNA
- testing for an euploidy in prenatal care. Prenat Diagn. 2013;33(Supp 1):S72.
- 620 54. Finning K, Martin P, Summers J, Daniels G. Fetal genotyping for the K (Kell) and Rh C, c
- 621 and E blood groups on cell-free fetal DNA in maternal plasma. Transfusion. 2007;47:2126-33.
- 622 55. Gautier E, Benachi A, Giovangrandi Y, Ernault P, Olivi M, Gaillon T, et al. Fetal RhD
- 623 genotyping by maternal serum analysis: A two-year experience. Am J Obstet Gynecol.
- 624 2005;192(3):666-9.
- 625 56. Ge Q, Bai Y, Liu Z, Liu Q, Yan L, Lu Z. Detection of fetal DNA in maternal plasma by
- microarray coupled with emulsions PCR. Clin Chim Acta. 2006;369:82-8.
- 627 57. Ghanta S, Mitchell M, Ames M, Hidestrand M, Simpson P, Goetsch M, et al. Non-
- 628 invasive prenatal detection of trisomy 21 using tandem single nucleotide polymorphisms. PloS
- 629 one. 2010;5(10):1.
- 630 58. Gorduza E, Popescu R, Caba L, Ivanov I, Martiniuc V, Nedelea F, et al. Prenatal
- diagnosis of 21 trisomy by quantification of methylated fetal DNA in maternal blood: study on
- 632 10 pregnancies. Rev Romana Med Lab. 2013;21:275-84.
- 633 59. Grill S, Banzola I, Li Y, Rekhviashvili T, Legler TJ, Muller SP, et al. High throughput non-
- 634 invasive determination of foetal Rhesus D status using automated extraction of cell-free foetal
- DNA in maternal plasma and mass spectrometry. Arch Gynecol Obstet. 2009;279(4):533-7.

- 636 60. Gunel T, Kalelioglu I, Ermis H, Aydinli K. Detection of fetal RhD gene from maternal
- 637 blood. J Turk Ger Gynecol Assoc. 2010;11(2):82-5.
- 638 61. Gutensohn K, Müller S, Thomann K, Stein W, Suren A, Körtge-Jung S, et al. Diagnostic
- 639 accuracy of noninvasive polymerase chain reaction testing for the determination of fetal
- rhesus C, c and E status in early pregnancy. BJOG. 2010;117(6):722-9.
- 641 62. Han S, Ryu J, Bae S, Kim Y, Yang Y, Lee K. Noninvasive fetal RhD genotyping using
- circulating cell-free fetal DNA from maternal plasma in RhD-negative pregnant women. J Mol
- 643 Diagn. 2012;14(6):648.
- 644 63. Hill M, Finning K, Martin P, Hogg J, Meaney C, Norbury G, et al. Non-invasive prenatal
- determination of fetal sex: translating research into clinical practice. Clin Genet. 2011;80(1):68-
- 646 75.
- 647 64. Ho SS, Damayanti Z, Chua WY, Ng BL, Peh CM, Biswas A, et al. Non-invasive prenatal
- diagnosis of fetal gender using real-time polymerase chain reaction amplification of SRY in
- maternal plasma. Ann Acad Med Singapore. 2004;33(5):S61-2.
- 650 65. Hofmann W, Entezami M, Haug K, Blank C, Wustemann M, Schulze B. Diagnostic
- 651 accuracy for the noninvasive prenatal detection of common autosomal aneuploidies. Prenat
- 652 Diagn. 2013;33(Supp 1):75.
- 653 66. Hromadnikova I, Vechetova L, Vesela K, Benesova B, Doucha J, Vlk R. Non-invasive fetal
- 654 RHD and RHCE genotyping using real-time PCR testing of maternal plasma in RhD-negative
- pregnancies. J Histochem Cytochem. 2005;53(3):301-5.
- 656 67. Hromadnikova I, Vesela K, Doucha J, Nekovarova K, Duskova D, Schrollova R, et al.
- 657 Non-invasive determination of fetal c and E allele of RHCE gene via real-time PCR testing of
- extracellular DNA extracted from maternal plasma samples using QIAamp DSP virus kit. Journal
- of the Turkish German Gynecology Association. 2007;8(2):140-5.
- 660 68. Hwa H, Ko T, Yen M, Chiang Y. Fetal gender determination using real-time quantitative
- 661 polymerase chain reaction of maternal plasma. J Formos Med Assoc. 2004;103(5):364-68.
- 662 69. Hyett J, Gardener G, Stojilkovic-Mikic T, Finning K, Martin P, Rodeck C, et al. Reduction
- in diagnostic and therapeutic interventions by non-invasive determination of fetal sex in early
- pregnancy. Prenatal Diagnosis. 2005;25(12):1111-6.
- 665 70. Hyland C, Gardener G, Davies H, Ahvenainen M, Flower R, Irwin D, et al. Evaluation of
- 666 non-invasive prenatal RHD genotyping of the fetus. Med J Aust. 2009;191(1):21-5.
- 667 71. Kim SY, Lim JH, Park SY, Kim MY, Choi JS, Ryu HM. Non-invasive prenatal determination
- of fetal gender using QF-PCR analysis of cell-free fetal DNA in maternal plasma. Clin Chim Acta.
- 669 2012;413(5-6):600-4.
- 670 72. Kolialexi A, Tounta G, Apostolou P, Vrettou C, Papantoniou N, Kanavakis W, et al. Early
- 671 non-invasive detection of fetal Y chromosome sequences in maternal plasma using multiplex
- 672 PCR. Eur J Obstet Gynecol Reprod Biol. 2012;161(1):34-7.
- 673 73. Lau TK, Chen F, Pan X, Pooh RK, Jiang F, Li Y. Noninvasive prenatal diagnosis of
- 674 common fetal chromosomal aneuploidies by maternal plasma DNA sequencing. J Matern Fetal
- 675 Neonatal Med. 2012;25:1370-74.
- 676 74. Li Y, Edoardo Di N, Vitucci A, Zimmermann B, et al. Detection of Paternally Inherited
- 677 Fetal Point Mutations for [beta]-Thalassemia Using Size-Fractionated Cell-Free DNA in
- 678 Maternal Plasma. JAMA. 2005;293(7):843-9.
- 679 75. Li Y, Finning K, Daniels G, Hahn S, Zhong X, Holzgreve W. Noninvasive genotyping fetal
- 680 Kell blood group (KEL1) using cell-free fetal DNA in maternal plasma by MALDI-TOF mass
- 681 spectrometry. Prenat Diagn. 2008;28(3):203-8.
- 682 76. Li P-Q, XZhang J, Fan J-H, Zhang Y-Z, Hou H-Y. Development of noninvasive prenatal
- diagnosis of trisomy 21 by RT-MLPA with a new set of SNP markers. Arch Gynecol Obstet.
- 684 2014;289:67-73.
- 685 77. Liao C, Fu Y-G, Huang S-Y, Fu F, Xie G-E. Rapid noninvasive prenatal diagnosis of Down
- 686 syndrome with Ion Proton. Prenat Diagn. 2013;33(Supp 1):76.

- 687 78. Liao C, Yin A-H, Peng C-F, Fu F, Yang J-X, Li R, et al. Noninvasive prenatal diagnosis of
- common aneuploidies by semiconductor sequencing. PNAS. 2014;111:7415-20.
- 689 79. Lim J, Park S, Kim S, Kim D, Choi J, Kim M, et al. Effective detection of fetal sex using
- 690 circulating fetal DNA in first-trimester maternal plasma. FASEB J. 2012;26(1):250-8.
- 691 80. Liu F-M, Wang X-Y, Feng X, Wang W, Ye Y-X, Chen H. Feasibility study of using fetal
- 692 DNA in maternal plasma for non-invasive prenatal diagnosis. Acta Obstet Gynecol Scand.
- 693 2007;86(5):535-41.
- 694 81. Lo Y, Hjelm N, Fidler C, Sargent I, Murphy M, Chamberlain P, et al. Prenatal diagnosis of
- 695 fetal RhD status by molecular analysis of maternal plasma. N Engl J Med. 1998;339(24):1734-8.
- 696 82. Machado I, Castilho L, Pellegrino Jr J, Barini R. Fetal rhd genotyping from maternal
- 697 plasma in a population with a highly diverse ethnic background. Rev Assoc Med Bras
- 698 2006;52(4):232-5.
- 699 83. Manzanares S, Entrala C, Sanchez-Gila M, Fernandez-Rosado F, Cobo D, Martinez E, et
- al. Noninvasive fetal RhD status determination in early pregnancy. Fetal Diagn Ther. 2013;35:7-
- 701 12.
- 702 84. Martinhago C, de Oliveira R, Tomitão Canas M, Vagnini L, Alcantara Oliveira J, Petersen
- 703 C, et al. Accuracy of fetal gender determination in maternal plasma at 5 and 6 weeks of
- 704 pregnancy. Prenat Diagn. 2006;26(13):1219-23.
- 705 85. Minon JM, Gerard C, Senterre JM, Schaaps JP, Foidart JM. Routine fetal RHD
- 706 genotyping with maternal plasma: a four-year experience in Belgium. Transfusion.
- 707 2008;48(2):373-81.
- 708 86. Mohammed N, Kakal F, Somani M, Zafar W. Non-invasive prenatal determination of
- 709 fetal RhD genotyping from maternal plasma: a preliminary study in Pakistan. J Coll Physicians
- 710 Surg Pak. 2010;20(4):246-9.
- 711 87. Moise K, Boring N, O'Shaughnessy R, Simpson L, Wolfe H, Baxter J, et al. Circulating
- 712 cell-free fetal DNA for the detection of RHD status and sex using reflex fetal identifiers. Prenat
- 713 Diagn. 2013;33:95-101.
- 714 88. Mortarino M, Garagiola I, Lotta L, Siboni S, Semprini A, Peyvandi F. Non-invasive tool
- 715 for foetal sex determination in early gestational age. Haemophilia. 2011;17(6):952-6.
- 716 89. New M, Tong Y, Yuen T, Jiang P, Pina C, Chan K, et al. Noninvasive prenatal diagnosis of
- 717 congenital adrenal hyperplasia using cell-free fetal DNA in maternal plasma. J Clin Endocinol
- 718 Metab. 2014;99:1022-30.
- 719 90. Nicolaides KH, Syngelaki A, Gil M, Atanasova V, Markova D. Validation of targeted
- 720 sequencing of single-nucleotide polymorphisms for non-invasive prenatal detection of
- 721 aneuploidy of chromosomes 13, 18, 21, X, and Y. Prenat Diagn. 2013;33(6):575-9.
- 722 91. Nicolaides K, Syngelaki A, Ashoor G, Birdir C, Touzet G. Noninvasive prenatal testing for
- fetal trisomies in a routinely screened first-trimester population. Am J Obstet Gynecol.
- 724 2012;207(5):374.e1-.e6.
- 725 92. Norton M, Brar H, Weiss J, Karimi A, Laurent L, Caughey A, et al. Non-Invasive
- 726 Chromosomal Evaluation (NICE) Study: results of a multicenter prospective cohort study for
- 727 detection of fetal trisomy 21 and trisomy 18. Am J Obstet Gynecol. 2012;207(2):137.e1-.e8.
- 728 93. Norton M, Jacobsson B, Swamy G, Laurent L, Ranzini A, Brar H, et al. Cell-free DNA
- Analysis for Noninvasive Examination of Trisomy. N Engl J Med. 2015;0:null.
- 730 94. Pergament E, Cuckle H, Zimmermann B, Banjevic M, Sigurjonsson S, Ryan A, et al.
- 731 Single-nucleotide polymorphism-based noninvasive prenatal screening in a high-risk and low-
- 732 risk cohort. Obstet Gynecol. 2014;124:210-18.
- 733 95. Perlado-Marina A, Bustamente-Aragones A, Horcajada L, Trujillo-Tiebas M, Lorda-
- 734 Sanchez I, Ramos M, et al. Overview of five-years of experience performing non-invasive fetal
- 735 sex assessment in maternal blood. Diagnostics. 2013;3:283-90.
- 736 96. Picchiassi E, Coata G, Fanetti A, Centra M, Pennacchi L, Di Renzo G. The best approach
- for early prediction of fetal gender by using free fetal DNA from maternal plasma. Prenat
- 738 Diagn. 2008;28(6):525-30.

- 739 97. Polin H, Reiter A, Brisner M, Danzer M, Weinberger J, Gabriel C. Clinical application of
- 740 non-invasive fetal blood group genotyping in Upper Austria. Transfus Med Hemother.
- 741 2013;40(Supp 1):36-7.
- 742 98. Porreco R, Garite T, Maurel K, Marusiak B, Ehrich M, van den Boom D. Noninvasive
- prenatal screening for fetal trisomies 21, 18, 13 and common sex chromosome aneuploidies
- 744 from maternal blood using massively parallel genomic sequencing of DNA. Am J Obstet
- 745 Gynecol. 2014;211:e1-12.
- 746 99. Quezada M, Gil M, Francisco C, Orosz G, Nicolaides K. Screening for trisomies 21, 18
- 747 and 13 by cell-free DNA analysis of maternal blood at 11-13 weeks' gestation and the
- 748 combined test at 11-13 weeks. Ultrasound Obstet Gynecol. 2015;45:36-41.
- 749 100. Rijnders R, Christiaens G, Bossers B, van de Smagt J, van der Schoot, E, de Haas M.
- 750 Clinical applications of cell-free fetal DNA from maternal plasma. Obstet Gynecol.
- 751 2004;130:157-64.
- 752 101. Rijnders R, van der School CE, Bossers B, de Vroede M, Christiaens G. Fetal sex
- determination from maternal plasma in pregnancies at risk for congenital adrenal hyperplasia.
- 754 Obstet Gynecol. 2001;98:374-78.
- 755 102. Rong Y, Gao JJ, Jiang XQ, Zheng F. Multiplex PCR for 17 Y-Chromosome Specific Short
- 756 Tandem Repeats (STR) to Enhance the Reliability of Fetal Sex Determination in Maternal
- 757 Plasma. Int J Mol Sci. 2012;13(5):5972-81.
- 758 103. Rouillac-Le S, Sérazin V, Brossard Y, Oudin O, Le Van Kim C, Colin Y, et al. Noninvasive
- 759 fetal RHD genotyping from maternal plasma: Use of a new developed Free DNA Fetal Kit RhD®.
- 760 Transfus Clin Biol. 2007;14(6):572-7.
- 761 104. Sbarsi I, Isernia P, Montanari L, Badulli C, Martinetti M, Salvaneschi L. Implementing
- 762 non-invasive RHD genotyping on cell-free foetal DNA from maternal plasma: the Pavia
- 763 experience. Blood transfus. 2012;10(1).
- 764 105. Scheffer PG, van der School CE, Page-Christiaens G, Bossers B, van Erp F, de Haas M.
- 765 Reliability of Fetal Sex Determination Using Maternal Plasma. Obstet Gynecol.
- 766 2010;115(1):117-26.
- 767 106. Scheffer P, Ait Soussan A, Verhagen O, Page-Christiaens G, Oepkes D, de Haas M, et al.
- 768 Noninvasive fetal genotyping of human platelet antigen-1a. BJOG. 2011;118(11):1392-5.
- 769 107. Sehnert AJ, Rhees B, Comstock D, de Feo E, Heilek G, Burke J, et al. Optimal Detection
- 770 of Fetal Chromosomal Abnormalities by Massively Parallel DNA Sequencing of Cell-Free Fetal
- 771 DNA from Maternal Blood. Clin Chem. 2011;57(7):1042-9.
- 772 108. Sekizawa A, Kondo T, Iwasaki M, Watanabe A, Jimbo M, Saito H, et al. Accuracy of fetal
- 773 gender determination by analysis of DNA in maternal plasma. Clin Chem. 2001;47:1856-58.
- 774 109. Sesarini C, Gimenez M, Redal M, Izbizky G, Aiello H, Argibay P, et al. Non invasive
- prenatal genetic diagnosis of fetal RhD and sex through the analysis of free fetal DNA in
- maternal plasma. Arch Argent Pediatr. 2009;107(5):405-9.
- 777 110. Shaw S, Chen C-Y, Hsiao C-H, Ren Y, Tian F, Tsai C. Non-invasive prenatal testing for
- 778 whole fetal chromosome aneuploidies: a multi-center prospective cohort trial in Taiwan.
- 779 Prenat Diagn. 2013;33(supp1):81.
- 780 111. Sirichotiyakul S, Charoenkwan P, Sanguansermsri T. Prenatal diagnosis of homozygous
- 781 alpha-thalassaemia-1 by cell-free fetal DNA in maternal plasma. Prenat Diagn. 2011;32:45-9.
- 782 112. Siva S, Johnson S, McCracken S, Morris J. Evaluation of the clinical usefulness of
- isolation of fetal DNA from the maternal circulation. Aust N Z J Obstet Gynaecol.
- 784 2003;43(1):10-5.
- 785 113. Song K, Ashoor G, Syngelaki A, Wagner M, Birdir C, Struble C, et al. Clinical evaluation
- 786 of a directed cfDNA analysis method for non-invasive prenatal fetal trisomy detection. Prenat
- 787 Diagn. 2012;32:1-35.
- 788 114. Song Y, Liu C, Qi H, Zhang Y, Bian X, Liu J. Noninvasive prenatal testing of fetal
- 789 aneuploidies by massively parallel sequencing in a prospective Chinese population. Prenat
- 790 Diagn. 2013;33(7):700-6.

- 791 115. Sparks A, Struble C, Wang E, Song K, Oliphant A. Noninvasive prenatal detection and
- 792 selective analysis of cell-free DNA obtained from maternal blood: evaluation for trisomy 21
- 793 and trisomy 18. Am J Obstet Gynecol. 2012;206(4):319.e1-.e9.
- 794 116. Stumm M, Entezami M, Trunk N, Beck M, Löcherbach J, Wegner R-D, et al. Noninvasive
- 795 prenatal detection of chromosomal aneuploidies using different next generation sequencing
- rategies and algorithms. Prenat Diagn. 2012;32(6):569-77.
- 797 117. Stumm M, Entezami M, Haug K, Blank C, Wustemann C, Schulze B, et al. Diagnostic
- 798 accuracy of random massively parallel sequencing for non-invasive prenatal detection of
- 799 common autosomal aneuploidies: a collaborative study in Europe. Prenat Diagn. 2014;34:185-
- 800 91.
- 118. Tong Y, Jin S, Chiu R, Ding C, Chan K, Leung T, et al. Noninvasive Prenatal Detection of
- 802 Trisomy 21 by an Epigenetic-Genetic Chromosome-Dosage Approach. Clinical Chemistry.
- 803 2010;56(1):90-8.
- 804 119. Tsang JCH, Charoenkwan P, Chow KCK, Jin Y, Wanapirak C, Sanguansermsri T, et al.
- 805 Mass spectrometry-based detection of hemoglobin E mutation by allele-specific base
- 806 extension reaction. Clin Chem. 2007;53(12):2205-9.
- 807 120. Tungwiwat W, Fucharoen G, Fucharoen S, Ratanasiri T, Sanchaisuriya K, Sae-Ung N.
- 808 Application of maternal plasma DNA analysis for noninvasive prenatal diagnosis of Hb E-β-
- 809 thalassemia. Transl Res. 2007;150(5):319-25.
- 810 121. Tungwiwat W, Fucharoen S, Fucharoen G, Ratanasiri T, Sanchaisuriya K. Accuracy of
- fetal gender detection using a conventional nested PCR assay of maternal plasma in daily
- 812 practice. Aust N Z J Obstet Gynaecol. 2008;48(5):501-4.
- 813 122. Turner M, Martin C, O'Leary J. Detection of fetal Rhesus D gene in whole blood of
- women booking for routine antenatal care. Eur J Obstet Gynecol Reprod Biol. 2003;108(1):29-
- 815 32.
- 816 123. Tynan J, Angkachatchai V, Ehrich M, Paladino T, van den Boom D, Oeth P. Multiplexed
- 817 analysis of circulating cell-free fetal nucleic acids for noninvasive prenatal diagnostic RHD
- 818 testing. Am J Obstet Gynecol. 2011;204(3):251.e1-.e6.
- 819 124. Van den Oever JME, Balkassmi S, Johansson LF, van Scheltema PNA, Suijkerbuijk RF,
- 820 Hoffer MJV, et al. Successful Noninvasive Trisomy 18 Detection Using Single Molecule
- 821 Sequencing. Clin Chem. 2013;59(4):705-9.
- 822 125. Van den Oever JME, Balkassmi S, Verweij EJ, van Iterson M, van Scheltema PNA,
- 823 Oepkes D, et al. Single Molecule Sequencing of Free DNA from Maternal Plasma for
- Noninvasive Trisomy 21 Detection. Clin Chem. 2012;58(4):699-706.
- 825 126. Vecchione G, Tomaiuolo M, Sarno M, Colaizzo D, Petraroli R, Matteo M, et al. Fetal Sex
- 826 Identification in Maternal Plasma by Means of Short Tandem Repeats on Chromosome X. Ann
- 827 N Y Acad Sci. 2008;1137(1):148-56.
- 828 127. Verweij E, deBoer M, van Scheltema P, van den oever J, Boon E, Oepkes D. Non-
- 829 invasive prenantal diagnosis of trisomy 21: replacing invasive testing or replacing screening?
- 830 Am J Obstet Gynecol. 2012:S313.
- 831 128. Vora N, Johnson K, Peter I, Tighiouart H, Ralston S, Craigo S, et al. Circulating cell-free
- 832 DNA levels increase variably following chorionic villus sampling. Prenat Diagn. 2010;30(4):325-
- 833 8.
- 834 129. Wagner J, Džijan S, Pavan-Jukić D, Wagner J, Lauc G. Analysis of multiple loci can
- 835 increase reliability of detection of fetal Y-chromosome DNA in maternal plasma. Prenat Diagn.
- 836 2008;28(5):412-6.
- 837 130. Wang X, Wang B, Ye S, Liao Y, Wang L, He Z. Non-invasive foetal RHD genotyping via
- 838 real-time PCR of foetal DNA from Chinese RhD-negative maternal plasma. Eur J Clin Invest.
- 839 2009;39(7):607-17.
- 840 131. Wei C, Saller D, Sutherland J. Detection and Quantification by Homogeneous PCR of
- 841 Cell-free Fetal DNA in Maternal Plasma. Clin Chem. 2001;47(2):336-8.

- 842 132. Zadeh NM, Mesbah-Namin A, Ala F. Noninvasive prenatal diagnosis of fetal sex by a
- new highly sensitive Real-time PCR. Clin Biochem. 2011;44(13):S100-01.
- 244 133. Zhang H, Gao Y, Jiang F, Fu M, Yuan Y, Guo Y, et al. Non-invasive prenatal testing for
- 845 trisomies 21, 18 and 13: clinical experience from 146 958 pregnancies. Ultrasound Obstet
- 846 Gynecol. 2015;45(5):530-8.
- 847 134. Zhong X, Holzgreve W, Hahn S. Risk free simultaneous prenatal identification of fetal
- 848 Rhesus D status and sex by multiplex real-time PCR using cell free fetal DNA in maternal
- 849 plasma. Swiss Med Wkly. 2001;131:70-4.
- 850 135. Zhu B, Sun Q-W, Lu Y-C, Sun M-M, Wang L-J, Huang X-H. Prenatal fetal sex diagnosis by
- detecting amelogenin gene in maternal plasma. Prenat Diagn. 2005;25(7):577-81.
- 852 136. Zhou L, Thorson JA, Nugent C, Davenport RD, Butch SH, Judd WJ. Noninvasive prenatal
- 853 RHD genotyping by real-time polymerase chain reaction using plasma from D-negative
- 854 pregnant women. Am J Obstet Gynecol. 2005;193(6):1966-71.
- 855 137. Zimmermann B, Hill M, Gemelos G, Demko Z, Banjevic M, Baner J, et al. Noninvasive
- 856 prenatal aneuploidy testing of chromosomes 13, 18, 21, X, and Y, using targeted sequencing of
- 857 polymorphic loci. Prenat Diagn. 2012;32(13):1233-41.
- 858 138. Zolotukhina TV, Shilova NV, Voskoboeva EY. Analysis of cell-free fetal DNA in plasma
- and serum of pregnant women. J Histochem Cytochem. 2005;53(3):297-9.
- 860 139. Illanes S, Denbow M, Kailasam C, Finning K, Soothill PW. Early detection of cell-free
- fetal DNA in maternal plasma. Early Hum Dev. 2007;83(9):563-6.
- 862 140. Santacroce R, Vecchione G, Tomaiyolo M, Sessa F, Sarno M, Colaizzo D, et al.
- 863 Identification of fetal gender in maternal blood is a helpful tool in the prenatal diagnosis of
- 864 haemophilia. Haemophilia. 2006;12(4):417-22.
- 865 141. Smid M, Lagona F, de Benassuti L, Ferrari A, Ferrari M, Cremonesi L. Evaluation of
- 866 Different Approaches for Fetal DNA Analysis from Maternal Plasma and Nucleated Blood Cells.
- 867 Clin Chem. 1999;45(9):1570-2.
- 868 142. Song Y, Huang S, Zhou X, Jiang Y, Qi Q, Bian X, et al. Non-invasive prenatal testing for
- fetal aneuploidies in the first trimester of pregnancy. Ultrasound Obstet Gynecol.
- 870 2015;45(1):55-60.
- 871 143. Zhao X, Suzumori N, Ozaki Y, Sato T, Suzumori K. Examination of fetal cells and cell-free
- 872 fetal DNA in material blood for fetal gender determination. Gynecol Obstet Invest. 2004;58:57-
- 873 60.
- 874 144. Morris J, Springett A. The National Down Syndrome Cytogenetic Register for England
- 875 and Wales 2012 Annual Report: Queen Mary University of London, Barts and The London
- 876 School of Medicine and Dentistry 2014.
- 877 145. Programmes NS. Fetal Anomaly Screening Programme (FASP) Standards. London:
- 878 Public Health England; 2015.
- 879 146. Morris S, Karlsen S, Chung N, Hill M, Chitty L. Model-based analysis of costs and
- outcomes of non-invasive prenatal testing for Down's Syndrome using cell free fetal DNA in
- the UK National Health Service. Plos One. 2014;9(4):e935559.
- 882 147. Palomaki GE, Kloza EM, Lambert-Messerlian GM, van den Boom D, Ehrich M, Deciu C,
- 883 et al. Circulating cell free DNA testing: are some test failures informative? Prenatal Diagnosis.
- 884 2015;35(3):289-93.
- 885 148. Mennuti M, Cherry A, Morrissette J, Dugoff L. Is it time to sound an alarm about false-
- 886 positive cell free DNA testing for fetal aneuploidy? Am J Obstet Gynecol. 2013;209:415-9.
- 887 149. Ma J, Pan H, Fu J, Yu L, Yang H. Perspective study of non-invasive prenatal testing using
- 888 cell-free fetal DNA in high-risk population. Zhonghua Yi Xue Za Zhi. 2015;95(11):849-52.
- 889 150. Wax J, Cartin A, Chard D, Lucas F, Pinette M. Noninvasive prenatal testing: impact of
- 890 genetic counselling, invasive prenatal diagnosis, and trimsomy 21 detection. J Clin Ultrasound.
- 891 2015;43(1):1-6.

- 892 151. Ke WL, Zhao WH, Wany XY. Detection of fetal cell-free DNA in maternal plasma for
- 893 Down Syndrome, Edward Syndrome and Patau syndrome of high risk fetus. Int J Clin Exp Med.
- 894 2015;8(6):9525-30.
- 895 152. Ahmadi M, Amirizadeh N, Azarkeyvan A, Valikhani A, Sayyadipoor F, Navirouyan M.
- 896 Fetal RHD genotyping in plasma of RH negative pregnant women by real time PCR. Vox Sang.
- 897 2015;109:302.
- 898 153. Finning K, Tovey S, Desay K, Latham T, Daniels G. UK NHS blood and transplant fetal
- 899 RHD screening Giving anti-D only to those who need it! Vox Sang. 2015;109:282.
- 900 154. Gonenc G, Isci H, Yititer A, Hancer V, Buyukdotan M, Guducu N, et al. Non-invasive
- 901 prenatal diagnosis of fetal RhD by using free fetal DNA. Clin Exp Obstet Gynecol.
- 902 2015;42(3):344-46.
- 903 155. Hernandez-Gomez M, Ramirez-Arroyo E, Melendez-Hernandez R, Garduno-Zarazua L,
- 904 Mayen-Molina D. Non-invasive prenatal test (NIPT) in maternal blood by parallel massive
- 905 sequencing, initial experience in Mexican women and literature review. Ginecol Obstet Mex.
- 906 2015;83(5):277-88.
- 907 156. Sago H, Sekizawa A. Nationwide demonstration project of next-generation sequencing
- of cell-free DNA in maternal plasma in Japan: 1-year experience. Prenat Diagn. 2015;35(4):331-
- 909 6.
- 910 157. Picchiassi E, Di Renzo G, Tarquini F, Bini V, Centra M, Pennacchi L, et al. Non-invasive
- 911 prenatal RHD genotypiing using cell-free fetal FNA from amternal plasma: An Intalian
- 912 experience. Transfus Med Hemother. 2015;42(1):22-8.
- 913 158. Tarquini F, Picchiassi E, Centra M, Pennacchi L, Galeone F, Bini V, et al. Maternal
- 914 smoking does not affect the amount of cell-free fetal DNA in maternal plasma during the 1st
- 915 trimester of pregnancy. J Obstet Gynecol. 2015;35(1):42-5.
- 916 159. Grace M, Hardisty E, Green N, Davidson E, Stuebe A, Vora N. Cell free DNA testing-
- 917 interpretation of results using an online calculator. Am J Obstet Gynecol. 2015;213(1):30.e1-
- 918 .e4.
- 919 160. Bianchi D. Pregnancy: prepare for unexpected prenatal test results. Nature.
- 920 2015;522:29-30.
- 921 161. Chitty L, Kroese M. Realising the promise of non-invasive prenatal testing. BMJ.
- 922 2015;350.
- 923
- 924 Supplementary material legends
- 925 **Figure S1** Study selection from initial search
- 926 **Figure S2** HSROC curves for bivariate analyses
- 927 **Figure S3** Forest plots of studies bivariate not possible
- 928 **Figure S4** Bar chart demonstrating quality assessment of included studies from
- 929 QUADAS-2 risk of bias assessment
- 930 **Table S1** Characteristics of included studies
- 931 **Table S2** Bivariate results
- 932 **Table S3** Reasons for false positives and false negatives and inconclusive results
- 933 **Table S4** Clinical application for Trisomy 21

Appendix S1 Search strategy
 Appendix S2 Additional statistical methods
 Appendix S3 Quality assessment results

Figure 1: Forest plot of studies testing fetal sex using cell-free fetal DNA

Cturdu	TD	ED	EN	TN	Canaitivity (DEN CI)	Consider to EN CIV	Considerate (DEN CI)	Consider to EN CD
Study	TP	FP		TN		Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Smid 1999	16	1	0	10	1.00 [0.79, 1.00]	0.91 [0.59, 1.00]		
Lo 1997	24	0	6	13	0.80 [0.61, 0.92]	1.00 [0.75, 1.00]		
Chen 2001	44	0	2	19	0.96 [0.85, 0.99]	1.00 [0.82, 1.00]		
Al-Yatama 2001	53	3	2	22	0.96 [0.87, 1.00]	0.88 [0.69, 0.97]		
Costa 2001	61	0	0	60	1.00 [0.94, 1.00]	1.00 [0.94, 1.00]	•	•
Rjinders 2001	23	0	1	21	0.96 [0.79, 1.00]	1.00 [0.84, 1.00]	_	_
Sekizawa 2001	139	0	4	159	0.97 [0.93, 0.99]	1.00 [0.98, 1.00]	•	•
Wei 2001	19	0	0	11	1.00 [0.82, 1.00]	1.00 [0.72, 1.00]	_	
Zhong 2001	16	0	0	18	1.00 [0.79, 1.00]	1.00 [0.81, 1.00]		
Siva 2003	10	3	0	11	1.00 [0.69, 1.00]	0.79 [0.49, 0.95]		
Cremonesi 2004	183	0	5	168	0.97 [0.94, 0.99]	1.00 [0.98, 1.00]	•	•
Rjinders 2004	35	0	1	29	0.97 [0.85, 1.00]	1.00 [0.88, 1.00]	-	-
Hwa 2004	20	0	3	33	0.87 [0.66, 0.97]	1.00 [0.89, 1.00]	-	-
Ho 2004	13	0	0	10	1.00 [0.75, 1.00]	1.00 [0.69, 1.00]		
Zhao 2004	29	4	0	11	1.00 [0.88, 1.00]	0.73 [0.45, 0.92]		
Hyett 2005	13	0	0	15	1.00 [0.75, 1.00]	1.00 [0.78, 1.00]		
Zhu 2005	17	2	0	13	1.00 [0.80, 1.00]	0.87 [0.60, 0.98]		
Zolotukhina 2005	34	3	2	21	0.94 [0.81, 0.99]	0.88 [0.68, 0.97]	-	-
Zhou 2005	68	0	4	26	0.94 [0.86, 0.98]	1.00 [0.87, 1.00]	-	-
Ge 2006	40	0	2	34	0.95 [0.84, 0.99]	1.00 [0.90, 1.00]	-	-
Deng 2006	30	0	0	34	1.00 [0.88, 1.00]	1.00 [0.90, 1.00]	-	-
Davalieva 2006	25	0	3	18	0.89 [0.72, 0.98]	1.00 [0.81, 1.00]	-	-
Chi 2006	6	0	0	4	1.00 [0.54, 1.00]	1.00 [0.40, 1.00]		
Martinhago 2006	36	0	3	40	0.92 [0.79, 0.98]	1.00 [0.91, 1.00]	-	-
Santacroce 2006	22	0	0	18	1.00 [0.85, 1.00]	1.00 [0.81, 1.00]	-	-
Al-Yatama 2007	25	2	1	20	0.96 [0.80, 1.00]	0.91 [0.71, 0.99]	-	-
Boon 2007	44	2	0	52	1.00 [0.92, 1.00]	0.96 [0.87, 1.00]	-	-
Liu 2007	13	0	0	17	1.00 [0.75, 1.00]	1.00 [0.80, 1.00]		-
Illanes 2007	15	1	0	10	1.00 [0.78, 1.00]	0.91 [0.59, 1.00]		
Bustamante-Aragones 2008	177	0	6	163	0.97 [0.93, 0.99]	1.00 [0.98, 1.00]	•	•
Picchiassi 2008	81	2	1	61	0.99 [0.93, 1.00]	0.97 [0.89, 1.00]	•	-
Tungwiwat 2008	95	0	2	71	0.98 [0.93, 1.00]	1.00 [0.95, 1.00]	-	-
Minon 2008	278	0	0	266	1.00 [0.99, 1.00]	1.00 [0.99, 1.00]	•	•
Vecchione 2008	8	0	0	18	1.00 [0.63, 1.00]	1.00 [0.81, 1.00]		
Wagner 2008	154	0	- 5	129	0.97 [0.93, 0.99]	1.00 [0.97, 1.00]	•	•
Sesarini 2009	33	2	1	20	0.97 [0.85, 1.00]	0.91 [0.71, 0.99]	-	
Hyland 2009	24	0	0	16	1.00 [0.86, 1.00]	1.00 [0.79, 1.00]	-	-
Wang 2009	41	0	0	37	1.00 [0.91, 1.00]	1.00 [0.91, 1.00]	-	-
Akolekar 2010	90	1	1	119	0.99 [0.94, 1.00]	0.99 [0.95, 1.00]	-	•
Scheffer 2010	105	0	0	81	1.00 [0.97, 1.00]	1.00 [0.96, 1.00]	•	•
Vora 2010	30	0	0	22	1.00 [0.88, 1.00]	1.00 [0.85, 1.00]	-	-
Hill 2011	209	0	2	194	0.99 [0.97, 1.00]	1.00 [0.98, 1.00]	•	•
Tynan 2011	74	5	1	70	0.99 [0.93, 1.00]	0.93 [0.85, 0.98]	-	-
Sehnert 2011	23	0	0	19	1.00 [0.85, 1.00]	1.00 [0.82, 1.00]	-	-
Sirichotiyakul 2011	72	0	0	86	1.00 [0.95, 1.00]	1.00 [0.96, 1.00]	-	•
Mortarino 2011	63	1	14	103	0.82 [0.71, 0.90]	0.99 [0.95, 1.00]	-	-
Zadeh 2011	6	0	0	9	1.00 [0.54, 1.00]	1.00 [0.66, 1.00]		
Aghanoori 2012	92	2	6	100	0.94 [0.87, 0.98]	0.98 [0.93, 1.00]	-	-
Fernandez-Martinez 2012	220	1	0	183	1.00 [0.98, 1.00]	0.99 [0.97, 1.00]	•	•
Rong 2012	25	0	0	15	1.00 [0.86, 1.00]	1.00 [0.78, 1.00]	-	-
Sbarsi 2012	10	0	0	10	1.00 [0.69, 1.00]	1.00 [0.69, 1.00]		
Kolialexi 2012	10	0	0	5	1.00 [0.69, 1.00]	1.00 [0.48, 1.00]		
Kim 2012	81	3	1	77	0.99 [0.93, 1.00]	0.96 [0.89, 0.99]	-	-
Lim 2012	99	0	0	104	1.00 [0.96, 1.00]	1.00 [0.97, 1.00]	-	•
Lau 2012	61	Ō	0	47	1.00 [0.94, 1.00]	1.00 [0.92, 1.00]	-	-
Perlado-Marina 2013	111	ō	0	115	1.00 [0.97, 1.00]	1.00 [0.97, 1.00]	•	•
Moise 2013	167	1	1	168	0.99 [0.97, 1.00]	0.99 [0.97, 1.00]		
Nicolaides 2013	116	Ö	Ö	109	1.00 [0.97, 1.00]	1.00 [0.97, 1.00]	•	•
Porreco 2014	1634	4	3	1681	1.00 [0.99, 1.00]	1.00 [0.99, 1.00]	•	
Pergament 2014	418	Ö	0	358	1.00 [0.99, 1.00]	1.00 [0.99, 1.00]	<u></u>	<u></u>
-	_			_		. ,,	0 0.2 0.4 0.6 0.8 1	0 0.2 0.4 0.6 0.8 1

Figure 2: Forest plot of studies testing Rhesus D status using cell-free fetal DNA

Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Lo 1998	37	0	2	18	0.95 [0.83, 0.99]	1.00 [0.81, 1.00]	-	-
Zhong 2001	26	0	1	7	0.96 [0.81, 1.00]	1.00 [0.59, 1.00]	-	
Costa 2002	62	0	0	40	1.00 [0.94, 1.00]	1.00 [0.91, 1.00]	-	-
Siva 2003	17	2	3	4	0.85 [0.62, 0.97]	0.67 [0.22, 0.96]		
Turner 2003	30	0	4	14	0.88 [0.73, 0.97]	1.00 [0.77, 1.00]	-	
Rjinders 2004	43	1	0	28	1.00 [0.92, 1.00]	0.97 [0.82, 1.00]	-	_
Hromadnikova 2005	24	0	0	21	1.00 [0.86, 1.00]	1.00 [0.84, 1.00]	-	_
Gautier 2005	170	0	0	102	1.00 [0.98, 1.00]	1.00 [0.96, 1.00]	•	•
Zhou 2005	68	0	0	26	1.00 [0.95, 1.00]	1.00 [0.87, 1.00]	-	-
Machado 2006	58	1	1	15	0.98 [0.91, 1.00]	0.94 [0.70, 1.00]	-	
Al-Yatama 2007	21	0	5	28	0.81 [0.61, 0.93]	1.00 [0.88, 1.00]		-
Rouillac-Le 2007	229	2	0	77	1.00 [0.98, 1.00]	0.97 [0.91, 1.00]	•	-
Minon 2008	359	1	0	185	1.00 [0.99, 1.00]	0.99 [0.97, 1.00]	•	•
Hyland 2009	95	0	0	40	1.00 [0.96, 1.00]	1.00 [0.91, 1.00]	-	-
Grill 2009	122	2	5	49	0.96 [0.91, 0.99]	0.96 [0.87, 1.00]	-	-
Wang 2009	60	5	0	10	1.00 [0.94, 1.00]	0.67 [0.38, 0.88]	-	
Sesarini 2009	34	6	2	18	0.94 [0.81, 0.99]	0.75 [0.53, 0.90]	-	
Aykut 2010	21	0	0	8	1.00 [0.84, 1.00]	1.00 [0.63, 1.00]	-	
Mohammed 2010	12	3	1	5	0.92 [0.64, 1.00]	0.63 [0.24, 0.91]		
Gunel 2010	7	0	0	12	1.00 [0.59, 1.00]	1.00 [0.74, 1.00]		
Achargui 2011	83	1	5	31	0.94 [0.87, 0.98]	0.97 [0.84, 1.00]	-	-
Bombard 2011	278	3	4	121	0.99 [0.96, 1.00]	0.98 [0.93, 0.99]	•	•
Tynan 2011	86	0	0	62	1.00 [0.96, 1.00]	1.00 [0.94, 1.00]	-	-
Han 2012	24	0	0	8	1.00 [0.86, 1.00]	1.00 [0.63, 1.00]	-	
Clausen 2012	1368	6	2	862	1.00 [0.99, 1.00]	0.99 [0.99, 1.00]	•	•
Sbarsi 2012	13	0	0	7	1.00 [0.75, 1.00]	1.00 [0.59, 1.00]		
Moise 2013	220	3	1	96	1.00 [0.98, 1.00]	0.97 [0.91, 0.99]	•	-
Manzanares 2013	73	1	1	40	0.99 [0.93, 1.00]	0.98 [0.87, 1.00]	-	-
Polin 2013	88	0	0	34	1.00 [0.96, 1.00]	1.00 [0.90, 1.00]	•	-
Chitty 2014	2563	18	19	1920	0.99 [0.99, 1.00]	0.99 [0.99, 0.99]		
							0 0.2 0.4 0.6 0.8 1	0 0.2 0.4 0.6 0.8 1

Figure 3A: Forest plot of studies testing Trisomy 21 using cell-free fetal DNA

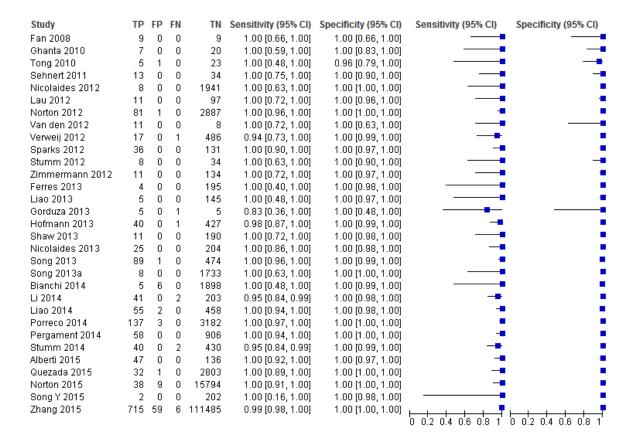


Figure 3B: Forest plot of studies testing Trisomy 18 using cell-free fetal DNA

Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Fan 2008	2	0	0	16	1.00 [0.16, 1.00]	1.00 [0.79, 1.00]		-
Ghanta 2010	2	0	0	25	1.00 [0.16, 1.00]	1.00 [0.86, 1.00]		-
Sehnert 2011	8	0	0	39	1.00 [0.63, 1.00]	1.00 [0.91, 1.00]		-
Nicolaides 2012	2	2	0	1945	1.00 [0.16, 1.00]	1.00 [1.00, 1.00]		•
Lau 2012	10	0	0	98	1.00 [0.69, 1.00]	1.00 [0.96, 1.00]		•
Norton 2012	37	2	1	2886	0.97 [0.86, 1.00]	1.00 [1.00, 1.00]	-	•
Zimmermann 2012	3	0	0	142	1.00 [0.29, 1.00]	1.00 [0.97, 1.00]		•
Sparks 2012	8	0	0	159	1.00 [0.63, 1.00]	1.00 [0.98, 1.00]		•
Hofmann 2013	8	0	0	460	1.00 [0.63, 1.00]	1.00 [0.99, 1.00]		•
Nicolaides 2013	3	0	0	226	1.00 [0.29, 1.00]	1.00 [0.98, 1.00]		•
Van den 2013	9	0	0	8	1.00 [0.66, 1.00]	1.00 [0.63, 1.00]		
Shaw 2013	8	0	0	193	1.00 [0.63, 1.00]	1.00 [0.98, 1.00]		•
Song 2013a	2	1	0	1738	1.00 [0.16, 1.00]	1.00 [1.00, 1.00]		•
Song 2013	57	1	1	505	0.98 [0.91, 1.00]	1.00 [0.99, 1.00]	-	•
Liao 2014	16	4	0	495	1.00 [0.79, 1.00]	0.99 [0.98, 1.00]	_	•
Pergament 2014	24	1	1	938	0.96 [0.80, 1.00]	1.00 [0.99, 1.00]	-	•
Bianchi 2014	2	3	0	1900	1.00 [0.16, 1.00]	1.00 [1.00, 1.00]		•
Bijok 2014	1	0	1	7	0.50 [0.01, 0.99]	1.00 [0.59, 1.00]		
Porreco 2014	36	0	3	3283	0.92 [0.79, 0.98]	1.00 [1.00, 1.00]	-	•
Stumm 2014	8	1	0	463	1.00 [0.63, 1.00]	1.00 [0.99, 1.00]		•
Norton 2015	9	1	1	15830	0.90 [0.55, 1.00]	1.00 [1.00, 1.00]		•
Zhang 2015	167	51	3	112044	0.98 [0.95, 1.00]	1.00 [1.00, 1.00]	•	•
Quezada 2015	9	5	1	2821	0.90 [0.55, 1.00]	1.00 [1.00, 1.00]		•
Song Y 2015	1	0	0	203	1.00 [0.03, 1.00]	1.00 [0.98, 1.00]		
							0 0.2 0.4 0.6 0.8 1	0 0.2 0.4 0.6 0.8 1

Figure 3C: Forest plot of studies testing Monosomy X using cell-free fetal DNA

Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Tungwiwat 2007	0	0	0	0	Not estimable	Not estimable		
Sehnert 2011	2	0	0	45	1.00 [0.16, 1.00]	1.00 [0.92, 1.00]		-
Lau 2012	0	0	0	0	Not estimable	Not estimable		
Zimmermann 2012	1	0	0	144	1.00 [0.03, 1.00]	1.00 [0.97, 1.00]	-	•
Nicolaides 2013	2	0	0	227	1.00 [0.16, 1.00]	1.00 [0.98, 1.00]		•
Song 2013a	2	0	1	1737	0.67 [0.09, 0.99]	1.00 [1.00, 1.00]		•
Shaw 2013	3	0	1	197	0.75 [0.19, 0.99]	1.00 [0.98, 1.00]		•
Pergament 2014	9	1	1	953	0.90 [0.55, 1.00]	1.00 [0.99, 1.00]		•
Porreco 2014	9	11	0	3258	1.00 [0.66, 1.00]	1.00 [0.99, 1.00]	0 0.2 0.4 0.6 0.8 1	0 0.2 0.4 0.6 0.8 1

Figure 3D: Forest plot of studies testing Trisomy 13 using cell-free fetal DNA

Study	TP	FP	FN	TN	Sensitivity (95% CI)	Specificity (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)
Fan 2008	1	0	0	17	1.00 [0.03, 1.00]	1.00 [0.80, 1.00]		
Zimmermann 2012	2	0	0	143	1.00 [0.16, 1.00]	1.00 [0.97, 1.00]		•
Lau 2012	2	0	0	106	1.00 [0.16, 1.00]	1.00 [0.97, 1.00]		•
Van den 2013	1	4	3	9	0.25 [0.01, 0.81]	0.69 [0.39, 0.91]		
Nicolaides 2013	1	0	0	228	1.00 [0.03, 1.00]	1.00 [0.98, 1.00]		•
Shaw 2013	3	0	0	198	1.00 [0.29, 1.00]	1.00 [0.98, 1.00]		•
Song 2013a	1	0	0	1740	1.00 [0.03, 1.00]	1.00 [1.00, 1.00]		•
Hofmann 2013	5	0	0	463	1.00 [0.48, 1.00]	1.00 [0.99, 1.00]		•
Porreco 2014	14	0	2	3306	0.88 [0.62, 0.98]	1.00 [1.00, 1.00]		•
Pergament 2014	12	0	0	953	1.00 [0.74, 1.00]	1.00 [1.00, 1.00]		•
Stumm 2014	5	0	0	467	1.00 [0.48, 1.00]	1.00 [0.99, 1.00]		•
Liao 2014	3	0	0	512	1.00 [0.29, 1.00]	1.00 [0.99, 1.00]		•
Zhang 2015	22	45	0	112198	1.00 [0.85, 1.00]	1.00 [1.00, 1.00]	-	•
Song Y 2015	1	0	0	203	1.00 [0.03, 1.00]	1.00 [0.98, 1.00]		•
Norton 2015	2	2	0	11181	1.00 [0.16, 1.00]	1.00 [1.00, 1.00]		•
Quezada 2015	2	2	3	2829	0.40 [0.05, 0.85]	1.00 [1.00, 1.00]	0 0.2 0.4 0.6 0.8 1	0 0.2 0.4 0.6 0.8 1