# SOUTHEAST ASIAN OVALOCYTOSIS IN THE CAPE COLOURED POPULATION

Cheryl Anne Ziervogel (Student number 8916411)

A dissertation submitted to the Faculty of Health Sciences, University of Johannesburg, in fulfilment of the requirements for the degree of Master of Technology.

Supervisor: Prof. T.L. Coetzer ------

Co-supervisor: Dr. H. Abrahamse

Johannesburg 2005

## DECLARATION

I declare that this dissertation is my own, unaided work. It is being submitted for the Degree of Master of Technology at the University of Johannesburg. It has not been submitted before for any degree or examination in any other University or Technikon.

\_\_\_\_\_ day of \_\_\_\_\_

#### ABSTRACT

Southeast Asian Ovalocytosis (SAO) is an autosomal dominantly inherited, classically asymptomatic condition, that is widespread in Southeast Asian populations of Malasia, Indonesia, Papua New Guinea and the Philippines. Some regions have close to 30% prevalence and this is thought to be due to SAO providing partial protection against malaria.

SAO is characterized by rigid, spoonshaped, ovalocytic red blood cells. The underlying defect is a deletion of 27bp in the band 3 gene, resulting in the absence of 9 amino acids (400-408) at the boundary of the cytoplasmic and membrane domains of band 3, causing abnormal structure and function. SAO is tightly linked in all cases to the band 3 Memphis 1 polymorphism, which is a lysine 56 (AAG) – glutamic acid (GAG) substitution. This polymorphism can be inherited independently and the prevalence ranges from about 6-30% according to various populations studied.

The presence of SAO in a Cape Coloured family is a recent finding. The purpose of this study was to further investigate the prevalence of SAO and the band 3 Memphis 1 polymorphism in the Cape Coloured population. 20 unrelated individuals with SAO morphology were identified. DNA analysis revealed the 27bp deletion of exon 11 of the band 3 gene in all 20 subjects, which is diagnostic of SAO. This indicates a high occurrence of the SAO mutation in the Cape Coloured population, which is speculated to be due to a founder effect. Some of the clinical features differed from classically described SAO as some individuals showed evidence of haemolysis.

Protein analysis showed all 20 individuals to have a reduced band 3 mobility, indicating the band 3 Memphis 1 polymorphism. Detecting the band 3 Memphis 1 polymorphism on a protein level is time consuming and labour intensive, therefore

a PCR assay, which utilizes DNA, was developed for the rapid screening of this polymorphism. The PCR assay was based on a nucleotide mismatch which created a Taq 1 restriction site when combined with the band 3 Memphis 1 allele, but not with the wild type allele. Digestion of the PCR product with Taq 1 allowed differentiation between the two alleles. It was established that the band 3 Memphis 1 polymorphism has a high prevalence in the Cape Coloured population as it was detected in 108/326 (33%) of the individuals studied. Analysis indicated the Memphis allele is in Hardy-Weinberg equilibrium.

"Great things are not done by impulse, but by a series of small things brought together"

Vincent Van Gogh

In loving memory of my father Richard Cleveland Lyons 1932-1997

#### ACKNOWLEDGEMENTS

This work was performed in the laboratory of the Red Cell Membrane Unit, Department of Haematology, National Health Laboratory Services.

I would like to give thanks to:

- Prof. Thérèsa L. Coetzer (head of the Red Cell Membrane Unit), for her endless patience, supervision, guidance and encouragement during the preparation of this dissertation.
- Rhoda Essack and staff from NHLS Greenpoint Haematology laboratory, who faithfully and consistently submitted blood samples to the RBC membrane unit.
- Dr. H. Abrahamse (co-supervisor at the University of Johannesburg), for her endless patience and encouragement.
- My husband, Craig Ziervogel, for his constant patience and understanding.
- My sister and brother-in-law, Desireè and Dewald Waso, for their endless support.
- My mother and step dad, Sybil and Eddie Roos, for their support.
- My parents-in-law, David and Brenda Ziervogel for their encouragement.
- The students in the Red Cell Membrane Unit, Kuben, Sonja, Roberto,
   Lara, Marcel and Danny, for their invaluable assistance with computer software during the write up of this dissertation.
- My friends Desire and Yogi Groos for their kind assistance with MS Word.
- Mrs N. Pienaar and the Coagulation team (Lancet laboratories), for their love and support.
- National Health Laboratory Services, for funds provided.
- Medical Faculty Research Endowment Fund, for funds provided.

## **PUBLICATIONS**

#### Poster presentation at an international conference:

Lyons (Ziervogel) CA, Officer S, Coetzer TL.

Southeast Asian Ovalocytosis in the Cape Coloured population. Presented at: The 15<sup>th</sup> Meeting of the International Society of Haematology, African and European Division. Durban, South Africa, 1999.

#### Manuscript in preparation:

Coetzer TL, Ziervogel CA.

A novel PCR assay detects a high prevalence of band 3 Memphis 1 polymorphism in the Cape Coloured population.

## **TABLE OF CONTENTS**

		PAGE
Abst	tract	iii
Acknowledgements		
Publ	lications	vii
Tabl	le of Contents	viii
List of Figures		
List of Tables		xiii
List of Abbreviations		xiv
1.	INTRODUCTION	1
1.1	The Red Blood Cell	1
1.2	The Red Blood Cell Membrane	1
1.3	Band 3	5
	1.3.1 Function of band 3	5
	1.3.2 Structure of band 3	6
1.4	Band 3 Memphis 1 Polymorphism	6
1.5	Southeast Asian Ovalocytosis (SAO)	8
	1.5.1 Geographical distribution of SAO	8
	1.5.2 Clinical manifestations of SAO	10
	1.5.3 Protein defects of SAO	11
	1.5.4 Genetic defect of SAO	12
	1.5.5 SAO and malaria	12
1.6	Aims of the Study	14

## 2. METHODS AND MATERIALS

2.1	Subjects	16
2.2	Erythrocyte Membrane Protein Analysis	17
	2.2.1 Membrane protein preparation	17
	2.2.2 Erythrocyte membrane solubilization	18
	2.2.3 Protein concentration determination	18
	2.2.4 Fairbanks sodium dodecyl sulphate polyacrylamide	19
	gel electrophoresis (SDS-PAGE)	
	2.2.5 Laemmli SDS-PAGE	20
	2.2.6 Limited tryptic digest of RBC membranes	22
	2.2.7 Linear 15%-25% gradient Laemmli gel	22
	2.2.8 Immunoblot analysis of trypsin digested	23
	RBC membranes	
2.3	DNA Analysis	24
	2.3.1 Extraction of genomic DNA	24
	2.3.2 Quantitation of DNA	25
	2.3.3 Electrophoresis of DNA	25
	2.3.4 Detection of the SAO band 3 mutation using	26
	polymerase chain reaction (PCR)	
	2.3.5 Detection of band 3 Memphis 1 polymorphism	27
	using PCR	
3.	RESULTS	30
3.1	Subjects	30
3.2	Erythrocyte Membrane Protein Analysis	35
	3.2.1 Fairbanks SDS-PAGE analysis	35
	3.2.2 Laemmli SDS-PAGE	37
	3.2.3 Band 3 Memphis 1 protein analysis	42

16

42
42
46
46
49
54
59
()
63
63
63
63 63 64
63 63 64 64
63 63 64 64 66
63 63 64 64 66
63 64 64 66 68
63 64 64 66 68
<ul> <li>63</li> <li>64</li> <li>64</li> <li>66</li> <li>68</li> <li>69</li> </ul>
<ul> <li>63</li> <li>64</li> <li>64</li> <li>66</li> <li>68</li> <li>69</li> </ul>

## LIST OF FIGURES

## PAGE

## 1. INTRODUCTION

Figure 1.1	A schematic diagram representing the separation	3
	of the main red cell proteins on Fairbanks	
	SDS-PAGE (Fairbanks et al., 1971)	
Figure 1.2	Schematic model of the RBC membrane by Lux	4
	and Palek (1995)	
Figure 1.3	Organizational model of the human red cell anion	7
	exchange protein band 3	
Figure 1.4	The SAO band 3 defect and Memphis 1 polymorphism	9
Figure 1.5	Model of the role of the cytoplasmic domain of band 3	13
	in regulating membrane extensional rigidity	
	(Mohandas et al., 1992)	

## 2. **RESULTS**

Figure 3.1	Peripheral blood smears	31
Figure 3.2	Pedigree of the P kindred	32
Figure 3.3	3.5%-17% non-linear Fairbanks SDS-PAGE of the	36
	RBC membrane protein profile of SAO subject D	
	and a control	
Figure 3.4	12% Laemmli SDS-PAGE of the RBC membrane	40
	protein profile of SAO subject D and a control	
Figure 3.5	Reticulocyte % versus 4.1a/4.1b ratio	41
Figure 3.6	Limited tryptic digest of RBC membranes	43
	separated on 15%-25% Laemmli SDS-PAGE	
Figure 3.6	separated on 15%-25% Laemmli SDS-PAGE	4

Figure 3.7	Immunoblot analysis of limited trypsin digested	44
	RBC membranes	
Figure 3.8	1% agarose gel of electrophoresed 1µg aliquots of	45
	extracted human genomic DNA	
Figure 3.9	Partial gene sequence of band 3	47
Figure 3.10	0.8% Agarose gel separation of PCR amplification	48
	products of exon 11 of the band 3 gene	
Figure 3.11	Partial gene sequence of band 3	52
Figure 3.12	PCR products using P154 + P155 oligonucleotide	53
	primers with the wild type and band 3 Memphis 1 alleles	
Figure 3.13	2% Agarose gel separation of PCR amplification	55
	products of exon 4 of the band 3 gene	
Figure 3.14	2.5% Agarose gel separation of Tag 1 digested	56
	PCR amplicons of exon 4 of the band 3 gene	
Figure 3.15	P kindred pedigree depicting SAO and band 3	57
	Memphis 1 gene status	

#### LIST OF TABLES

#### 3. RESULTS

#### **Table 3.1** Clinical data of the P kindred 33 Table 3.2 Clinical data of seven unrelated individuals with SAO 34 Table 3.3 Quantitation of erythrocyte membrane proteins from control 38 and SAO individuals separated on Fairbanks SDS-PAGE 39 **Table 3.4** The mean ratios of 4.1a/4.1b proteins in relation to the reticulocyte count Table 3.5 Summary of the SAO and band 3 Memphis gene status 58 of 299 Cape Coloured controls and 27 individuals from 21 SAO families **Table 3.6** Relative genotype frequencies 59 Table 3.7 Observed versus expected genotype values 60 **Table 3.8** $\chi^2$ values for normal, SAO and band 3 Memphis 1 61 polymorphism individuals

PAGE

## LIST OF ABBREVIATIONS

А	Ampere
A <sub>260/280/595</sub>	absorbance at 260nm, 280nm and 595nm respectively
ACD	acid citrate dextrose
AE1	anion exchange protein 1
APS	ammonium persulphate
Arg	arginine
Asn	asparagine
Asp	aspartic acid
ATP	adenosine triphosphate
b	band
bis	N,N-methylene bisacrylamide
bp	base pair
BSA	bovine serum albumin
°C	degrees Celcius
DMSO	dimethyl sulphoxide
DNA	deoxyribonucleic acid
EDTA	ethylenediamine tetraacetic acid disodium salt
8	relative centrifugal force
g	gram
Glu	glutamic acid
Hb	haemoglobin
Hct	haematocrit
k	kilo
kb	kilo bases
kDa	kilodalton
1	litre
Lys	lysine
М	molar
μ	micro

m	milli
MCV	mean cell volume
nm	nanometer
OD	optical density
Р	primer
PAGE	polyacrylamide gel electrophoresis
PCR	polymerase chain reaction
PMSF	phenylmethylsulphonylfluoride
RBC	red blood cell
rpm	revolutions per minute
SAO	southeast asian ovalocytosis
SDS	sodium dodecyl sulphate
TEA	Tris EDTA acetate buffer
TBS	Tris buffered saline
TE	Tris EDTA buffer
TEMED	N,N,N,N-tetramethylethylenediamine
T <sub>m</sub>	melting temperature
Tris	2-amino-2-(hydroxymethyl)-1,3-propandiol
uv	ultra-violet
V	Volt
V	volume
W	weight
$\chi^2$	chi-squared