| 1 | |
|----|--|
| 2 | Testing peatland testate amoeba transfer functions: appropriate methods for |
| 3 | clustered training-sets. |
| 4 | |
| 5 | Richard J. Payne ^{1,2*} , Richard J. Telford ^{3*} , Jeffrey J. Blackford ² , Antony Blundell ⁴ , |
| 6 | Robert K. Booth ⁵ , Dan J. Charman ⁶ , Łukasz Lamentowicz ⁷ , Mariusz Lamentowicz ⁸ , |
| 7 | Edward A.D. Mitchell ⁹ , Genevieve Potts ² , Graeme T. Swindles ⁴ , Barry G.Warner ¹⁰ and |
| 8 | Wendy Woodland ¹¹ |
| 9 | |
| 10 | ¹ School of Science and the Environment, Manchester Metropolitan University, |
| 11 | Chester Street, Manchester, M1 5GD, UK. |
| 12 | |
| 13 | ² Geography, School of Environment and Development, The University of |
| 14 | Manchester, Oxford Road, Manchester, M13 9PL, UK. |
| 15 | |
| 16 | ³ Bjerknes Centre for Climate Change and Department of Biology, University of |
| 17 | Bergen, Post box 7820, N-5020 Bergen, Norway. |
| 18 | |
| 19 | ⁴ School of Geography, University of Leeds, Leeds, LS2 9JT, UK. |
| 20 | |
| 21 | ⁵ Earth and Environmental Sciences, Lehigh University, 31 Williams Drive, Bethlehem, |
| 22 | PA 18015, USA |
| 23 | |
| 24 | ⁶ Department of Geography, College of Life and Environmental Sciences, University |
| 25 | of Exeter, Exeter, Devon, EX4 4RJ, UK |
| 26 | |
| 27 | ⁷ Department of Hydrobiology, Adam Mickiewicz University, Umultowska 89, 61-614 |
| 28 | Poznan, Poland. |
| 29 | |
| 30 | ⁸ Dept. of Biogeography & Palaeoecology, Adam Mickiewicz University, Dziegielowa |
| 31 | 27, PL-61-680 Poznan, Poland. |
| 32 | |

| 33 | ⁹ Laboratoire de biologie du sol, Université de Neûchatel, Rue Emile-Argand 11, Case |
|----|---|
| 34 | postale 158, 2009 Neuchâtel, Switzerland. |
| 35 | |
| 36 | ¹⁰ Department of Earth and Environmental Sciences, University of Waterloo, 200 |
| 37 | University Avenue West, Waterloo, Ontario, N2L 3G1, Canada |
| 38 | |
| 39 | ¹¹ Department of Geography & Environmental Management, School of the Built and |
| 40 | Natural Environment, University of the West of England, Coldharbour Lane, Bristol, |
| 41 | BS16 1QY, UK. |
| 42 | |
| 43 | * Joint first authors, e-mails: r.payne@mmu.ac.uk, richard.telford@bio.uib.no |
| 44 | |
| 45 | ABSTRACT |
| 46 | |
| 47 | Transfer functions are widely used in palaeoecology to infer past environmental |
| 48 | conditions from fossil remains of many groups of organisms. In contrast to |
| 49 | traditional training-set design with one observation per site, some training sets, |
| 50 | including those for peatland testate amoeba-hydrology transfer functions, have a |
| 51 | clustered structure with many observations from each site. Here we show that this |
| 52 | clustered design causes standard performance statistics to be overly optimistic. |
| 53 | Model performance when applied to independent data sets is considerably weaker |
| 54 | than suggested by statistical cross-validation. We discuss the reasons for these |
| 55 | problems and describe leave-one-site-out cross-validation and the cluster bootstrap |
| 56 | as appropriate methods for clustered training sets. Using these methods we show |
| 57 | that the performance of most testate amoeba-hydrology transfer functions is worse |
| 58 | than previously assumed and reconstructions are more uncertain. |
| 59 | |
| 60 | KEYWORDS: Transfer functions; Palaeoclimate; Clustered data; Leave-one-site-out |
| 61 | cross-validation, Cluster bootstrap. |
| 62 | |
| 63 | |

64 Transfer functions are widely used to generate quantitative environmental 65 reconstructions in palaeoecology. Traditional training-set design (e.g. Birks et al. 66 1990) has one observation per site. An alternative design with many observations at 67 each site is used for some training-sets, including those for chironomid-lake depth 68 (Kurek and Cwynar 2009); coastal diatom-water chemistry (Saunders et al. 2008); 69 diatom- and foraminifera-sea level (Massey et al. 2006; Zong & Horton 1999; Leorri 70 et al. 2008); and testate amoeba-hydrology transfer functions (Charman 2001, 71 Mitchell et al. 2008). Although the implications of, and methods for, such clustered 72 data are well known in other branches of statistics (Walsh 1947), the implications of 73 this design have been neglected for transfer functions.

74 One motivation for developing clustered training-sets is the presence within 75 each site of substantial environmental gradients, which may be large relative to the 76 differences between sites. This contrasts with the traditional one observation per 77 site training-set where typically the environmental variable (e.g. lake-pH) is assumed 78 to be spatially homogeneous at each site. Standard methods for assessing the 79 performance of transfer functions assume that the observations are independent 80 and are thus inappropriate for clustered data. Lack of independence between 81 observations, either because of spatial autocorrelation or a clustered design, will 82 cause performance statistics to be over-optimistic (Telford and Birks, 2005). Telford 83 and Birks (2009) have developed cross-validation methods appropriate for spatially 84 autocorrelated training sets; here we consider the problem of clustered training sets 85 and develop appropriate cross-validation methods. We focus on testate amoeba-86 hydrology transfer functions from peatlands, which have become increasingly 87 important in shaping our understanding of Holocene climatic change (Charman et al. 88 2004, 2006).

89

90 Indications that standard tools are misleading

91 Training sets for peatland testate amoebae transfer functions have a highly
92 uneven spatial structure, with samples from individual sites often only separated by
93 a few metres, while sites may be separated by tens or hundreds of kilometres.
94 Ordinations of testate amoeba data frequently show distinct clustering of
95 observations from the same bog (e.g. Charman et al. 2007, Swindles et al. 2009) and

96 site identity typically explains a large proportion of variance in constrained97 ordinations (Fig. 1).

98 To provide an independent estimate of transfer function performance, we 99 apply five transfer functions to all comparable independent datasets with 100 appropriate corrections for taxonomic and methodological differences (Appendix I). 101 Table 1 shows that most transfer functions perform worse than suggested by leave-102 one-out (LOO) cross-validation when applied to independent data. Methodological 103 explanations for the poor model performance can largely be excluded. Differences in 104 time-discrete water-table measurements cannot explain the differences in rank-105 order shown by Spearman's p. Any differences in sample preparation and analysis, or 106 residual taxonomic biases cannot explain poor performance where these are closely 107 harmonised (e.g. Polish data). Performance is particularly poor for two datasets from 108 Scotland (Payne 2010a; Potts & Blackford unpublished data); in the case of the Moss 109 of Achnacree, this is likely to be due to the limited WTD range in a site which has 110 experienced hydrological modification. As previously presented tests with transfer functions from different regions have frequently (Charman et al. 2007; Booth et al. 111 112 2008; Payne 2011), but not universally (e.g. Swindles et al. 2009), shown 113 performance poorer than LOO cross-validation we conclude that model performance 114 in praxis appears to be weaker than suggested by conventional cross-validation.

115

116 Appropriate cross-validation methods for clustered data

117 Typically, transfer function model performance is assessed by either leave-118 one-out (LOO) or bootstrap cross-validation. In LOO, one observation at a time is 119 omitted from the training-set of size *n* and the environmental value predicted using 120 the remaining *n*-1 observations. For clustered data, this can be extended to leave-121 one-site-out cross-validation (LOSO), where data from one site is omitted from the 122 training set, and data from the remaining *m*-1 sites used to predict it. LOSO is also 123 known as leave-one-cluster-out cross-validation and sometimes as leave-one-group-124 out cross-validation (confusingly, this latter term is also used to refer to k-fold cross-125 validation in which k groups are created at random). 126 In standard bootstrap cross-validation, *n* observations are selected from the

127 training set with replacement, and used to predict the remaining observations and

128 new observations. There are several possible bootstrap schemes available for 129 clustered data including the cluster bootstrap, where *m* clusters are selected at 130 random with replacement, and the two-level bootstrap where *m* clusters are 131 selected at random and observations are selected at random from within each 132 cluster (Field and Welsh 2007). Here we use the cluster bootstrap following the 133 findings of Field and Welsh (2007) that the two-level bootstrap and the related 134 reverse-two-level bootstrap generate excessive variability.

135

136

Application to Testate Amoeba Training sets

137 We determine the performance of 14 published testate amoeba transfer 138 functions for water-table depth (WTD) using both robust cross-validation methods 139 and standard methods. In the case of the Jura training set (Mitchell et al. 1999) we 140 omit samples with estimated rather than measured water-table depths. For all 141 training sets, we use weighted averaging with inverse deshrinking as this transfer 142 function method is fairly robust to spatial autocorrelation (Telford and Birks, 2005) 143 and so should also be fairly robust to clustered data. Assemblage data were square 144 root transformed prior to analysis. All analyses were carried out in R (R Development 145 Core Team 2010) with the rioja library (Juggins 2010).

146 While differences are not always great, all transfer functions except for one 147 exhibit worse performance with LOSO than LOO cross-validation (Table 2). One 148 transfer function has an LOSO RMSEP greater than the standard deviation of WTD. There are several possible reasons for this deterioration in performance. It could be 149 150 simply an artefact because the estimates are based on fewer observations as more 151 observations are omitted during LOSO than LOO. We tested for the importance of 152 this factor by running a modified cross-validation scheme termed leave-many-out 153 (LMO) that omits as many observations as LOSO when making each prediction but 154 with the observations chosen at random rather than being from the same site. We 155 repeated this analysis 100 times to get a distribution of performance statistics and 156 tested if the observed LOSO RMSEP is worse than the 95th percentile of the leavemany-out RMSEP. Only the Poland (Lamentowicz & Mitchell 2005) training set had a 157 158 LOSO performance that was not statistically significantly worse than expected from 159 leaving out so many observations during cross-validation.

LOSO performance would be worse than LOO performance if each site only covered part of the environmental gradient. This factor is likely to be of minor importance, except in the Greece training set as all the other training sets have replication along the WTD gradient and variance partitioning shows only a small covariance between WTD and site for most of the training sets (Figure 1).

As for most training sets the WTD measurements are based on one-time spot measurements, there may be site-specific errors in the WTD measurements if heavyrainfall or prolonged drought occurs between sampling the first and last bog. Most training sets were collected within a short period of time, so major changes in WTD are unlikely to have occurred however a few training sets were acquired over a longer period of time and this may be an important factor (Charman et al. 2007; Lamentowicz et al. 2008b).

172 There are likely to be important non-hydrological controls on amoebae which 173 differ between sites such as pollutant loading with recent studies showing sulphur 174 (Payne et al. 2010), reactive nitrogen (Nguyen-Viet et al. 2004; Mitchell 2004), heavy metals (Nguyen-Viet et al. 2007; 2008) and particulate matter (Meyer et al. 2010) to 175 176 be important. Many transfer function studies have included sites of differing pH and 177 trophic status, and there is evidence for differences in amoeba communities and 178 their hydrological responses between fens and bogs (Payne 2011; Jassey et al. 2011). 179 Plant communities, which differ between sites in many studies, shape both the 180 physical and biotic environment of amoebae through processes such as root 181 exudation and allelopathy, particularly the production of phenolic compounds 182 (Jassey et al. 2011). The fundamental hydrological controls on amoeba communities 183 are poorly understood, while water table depth consistently explains the largest 184 proportion of variance in gradient studies it is clearly not water table depth per se 185 which is important to amoebae usually living well above the water table. Water table 186 depth is simply a robust measurement, which serves as a proxy for the hydrological 187 variables which do affect amoebae such as water film thickness and variability in the top few cm of moss where amoebae live (Sullivan et al. 2011). These variables may 188 189 be controlled by fine-scale structural details of the peat and plant communities. 190

191 <u>Predictors of LOSO relative performance</u>

192 In an attempt to understand the attributes of training sets that have a large 193 decrease in performance with LOSO cross-validation, we regress the decrease in 194 performance, standardised by dividing by the standard deviation of WTD, against the 195 number of sites and observations, the proportion of variance explained by WTD, site, 196 and the covariance between WTD and site (Fig. 2). Of these predictors, only the 197 proportion of variance explained by WTD is a statistically significant predictor of the 198 deterioration in performance. Although the regression is not statistically significant, 199 there appears to be an increased risk of a large reduction in performance for training 200 sets with few sites.

201

202 Error decomposition

The magnitude of the RMSEP is not necessarily a good guide to the utility of a transfer function. If, as is usually the case in testate amoeba palaeoecology, one is interested only in identifying relatively wet and dry phases, then the absolute value of the reconstruction is not very important. Thus, even transfer functions with a large RMSEP could potentially have utility.

For each site in the clustered training-set, we can decompose the total sum of squares of residuals into the proportion explained by site-specific offsets or biases and the residual variation. Table 3 shows that when LOSO is used instead of LOO, the site specific offset increases much more than the residual variation in both absolute and relative terms. This suggests that the absolute values of reconstructions are much more uncertain, but the relative values are only slightly more uncertain than LOO suggests.

215

216 <u>Reconstruction errors</u>

Sample-specific (s1; Birks et al., 1990; Birks, 1995) bootstrap errors for the
cluster bootstrap will always be larger than those from the standard bootstrap. Fig. 3
shows the WTD reconstruction for Jelenia Wyspa, Poland (Lamentowicz et al. 2007b)
using the Poland 2008 training set, with sample-specific bootstrap errors using both
bootstrap techniques. Bootstrap errors vary by sample but are in all cases greater
when using the cluster bootstrap and for some samples the errors are more than
double.

225 <u>Recommendations</u>

226 Given our results, improvements can be made in both the generation and 227 application of clustered training sets. We make four recommendations for 228 generating new training sets, which should be followed where it is practical to do so 229 and may not be possible to satisfy simultaneously. First, efforts should be made to 230 sample the full environmental gradient at each site, or at least to ensure that all 231 parts of the gradient are replicated in several sites. Ideally, the gradients should be 232 uniformly sampled at each site (Telford and Birks 2011). Second, approximately the 233 same number of observations should be made at each site, so that in LOSO cross-234 validation the number of observations omitted is close to constant. Third, a large 235 number of sites should be sampled, as the cluster bootstrap is not appropriate for 236 datasets with few clusters. Finally, the sites should be similar to each other with 237 respect to, for example, vegetation and climate, with the proviso that care is taken 238 to include sufficient diversity of sites to ensure that all fossil samples have good 239 analogues in the training set.

240 We recommend that the robust cross-validation methods developed here are 241 used when testing the performance of clustered training sets. We anticipate that the 242 performance statistics of transfer function methods robust to autocorrelation (e.g., 243 WA) will deteriorate less with robust cross-validation than methods more sensitive 244 to autocorrelation (e.g., WAPLS with several components). If there is a choice of training set that could be applied to the fossil data, we recommend, all else being 245 246 equal, using the training set with the smallest loss of performance when robust 247 cross-validation is used. Single-site training sets (e.g. Booth et al. 2008; Payne et al. 248 2008) will be immune to cluster problems but this may be offset by poor 249 reconstructive ability. As always in quantitative palaeoecology, caution should be 250 used in interpreting small changes in reconstructions and replication using multi-251 core, multi-proxy and multi-site records is desirable.

252

253 <u>Conclusions</u>

Published performance statistics of testate amoeba transfer functions are
 over-optimistic due to the clustered design of the training sets. LOO cross-validation

| 256 | is biased by the lack of independence of the observations. As amoeba communities |
|-----|--|
| 257 | in a sample tend to be more similar to other samples from the same site than to |
| 258 | samples from different sites, if samples from the same site remain in the training set |
| 259 | during cross-validation, then the model will generate unrealistically accurate |
| 260 | predictions of water-table depth in the training set. |
| 261 | |
| 262 | |
| 263 | |
| 264 | |
| 265 | |
| 266 | |
| 267 | |
| 268 | |
| 269 | |
| 270 | |
| 271 | |
| 272 | |
| 273 | |
| 274 | ACKNOWLEDGEMENTS |
| 275 | |
| 276 | RJP was supported by a Humanities Research Fellowship from the University of |
| 277 | Manchester and a Study Grant from the British Institute at Ankara. Norwegian |
| 278 | Research Council projects ARCTREC and PES helped support RJT. We thank H.J.B. |
| 279 | Birks for his comments on this manuscript. R-code for leave-one-site-out and cluster |
| 280 | bootstrap cross-validation has been implemented in the rioja library. This is |
| 281 | publication no. A358 from the Bjerknes Centre for Climate Research |
| 282 | Author contributions: |
| 283 | RJP conceived and coordinated the project, compiled the data and carried out the |
| 284 | tests with independent data-sets. RJT devised and implemented the cross-validation |
| 285 | procedures. RJP and RJT wrote the paper. Other authors contributed data, discussed |
| 286 | the taxonomic harmonisation issues and commented on the interpretation of the |
| 287 | results and manuscript. |

290 TABLES

- 291 Table 1. Transfer function performance for five training sets tested by leave-one-out
- 292 (LOO) cross-validation and application to independent test-sets, showing transfer
- 293 function method used, number of samples (*n*), root mean squared error of
- 294 prediction (RMSEP), R², and Spearman's p. Some values differ from previously
- 295 published values due to minor variation in sample selection and taxonomic
- 296 harmonisation. Values in round brackets show performance when small taxa are
- excluded to account for differences in the use of back-sieving (Appendix 1). R^2 and ρ

298 values in square brackets denote negative correlations.

| Training-set | Transfer | Test-set | Peatland | N | RMSEP | R ² | Р |
|-------------------------|--------------|---|------------------------|-----|--------|----------------|---------|
| | function | | type(s) | | (cm) | | |
| European | 2 component | LOO cross-validation | - | 119 | 5.63 | 0.71 | 0.90 |
| (Charman et WA-PLS | | | | | (5.80) | (0.69) | (0.89) |
| al. 2007) | | All test data | Bogs | 200 | 5.51 | 0.18 | 0.67 |
| | | Blythermo (Potts & Blackford, | Bog | 9 | 11.40 | 0.37 | 0.66 |
| | | unpublished) ² | | | | | |
| | | Loonan (Potts & Blackford, unpublished) ² | Bog | 11 | 13.02 | [0.12] | [-0.38] |
| | | Moss of Achnacree (Payne 2010a) ^{1,2} | Bog | 30 | 6.65 | [0.01] | [-0.01] |
| | | Moidach More (Payne et al. 2010b) ¹ | Bog | 150 | 4.38 | 0.53 | 0.75 |
| UK | WA-Tol | LOO cross-validation | - | 160 | 3.94 | 0.29 | 0.64 |
| (Woodland et | (inverse | | | | (3.91) | (0.30) | (0.64) |
| al. 1998) | deshrinking) | All test data | Bogs | 200 | 6.71 | 0.25 | 0.60 |
| | | Blythermo (Potts & Blackford, unpublished) ² | Bog | 9 | 13.18 | 0.56 | 0.82 |
| | | Loonan (Potts & Blackford, unpublished) ² | Bog | 11 | 17.05 | [0.13] | [-0.21] |
| | | Moss of Achnacree (Payne 2010a) ^{1,2} | Bog | 30 | 10.19 | 0.01 | 0.11 |
| | | Moidach More (Payne et al. 2010b) ¹ | Bog | 150 | 4.86 | 0.23 | 0.42 |
| Alaska | 2 component | LOO cross-validation | - | 91 | 9.99 | 0.53 | 0.81 |
| (Payne et al. 2006) | WA-PLS | Alaska (Markel et al. 2010) | Various | 126 | 16.52 | 0.42 | 0.61 |
| Alaska | 2 component | LOO cross-validation | - | 126 | 8.50 | 0.63 | 0.84 |
| (Markel et al. 2010) | WA-PLS | Alaska (Payne et al. 2006) | Various | 91 | 16.94 | 0.42 | 0.69 |
| Poland | WA-Tol | LOO cross-validation | - | 36 | 7.75 | 0.72 | 0.94 |
| (Lamentowicz | (inverse | All test data | Various | 213 | 11.23 | 0.20 | 0.48 |
| & Mitchell 2005) | deshrinking) | Jedwabna (Lamentowicz et al. 2008b) | Poor fen | 10 | 5.77 | 0.17 | 0.53 |
| | | Mietlica (Lamentowicz et al. 2008b) | Poor fen | 12 | 7.86 | 0.85 | 0.77 |
| | | Ostrowite (Lamentowicz et al. 2008b) | Bog | 7 | 13.41 | 0.82 | 0.85 |
| | | Rybie Oko (Lamentowicz et al. 2008b) | Bog | 16 | 6.35 | 0.80 | 0.84 |
| | | Skrzynka (Lamentowicz et al. 2008b) | Poor fen | 12 | 4.13 | 0.55 | 0.60 |
| | | , Stawek (Lamentowicz et al. 2008b) | Poor fen | 9 | 8.69 | 0.52 | 0.39 |
| | | Stążki (Lamentowicz et al. 2008b) | Moderately rich fen | 10 | 7.89 | 0.51 | 0.71 |

| Żabieniec (Lamentowicz et al. 2008b) | Schwingmoor | 8 | 3.83 | 0.76 | 0.96 |
|--|---------------------|----|-------|------|------|
| Chlebowo (Lamentowicz et al. 2007a, 2008a) | Poor fen | 27 | 5.96 | 0.27 | 0.54 |
| Linje (Lamentowicz et al. 2008b) | Bog and poor fen | 46 | 12.07 | 0.52 | 0.55 |
| Słowińskie Błota (Lamentowicz et al. 2008b) | Bog | 25 | 29.58 | 0.24 | 0.73 |
| Jeziorka Kozie (Lamentowicz et al. 2008b) | Poor fen | 31 | 11.34 | 0.00 | 0.27 |

300 ¹Back-sieving not used so small taxa excluded. 301 ²Lower counts of around 100 tests.

- 303 Table 2. Root mean squared error of prediction for 14 published training sets
- 304 calculated with leave-one-out (LOO), leave-one-site-out (LOSO), and leave-many-out
- 305 (LMO) cross-validation. The 95th percentile of the LMO distribution is shown. Results
- 306 are based on weighted averaging with inverse deshrinking on square root
- 307 transformed data. Also shown are the DWT range (cm), number of sites (m) and
- 308 observations (*n*), and the standard deviation of WTD (sd).

| | Range | т | n | LOO | LOSO | LMO | sd |
|---|---------|----|-----|------|------|------|------|
| | (cm) | | | | | 95% | |
| Europe (Charman et al. 2007) | -3-35 | 7 | 119 | 6.2 | 6.9 | 6.3 | 10.5 |
| Alaska 2006 (Payne et al. 2006) | 7-67 | 8 | 91 | 10.8 | 14.0 | 11.1 | 14.6 |
| Alaska 2010 (Markel et al. 2010) | -18-46 | 12 | 126 | 8.6 | 9.3 | 8.8 | 14.0 |
| Engadine (Lamentowicz et al. 2010) | -20-76 | 6 | 84 | 9.8 | 11.0 | 10.3 | 16.1 |
| Greece (Payne and Mitchell 2007) | -1-14.5 | 4 | 57 | 2.2 | 3.3 | 2.2 | 4.1 |
| Jura (Mitchell et al. 1999) | 3-53 | 4 | 36 | 9.5 | 12.4 | 10.4 | 13.4 |
| Minnesota/Ontario (Warner and Charman | 0-100 | 10 | 49 | 20.1 | 22.7 | 20.8 | 26.2 |
| Newfoundland (Charman and Warner 1997) | -4-46 | 6 | 57 | 7.2 | 8.1 | 7.6 | 11.8 |
| Northern Ireland (Swindles et al. 2009) | -10-38 | 3 | 81 | 5.3 | 6.0 | 5.6 | 12.2 |
| Rockies (Booth and Zygmunt 2005) | -5-50 | 14 | 139 | 7.5 | 8.0 | 7.6 | 16.1 |
| UK (Woodland et al. 1998) | 0-19 | 9 | 160 | 4.0 | 4.8 | 4.1 | 4.7 |
| North America (Booth 2008) | -13-75 | 31 | 403 | 8.1 | 8.2 | 8.2 | 17.1 |
| Poland 2008 (Lamentowicz et al. 2008b) | -25-84 | 15 | 249 | 14.0 | 16.3 | 14.1 | 17.8 |
| Poland 2005 (Lamentowicz and Mitchell 2005) | -3-55 | 3 | 36 | 9.6 | 9.3 | 11.8 | 14.7 |

- Table 3. Decomposition of the mean total sum of squares of the transfer function
- residuals into the portion explained by site-specific offsets and the residual variation
- for both LOO and LOSO cross-validation, and the ratio of the LOSO and LOO results.

| | - | | | | | | | | |
|---------------|-------|------|----------|-------|------|----------|-------|------|----------|
| | LOO | | | LOSO | | | LOSO/ | 'LOO | |
| | Total | Site | Residual | total | Site | Residual | total | Site | Residual |
| Europe | 38 | 9 | 29 | 48 | 16 | 32 | 1.26 | 1.89 | 1.08 |
| Alaska 2006 | 116 | 53 | 63 | 197 | 121 | 75 | 1.69 | 2.28 | 1.19 |
| Alaska 2010 | 75 | 13 | 61 | 86 | 25 | 60 | 1.14 | 1.88 | 0.98 |
| Engadine | 96 | 17 | 79 | 120 | 30 | 90 | 1.25 | 1.72 | 1.15 |
| Greece | 5 | 2 | 2 | 11 | 8 | 3 | 2.35 | 3.56 | 1.22 |
| Jura | 90 | 8 | 82 | 154 | 69 | 85 | 1.71 | 8.93 | 1.04 |
| Minnesota/Ont | 405 | 177 | 228 | 516 | 250 | 266 | 1.27 | 1.41 | 1.17 |
| ario | | | | | | | | | |
| Newfoundland | 52 | 15 | 37 | 66 | 29 | 37 | 1.26 | 1.87 | 1.01 |
| Northern | 28 | 5 | 24 | 35 | 9 | 26 | 1.25 | 2.04 | 1.10 |
| Ireland | | | | | | | | | |
| Rockies | 57 | 8 | 48 | 64 | 16 | 48 | 1.12 | 1.95 | 0.98 |
| UK | 16 | 4 | 12 | 23 | 11 | 11 | 1.44 | 2.74 | 0.98 |
| North America | 66 | 12 | 54 | 68 | 13 | 54 | 1.02 | 1.12 | 1.00 |
| Poland 2008 | 196 | 72 | 124 | 266 | 134 | 133 | 1.36 | 1.85 | 1.07 |
| Poland 2005 | 91 | 11 | 80 | 84 | 13 | 71 | 0.92 | 1.18 | 0.88 |

- 316 Figure 1.
- 317 Variance partitioning of the inertia in the different data-sets into components
- 318 explained by water table depth (light grey), site (dark grey), covariance between site
- and water table depth (black). Unexplained inertia is shown in white. See Table 2 for
- 320 data sources. Site is a statistically significant predictor for all training sets except
- 321 Poland 2005.



323 Figure 2. Scatter plots of the relative decrease in performance against different

324 predictors: a) number of sites; and proportion of variance explained by b) site, c)

water table depth and d) covariance between water table depth and site in a CCA.





Fig. 3. Water table reconstruction from Jelenia Wyspa, Poland (Lamentowicz et al.
2007b) calculated using weighted averaging with inverse deshrinking on square root
transformed data with the expanded Polish training set (Lamentowicz et al. 2008b).
Reconstructions (black) are based on 1000 bootstrap predictions (50 of which are
shown in grey) for a) conventional bootstrap and b) cluster bootstrap. The standard
deviation of the bootstrap predictions (error component s1) is shown with vertical
black lines).



| 337 | |
|-----|---|
| 338 | |
| 339 | REFERENCES |
| 340 | |
| 341 | Birks HJB, Line JM, Juggins S, Stevenson AC, ter Braak CJF (1990) Diatoms and pH |
| 342 | reconstruction. Philosophical Transactions of the Royal Society London B 327: |
| 343 | 263-278. |
| 344 | Birks HJB (1995) Quantitative palaeoenvironmental reconstructions, In: Maddy D and |
| 345 | Brew JS (eds.) Statistical modelling of quaternary science data. Technical Guide 5. |
| 346 | Cambridge: Quaternary Research Association. |
| 347 | Booth RK (2008) Testate amoebae as proxies for mean annual water-table depth in |
| 348 | Sphagnum-dominated peatlands of North America. Journal of Quaternary Science |
| 349 | 23: 43-57. |
| 350 | Booth RK, Sullivan ME, Sousa VA (2008) Ecology of testate amoebae in a North |
| 351 | Carolina pocosin and their potential use as environmental and |
| 352 | palaeoenvironmental indicators. Ecoscience 15: 277-289. |
| 353 | Booth RK, Zygmunt JR (2005) Biogeography and comparative ecology of testate |
| 354 | amoebae inhabiting Sphagnum-dominated peatlands in the Great Lakes and |
| 355 | Rocky Mountain regions of North America. Diversity and Distributions 11: 577- |
| 356 | 590. |
| 357 | Charman DJ (2001) Biostratigraphic and palaeoenvironmental applications of testate |
| 358 | amoebae. Quaternary Science Reviews 20, 1753-1764. |
| 359 | Charman DJ, Hendon D (2000) Long-term changes in soil water tables over the past |
| 360 | 4500 years: relationships with climate and North Atlantic atmospheric circulation |
| 361 | and sea surface temperature. Climatic Change 47: 45-59 |
| 362 | Charman DJ, Warner BG (1992) Relationship between testate amoebae (Protozoa: |
| 363 | Rhizopoda) and microenvironmental parameters on a forested peatland in |
| 364 | northeastern Ontario. Canadian Journal of Zoology 70: 2474-2482. |
| 365 | Charman DJ, Warner BG (1997) The ecology of testate amoebae (Protozoa: |
| 366 | Rhizopoda) and microenvironmental parameters in Newfoundland, Canada: |
| 367 | modeling hydrological relationships for palaeoenvironmental reconstruction. |
| 368 | Ecoscience 4: 555-562. |

369 Charman DJ, Brown A, Hendon D, Karofeld E (2004) Testing the relationship between 370 Holocene peatland palaeoclimate reconstructions and instrumental data at two 371 European sites. *Quaternary Science Reviews* 23: 137-143. 372 Charman DJ, Blundell A, Chiverrell RC, Hendon D, Langdon PG (2006) Compilation of 373 non-annually resolved Holocene proxy climate records: stacked Holocene 374 peatland palaeo-water table reconstructions from northern Britain. Quaternary 375 Science Reviews 25: 336–350. 376 Charman DJ, Blundell A, ACCROTELM members (2007) A new-European testate 377 amoebae transfer function for palaeohydrological reconstruction on 378 ombrotrophic peatlands. Journal of Quaternary Science 22: 209-221. 379 Field CA, Welsh AH (2007) Bootstrapping clustered data. Journal of the Royal 380 Statistical Society B 69: 369-390. 381 Hendon D, Charman DJ (2004) High-resolution peatland water-table changes for the 382 past 200 years: the influence of climate and implications for management. The 383 Holocene 14: 125-134. 384 Jassey VEJ, Chiapusio G, Mitchell EAD, Binet P, Toussaint M-L, Gilbert D (2011) Fine-385 scale horizontal and vertical micro-distribution patterns of testate amoebae 386 along a narrow fen/bog gradient. *Microbial Ecology* 61: 374-385. 387 Juggins S (2003) C2 user guide. Software for ecological and palaeoecological data 388 analysis and visualisation. University of Newcastle, Newcastle Upon Tyne. 389 Kurek J, Cwynar LC (2009) The potential of site-specific and local chironomid-based 390 inference models for reconstructing past lake levels. Journal of Paleolimnology 391 42: 37-50. 392 Lamentowicz M, Mitchell EAD (2005) The ecology of testate amoebae (Protists) in 393 Sphagnum in north-west Poland in relation to peatland ecology. Microbial 394 *Ecology* 50: 48-63. 395 Lamentowicz Ł, Gabka M, Lamentowicz M (2007a) Species composition of testate 396 amoebae (Protists) and environmental parameters in a Sphagnum peatland. 397 Polish Journal of Ecology 55: 749-759. Lamentowicz M, Tobolski K, Mitchell EAD (2007b) Palaeoecological evidence for 398 399 anthropogenic acidification of a kettle-hole peatland in northern Poland. The 400 Holocene 17: 1185-1196.

Lamentowicz Ł, Lamentowicz M, Gabka M (2008a) Testate amoebae ecology and a
local transfer function from a peatland in western Poland. *Wetlands* 28: 164-175.
Lamentowicz M, Obremska M, Mitchell EAD (2008b) Autogenic succession, land-use
change, and climatic influences on the Holocene development of a kettle-hole
mire in Northern Poland. *Review of Palaeobotany and Palynology* 151: 21-40.
Lamentowicz M, Lamentowicz Ł, van der Knaap WO, Gąbka M, Mitchell EAD (2010)

407 Contrasting species—environment relationships in communities of testate

amoebae, bryophytes and vascular plants along the fen-bog gradient. *Microbial Ecology* 59: 499-510.

410 Leorri E, Horton BP, Cearreta A (2008) Development of a foraminifera-based transfer

function in the Basque marshes, N. Spain: Implications for sea-level studies in the
Bay of Biscay. *Marine Geology* 251: 60-74.

413 Markel ER, Booth RK, Qin Y (2010) Testate amoebae and δ 13C of *Sphagnum* as

surface-moisture proxies in Alaskan peatlands. *The Holocene* 20: 463-475.

415 Massey A, Gehrels WR, Charman DJ, White SV (2006) An intertidal foraminifera-

416 based transfer function for reconstructing Holocene sea-level change in

417 southwest England. *Journal of Foraminiferal Research* 36: 215-232.

418 Mitchell EAD (2004) Response of testate amoebae (Protozoa) to N and P fertilization

419 in an Arctic wet sedge tundra. *Arctic Antarctic and Alpine Research* 36: 77–82.

420 Mitchell EAD, Buttler AJ, Warner BG, Gobat JM (1999) Ecology of testate amoebae

421 (Protozoa: Rhizopoda) in *Sphagnum* peatlands in the Jura mountains, Switzerland
422 and France. *Ecoscience* 6: 565-576.

Mitchell EAD, Charman DJ, Warner BG (2008) Testate amoebae analysis in ecological
and paleoecological studies of wetlands: past, present and future. *Biodiversity and Conservation* 17: 2115-2137.

426 Meyer C, Bernard N, Moskura M, Toussaint ML, Denayer F, Gilbert D (2010) Effects of

427 urban particulate deposition on microbial communities living in bryophytes: An
428 experimental study. *Ecotoxicology and Environmental Safety* 73: 1776-1784.

429 Nguyen-Viet H, Gilbert D, Bernard N, Mitchell EAD, Badot PM (2004) Relationship

430 between atmospheric pollution characterized by NO₂ concentrations and testate

431 amoebae density and diversity. *Acta Protozoologica* 43: 233–239.

- 432 Nguyen-Viet H, Bernard N, Mitchell EAD, Cortet J, Badot PM, Gilbert D (2007)
- 433 Relationship between testate amoeba (Protist) communities and atmospheric
- 434 heavy metals accumulated in Barbula indica (Bryophyta) in Vietnam. *Microbial*435 *Ecology* 53: 53–65.
- 436 Nguyen-Viet H, Bernard N, Mitchell EAD, Badot PM, Gilbert D (2008) Effect of lead
- 437 pollution on testate amoebae communities living in *Sphagnum fallax*: an
- 438 experimental study. *Ecotoxicology and Environmental Safety* 69: 130–138.
- Payne RJ (2009) The standard preparation method for testate amoebae leads to
 selective loss of the smallest shells. *Quaternary Newsletter* 119: 16-20.
- Payne RJ (2010) Testate amoeba response to acid deposition in a Scottish peatland.
 Aquatic Ecology 44, 373-385.
- Payne RJ (2011) Can testate amoeba-based palaeohydrology be extended to fens?
 Journal of Quaternary Science 26: 15-27.
- Payne RJ, Mitchell EAD (2007) Ecology of testate amoebae from mires in the Central
- 446 Rhodope Mountains, Greece and development of a transfer function for
- 447 paleohydrological reconstruction. *Protist* 158: 159-171.
- 448 Payne RJ, Mitchell EAD (2009) How many is enough? Determining optimal count
- totals for ecological and palaeoecological studies of testate amoebae. *Journal of Paleolimnology* 42: 483-495.
- 451 Payne RJ, Kishaba K, Blackford JJ, Mitchell EAD (2006) The ecology of testate
- 452 amoebae in southcentral Alaskan peatlands: Building transfer function models for
- 453 palaeoenvironmental inference. *The Holocene* 16: 403-414.
- 454 Payne RJ, Charman DJ, Matthews S, Eastwood W (2008) Testate amoebae as
- palaeoclimate proxies in Sürmene Ağaçbaşi Yaylasi peatland (Northeast Turkey).
 Wetlands 28: 311-323.
- 457 Payne RJ, Charman, DJ, Gauci V (2010): The impact of simulated sulfate deposition
- 458 on peatland testate amoebae. *Microbial Ecology* 59: 76-83.
- 459 Payne RJ, Lamentowicz M, Mitchell EAD (2011) The perils of taxonomic
- 460 inconsistency in quantitative palaeoecology: simulations with testate amoeba461 data. *Boreas* 40: 15-27.
- 462 R Development Core Team (2010) *R: A language and environment for statistical*
- 463 *computing*. R Foundation for Statistical Computing, Vienna, Austria.

| 464 | Saunders KM, Hodgson DA, Harrison J, McMinn A (2008) Palaeoecological tools for |
|--------------------------|---|
| 465 | improving the management of coastal ecosystems: a case study from Lake King |
| 466 | (Gippsland Lakes) Australia. Journal of Paleolimnology 40: 33-47. |
| 467 468 469 470 | Sullivan, M.E. & Booth, R.K. (2011) The potential influence of short-term environmental variability on the composition of testate amoeba communities in <i>Sphagnum</i> peatlands. <i>Microbial Ecology</i> 62: 80-93. Swindles GT, Charman DL, Roe HM, Sansum PA (2009) Environmental controls on |
| 471 | neatland testate amoebae (Protozoa: Rhizonoda) in the North of Ireland. |
| 472 | Implications for Holocene palaeoclimate studies. <i>Journal of Paleolimpology</i> 42: |
| 473 | 123–140. |
| 474 | Telford RJ, Birks HJB (2005) The secret assumptions of transfer functions: problems |
| 475 | with spatial autocorrelation in evaluating model performance. Quaternary |
| 476 | Science Reviews 24: 2173-2179. |
| 477 | Telford RJ, Birks HJB (2009) Evaluation of transfer functions in spatially structured |
| 478 | environments. Quaternary Science Reviews 28: 1309-1316. |
| 479 | Telford, R.J. Birks, H.J.B. (2011) Effect of uneven sampling along an environmental |
| 480 | gradient on transfer-function performance. Journal of Paleolimnology 46: 99-106. |
| 481 | Walsh JE (1947) Concerning the effect of intraclass correlation on certain significance |
| 482 | tests. Annals of Mathematical Statistics 18: 88-96. |
| 483 | Warner BG, Charman DJ (1994) Holocene changes on a peatland interpreted from |
| 484 | testate amoebae (Protozoa) analysis. <i>Boreas</i> 23: 270-280. |
| 485 | Woodland W, Charman DJ, Simms P (1998) Quantitative estimates of water tables |
| 486 | and soil moisture in Holocene peatlands from testate amoebae. The Holocene 8: |
| 487 | 261-273. |
| 488 | Zong Y, Horton BP (1999) Diatom-based tidal-level transfer functions as an aid in |
| 489 | reconstructing Quaternary history of sea-level movements in the UK. Journal of |
| 490 | Quaternary Science 14: 153-167. |
| 491 | |

- 493 Appendix 1. Details of taxonomic harmonisation showing groupings and
- 494 nomenclatural changes made to the original data. In addition to these changes small
- 495 taxa (Corythion spp., Trinema spp., Euglypha rotunda type, Euglypha cristata,
- 496 Cryptodifflugia oviformis, Difflugia pulex type and Pseudodifflugia fulva type) were
- 497 eliminated where there was a difference in preparation method between training
- 498 and test sets.
- 499

| Dataset | Taxa in original data | Taxa here |
|----------------------|---|---------------------------------|
| Moss of Achnacree | Centropyxis aerophila type | Centropyxis cassis type |
| (Payne 2010a) | Phryganella acropodia type | Cyclopyxis arcelloides type |
| | Corythion dubium, Trinema complanatum | Corythion-Trinema type |
| Moidach More | Phryganella acropodia type | Cyclopyxis arcelloides type |
| (Payne et al. 2010b) | Corythion dubium, Trinema complanatum | Corythion-Trinema type |
| UK | Nebela minor, Nebela tincta, Nebela | Nebela tincta type |
| (Woodland et al. | parvula | |
| 1998; Charman et al. | | |
| 2007; Potts & | | |
| Blackford | | |
| unpublished data) | | |
| Alaska | Arcella arenaria type, A. catinus type | Arcella catinus type |
| (Payne et al. 2006; | Centropyxis aerophila s.l., C. cassis type | Centropyxis aerophila type |
| Markel et al. 2010) | Centropyxis laevis, C. ecornis, C. ecornis | Centropyxis ecornis type |
| | type | |
| | Cyclopyxis arcelloides type, Phryganella | Cyclopyxis arcelloides type |
| | acropodia type, P. acropodia s.l. | |
| | Nebela dentistoma, N. vitraea | Argynnia <i>dentistoma</i> type |
| | Euglypha ciliata, E. compressa, E. strigosa, | Euglypha spp. |
| | E. rotunda s.l., E. tuberculata type, E. | |
| | <i>strigosa</i> type <i>, E. rotunda</i> type | |
| | Nebela tincta s.l., N. tincta, N. parvula | Nebela tincta type |
| | Placocista spinosa s.l., P. lens, P. spinosa | Placocista spinosa type |
| | Trigonopyxis arcula, T. minuta | Trigonopyxis arcula type |
| | Trinema spp., T. lineare | Trinema spp. |