

AGNP Consensus Guidelines for Therapeutic Drug Monitoring in Psychiatry: Update 2011

Authors

C. Hiemke¹, P. Baumann², N. Bergemann³, A. Conca⁴, O. Dietmaier⁵, K. Egberts⁵, M. Fric⁷, M. Gerlach⁶, C. Greiner⁸, G. Gründer⁹, E. Haen¹⁰, U. Havemann-Reinecke¹¹, E. Jaquenoud Siro¹², H. Kirchherr¹³, G. Laux⁷, U. C. Lutz¹⁴, T. Messer¹⁵, M. J. Müller¹⁶, B. Pfuhlmann¹⁷, B. Rambeck¹⁸, P. Riederer¹⁷, B. Schoppek¹⁹, J. Stingl²⁰, M. Uhr²¹, S. Ulrich²², R. Waschgler²³, G. Zernig²⁴

Affiliations

Affiliation addresses are listed at the end of the article

Key words

- consensus guidelines
- drug analysis
- pharmacokinetics
- psychotropic drugs
- reference ranges
- therapeutic drug monitoring
- therapeutic window

Abstract

Therapeutic drug monitoring (TDM), i.e., the quantification of serum or plasma concentrations of medications for dose optimization, has proven a valuable tool for the patient-matched psychopharmacotherapy. Uncertain drug adherence, suboptimal tolerability, non-response at therapeutic doses, or pharmacokinetic drug-drug interactions are typical situations when measurement of medication concentrations is helpful. Patient populations that may predominantly benefit from TDM in psychiatry are children, pregnant women, elderly patients, individuals with intelligence disabilities, forensic patients, patients with known or suspected genetically determined pharmacokinetic abnormalities or individuals with pharmacokinetically relevant comorbidities. However, the potential benefits of TDM for optimization of pharmacotherapy can only be obtained if the method is adequately integrated into the clinical treatment process. To promote an appropriate use of TDM, the TDM expert group of the Arbeitsgemeinschaft für Neuropsychopharmakologie und Pharmakopsychiatrie (AGNP) issued guidelines for TDM in psychiatry in 2004. Since then, knowledge has advanced significantly, and new psychopharma-

cologic agents have been introduced that are also candidates for TDM. Therefore the TDM consensus guidelines were updated and extended to 128 neuropsychiatric drugs. 4 levels of recommendation for using TDM were defined ranging from “strongly recommended” to “potentially useful”. Evidence-based “therapeutic reference ranges” and “dose related reference ranges” were elaborated after an extensive literature search and a structured internal review process. A “laboratory alert level” was introduced, i.e., a plasma level at or above which the laboratory should immediately inform the treating physician. Supportive information such as cytochrome P450 substrate- and inhibitor properties of medications, normal ranges of ratios of concentrations of drug metabolite to parent drug and recommendations for the interpretative services are given. Recommendations when to combine TDM with pharmacogenetic tests are also provided. Following the guidelines will help to improve the outcomes of psychopharmacotherapy of many patients especially in case of pharmacokinetic problems. Thereby, one should never forget that TDM is an interdisciplinary task that sometimes requires the respectful discussion of apparently discrepant data so that, ultimately, the patient can profit from such a joint effort.

Bibliography

DOI <http://dx.doi.org/10.1055/s-0031-1286287>
Pharmacopsychiatry 2011; 44: 195–235
© Georg Thieme Verlag KG
Stuttgart · New York
ISSN 0176-3679

Correspondence

C. Hiemke, PhD, Univ.-Prof.
Department of Psychiatry and Psychotherapy
University Medical Center,
Mainz
D-55101 Mainz
Germany
Tel.: +49/6131/177 131
Fax: +49/6131/176 789
hiemke@uni-mainz.de

Introduction

In psychiatry, around 130 drugs are now available which have been detected and developed during the last 60 years [54]. These drugs are effective and essential for the treatment of many psychiatric disorders and symptoms. Despite enormous medical and economic benefits, however, therapeutic outcomes are still far from satisfactory for many patients [5,6,396,661]. Therefore, after having focused clinical research on the development of new drugs during more

than 5 decades [521,522], growing evidence suggests that improving the way the available medications are administered may bring substantial benefit to patients [45]. Evidence-based guidelines for optimum treatment have been published during the last decade [23,46,101,204,205,221,234,254,276,284,582,585,748]. A valuable tool for tailoring the dosage of the prescribed medication(s) to the individual characteristics of a patient is therapeutic drug monitoring (TDM). The major reason to use TDM for the guidance of psychopharmacotherapy is the

considerable interindividual variability in the pharmacokinetic properties of the patient [524,526]. At the very same dose, a more than 20-fold interindividual variation in the medication's steady state concentration in the body may result, as patients differ in their ability to absorb, distribute, metabolize and excrete drugs due to concurrent disease, age, concomitant medication or genetic peculiarities [61,94,310,311,334,335,374]. Different formulations of the same medication may also influence the degree and temporal pattern of absorption and, hence, medication concentrations in the body. TDM uses the quantification of drug concentrations in blood plasma or serum to titrate the dose of individual patients so that a drug concentration associated with highest possible probability of response and tolerability and a low risk of toxicity can be obtained. Moreover, TDM has the possible and widely unexploited potential to improve cost-effectiveness of psychopharmacotherapy [527,660]. For a considerable number of psychopharmacologic compounds, the quantification of the medications' plasma concentration has become clinical routine for dose adjustment. Clear evidence of the benefits of TDM has been given for tricyclic antidepressants, a number of old and new antipsychotic drugs and for conventional mood stabilizing drugs [51,459,505]. For lithium, TDM has become a standard of care due to its narrow therapeutic range [133,395].

The benefits of TDM regarding the optimization of pharmacotherapy, however, can only be obtained if the method is adequately integrated into the clinical treatment process. Current TDM use in psychiatric care is obviously suboptimal [134,700,742]. Similar to other medical disciplines, systematic studies have demonstrated that the inappropriate use of TDM is widespread. Inappropriate TDM testing wastes laboratory resources and also bears the risk that misleading results will adversely influence clinical decision making [122]. A study on the clinical use of TDM for tricyclic antidepressants in psychiatric university hospital settings showed that between 25 and 40% of the requests for TDM were inappropriate and the interpretation of the results led to about 20% of inappropriate therapeutic adjustments [700,742]. Other typical errors were absence of steady-state conditions and transcription errors on the request form [700,743]. Studies on TDM for antidepressant and mood stabilizing drugs further specified the information on the inappropriate use of TDM [420,421].

Against this background, the TDM group of the Arbeitsgemeinschaft für Neuropsychopharmakologie und Pharmakopsychiatrie (AGNP) issued best practice guidelines for TDM in psychiatry in 2004 [51]. These guidelines were widely accepted by many laboratories and practicing clinicians. They have been cited more than 200 times in the literature and were translated into German [312] and French [50]. Moreover, they were summarized for depression [52]. The AGNP-TDM consensus guidelines have also been implemented in recent international guidelines on the treatment of mental diseases [582]. Since 2004, knowledge on TDM has advanced significantly. New psychotropic medications have been introduced which are also candidates for TDM. The TDM group of the AGNP therefore decided to prepare an updated version of their guidelines.

Objectives of this Consensus Document

This document addresses topics related to the theory and practice of TDM in psychiatry. The first part deals with theoretical

aspects of monitoring drug plasma concentrations. The second part defines indications for TDM and gives reference drug plasma concentrations for dose optimization. The third part describes the best practice of the process of TDM, which starts with the request and ends with the clinical decision to either continue or change the pre-TDM pharmacotherapy.

Aiming to optimise the practice of TDM the following topics were addressed:

- ▶ definition of indications to utilize TDM in psychiatry
- ▶ definition of graded levels of recommendations to use TDM
- ▶ definition of therapeutic reference ranges (“therapeutic windows”) and dose-related reference ranges that laboratories can quote and clinicians can use to guide the psychopharmacotherapy
- ▶ definition of alert levels for laboratories to warn the treating physician when plasma concentrations are considered to be too high and potentially harmful
- ▶ recommendations and help for interpretative services
- ▶ recommendations concerning the combination of TDM with pharmacogenetic tests

Preparation of the Consensus Document

The updated consensus guidelines were prepared by the interdisciplinary TDM group of the AGNP consisting of clinical psychiatrists, pharmacologists, biochemists, pharmacists and chemists from academic and non academic hospitals and institutions of Germany, Switzerland, Austria and Italy, who have been involved for many years in the development and implementation of TDM for psychotropic medications in everyday clinical practice. The experts compiled information from the literature and worked out the present best practice guidelines aiming at promoting the appropriate use of TDM in psychiatry. Because TDM is widely used in daily clinical practice for antidepressant, antipsychotic and mood stabilizing drugs, these 3 pharmacologic classes are extensively represented in the present guidelines. Anxiolytic and hypnotic drugs, antimentia drugs, drugs for treatment of substance abuse related disorders and other psychotropic drugs are also candidates for TDM and are thus covered in the present guidelines. In special situations, the measurement of drug plasma concentrations can be helpful for any drug. Many patients are simultaneously treated for neurologic and psychiatric disorders. Therefore, the updated guidelines also contain information on anticonvulsant and antiparkinson drugs which are also more or less well established candidates for TDM [481,499] and were thus extended from 65 psychiatric drugs in 2004 [51] to 128 neuropsychiatric drugs at present.

Data published in the AGNP consensus guidelines 2004 [51] and other guidelines and recommendations for TDM of primarily antidepressant and antipsychotic drugs [317,400,488–490,504,505] were initially used as the basis for this update. An extensive literature search was conducted, primarily in MEDLINE, to identify TDM-related information for the surveyed 128 neuropsychiatric drugs. The search concentrated on reports on “optimum plasma concentrations”, “dose related drug plasma concentrations”, “cytochrome P450 substrate, inducer and inhibitor properties” and on “ratios of concentrations of drug metabolites to parent drugs”. Relevant reports were also searched by hand in pharmacologic and clinical chemical journals dealing with TDM. Over 1 000 articles were assessed and

analysed. Extracted data on reference ranges were listed in tables by 7 authors (CH, EH, CG, BR, PR, HK). Results of the literature search and analyses were sent out for review to 20 members of the TDM group with inclusion of a checklist how to extract and analyse the data. An internet based and password-protected platform was built up for the reviewers to have access to relevant articles. The reviewers' protocols and commentaries were distributed to all authors of these guidelines. Final decisions on data reported in this document were made during 2 consensus conferences and by e-mail communication. Consensus making also included definitions of reference ranges, alert levels and graded levels of recommendations to utilize TDM.

Theoretical Aspects of TDM in Psychiatry

Pharmacokinetics, metabolism and pharmacogenetics of neuropsychiatric drugs

Most psychotropic drugs share a number of pharmacokinetic characteristics

- ▶ good absorption from the gastrointestinal tract within plasma concentrations reaching a maximum within 1–6h
- ▶ highly variable first-pass metabolism (systemic bioavailability ranging 5–90%)
- ▶ fast distribution from plasma to the central nervous system with 2- to 40-fold higher levels in brain than in blood
- ▶ high apparent volume of distribution (about 10–50L/kg)
- ▶ low trough plasma concentrations under steady-state (about 0.1–500 ng/mL for psychoactive drugs and up to 20 µg/mL for neurological drugs)
- ▶ slow elimination from plasma (half-life 12–36h) mainly by hepatic metabolism
- ▶ linear pharmacokinetics at therapeutic doses which has the consequence that doubling the daily dose will result in a doubling of the plasma level
- ▶ low renal excretion with small effect of renal insufficiency on the plasma concentrations of parent drug and active metabolites
- ▶ cytochrome P450 (CYP) and UDP-glucuronosyltransferases as major metabolic enzyme systems

There are, however, numerous exceptions. For example, venlafaxine, nefazodone, trazodone, tranylcypromine, moclobemide, quetiapine, rivastigmine and ziprasidone display short (about 2–10h) elimination half-lives, whereas aripiprazole and fluoxetine have long elimination half-lives (72h for aripiprazole and 3–15 days for fluoxetine, taking into account its active metabolite norfluoxetine). Amisulpride, milnacipran, memantine, gabapentin, or sulpiride are not or only poorly metabolised in the liver but also mainly excreted renally. Paroxetine exhibits non-linear pharmacokinetics, due to the inhibition of its own metabolism by a metabolite which is irreversibly bound to the enzyme (mechanism based inhibition) resulting in its inactivation [69].

Many psychotropic drugs are used as racemic compounds, and their enantiomers differ markedly in their pharmacology, metabolism and pharmacokinetics [53,605]. So far however, methadone, methylphenidate and flupentixol are at present the only racemic psychotropic compounds for which TDM of the enantiomers has been introduced [39,189]. The active principles of racemic methadone and fluoxetine are (R)-methadone and cis-(Z)-flupentixol, respectively. For research projects and other special situations, stereoselective analysis should be considered, e.g., for citalopram, fluoxetine, reboxetine, venlafaxine, paliperidone or amitriptyline metabolites.

Most psychotropic drugs undergo phase-I metabolism by oxidative (e.g., hydroxylation, dealkylation, oxidation to N-oxides, S-oxidation to sulfoxides or sulfones), reductive (e.g., carbonyl reduction to secondary alcohols) or hydrolytic reactions, dealkylation, oxidation to N-oxides, carbonyl reduction to secondary alcohols or S-oxidation to sulfoxides or sulfones. The phase-I reactions are predominantly catalysed by cytochrome P450 (CYP) enzymes which comprise more than 200 isoenzymes. The most important isoenzymes for psychotropic medications are CYP1A2, CYP2B6, CYP2D6, CYP2C9, CYP2C19 and CYP3A4/5 (● **Table 1**) [745–747]. In general, phase-I reactions introduce a polar functional group that enables a phase-II conjugation reaction with highly polar molecules such as glucuronic or sulphuric acid. For psychotropic compounds possessing functional groups in the parent compound, glucuronidation of a hydroxyl group (for example oxazepam or lorazepam) or an N-H group (for example olanzapine) may represent the essential metabolic pathway. In addition, tertiary amine groups can be conjugated with the formation of quaternary ammonium glucuronides. Actually, phase II enzymes are poorly characterised with regard to substrate specificity, and there is much overlap between the isozymes regarding affinity for substrates [143].

Other enzymatic systems may also be involved, such as ketoaldehyde oxidases [43], which have been shown to reduce ziprasidone to its dihydro-derivative [58] or naltrexone to naltrexol [92], or MAO-A and MAO-B, which deaminate citalopram stereoselectively to an apparently inactive acidic metabolite [562].

Drugs are metabolised mainly in the liver and, to a minor degree, in extrahepatic tissues such as the intestinal mucosa or the brain [59,238,444]. Inter- and intra-individual differences in plasma concentrations of psychotropic drugs (i.e., the pharmacokinetic variability) are caused by different activities of drug-metabolising enzymes. The enzyme activity may decrease with age [374] and can be modified by renal and hepatic diseases. Gender differences have been reported for psychotropic drugs, but the findings are inconsistent and the clinical relevance is not clear [7–9,608].

For a number of psychoactive drugs, metabolites actively contribute to the overall clinical effect of the parent compound. For this reason, TDM must include the quantification of active metabolites, e.g., in the case of clomipramine (norclomipramine), doxepin (nordoxepin), fluoxetine (norfluoxetine) or risperidone (9-hydroxyrisperidone). For drugs like sertraline or clozapine, the clinical relevance of their metabolites norsesertraline and norclozapine, respectively, is still a matter of debate. The analysis of pharmacologically inactive metabolites, however, may give useful information on the metabolic state of the patient or on his/her compliance [105,569]. ● **Table 2** shows the “normal” ratios of concentrations of metabolites to parent drugs. Calculated ranges contain 68% of the ratios expected under standard dosages, i.e., ratios within the range of the mean ± 1 SD assuming normal distribution. A ratio above or below the “normal ratio” (● **Table 2**) can indicate problems of drug adherence [546] or metabolic abnormalities due to a genetic variation [157,159,350,592] or a drug-drug interaction. Spina and co-workers [618] have shown this for the conversion of 2-hydroxydesipramine to desipramine. With regard to drug-drug interactions, ratios increase if the enzymatic conversion of the parent medication is induced by concurrent psychotropic or non-psychotropic medications or pharmacokinetically relevant activities such as smoking (● **Table 3**). Other co-medications and food

Table 1 Psychopharmacologic medications and enzymes involved in their metabolism.

Drug (active metabolite)	Enzymes	Reference
Acamprosate	not involved (not metabolized)	[578]
Agomelatine	CYP1A2 , CYP2C19	[78]
Amantadine	merely involved (90% excreted unmetabolized)	[24]
Alprazolam	CYP3A4/5	[17, 496]
Amisulpride	merely involved (more than 90% is excreted unmetabolized via the kidney)	[566]
Amitriptyline and amitriptyline oxide (amitriptyline, nortriptyline)	CYP1A2, CYP2C9, CYP2C19 , CYP2D6 , CYP3A4	[90, 650, 713]
Aripiprazole (dehydroaripiprazole)	CYP2D6 , CYP3A4	[306, 701]
Asenapine	Glucuronosyltransferase and CYP1A2	[707]
Atomoxetine	CYP2D6	[446]
Benperidol	unclear	[589]
Benserazide	hydroxylation , COMT	[347]
Biperiden	hydroxylation	[628]
Bromocriptine	CYP3A4	[513]
Bromperidol	CYP3A4	[230, 633, 645, 736]
Brotizolam	CYP3A4	[655]
Buprenorphine (norbuprenorphine)	CYP2C8, CYP3A4	[79, 454]
Bupropion (hydroxybupropion)	CYP2B6	[309]
Buspirone	CYP3A4	[416]
Cabergoline	hydrolysis , CYP3A4	[167]
Carbidopa	unknown metabolic pathways 1/3 unmetabolized	[575]
Carbamazepine, CBZ (CBZ-10,11-epoxide)*	CYP1A2, CYP2B6, CYP2C8, CYP3A4/5	[360, 497]
Chlorpromazine	CYP1A2 , CYP2D6	[724]
Citalopram	CYP2C19 , CYP2D6, CYP3A4	[97, 227, 739]
Clomipramine (norclomipramine)	CYP1A2, CYP2C19 , CYP2D6 , CYP3A4	[244]
Clomethiazol	CYP2A6, CYP2B6, CYP3A4	[116]
Clozapine	CYP1A2 , CYP2C19 , CYP3A4	[334, 487]
Desipramine	CYP2D6	[244]
Diazepam (nordazepam, oxazepam, temazepam)	CYP2B6, CYP2C19 , CYP3A4	[228, 704]
Dihydroergocryptine	CYP3A4	[19, 162]
Diphenhydramine	CYP2D6	[13]
Disulfiram	CYP1A2, CYP2B6, CYP2E1, CYP3A4	[412]
Donepezil	CYP2D6 , CYP3A4	[681]
Dothiepin = Dosulepin	CYP2C19 , CYP2D6	[740]
Doxepin (nordoxepin)	CYP2C9 , CYP2C19 , CYP2D6	[295, 365]
Duloxetine	CYP1A2 , CYP2D6	[405]
Entacapone	Glucuronosyltransferase	[387]
Escitalopram	CYP2C19 , CYP2D6, CYP3A4	[662, 697]
Fluoxetine (norfluoxetine)	CYP2B6, CYP2C9 , CYP2C19 , CYP2D6	[404, 588]
Flupenthixol	CYP2D6	[148, 365]
Fluphenazine	CYP2D6	[746]
Fluvoxamine	CYP2D6 , CYP1A2	[354, 450]
Galantamine	CYP2D6, CYP3A4	[34]
Gabapentin	unmetabolized renal excretion	[77]
Haloperidol	CYP2D6, CYP3A4	[93, 645]
lloperidone	CYP2D6 , CYP3A4	[106]
Imipramine (desipramine)	CYP1A2 , CYP2C19 , CYP2D6 , CYP3A4	[244, 413]
Lamotrigine	Glucuronosyltransferase, CYP2A6	[121]
Levodopa	Dopadecarboxylase , COMT, MAO	[575]
Levomepromazine	CYP1A2, CYP2D6	[36]
Levomethadon	CYP19, CYP2B6, CYP3A4 , CYP2D6	[145]
Lisuride	CYP3A4, CYP2D6	[539]
Lithium	no metabolism, renal clearance	[256, 619]
Lorazepam	Glucuronosyltransferase	[164, 196]
Maprotiline	CYP2D6 , CYP1A2	[86]
Melatonin	CYP1A2	[296]
Memantine	merely metabolized	[251]
Methadone	CYP2B6 , CYP2C19, CYP3A4, CYP2D6	[145]
Methylphenidate	Carboxylesterase 1	[468]
Mianserine	CYP2D6 , CYP1A2, CYP3A4	[379]
Midazolam	CYP3A4	[220]
Milnacipran	no CYP related metabolism	[495, 533]

Table 1 Continued.

Drug (active metabolite)	Enzymes	Reference
Mirtazapine	CYP3A4, CYP1A2, CYP2B6, CYP2D6	[397, 630]
Moclobemide	CYP2C19 , CYP2D6	[255]
Modafinil	Amide hydrolysis, CYP3A4	[561]
Naltrexone	Aldoketoreductase AKR1C4	[92]
Nortriptyline	CYP2D6	[385, 485, 687]
Olanzapine	N-Glucuronosyltransferase, Flavin monooxygenase, CYP1A2 , CYP2D6	[107]
Opipramol	unclear	
Paliperidone (= 9-Hydroxyrisperidone)	60% excreted unmetabolized, different pathways	[161]
Paroxetine	CYP1A2, CYP2D6 , CYP3A4	[209, 349, 691]
Perazine	CYP1A2, CYP2C19 , CYP3A4, Flavin monooxygenase	[629, 725]
Pergolide	CYP3A4	[731]
Perphenazine	CYP1A2, CYP2C19, CYP2D6 , CYP3A4	[12, 77, 168, 486]
Pregabalin	unmetabolized renal excretion	[77]
Piripedit	demethylation, p-hydroxylation, and N-oxidation	[168]
Pimozide	CYP1A2, CYP3A4	[171]
Pramipexole	not metabolized	[62]
Promazine	CYP1A2, CYP2C19, CYP3A4	[726]
Promethazine	CYP2D6	[465]
Quetiapine	CYP3A4 , CYP2D6	[38]
Rasagiline	CYP1A2	[277]
Reboxetine	CYP3A4	[307, 716]
Risperidone (9-Hydroxyrisperidone)	CYP2D6 , CYP3A4	[732]
Ropinirole	CYP1A2	[357]
Rotigotine	Glucuronosyltransferase, several other unknown pathways	[115]
Selegiline	CYP2B6	[60]
Sertindole	CYP3A4 , CYP2D6	[729]
Sertraline	CYP2B6 , CYP2C19 , CYP2C9, CYP2D6	[482, 705]
Thioridazine	CYP1A2 , CYP2C19, CYP2D6 , CYP3A4	[648, 714]
Tiapride	mainly not metabolized	[477]
Tolcapone	Glucuronosyltransferase	[387]
Trimipramine (nortrimipramine)	CYP2C19 , CYP2D6 , CYP2C9	[187]
Tranylcypromine	monoamine oxidase, unclear	[37]
Trazodone	CYP3A4 , CYP2D6	[268, 567]
Valproic acid	Glucuronosyltransferase , CYP2A6, CYP2B6, CYP2C9, beta-oxidation	[641]
Venlafaxine (O-desmethylvenlafaxine)	CYP2C19, CYP2D6 , CYP3A4	[217, 434]
Zaleplone	Aldehyde oxidase , CYP3A4	[554]
Ziprasidone	CYP3A4, Aldehyde oxidase	[58, 519]
Zolpidem	CYP1A2, CYP2C9, CYP3A4	[698]
Zopiclone	CYP2C8, CYP3A4	[57, 659]
Zotepine	CYP1A2, CYP2D6, CYP3A4	[596]
Zuclopenthixol	CYP2D6	[330]

Inhibition of enzymes indicated in bold will significantly increase the plasma concentrations of the drug, induction (CYP1A2, CYP3A4) will lead to decreased plasma concentrations (See ● Table 2). Prepared by CH, reviewed and supplemented by EJS

which inhibit metabolic enzymes may decrease the ratio. ● Table 3 summarizes drugs that are inhibitors or inducers of CYP enzymes and thus may lead to clinically relevant pharmacokinetic drug-drug interactions.

Pharmacogenetic aspects

The clinical importance of pharmacogenetic factors in the pharmacokinetics and pharmacodynamics of psychotropic drugs is increasingly recognised [156, 199, 457]. Drug-metabolising enzymes, especially CYP isoenzymes, exhibit genetic variability [745–747]. When the frequency of a deviation in the alleles is at least 1% of the population, it is considered a genetic polymorphism. The number of active alleles in a gene determines how much of the enzyme is expressed (phenotype). Poor metabolisers (PM) lack functional alleles. Intermediate metabolisers (IM) are either genetically heterozygous, carrying an active and an inactive allele (or an allele with reduced activity) or have 2 alle-

les with reduced activity. Extensive metabolisers (EM) are wild-type with 2 active alleles, and ultra-rapid metabolisers (UM) have an amplification of functional alleles [66]. Genetic polymorphisms of drug-metabolising enzymes may be clinically important, because unexpected adverse reactions and toxicity may occur in PM due to increased plasma concentrations and non-response may occur in UM due to subtherapeutic plasma concentrations [160]. Prodrugs are activated by metabolism such as codeine by CYP2D6 to morphine or clopidogrel by CYP2C19 to 2-oxoclopidogrel. PM patients will not be able to produce pharmacologically active metabolites. Other enzyme systems such as UDP-glucuronosyltransferases also display genetic polymorphism [155], but their clinical relevance in pharmacopsychiatry is unclear.

CYP genotyping methods are becoming more and more available, and guidelines have been published for their use in clinical practice [675]. The functional significance of many genotypes,

Table 2 Ranges of metabolite-to-parent concentration ratios for psychopharmacologic medications. Reported ranges contain 68% of ratios determined under “normal” conditions in the blood of patients or healthy subjects.

Drug	Metabolite	Ratios of concentrations metabolite: parent drug (Mean – SD – Mean + SD)	Reference
Amitriptyline	Nortriptyline*	0.2–1.8 (n = 83)	[545]
Aripiprazole	Dehydroaripiprazole(*)	0.3–0.5 PM of CYP2D6: 0.2	[306, 368, 452]
Bromperidol	Reduced bromperidol	0.11–0.51 (n = 31)	[609, 633]
Buprenorphine	Norbuprenorphine	0.8–2.0 (n = 5)	[383]
Bupropion	Hydroxybupropion	5–47 (24 h, n = 9) 6–30 (12 h, n = 9)	[152, 253, 336]
Buspirone	6-Hydroxybuspirone	25–53 (n = 20)	[178]
Carbamazepine	Carbamazepine-10,11-epoxide	0.07–0.25 (n = 14)	[338]
Citalopram	N-Desmethylcitalopram	0.31–0.60 (n = 2 330)	[549]
Clomipramine	Norclomipramine*	0.8–2.6 (n = 115)	[545]
Clozapine	Norclozapine(*)	nonsmokers (n = 98) 0.5–0.6 smokers (n = 198) 0.4–0.7	[140, 308, 500]
Dothiepin	Nordothiepin	0–1.4 (n = 50)	[325]
Doxepin	Nordoxepin	0.6–1.6 (n = 12) PM CYP2C19: 1.8 (n = 4) PM CYP2D6: 0.8 (n = 6)	[172, 363]
Escitalopram	N-Demethylescitalopram	0.3–1.0 (n = 243)	[548]
Fluoxetine	Norfluoxetine*	0.7–1.9 (n = 334)	[545]
Fluvoxamine	Fluvoxamine acid	0–1.2 (n = 49)	[237]
Haloperidol	Reduced haloperidol	mean 0.6	[673]
Imipramine	Desipramine	0.6–3.2 (n = 14) PM CYP2D6 4.1 (n = 2)	[95, 96, 632]
Maprotiline	Desmethylmaprotiline	1.1–3.7 (n = 76) PM CYP2D6 4.9	[699]
Mianserin	N-Desmethylmianserin	0.5–0.8 (n = 182)	[545]
Mirtazapine	N-Desmethylmirtazapine	0.2–1.2 (n = 100)	[591]
Moclobemide	Moclobemide N-oxide	0.8–2.5 (n = 6)	[291]
Olanzapine	N-Demethylolanzapine	non smokers: 0.1–0.3 (n = 76) smokers: 0.2–0.4 (n = 69)	[602]
Perazine	Desmethylperazine	1.1–3.3 (n = 27)	[91]
Perphenazine	N-Dealkylperphenazine	0.6–2.8 (n = 54)	[637]
Quetiapine	Norquetiapine	0.1–3.8 (n = 25) (calculated for 400 mg)	[723]
Reboxetine	O-Desethylreboxetine	<0.1	[484]
Risperidone	9-Hydroxyrisperidone*	EM or IM CYP2D6: 1.5–10.0 PM CYP2D6: ≤ 1	[159, 677]
Risperidone depot	9-Hydroxyrisperidone*	EM: 1.2–4.3	[469]
Sertindole	Dehydrosertindole	1.1–2.7 (n = 6) 1.0 in PM of CYP2D6	[729]
Sertraline	Norserttraline	1.7–3.4 (n = 348)	[546]
Trazodone	m-Chlorophenylpiperazine (mCPP)	0.04–0.22 (total range)	[328]
Trimipramine	Nortrimipramine*	0–12.0 (n = 17)	[142]
Venlafaxine	O-Desmethylvenlafaxine*	EM or IM CYP2D6: 0.3–5.2 PM CYP2D6: ≤ 0.3 UM CYP2D6: > 5.2	[592]
	N-Desmethylvenlafaxine	0.46–1.48	

* pharmacologically active metabolite, (*) active metabolite in vitro but unclear under in vivo conditions

When SD values of ranges of ratios (SD ratio) were not reported in the literature, SD ratios were calculated in accordance with Gaussian's law for the propagation of errors: SD ratio = [(SD parent drug x mean metabolite) + (SD metabolite x mean parent drug)] / ((mean metabolite)²)

Prepared by CH, reviewed by Sonja Brünen, Christiane Knoth, Elnaz Ostad Haji and Viktoria Stieffenhofer

however, is unclear. For some enzymes, a genetic polymorphism is not clearly demonstrated despite the fact that they display a wide interindividual variability in their activity. Therefore it may be advantageous to use phenotyping methods with probe drugs such as caffeine for CYP1A2, omeprazole for CYP2C19, dextromethorphan for CYP2D6, or midazolam for CYP3A4/5 [403, 643]. Phenotyping measures the metabolic situation of the

patient at the moment of the test, and allows to follow its evolution. The measurement, however, may be influenced by environmental factors such as smoking or comedications [201, 601, 749]. The clear advantage of genotyping is that it represents a “trait marker” and that its result is not influenced by environmental factors. It can be carried out in any situation and its result has a lifetime value.

Table 3 Inhibitors and inducers of enzymes involved in the metabolism of drug.

Inhibiting drugs	Inhibited enzymes	Inducing drugs	Induced enzymes
Amiodarone	CYP2C9, CYP2D6, CYP3A4	Carbamazepine	CYP1A2, CYP2B6, CYP2C9, CYP3A4
Bupropion	CYP2D6	Dexamethason	CYP2C9, CYP3A4
Bromocriptine	CYP3A4	Efavirenz	CYP2B6, CYP3A4
Chinidine	CYP2D6	Ethanol	CYP2E1
Cimetidin	CYP1A2, CYP2D6, CYP3A4	Ginkgo biloba	CYP2C19
Ciprofloxacin	CYP1A2	Isoniazide	CYP2E1
Clarithromycin	CYP3A4	St. John's wort	CYP2C19, CYP3A4
Clopidogrel	CYP2B6	Oxybutynin	CYP3A4
Disulfiram	CYP2E1	Phenobarbital	CYP2C9, CYP2C19, CYP3A4
Duloxetine	CYP2D6	Phenytoin	CYP2B6, CYP2C9, CYP2C19, CYP3A4
Enoxacin	CYP1A2	Primidon	CYP2C9, CYP2C19, CYP3A4
Erythromycin	CYP3A4	Smoke	CYP1A2
Esomeprazole	CYP2C19	Rifabutin	CYP3A4
Felbamate	CYP2C19	Rifampicin	CYP1A2, CYP2B6, CYP2C9, CYP2C19
Fluconazole	CYP2C19, CYP2C9, CYP3A4	Ritonavir	CYP3A4, CYP2C9, CYP3A4 (high dose)
Fluoxetine and norfluoxetine	CYP2D6, CYP2C19		
Fluvoxamine	CYP1A2, CYP2C9, CYP2C19, CYP3A4		
Indinavir	CYP3A4		
Isoniazid	CYP1A2, CYP2A6, CYP2C19, CYP3A4		
Itraconazol	CYP2B6, CYP3A4		
Ketoconazol	CYP3A4		
Levomepromazine	CYP2D6		
Melperone	CYP2D6		
Metoclopramide	CYP2D6		
Metoprolol	CYP2D6		
Miconazol	CYP2C9, CYP2C19		
Mifepriston	CYP3A4		
Moclobemide	CYP2C19, CYP2D6		
Nelfinavir	CYP3A4		
Norfloxacin	CYP1A2		
Omeprazole	CYP2C19		
Paroxetine	CYP2D6		
Perazine	CYP1A2		
Pergolide	CYP2D6		
Perphenazin	CYP2D6		
Propafenon	CYP1A2, CYP2D6		
Propranolol	CYP2D6		
Ritonavir	CYP2D6, CYP3A4		
Saquinavir	CYP3A4, CYP2C9		
Troleandomycin	CYP3A4		
Valproate	CYP2C9		
Verapamil	CYP3A4		
Voriconazol	CYP2C9, CYP3A4		

Combination of psychoactive drugs with these inhibitors or inducers can lead to clinically relevant drug-drug interactions (www.mediq.ch or www.psiac.de)

Prepared by CH, reviewed by EJS

Recent investigations indicate that the drug efflux transporter P-glycoprotein (P-gp) in the intestinal mucosa and blood-brain-barrier is also relevant for the pharmacokinetic variability of psychotropic medications [1]. This protein, a member of the ATP-cassette binding (ABC) transporter protein family, is encoded by the multidrug resistance gene (*MDR1*; *ABCB1*). It displays a genetic polymorphism, but as yet, mainly genotyping but not phenotyping (e.g., with digoxin) is more commonly used [129, 183, 210, 389]. Genetic polymorphism of P-gp may be of the same considerable clinical relevance as has been demonstrated for drug-metabolizing enzymes. For antidepressant drugs that are substrates of P-gp, a genotype dependent association of drug response was found [668, 669]. Both plasma concentrations of quetiapine and its clinical effectiveness have been shown to depend on the P-gp genotype of patients suffering from schizophrenia [470]. With regard to the occurrence of

wanted or unwanted clinical effects of psychoactive drugs, some first reports suggest the influence of the genetic polymorphism of P-gp [279, 560]. However, further research is needed to evaluate the clinical relevance of the genetic polymorphisms of drug transporters.

Dose and drug concentration in blood

In most situations that use TDM for dose optimization, drugs are administered in a series of repeated doses to attain a steady-state concentration within a given therapeutic reference range. Steady-state is attained when the rate of medication input equals the rate of medication loss, i.e., approximately after 4 times the elimination half life. With multiple dosing, 94% of the steady state are achieved after 4 and 97% after 5 elimination half-lives. For more than 90% of all psychoactive medications, such a steady-state is reached within 1 week of maintenance

Table 4 Total clearance (Cl_t), bioavailability (F), dosing intervals (τ) and factors (C/D_{low} and C/D_{high}) for calculation of dose-related plasma concentrations (C/D) for psychotropic drugs.

Drug	n	$Cl_t - SD - Cl_t + SD$ [mL/min]	F	τ [h]	C/D_{low} [ng/mL/mg]	C/D_{high} [ng/mL/mg]	Reference
Antidepressant drugs							
Amitriptyline	8	198–373	0.5	24	1.03	1.68	[165]
Amitriptyline oxide	12	331–539	0.8	24	0.93	1.75	[384]
Bupropion	17	2500–11 300	1.0	24	0.06	0.28	[665]
Citalopram	8	367–545	0.8	24	1.02	1.51	[616]
Clomipramine	9	583–933	0.5	24	0.37	0.60	[198]
Desipramine	12	1 633–2 333	0.5	24	0.15	0.21	[2]
Desvenlafaxine	7	233–396	1.0	24	1.75	2.98	[520]
Dothiepin = Dosulepin	22	674–3 960	0.3	24	0.05	0.31	[740]
Doxepin	85	769–2 644	1.0	24	0.18	0.27	[100]
Duloxetine	12	610–1 733	0.5	24	0.20	0.57	[600]
Escitalopram	24	360–960	0.8	24	0.58	1.54	[607]
Fluoxetine	n.r.	600–833	0.7	24	0.60	0.83	[18]
Fluvoxamine	6	807–1 960	1.0	24	0.35	0.86	[163]
Imipramine	n.r.	791–1 029	0.4	24	0.28	0.37	[100]
Maprotiline	6	503–1 747	0.8	24	0.32	1.10	[415]
Mianserin	n.r.	843–1 948	0.3	24	0.11	0.25	[137]
Mirtazapine	10	455–945	0.5	24	0.37	0.85	[651]
Nordoxepin	85	504–2 738	1.0	24	0.25	1.38	[445]
Nortriptyline	n.r.	300–1 117	0.5	24	0.31	1.16	[664]
Paroxetine	30	1 561–10 856	1.0	24	0.06	0.44	[213]
Reboxetine	n.r.	22–51	1.0	24	12.55	31.10	[141]
Sertraline	11 (m)	1 313–2 213 (m)	1.0	24	0.31	0.53	[565]
	11 (f)	793–2 357 (f)	1.0	24	0.29	0.88	
Trazodone	8	73–103	1.0	24	6.72	9.47	[473]
Trimipramine	12	898–1 215	0.40	24	0.23	0.31	[165, 364]
Venlafaxine	18	747–1 540	1.0	24	0.45	0.93	[372]
O-Desmethylenlafaxine		315–618	1.0	24	1.12	2.2	
Antipsychotic drugs							
Amisulpride	78	520–693	0.5	24	0.50	0.67	[566]
Asenapine	n.r.	867	0.35	24	0.28		[707]
Aripiprazole	6	47–70	0.9	24	8.63	12.85	[417]
Benperidol	14	1 073–2 240	0.5	24	0.15	0.31	[589]
Bromperidol	14	3 570–7 938	1.0	24	0.09	0.19	[390]
Chlorpromazine	11	1 043–1 510	0.1	24	0.05	0.07	[738]
Chlorprothixene	3	918–1 448	0.2	24	0.10	0.15	[534]
Clozapine	16	258–728	0.5	24	0.40	0.80	[128, 176, 332]
Flupentixol	3	440–490	0.6	24	0.78	0.87	[348]
Fluphenazine decanoate	12	2 380–3 940	1.0	24	0.18	0.29	[197]
Haloperidol	6	420–680	0.6	24	0.61	0.99	[123]
Haloperidol decanoate		420–680	1.0	336	0.073	0.118	[123]
				672	0.036	0.059	
Melperone	6	1 484–2 898	0.6	24	0.14	0.28	[83]
Levomepromazine	8	913–4 737	0.5	24	0.07	0.38	[149]
Olanzapine	491	233–637	0.8	24	0.87	2.38	[67]
Paliperidone	n.r.	31–98	0.3	24	1.99	6.31	[161]
Perphenazine	8	1 009–2 566	0.4	24	0.11	0.28	[195]
Pimozide	7	21–553	0.5	24	0.64	16.53	[581]
Quetiapine	10	1 146–2 421	1.0	24	0.13	0.21	[7, 435]
Risperidone, oral	8	91–171	0.7	24	3.50	14.00	[159]
Risperidone, depot	n.r.	91–171	1.0	336	0.29	0.55	[606]
					active moiety	active moiety	
					active moiety	active moiety	
Sertindole	6	133–600	1.0	24	1.16	5.22	[728]
Supiride	6	331–499	0.25	24	0.35	0.52	[717]
Thiordazine	11	404–982	0.60	24	0.42	1.03	[117]
Zotepine	14	467–10 267	1.0	24	0.07	1.49	[642]
Ziprasidone	12	303–397	0.6	24	1.05	1.36	SPC
Zuclopenthixol	8	867–2 300	0.4	24	0.13	0.35	[337]

Table 4 Continued.

Drug	n	Cl _t – SD – Cl _t + SD [mL/min]	F	τ [h]	C/D _{low} [ng/mL/mg]	C/D _{high} [ng/mL/mg]	Reference
Anticonvulsant drugs Mood stabilizers							
Carbamazepine	n.r.	58–74	1.0	24	9.40	11.93	SPC
Felbamate	10	29.1–33.3	1.0	24	20.85	23.86	[556]
Lamotrigine	129	22–49	1.0	24	14.09	31.28	[118]
Levetiracetam	216	52–72	1.0	24	9.65	13.35	[535]
Lithium	n.r.	10–40	1.0	24	17.36	69.44	[706]
Oxcarbazepine	7	1703–5063	1.0	24	0.14	0.41	[319, 694]
Primidone	8	30–47	1.0	24	14.78	23.15	[423]
Topiramate	6	21–31	1.0	24	22.47	33.55	[179]
Valproic acid	9	4.5–9.8	1.0	24	71.23	154.32	[682]
Anxiolytic and hypnotic drugs							
Alprazolam	6	34–83	0.8	24	6.73	16.53	[496, 604]
Bromazepam	10	50–91	1.0	24	7.67	13.95	[352]
Brotizolam	8	85–141	0.7	24	4.93	8.17	[341]
Buspirone	41	1260–2702	0.04	24	0.01	0.02	[41]
Clonazepam	9	63–90	0.8	24	5.43	7.69	[259]
Diazepam	48	10–43	0.9	24	13.01	52.91	[264]
Lorazepam	15	36–109	0.8	24	5.98	17.93	[266]
Oxazepam	18 (m) 20 (w)	36–167 29–109	0.8 0.8	24 24	3.33 5.12	15.22 18.90	[260]
Triazolam	13	326–584	0.9	24	1.01	1.81	[263]
Zaleplon	10	868–1330	0.3	24	0.16	0.25	[265]
Zolpidem	10	266–364	0.67	24	1.02	2.14	[265]
Zopiclone	10	250–883	1	24	0.79	2.78	[411]
Antidementia drugs							
Donepezil	14	112–217	1.0	24	3.20	6.20	[463]
Galantamine	8	268–400	1.0	24	1.74	2.59	[744]
Rivastigmine	20	29–64 (patch)	0.5	24	0.18	0.74	[391]
Drugs for treatment of substance related disorders							
Acamprosate	24	1741–4221	1.0	24	0.16	0.40	[287]
Buprenorphin							no data available
Bupropion	17	2500–11300	1.0	24	0.06	0.28	[665]
Methadone	12	75–148	0.95	24	4.46	8.80	[474, 727]
Naltrexone	453	2077–2590	1.0	24	0.27	0.33	[182]
6β-naltrexol		928–1242			0.56	0.75	
Varenicline	1878	170–176	1.0	24	3.95	4.08	[540]

SPC: Summary of product characteristics; n.r.: not reported; active moiety: risperidone plus 9-hydroxyrisperidone; n: number of individuals; SD: standard deviation

Dose related ranges are obtained by multiplying C/D_{low} and C/D_{high} by the dose. Drugs listed in Table 5 were not included in this table, when clearance data were not available from the literature.

Prepared by EH and CG, reviewed and supplemented by CH

dosing. The dose required to attain a steady-state concentration of a drug in plasma can be calculated if the dosing interval (τ), the clearance (Cl) and the bioavailability (F) for the drug in a particular patient are known. The calculation is based on the direct correlation of the drug dose D_e (constant dose per day at steady-state) to its blood concentration c, with the total clearance of the drug (Cl_t) being the correlation coefficient:

$$D_e = D \times F / \tau = c \times Cl_t$$

Based on this information it is possible to calculate the dose-related plasma concentration of a drug that may be expected in blood specimens of patients under medication with a given dose [285]:

$$c = D_e / Cl_t$$

For psychoactive medications, such data are available from studies in which drug concentrations were measured in plasma of healthy volunteers or patients treated with fixed doses. When the clearance is taken as arithmetic mean ± standard deviation

from clinical trials of the drug, a dose related reference range can be calculated [285].

Definition

The “dose-related reference range” reported in the present guidelines is calculated as a concentration range within that a drug concentration is expected according to pharmacokinetic studies in human blood specimens from subjects under medication with a given dose of the drug. It contains 68% of all the drug concentrations determined under normal conditions in the blood of a “normal” patient or subject, “normal” being defined by the population in the respective clinical trial. It usually consists of individuals 18–65 years of age without relevant comorbidity, comedication, and genetic abnormalities in drug metabolism.

Table 4 lists factors for calculation of dose-related reference ranges for the most relevant psychoactive drugs. Dose-related reference ranges are calculated by multiplying C/D_{low} and C/D_{high}

by the daily dose. One must be aware, however, that many patients encountered in the clinical context do not fulfil all the abovementioned conditions.

Drug concentration in blood and brain

The pharmacological activity of a psychotropic drug depends on its availability in the target organ, the brain. However, the latter is separated from the blood by 2 barriers, which have to be crossed by the drug, the blood-brain barrier (BBB) and the blood-cerebrospinal fluid barrier [154]. Most psychoactive drugs enter the brain due to their high lipid solubility by passive diffusion and thereby cross the barriers. The BBB is a physical barrier that separates circulating blood and the central nervous system, and it consists of endothelial cells around the capillaries joined together by tight junctions [154]. It efficiently restricts the exchange of solutes between the blood and the brain extracellular fluid. Functionally, it protects the brain against potentially harmful chemicals. As mentioned above, a number of psychoactive drugs, such as risperidone, aripiprazole or venlafaxine are substrates of P-gp [180,370,668]. As a consequence, brain to plasma concentration ratios vary widely for psychotropic drugs with similar physicochemical properties. Animal studies found ratios from 0.22 for risperidone [29] to 34 for fluphenazine [27]. In spite of highly variable ratios of brain to plasma concentrations of the different psychotropic drugs, animal studies have shown that steady-state plasma concentrations of psychoactive drugs correlate well with concentrations in brain, much better than doses. This has been shown for tricyclic antidepressants [249], trazodone [173], or olanzapine [28]. Drug concentrations in plasma can therefore be considered as a valid surrogate marker of concentrations in brain.

Drug concentration in blood and target structure occupancy in brain

Positron emission tomography (PET) enables analysis of central nervous receptor occupancy in vivo [207,274,275]. Antipsychotic drugs exert most of their therapeutic actions by blockade of dopamine D2-like receptors. Blockade of D2 receptors by antipsychotic drugs reduces the binding of radioactive PET ligands [207,272,275]. Using this approach and quantification of the displacement of dopamine receptor radioligands, it has been shown that plasma concentrations of antipsychotic drugs correlate well with receptor occupancy. In accordance with the high variability of drug concentrations in plasma under same doses it was found that receptor occupancy correlates better with plasma concentrations than with daily doses [313]. Optimal response was seen at 70–80% receptor occupancy, and 80% receptor occupancy was defined as the threshold for the occurrence of extrapyramidal side effects [207,480]. PET was also used to characterize in vivo serotonin transporter occupancy by SSRIs [442,443]. Using a serotonin transporter radioligand, plasma concentrations of citalopram, paroxetine, fluoxetine and sertraline were shown to correlate well with serotonin transporter occupancy. It was found that at least 80% occupancy should be attained for optimal clinical outcome [442,443]. PET studies have thus brought about highly relevant information for the determination of optimal plasma concentrations of a considerable number of psychotropic drugs which is reviewed in this special issue by Gründer and co-workers [274].

“Therapeutic window” – therapeutic reference range

TDM is based on the assumption that there is a relationship between plasma concentrations and clinical effects (therapeutic improvement, side effects and adverse effects). It also assumes that there is a plasma concentration range of the drug which is characterized by maximal effectiveness and maximal safety, the so-called “therapeutic window”. Studies on relations between plasma concentration and clinical improvement have supported this concept since the sixties for lithium, tricyclic antidepressants and classical antipsychotic drugs. Systematic reviews and meta-analyses that were based on adequately designed studies led to convincing evidence of a significant relationship between clinical outcomes and plasma concentrations for nortriptyline, imipramine and desipramine which are associated with a high probability of response [51]. For amitriptyline as a model compound, a meta-analysis of 45 studies has shown that various statistical approaches provided almost identical results [672,674]. For new antipsychotic drugs like aripiprazole [612], olanzapine [509] or risperidone [737] relationships between plasma concentration and clinical effectiveness have been reported. For the “therapeutic window” there are many synonymous terms like “therapeutic reference range”, “therapeutic range”, “optimal plasma concentration”, “effective plasma concentration”, “target range”, “target concentration”, or “orienting therapeutic range”, the term used in the first consensus [51]. The present consensus uses the term “therapeutic reference range” in accordance with the guidelines on TDM for antiepileptic drugs [499]. The “therapeutic reference range” was defined in this consensus guideline for neuropsychiatric drugs as follows:

Definition

The “**therapeutic reference ranges**” reported in this guideline (○ **Table 5**) define ranges of medication concentrations which specify a **lower limit** below which a drug induced therapeutic response is relatively unlikely to occur and an **upper limit** above which tolerability decreases or above which it is relatively unlikely that therapeutic improvement may be still enhanced. The therapeutic reference range is an orienting, population based range which may not necessarily be applicable to all patients. Individual patients may show optimal therapeutic response under a drug concentration that differs from the therapeutic reference range. Ultimately, psychopharmacotherapy can be best guided by identification of the patient’s “individual therapeutic concentration”.

The therapeutic reference ranges as recommended by the TDM group of the AGNP are given in ○ **Table 5**. They were evidence-based and derived from the literature by the structured review process described above. For only 15 neuropsychiatric drugs therapeutic reference ranges based on randomized clinical trials were found in the literature. For most drugs, reference ranges were obtained from studies with therapeutically effective doses. Therefore, there is a need for further studies to define therapeutic ranges.

The reference ranges listed in ○ **Table 5** are generally those for the primary indication. A number of drugs, however, are recommended for several indications. For example, antidepressant drugs are also used for the treatment of anxiety states, and antipsychotic drugs are increasingly used to treat mania. Little information is available on optimum plasma concentrations in these situations. Exceptions are carbamazepine, lamotrigine and

Table 5 Recommended reference ranges, laboratory alert levels and levels of recommendation for TDM.

Drugs and active metabolites	Therapeutic reference range/recommended drug concentration	t _{1/2}	Laboratory alert level	Level of recommendation to use TDM (consensus)	Conversion factor (CF, see below)	Reference	Comments
Antidepressant drugs							
Agomelatine	7–300 ng/mL 1–2 h after 50 mg	1–2 h	600 ng/mL	4	4.11	[78]	Because of rapid elimination, trough drug concentrations are not measurable under chronic treatment. Determinations, preferentially of C _{max} , should be restricted to specific indications.
Amiripryline plus nortriptyline	80–200 ng/mL	10–28 h 30 h	300 ng/mL	1	3.41 3.61	[282, 502, 672]	
Bupropion plus hydroxybupropion	225–1 500 ng/mL	8–26 h 17–47 h	2 000 ng/mL	3	4.17 3.91	[151, 152, 336, 529, 636]	Bupropion, and to a lesser degree its metabolite, are unstable, plasma or serum must be stored frozen (–20°C)
Citalopram	50–110 ng/mL	33 h	220 ng/mL	2	3.08	[42, 73, 111, 339, 388, 442, 471, 491, 549, 598]	N-Demethylated metabolites do not contribute to pharmacological actions
Clomipramine plus norclomipramine	230–450 ng/mL	16–60 h 36 h	450 ng/mL	1	3.18 3.32	[239]	
Desipramine	100–300 ng/mL	15–18 h	300 ng/mL	2	3.75	[502]	Delayed elimination in PM of CYP2D6
Desvenlafaxine	100–400 ng/mL	11 h	600 ng/mL	2	3.80	[520]	
Dosulepin = Dothiepin	45–100 ng/mL	18–21 h	200 ng/mL	2	3.39	[102, 325, 414, 541]	
Doxepin plus nordoxepin	50–150 ng/mL	15–20 h	300 ng/mL	2	3.58 3.77	[172, 321, 393, 445]	
Duloxetine	30–120 ng/mL	9–19 h	240 ng/mL	2	3.36	[21, 640, 703]	No active metabolites
Escitalopram	15–80 ng/mL	30 h	160 ng/mL	2	3.08	[409, 679]	N-Demethylated metabolites do not contribute to pharmacological actions
Fluoxetine plus norfluoxetine	120–500 ng/mL	4–6 days 4–16 days	1 000 ng/mL	2	3.23 3.39	[84, 187, 410, 442, 545]	lower level of the reference range was calculated from a PET study (80% 5HTT occupancy) [409], upper level from the SPC
Fluvoxamine	60–230 ng/mL	20 h	500 ng/mL	2	3.14	[353, 587, 631, 634, 639]	Long elimination half life of norfluoxetine (mean 14 days) and long-lasting potent inhibition of CYP2D6
Imipramine plus desipramine	175–300 ng/mL	11–25 h 15–18 h	300 ng/mL	1	3.57 3.75	[72, 229, 245, 510, 538]	Inhibition of CYP1A2, CYP2C19 Hydroxylated metabolites
Maprotiline	75–130 ng/mL	20–58 h	220 ng/mL	2	3.60	[231, 321, 384]	Active metabolite N-desmethylinaprotiline
Mianserine	15–70 ng/mL	14–33 h	140 ng/mL	3	3.78	[191, 192, 453]	
Milnacipran	50–110 ng/mL	5–8 h	220 ng/mL	2	2.24	[206, 315]	
Mirtazapine	30–80 ng/mL	20–40 h	160 ng/mL	2	3.77	[257, 367, 397, 440, 552, 591]	N-Demethylated metabolite does not contribute to pharmacological actions
Moclobemide	300–1 000 ng/mL	2–7 h	2 000 ng/mL	3	3.72	[225, 291, 327]	Metabolites are pharmacologically inactive
Nortriptyline	70–170 ng/mL	30 h	300 ng/mL	1	3.80	[30, 31, 504, 506, 510]	Hydroxylated metabolites
Paroxetine	30–120 ng/mL	12–44 h	240 ng/mL	3	3.04	[242, 243, 410, 443]	
Reboxetine	60–350 ng/mL	13–30 h	700 ng/mL	3	3.19	[483, 484]	
Sertraline	10–150 ng/mL	26 h	300 ng/mL	2	3.27	[15, 49, 258, 281, 410, 443, 545, 696]	N-Demethylated metabolite has a 2-fold longer elimination half life than sertraline, but only 1/20 of the activity of sertraline Due to irreversible inhibition of monoamine oxidase, plasma concentrations do not correlate with drug actions
Tranylcypromin	≤ 50 ng/mL	1–3 h	100 ng/mL	4	7.51	[103, 329]	
Trazodone	700–1 000 ng/mL	4–11 h	1 200 ng/mL	2	2.69	[250, 262, 268, 447, 590]	
Trimipramine	150–300 ng/mL	23 h	600 ng/mL	2	3.40	[142, 187, 223, 326]	Active metabolite N-desmethylnortriptyline
Venlafaxine plus O-desmethylvenlafaxine	100–400 ng/mL	5 h 11 h	800 ng/mL	2	3.61 3.80	[85, 241, 316, 443, 545, 550, 592, 684, 696]	In most patients O-desmethylvenlafaxine is the active principle in vivo, N-demethylated venlafaxine does not contribute to pharmacological actions. At low concentrations, the drug acts predominantly as an SSRI

Table 5 Continued.

Drugs and active metabolites	Therapeutic reference range/recommended drug concentration	t _{1/2}	Laboratory alert level	Level of recommendation to use TDM (consensus)	Conversion factor (CF, see below)	Reference	Comments
Antipsychotic drugs							
Amisulpride	100–320 ng/mL	12–20 h	640 ng/mL	1	2.71	[64, 89, 441, 461, 531, 613, 690]	No metabolites
Aripiprazole	150–500 ng/mL	60–80 h	1 000 ng/mL	2	2.23	[33, 273, 306, 368, 452, 612]	The metabolite dehydroanipiprazole is active in vitro, it remains unclear to which extend it contributes to clinical effects
Asenapine	2–5 ng/mL	24 h	10 ng/mL	4	3.50	[707]	
Benperidol	1–10 ng/mL	5 h	20 ng/mL	3	2.62	[472, 589]	Higher levels may be tolerated in patients under long-term high-dose therapy due to adaptive changes.
Bromperidol	12–15 ng/mL	20–36 h	30 ng/mL	2	4.38	[609, 656, 735]	
Chlorpromazine	30–300 ng/mL	15–30 h	600 ng/mL	2	3.14	[127, 559]	
Chlorprothixene	20–300 ng/mL	8–12 h	400 ng/mL	3	3.17	[542]	
Clozapine	350–600 ng/mL	12–16 h	1 000 ng/mL	1	3.06	[175, 507, 493, 507, 678]	Major metabolite N-desmethylclozapine with unclear antipsychotic activity
Flupenthixol	1–10 ng/mL	20–40 h	15 ng/mL	2	2.30	[40, 543, 564]	
Fluphenazine	1–10 ng/mL	16 h	15 ng/mL	1	2.29	[564, 680]	
Fluspirilen	0.1–2.2 ng/mL	7–14 days	4.4 ng/mL	2	2.10	[611]	
Haloperidol	1–10 ng/mL	12–36 h	15 ng/mL	1	2.66	[74, 214, 480, 494, 508, 674, 680]	Higher levels can be tolerated in patients under long-term high-dose therapy due to adaptive changes.
Iloperidone	5–10 ng/mL	18–33 h	20 ng/mL	3	2.34	[476, 576]	
Levomepromazine	30–160 ng/mL	16–78 h	320 ng/mL	3	3.04	[656]	
Melperone	30–100 ng/mL	4–6 h	200 ng/mL	3	3.80	[83, 324]	Inhibitor of CYP2D6
Olanzapine	20–80 ng/mL	30–60 h	150 ng/mL	1	3.20	[32, 56, 63, 132, 208, 240, 418, 478, 509, 602, 711]	Under olanzapine pamoate, patients exhibited a post injection syndrome when drug concentrations exceeded 150 ng/mL
Paliperidone	20–60 ng/mL	23 h	120 ng/mL	2	2.35	[26, 70, 131, 466]	Paliperidone=9-hydroxyrisperidone
Perazine	100–230 ng/mL	8–16 h	460 ng/mL	1	2.95	[91]	
Perphenazine	0.6–2.4 ng/mL	8–12 h	5 ng/mL	1	2.48	[564, 637, 680]	
Pimozide	15–20 ng/mL	23–43 h	20 ng/mL	3	2.17	[649]	
Pipamperone	100–400 ng/mL	17–22 h	500 ng/mL	3	2.66	[82, 517]	
Prothipendyl	5–10 ng/mL	2–3 h	20 ng/mL	4	3.35	[436] SPC	
Quetiapine	100–500 ng/mL	7 h	1 000 ng/mL	2	2.61	[112, 212, 236, 299, 498, 603, 627, 689, 723]	When the patient has taken the extended release (XR) formulation in the evening and blood was withdrawn in the morning, expected plasma concentrations are 2-fold higher than trough levels
Risperidone	20–60 ng/mL	3 h	120 ng/mL	2	2.44	[150, 406, 426, 437, 469, 475, 553, 557, 617, 729, 737]	
Sertindole	50–100 ng/mL	55–90 h	200 ng/mL	2	2.27	[71, 109, 110, 653, 728, 729]	Active metabolite dehydrosertindole (concentration at therapeutic doses 40–60 ng/mL), concentration dependent increase of QT interval by blockade of potassium channels
Sulpiride	200–1 000 ng/mL	8–14 h	1 000 ng/mL	2	2.93	[460, 656]	No metabolites, renal elimination
Thioridazine	100–200 ng/mL	30 h	400 ng/mL	1	2.70	[190, 656]	Contraindicated in poor metabolizers of CYP2D6
Ziprasidone	50–200 ng/mL	6 h	400 ng/mL	2	2.55	[126, 419, 427, 688, 695]	The drug should be taken with a meal, otherwise absorption is reduced and plasma concentrations will be lower than expected
Zotepine	10–150 ng/mL	13–16 h	300 ng/mL	3	3.01	[376, 642]	
Zuclopentixol	4–50 ng/mL	15–25 h	100 ng/mL	3	2.49	[330, 371, 692]	

Table 5 Continued.

Drugs and active metabolites	Therapeutic reference range/recommended drug concentration	t _{1/2}	Laboratory alert level	Level of recommendation to use TDM (consensus)	Conversion factor (CF, see below)	Reference	Comments
Mood stabilizing drugs							
Carbamazepine	4–10 µg/mL	10–20 h	20 µg/mL	2	4.23	[512]	Active 10,11-epoxide metabolite contributes to clinical effects
Lamotrigine	3–14 µg/mL	7–23 h	30 µg/mL	2	3.90	[455, 558]	So far no specific reference range for mood stabilizing effect, valproate increases elimination half life to 48–70 h
Lithium	0.5–1.2 mmol/l (4–8 µg/mL)	24 h	1.2 mmol/l (8 µg/mL)	1	1.25.8	[593, 721]	Age dependent increase of elimination half life
Valproic acid	50–100 µg/mL	18 h	120 µg/mL	2	6.93	[16, 216, 301, 683]	In individual cases 120 µg/mL are also tolerated in acute mania.
Anticonvulsant drugs							
Carbamazepine	4–12 µg/mL	10–20 h	20 µg/mL	2	4.25	[87, 338, 499]	Active 10,11-epoxide metabolite contributes to clinical effects
Clobazam and N-desmethylclobazam	30–300 ng/mL 300–3 000 ng/mL	18–42 h	500 ng/mL 5 000 ng/mL	2	3.33 3.49	[278, 499]	Active N-demethylated metabolite contributes to clinical effects
Clonazepam	20–70 ng/mL	40 h	80 ng/mL	2	3.17	[44, 464, 499]	7-Amino metabolite retains some activity
Ethosuximide	40–100 µg/mL	33–55 h	120 µg/mL	2	7.08	[88, 499]	
Felbamate	30–60 µg/mL	15–23 h	100 µg/mL	2	4.20	[290, 343, 499]	
Gabapentin	2–20 µg/mL	6 h	25 µg/mL	3	5.84	[75–77, 343, 398, 499]	
Lacosamide	1–10 µg/mL	13 h	20 µg/mL	3	2.66	[47]	
Lamotrigine	3–14 µg/mL	7–23 h	20 µg/mL	2	3.90	[88, 343, 455, 456, 499, 610]	Valproate increases elimination half life to 48–70 h
Levetiracetam	10–40 µg/mL	6–8 h	100 µg/mL (morning levels)	2	3.87	[88, 343, 430, 499]	
Methsuximide plus methsuximide	10–40 µg/mL	1–3 h 36–45 h	45 µg/mL	2	4.92 and 5.29	[88]	The metabolite is the active principle in vivo
Oxcarbazepine plus 10-hydroxycarbazepine	10–35 µg/mL	5 h 10–20 h	40 µg/mL	2	3.96 and 3.73	[88, 343, 428, 499]	
Phenobarbital	10–40 µg/mL	80–120 h	50 µg/mL	1	4.31	[88, 499]	
Phenytoin	10–20 µg/mL	20–60 h	25 µg/mL	1	3.96	[88, 380, 499]	
Pregabalin	2–5 µg/mL	6 h	10 µg/mL	3	6.28	[68, 77, 88, 343, 432, 499]	
Primidone (active metabolite phenobarbital)	5–10 µg/mL	14–15 h	25 µg/mL	2	4.58	[88, 499]	Data given are restricted to primidone, for the active metabolite phenobarbital recommended plasma concentrations are 10–40 µg/mL
Rufinamid	5–30 µg/mL	7 h	40 µg/mL	2	4.20	[511]	
Stiripentol	1–10 µg/mL	4–13 h	15 µg/mL	2	4.27	[503]	
Sulfthiame	2–8 µg/mL	3–30 h	12 µg/mL	2	3.46	[88, 375, 429]	
Tiagabine	20–200 ng/mL	7–9 h	300 ng/mL	2	2.66	[88, 235, 343, 499]	
Topiramate	2–8 µg/mL (morning levels)	21 h	16 µg/mL	3	2.95	[88, 226, 343, 431, 499]	
Valproic acid	50–100 µg/mL	18 h	120 µg/mL	2	6.93	[16, 88, 216, 301, 499, 682, 683]	
Vigabatrin	2–10 µg/mL	5–8 h	20 µg/mL	4	7.74	[88, 342, 398, 499, 719]	
Zonisamide	10–40 µg/mL	60 h	40 µg/mL	2	4.71	[247, 448, 449]	
Anxiolytic/hypnotic drugs							
Alprazolam	5–50 ng/mL	12–15 h	100 ng/mL [§]	4	3.22	[585, 686]	In chronic users of benzodiazepines, effective plasma concentrations can be markedly higher than in non users.
Bromazepam	50–200 ng/mL	15–35 h	300 ng/mL [§]	4	3.16	[218, 286, 586]	
Brotizolam	4–10 ng/mL (Cmax)	3–6 h	20 ng/mL	4	2.53	[341, 669]	
Buspirone (active metabolite 6-hydroxybuspirone)	1–4 ng/mL	2–3 h	8 ng/mL [§]	3	2.59 2.49	[178, 580, 586]	

Table 5 Continued.

Drugs and active metabolites	Therapeutic reference range/recommended drug concentration	t _{1/2}	Laboratory alert level	Level of recommendation to use TDM (consensus)	Conversion factor (CF, see below)	Reference	Comments
Chlordiazepoxide	400–3 000 ng/mL	5–30 h	3 500 ng/mL	4	3.48	[408, 586]	
Clonazepam	4–80 ng/mL	19–30 h	100 ng/mL	4	3.17	[181, 467, 586]	
Diazepam and metabolites	200–2 500 ng/mL	24–48 h	3 000 ng/mL	4	3.51	[224, 261, 264, 586]	Active metabolites are nordazepam, oxazepam and temazepam
Flunitrazepam	5–15 ng/mL	10–30 h	50 ng/mL	4	3.20	[80, 425]	
Lorazepam	10–15 ng/mL	12–16 h	30 ng/mL	4	3.20	[164, 196, 218, 267]	
Lormetazepam	2–10 ng/mL	8–14 h	100 ng/mL	4	2.98	[3, 515]	
Midazolam	6–15 ng/mL Cmax: 60–80 ng/mL	1–3 h	1 000 ng/mL	4	3.06	[35, 261, 323]	
Nitrazepam	30–100 ng/mL	18–30 h	200 ng/mL	4	3.56	[467, 586]	
Nordazepam	20–800 ng/mL	50–90 h	1 500 ng/mL	4	3.69	[586]	
Opipramol	50–500 ng/mL	11 h	1 000 ng/mL	3	2.87	[386]	
Oxazepam	200–1 500 ng/mL	4–15 h	2 000 ng/mL	4	3.49	[586]	
Pregabalin	2–5 µg/mL	6 h	10 µg/mL	3	6.28	[76, 77]	
Temazepam	20–900 ng/mL	5–13 h	1 000 ng/mL	4	3.51	[586]	
Triazolam	2–20 ng/mL	1–5 h	40 ng/mL [§]	4	4.12	[586]	
Zolpidem	80–150 ng/mL	1–4 h	300 ng/mL	4	3.23	[586]	
Zopiclone	10–50 ng/mL	5 h	150 ng/mL	4	3.48	[586]	Unstable at room temperature
Antidementia Drugs							
Donepezil	30–75 ng/mL	70–80 h	75 ng/mL	2	2.64	[492, 563, 652]	
Galantamine	30–60 ng/mL	8 h	90 ng/mL	3	3.48	[322, 333, 734]	
Memantine	90–150 ng/mL	60–100 h	300 ng/mL	3	5.58	[251, 378]	
Rivastigmine	oral 8–20 ng/mL (1–2 h after dose) Patch 5–13 ng/mL (1 h before application of a new patch)	1–2 h	40 ng/mL	3	4.00	[597] 147, 391]	
Drugs for treatment of substance related disorders							
Acamprosate	250–700 ng/mL	13 h	1 000 ng/mL	3	8.68	[287, 288, 424]	
Buprenorphine	0.7–1.6 ng/mL Cmax: < 9 ng/mL after 24 mg	2–5 h	10 ng/mL (Cmax)	2	2.38	[120, 130, 383]	
Bupropion plus Hydroxybupropion	550–1 500 ng/mL	20 h 20 h	2 000 ng/mL	2	4.17 3.91	[345]	Bupropion is unstable, plasma or serum must be stored frozen (–20°C) after blood withdrawal In a clinical trial 300 mg was the most effective dose with resulting plasma concentrations as indicated
Clomethiazol	100–5 000 ng/mL	2–5 h	500 ng/mL	4	6.19	[672]	In alcohol dependent patients much higher plasma concentrations may be tolerated than in healthy subjects
Disulfiram	50–400 ng/mL	7 h	500 ng/mL	3	3.37	[203, 344, 586]	Disulfiram (DSF) is a prodrug, its active metabolite diethylthio-methyl-carbamate (DDTC-Me) has been suggested as a possible marker for proper dose titration of disulfiram [344]. In a pharmacokinetic study under 300 DSF mean±SD steady state concentrations of DSF amounted to 170±10 ng/mL those of DDTC-Me to 290±20 ng/mL.
Levomethadone	250–400 ng/mL	14–55 h	400 ng/mL 100 ng/mL [§]	2	3.23	[146]	[§] In non users of opiates, effective or toxic plasma concentrations are markedly lower than in users. Chronic users may even need "toxic" concentrations in blood to avoid the occurrence of withdrawal symptoms.
Methadone	400–600 ng/mL	24–48 h	600 ng/mL 300 ng/mL [§]	2	3.23	[146, 188, 595]	

Table 5 Continued.

Drugs and active metabolites	Therapeutic reference range/recommended drug concentration	t _{1/2}	Laboratory alert level	Level of recommendation to use TDM (consensus)	Conversion factor (Cf, see below)	Reference	Comments
Naltrexone plus 6β-naltrexol	25–100 ng/mL	4h 13h 24h	200 ng/mL	2	3.06 3.04 4.73	[99, 211, 252, 424]	
Varenicline	4–5 ng/mL		10 ng/mL	3	4.73	[202, 532]	
Antiparkinson drugs							
Amantadine	0.3–0.6 µg/mL	10–14h	1.2 µg/mL	3	5.98	[320]	
Biperiden	Cmax. 1–6.5 ng/mL 0.5–2 h after 4 mg	18–24h	13 ng/mL	3	3.21	[270]	
Bornaprine	Cmax. 0.7–7.2 ng/mL 1–2h after 4mg	30h	14 ng/mL	3	3.04	[433]	
Bromocriptine	Low dose (2.5mg): 0.1–0.3 ng/mL Max. dose (25 mg): 1.0–4.0 ng/mL	38h	8 ng/mL	3	1.53	[168]	
Cabergoline	Cmax. 58–144 pg/mL at 0.5–4h after drug intake for 4 weeks	63–68h	390 pg/mL	3	2.21	[168]	Unstable at room temperature, plasma or serum should be stored frozen (< -20 °C)
Carbidopa	Cmax. 20–200 ng/mL after 2 h	2h	400 ng/mL	3	4.42	[574]	Unstable at room temperature, plasma or serum should be stored frozen (< -20 °C)
Levodopa O-Methyldopa	Cmax. 0.9–2.0 µg/mL 0.6–0.9 h after 250 mg combined with 25 mg carbidopa 0.7–10.9 µg/mL	1–3h	5 µg/mL	3	5.07	[4, 135, 394, 479, 574]	Unstable at room temperature, plasma or serum should be stored frozen (< -20 °C) Elimination half-life and plasma concentrations increases under comedication with carbidopa or benserazide
Entacapone	Cmax. 0.4–1.0 µg/mL	0.5h	2 µg/mL	3	3.28	[304, 570]	Unstable at room temperature, plasma or serum should be stored frozen (< -20 °C)
Pramipexole	0.39–7.17 ng/mL	8–12h	15 ng/mL	3	4.73	[730]	
Ropinireole	0.4–6.0 ng/mL	3–10h	12 ng/mL	3	3.84	[657]	
Tiaprude	Cmax. 1–2 µg/mL	3–4h	4 µg/mL	3	3.05	[108]	
Tolcapone	Cmax. 3–6 µg/mL	2h	12 µg/mL	3	3.66	[177, 346]	
Other Drugs							
Atomoxetine	200–1000 ng/mL 60–90 min after intake of 1.2 mg/kg/day	4h	2000 ng/mL	3	3.91	[233, 302, 446, 583]	Recommended reference ranges indicate Cmax measured in remitters. Elimination half-life is 21 h in PM of CYP2D6
Dexmethylphenidate	13–23 ng/mL 4h after 20 mg	2	44	2	4.29	[663]	5.2–5.5 ng/mL are associated with 50% dopamine transporter blockade [614]
Methylphenidate	13–22 ng/mL d-methylphenidate 2 h after 20 mg immediate release or 6–8h after 40 mg extended release	2h	44 ng/mL	2	4.29	[331, 422, 614, 615]	Methylphenidate is unstable at room temperature, recommended reference range indicates Cmax
Modafinil	1000–1700 ng/mL after 200 mg/day	10–12h	3400 ng/mL	3	4.21	[733]	

Plasma concentrations given in mass units can be converted to molar units by multiplication with the conversion factor (Cf) nmol/L = ng/mL x Cf
% Active metabolite contributes to wanted and unwanted effects. Indicated reference ranges and laboratory alert levels refer to the mother compound only.
For bupropion, carbamazepine, lamotrigine and valproic acid recommended reference ranges were listed twice in accordance with the 2 different indications.
Prepared by CH, PB, SU, BR and HK, reviewed by AC, OD, KE, MF, MG, CG, GG, EH, UHR, CH, EJS, HK, UL, TM, BP, BS, MU, SU, GZ

valproic acid, which are therefore listed twice in **Table 5**. Moreover, it should be mentioned that studies are on the way to evaluate therapeutic reference ranges for children or adolescent patients and for elderly patients.

Estimation of the lower limit of the therapeutic reference range

Estimation of a therapeutic reference range (TRR) requires estimation of a lower and an upper limit of drug concentration in plasma. A generally accepted method for calculation of these limits does not exist. Whenever possible the lower limit of a drug's therapeutic range should be based on studies on the relationship between a drug's plasma concentration and clinical effectiveness. Below this limit, therapeutic effects are not significantly different from placebo. The optimum study design for evaluation of the lower limit of the therapeutic range is a prospective double-blind study where patients are treated with drug doses which lead to a defined plasma concentration range of the drug. Such a design was applied by Van der Zwaag and co-workers for patients treated with clozapine [678]. Patients were titrated to 3 different plasma concentrations of the antipsychotic drug. Significant superiority was found in patients with middle and high plasma concentration compared with low concentrations of clozapine. A similar design was applied for a blood-level study comparing imipramine and mirtazapine [98]. To conduct such studies, however, is a considerable logistic challenge. Fixed dose studies are therefore preferred for evaluation of the lower limit of the therapeutic reference range [672,674].

For the estimation of threshold values of the therapeutic reference range, receiver operating characteristic (ROC) analysis has proven helpful [289]. A ROC plot allows the identification of a cut-off value that separates responders from non-responders and estimates the sensitivity and specificity of the parameter "medication plasma concentration". The usefulness of the ROC analysis has been demonstrated for a number of antipsychotic and antidepressant drugs [461,505,510,703].

Estimation of the upper limit of the therapeutic reference range

In the first study on TDM in psychiatry [31] an U-shaped relationship between plasma concentration and clinical effect was reported for nortriptyline. The lack of effect at high concentrations was attributed to the mechanism of action of the tricyclic antidepressant drug on monoaminergic neurons. According to actual knowledge, however, it seems more likely that reduced amelioration at high concentrations is due to side effects. The upper limit of the therapeutic range is therefore defined by the occurrence of side effects, also in this guideline. For most side effects (type A adverse reactions), it is also assumed that they are a function of dose and drug concentration in the body [335]. This assumption has been confirmed for motor side effects of antipsychotic drugs [536] and for unwanted side effects of tricyclic antidepressant drugs [153,282]. For paroxetine, a positive correlation was found between drug concentration in plasma and serotonin syndrome symptoms [303]. When such data are available, it is possible to apply ROC analysis for the calculation of the upper limit of the therapeutic range [461]. For many psychotropic drugs listed in **Table 5**, however, valid data on both plasma concentration and the incidence of side effects are lacking. Case reports on tolerability problems or intoxications do often not include drug concentration measurements in plasma. Sporadic reports on fatal cases and intoxications are of limited

value. Most blood concentrations reported to have caused death are far above drug concentrations that are associated with maximum therapeutic effects [544,622]. Post mortem redistribution of medications from or into the blood can lead to dramatic changes in blood levels [382,518], and the direction of the change does not follow a general rule [359]. Estimation of an upper threshold level above which tolerability decreases or the risk of intoxication increases is therefore more difficult than estimation of the lower threshold level, especially for drugs with a broad therapeutic index like SSRIs.

Estimation and definition of a laboratory alert level

As explained above, plasma concentrations with an increased risk of toxicity are normally much higher than the upper threshold levels of the therapeutic reference ranges for most psychotropic drugs shown in **Table 5**. For the present guidelines, we therefore defined an upper plasma concentration limit above which it seems unlikely that therapeutic effects may be enhanced and added a "laboratory alert level" which was defined as follows:

Definition

The "laboratory alert levels" reported in this guideline (**Table 5**) indicate drug concentrations above the recommended reference range that causes the laboratory to feedback immediately to the prescribing physician. The alert levels are based on reports on intolerance or intoxications and plasma concentration measurements. In most cases, however, it was arbitrarily defined as a plasma concentration that is 2-fold higher than the upper limit of the therapeutic reference range. The laboratory alert should lead to dose reduction when the patient exhibits signs of intolerance or toxicity. When the high drug concentration is well tolerated by the patient and if dose reduction bears the risk of symptom exacerbation, the dose should remain unchanged. The clinical decision, especially in case of unchanged dose needs to be documented in the medical file.

From population-based to subject-based reference values

All therapeutic reference ranges listed in **Table 5** are orienting, population-based ranges. The population-derived ranges constitute descriptive statistical values which may not necessarily be applicable to all patients. Individual patients may show the optimum therapeutic response under a drug concentration that differs from the therapeutic reference range. Psychopharmacotherapy should therefore try to identify a patient's "individual therapeutic concentration" to guide the treatment [61,523]. For lithium it has been shown that the recommended plasma concentration range depends on whether the patient is in an acute manic episode or needs maintenance therapy [593]. For clozapine, Gaertner and colleagues [232] determined optimal plasma concentrations required for stable remission of individual patients under maintenance therapy in a relapse prevention study.

Recommendations for measuring plasma concentrations of psychoactive drugs

The usefulness of TDM varies with the clinical situation and the particular drug involved. In case of suspected non-adherence to medication or intoxications, quantifying plasma concentrations is a generally accepted tool for all drugs and groups of patients. However, it is still a matter of debate if TDM should be imple-

mented in clinical routine. Based on empirical evidence, 5 levels of recommendation to use TDM were defined in the guidelines 2004 for 65 psychotropic drugs. These definitions were revised and grading reduced to 4 levels of recommendation, now ranging from “strongly recommended” to “potentially useful” as follows:

Definitions

Level 1: Strongly recommended

Evidence: Reported drug concentrations are established and evaluated therapeutic reference ranges. Controlled clinical trials have shown beneficial effects of TDM, reports on decreased tolerability or intoxications.

Recommendation: TDM is strongly recommended for dose titration and for special indications. For lithium, TDM is a standard of care.

Clinical consequences: At therapeutic plasma concentrations highest probability of response or remission; at “subtherapeutic” plasma concentrations: response rate similar to placebo under acute treatment and risk of relapse under chronic treatment; at “supratherapeutic” plasma concentrations: risk of intolerance or intoxication.

Level 2: Recommended

Evidence: Reported drug concentrations were obtained from plasma concentrations at therapeutically effective doses and related to clinical effects; reports on decreased tolerability or intoxications at “supratherapeutic” plasma concentrations.

Recommendation: TDM is recommended for dose titration and for special indications or problem solving.

Clinical consequences: TDM will increase the probability of response in non-responders. At “subtherapeutic” plasma concentrations: risk of poor response; at “supratherapeutic” plasma concentrations: risk of intolerance or intoxication.

Level 3: Useful

Evidence: Reported drug concentrations were calculated from plasma concentrations at effective doses obtained from pharmacokinetic studies. Plasma concentrations related to pharmacodynamic effects are either not yet available or based on retrospective analysis of TDM data, single case reports or non-systematic clinical experience.

Recommendation: TDM is useful for special indications or problem solving.

Clinical consequences: TDM can be used to control whether plasma concentrations are plausible for a given dose, or clinical improvement may be attained by dose increase in non-responders who display too low plasma concentrations.

Level 4: Potentially useful

Evidence: Plasma concentrations do not correlate with clinical effects due to unique pharmacology of the drug, e.g., irreversible blockade of an enzyme, or dosing can be easily guided by clinical symptoms, e.g., sleep induction by a hypnotic drug.

Recommendation: TDM is not recommended for dose titration but may be potentially useful for special indications or problem solving.

Clinical consequences: TDM should be restricted to special indications.

According to our literature-based evaluations, TDM was graded as “strongly recommended” for 15 of the 128 surveyed neuropsychiatric compounds, “recommended” for 52 medications, “useful” for 44 drugs and “potentially useful” for 19 drugs (Table 5).

TDM is highly recommended for most tricyclic **antidepressants**. It reduces the risk of intoxications [103, 381, 459, 510, 525, 527, 528, 530, 718], and for many tricyclic antidepressants, a plasma concentration – clinical effectiveness relationship has been shown. For SSRIs, TDM is of little clinical importance in clinical practice [6, 537, 644]. Toxicity of this type of antidepressants is low in comparison to most of the pre-SSRI antidepressants [48, 166, 314, 646, 715]. Data from Sweden revealed that TDM of SSRIs is cost-effective in elderly patients where it helped to use minimum effective doses [410]. For citalopram a recent observational study revealed that plasma concentrations on day 7 of treatment are predictive for later non-response [491]. Patients exhibiting citalopram plasma concentrations below 50 ng/mL had a significantly reduced improvement on the Hamilton rating scale for depression. Evidence for a statistically significant relationship between drug concentration and therapeutic outcome is lacking for the tetracyclic antidepressants maprotiline, mianserin and mirtazapine and also for trazodone and reboxetine, the monoamine oxidase inhibitors moclobemide and tranlycypromine.

TDM is strongly recommended for the **typical antipsychotic drugs** haloperidol, perphenazine and fluphenazine, and for the atypical antipsychotics amisulpride, clozapine, olanzapine, and risperidone (Table 5). Overdosing may lead to extrapyramidal side effects. In the case of clozapine, there is a strong correlation between clozapine plasma levels and incidence of seizures. Avoiding overdosing of typical antipsychotic drugs by TDM is for the majority of patients a matter of quality of life rather than safety [136]. TDM of antipsychotics is also useful when medication is switched from the oral to the depot form, or vice versa.

With regard to the **mood stabilizing** and/or **antimanic drugs** lithium, valproic acid and carbamazepine, therapeutic reference ranges and toxic levels are well defined. Therefore TDM is strongly recommended for these drugs (Table 5). For lithium TDM is even the standard of care [133, 170, 185, 280, 395, 593, 706, 721]. For its long-term use, plasma concentrations of 0.5–0.8 nmol/L are advised. For an acute treatment with lithium, it may be justified to increase its concentrations up to 1.2 mmol/L. Compounds that have been shown to be effective as **antidementia drugs** are donepezil, rivastigmine, galantamine and memantine. TDM is rarely used for the treatment of dementia, though there is evidence that it can be useful. For donepezil, it has been shown that the patients' improvement was significantly better if their plasma concentrations were above 50 ng/mL as compared to patients that showed lower drug concentrations [563].

Most **anxiolytic** and **hypnotic drugs** belong to the class of benzodiazepines. Anxiolytic and hypnotic effects are rapid. Treatment can therefore be guided by immediate clinical impression rather than by TDM. In case of lack of therapeutic effects under usual doses, however, TDM may clarify if non-response was due to drug abuse that has led to tolerance or due to pharmacokinetic abnormalities. For alprazolam, TDM may be useful to suppress panic attacks [722].

The **opiate agonists** methadone, R-methadone (levomethadone), buprenorphine, 1- α -acetylmethadol (LAAM) and slow-release formulations of morphine are used for the treatment of opioid addiction. TDM is indicated for patients treated with methadone or R-methadone. The usefulness of TDM for monitoring treatment with “anti-craving” medications such as acamprosate or naltrexone, employed for the treatment of alcohol use disorders, has recently been reviewed elsewhere [99]. TDM was recommended to enhance the moderate efficacy of these drugs.

For **anticonvulsant drugs**, TDM is well established, especially for old drugs which are more toxic than the new ones [499]. For **antiparkinson drugs**, TDM has not been established so far. For the dopamine agonists, data on reference ranges are scarce. For L-dopa, there is an imperfect correlation between plasma concentrations and short-term clinical response [479]. Nevertheless, we considered the pharmacologic properties of these neurological drugs (◉ **Table 1, 5**), since psychiatric patients may receive antiparkinson drugs that possibly interfere with the action of psychotropic medication. For most of these drugs C_{max} values are given.

Indications for measuring plasma concentrations of psychoactive drugs

◉ **Table 6** presents a list of indications for TDM in psychiatry. The validity of these indications has to be examined on an individual basis and evaluated for each case individually. Similar to any diagnostic test, TDM should only be requested when there is evidence that the result will provide an answer to a well defined question.

For drugs with well defined therapeutic reference ranges or with a narrow therapeutic index it makes sense to measure plasma levels for dose titration after initial prescription or after dose change. Even without a specific problem, there is sufficient evidence that TDM has beneficial effects for patients treated with these drugs. This holds true for lithium, tricyclic antidepressants, several antipsychotics or anticonvulsants (◉ **Table 5**). For lithium, TDM is even mandatory for safety reasons.

In case of **suspected non-adherence** or lack of clinical improvement under recommended doses: TDM is a valid tool for treatment with all drugs considered in these guidelines. Loss of adherence is a major problem of long-term medication [10,55,401]. In patients with schizophrenia [55,351] and in patients with unipolar or bipolar disorders non-adherence ranges from 10 to 69% [401,439]. Methods used to measure adherence include pill counting, examining case-note recordings, interviewing patients or noting the attending physicians' clinical judgement about adherence [11,355,685,708]. Studies have shown that clinicians cannot reliably predict their patients' adherence [104,579]. TDM is advantageous, since it is an objective method and tells the prescribing physician if the drug is in the body at a concentration that is potentially sufficient for the expected clinical response. Deviations from the expected dose-related reference range (◉ **Table 4**) indicate if the patient has taken his/her medication, and concomitant determination of metabolites is another approach to clarify if the drug was taken continuously as recommended. For interpretation, however, possible interactions with co-medications exhibiting enzyme inhibiting or inducing properties must be considered (◉ **Table 3**). Reis and coworkers [546,547] analysed the compliance of patients who were treated with sertraline by repeated determination of serum drug concentrations of the parent compound and of the metabolite. Variations of the ratios of concentrations of norsesertraline to sertraline were highly indicative for hidden and partial non-adherence. To be able to use this approach, these guidelines were supplemented with data on ratios of concentrations for 32 psychoactive drugs (◉ **Table 2**). By taking several blood samples per day and by calculation the observed and expected time dependent plasma concentrations it can be differentiated if a low plasma concentration is due to reduced bioavailability, enhanced degradation or poor adherence. Pharmacokinetic modelling of the expected time dependent

plasma concentration thereby considers a drug's basic pharmacokinetic properties [4, 78, 340, 626, 654].

When **clinical improvement** under recommended doses is **insufficient** and the drug is well tolerated, TDM will clarify if the drug concentration is too low and if it makes sense to increase the dose.

When **adverse effects** are associated with clinical improvement under recommended doses, measurement of the plasma concentration may clarify if side effects are related to excessively high drug levels in the blood and if the dose should be decreased. When **combining medications** that are inhibitors or inducers of drug metabolizing enzymes (◉ **Table 1**), pharmacokinetic drug interactions will occur if the comedication is a substrate of the inhibited or induced enzyme (◉ **Table 3**). Dose adaptation should be guided by TDM in combination with an inducer or inhibitor and avoid loss of action, poor tolerability or intoxication due to a pharmacokinetic drug-drug interaction [215, 244, 594]. With regard to environmental factors smoking is of high clinical relevance for drugs that are substrates of CYP1A2 (◉ **Table 1**). The isoenzyme is dose dependently induced by constituents of cigarette smoke (polycyclic aromatic hydrocarbons, not nicotine). Its activity increases by 1.2-fold, 1.5-fold for 1.7-fold for 1–5, 6–10 and > 10 cigarettes smoked per day [201]. On the other hand, CYP1A2 activity decreases until the fourth day immediately on cessation of heavy smoking [200]. Smoking effects should therefore be considered when patients are under therapy with a CYP1A2 substrate (◉ **Table 1**) such as clozapine [81,676], duloxetine [222] or olanzapine [749]. It should also be mentioned that many pharmacokinetic drug-drug interactions have been found by TDM either by chance or by retrospective analysis of TDM data bases [112,537].

In **pharmacovigilance programs**, the safety of drug use is supervised under naturalistic conditions [271,285]. In case of observed adverse events, measurement of plasma concentrations is most helpful for clarification [335].

Relapse prevention is a major goal of maintenance treatment. Reduction of relapse rates by TDM is highly cost-effective, as relapses can lead to hospitalization [377]. In schizophrenic patients, it has been shown that fluctuations of clozapine plasma concentrations are predictive for relapses [232,670] and rehospitalizations [627]. In these patients, TDM may help reduce the risk of relapse or recurrence by increasing adherence to the medication. One day in the hospital is 4–16 times more expensive than a single drug concentration measurement in the laboratory.

Recommendation

Though clinical evidence is still scarce, we recommend regular monitoring of plasma concentrations under maintenance therapy, at least every 3–6 months, to prevent relapses and rehospitalizations. The frequency of TDM requests may be increased if patients are known to be non-adherent to the medication or in case of changes of co-medications or of smoking that affect the pharmacokinetics of the drug.

In patients exhibiting **genetic peculiarities** of drug metabolizing enzymes, doses must be adapted. Kirchheiner and coworkers [362,365] calculated doses for PM or UM of CYP2D6 based on pharmacokinetic and pharmacodynamic findings. However, even in the case of a confirmed abnormal CYP genotype, TDM is recommended, because genotyping can only roughly predict to

which extent the plasma concentration may be changed in the individual patient [496,497,625].

For **special groups of patients**, such as pregnant or breastfeeding patients, children or adolescent patients [22,373,194], individuals with intellectual disabilities [158,300], or elderly patients, especially patients aged above 75 years [374], TDM is highly recommended.

Any psychopharmacologic therapy of pregnant or breastfeeding women should assure that the plasma concentration of the drug is in the therapeutic reference range to minimize the risk of relapse on the mother's side and, at the same time, to minimize risks associated with drug exposure of the fetus or the child [169,174]. Renal clearance and the activity of the CYP isoenzymes 3A4, 2D6 and 2C9, and uridine 5'-diphosphate glucuronosyltransferase are increased during pregnancy, whereas activities of CYP1A2 and 2C19 decrease [21]. TDM in pregnant women and/or mothers should be carried out at least once per trimester and within 24 h after delivery [65].

Many psychoactive drugs are not approved for use in children or adolescents [248]. Pharmacokinetics and pharmacodynamics change during development [194,438,514,516]. In adolescents suffering from psychotic disorders, comorbid drug abuse is very common, and compliance with an antipsychotic treatment is generally marginal [318]. Therefore, TDM is recommended in these patients. To raise data on the effectiveness and tolerability of psychoactive drugs under every day conditions, a TDM network was established for child and adolescent patients [see <http://www.tdm-kjp.de/eng/contact.html>].

In **elderly patients**, who frequently are hypersensitive to medication, TDM is helpful to distinguish between pharmacokinetic and pharmacodynamic factors when adverse effects occur [666]. Ageing involves progressive impairments of the functional reserve of multiple organs [407], especially renal excretion, and body composition changes significantly [361,374]. Hepatic clearance can be reduced by up to 30%. Phase I reactions are more likely to be impaired than phase II reactions. On the other hand, there are no age-dependent changes in CYP isoenzyme activity [374]. Age-related changes in physiologic and pharmacokinetic functions as well as the comorbidity and polypharmacy complicate pharmacotherapy in the elderly [125]. Therefore, TDM should be used for these patients to improve safety and tolerability of psychopharmacotherapy.

In **individuals with intellectual disabilities**, new generation antipsychotic drugs are frequently used. Recently published guidelines recommend TDM for these patients, at least when treated with risperidone or olanzapine [158]. For ethical and legal reasons, patients with intellectual disabilities are excluded from clinical trials. On the other hand, many of these patients need medication. In these individuals, it may be difficult to differentiate between moribogenic and pharmacogenic reasons for symptom aggravation. Though evidence is poor, TDM is recommended to guide the pharmacotherapy of these patients [158].

In **forensic psychiatry** the primary aim of pharmacotherapy, consisting mostly antipsychotic drugs, is reduction of dangerous behaviour [458,462]. To consistently reduce the risk of violence and aggression, adherence to the prescribed medication is essential [658]. Therefore, TDM is recommended for this group of psychiatric patients. It is, however, not clear if effective plasma concentrations are identical in forensic and general psychiatry patients. Castberg and Spigset [113] analyzed data obtained by survey in a high security forensic unit and found higher doses in forensic patients than in a control group. The dose related

Table 6 Typical indications for measuring plasma concentrations of medications in psychiatry.

- Dose optimization after initial prescription or after dose change
- Drugs, for which TDM is mandatory for safety reasons (e. g., lithium)
- Suspected complete or partial non-adherence (non-compliance) to medication
- Lack of clinical improvement under recommended doses
- Adverse effects and clinical improvement under recommended doses
- Combination treatment with a drug known for its interaction potential or suspected drug interaction
- TDM in pharmacovigilance programs
- Relapse prevention under maintenance treatment
- Recurrence under adequate doses
- Presence of a genetic particularity concerning drug metabolism (genetic deficiency, gene multiplication)
- Pregnant or breast feeding patient
- Children and adolescent patient
- Elderly patient (>65 y)
- Individuals with intellectual disabilities
- Patients with pharmacokinetically relevant comorbidities (hepatic or renal insufficiency, cardiovascular disease)
- Forensic patient
- Problems occurring after switching from an original preparation to a generic form (and vice versa)

plasma concentrations were significantly lower for olanzapine but higher for quetiapine in the forensic patients than in the control group.

The indication **"problem occurring after switching from an original preparation to a generic form (and vice versa)"** is still under-investigated and data are scarce [124,139].

Another potential indication for TDM not listed in **Table 6** is the increasing availability of counterfeit drugs on the internet [599]. WHO launched a program in 2006 to combat this illegal industry. There are no data published on this type of market concerning psychotropic drugs, but patients may be co-medicated (mostly auto-medication) with other drugs obtained from this source. The counterfeit medications may not comply with purity and dosage standards and therefore increase the risk for interactions.

Practical Aspects for TDM in Psychiatry



Essential for an effective TDM service is the availability of appropriate analytical methods that produce results within a reasonable time, i. e., 48 h, and advice from someone who understands pharmacokinetics and therapeutics [184]. As shown in **Fig. 1**, the TDM process starts with the request and ends with the final decision about how to adjust a given patient's therapeutic regimen by the health care professional.

Request for plasma concentration quantification

As mentioned above, TDM should only be requested when there is evidence that the result will provide an answer to a specific question. If it is not clear what the question is, the answer is of little value. Typical indications are listed in **Table 6**. A single measurement is often insufficient for problem solving. For example, a series of measurements may be required at appropriate intervals to clarify if a low plasma concentration is either due to poor compliance, reduced bioavailability or abnormally rapid elimination.

Pre-TDM: Indication for TDM? - Availability of laboratory and pharmacological advise?

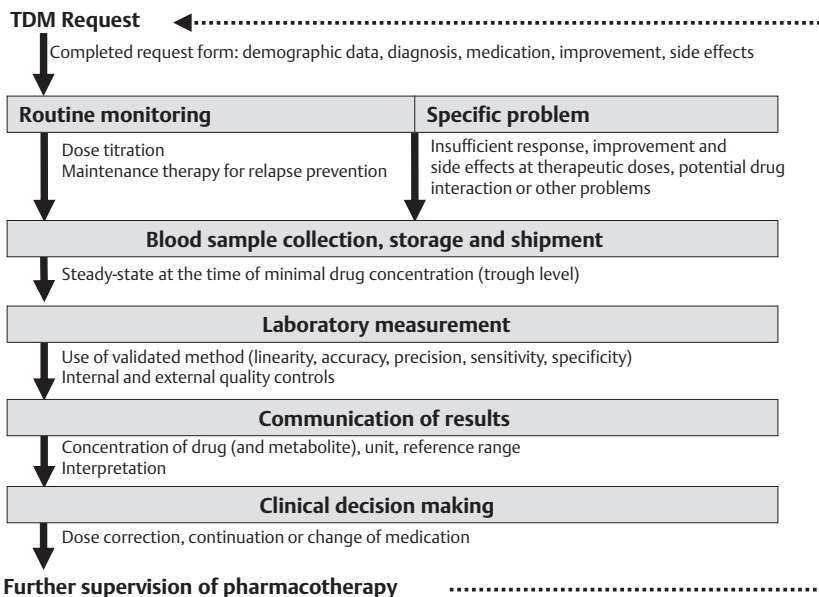


Fig. 1 Schematic overview of the TDM process as a guide for psychopharmacotherapy. Routine TDM is primarily applied to drugs with a narrow therapeutic index and a well-defined therapeutic reference range. However, TDM is useful for any psychotropic drug when addressing special therapeutic problems such as “therapy refractoriness” or side effects under recommended dosage.

LABORATORY

Address
Phone
Fax

REQUESTING HOSPITAL / DOCTOR

Address
Phone in case of alert
Fax

PATIENT DETAILS	Name or Code	<input type="checkbox"/> Inpatient <input type="checkbox"/> Outpatient	Date and time of blood withdrawal
Date of birth	Sex	Diagnosis / Symptom(s)	
<input type="checkbox"/> HIV-patient	Weight (kg)	Smoker <input type="checkbox"/> No <input type="checkbox"/> Moderate (<10 cig/day) <input type="checkbox"/> Heavy (>10cig/day)	Genotype to be considered (e.g. CYP2D6, CYP2C9, CYP2C19): _____

REASON FOR REQUEST (tick more than one if applicable)	<input type="checkbox"/> Dose adaptation	<input type="checkbox"/> Drug-drug interaction
<input type="checkbox"/> Control of adherence	<input type="checkbox"/> Insufficient improvement	<input type="checkbox"/> Control under maintenance therapy
	<input type="checkbox"/> Adverse effects (specify below)	<input type="checkbox"/> Other reason (to be specified)

SEVERITY OF ILLNESS (CGI-S) <i>How mentally ill is the patient at this time?</i>	IMPROVEMENT (CGI-I) <i>Change compared to condition at admission?</i>	SIDE EFFECTS (UKU) <input type="checkbox"/> not at all (0) <input type="checkbox"/> a little (1) <input type="checkbox"/> moderate (2) <input type="checkbox"/> severe (3)
<input type="checkbox"/> Not at all ill (1) <input type="checkbox"/> Borderline mentally ill (2) <input type="checkbox"/> Mildly ill (3) <input type="checkbox"/> Moderately ill (4) <input type="checkbox"/> Markedly ill (5) <input type="checkbox"/> Severely ill (6) <input type="checkbox"/> Extremely ill (7)	<input type="checkbox"/> Very much improved (1) <input type="checkbox"/> Much improved (2) <input type="checkbox"/> Minimally improved (3) <input type="checkbox"/> No change (4) <input type="checkbox"/> Minimally worse (5) <input type="checkbox"/> Much worse (6) <input type="checkbox"/> Very much worse (7)	<input type="checkbox"/> Concentration difficulties <input type="checkbox"/> Asthenia <input type="checkbox"/> Sleepiness/Sedation <input type="checkbox"/> Tension/inner unrest <input type="checkbox"/> Sleep disturbances <input type="checkbox"/> Emotional indifference <input type="checkbox"/> Dystonia <input type="checkbox"/> Rigidity <input type="checkbox"/> Hypokinesia/Akinesia <input type="checkbox"/> Hyperkinesia <input type="checkbox"/> Tremor <input type="checkbox"/> Akathisia <input type="checkbox"/> Epilepticseizures <input type="checkbox"/> Paresthesias <input type="checkbox"/> Headache <input type="checkbox"/> Accomodation disturbance <input type="checkbox"/> Increased salivation <input type="checkbox"/> Dry mouth <input type="checkbox"/> Nausea/Vomiting <input type="checkbox"/> Diarrhoea <input type="checkbox"/> Constipation <input type="checkbox"/> Micturation disturbance <input type="checkbox"/> Polyuria/Polydypsia <input type="checkbox"/> Increased sweating <input type="checkbox"/> Galactorrhoea <input type="checkbox"/> Weight gain <input type="checkbox"/> Sexual dysfunction <input type="checkbox"/> Other (to be specified) Causal relationship: <input type="checkbox"/> improbable <input type="checkbox"/> possible <input type="checkbox"/> probable

Drug(s) to be assayed	Formulation	Daily dose	Date started	Time of last dose

Other medications (include herbals, over-the-counter drugs etc)

TDM request : Blood should be withdrawn under steady-state conditions, preferably in the morning BEFORE taking the morning dose. Return the completed form, together with a minimum of 2 ml serum or plasma.

Date of sample receipt: _____

Signature: _____

Fig. 2 Request form for therapeutic drug monitoring in psychiatry.

TDM requests must include a completed request form (● Fig. 2) which is essential for effective drug concentration measurements and an adequate interpretation of the results [501,635]. The form should contain the patient name or code, demographic data, diagnosis, medication, reason for the request, the commercial and the generic name of the drug and its dose, the galenic formulation, the time of the last change of the dose, time of blood withdrawal. A brief comment on the clinical situation should be given for interpretation of the results. We recommend to use objective symptom rating, e.g., application of the clinical global impression (CGI) scale [283], to measure severity of illness and therapeutic improvement. The summary form of the UKU scale is useful to evaluate the occurrence and severity of side effects [402]. However, documented feedback to questionnaires indicates that clinicians often do NOT want to put that much information on the form. Moreover, the filled-in information is often not accurate. As an alternative, feedback by phone may be offered for interested physicians.

When interpretation of the results is requested from the laboratory, it is absolutely necessary to fill out the request forms adequately and completely. Thereby computerized ordering of TDM has advantages. It is inexpensive and it guides the ordering physician to give the relevant information required for interpretation in a comfortable way. Computerized ordering, however, is still not widely used. But effective packages are on the way to become available (e.g., www.konbest.de).

Blood sample collection

Generally, TDM is carried out in plasma or serum samples. The analysis of whole blood, which is long established for immunosuppressant drugs by using immunoassays [693], has been abandoned for TDM in psychiatry. There is no consensus whether plasma or serum should be preferred. Definite experimental data are still lacking which clearly demonstrate differences in the drug concentrations using either plasma or serum. The few available comparisons indicate that values obtained from serum or plasma can be used interchangeably [308]. Most psychoactive drugs are intensively bound to blood cells of plasma proteins. Concentrations of neuropsychiatric drugs reported in this guideline refer to the total drug fraction in accordance with the literature. For imipramine, it has been shown that the drug is rapidly and almost totally cleared by the brain through a single passage in the microvasculature [555]. The extraction was not significantly affected in the presence of albumin, lipoproteins or erythrocytes. For nortriptyline, statistical relationships between free levels of drug and clinical response were found to be insignificant [506]. Therefore it seems likely that the clinical response depends on the total drug fraction. Analysis of psychotropic medications in other materials such as urine, spinal fluid, tears, hairs or maternal milk have not been introduced for TDM purposes, and no validated data are available which deal with therapeutic concentrations. Saliva offers the advantage of non-invasive collection [20,25,356]. However, the drug concentration in saliva corresponds to the free (i.e., non-protein-bound) fraction of the drug in blood – which is for most psychopharmacologic medications only 10% or less of the total concentration. Thus detection problems may occur when using saliva instead of blood plasma or serum. In any case, more data will have to be obtained for saliva as a matrix for measurement of drug concentrations.

With few exceptions, TDM relies on trough steady-state plasma concentrations. Blood should therefore be collected after at least

4 drug elimination half-lives after the start of or a change in dosage and during the terminal β -elimination phase. For most psychotropic drugs, elimination half-lives vary between 12 and 36 h (● Table 5). Notable exceptions are quetiapine, trazodone, or venlafaxine, which display elimination half-lives around 6 h. Fluoxetine and aripiprazole have longer elimination half-lives. In clinical practice, the appropriate sampling time for most psychoactive drugs is one week after stable daily dosing and immediately before ingestion of the morning dose, which usually is 12–16 h (or 24 h if the drug is given once daily in the morning) after the last medication. If, for logistics reasons, blood can only be collected late in the morning, the patient should not be medicated before blood withdrawal. In an outpatient setting it is important to indicate exactly the time of administration of the last dose for interpretation. Trough levels can then be extrapolated by pharmacokinetic modelling.

In patients treated with a depot preparation of an antipsychotic drug, blood should be sampled immediately before the next injection. Formulations of antipsychotic drugs such as haloperidol decanoate or risperidone microspheres are characterised by a slow absorption after intramuscular administration. Maximum plasma concentration of first generation depot antipsychotics are reached after 1–14 days after injection, and the apparent elimination half-life is 2–3 weeks [647]. Similar properties exhibits the newly introduced paliperidone palmitate [131]. For risperidone microspheres the mean time to peak concentrations is 4 weeks and its plasma half life 4–6 days [647]. For other drugs delivered in extended or retarded release formulations like paliperidone [70] or quetiapine [212], special attention has to be given to the time of drug intake for correct interpretation (see ● Table 5). In these formulations, the time of maximum plasma concentration is delayed but the elimination half-life remains essentially unchanged. The long acting olanzapine pamoate is a new depot formulation [399]. The salt slowly releases olanzapine from the injection site into the muscle tissue. However, it dissolves rapidly when it is in contact with blood or plasma. The latter results in high plasma concentrations and may lead to marked sedation and delirium, the so called post-injection syndrome [399,647]. Considering this special problem it could be useful to control plasma concentrations of olanzapine shortly (i.e., about 2 h) after the i.m. injection to monitor if plasma concentrations increase. This approach, however, relies on the rapid quantification of olanzapine.

TDM may of course be carried at any time after drug ingestion if unexpected side effects are observed. It is not necessary to measure trough levels, but the dosing schedule should be reported for interpretation.

Storage and shipment of blood samples

When samples must be stored and sent frozen, it is required to prepare serum or plasma before freezing, since it is not possible to prepare serum or plasma from frozen blood. With few exceptions, serum or plasma samples can be stored in the dark (at 4°C) for at least 24 h, and most drug samples can be sent without freezing [305]. Exceptions are light and/or oxygen sensitive substances. For the determination of bupropion or methylphenidate, however, serum samples must be frozen or extracted and stabilized immediately after blood withdrawal and centrifugation (see ● Table 5). Olanzapine must be stored frozen (–20°C) if not analysed within 72 h [305]. The laboratory should give instructions on its web site or the request form how

to collect (plasma volume, labelling of the samples), store and mail the sample.

Laboratory measurements

Selective and sensitive analytical methods for the quantitative evaluation of drugs and their metabolites (analytes) are essential for the successful conduct of TDM. Methods must be validated which includes all of the procedures that demonstrate that a particular method used for quantitative measurement of analytes in a given biological matrix is reliable and reproducible for the intended use. The fundamental parameters for this validation include (1) accuracy, (2) precision, (3) selectivity, (4) sensitivity, (5) reproducibility and (6) stability. Validation involves documenting, through the use of specific laboratory investigations, that the performance characteristics of the method are suitable and reliable for the intended analytical applications. The acceptability of analytical data corresponds directly to the criteria used to validate the method [114,219].

For psychoactive drugs, chromatographic techniques (gas chromatography (GC), and high-performance liquid chromatography (HPLC), in combination with suitable detection methods, are preferred [186]. They are sufficiently precise, accurate and robust and can be adapted to the analysis of a huge number of drugs. A disadvantage is the need for sample preparation before chromatographic separation and hence a limited sample throughput. Throughput can be enhanced by automated sample preparation prior to GC or HPLC. Some laboratories have introduced HPLC with column switching which allows direct injection of plasma or serum into the HPLC system. Such procedures are available for a number of antidepressant [269,292–294,297,298,702,710] and antipsychotic drugs [368,369,571–573,709–712]. Another high-throughput chromatographic method is liquid chromatography coupled with mass spectroscopy (LC-MS) especially tandem MS (LC-MS/MS). LC/MSMS methods can be applied to almost any psychotropic drug including metabolites [577]. They are most sensitive and selective and can be used without time-consuming sample preparation. Many compounds can be analysed simultaneously. An excellent example is the LC-MS/MS method described by Kirchherr and Kühn-Felten [366]. This method was validated for over 50 psychoactive drugs. Disadvantageous for LC-MS/MS methods are high costs. Moreover, quantification can be problematic due to ion suppression and the availability of suitable calibration standards, preferentially deuterated analogues [584].

In case of suspected intoxications, TDM methods should enable drug analysis within 1–2 h [215]. For this purpose automated methods are advantageous.

The laboratory should not only analyse the drug but also its active metabolites, e.g., bupropion plus hydroxybupropion, clomipramine plus desmethylclomipramine, fluoxetine plus norfluoxetine, naltrexone plus naltrexol, risperidone plus 9-hydroxyrisperidone or venlafaxine plus O-desmethylvenlafaxine (● **Table 5**). For some drugs, the determination of metabolites that do not contribute to the overall clinical effect (e.g., norsertaline, normirtazapine, norcitalopram) is also useful to monitor drug adherence of the patient [546], to get information on his/her capacity to metabolise drugs, or to interpret drug-drug interactions when drugs are involved exhibiting enzyme inhibiting or inducing properties (● **Table 2**). “Normal” ratios of concentrations of metabolites to parent drugs that are expected in 68.3% of the patients are listed in ● **Table 3**. Any ratio outside the reported “normal” range should be considered as a signal

pointing to individual abnormalities due to a drug-drug-interaction, gene polymorphism, altered liver function, non-adherence or drug intake few hours before blood withdrawal.

The assay of enantiomers of chiral compounds requires either stereoselective derivatisation of the drugs prior to their quantification, or their separation by chiral chromatographic GC or HPLC columns. LC-MS/MS may be the method of choice. As an example, the TDM of the enantiomers of methadone using a classical detection method such as fluorescence or ultraviolet light absorption is often jeopardized by comedication or by coconsumption drugs of abuse. These problems may be circumvented by use of a mass detector, preferably a tandem mass spectrometer.

Within the therapeutic reference range, intraday- and interday precision should not exceed 15% (coefficient of variation) and accuracy should not deviate more than 15% from the nominal value [114,219].

To ensure quality and reliability of plasma concentrations assays, internal and external quality control procedures are mandatory. Samples must contain suitable internal standards, and each series of samples must include internal control samples. If standards are not available commercially, they should be prepared by personnel other than those performing the assays and by separate weighing of reference material. Reporting of results requires that the results of the quality controls are within the expected range. If quality controls are outside the expected range, the reason underlying the outlier needs to be clarified and documented.

The laboratory has to participate in an **external quality assessment scheme**, although this is not a legal requirement in all countries. For neuropsychiatric drugs, the first external quality program was introduced by Cardiff Bioanalytical Services Ltd in 1972 [720]. It has currently 450 participants from 36 countries (www.heathcontrol.com). Instand e.V. (www.instanddev.de/ringversuche/) is another recommended provider of external control, the external quality control scheme was recently expanded to multiple psychoactive drugs samples. Moreover, reference materials are also available from forensic chemistry (<http://www.pts-gtfch.de/>).

Communication of results

The concentration of the psychoactive drug as well as that of active metabolites contributing to the therapeutic action should be reported with reference ranges (● **Table 5**) either in mass or molar units. We recommend the use of mass units to relate concentration to dose. Laboratories vary in the presentation of their results. The clinician should take note of the units (i.e., ng/mL, µg/L, µmol/L, or nmol/L) in which the results of the analysis are expressed. This is especially recommended for comparisons of TDM values obtained from different laboratories or with those in the literature. To transform molar units into mass units and vice versa conversion factors are given in ● **Table 5**.

When drug concentrations are below the limit of quantification (LOQ), which refers to the lowest concentration of the standard curve that can be measured with at least 20% accuracy and precision, this limit should be indicated.

The results should be available for decision making within a clinically meaningful time. Although 24 h TDM service would be desirable, 48 h turnaround time is sufficient in most cases. In case of suspected intoxications, a few hours service is necessary [215]. To assist rapid intervention in patients at risk for toxicity or loss of tolerability, prompt information (phone call) of the

treating physician is required when the laboratory measures drug concentrations above the “laboratory alert level” which was newly defined (see above) in the present consensus guidelines (◉ **Table 5**).

Interpretation of results

We recommend that interpretation and pharmacologic advice are provided with every report. Expert interpretation of a drug concentration measurement and the adequate use of the information are essential to ensure the full clinical benefit of TDM. Reporting of results with inclusion of dose recommendations and other comments must be guided by the best available evidence. Expert knowledge may be necessary to calculate dose corrections or to analyse drug-drug interactions. It is therefore advantageous for the clinician to choose a laboratory that offers this service. Otherwise, the treating physician, a clinical pharmacologist or a trained expert of the clinic has to interpret the results. Access to specialist advice is also necessary if TDM results suggest that genotyping may be advisable [335].

Diagnosis and drug dose are important information for interpretation, since they permit a judgement on whether a result is plausible or not. Moreover, it must be controlled if blood samples were collected under recommended conditions, especially when the plasma concentration is unexpectedly high in an out-patient. When the drug was taken a few hours before blood sampling the drug concentration can be several-fold higher than the trough level.

For the interpretation of the results, it should not only be considered whether the plasma concentration of the drug is within the “therapeutic reference range” (◉ **Table 5**). It must also be considered if the drug plasma concentration is consistent with the dose (◉ **Table 4**). A plasma concentration may be outside the therapeutic reference range, just because a low or high dose was taken. In addition, it is wise to take into account the level of evidence underlying the “therapeutic reference range” of the particular drug (◉ **Table 5**). It should also be considered if the daily drug dose was given as a single or a multiple dose.

Often it is necessary to deal with pharmacokinetic properties such as metabolic pathways, enzymes involved and substrate and inhibitor properties of all drugs taken by the patient for interpretation of the results. Supportive information is therefore given in the present updated guidelines showing literature based substrate (◉ **Table 1**) and inhibitor or inducer properties of drugs (◉ **Table 3**) to deal with possible drug-drug interactions.

Any drug concentration outside its dose-related reference range (◉ **Table 5**) should alert the TDM laboratory to actively look for non-average pharmacokinetic drug disposition of the patient, drug-drug-interactions, gene polymorphisms that give rise to poor or ultra rapid metabolism, altered function of the excretion organs liver and kidneys, age and/or disease-related changes in the patient's pharmacokinetics, compliance (adherence) problems, a non-steady state and even signal interference from other medications that the patient may not have declared to the prescribing physician (e.g., St. John's wort) in the laboratory analysis. It may also be informative to calculate the dose related reference range (◉ **Table 4**) if the drug concentration lies outside the recommended therapeutic reference range (◉ **Table 5**) [285].

Plasma concentrations must be interpreted with the clinical presentation in mind. Recommendations on dosage changes constitute the most frequent advice. Other information which

could be of help for the physician are those related to genetic polymorphisms, risks of pharmacokinetic interactions in the case of polypragmasy, pharmacokinetic properties of the drug in patients belonging to a “special population”, e.g., elderly patients, or patients with hepatic or renal insufficiency. For the treatment of pain, relatively low plasma concentrations of tricyclic antidepressants may be sufficient. They may be within the “dose related reference range” (◉ **Table 4**) but outside the “therapeutic reference range” of ◉ **Table 5** which was established for the indication of depression.

A laboratory may recommend that an additional sample should be taken after a certain period, because in cases with unusually low or high plasma concentrations, repeated measurements may help to decide whether the patient's adherence is inconsistent (irregular intake of the drug) or whether the patient is an abnormal metabolizer.

Since the interpretation of TDM results relies on complex quantitative relationships, training in clinical psychopharmacology and pharmacokinetics and the application of TDM is essential. Regular conferences with discussion of the interpretation of real cases are most helpful for learning. It is also recommended that junior psychiatrists interpret the results under supervision of an expert.

Clinical decision making

A TDM result is a guide to proper dosing of the individual patient. The physician has to be aware that, under optimal conditions, reporting of results with inclusion of dose recommendations and other comments by the laboratory is guided by the best available evidence [310]. The laboratory, however, has only a restricted knowledge of the clinical situation. On the other hand, most treating physicians have limited pharmacokinetic knowledge. Therefore it is essential to be aware that optimal TDM is an interdisciplinary task that requires close communication between laboratory and clinical experts.

If the plasma concentration of the drug is within the therapeutic reference range, an adaptation of the dose is, of course only recommended when clinical reasons, such as adverse effects or non-response clearly justify such a decision. Evidently, the treating physician has to decide whether the treatment strategy is to be changed or not. On the other hand, when the advice given on the TDM report is not followed, the reason for this course of action must be substantiated to allow evaluation of the treating physician's decision should the patient come to harm. Recommendations for such an evaluation in a court of law have been recently published by the TDM-AGNP group [741].

In patients with abnormally rapid elimination it may be useful to prescribe a dose above the maximal recommended dose, since such patients can exhibit drug concentrations below the reference range under standard doses. However, the medication should be changed if the patient exhibited sufficiently high drug concentrations for a sufficiently long treatment period, i.e., for at least 2 weeks, and did not improve by at least 20%.

When **adverse effects** are associated with clinical improvement under recommended doses, measurement of the plasma concentration may clarify if side effects are related to exceedingly high drug levels in the blood. In this situation, the dose can be decreased, normally without risk of loss of action.

For the treatment with antidepressant or antipsychotic drugs, there is good evidence that clinical non-improvement at week 2 is highly predictive for later response and remission [119, 138, 392, 620, 621, 638]. Especially the absence of early improvement

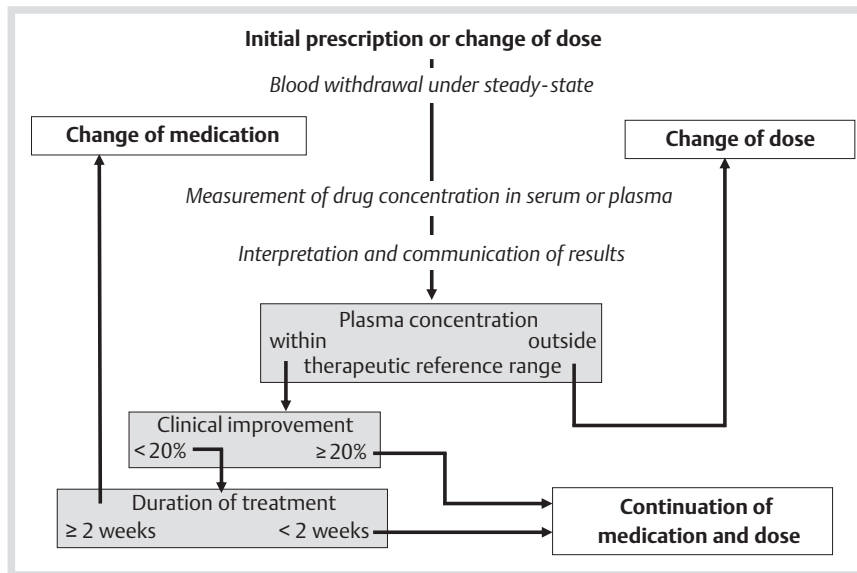


Fig. 3 TDM-guided dose titration of antidepressant or antipsychotic drug treatment (adapted from [311]). Clinical decision making has to consider the clinical improvement, the duration of treatment, and steady-state concentration of the drug in plasma or serum. The steady-state is reached by 94% after 4 elimination half-lives of the drug or active metabolites (see Table 5).

appears to be a highly reliable predictor of later non-response [358]. For dose titration with antidepressant and antipsychotic drugs we therefore recommend to include symptom rating by the treating physician [138] at baseline and at week 2 in addition to drug concentration measurements. Fig. 3 summarizes the above recommendations in a flow chart.

When further plasma concentration measurements are recommended after a modification of the dose or after prescription of a comedication that is known to inhibit or enhance the metabolism of the drug to be measured, the next TDM should be delayed until steady-state conditions are reached again. For this, the terminal elimination half-life of the drug has to be considered (Table 5).

Pharmacogenetic tests in addition to TDM

Concentrations outside the reference range may be due to gene polymorphisms that give rise to slow/rapid metabolizers. As a consequence, the laboratory may also suggest that a pharmacogenetic test should be carried out [14, 144, 158, 193, 335, 362, 365, 377, 623, 624, 675]. Genotyping, however, is not available in all TDM laboratories, and we recommend consultation of specialized laboratories for interpretation of the results.

Situations and cases where pharmacogenetic tests could advantageously be combined with TDM are explained in more detail by Jaquenoud Sirot and coworkers [335]. Some of the most important indications for the combination of genotyping with TDM are the following:

- ▶ the patient is treated with a substrate the metabolism of which shows a wide interindividual variability;
- ▶ a drug is characterized by a small therapeutic index: risk of toxicity in the case of a genetically impaired metabolism, or on the other hand, risk of non-response due to an ultra-rapid metabolism and the inability to reach therapeutic drug levels;
- ▶ the patient presents unusual plasma concentrations of the drug or its metabolite(s) and genetic factors are suspected to be responsible;
- ▶ the patient suffers from a chronic illness, which requires life-long treatment.

In a patient who is genotyped as a PM or UM, the medication should not automatically be replaced by another as suggested by

some authors, but the dose can often be adapted, using clinical judgement and TDM.

Conclusions and Perspectives

The choice of pharmacologic treatment should always take into account the clinical presentation of the patient and consider psychopathology and drug history. TDM is, if used appropriately, a valid tool for optimising pharmacotherapy. During the past decades, knowledge on the metabolic fate and actions of psychotropic drugs in the human body has markedly advanced. Pharmacogenetic and environmental factors have been identified and summarized in the first part of this review. The present updated AGNP guidelines describe the best practice of TDM in psychiatry in order to promote the appropriate use of TDM. Although a considerable body of data for plasma concentrations of psychotropic drugs has been accumulated and although our knowledge about the quantitative relationship between plasma concentration and therapeutic response has improved, there is still a need to conduct further controlled and randomised concentration-response studies to improve the quality of data on therapeutic reference ranges. We also recommend inclusion of pharmacokinetic measurements during phase III and IV studies. Product information should be supplemented with TDM related data to enhance the therapeutic effectiveness of psychoactive drugs. Analyses of German [671] and French [568] summaries of product characteristics (SPC) revealed that many SPC do not contain TDM related information in spite of available valid clinical-scientific evidence. Another need for research is to study cost-effectiveness of TDM when the method is used in an appropriate way. Polypharmacy is very common in psychiatry while essentially all TDM recommendations are based on single-medication trials. Thus, the efficacy of drug combinations constitutes a severely under-investigated area of TDM. Finally, one should never forget that TDM is an interdisciplinary task that sometimes requires the respectful discussion of apparently discrepant data so that, ultimately, the patient can profit from such a joint effort.

Conflicts of Interest

Christoph Hiemke has received speaker's or consultancy fees from the following pharmaceutical companies: Bristol-Myers Squibb, Pfizer, Lilly and Servier. He is managing director of the psiac GmbH which provides an internet based drug-drug interaction program for psychopharmacotherapy. He reports no conflict of interest with this publication. Pierre Baumann has received speaker's or consultancy fees from almost all pharmaceutical companies selling psychotropic drug in Switzerland. He reports no conflict of interest with this publication. Niels Bergemann, Mirjam Fric, Christine Greiner, Hartmut Kirchherr, Ulrich C Lutz, Bernhard Rambeck, Bernd Schoppek, Julia C Stingl, Manfred Uhr and Roland Waschglar have no conflict of interest to declare. Andreas Conca has served as a consultant for Lilly, BMS, Pfizer. He has served on the speakers' bureau of Lilly, BMS, AstraZeneca, Lundbeck, Italfarma, Janssen. He reports no conflict of interest with this publication. Otto Dietmaier has received speaker's or consultancy fees from Bristol-Myers Squibb, Janssen, Eli Lilly and Lundbeck. He reports no conflict of interest with this publication. Ursula Havemann-Reinecke has received speaker's or consultancy fees or unrestricted educational grants from AstraZeneca, Bristol-Myers Squibb, Cephalon, Essex, Janssen Cilag, Lundbeck, Pfizer, Schering-Plough, Wyeth. She reports no conflict of interest with this publication. Ekkehard Haen has served as a consultant and received speaker's fees from Janssen-Cilag, Lilly, Pfizer, GlaxoSmithKline, AstraZeneca, Bristol-Myers Squibb, Otsuka, Bayer Vital, Servier and Südmedica GmbH. He reports no conflict of interest with this publication. Karin Egberts has received speaker's fees or travel grants from Wyeth and Medice. She participated in performing clinical trials for AstraZeneca, Janssen-Cilag, Lilly and Shire. She reports no conflict of interest with this publication. Gerhard Gründer has served as a consultant for AstraZeneca, Bristol-Myers Squibb, Eli Lilly, Johnson & Johnson, and Otsuka. He has served on the speakers' bureau of AstraZeneca, Bristol-Myers Squibb, Eli Lilly, Janssen Cilag, Otsuka, Pfizer, Servier, and Wyeth. He has received grant support from Alkermes, Bristol-Myers Squibb, Eli Lilly, and Johnson & Johnson. He is co-founder of Pharma-Image – Molecular Imaging Technologies GmbH. He reports no conflict of interest with this publication. Eveline Jaquenoud Sirot is managing director of mediQ which sells an internet based drug-drug interaction program for psychiatry. She reports no conflict of interest with this publication. Gerd Laux has received speaker's or consultancy fees or unrestricted educational grants from AstraZeneca, Bayer, Eli Lilly, Lundbeck, Merz, Pfizer, Servier and Wyeth. He reports no conflict of interest. Bruno Pfuhlmann has received speaker's or consultancy fees from AstraZeneca, Janssen and Pfizer. He reports no conflict of interest with this publication. Manfred Gerlach has received speaker's or consultancy honoraria or restricted research grants from Boehringer Ingelheim Pharma GmbH & Co. KG, Desitin Arzneimittel GmbH, Janssen Cilag GmbH, Lundbeck GmbH and Merz Pharmaceuticals GmbH. He reports no conflict of interest with this paper. Thomas Messer has received speaker's or consultancy fees or unrestricted educational grants from Eli Lilly, Bristol-Myers Squibb, Janssen, Servier, Pfizer, Lundbeck and Bayer Vital Health Care. He reports no conflict of interest with this publication. Matthias J. Müller has received speaker's or consultancy fees from Janssen, Servier, Pfizer, and Astra-Zeneca. He reports no conflict of interest with this publication. Sven Ulrich is an employe of Ariston Pharma GmbH, Berlin, Germany. He reports

no conflict of interest with this publication. Gerald Zernig has received speaker's or consultancy fees or unrestricted educational grants from AlcaSynn, AstraZeneca, Bio-Rad, Bristol-Myers Squibb, Eli Lilly, Lundbeck, Mundipharma, Novartis, Pfizer, and Wyeth. He reports no conflict of interest with this publication.

Acknowledgements

The authors thank Sonja Brünen, Elnaz Ostad Haji, Christiane Knoth and Viktoria Stieffenhofer for helping us to calculate ratios of plasma concentrations of metabolite and parent compound reported in the literature and shown in **Table 2**. We thank Ralf Köber for his help in evaluating the therapeutic reference ranges of antimentia drugs. We thank Michaela Jahnke, Christiane Kobelt and Nina Wenzel for most helpful editorial assistance, especially for organization of the long list of references.

Affiliations

- ¹ Department of Psychiatry and Psychotherapy, University Medical Center of Mainz, Germany
- ² Department of Psychiatry, University of Lausanne, Prilly-Lausanne, Switzerland
- ³ Psychiatric Hospital, Bad Arolsen, Germany
- ⁴ Psychiatric Hospital, Bolzano, Italy
- ⁵ Psychiatrc Hospital, Weinsberg, Germany
- ⁶ Department Child and Adolescent Psychiatry, University Hospital of Würzburg, Germany
- ⁷ Kliniken des Bezirks Oberbayern (kbo) Salzach-Inn-Klinikum, Wasserburg a. Inn, Germany
- ⁸ Federal Institute for Drugs and Medical Devices (BfArM), Bonn, Germany
- ⁹ Department of Psychiatry and Psychotherapy, University of Aachen, Germany
- ¹⁰ Clinical Pharmacology, Department of Psychiatry and Psychosomatics, University of Regensburg, Germany
- ¹¹ Department of Psychiatry and Psychosomatics, University of Göttingen, Germany
- ¹² Psychiatric Hospital, Königfelden, Brugg, Aargau, Switzerland
- ¹³ Medical Laboratory Bremen, Germany
- ¹⁴ Department of Psychiatry and Psychotherapy, University of Tübingen, Germany
- ¹⁵ Psychiatric Hospital, Pfaffenhofen, Germany
- ¹⁶ Psychiatric Hospital, Marburg and Gießen, Germany
- ¹⁷ Department of Psychiatry, Psychotherapy and Psychosomatics, University Hospital of Würzburg, Germany
- ¹⁸ Center of Epilepsy, Bielefeld, Germany
- ¹⁹ Psychiatric Hospital, Haar, Germany
- ²⁰ Department of Pharmacology of Natural Products and Clinical Pharmacology, University of Ulm, Germany
- ²¹ Max Planck Institute of Psychiatry, Munich, Germany
- ²² Aristo Pharma GmbH, Berlin, Germany
- ²³ Psychiatric Hospital, Feldkirch, Austria
- ²⁴ Experimental Psychiatry Unit, Department of Psychiatry and Psychotherapy, Medical University of Innsbruck, Austria

References

- 1 Abbott NJ, Patabendige AA, Dolman DE et al. Structure and function of the blood-brain barrier. *Neurobiol Dis* 2010; 37: 13–25
- 2 Abernethy DR, Greenblatt DJ, Shader RI. Imipramine and desipramine disposition in the elderly. *J Pharmacol Exp Ther* 1985; 232: 183–188
- 3 Adam K, Oswald I. Effects of lormetazepam and of flurazepam on sleep. *Br J Clin Pharmacol* 1984; 17: 531–538
- 4 Adamiak U, Kaldonska M, Klodowska-Duda G et al. Pharmacokinetic-pharmacodynamic modeling of levodopa in patients with advanced Parkinson disease. *Clin Neuropharmacol* 2010; 33: 135–141
- 5 Addington D. Best practices: improving quality of care for patients with first-episode psychosis. *Psychiatr Serv* 2009; 60: 1164–1166
- 6 Adli M, Baethge C, Heinz A et al. Is dose escalation of antidepressants a rational strategy after a medium-dose treatment has failed? *Eur Arch Psychiatry Clin Neurosci* 2005; 55: 387–400

- 7 Aichhorn W, Marksteiner J, Walch T et al. Influence of age, gender, body weight and valproate comedication on quetiapine plasma concentrations. *Int Clin Psychopharmacol* 2006; 21: 81–85
- 8 Aichhorn W, Weiss U, Marksteiner J et al. Influence of age and gender on risperidone plasma concentrations. *J Psychopharmacol* 2005; 19: 395–401
- 9 Aichhorn W, Whitworth AB, Weiss ME et al. Second-generation antipsychotics: Is there evidence for sex differences in pharmacokinetic and adverse effect profiles? *Drug Saf* 2006; 29: 587–598
- 10 Åkerblad AC, Bengtsson F, Ekselius L et al. Effects of an educational compliance enhancement programme and therapeutic drug monitoring on treatment adherence in depressed patients managed by general practitioners. *Int Clin Psychopharmacol* 2003; 18: 347–354
- 11 Åkerblad AC, Bengtsson F, Holgersson M et al. Identification of primary care patients at risk of nonadherence to antidepressant treatment. *Patient Prefer Adherence* 2008; 2: 379–386
- 12 Akhlu E, Kalow W, Endrenyi L et al. CYP2D6 and DRD2 genes differentially impact pharmacodynamic sensitivity and time course of prolactin response to perphenazine. *Pharmacogenet Genomics* 2007; 17: 989–993
- 13 Akutsu T, Kobayashi K, Sakurada K et al. Identification of human cytochrome p450 isozymes involved in diphenhydramine N-demethylation. *Drug Metab Dispos* 2007; 35: 72–78
- 14 Albers LJ, Ozdemir V, Marder SR et al. Low-dose fluvoxamine as an adjunct to reduce olanzapine therapeutic dose requirements: a prospective dose-adjusted drug interaction strategy. *J Clin Psychopharmacol* 2005; 25: 170–174
- 15 Alderman J, Wolkow R, Fogel IM. Drug concentration monitoring with tolerability and efficacy assessments during open-label, long-term sertraline treatment of children and adolescents. *J Child Adolesc Psychopharmacol* 2006; 16: 117–129
- 16 Allen MH, Hirschfeld RM, Wozniak PJ et al. Linear relationship of valproate serum concentration to response and optimal serum levels for acute mania. *Am J Psychiatry* 2006; 163: 272–275
- 17 Allqvist A, Miura J, Bertilsson L et al. Inhibition of CYP3A4 and CYP3A5 catalyzed metabolism of alprazolam and quinine by ketoconazole as racemate and four different enantiomers. *Eur J Clin Pharmacol* 2007; 63: 173–179
- 18 Altamura AC, Moro AR, Percudani M. Clinical pharmacokinetics of fluoxetine. *Clin Pharmacokinet* 1994; 26: 201–214
- 19 Althaus M, Retzow A, Castell JV et al. In vitro identification of the cytochrome P450 isoform responsible for the metabolism of alpha-dihydroergocryptine. *Xenobiotica* 2000; 30: 1033–1045
- 20 Aman MG, Vinks AA, Remmerie B et al. Plasma pharmacokinetic characteristics of risperidone and their relationship to saliva concentrations in children with psychiatric or neurodevelopment disorders. *Clin Therap* 2007; 29: 1476–1486
- 21 Anderson D, Reed S, Lintemoot J et al. A first look at duloxetine (Cymbalta®) in a post-mortem laboratory. *J Analyt Toxicology* 2006; 30: 576–579
- 22 Anderson GD. Pregnancy-induced changes in pharmacokinetics: a mechanistic-based approach. *Clin Pharmacokinet* 2005; 44: 989–1008
- 23 Anderson IM, Ferrier IN, Baldwin RC et al. Evidence-based guidelines for treating depressive disorders with antidepressants: a revision of the 2000 British Association for Psychopharmacology guidelines. *J Psychopharmacol* 2008; 22: 343–396
- 24 Aoki FY, Sitar DS. Clinical pharmacokinetics of amantadine hydrochloride. *Clin Pharmacokinet* 1988; 14: 35–51
- 25 Aps JK, Martens LC. Review: The physiology of saliva and transfer of drugs into saliva. *Forensic Sci Int* 2005; 150: 119–131
- 26 Arakawa R, Ito H, Takano A et al. Dose-finding study of paliperidone ER based on striatal and extrastriatal dopamine D2 receptor occupancy in patients with schizophrenia. *Psychopharmacology (Berl)* 2008; 197: 229–235
- 27 Aravagiri M, Marder SR, Yuwiler A et al. Distribution of fluphenazine and its metabolites in brain regions and other tissues of the rat. *Neuropsychopharmacology* 1995; 13: 235–247
- 28 Aravagiri M, Teper Y, Marder SR. Pharmacokinetics and tissue distribution of olanzapine in rats. *Biopharm Drug Dispos* 1999; 20: 369–377
- 29 Aravagiri M, Yuwiler A, Marder SR. Distribution after repeated oral administration of different dose levels of risperidone and 9-hydroxy-risperidone in the brain and other tissues of rat. *Psychopharmacology* 1998; 139: 356–363
- 30 Åsberg M, Crönholm B, Sjöqvist F et al. Correlation of subjective side effects with plasma concentrations of nortriptyline. *Br Med J* 1970; 5726: 18–21
- 31 Åsberg M, Crönholm B, Sjöqvist F et al. Relationship between plasma level and therapeutic effect of nortriptyline. *Br Med J* 1971; 3: 331–334
- 32 Bachmann CJ, Haberhausen M, Heinzel-Gutenbrunner M et al. Large intraindividual variability of olanzapine serum concentrations in adolescent patients. *Ther Drug Monit* 2008; 30: 108–112
- 33 Bachmann CJ, Rieger-Gies A, Heinzel-Gutenbrunner M et al. Large variability of aripiprazole and dehydroaripiprazole serum concentrations in adolescent patients with schizophrenia. *Ther Drug Monit* 2008; 30: 462–466
- 34 Bachus R, Bickel U, Thomsen T et al. The O-demethylation of the anti-dementia drug galanthamine is catalysed by cytochrome P450 2D6. *Pharmacogenetics* 1999; 9: 661–668
- 35 Backman JT, Olkkola KT, Ojala M et al. Concentrations and effects of oral midazolam are greatly reduced in patients treated with carbamazepine or phenytoin. *Epilepsia* 1996; 37: 253–257
- 36 Bagli M, Höflich G, Rao ML et al. Bioequivalence and absolute bioavailability of oblong and coated levomepromazine tablets in CYP2D6 phenotyped subjects. *Int J Clin Pharmacol Ther* 1995; 33: 646–652
- 37 Baker GB, Urchuk LJ, McKenna KF et al. Metabolism of monoamine oxidase inhibitors. *Cell Mol Neurobiol* 1999; 19: 411–426
- 38 Bakken GV, Rudberg I, Christensen H et al. Metabolism of quetiapine by CYP3A4 and CYP3A5 in presence or absence of cytochrome B5. *Drug Metab Dispos* 2009; 37: 254–258
- 39 Balant LP, Balant-Gorgia AE, Eisele R et al. Clinical and pharmacokinetic evaluation of zuclopentixol acetate in Viscolear. *Pharmacopsychiatry* 1989; 22: 250–254
- 40 Balant-Gorgia AE, Eisele R, Aeschlimann JM et al. Plasma flupentixol concentrations and clinical response in acute schizophrenia. *Ther Drug Monit* 1985; 7: 411–414
- 41 Barbhaiya RH, Shukla UA, Pfeiffer M et al. Disposition kinetics of buspirone in patients with renal or hepatic impairment after administration of single and multiple doses. *Eur J Clin Pharmacol* 1994; 46: 41–47
- 42 Bareggi SR, Bianchi L, Cavallaro R et al. Citalopram concentrations and response in obsessive-compulsive disorder – Preliminary results. *CNS Drugs* 2004; 18: 329–335
- 43 Barski OA, Tipparaju SM, Bhatnagar A. The aldo-keto reductase superfamily and its role in drug metabolism and detoxification. *Drug Metab Rev* 2008; 40: 553–624
- 44 Baruzzi A, Bordo B, Bossi L et al. Plasma levels of di-n-propylacetate and clonazepam in epileptic patients. *Int J Clin Pharmacol Biopharm* 1977; 15: 403–408
- 45 Bates DW, Gawande AA. Improving safety with information technology. *N Engl J Med* 2003; 348: 2526–2534
- 46 Bauer M, Whybrow PC, Angst J et al. World Federation of Societies Biological Psychiatry Task Force on Treatment Guidelines for Unipolar Depressive Disorders. World Federation of Societies of Biological Psychiatry (WFSBP) Guidelines for Biological Treatment of Unipolar Depressive Disorders, Part 1: Acute and continuation treatment of major depressive disorder. *World J Biol Psychiatry* 2002; 3: 5–43
- 47 Bauer S, David Rudd G, Mylius V et al. Lacosamide intoxication in attempted suicide. *Epilepsy Behav* 2010; 17: 549–551
- 48 Baumann P. Pharmacokinetic-pharmacodynamic relationship of the selective serotonin reuptake inhibitors. *Clin Pharmacokinet* 1996; 31: 444–469
- 49 Baumann P, Barbe R, Vabre-Bogdalova A et al. Epileptiform seizure after sertraline treatment in an adolescent experiencing obsessive-compulsive disorder and presenting a rare pharmacogenetic status. *J Clin Psychopharmacol* 2006; 26: 679–681
- 50 Baumann P, Hiemke C, Ulrich S et al. Le dosage plasmatique des médicaments psychotropes à des fins thérapeutiques: recommandations du groupe d'experts AGNP-TDM. *Rev Med Suisse* 2006; 2: 1413–1418
- 51 Baumann P, Hiemke C, Ulrich S et al. The AGNP-TDM expert group consensus guidelines: therapeutic drug monitoring in psychiatry. *Pharmacopsychiatry* 2004; 37: 243–265
- 52 Baumann P, Ulrich S, Eckermann G et al. The AGNP-TDM Expert Group Consensus Guidelines: focus on therapeutic monitoring of antidepressants. *Dialogues Clin Neurosci* 2005; 7: 231–247
- 53 Baumann P, Zullino DF, Eap CB. Enantiomers' potential in psychopharmacology – a critical analysis with special emphasis on the antidepressant escitalopram. *Eur Neuropsychopharmacol* 2002; 12: 433–444
- 54 Bazire S. *Psychotropic Drug Directory* 2010. Aberdeen: Healthcomm UK Ltd, 2011

- 55 Beasley CM Jr, Stauffer VL, Liu-Seifert H et al. All-cause treatment discontinuation in schizophrenia during treatment with olanzapine relative to other antipsychotics: an integrated analysis. *J Clin Psychopharmacol* 2007; 27: 252–258
- 56 Bech P, Gex-Fabry M, Aubry JM et al. Olanzapine plasma level in relation to antimanic effect in the acute therapy of manic states. *Nord J Psychiatry* 2006; 60: 181–182
- 57 Becquemont L, Mouajjah S, Escaffre O et al. Cytochrome P-450 3A4 and 2C8 are involved in zopiclone metabolism. *Drug Metab Dispos* 1999; 27: 1068–1073
- 58 Beedham C, Miceli JJ, Obach RS. Ziprasidone metabolism, aldehyde oxidase, and clinical implications. *J Clin Psychopharmacol* 2003; 23: 229–232
- 59 Benedetti MS, Whomsley R, Poggesi I et al. Drug metabolism and pharmacokinetics. *Drug Metab Rev* 2009; 41: 344–390
- 60 Benetton SA, Fang C, Yang YO et al. P450 phenotyping of the metabolism of selegiline to desmethylselegiline and methamphetamine. *Drug Metab Pharmacokinet* 2007; 22: 78–87
- 61 Bengtsson F. Therapeutic drug monitoring of psychotropic drugs. TDM “nouveau”. *Ther Drug Monit* 2004; 26: 145–151
- 62 Bennett JP Jr, Piercey MF. Pramipexole: a new dopamine agonist for the treatment of Parkinson's disease. *J Neurol Sci* 1999; 163: 25–31
- 63 Bergemann N, Frick A, Parzer P et al. Olanzapine plasma concentration, average daily dose, and interaction with co-medication in schizophrenic patients. *Pharmacopsychiatry* 2004; 37: 63–68
- 64 Bergemann N, Kopitz J, Kress KR et al. Plasma amisulpride levels in schizophrenia or schizoaffective disorder. *Eur Neuropsychopharmacology* 2004; 14: 245–250
- 65 Bergemann N, Rommel F, Conca A. Therapeutisches Drug-Monitoring von Psychopharmaka in der Schwangerschaft. *J Neurol Neurochir Psychiatr* 2009; 10: 38–40
- 66 Bergmann TK, Bathum L, Brösen K. Duplication of CYP2D6 predicts high clearance of desipramine but high clearance does not predict duplication of CYP2D6. *Eur J Clin Pharmacol* 2001; 57: 123–127
- 67 Bergstrom RF, Cerimele BJ. Olanzapine in subjects with and without renal failure (data on file). Lilly Laboratory for Clinical Research. Eli Lilly and Co. 1996
- 68 Berry D, Millington C. Analysis of pregabalin at therapeutic concentrations in human plasma/serum by reversed-phase HPLC. *Ther Drug Monit* 2005; 27: 451–456
- 69 Bertelsen KM, Venkatakrishnan K, von Moltke LL et al. Apparent mechanism-based inhibition of human CYP2D6 in vitro by paroxetine: comparison with fluoxetine and quinidine. *Drug Metab Dispos* 2003; 31: 289–293
- 70 Berwaerts J, Cleton A, Rossenu S et al. A comparison of serum prolactin concentrations after administration of paliperidone extended-release and risperidone tablets in patients with schizophrenia. *J Psychopharmacol* 2010; 24: 1011–1018
- 71 Bigliani V, Mulligan RS, Acton PD et al. Striatal and temporal cortical D2/D3 receptor occupancy by olanzapine and sertindole in vivo: a [123I]epidepride single photon emission tomography (SPET) study. *Psychopharmacology (Berl)* 2000; 150: 132–140
- 72 Birkenhäger TK, van den Broek WW, Moleman P et al. Imipramine dose in relation to therapeutic plasma level: are clinical trials using imipramine as a positive control flawed? *Psychopharmacology (Berl)* 2005; 181: 595–599
- 73 Bjerkenstedt L, Flyckt L, Overø KF et al. Relationship between clinical effects, serum drug concentration and serotonin uptake inhibition in depressed patients treated with citalopram. A double-blind comparison of three dose levels. *Eur J Clin Pharmacol* 1985; 28: 553–557
- 74 Bjørndal N, Bjerre M, Gerlach J et al. High dosage haloperidol therapy in chronic schizophrenic patients: a double-blind study of clinical response, side effects, serum haloperidol, and serum prolactin. *Psychopharmacology (Berl)* 1980; 67: 17–23
- 75 Bockbrader HN. Clinical pharmacokinetics of gabapentin. *Drugs Today* 1995; 31: 613–619
- 76 Bockbrader HN, Burger P, Knapp L et al. Population pharmacokinetics of pregabalin in healthy subjects and patients with chronic pain or partial seizures. *Epilepsia* 2011; 52: 248–257
- 77 Bockbrader HN, Wesche D, Miller R et al. A comparison of the pharmacokinetics and pharmacodynamics of pregabalin and gabapentin. *Clin Pharmacokinet* 2010; 49: 661–669
- 78 Bogaards JJ, Hissink EM, Briggs M et al. Prediction of interindividual variation in drug plasma levels in vivo from individual enzyme kinetic data and physiologically based pharmacokinetic modeling. *Eur J Pharm Sci* 2000; 12: 117–124
- 79 Bomsien S, Aderjan R, Mattern R et al. Effect of psychotropic medication on the in vitro metabolism of buprenorphine in human cDNA-expressed cytochrome P450 enzymes. *Eur J Clin Pharmacol* 2006; 62: 639–643
- 80 Bond A, Seijas D, Dawling S et al. Systemic absorption and abuse liability of snorted flunitrazepam. *Addiction* 1994; 89: 821–830
- 81 Bondolfi G, Morel F, Crettol et al. Increased clozapine plasma concentrations and side effects induced by smoking cessation in 2 CYP1A2 genotyped patients. *Ther Drug Monit* 2005; 27: 539–543
- 82 Bont L, Bosker HA, Brus F et al. Torsade de pointes after pipamperone intoxication. *Pharm World Sci* 1998; 20: 137
- 83 Borgstrom L, Larsson H, Molander L. Pharmacokinetics of parenteral and oral melperone in man. *Eur J Clin Pharmacol* 1982; 23: 173–176
- 84 Borys DJ, Setzer SC, Ling LJ et al. Acute fluoxetine overdose: a report of 234 cases. *Am J Emerg Med* 1992; 10: 115–120
- 85 Bosse GM, Spiller HA, Collins AM. A fatal case of venlafaxine overdose. *J Med Toxicol* 2008; 4: 18–20
- 86 Brachtendorf L, Jetter A, Beckurts KT et al. Cytochrome P450 enzymes contributing to demethylation of maprotiline in man. *Pharmacol Toxicol* 2002; 90: 144–149
- 87 Brahmi N, Kouraichi N, Abderrazek H et al. Clinical experience with carbamazepine overdose: relationship between serum concentration and neurological severity. *J Clin Psychopharmacol* 2008; 28: 241–243
- 88 Brandt C, Baumann P, Eckermann G et al. Therapeutic drug monitoring in Epileptologie und Psychiatrie (Therapeutic drug monitoring in epileptology and psychiatry). *Nervenarzt* 2008; 79: 167–174
- 89 Bressan RA, Erlandsson K, Jones HM et al. Is regionally selective D2/D3 dopamine occupancy sufficient for atypical antipsychotic effect? An in vivo quantitative [123I] epidepride SPET study of amisulpride-treated patients. *Am J Psychiatry* 2003; 160: 1413–1420
- 90 Breyer-Pfaff U. The metabolic fate of amitriptyline, nortriptyline and amitriptyline oxide in man. *Drug Metab Rev* 2004; 36: 723–746
- 91 Breyer-Pfaff U, Brinkschulte M, Rein W et al. Prediction and evaluation criteria in perazine therapy of acute schizophrenics pharmacokinetic data. *Pharmacopsychiatry* 1983; 16: 160–165
- 92 Breyer-Pfaff U, Nill K. Carbonyl reduction of naltrexone and dolasetron by oxidoreductases isolated from human liver cytosol. *J Pharm Pharmacol* 2004; 56: 1601–1606
- 93 Brockmüller J, Kirchheiner J, Schmider J et al. The impact of the CYP2D6 polymorphism on haloperidol pharmacokinetics and on the outcome of haloperidol treatment. *Clin Pharmacol Ther* 2002; 72: 438–452
- 94 Brösen K. Drug-metabolizing enzymes and therapeutic drug monitoring in psychiatry. *Ther Drug Monit* 1996; 18: 393–396
- 95 Brösen K, Gram LF, Klysner R et al. Steady-state levels of imipramine and its metabolites: significance of dose-dependent kinetics. *Eur J Clin Pharmacol* 1986; 30: 43–49
- 96 Brösen K, Klysner R, Gram LF et al. Steady-state concentrations of imipramine and its metabolites in relation to the sparteine/debrisoquine polymorphism. *Eur J Clin Pharmacol* 1986; 30: 679–684
- 97 Brösen K, Naranjo CA. Review of pharmacokinetic and pharmacodynamic interaction studies with citalopram. *Eur Neuropsychopharmacol* 2001; 11: 275–283
- 98 Bruijn JA, Moleman P, Mulder PG et al. A double-blind, fixed blood-level study comparing mirtazapine with imipramine in depressed in-patients. *Psychopharmacology (Berl)* 1996; 127: 231–237
- 99 Brünen S, Vincent DP, Baumann P et al. Therapeutic Drug Monitoring (TDM) for drugs used in the treatment of substance related disorders. Literature review using a TDM appropriateness rating scale. *Ther Drug Monit* 2011 in press
- 100 Brunton LL, Lazo JS, Parker KL. Goodman and Gilman's The Pharmacological Basis of Therapeutics. 11th edn. McGraw-Hill, New York 2006
- 101 Buchanan RW, Kreyenbuhl J, Kelly DL et al. Schizophrenia Patient Outcomes Research Team (PORT). The 2009 schizophrenia PORT psychopharmacological treatment recommendations and summary statements. *Schizophr Bull* 2010; 36: 71–93
- 102 Buckley NA, Dawson AH, Whyte IM et al. Greater toxicity in overdose of dothiepin than of other tricyclic antidepressants. *Lancet* 1994; 343: 159–162
- 103 Burke MJ, Preskorn SH. Therapeutic drug monitoring of antidepressants – cost implications and relevance to clinical practice. *Clin Pharmacokinet* 1999; 37: 147–165
- 104 Byerly MJ, Thompson A, Carmody T et al. Validity of electronically monitored medication adherence and conventional adherence measures in schizophrenia. *Psychiatr Serv* 2007; 58: 844–847

- 105 Caccia S, Garattini S. Pharmacokinetic and pharmacodynamic significance of antidepressant drug metabolites. *Pharmacol Res* 1992; 26: 317–329
- 106 Caccia S, Pasia L, Nobili L. New atypical antipsychotics for schizophrenia: iloperidone. *Drug Des Devel Ther* 2010; 4: 33–48
- 107 Callaghan JT, Bergstrom RF, Ptak LR et al. Olanzapine. Pharmacokinetic and pharmacodynamic profile. *Clin Pharmacokinet* 1999; 37: 177–193
- 108 Canal M, Desanti CR, Santoni JP. A new oral formulation of tiapride (drops): pharmacokinetic profile and therapeutic applications. *Clin Drug Investig* 1998; 15: 455–460
- 109 Canal-Raffin M, Déridet E, Titier K et al. Simplified ultraviolet liquid chromatographic method for determination of sertindole, dehydro-sertindone and norsertindole, in human plasma. *J Chromatography B* 2005; 814: 61–67
- 110 Canal-Raffin M, Titier K, Déridet E et al. Myocardium distribution of sertindole and its metabolite dehydrosertindole in guinea-pigs. *Biopharm Drug Dispos* 2006; 27: 171–179
- 111 Carlsson B, Olsson G, Reis M et al. Enantioselective analysis of citalopram and metabolites in adolescents. *Ther Drug Monit* 2001; 23: 658–664
- 112 Castberg I, Skogvoll E, Spigset O. Quetiapine and drug interactions: evidence from a routine therapeutic drug monitoring service. *J Clin Psychiatry* 2007; 68: 1540–1545
- 113 Castberg I, Spigset O. Prescribing pattern and use of therapeutic drug monitoring of psychotropic medication in a psychiatric high-security unit. *Ther Drug Monit* 2008; 30: 597–603
- 114 Causon R. Validation of chromatographic methods in biomedical analysis. Viewpoint and discussion. *J Chromatogr B Biomed Sci Appl* 1997; 689: 175–180
- 115 Cawello W, Braun M, Boekens H. Absorption, disposition, metabolic fate, and elimination of the dopamine agonist rotigotine in man: administration by intravenous infusion or transdermal delivery. *Drug Metab Dispos* 2009; 37: 2055–2060
- 116 Centerholt C, Ekblom M, Odergren T et al. Pharmacokinetics and sedative effects in healthy subjects and subjects with impaired liver function after continuous infusion of clomethiazole. *Eur J Clin Pharmacol* 2003; 59: 117–122
- 117 Chakraborty BS, Midha KK, McKay G et al. Single dose kinetics of thioridazine and its two psychoactive metabolites in healthy humans: a dose proportionality study. *J Pharm Sci* 1989; 78: 796–801
- 118 Chan V, Morris RG, Ilett KF et al. Population pharmacokinetics of lamotrigine. *Ther Drug Monit* 2001; 23: 630–635
- 119 Chang YC, Lane HY, Yang KH et al. Optimizing early prediction for antipsychotic response in schizophrenia. *J Clin Psychopharmacol* 2006; 26: 554–559
- 120 Chawarski MC, Schottenfeld RS, O'Connor PG et al. Plasma concentrations of buprenorphine 24 to 72 hours after dosing. *Drug Alcohol Depend* 1999; 55: 157–163
- 121 Chen H, Grover S, Yu L et al. Bioactivation of lamotrigine in vivo in rat and in vitro in human liver microsomes, hepatocytes, and epidermal keratinocytes: characterization of thioether conjugates by liquid chromatography/mass spectrometry and high field nuclear magnetic resonance spectroscopy. *Chem Res Toxicol* 2010; 23: 159–170
- 122 Chen P, Tanasijevic MJ, Schoenenberger RA et al. A computer-based intervention for improving the appropriateness of antiepileptic drug level monitoring. *Am J Clin Pathol* 2003; 119: 432–438
- 123 Cheng YF, Paalzow LK, Bondesson U et al. Pharmacokinetics of haloperidol in psychotic patients. *Psychopharmacology (Berl)* 1987; 91: 410–414
- 124 Chen F, Batten LA, Zernig G et al. Comparison of pharmacokinetic profiles of brand-name and generic formulations of citalopram and venlafaxine: a crossover study. *J Clin Psychiatry* 2009; 70: 958–966
- 125 Chermá MD, Löfgren UB, Almkvist G et al. Assessment of the prescription of antidepressant drugs in elderly nursing home patients. *J Clin Psychopharmacol* 2008; 28: 424–431
- 126 Chermá MD, Reis M, Hägg S et al. Therapeutic drug monitoring of ziprasidone in a clinical treatment setting. *Ther Drug Monit* 2008; 30: 682–688
- 127 Chetty M, Gouws E, Miller R et al. The use of a side effect as a qualitative indicator of plasma chlorpromazine levels. *Eur Neuropsychopharmacol* 1999; 9: 77–82
- 128 Choc MG, Hsuan F, Honigfeld G et al. Single- vs. multiple-dose pharmacokinetics of clozapine in psychiatric patients. *Pharm Res* 1990; 7: 347–351
- 129 Choong E, Dobrin M, Carrupt PA et al. The permeability P-glycoprotein: a focus on enantioselectivity and brain distribution. *Expert Opin Drug Metab Toxicol* 2010; 6: 953–965
- 130 Ciraulo DA, Hitzemann RJ, Somoza E et al. Pharmacokinetics and pharmacodynamics of multiple sublingual buprenorphine tablets in dose-escalation trials. *J Clin Pharmacol* 2006; 46: 179–192
- 131 Citrome L. Paliperidone palmitate – review of the efficacy, safety and cost of a new second-generation depot antipsychotic medication. *Int J Clin Pract* 2010; 64: 216–239
- 132 Citrome L, Stauffer VL, Chen L et al. Olanzapine plasma concentrations after treatment with 10, 20, and 40 mg/d in patients with schizophrenia. *J Clin Psychopharmacol* 2009; 29: 278–283
- 133 Collins N, Barnes TR, Shingleton-Smith A et al. Standards of lithium monitoring in mental health trusts in the UK. *BMC Psychiatry* 2010; 10: 80
- 134 Conca A, Schmidt E, Pastore M et al. Therapeutic drug monitoring in Italian psychiatry. *Pharmacopsychiatry* 2011; 44: 259–262
- 135 Contin M, Riva R, Martinelli P et al. Effect of meal timing on the kinetic-dynamic profile of levodopa/carbidopa controlled release in parkinsonian patients. *Eur J Clin Pharmacol* 1998; 54: 303–308
- 136 Cooper TB. Plasma level monitoring of antipsychotic drugs. *Clin Pharmacokinet* 1978; 3: 14–38
- 137 Coppen A, Kopera H. Workshop on the clinical pharmacology and efficacy of mianserin. *Br J Clin Pharmacol* 1978; 5: 91S–99S
- 138 Correll CU, Malhotra AK, Kaushik S et al. Early prediction of antipsychotic response in schizophrenia. *Am J Psychiatry* 2003; 160: 2063–2065
- 139 Couchman L, Morgan PE, Spencer EP et al. Plasma clozapine and norclozapine in patients prescribed different brands of clozapine (Clozaril, Denzapine, and Zaponex). *Ther Drug Monit* 2010; 32: 624–627
- 140 Couchman L, Morgan PE, Spencer EP et al. Plasma clozapine, norclozapine, and the clozapine: norclozapine ratio in relation to prescribed dose and other factors: data from a therapeutic drug monitoring service, 1993–2007. *Ther Drug Monit* 2010; 32: 438–447
- 141 Coulomb F, Ducret F, Laneury JP et al. Pharmacokinetics of single-dose reboxetine in volunteers with renal insufficiency. *J Clin Pharmacol* 2000; 40: 482–487
- 142 Cournoyer G, De Montigny C, Ouellette J et al. A comparative double-blind controlled study of trimipramine and amitriptyline in major depression: lack of correlation with 5-hydroxytryptamine reuptake blockade. *J Clin Psychopharmacol* 1987; 7: 385–393
- 143 Court MH. Interindividual variability in hepatic drug glucuronidation: studies into the role of age, sex, enzyme inducers, and genetic polymorphism using the human liver bank as a model system. *Drug Metab Rev* 2010; 42: 202–217
- 144 Crettol S, Besson J, Croquette-Krokar M et al. Association of dopamine and opioid receptor genetic polymorphisms with response to methadone maintenance treatment. *Prog Neuropsychopharmacol Biol Psychiatry* 2008; 32: 1722–1727
- 145 Crettol S, Déglon JJ, Besson J et al. ABCB1 and cytochrome P450 genotypes and phenotypes: influence on methadone plasma levels and response to treatment. *Clin Pharmacol Ther* 2006; 80: 668–681
- 146 Crettol S, Déglon JJ, Besson J et al. Methadone enantiomer plasma levels, CYP2B6, CYP2C19, and CYP2C9 genotypes, and response to treatment. *Clin Pharmacol Ther* 2005; 78: 593–604
- 147 Cummings J, Lefèvre G, Small G et al. Pharmacokinetic rationale for the rivastigmine patch. *Neurology* 2007; 69 (Suppl 1): S10–S13
- 148 Dahl SG. Active metabolites of neuroleptic drugs: possible contribution to therapeutic and toxic effects. *Ther Drug Monit* 1982; 4: 33–40
- 149 Dahl SG, Strandjord RE, Sigfusson S. Pharmacokinetics and relative bioavailability of levomepromazine after repeated administration of tablets and syrup. *Eur J Clin Pharmacol* 1977; 11: 305–310
- 150 Darby JK, Pasta DJ, Wilson MG et al. Long-term therapeutic drug monitoring of risperidone and olanzapine identifies altered steady-state pharmacokinetics: a clinical, two-group, naturalistic study. *Clin Drug Investig* 2008; 28: 553–564
- 151 Daviss WB, Perel JM, Birmaher B et al. Steady-state clinical pharmacokinetics of bupropion extended-release in youths. *J Am Acad Child Adolesc Psychiatry* 2006; 45: 1503–1509
- 152 Daviss WB, Perel JM, Brent DA et al. Acute antidepressant response and plasma levels of bupropion and metabolites in a pediatric-aged sample: an exploratory study. *Ther Drug Monit* 2006; 28: 190–198
- 153 Dawling S. Monitoring of tricyclic antidepressant therapy. *Clin Biochem* 1982; 15: 56–61
- 154 de Lange EC. Potential role of ABC transporters as a detoxification system at the blood-CSF barrier. *Adv Drug Deliv Rev* 2004; 56: 1793–1809

- 155 *de Leon J*. Glucuronidation enzymes, genes and psychiatry. *Int J Neuropsychopharmacol* 2003; 6: 57–72
- 156 *de Leon J*. Incorporating pharmacogenetics into clinical practice: reality of a new tool in psychiatry. *Current issues in clinical implementation*. *CNS Spectr* 2006; 11 (Suppl 3): 8–12
- 157 *de Leon J*. The crucial role of the therapeutic window in understanding the clinical relevance of the poor versus the ultrarapid metabolizer phenotypes in subjects taking drugs metabolized by CYP2D6 or CYP2C19. *J Clin Psychopharmacol* 2007; 27: 241–245
- 158 *de Leon J, Greenlee B, Barber J et al*. Practical guidelines for the use of new generation antipsychotic drugs (except clozapine) in adult individuals with intellectual disabilities. *Res Dev Disabil* 2009; 30: 613–669
- 159 *de Leon J, Susce MT, Pan RM et al*. A study of genetic (CYP2D6 and ABCB1) and environmental (drug inhibitors and inducers) variables that may influence plasma risperidone levels. *Pharmacopsychiatry* 2007; 40: 93–102
- 160 *de Leon J, Susce MT, Pan RM et al*. The CYP2D6 poor metabolizer phenotype may be associated with risperidone adverse drug reactions and discontinuation. *J Clin Psychiatry* 2005; 66: 15–27
- 161 *de Leon J, Wynn G, Sandson NB*. The pharmacokinetics of paliperidone versus risperidone. *Psychosomatics* 2010; 51: 80–88
- 162 *de Mey C, Althaus M, Ezan E et al*. Erythromycin increases plasma concentrations of alpha-dihydroergocryptine in humans. *Clin Pharmacol Ther* 2001; 70: 142–148
- 163 *de Vries MH, Raghoobar M, Mathlener IS et al*. Single and multiple oral dose fluvoxamine kinetics in young and elderly subjects. *Ther Drug Monit* 1992; 14: 493–498
- 164 *de Wit M, Best AM, Epstein SK et al*. Lorazepam concentrations, pharmacokinetics and pharmacodynamics in a cohort of mechanically ventilated ICU patients. *Int J Clin Pharmacol Ther* 2006; 44: 466–473
- 165 *Degen J, Wölke E, Seiberling M et al*. Comparative study of the pharmacokinetics of amitriptyline oxide and trimipramine after single administration in healthy male probands and patients with renal failure. *Med Klin (Munich)* 1993; 88: 129–133
- 166 *Degner D, Grohmann R, Kropp S et al*. Severe adverse drug reactions of antidepressants: results of the German multicenter drug surveillance program AMSP. *Pharmacopsychiatry* 2004; 37 (Suppl 1): S39–S45
- 167 *Del Dotto P, Bonuccelli U*. Clinical pharmacokinetics of cabergoline. *Clin Pharmacokinet* 2003; 42: 633–645
- 168 *Deleu D, Northway MG, Hanssens Y*. Clinical pharmacokinetic and pharmacodynamic properties of drugs used in the treatment of Parkinson's disease. *Clin Pharmacokinet* 2002; 41: 261–309
- 169 *Deligiannidis KM*. Therapeutic drug monitoring in pregnant and postpartum women: recommendations for SSRIs, lamotrigine, and lithium. *J Clin Psychiatry* 2010; 71: 649–650
- 170 *Delva NJ, Hawken ER*. Preventing lithium intoxication. *Guide for physicians*. *Can Fam Physician* 2001; 47: 1595–1600
- 171 *Desta Z, Kerbusch T, Soukhova N et al*. Identification and characterization of human cytochrome P450 isoforms interacting with pimozone. *J Pharmacol Exp Ther* 1998; 285: 428–437
- 172 *Deuschle M, Härterter S, Hiemke C et al*. Doxepin and its metabolites in plasma and cerebrospinal fluid in depressed patients. *Psychopharmacology (Berl)* 1997; 131: 19–22
- 173 *DeVane CL, Boulton DW, Miller LF et al*. Pharmacokinetics of trazodone and its major metabolite m-chlorophenylpiperazine in plasma and brain of rats. *Int J Neuropsychopharmacol* 1999; 2: 17–23
- 174 *DeVane CL, Stowe ZN, Donovan JL et al*. Therapeutic drug monitoring of psychoactive drugs during pregnancy in the genomic era: challenges and opportunities. *J Psychopharmacol* 2006; 20 (Suppl): 54–59
- 175 *Diaz FJ, de Leon J, Josiassen RC et al*. Plasma clozapine concentration coefficients of variation in a long-term study. *Schizophr Res* 2005; 72: 131–135
- 176 *Diaz FJ, Santoro V, Spina E et al*. Estimating the size of the effects of co-medications on plasma clozapine concentrations using a model that controls for clozapine doses and confounding variables. *Pharmacopsychiatry* 2008; 41: 81–91
- 177 *Dingemans J, Jorga K, Zürcher G et al*. Multiple-dose clinical pharmacology of the catechol-O-methyl-transferase inhibitor tolcapone in elderly subjects. *Eur J Clin Pharmacol* 1996; 50: 47–55
- 178 *Dockens RC, Salazar DE, Fulmor IE et al*. Pharmacokinetics of a newly identified active metabolite of buspirone after administration of buspirone over its therapeutic dose range. *J Clin Pharmacol* 2006; 46: 1308–1312
- 179 *Doose DR, Walker SA, Gisclon LG et al*. Single-dose pharmacokinetics and effect of food on the bioavailability of topiramate, a novel antiepileptic drug. *J Clin Pharmacol* 1996; 36: 884–891
- 180 *Doran A, Obach RS, Smith BJ et al*. The impact of P-glycoprotein on the disposition of drugs targeted for indications of the central nervous system: evaluation using the MDR1A/1B knockout mouse model. *Drug Metab Dispos* 2005; 33: 165–174
- 181 *dos Santos FM, Gonçalves JC, Caminha R et al*. Pharmacokinetic/pharmacodynamic modeling of psychomotor impairment induced by oral clonazepam in healthy volunteers. *Ther Drug Monit* 2009; 31: 566–574
- 182 *Dunbar JL, Turncliff RZ, Hayes SC et al*. Population pharmacokinetics of extended-release injectable naltrexone (XR-NTX) in patients with alcohol dependence. *J Stud Alcohol Drugs* 2007; 68: 862–870
- 183 *Duthheil F, Jacob A, Dauchy S et al*. ABC transporters and cytochromes P450 in the human central nervous system: influence on brain pharmacokinetics and contribution to neurodegenerative disorders. *Expert Opin Drug Metab Toxicol* 2010; 6: 1161–1174
- 184 *Dvorchik BH, Vesell ES*. Pharmacokinetic interpretation of data gathered during therapeutic drug monitoring. *Clin Chem* 1976; 22: 868–878
- 185 *Eagles JM, McCann I, MacLeod TN et al*. Lithium monitoring before and after the distribution of clinical practice guidelines. *Acta Psychiatr Scand* 2000; 101: 349–353
- 186 *Eap CB, Baumann P*. Analytical methods for the quantitative determination of selective serotonin reuptake inhibitors for therapeutic drug monitoring purposes in patients. *J Chromatogr B Biomed Appl* 1996; 686: 51–63
- 187 *Eap CB, Bender S, Gastpar M et al*. Steady state plasma levels of the enantiomers of trimipramine and of its metabolites in CYP2D6-, CYP2C19- and CYP3A4/5-phenotyped patients. *Ther Drug Monit* 2000; 22: 209–214
- 188 *Eap CB, Bertschy G, Baumann P et al*. High interindividual variability of methadone enantiomer blood levels to dose ratios. *Arch Gen Psychiatry* 1998; 55: 89–90
- 189 *Eap CB, Buclin T, Baumann P*. Interindividual variability of the clinical pharmacokinetics of methadone: implications for the treatment of opioid dependence. *Clin Pharmacokinet* 2002; 41: 1153–1193
- 190 *Eap CB, Finkbeiner T, Gastpar M et al*. Replacement of R-methadone by a double dose of (R,S)-methadone in addicts: interindividual variability of the R/S ratios and evidence of adaptive changes in methadone pharmacokinetics. *Eur J Clin Pharmacol* 1996; 50: 385–389
- 191 *Eap CB, Koeb L, Baumann P*. Determination of trimipramine and its demethylated and hydroxylated metabolites in plasma by gas chromatography-mass spectrometry. *J Chromatogr* 1994; 652: 97–103
- 192 *Eap CB, Lima CA, Macciardi F et al*. Steady state concentrations of the enantiomers of mianserin and desmethylmianserin in poor and in homozygous and heterozygous extensive metabolizers of debrisoquine. *Ther Drug Monit* 1998; 20: 7–13
- 193 *Eichelbaum M, Ingelman-Sundberg M, Evans WE*. Pharmacogenomics and individualized drug therapy. *Annu Rev Med* 2006; 57: 119–137
- 194 *Egberts K, Mehler-Wex C, Gerlach M*. 2011; Therapeutic drug monitoring in child and adolescent psychiatry. *Pharmacopsychiatry* 2011; 44: 249–253
- 195 *Eggert Hansen C, Rosted Christensen T, Elley J et al*. Clinical pharmacokinetic studies of perphenazine. *Br J Clin Pharmacol* 1976; 3: 915–923
- 196 *Ellinwood EHJR, Heatherly DG, Nikaido AM et al*. Comparative pharmacokinetics and pharmacodynamics of lorazepam, alprazolam and diazepam. *Psychopharmacology* 1985; 86: 392–399
- 197 *Ereshefsky L, Jann MW, Saklad SR et al*. Effects of smoking on fluphenazine clearance in psychiatric inpatients. *Biol Psychiatry* 1985; 20: 329–332
- 198 *Evans LE, Bett JH, Cox JR et al*. The bioavailability of oral and parenteral chlorimipramine (Anafranil). *Prog Neuropsychopharmacol* 1980; 4: 293–302
- 199 *Evans WE, Relling MV*. Pharmacogenomics: translating functional genomics into rational therapeutics. *Science* 1999; 286: 487–491
- 200 *Faber MS, Fuhr U*. Time response of cytochrome P450 1A2 activity on cessation of heavy smoking. *Clin Pharmacol Ther* 2004; 76: 178–184
- 201 *Faber MS, Jetter A, Fuhr U*. Assessment of CYP1A2 activity in clinical practice: why, how, and when? *Basic Clin Pharmacol Toxicol* 2005; 97: 125–134
- 202 *Faessel HM, Gibbs MA, Clarc DJ et al*. Multiple-doses pharmacokinetics of the selective nicotinic receptor partial agonist, varenicline, in healthy smokers. *J Clin Pharmacol* 2006; 46: 1439–1448

- 203 Faiman MD, Jensen JC, Lacoursiere RB. Elimination kinetics of disulfiram in alcoholics after single and repeated doses. *Clin Pharmacol Ther* 1984; 36: 520–526
- 204 Falkai P, Wobrock T, Lieberman J et al. World Federation of Societies of Biological Psychiatry (WFSBP) guidelines for biological treatment of schizophrenia, Part 1: acute treatment of schizophrenia. *World J Biol Psychiatry* 2005; 6: 132–191
- 205 Falkai P, Wobrock T, Lieberman J et al. World Federation of Societies of Biological Psychiatry (WFSBP) guidelines for biological treatment of schizophrenia, part 2: long-term treatment of schizophrenia. *World J Biol Psychiatry* 2006; 7: 5–40
- 206 Fanton L, Bévalot F, Grait H et al. Fatal intoxication with milnacipran. *J Forensic Leg Med* 2008; 15: 388–390
- 207 Farde L, Nordström AL, Wiesel FA et al. Positron emission tomographic analysis of central D1 and D2 dopamine receptor occupancy in patients treated with classical neuroleptics and clozapine. *Arch Gen Psychiatry* 1992; 49: 538–544
- 208 Feng Y, Pollock BG, Coley K et al. Population pharmacokinetic analysis for risperidone using highly sparse sampling measurements from the CATIE study. *J Clin Pharmacol* 2008; 66: 629–639
- 209 Feng Y, Pollock BG, Ferrell RE et al. Paroxetine: population pharmacokinetic analysis in late-life depression using sparse concentration sampling. *Br J Clin Pharmacol* 2006; 61: 558–569
- 210 Fenner KS, Troutman MD, Kempshall S et al. Drug-drug interactions mediated through P-glycoprotein: clinical relevance and in vitro-in vivo correlation using digoxin as a probe drug. *Clin Pharmacol Ther* 2009; 85: 173–181
- 211 Ferrari A, Bertolotti M, Dell Utri A et al. Serum time course of naltrexone and 6 β -naltrexol levels during long term treatment in drug addicts. *Drug Alcohol Depend* 1998; 52: 211–220
- 212 Figueroa C, Brecher M, Hamer-Maansson JE et al. Pharmacokinetic profiles of extended release quetiapine fumarate compared with quetiapine immediate release. *Prog Neuropsychopharmacol Biol Psychiatry* 2009; 33: 199–204
- 213 Findling RL, Reed MD, Myers C et al. Paroxetine pharmacokinetics in depressed children and adolescents. *J Am Acad Child Adolesc Psychiatry* 1999; 38: 952–959
- 214 Fitzgerald PB, Kapur S, Remington G et al. Predicting haloperidol occupancy of central dopamine D2 receptors from plasma levels. *Psychopharmacology (Berl)* 2000; 149: 1–5
- 215 Flanagan RJ. Developing an analytical toxicology service: principles and guidance. *Toxicol Rev* 2004; 23: 251–263
- 216 Fleming J, Chetty M. Therapeutic monitoring of valproate in psychiatry: how far have we progressed? *Clin Neuropharmacol* 2006; 29: 350–360
- 217 Fogelman SM, Schmider J, Venkatakrishnan K et al. O- and N-demethylation of venlafaxine in vitro by human liver microsomes and by microsomes from cDNA-transfected cells: effect of metabolic inhibitors and SSRI antidepressants. *Neuropsychopharmacology* 1999; 20: 480–490
- 218 Fontaine R, Mercier P, Beaudry P et al. Bromazepam and lorazepam in generalized anxiety: a placebo-controlled study with measurement of drug plasma concentrations. *Acta Psychiatr Scand* 1986; 74: 451–458
- 219 Food and Drug Administration. Guidance for industry: bioanalytical method validation. 2001; <http://www.fda.gov/cvm>
- 220 Foti RS, Rock DA, Wienkers LC et al. Selection of alternative CYP3A4 probe substrates for clinical drug interaction studies using in vitro data and in vivo simulation. *Drug Metab Dispos* 2010; 38: 981–987
- 221 Fountoulakis KN. An update of evidence-based treatment of bipolar depression: where do we stand? *Curr Opin Psychiatry* 2010; 23: 19–24
- 222 Fric M, Pfuhlmann B, Laux G et al. The influence of smoking on the serum level of duloxetine. *Pharmacopsychiatry* 2008; 41: 151–155
- 223 Frieboes RM, Sonntag A, Yassouridis A et al. Clinical outcome after trimipramine in patients with delusional depression – a pilot study. *Pharmacopsychiatry* 2003; 36: 12–17
- 224 Friedman H, Greenblatt DJ, Peters GR et al. Pharmacokinetics and pharmacodynamics of oral diazepam: effect of dose, plasma concentration, and time. *Clin Pharmacol Ther* 1992; 52: 139–150
- 225 Fritze J, Laux G, Sofic E et al. Plasma moclobemide and metabolites: lack of correlation with clinical response and biogenic amines. *Psychopharmacology (Berl)* 1989; 99: 252–256
- 226 Fröscher W, Schier KR, Hoffmann M et al. Topiramate: a prospective study on the relationship between concentration, dosage and adverse events in epileptic patients on combination therapy. *Epileptic Disord* 2005; 7: 237–248
- 227 Fudio S, Borobia AM, Piñana E et al. Evaluation of the influence of sex and CYP2C19 and CYP2D6 polymorphisms in the disposition of citalopram. *Eur J Pharmacol* 2010; 626: 200–204
- 228 Fukasawa T, Suzuki A, Otani K. Effects of genetic polymorphism of cytochrome P450 enzymes on the pharmacokinetics of benzodiazepines. *J Clin Pharm Ther* 2007; 32: 333–341
- 229 Furlanut M, Montanari G, Benetello P et al. Steady-state serum concentrations of imipramine, its main metabolites and clinical response in primary enuresis. *Pharmacol Res* 1989; 21: 561–566
- 230 Furukori H, Kondo T, Yasui N et al. Effects of itraconazole on the steady-state plasma concentrations of bromperidol and reduced bromperidol in schizophrenic patients. *Psychopharmacology (Berl)* 1999; 145: 189–192
- 231 Gaertner HJ, Golfinopoulos G, Breyer-Pfaff U. Response to Maprotiline treatment in depressive patients, relationship to urinary MHPG excretion, and plasma drug level. *Pharmacopsychiatry* 1982; 15: 170–174
- 232 Gaertner I, Gaertner HJ, Vonthein R et al. Therapeutic drug monitoring of clozapine in relapse prevention: a five-year prospective study. *J Clin Psychopharmacol* 2001; 21: 305–310
- 233 Garnock-Jones KP, Keating GM. Atomoxetine: a review of its use in attention-deficit hyperactivity disorder in children and adolescents. *Paediatr Drugs* 2009; 11: 203–226
- 234 Gelenberg AJ. A review of the current guidelines for depression treatment. *J Clin Psychiatry* 2010; 71: e15
- 235 Genton P, Guerrini R, Perucca E. Tiagabine in clinical practice. *Epilepsia* 2001; 42 (Suppl 3): 42–45
- 236 Gerlach M, Hünnerkopf R, Rothenhöfer S et al. Therapeutic drug monitoring of quetiapine in adolescents with psychotic disorders. *Pharmacopsychiatry* 2007; 40: 72–76
- 237 Gerstenberg G, Aoshima T, Fukasawa T et al. Relationship between clinical effects of fluvoxamine and the steady-state plasma concentrations of fluvoxamine and its major metabolite fluvoxaminic acid in Japanese depressed patients. *Psychopharmacology (Berl)* 2003; 167: 443–448
- 238 Gervasini G, Carrillo JA, Benitez J. Potential role of cerebral cytochrome P450 in clinical pharmacokinetics: modulation by endogenous compounds. *Clin Pharmacokinet* 2004; 43: 693–706
- 239 Gex-Fabry M, Balant-Gorgia AE, Balant LP. Clomipramine concentration as a predictor of delayed response: a naturalistic study. *Eur J Clin Pharmacol* 1999; 54: 895–902
- 240 Gex-Fabry M, Balant-Gorgia AE, Balant LP. Therapeutic drug monitoring of olanzapine: the combined effect of age, gender, smoking, and comedication. *Ther Drug Monit* 2003; 25: 46–53
- 241 Gex-Fabry M, Balant-Gorgia AE, Balant LP et al. Time course of clinical response to venlafaxine: relevance of plasma level and chirality. *Eur J Clin Pharmacol* 2004; 59: 883–891
- 242 Gex-Fabry M, Gervasoni N, Eap CB et al. Time course of response to paroxetine: influence of plasma level. *Prog Neuropsychopharmacol Biol Psychiatry* 2007; 31: 892–900
- 243 Gilles M, Deuschle M, Kellner S et al. Paroxetine serum concentrations in depressed patients and response to treatment. *Pharmacopsychiatry* 2005; 38: 118–121
- 244 Gillman PK. Tricyclic antidepressant pharmacology and therapeutic drug interactions updated. *Br J Pharmacol* 2007; 151: 737–748
- 245 Glassman AH, Perel JM, Shostak M et al. Clinical implications of imipramine plasma levels for depressive illness. *Arch Gen Psychiatry* 1977; 34: 197–204
- 246 Glassmann AH, Schildkraut JJ, Orsulak PJ et al. Tricyclic antidepressants, blood level measurements and clinical outcome: an APA task force report. *Am J Psychiatr* 1985; 142: 155–162
- 247 Glauser TA, Pippenger CE. Controversies in blood-level monitoring: reexamining its role in the treatment of epilepsy. *Epilepsia* 2000; 41 (Suppl 8): S6–S15
- 248 Gleason MM, Egger HL, Emslie GJ et al. Psychopharmacological treatment for very young children: contexts and guidelines. *J Am Acad Child Adolesc Psychiatry* 2007; 46: 1532–1572
- 249 Glotzbach RK, Preskorn SH. Brain concentrations of tricyclic antidepressants: Single-dose kinetics and relationship to plasma concentrations in chronically dosed rats. *Psychopharmacology* 1982; 78: 25–27
- 250 Goeringer KE, Raymon L, Christian GD et al. Postmortem forensic toxicology of selective serotonin reuptake inhibitors: a review of pharmacology and report of 168 cases. *J Forensic Sci* 2000; 45: 633–648
- 251 Gomolin IH, Smith C, Jeitner TM. Once-daily memantine: pharmacokinetic and clinical considerations. *J Am Geriatr Soc* 2010; 58: 1812–1813

- 252 Gonzalez JP, Brogden RN. Naltrexone. A review of its pharmacodynamic and pharmacokinetic properties and therapeutic efficacy in the management of opioid dependence. *Drugs* 1988; 35: 192–213
- 253 Goodnick PJ, Dominguez RA, DeVane CL et al. Bupropion slow-release response in depression: diagnosis and biochemistry. *Biol Psychiatry* 1998; 44: 629–632
- 254 Goodwin GM. Consensus Group of the British Association for Psychopharmacology. Evidence-based guidelines for treating bipolar disorder: revised second edition-recommendations from the British Association for Psychopharmacology. *J Psychopharmacol* 2009; 23: 346–388
- 255 Gram LF, Guentert TW, Grange S et al. Moclobemide, a substrate of CYP2C19 and an inhibitor of CYP2C19, CYP2D6, and CYP1A2: a panel study. *Clin Pharmacol Ther* 1995; 57: 670–677
- 256 Grandjean EM, Aubry JM. Lithium: updated human knowledge using an evidence-based approach. Part II: Clinical pharmacology and therapeutic monitoring. *CNS Drugs* 2009; 23: 331–349
- 257 Grasmäder K, Verwohlt PL, Kühn KU et al. Relationship between mirtazapine dose, plasma concentration, response, and side effects in clinical practice. *Pharmacopsychiatry* 2005; 38: 113–117
- 258 Grasmäder K, Verwohlt PL, Rietschel M et al. Impact of polymorphisms of cytochrome-P450 isoenzymes 2C9, 2C19 and 2D6 on plasma concentrations and clinical effects of antidepressants in a naturalistic clinical setting. *Eur J Clin Pharmacol* 2004; 60: 329–336
- 259 Greenblatt DJ, Blaskovich PD, Nuwaysir ES et al. Clonazepam pharmacokinetics: comparison of subcutaneous microsphere injection with multiple-dose oral administration. *J Clin Pharmacol* 2005; 45: 1288–1293
- 260 Greenblatt DJ, Divoll M, Harmatz JS et al. Oxazepam kinetics: effects of age and sex. *J Pharmacol Exp Ther* 1980; 215: 86–91
- 261 Greenblatt DJ, Ehrenberg BL, Gunderman J et al. Pharmacokinetic and electroencephalographic study of intravenous diazepam, midazolam, and placebo. *Clin Pharmacol Ther* 1989; 45: 356–365
- 262 Greenblatt DJ, Friedman H, Burstein ES et al. Trazodone kinetics: effect of age, gender, and obesity. *Clin Pharmacol Ther* 1987; 42: 193–200
- 263 Greenblatt DJ, Gan L, Harmatz JS et al. Pharmacokinetics and pharmacodynamics of single-dose triazolam: electroencephalography compared with the Digit-Symbol Substitution Test. *Br J Clin Pharmacol* 2005; 60: 244–248
- 264 Greenblatt DJ, Harmatz JS, Friedman H et al. A large-sample study of diazepam pharmacokinetics. *Ther Drug Monit* 1989; 11: 652–657
- 265 Greenblatt DJ, Harmatz JS, von Moltke LL et al. Comparative kinetics and dynamics of zaleplon, zolpidem, and placebo. *Clin Pharmacol Ther* 1998; 64: 553–561
- 266 Greenblatt DJ, Shader RI, Franke K et al. Pharmacokinetics and bioavailability of intravenous, intramuscular, and oral lorazepam in humans. *J Pharm Sci* 1979; 68: 57–63
- 267 Greenblatt DJ, von Moltke LL, Ehrenberg BL et al. Kinetics and dynamics of lorazepam during and after continuous intravenous infusion. *Crit Care Med* 2000; 28: 2750–2757
- 268 Greenblatt DJ, von Moltke LL, Harmatz JS et al. Short-term exposure to low-dose ritonavir impairs clearance and enhances adverse effects of trazodone. *J Clin Pharmacol* 2003; 43: 414–422
- 269 Greiner C, Hiemke C, Bader W et al. Determination of citalopram and escitalopram together with their active main metabolites desmethyl(es)-citalopram in human serum by column-switching high performance liquid chromatography (HPLC) and spectrophotometric detection. *J Chromatogr B Analyt Technol Biomed Life Sci* 2007; 848: 391–394
- 270 Grimaldi R, Perucca E, Ruberto G et al. Pharmacokinetic and pharmacodynamic studies following the intravenous and oral administration of the antiparkinsonian drug biperiden to normal subjects. *Eur J Clin Pharmacol* 1986; 29: 735–737
- 271 Grohmann R, Engel RR, Rütther E et al. The AMSP drug safety program: methods and global results. *Pharmacopsychiatry* 2004; 37 (Suppl 1): S4–S11
- 272 Gründer G, Carlsson A, Wong DF. Mechanism of new antipsychotic medications. Occupancy is not just antagonism. *Arch Gen Psychiatry* 2003; 60: 974–977
- 273 Gründer G, Fellows C, Janouschek H et al. Brain and plasma pharmacokinetics of aripiprazole in patients with schizophrenia: an [18F] fallypride PET study. *Am J Psychiatry* 2008; 165: 988–995
- 274 Gründer G, Hiemke C, Paulzen M et al. Therapeutic drug concentrations of antidepressants and antipsychotics: Guidance from PET imaging. *Pharmacopsychiatry* 2011; 44: 236–248
- 275 Gründer G, Hippus H, Carlsson A. The 'atypicality' of antipsychotics: a concept re-examined and re-defined. *Nat Rev Drug Discov* 2009; 8: 197–202
- 276 Grunze H, Vieta E, Goodwin GM. The World Federation of Societies of Biological Psychiatry (WFSBP) guidelines for the biological treatment of bipolar disorders: update 2009 on the treatment of acute mania. *World J Biol Psychiatry* 2009; 10: 85–116
- 277 Guay DR. Rasagiline (TVP-1012): a new selective monoamine oxidase inhibitor for Parkinson's disease. *Am J Geriatr Pharmacother* 2006; 4: 330–346
- 278 Guberman A, Couture M, Blaschuk K et al. Add-on trial of clobazam in intractable adult epilepsy with plasma level correlations. *Can J Neurol Sci* 1990; 17: 311–316
- 279 Gunes A, Spina E, Dahl ML et al. ABCB1 polymorphisms influence steady-state plasma levels of 9-hydroxyrisperidone and risperidone active moiety. *Ther Drug Monit* 2008; 30: 628–633
- 280 Gupta N. Guidelines for lithium monitoring: are they ideal? *Acta Psychiatr Scand* 2001; 104: 76–77
- 281 Gupta RN, Dziurdzy SA. Therapeutic monitoring of sertraline. *Clin Chem* 1994; 40: 498–499
- 282 Gupta SK, Shah JC, Hwang SS. Pharmacokinetic and pharmacodynamic characterization of OROS and immediate-release amitriptyline. *Br J Clin Pharmacol* 1999; 48: 71–78
- 283 Guy W. editor. ECDEU Assessment Manual for Psychopharmacology. Rockville, MD. U.S. Department of Health, Education, and Welfare, 1976
- 284 Haberstroh J, Hampel H, Pantel J. Optimal management of Alzheimer's disease patients: Clinical guidelines and family advice. *Neuropsychiatr Dis Treat* 2010; 6: 243–253
- 285 Haen E, Greiner C, Bader W et al. Wirkstoffkonzentrationsbestimmungen zur Therapieleitung. Ergänzung therapeutischer Referenzbereiche durch dosisbezogene Referenzbereiche. *Nervenarzt* 2008; 79: 558–566
- 286 Hallett C, Dean BC. Bromazepam: acute benefit-risk assessment in general practice. *Curr Med Res Opin* 1984; 8: 683–688
- 287 Hammarberg A, Beck O, Eksborg S et al. Acamprosate determinations in plasma and cerebrospinal fluid after multiple dosing measured by liquid chromatography-mass spectroscopy: a pharmacokinetic study in healthy volunteers. *Ther Drug Monit* 2010; 32: 489–496
- 288 Hammarberg A, Jayaram-Lindström N, Berck O et al. The effects of acamprosate on alcohol-cue reactivity and alcohol priming in dependent patients: a randomized controlled trial. *Psychopharmacol* 2009; 205: 53–62
- 289 Hanley JA, McNeil BJ. The meaning and use of the area under a receiver operating characteristic (ROC) curve. *Radiology* 1982; 143: 29–36
- 290 Harden CL, Trifiletti R, Kutt H. Felbamate levels in patients with epilepsy. *Epilepsia* 1996; 37: 280–283
- 291 Härtter S, Dingemans J, Baier D et al. The role of cytochrome P450 2D6 in the metabolism of moclobemide. *Eur Neuropsychopharmacol* 1996; 6: 225–230
- 292 Härtter S, Hermes B, Hiemke C. Automated determination of trimipramine and N-desmethyl-trimipramine in human plasma or serum by HPLC with on-line solid phase extraction. *J Liq Chromatogr* 1995; 18: 3495–3505
- 293 Härtter S, Hermes B, Szegedi A et al. Automated determination of paroxetine and its main metabolite by column switching and on-line high-performance liquid chromatography. *Ther Drug Monit* 1994; 16: 400–406
- 294 Härtter S, Hiemke C. Column switching and high-performance liquid chromatography in the analysis of amitriptyline, nortriptyline and hydroxylated metabolites in human plasma or serum. *J Chromatogr* 1992; 578: 273–282
- 295 Härtter S, Tybring G, Friedberg T et al. The N-demethylation of the doxepin isomers is mainly catalyzed by the polymorphic CYP2C19. *Pharm Res* 2002; 19: 1034–1037
- 296 Härtter S, Wang X, Weigmann H et al. Differential effects of fluvoxamine and other antidepressants on the biotransformation of melatonin. *J Clin Psychopharmacol* 2001; 21: 167–174
- 297 Härtter S, Weigmann H, Hiemke C. Automated determination of reboxetine by high-performance liquid chromatography with column-switching and ultraviolet detection. *J Chromatogr B Biomed Sci Appl* 2000; 740: 135–140
- 298 Härtter S, Wetzel H, Hiemke C. Automated determination of fluvoxamine in plasma by column-switching high-performance liquid chromatography. *Clin Chem* 1992; 38: 2082–2086
- 299 Hasselström J, Linnet K. Quetiapine serum concentrations in psychiatric patients: the influence of comedication. *Ther Drug Monit* 2004; 26: 486–491

- 300 Hässler F, Reis O. Pharmacotherapy of disruptive behavior in mentally retarded subjects: A review of the current literature. *Dev Disabil Res Rev* 2010; 16: 265–272
- 301 Haymond J, Ensom MH. Does valproic acid warrant therapeutic drug monitoring in bipolar affective disorder? *Ther Drug Monit* 2010; 32: 19–29
- 302 Hazell P, Becker K, Nikkanen EA *et al.* Relationship between atomoxetine plasma concentration, treatment response and tolerability in attention-deficit/hyperactivity disorder and comorbid oppositional defiant disorder. *Atten Defic Hyperact Disord* 2009; 1: 201–210
- 303 Hegerl U, Bottlender R, Gallinat J *et al.* The serotonin syndrome scale: first results on validity. *Eur Arch Psychiatry Clin Neurosci* 1998; 248: 96–103
- 304 Heikkinen H, Saraheimo M, Antila S *et al.* Pharmacokinetics of entacapone, a peripherally acting catechol-O-methyltransferase inhibitor, in man. A study using a stable isotope technique. *Eur J Clin Pharmacol* 2001; 56: 821–826
- 305 Heller S, Hiemke C, Stroba G *et al.* Assessment of storage and transport stability of new antidepressant and antipsychotic drugs for a nationwide TDM service. *Ther Drug Monit* 2004; 26: 459–461
- 306 Hensset M, Hermann M, Lunde H *et al.* Impact of the CYP2D6 genotype on steady-state serum concentrations of aripiprazole and dehydroaripiprazole. *Eur J Clin Pharmacol* 2007; 63: 1147–1151
- 307 Herman BD, Fleishaker JC, Brown MT. Ketoconazole inhibits the clearance of the enantiomers of the antidepressant reboxetine in humans. *Clin Pharmacol Ther* 1999; 66: 374–379
- 308 Hermida J, Paz E, Tutor JC. Clozapine and norclozapine concentrations in serum and plasma samples from schizophrenic patients. *Ther Drug Monit* 2008; 30: 41–45
- 309 Hesse LM, He P, Krishnaswamy S *et al.* Pharmacogenetic determinants of interindividual variability in bupropion hydroxylation by cytochrome P450 2B6 in human liver microsomes. *Pharmacogenetics* 2004; 14: 225–238
- 310 Hiemke C. Clinical utility of drug measurement and pharmacokinetics – therapeutic drug monitoring in psychiatry. *Eur J Clin Pharmacol* 2008; 64: 159–166
- 311 Hiemke C. Therapeutic drug monitoring in neuropharmacology: does it hold its promises? *Eur Arch Psychiatry Clin Neurosci* 2008; 258 (Suppl 1): 21–27
- 312 Hiemke C, Baumann P, Laux G *et al.* Therapeutisches Drug-Monitoring in der Psychiatrie. Konsensus-Leitlinie der AGNP. *Psychopharmakotherapie* 2005; 12: 166–182
- 313 Hiemke C, Dragicevic A, Gründer G *et al.* Therapeutic monitoring of new antipsychotic drugs. *Ther Drug Monit* 2004; 26: 156–160
- 314 Hiemke C, Härtter S. Pharmacokinetics of selective serotonin reuptake inhibitors. *Pharmacol Ther* 2000; 85: 11–28
- 315 Higuchi H, Yoshida K, Takahashi H *et al.* Milnacipran plasma levels and antidepressant response in Japanese major depressive patients. *Hum Psychopharmacol* 2003; 18: 255–259
- 316 Höjer J, Hulting J, Salmonson H. Fatal cardiotoxicity induced by venlafaxine overdose. *Clin Toxicol (Phila)* 2008; 46: 336–337
- 317 Holbrook JM, Parks-Veal P, Mimbs J. Clinical monitoring guidelines for neuroleptic and antidepressant drugs. Central State Hospital, Milledgeville, Georgia. *Hosp Pharm* 1991; 26: 783–784, 787–793
- 318 Holzer L, Preuss U, Baumgartner L *et al.* Quetiapine in adolescents with non-affective psychotic disorders: An open-label trial. *Pharmacopsychiatry* 2011; 44: 87–95
- 319 Hooper WD, Dickinson RG, Dunstan PR *et al.* Oxcarbazepine: preliminary clinical and pharmacokinetic studies on a new anticonvulsant. *Clin Exp Neurol* 1987; 24: 105–112
- 320 Horadam VW, Sharp JG, Smilack JD *et al.* Pharmacokinetics of amantadine hydrochloride in subjects with normal and impaired renal function. *Ann Intern Med* 1981; 94: 454–458
- 321 Hrdina PD, Lapierre YD. Plasma levels of maprotiline and zimelidine and their relationship to clinical response in depressed patients. *Ther Drug Monit* 1986; 8: 400–406
- 322 Hsieh YH, Yang YH, Yeh HH *et al.* Simultaneous determination of galantamine, rivastigmine and NAP 226-90 in plasma by MEKC and its application in Alzheimer's disease. *Electrophoresis* 2009; 30: 644–653
- 323 Hughes J, Gill AM, Mulhearn H *et al.* Steady-state plasma concentrations of midazolam in critically ill infants and children. *Ann Pharmacother* 1996; 30: 27–30
- 324 Hui WK, Mitchell LB, Kavanagh KM *et al.* Melperone: electrophysiologic and antiarrhythmic activity in humans. *J Cardiovasc Pharmacol* 1990; 15: 144–149
- 325 Ilett KF, Blythe TH, Hackett LP *et al.* Plasma concentrations of dothiepin and its metabolites are not correlated with clinical efficacy in major depressive illness. *Ther Drug Monit* 1993; 15: 351–357
- 326 Isacson G, Holmgren P, Druid H *et al.* The utilization of antidepressants – a key issue in the prevention of suicide: an analysis of 5 281 suicides in Sweden during the period 1992–1994. *Acta Psychiatr Scand* 1997; 96: 94–100
- 327 Isbister GK, Hackett LP, Dawson AH *et al.* Moclobemide poisoning: toxicokinetics and occurrence of serotonin toxicity. *Br J Clin Pharmacol* 2003; 56: 441–450
- 328 Ishida M, Otani K, Kaneko S *et al.* Effects of various factors on steady state plasma concentrations of trazodone and its active metabolite m-chlorophenylpiperazine. *Int Clin Psychopharmacol* 1995; 10: 143–146
- 329 Iwersen S, Schmoldt A. One fatal and one nonfatal intoxication with tranlycypromine. Absence of amphetamines as metabolites. *J Anal Toxicol* 1996; 20: 301–304
- 330 Jaanson P, Marandi T, Kiivet RA *et al.* Maintenance therapy with zuclopentixol decanoate: associations between plasma concentrations, neurological side effects and CYP2D6 genotype. *Psychopharmacology (Berl)* 2002; 162: 67–73
- 331 Janis GC, Markowitz JS. Influence of ethanol and gender on methylphenidate pharmacokinetics and pharmacodynamics. *Clin Pharmacol Ther* 2007; 81: 346–353
- 332 Jann MW, Grimsley SR, Gray EC *et al.* Pharmacokinetics and pharmacodynamics of clozapine. *Clin Pharmacokinet* 1993; 24: 161–176
- 333 Jann MW, Shirley KL, Small GW. Clinical pharmacokinetics and pharmacodynamics of cholinesterase inhibitors. *Clin Pharmacokinet* 2002; 41: 719–739
- 334 Jaquenoud Sirot E, Knezevic B, Morena GP *et al.* ABCB1 and cytochrome P450 polymorphisms: clinical pharmacogenetics of clozapine. *J Clin Psychopharmacol* 2009; 29: 319–326
- 335 Jaquenoud Sirot E, van der Velden JW, Rentsch K *et al.* Therapeutic drug monitoring and pharmacokinetic rests as tools in pharmacovigilance. *Drug Safety* 2006; 29: 735–768
- 336 Jefferson JW, Pradko JF, Muir KT. Bupropion for major depressive disorder: pharmacokinetic and formulation considerations. *Clin Therap* 2005; 27: 1685–1695
- 337 Jerling M, Dahl ML, Aberg-Wistedt A *et al.* The CYP2D6 genotype predicts the oral clearance of the neuroleptic agents perphenazine and zuclopentixol. *Clin Pharmacol Ther* 1996; 59: 423–428
- 338 Ji P, Damle B, Xie J *et al.* Pharmacokinetic interaction between efavirenz and carbamazepine after multiple-dose administration in healthy subjects. *J Clin Pharmacol* 2008; 48: 948–956
- 339 Jimmink A, Caminada K, Hunfeld NG *et al.* Clinical toxicology of citalopram after acute intoxication with the sole drug or in combination with other drugs: overview of 26 cases. *Ther Drug Monit* 2008; 30: 365–371
- 340 Jin Y, Pollock BG, Frank E *et al.* The effect of reporting methods for dosing times on the estimation of pharmacokinetic parameters of escitalopram. *J Clin Pharmacol* 2009; 49: 176–184
- 341 Jochemsen R, Wesselman JG, Hermans J *et al.* Pharmacokinetics of brotizolam in healthy subjects following intravenous and oral administration. *Br J Clin Pharmacol* 1983; 16 (Suppl 2): 285S–290S
- 342 Johannessen SI, Battino D, Berry DJ *et al.* Therapeutic drug monitoring of the newer antiepileptic drugs. *Ther Drug Monitor* 2003; 25: 347–363
- 343 Johannessen SI, Tomson T. Pharmacokinetic variability of newer antiepileptic drugs: when is monitoring needed? *Clin Pharmacokinet* 2006; 45: 1061–1075
- 344 Johansson B. A review of the pharmacokinetics and pharmacodynamics of disulfiram and its metabolites. *Acta Psychiatr Scand Suppl* 1992; 369: 15–26
- 345 Johnston JA, Fiedler-Kelly J, Glover ED *et al.* Relationship between drug exposure and the efficacy and safety of bupropion sustained release for smoking cessation. *Nicotine Tob Res* 2001; 3: 131–140
- 346 Jorga KM, Fotteler B, Heizmann P *et al.* Pharmacokinetics and pharmacodynamics after oral and intravenous administration of tolcapone, a novel adjunct to Parkinson's disease therapy. *Eur J Clin Pharmacol* 1998; 54: 443–447
- 347 Jorga KM, Larsen JP, Beiske A *et al.* The effect of tolcapone on the pharmacokinetics of benserazide. *Eur J Neurol* 1999; 6: 211–219
- 348 Jørgensen A. Pharmacokinetic studies in volunteers of intravenous and oral cis(Z)-flupentixol and intramuscular cis (Z)-flupentixol decanoate in Viscoleo. *Eur J Clin Pharmacol* 1980; 18: 355–360
- 349 Jornil J, Jensen KG, Larsen F *et al.* Identification of cytochrome P450 isoforms involved in the metabolism of paroxetine and estimation of their importance for human paroxetine metabolism using a population-based simulator. *Drug Metab Dispos* 2010; 38: 376–385

- 350 Kandasamy M, Srinivas P, Subramaniam K et al. Differential outcomes from metabolic ratios in the identification of CYP2D6 phenotypes – focus on venlafaxine and O-desmethylvenlafaxine. *Eur J Clin Pharmacol* 2010; 66: 879–887
- 351 Kane JM, Leucht S, Carpenter D et al. The expert consensus guideline series. Optimizing pharmacologic treatment of psychotic disorders. Introduction: methods, commentary, and summary. *J Clin Psychiatry* 2003; 12 (Suppl): 5–19
- 352 Kaplan SA, Jack ML, Weinfeld RE et al. Biopharmaceutical and clinical pharmacokinetic profile of bromazepam. *J Pharmacokinet Biopharm* 1976; 4: 1–16
- 353 Kasper S, Dötsch M, Kick H et al. Plasma concentrations of fluvoxamine and maprotiline in major depression: implications on therapeutic efficacy and side effects. *Eur Neuropsychopharmacol* 1993; 3: 13–21
- 354 Katoh Y, Uchida S, Kawai M et al. Effects of cigarette smoking and cytochrome P450 2D6 genotype on fluvoxamine concentration in plasma of Japanese patients. *Biol Pharm Bull* 2010; 33: 285–288
- 355 Katon W, Cantrell CR, Sokol MS et al. Impact of antidepressant drug adherence on comorbid medication use and resource utilization. *Arch Intern Med* 2005; 165: 2497–2503
- 356 Kaufman E, Lamster IB. The diagnostic applications of saliva – a review. *Crit Rev Oral Biol Med* 2002; 13: 197–212
- 357 Kaye CM, Nicholls B. Clinical pharmacokinetics of ropinirole. *Clin Pharmacokinet* 2000; 39: 243–254
- 358 Kemp DE, Ganocy SJ, Brecher M et al. Clinical value of early partial symptomatic improvement in the prediction of response and remission during short-term treatment trials in 3369 subjects with bipolar I or II depression. *J Affect Disord* 2011; 130: 171–179
- 359 Kennedy MC. Post mortem drug concentrations. *Intern Med J* 2010; 40: 183–187
- 360 Kerr BM, Thummel KE, Wurden CJ et al. Human liver carbamazepine metabolism. Role of CYP3A4 and CYP2C8 in 10,11-epoxide formation. *Biochem Pharmacol* 1994; 47: 1969–1979
- 361 Kinirons MT, O'Mahony MS. Drug metabolism and ageing. *Br J Clin Pharmacol* 2004; 57: 540–544
- 362 Kirchheiner J. CYP2D6 phenotype prediction from genotype: which system is the best? *Clin Pharmacol Ther* 2008; 83: 225–227
- 363 Kirchheiner J, Meineke I, Müller G et al. Contributions of CYP2D6, CYP2C9 and CYP2C19 to the biotransformation of E- and Z-doxepin in healthy volunteers. *Pharmacogenetics* 2002; 12: 571–580
- 364 Kirchheiner J, Müller G, Meineke I et al. Effects of polymorphisms in CYP2D6, CYP2C9, and CYP2C19 on trimipramine pharmacokinetics. *J Clin Psychopharmacol* 2003; 23: 459–466
- 365 Kirchheiner J, Nickchen K, Bauer M et al. Pharmacogenetics of antidepressants and antipsychotics: the contribution of allelic variations to the phenotype of drug response. *Mol Psychiatry* 2004; 9: 442–473
- 366 Kirchherr H, Kühn-Velten WN. Quantitative determination of forty-eight antidepressants and antipsychotics in human serum by HPLC tandem mass spectrometry: a multi-level, single-sample approach. *J Chromatogr B* 2006; 843: 100–113
- 367 Kirkton C, McIntyre IM. Therapeutic and toxic concentrations of mirtazapine. *J Anal Toxicol* 2006; 30: 687–691
- 368 Kirschbaum KM, Müller MJ, Malevani J et al. Serum levels of aripiprazole and dehydroaripiprazole, clinical response and side effects. *World J Biol Psychiatry* 2008; 9: 212–218
- 369 Kirschbaum KM, Müller MJ, Zernig G et al. Therapeutic monitoring of aripiprazole by HPLC with column-switching and spectrophotometric detection. *Clin Chem* 2005; 51: 1718–1721
- 370 Kirschbaum KM, Uhr M, Holthoewer D et al. Pharmacokinetics of acute and sub-chronic aripiprazole in P-glycoprotein deficient mice. *Neuropharmacology* 2010; 59: 474–479
- 371 Kjolbye M, Thomsen K, Rogne T et al. Search for a therapeutic range for serum zuclopenthixol concentrations in schizophrenic patients. *Ther Drug Monit* 1994; 16: 541–547
- 372 Klamerus KJ, Maloney K, Rudolph RL et al. Introduction of a composite parameter to the pharmacokinetics of venlafaxine and its active O-desmethyl metabolite. *J Clin Pharmacol* 1992; 32: 716–724
- 373 Klampfl K, Taurines R, Preuss A et al. Serum concentrations, therapeutic response and side effects in children and adolescents with impulsive-aggressive symptoms during risperidone therapy. *Pharmacopsychiatry* 2010; 43: 58–65
- 374 Klotz U. Pharmacokinetics and drug metabolism in the elderly. *Drug Metab Rev* 2009; 41: 67–76
- 375 Koepf MJ, Patsalos PN, Sander JW. Sulthiame in adults with refractory epilepsy and learning disability: an open trial. *Epilepsy Res* 2002; 50: 277–282
- 376 Kondo T, Otani K, Ishida M et al. Adverse effects of zotepine and their relationship to serum concentrations of the drug and prolactin. *Ther Drug Monit* 1994; 16: 120–124
- 377 Koostra-Ros JE, Van Weelden MJ, Hinrichs JW et al. Therapeutic drug monitoring of antidepressants and cytochrome P450 genotyping in general practice. *J Clin Pharmacol* 2006; 46: 1320–1327
- 378 Kornhuber J, Quack G, Danyasz W et al. Therapeutic brain concentration of the NMDA receptor antagonist amantadine. *Neuropharmacology* 1995; 34: 713–721
- 379 Koyama E, Chiba K, Tani M et al. Identification of human cytochrome P450 isoforms involved in the stereoselective metabolism of mianserin enantiomers. *J Pharmacol Exp Ther* 1996; 278: 21–30
- 380 Kozer E, Parvez S, Minassian BA et al. How high can we go with phenytoin? *Ther Drug Monit* 2002; 24: 386–389
- 381 Kraska J, Corner DA. Serum drug level monitoring in affective disorders. *J Clin Pharm Ther* 1992; 17: 357–363
- 382 Kugelberg FC, Druid H, Carlsson B et al. Postmortem redistribution of the enantiomers of citalopram and its metabolites: an experimental study in rats. *J Anal Toxicol* 2004; 28: 631–637
- 383 Kuhlman JJJR, Levine B, Johnson RE et al. Relationship of plasma buprenorphine and norbuprenorphine to withdrawal symptoms during dose induction, maintenance and withdrawal from sublingual buprenorphine. *Addiction* 1998; 93: 549–559
- 384 Kuss HJ, Feistenauer E. Quantitative high-performance liquid chromatographic assay for the determination of maprotiline and oxaprotiline in human plasma. *J Chromatogr* 1981; 204: 349–353
- 385 Kvist EE, Al-Shurbaji A, Dahl ML et al. Quantitative pharmacogenetics of nortriptyline: a novel approach. *Clin Pharmacokinet* 2001; 40: 869–877
- 386 Lappenberg-Pelzer M. Identification and determination of opiapramol metabolites in plasma and urine. *J Anal Toxicol* 1998; 22: 215–219
- 387 Lautala P, Ethell BT, Taskinen J et al. The specificity of glucuronidation of entacapone and tolcapone by recombinant human UDP-glucuronosyltransferases. *Drug Metab Dispos* 2000; 28: 1385–1389
- 388 Le Bloch Y, Woggon B, Weissenrieder H et al. Routine therapeutic drug monitoring in patients treated with 10–360 mg/day citalopram. *Ther Drug Monit* 2003; 25: 600–608
- 389 Lee CA, Cook JA, Reyner EL et al. P-glycoprotein related drug interactions: clinical importance and a consideration of disease states. *Expert Opin Drug Metab Toxicol* 2010; 6: 603–619
- 390 Lee SY, Kim YG, Kim HG et al. Pharmacokinetic parameters of bromperidol in Korean subjects. *Hum Psychopharmacol* 2006; 21: 409–412
- 391 Lefèvre G, Büche M, Sedek G et al. Similar rivastigmine pharmacokinetics and pharmacodynamics in Japanese and white healthy participants following the application of novel rivastigmine patch. *J Clin Pharmacol* 2009; 49: 430–443
- 392 Leucht S, Busch R, Kissling W et al. Early prediction of antipsychotic nonresponse among patients with schizophrenia. *J Clin Psychiatry* 2007; 68: 352–360
- 393 Leucht S, Steimer W, Kreuz S et al. Doxepin plasma concentrations: is there really a therapeutic range? *J Clin Psychopharmacol* 2001; 21: 432–439
- 394 LeWitt PA, Jennings D, Kelly EL et al. Pharmacokinetic-pharmacodynamic crossover comparison of two levodopa extension strategies. *Mov Disord* 2009; 24: 1319–1324
- 395 Licht RW, Vestergaard P, Kessing LV et al. Psychopharmacological treatment with lithium and antiepileptic drugs: suggested guidelines from the Danish Psychiatric Association and the Child and Adolescent Psychiatric Association in Denmark. *Acta Psychiatr Scand Suppl* 2003; 419: 1–22
- 396 Lieberman JA, Stroup TS, McEvoy JP et al. Effectiveness of Antipsychotic Drugs in Patients with Chronic Schizophrenia. *N Engl J Med* 2005; 353: 1209–1223
- 397 Lind AB, Reis M, Bengtsson F et al. Steady-state concentrations of mirtazapine, N-desmethylmirtazapine, 8-hydroxymirtazapine and their enantiomers in relation to cytochrome P450 2D6 genotype, age and smoking behaviour. *Clin Pharmacokinet* 2009; 48: 63–70
- 398 Lindberger M, Luhr O, Johannessen SI et al. Serum concentrations and effects of gabapentin and vigabatrin: observations from a dose titration study. *Ther Drug Monit* 2003; 25: 457–462
- 399 Lindenmayer J. Long-acting injectable antipsychotics: focus on Olanzapine pamoate. *Neuropsychiatr Dis Treat* 2010; 6: 261–267
- 400 Linder MW, Keck PEJR. Standards of laboratory practice: antidepressant drug monitoring. *National Academy of Clinical Biochemistry*. *Clin Chem* 1998; 44: 1073–1084
- 401 Lingam R, Scott J. Treatment non-adherence in affective disorders. *Acta Psychiatr Scand* 2002; 105: 164–172

- 402 Lingjaerde O, Ahlfors UG, Bech P et al. The UKU side effect rating scale. A new comprehensive rating scale for psychotropic drugs and a cross-sectional study of side effects in neuroleptic-treated patients. *Acta Psychiatr Scand Suppl* 1987; 334: 1–100
- 403 Liu Y, Jiao J, Zhang C et al. A simplified method to determine five cytochrome p450 probe drugs by HPLC in a single run. *Biol Pharm Bull* 2009; 32: 717–720
- 404 Liu ZQ, Shu Y, Huang SL et al. Effects of CYP2C19 genotype and CYP2C9 on fluoxetine N-demethylation in human liver microsomes. *Acta Pharmacol Sin* 2001; 22: 85–90
- 405 Lobo ED, Bergstrom RF, Reddy S et al. In vitro and in vivo evaluations of cytochrome P450 1A2 interactions with duloxetine. *Clin Pharmacokinet* 2008; 47: 191–202
- 406 Locatelli I, Kstelic M. Kores-Plesnicar et al. A population pharmacokinetic evaluation of the influence of CYP2D6 genotype on risperidone metabolism in patients with acute episode schizophrenia. *Eur J Pharm Sci* 2010; 41: 289–298
- 407 Lotrich FE, Pollock BG. Aging and clinical pharmacology: implications for antidepressants. *J Clin Pharmacol* 2005; 45: 1106–1122
- 408 Lucek R, Dixon R. Chlordiazepoxide concentrations in saliva and plasma measured by radioimmunoassay. *Res Commun Chem Pathol Pharmacol* 1980; 27: 397–400
- 409 Lundberg J, Christophersen JS, Peteresen KB et al. *Int J Neuropsychopharmacol* 2007; 10: 777–785
- 410 Lundmark J, Bengtsson F, Nordin C et al. Therapeutic drug monitoring of selective serotonin reuptake inhibitors influences clinical dosing strategies and reduces drug costs in depressed elderly patients. *Acta Psychiatr Scand* 2000; 101: 354–359
- 411 Luurila H, Olkkola KT. Pharmacokinetic-pharmacodynamic modeling of zopiclone effects on human central nervous system. *Pharmacol Toxicol* 1996; 78: 348–353
- 412 Madan A, Parkinson A, Faiman MD. Identification of the human P-450 enzymes responsible for the sulfoxidation and thiono-oxidation of diethylthiocarbamate methyl ester: role of P-450 enzymes in disulfiram bioactivation. *Alcohol Clin Exp Res* 1998; 22: 1212–1219
- 413 Madsen H, Nielsen KK, Brøsen K. Imipramine metabolism in relation to the sparteine and mephenytoin oxidation polymorphisms – a population study. *Br J Clin Pharmacol* 1995; 39: 433–439
- 414 Maguire KP, Burrows GD, Norman TR et al. Metabolism and pharmacokinetics of dothiepin. *Br J Clin Pharmacol* 1981; 12: 405–409
- 415 Maguire KP, Norman TR, Burrows GD et al. An evaluation of maprotiline intravenous kinetics and comparison of two oral doses. *Eur J Clin Pharmacol* 1980; 18: 249–254
- 416 Mahmood I, Sahajwalla C. Clinical pharmacokinetics and pharmacodynamics of buspirone, an anxiolytic drug. *Clin Pharmacokinet* 1999; 36: 277–287
- 417 Mallikaarjun S, Salazar DE, Bramer SL. Pharmacokinetics, tolerability, and safety of aripiprazole following multiple oral dosing in normal healthy volunteers. *J Clin Pharmacol* 2004; 44: 179–187
- 418 Mamo D, Kapur S, Keshavan M et al. D2 receptor occupancy of olanzapine pamoate depot using positron emission tomography: an open-label study in patients with schizophrenia. *Neuropsychopharmacology* 2008; 33: 298–304
- 419 Mamo D, Kapur S, Shammi CM et al. A PET study of dopamine D2 and serotonin 5-HT2 receptor occupancy in patients with schizophrenia treated with therapeutic doses of ziprasidone. *Am J Psychiatry* 2004; 161: 818–825
- 420 Mann K, Hiemke C, Lotz J et al. Appropriateness of plasma level determinations for lithium and valproate in routine care of psychiatric inpatients with affective disorders. *J Clin Psychopharmacol* 2006; 26: 671–673
- 421 Mann K, Hiemke C, Schmidt LG et al. Appropriateness of therapeutic drug monitoring for antidepressants in routine psychiatric inpatient care. *Ther Drug Monit* 2006; 28: 83–88
- 422 Markowitz J, Patrick K. Differential pharmacokinetics and pharmacodynamics of methylphenidate enantiomers: does chirality matter? *J Clin Psychopharmacol* 2008; 28 (Suppl 2): S54–S61
- 423 Martines C, Gatti G, Sasso E et al. The disposition of primidone in elderly patients. *Br J Clin Pharmacol* 1990; 30: 607–611
- 424 Mason BJ, Goodman AM, Dixon RM et al. A pharmacokinetic and pharmacodynamic drug interaction study of acamprosate and naltrexone. *Neuropsychopharmacology* 2002; 27: 596–606
- 425 Mattila MA, Larni HM. Flunitrazepam: a review of its pharmacological properties and therapeutic use. *Drugs* 1980; 20: 353–374
- 426 Mauri MC, Laini V, Boscati L et al. Long-term treatment of chronic schizophrenia with risperidone: a study with plasma levels. *Eur Psychiatry* 2001; 16: 57–63
- 427 Mauri MC, Volonteri LS, Colasanti A et al. Clinical pharmacokinetics of atypical antipsychotics. A critical review of the relationship between plasma concentrations and clinical response. *Clin Pharmacokinet* 2007; 46: 359–388
- 428 May TW, Korn-Merker E, Rambeck B. Clinical pharmacokinetics of oxcarbazepine. *Clin Pharmacokinet* 2003; 42: 1023–1042
- 429 May TW, Korn-Merker E, Rambeck B et al. Pharmacokinetics of sulthiame in epileptic patients. *Ther Drug Monit* 1994; 16: 251–257
- 430 May TW, Rambeck B, Jürgens U. Serum concentrations of levetiracetam in epileptic patients: the influence of dose and co-medication. *Ther Drug Monit* 2003; 25: 690–699
- 431 May TW, Rambeck B, Jürgens U. Serum concentrations of topiramate in patients with epilepsy: influence of dose, age, and comedication. *Ther Drug Monit* 2002; 24: 366–374
- 432 May TW, Rambeck B, Neb R et al. Serum concentrations of pregabalin in patients with epilepsy: the influence of dose, age, and comedication. *Ther Drug Monit* 2007; 29: 789–794
- 433 Mayo BC, Biggs SR, Chasseaud LF et al. The metabolic fate of Sormodren (bornaprine hydrochloride) in animals and humans. *Xenobiotica* 1980; 10: 873–888
- 434 McAlpine DE, Biernacka JM, Mrazek DA et al. Effect of cytochrome P450 enzyme polymorphisms on pharmacokinetics of venlafaxine. *Ther Drug Monit* 2011; 33: 14–20
- 435 McConville BJ, Arvanitis LA, Thyrum PT et al. Pharmacokinetics, tolerability, and clinical effectiveness of quetiapine fumarate: an open-label trial in adolescents with psychotic disorders. *J Clin Psychiatry* 2000; 61: 252–260
- 436 McKenzie ME, Roswell-Harris D. A controlled trial of prothipendyl (tolnate) in mentally subnormal patients. *Br J Psychiatry* 1966; 112: 95–100
- 437 Medori R, Mannaert E, Gründer G. Plasma antipsychotic concentration and receptor occupancy, with special focus on risperidone long-acting injectable. *Eur Neuropsychopharmacol* 2006; 16: 233–240
- 438 Mehler-Wex C, Kölch M, Kirchheiner J et al. Drug monitoring in child and adolescent psychiatry for improved efficacy and safety of psychopharmacotherapy. *Child Adolesc Psychiatry Ment Health* 2009; 3: 14
- 439 Meijer WE, Bouvy ML, Heerdink ER et al. Spontaneous lapses in dosing during chronic treatment with selective serotonin reuptake inhibitors. *Br J Psychiatry* 2001; 179: 519–522
- 440 Meineke I, Kress I, Poser W et al. Therapeutic drug monitoring and its metabolite desmethylmirtazapine by HPLC with fluorescence detection. *Ther Drug Monit* 2004; 26: 277–283
- 441 Meisenzahl EM, Schmitt G, Gründer G et al. Striatal D2/D3 receptor occupancy, clinical response and side effects with amisulpride: an iodine-123-iodobenzamide SPET study. *Pharmacopsychiatry* 2008; 41: 169–175
- 442 Meyer JH. Imaging the serotonin transporter during major depressive disorder and antidepressant treatment. *J Psychiatry Neurosci* 2007; 32: 86–102
- 443 Meyer JH, Wilson AA, Sagrati S et al. Serotonin transporter occupancy of five selective serotonin reuptake inhibitors at different doses: an [¹¹C]DASB positron emission tomography study. *Am J Psychiatry* 2004; 161: 826–835
- 444 Meyer RP, Gehlhaus M, Knoth R et al. Expression and function of cytochrome p450 in brain drug metabolism. *Curr Drug Metab* 2007; 8: 297–306
- 445 Meyer-Barner M, Meineke I, Schreeb KH et al. Pharmacokinetics of doxepin and desmethyldoxepin: an evaluation with the population approach. *Eur J Clin Pharmacol* 2002; 58: 253–257
- 446 Michelson D, Read HA, Ruff DD et al. CYP2D6 and clinical response to atomoxetine in children and adolescents with ADHD. *J Am Acad Child Adolesc Psychiatry* 2007; 46: 242–251
- 447 Mihara K, Yasui-Furukori N, Kondo T et al. Relationship between plasma concentrations of trazodone and its active metabolite, m-chlorophenylpiperazine, and its clinical effect in depressed patients. *Ther Drug Monit* 2002; 24: 563–566
- 448 Mimaki T. Clinical pharmacology and therapeutic drug monitoring of zonisamide. *Ther Drug Monit* 1998; 20: 593–597
- 449 Miura H. Zonisamide monotherapy with once-daily dosing in children with cryptogenic localization-related epilepsies: clinical effects and pharmacokinetic studies. *Seizure* 2004; 13 (Suppl 1): S17–S23
- 450 Miura M, Ohkubo T. Identification of human cytochrome P450 enzymes involved in the major metabolic pathway of fluvoxamine. *Xenobiotica* 2007; 37: 169–179

- 451 *Moffat AC*, editor. *Clarke's analysis of drugs and poisons*. London: Pharmaceutical Press, 2003; 1468–1469
- 452 *Molden E, Lunde H, Lunder N et al*. Pharmacokinetic variability of aripiprazole and the active metabolite dehydroaripiprazole in psychiatric patients. *Ther Drug Monit* 2006; 28: 744–749
- 453 *Montgomery SA, McAuley R, Montgomery DB*. Relationship between mianserin plasma levels and antidepressant effect in a double-blind trial comparing a single night-time and divided daily dose regimens. *Br J Clin Pharmacol* 1978; 5: 715–765
- 454 *Moody DE, Chang Y, Huang W et al*. The in vivo response of novel buprenorphine metabolites, M1 and M3, to antiretroviral inducers and inhibitors of buprenorphine metabolism. *Basic Clin Pharmacol Toxicol* 2009; 105: 211–215
- 455 *Morris RG, Black AB, Harris AL et al*. Lamotrigine and therapeutic drug monitoring: retrospective survey following the introduction of a routine service. *Br J Clin Pharmacol* 1998; 46: 547–551
- 456 *Morris RG, Lee MY, Cleanthous X et al*. Long-term follow-up using a higher target range for lamotrigine monitoring. *Ther Drug Monit* 2004; 26: 626–632
- 457 *Mrazek DA*. Psychiatric pharmacogenomic testing in clinical practice. *Dialogues Clin Neurosci* 2010; 12: 69–76
- 458 *Müller H, Eusterschulte B, Havemann-Reinecke U et al*. Forensische Aspekte des therapeutischen Drug-Monitorings (TDM) in der Psychiatrie. *Psychopharmakotherapie* 2009; 16: 52–56
- 459 *Müller MJ, Dragicevic A, Fric M et al*. Therapeutic drug monitoring of tricyclic antidepressants: how does it work under clinical conditions? *Pharmacopsychiatry* 2003; 36: 98–104
- 460 *Müller MJ, Härtter S, Köhler D et al*. Serum levels of sulpiride enantiomers after oral treatment with racemic sulpiride in psychiatric patients: a pilot study. *Pharmacopsychiatry* 2001; 34: 27–32
- 461 *Müller MJ, Regenbogen B, Härtter S et al*. Therapeutic drug monitoring for optimizing amisulpride therapy in patients with schizophrenia. *J Psychiat Res* 2007; 41: 673–679
- 462 *Müller-Isberner R, Freese R, Jöckel D et al*. Forensic psychiatric assessment and treatment in Germany. Legal framework, recent developments, and current practice. *Int J Law Psychiatry* 2000; 23: 467–480
- 463 *Nagy CF, Kumar D, Cullen EI et al*. Steady-state pharmacokinetics and safety of donepezil HCl in subjects with moderately impaired renal function. *Br J Clin Pharmacol* 2004; 58 (Suppl 1): 18–24
- 464 *Naito H, Wachi M, Nishida M*. Clinical effects and plasma concentrations of long-term clonazepam monotherapy in previously untreated epileptics. *Acta Neurol Scand* 1987; 76: 58–63
- 465 *Nakamura K, Yokoi T, Inoue K et al*. CYP2D6 is the principal cytochrome P450 responsible for metabolism of the histamine H1 antagonist promethazine in human liver microsomes. *Pharmacogenetics* 1996; 6: 449–457
- 466 *Nazirizadeh Y, Vogel F, Bader W et al*. Serum concentrations of paliperidone versus risperidone and clinical effects. *Eur J Clin Pharmacol* 2010; 66: 797–803
- 467 *Neels HM, Sierens AC, Naelaerts K et al*. Therapeutic drug monitoring of old and newer anti-epileptic drugs. *Clin Chem Lab Med* 2004; 42: 1228–1255
- 468 *Nemoda Z, Angyal N, Tarnok Z et al*. Carboxylesterase 1 gene polymorphism and methylphenidate response in ADHD. *Neuropharmacology* 2009; 57: 731–733
- 469 *Nesvag R, Hendset M, Refsum H et al*. Serum concentrations of risperidone and 9-OH risperidone following intramuscular injection of long-acting risperidone compared with oral risperidone medication. *Acta Psychiatr Scand* 2006; 114: 21–26
- 470 *Nikisch G, Baumann P, Oneda B et al*. Cytochrome P450 and ABCB1 genetics: association with quetiapine and norquetiapine plasma and cerebrospinal fluid concentrations and with clinical response in patients suffering from schizophrenia. A pilot study. *J Psychopharmacol* 2010; Dec 8 [Epub ahead of print]
- 471 *Nikisch G, Mathé AA, Czernik A et al*. Stereoselective metabolism of citalopram in plasma and cerebrospinal fluid of depressive patients: relationship with 5-HIAA in CSF and clinical response. *J Clin Psychopharmacol* 2004; 24: 283–290
- 472 *Nikolaus S, Larisch R, Beu M et al*. In vivo measurement of D2 receptor density and affinity for 18F-(3-N-methyl)benperidol in the rat striatum with a PET system for small laboratory animals. *J Nucl Med* 2003; 44: 618–624
- 473 *Nilsen OG, Dale O*. Single dose pharmacokinetics of trazodone in healthy subjects. *Pharmacol Toxicol* 1992; 71: 150–153
- 474 *Nilsson MI, Meresaar U, Ånggård E*. Clinical pharmacokinetics of methadone. *Acta Anaesthesiol Scand Suppl* 1982; 74: 66–69
- 475 *Nishikage H, Nakanishi T, Takamitsu Y et al*. Sequential changes in the plasma concentration of risperidone following intentional overdose. *Clin Neuropharmacol* 2002; 25: 307–309
- 476 *Nnadi CU, Malhotra AK*. Clinical and pharmacogenetic studies of iloperidone. *Per Med* 2008; 5: 367–375
- 477 *Norman T, Chiu E, James RH et al*. Single oral dose pharmacokinetics of tiapride in patients with Huntington's disease. *Eur J Clin Pharmacol* 1987; 32: 583–586
- 478 *Nozawa M, Ohnuma T, Matsubara Y et al*. The relationship between the response of clinical symptoms and plasma olanzapine concentration, based on pharmacogenetics. *Ther Drug Monit* 2008; 30: 35–40
- 479 *Nutt JG, Fellman JH*. Pharmacokinetics of levodopa. *Clin Neuropharmacol* 1984; 7: 35–49
- 480 *Nyberg S, Nordström AL, Halldin C et al*. Positron emission tomography studies on D2 dopamine receptor occupancy and plasma antipsychotic drug levels in man. *Int Clin Psychopharmacol* 1995; 10 (Suppl 3): 81–85
- 481 *Nyholm D*. Pharmacokinetic optimisation in the treatment of Parkinson's disease: an update. *Clin Pharmacokinet* 2006; 45: 109–136
- 482 *Obach RS, Cox LM, Tremaine LM*. Sertraline is metabolized by multiple cytochrome P450 enzymes, monoamine oxidases, and glucuronyl transferses in human: an in vitro study. *Drug Metab Dispos* 2005; 33: 262–270
- 483 *Öhman D, Cherma MD, Norlander B et al*. Determination of serum reboxetine enantiomers in patients on chronic medication with racemic reboxetine. *Ther Drug Monit* 2003; 25: 174–182
- 484 *Öhman D, Norlander B, Peterson C et al*. Bioanalysis of racemic reboxetine and its desethylated metabolite in a therapeutic drug monitoring setting using solid phase extraction and HPLC. *Ther Drug Monit* 2001; 23: 27–34
- 485 *Olesen OV, Linnet K*. Hydroxylation and demethylation of the tricyclic antidepressant nortriptyline by cDNA-expressed human cytochrome P-450 isozymes. *Drug Metab Dispos* 1997; 25: 740–744
- 486 *Olesen OV, Linnet K*. Identification of the human cytochrome P450 isoforms mediating in vitro N-dealkylation of perphenazine. *Br J Clin Pharmacol* 2000; 50: 563–571
- 487 *Olesen OV, Linnet K*. Contributions of five human cytochrome P450 isoforms to the N-demethylation of clozapine in vitro at low and high concentrations. *J Clin Pharmacol* 2001; 41: 823–832
- 488 *Orsulak PJ*. Therapeutic monitoring of antidepressant drugs: current methodology and applications. *J Clin Psychiatry* 1986; 47 (Suppl): 39–52
- 489 *Orsulak PJ*. Therapeutic monitoring of antidepressant drugs: guidelines updated. *Ther Drug Monit* 1989; 11: 497–507
- 490 *Orsulak PJ, Schildkraut JJ*. Guidelines for therapeutic monitoring of tricyclic antidepressant plasma levels. *Ther Drug Monit* 1979; 1: 199–208
- 491 *Ostad Haji E, Tadić A, Wagner S et al*. Association between citalopram serum levels and clinical improvement of patients with major depression. *J Clin Psychopharmacol* 2011; 31: 281–286
- 492 *Ota T, Shinotoh H, Fukushi K et al*. Estimation of plasma IC50 of donepezil for cerebral acetylcholinesterase inhibition in patients with Alzheimer's disease using positron emission tomography. *Clin Neuropharmacol* 2010; 33: 74–78
- 493 *Palego L, Biondi L, Giannaccini G et al*. Clozapine, norclozapine plasma levels, their sum and ratio in 50 psychotic patients: influence of patient-related variables. *Prog Neuropsychopharmacol Biol Psychiatry* 2002; 26: 473–480
- 494 *Panagiotidis G, Arthur HW, Lindh JD et al*. Depot haloperidol treatment in outpatients with schizophrenia on monotherapy: impact of CYP2D6 polymorphism on pharmacokinetics and treatment outcome. *Ther Drug Monit* 2007; 29: 417–422
- 495 *Paris BL, Ogilvie BW, Scheinkoenig JA et al*. In vitro inhibition and induction of human liver cytochrome p450 enzymes by milnacipran. *Drug Metab Dispos* 2009; 37: 2045–2054
- 496 *Park JY, Kim KA, Park PW et al*. Effect of CYP3A5*3 genotype on the pharmacokinetics and pharmacodynamics of alprazolam in healthy subjects. *Clin Pharmacol Ther* 2006; 79: 590–599
- 497 *Park PW, Seo YH, Ahn JY et al*. Effect of CYP3A5*3 genotype on serum carbamazepine concentrations at steady-state in Korean epileptic patients. *J Clin Pharm Ther* 2009; 34: 569–574
- 498 *Parker DR, McIntyre IM*. Case studies of post-mortem quetiapine: therapeutic or toxic concentrations? *J Analyt Toxicol* 2005; 29: 407–412
- 499 *Patsalos PN, Berry DJ, Bourgeois BF et al*. Antiepileptic drugs – best practice guidelines for therapeutic drug monitoring: a position paper by the subcommission on therapeutic drug monitoring, ILAE Commission on Therapeutic Strategies. *Epilepsia* 2008; 49: 1239–1276

- 500 Paz E, Bouzas L, Hermida J *et al*. Evaluation of three dosing models for the prediction of steady-state trough clozapine concentrations. *Clin Biochem* 2008; 41: 603–606
- 501 Pearce GA, Day RO. Compliance with criteria necessary for effective drug concentration monitoring. *Ther Drug Monit* 1990; 12: 250–257
- 502 Pedersen OL, Gram LF, Kristensen CB *et al*. Overdosage of Antidepressants: Clinical and Pharmacokinetic Aspects. *Eur J Clin Pharmacol* 1982; 23: 513–521
- 503 Perez J, Chiron C, Musial C *et al*. Stiripentol: efficacy and tolerability in children with epilepsy. *Epilepsia* 1999; 40: 1618–1626
- 504 Perry PJ. The relationship between antidepressant response and tricyclic antidepressant plasma concentrations: a retrospective analysis of the literature using logistic regression analysis. *Clin Pharmacokinet* 1987; 13: 381–392
- 505 Perry PJ. Therapeutic drug monitoring of antipsychotics. *Psychopharmacol Bull* 2001; 35: 19–29
- 506 Perry PJ, Browne JL, Alexander B *et al*. Relationship of free nortriptyline levels to therapeutic response. *Acta Psychiatr Scand* 1985; 72: 120–125
- 507 Perry PJ, Miller DD, Arndt SV *et al*. Clozapine and norclozapine plasma concentrations and clinical response of treatment-refractory schizophrenic patients. *Am J Psychiatry* 1991; 148: 231–235
- 508 Perry PJ, Miller DD, Arndt SV *et al*. Haloperidol dosing requirements: the contribution of smoking and nonlinear pharmacokinetics. *J Clin Psychopharmacol* 1993; 13: 46–51
- 509 Perry PJ, Sanger T, Beasley C. Olanzapine plasma concentrations and clinical response in acutely ill schizophrenic patients. *J Clin Psychopharmacol* 1997; 17: 472–477
- 510 Perry PJ, Zeilmann C, Arndt S. Tricyclic antidepressant concentrations in plasma: an estimate of their sensitivity and specificity as a predictor of response. *J Clin Psychopharmacol* 1994; 14: 230–240
- 511 Perucca E, Cloyd J, Critchley D *et al*. Rufinamide: clinical pharmacokinetics and concentration–response relationships in patients with epilepsy. *Epilepsia* 2008; 49: 1123–1141
- 512 Petit P, Lonjon R, Cociglio M *et al*. Carbamazepine and its 10,11-epoxide metabolite in acute mania: clinical and pharmacokinetic correlates. *Eur J Clin Pharmacol* 1991; 41: 541–546
- 513 Peyronneau MA, Delaforge M, Riviere R *et al*. High affinity of ergopeptides for cytochromes P450 3A. Importance of their peptide moiety for P450 recognition and hydroxylation of bromocriptine. *Eur J Biochem* 1994; 223: 947–956
- 514 Pichini S, Papaseit E, Joya X *et al*. Pharmacokinetics and therapeutic drug monitoring of psychotropic drugs in pediatrics. *Ther Drug Monit* 2009; 31: 283–318
- 515 Pierce DM, Franklin RA, Harry TV *et al*. Pharmacodynamic correlates of modified absorption: studies with lorazepam. *Br J Clin Pharmacol* 1984; 18: 31–35
- 516 Poggesi I, Benedetti MS, Whomsley R *et al*. Pharmacokinetics in special populations. *Drug Metab Rev* 2009; 41: 422–454
- 517 Potgieter GE, Groenewoud G, Jordaan PJ *et al*. Pharmacokinetics of pipamperone from three different tablet formulations. *Arzneimittelforschung* 2002; 52: 430–444
- 518 Pounder DJ, Jones GR. Post-mortem drug redistribution – a toxicological nightmare. *Forensic Sci Int* 1990; 45: 253–263
- 519 Prakash C, Kamel A, Cui D *et al*. Identification of the major human liver cytochrome P450 isoform(s) responsible for the formation of the primary metabolites of ziprasidone and prediction of possible drug interactions. *Br J Clin Pharmacol* 2000; 49 (Suppl 1): 35S–42S
- 520 Preskorn S, Patroneva A, Silman H *et al*. Comparison of the pharmacokinetics of venlafaxine extended release and desvenlafaxine in extensive and poor cytochrome P450 metabolizers. *J Clin Psychopharmacol* 2009; 29: 39–43
- 521 Preskorn SH. CNS drug development: part I: the early period of CNS drugs. *J Psychiatr Pract* 2010; 16: 334–339
- 522 Preskorn SH. CNS drug development: Part II: Advances from the 1960s to the 1990s. *J Psychiatr Pract* 2010; 16: 413–415
- 523 Preskorn SH. Patients who do not respond to the “usual” dose: why Terry fell off the dose-response curve. *J Psychiatr Pract* 2009; 15: 460–466
- 524 Preskorn SH. Practical application of therapeutic drug monitoring: a tale of two patients. *J Psychiatr Pract* 2008; 14: 301–306
- 525 Preskorn SH. Tricyclic antidepressant plasma level monitoring: an improvement over the dose–response approach. *J Clin Psychiatry* 1986; 47 (Suppl 1): 24–30
- 526 Preskorn SH, Burke MJ, Fast GA. Therapeutic drug monitoring: Principles and practice. *Ther Drug Monit* 1993; 16: 611–641
- 527 Preskorn SH, Fast GA. Therapeutic drug monitoring for antidepressants: efficacy, safety, and cost effectiveness. *J Clin Psychiatr* 1991; 52 (Suppl): 23–33
- 528 Preskorn SH, Fast GA. Tricyclic antidepressant-induced seizures and plasma drug concentration. *J Clin Psychiatry* 1992; 53: 160–162
- 529 Preskorn SH, Fleck RJ, Schroeder DH. Therapeutic drug monitoring of bupropion. *Am J Psychiatry* 1990; 147: 1690–1691
- 530 Preskorn SH, Jerkovich GS. Central nervous system toxicity of tricyclic antidepressants: phenomenology, course, risk factors, and role of therapeutic drug monitoring. *J Clin Psychopharmacol* 1990; 10: 88–95
- 531 Puech A, Fleuret O, Rein W. Amisulpride, an atypical antipsychotic, in the treatment of acute episodes of schizophrenia: a dose-ranging study vs. haloperidol. *Acta Psychiatr Scand* 1998; 98: 65–72
- 532 Pumariega AJ, Nelson R, Rotenberg L. Varenicline-induced mixed mood and psychotic episode in a patient with a past history of depression. *CNS Spectr* 2008; 13: 511–514
- 533 Puozzo C, Albin H, Vinçon G *et al*. Pharmacokinetics of milnacipran in liver impairment. *Eur J Drug Metab Pharmacokinet* 1998; 23: 273–279
- 534 Raaflaub J. On the pharmacokinetics of chlorprothixene in man. *Experientia* 1975; 31: 557–558
- 535 Radtke RA. Pharmacokinetics of levetiracetam. *Epilepsia* 2001; 42 (Suppl 4): 24–27
- 536 Rao VA, Bishop M, Coppen A. Clinical state, plasma levels of haloperidol and prolactin: a correlation study in chronic schizophrenia. *Br J Psychiatry* 1980; 137: 518–521
- 537 Rasmussen BB, Brøsen K. Is therapeutic drug monitoring a case for optimizing clinical outcome and avoiding interactions of the selective serotonin reuptake inhibitors? *Ther Drug Monit* 2000; 22: 143–154
- 538 Rasmussen PV, Jensen TS, Sindrup SH *et al*. TDM-based imipramine treatment in neuropathic pain. *Ther Drug Monit* 2004; 26: 352–360
- 539 Rauschenbach R, Gieschen H, Husemann M *et al*. Stable expression of human cytochrome P450 3A4 in V79 cells and its application for metabolic profiling of ergot derivatives. *Eur J Pharmacol* 1995; 293: 183–190
- 540 Ravva P, Gastonguay MR, Tensfeldt TG *et al*. Population pharmacokinetic analysis of varenicline in adult smokers. *Br J Clin Pharmacol* 2009; 68: 669–681
- 541 Rees JA. Clinical interpretation of pharmacokinetic data on dothiepine hydrochloride (Dosulepin, Prothiaden). *J Int Med Res* 1981; 9: 98–102
- 542 Regenthal R, Krueger M, Koepfel C *et al*. Drug levels: therapeutic and toxic serum/plasma concentrations of common drugs. *J Clin Monit Comput* 1999; 15: 529–544
- 543 Reimold M, Solbach C, Noda S *et al*. Occupancy of dopamine D(1), D(2) and serotonin (2A) receptors in schizophrenic patients treated with flupentixol in comparison with risperidone and haloperidol. *Psychopharmacology (Berl)* 2007; 190: 241–249
- 544 Reis M, Aamo T, Ahlner J *et al*. Reference concentrations of antidepressants. A compilation of post-mortem and therapeutic levels. *J Analyt Toxicol* 2007; 31: 254–264
- 545 Reis M, Aamo T, Spigset O *et al*. Serum concentrations of antidepressant drugs in a naturalistic setting: compilation based on a large therapeutic drug monitoring database. *Ther Drug Monit* 2009; 31: 42–56
- 546 Reis M, Åberg-Wistedt A, Ågren H *et al*. Compliance with SSRI medication during 6 months of treatment for major depression: an evaluation by determination of repeated serum drug concentrations. *J Affect Disorders* 2004; 82: 443–446
- 547 Reis M, Akerblad AC, Ekselius L *et al*. Partial compliance as determined from plasma levels of sertraline and its metabolite in depressed patients in primary care. *J Clin Psychopharmacol* 2010; 30: 746–748
- 548 Reis M, Chermá MD, Carlsson B *et al*. On behalf of the task force for TDM of escitalopram in Sweden. Therapeutic drug monitoring of escitalopram in an outpatient setting. *Ther Drug Monit* 2007; 29: 758–766
- 549 Reis M, Lundmark J, Bengtsson F. Therapeutic drug monitoring of racemic citalopram: a 5-year experience in Sweden, 1992–1997. *Ther Drug Monit* 2003; 25: 183–191
- 550 Reis M, Lundmark J, Björk H *et al*. Therapeutic drug monitoring of racemic venlafaxine and its main metabolites in an everyday clinical setting. *Ther Drug Monit* 2002; 24: 545–553
- 551 Reis M, Olsson G, Carlsson B *et al*. Serum levels of citalopram and its main metabolites in adolescent patients treated in a naturalistic clinical setting. *J Clin Psychopharmacol* 2002; 22: 406–413
- 552 Reis M, Prochazka J, Sitsen A *et al*. Inter- and intraindividual pharmacokinetic variations of mirtazapine and its N-demethyl metabolite in patients treated for major depressive disorder: a 6-month therapeutic drug monitoring study. *Ther Drug Monit* 2005; 27: 469–477

- 553 Remington G, Mamo D, Labelle A et al. A PET study evaluating dopamine D₂ receptor occupancy for long-acting injectable risperidone. *Am J Psychiatry* 2006; 163: 396–401
- 554 Renwick AB, Mistry H, Ball SE et al. Metabolism of Zaleplon by human hepatic microsomal cytochrome P450 isoforms. *Xenobiotica* 1998; 28: 337–348
- 555 Riant P, Urien S, Albengres E et al. Effects of the binding of imipramine to erythrocytes and plasma proteins on its transport through the rat blood-brain barrier. *J Neurochem* 1988; 51: 421–425
- 556 Richens A, Banfield CR, Salvi M et al. Single and multiple dose pharmacokinetics of felbamate in the elderly. *Br J Clin Pharmacol* 1997; 44: 129–134
- 557 Riedel M, Schwarz MJ, Strassnig M et al. Risperidone plasma levels, clinical response and side-effects. *Eur Arch Psychiatry Clin Neurosci* 2005; 255: 261–268
- 558 Rivas N, Buelga DS, Elger CE et al. Population pharmacokinetics of lamotrigine with data from therapeutic drug monitoring in German and Spanish patients with epilepsy. *Ther Drug Monit* 2008; 30: 483–489
- 559 Rivera-Calimlim L, Castañeda L, Lasagna L. Effects of mode of management on plasma chlorpromazine in psychiatric patients. *Clin Pharmacol Ther* 1973; 14: 978–986
- 560 Roberts RL, Joyce PR, Mulder RT et al. A common P-glycoprotein polymorphism is associated with nortriptyline-induced postural hypotension in patients treated for major depression. *Pharmacogenomics J* 2002; 2: 191–196
- 561 Robertson PJR, Hellriegel ET. Clinical pharmacokinetic profile of modafinil. *Clin Pharmacokinet* 2003; 42: 123–137
- 562 Rochat B, Kosel M, Boss G et al. Stereoselective biotransformation of the selective serotonin reuptake inhibitor, citalopram, and its demethylated metabolites by monoamine oxidases in human liver. *Biochem Pharmacol* 1998; 56: 15–23
- 563 Rogers SL, Friedhoff LT. The efficacy and safety of donepezil in patients with Alzheimer's disease: results of a US multicentre, randomized, double-blind, placebo-controlled trial. The Donepezil Study Group. *Dementia* 1996; 7: 293–303
- 564 Roman M, Kronstrand R, Lindstedt D et al. Quantitation of seven low-dosage antipsychotic drugs in human postmortem blood using LC-MS-MS. *J Anal Toxicol* 2008; 32: 147–155
- 565 Ronfeld RA, Tremaine LM, Wilner KD. Pharmacokinetics of sertraline and its N-demethyl metabolite in elderly and young male and female volunteers. *Clin Pharmacokinet* 1997; 32 (Suppl 1): 22–30
- 566 Rosenzweig P, Canal M, Patat A et al. A review of the pharmacokinetics, tolerability and pharmacodynamics of amisulpride in healthy volunteers. *Hum Psychopharmacol* 2002; 17: 1–13
- 567 Rotzinger S, Fang J, Baker GB. Trazodone is metabolized to m-chlorophenylpiperazine by CYP3A4 from human sources. *Drug Metab Dispos* 1998; 26: 572–575
- 568 Rougemont M, Ulrich S, Hiemke C et al. French summaries of product characteristics: content in relation to therapeutic monitoring of psychotropic drugs. *Fundam Clin Pharmacol* 2010; 24: 377–384
- 569 Rudorfer V, Potter WZ. The role of metabolites of antidepressants in the treatment of depression. *CNS Drugs* 1997; 7: 273–312
- 570 Ruottinen HM, Rinne UK. Effect of one month's treatment with peripherally acting catechol-O-methyltransferase inhibitor, entacapone, on pharmacokinetics and motor response to levodopa in advanced parkinsonian patients. *Clin Neuropharmacol* 1996; 19: 222–233
- 571 Sachse J, Härtter S, Hiemke C. Automated determination of ziprasidone by HPLC with column switching and spectrophotometric detection. *Ther Drug Monit* 2005; 27: 158–162
- 572 Sachse J, Härtter S, Weigmann H et al. Automated determination of amisulpride by liquid chromatography with column switching and spectrophotometric detection. *J Chromatogr B Analyt Technol Biomed Life Sci* 2003; 784: 405–410
- 573 Sachse J, Köller J, Härtter S et al. Automated analysis of quetiapine and other antipsychotic drugs in human blood by high performance-liquid chromatography with column-switching and spectrophotometric detection. *J Chromatogr B Analyt Technol Biomed Life Sci* 2006; 830: 342–348
- 574 Sagar KA, Smyth MR. Bioavailability studies of oral dosage forms containing levodopa and carbidopa using column-switching chromatography followed by electrochemical detection. *Analyst* 2000; 125: 439–445
- 575 Sage JJ, Mark MH. Pharmacokinetics of continuous-release carbidopa/levodopa. *Clin Neuropharmacol* 1994; 17 (Suppl 2): 1–6
- 576 Sainati SM, Hubbard JW, Chi E et al. Safety, tolerability, and effect of food on the pharmacokinetics of iloperidone (HP 873), a potential atypical antipsychotic. *J Clin Pharmacol* 1995; 35: 713–720
- 577 Saint-Marcoux F, Sauvage FL, Marquet P. Current role of LC-MS in therapeutic drug monitoring. *Anal Bioanal Chem* 2007; 388: 1327–1349
- 578 Saivin S, Hulot T, Chabac S et al. Clinical pharmacokinetics of acamprosate. *Clin Pharmacokinet* 1998; 35: 331–345
- 579 Sajatovic M, Velligan DI, Weiden PJ et al. Measurement of psychiatric treatment adherence. *J Psychosom Res* 2010; 69: 591–599
- 580 Salazar DE, Frackiewicz EJ, Dockens R et al. Pharmacokinetics and tolerability of buspirone during oral administration to children and adolescents with anxiety disorder and normal healthy adults. *J Clin Pharmacol* 2001; 41: 1351–1358
- 581 Sallee FR, Pollock BG, Stiller RL et al. Pharmacokinetics of pimozide in adults and children with Tourette's syndrome. *J Clin Pharmacol* 1987; 27: 776–781
- 582 Sartorius N, Baghai TC, Baldwin DS et al. Antidepressant medications and other treatments of depressive disorders: a CINP Task Force report based on a review of evidence. *Int J Neuropsychopharmacol* 2007; 10 (Suppl 1): S1–S207
- 583 Sauer JM, Ring BJ, Witcher JW. Clinical Pharmacokinetics of atomoxetine. *Clin Pharmacokinet* 2005; 44: 571–590
- 584 Sauvage FL, Gaulier JM, Lachâtre G et al. Pitfalls and prevention strategies for liquid chromatography-tandem mass spectrometry in the selected reaction-monitoring mode for drug analysis. *Clin Chem* 2008; 54: 1519–1527
- 585 Schulberg HC, Katon W, Simon GE et al. Treating major depression in primary care practice: an update of the Agency for Health Care Policy and Research Practice Guidelines. *Arch Gen Psychiatry* 1998; 55: 1121–1127
- 586 Schulz M, Schmoltdt A. Therapeutic and toxic blood concentrations of more than 800 drugs and other xenobiotics. *Pharmazie* 2003; 58: 447–474
- 587 Schwarzenbach F, Netillard C, Demoly P et al. Antidepressant response and fluvoxamine plasma concentrations: a pilot study. *Pharm World Sci* 2003; 25: 27–29
- 588 Scordo MG, Spina E, Dahl ML et al. Influence of CYP2C9, 2C19 and 2D6 genetic polymorphisms on the steady-state plasma concentrations of the enantiomers of fluoxetine and norfluoxetine. *Basic Clin Pharmacol Toxicol* 2005; 97: 296–301
- 589 Seiler W, Wetzel H, Hillert A et al. Pharmacokinetics and bioavailability of benperidol in schizophrenic patients after intravenous and two different kinds of oral application. *Psychopharmacology (Berl)* 1994; 116: 457–463
- 590 Service JA, Waring WS. QT Prolongation and delayed atrioventricular conduction caused by acute ingestion of trazodone. *Clin Toxicol (Phila)* 2008; 46: 71–73
- 591 Shams M, Hiemke C, Härtter S. Therapeutic drug monitoring of antidepressant mirtazapine and its N-demethylated metabolite in human serum. *Ther Drug Monit* 2004; 26: 78–84
- 592 Shams ME, Arneith B, Hiemke C et al. CYP2D6 polymorphism and clinical effect of the antidepressant venlafaxine. *J Clin Pharm Ther* 2006; 31: 493–502
- 593 Sharma S, Joshi S, Chadda RK. Therapeutic drug monitoring of lithium in patients with bipolar affective disorder: experiences from a tertiary care hospital in India. *Am J Ther* 2009; 16: 393–397
- 594 Shin JG, Soukova N, Flockart DA. Effect of antipsychotic drugs on human liver cytochrome P-450 (CYP) isoforms in vitro: Preferential inhibition of CYP2D6. *Drug Metab Dispos* 1999; 27: 1078–1084
- 595 Shinderman M, Maxwell S, Brawand-Amey M et al. Cytochrome P4503A4 metabolic activity, methadone blood concentrations, and methadone doses. *Drug Alcohol Depend* 2003; 69: 205–211
- 596 Shiraga T, Kaneko H, Iwasaki K et al. Identification of cytochrome P450 enzymes involved in the metabolism of zotepine, an antipsychotic drug, in human liver microsomes. *Xenobiotica* 1999; 29: 217–229
- 597 Shua-Haim J, Smith J, Picard F et al. Steady-state pharmacokinetics of rivastigmine in patients with mild to moderate Alzheimer's disease not affected by co-administration of memantine: an open-label, crossover, single-centre study. *Clin Drug Investig* 2008; 28: 361–374
- 598 Sidhu J, Priskorn M, Poulsen M et al. Steady-state pharmacokinetics of the enantiomers of citalopram and its metabolites in human. *Chirality* 1997; 9: 686–692
- 599 Siva N. Tackling the booming trade in counterfeit drugs. *Lancet* 2010; 376: 1725–1726
- 600 Skinner MH, Kuan HY, Skerjanec A et al. Effect of age on the pharmacokinetics of duloxetine in women. *Br J Clin Pharmacol* 2004; 57: 54–61

- 601 Skogh E, Bengtsson F, Nordin C. Could discontinuing smoking be hazardous for patients administered clozapine medication? A case report. *Ther Drug Monit* 1999; 21: 580–582
- 602 Skogh E, Reis M, Dahl ML *et al.* Therapeutic drug monitoring data on olanzapine and its N-demethyl metabolite in the naturalistic clinical setting. *Ther Drug Monit* 2002; 24: 518–526
- 603 Small JG, Hirsch SR, Arvanitis LA *et al.* Quetiapine in patients with schizophrenia – a high- and low-dose double-blind comparison with placebo. Seroquel study group. *Arch Gen Psychiatry* 1997; 54: 549–557
- 604 Smith RB, Kroboth PD, Vanderlugt JT *et al.* Pharmacokinetics and pharmacodynamics of alprazolam after oral and IV administration. *Psychopharmacology (Berl)* 1984; 84: 452–456
- 605 Smith SW. Chiral toxicology: it's the same thing...only different. *Toxicol Sci* 2009; 110: 4–30
- 606 Snoeck E, Van Peer A, Sack M *et al.* Influence of age, renal and liver impairment on the pharmacokinetics of risperidone in man. *Psychopharmacology (Berl)* 1995; 122: 223–229
- 607 Sogaard B, Mengel H, Rao N *et al.* The pharmacokinetics of escitalopram after oral and intravenous administration of single and multiple doses to healthy subjects. *J Clin Pharmacol* 2005; 45: 1400–1406
- 608 Soldin P, Mattison DR. Sex differences in pharmacokinetics and pharmacodynamics. *Clin Pharmacokinet* 2009; 48: 143–157
- 609 Someya T, Muratake T, Hirokane G *et al.* Interindividual variation in bromperidol metabolism and relationship to therapeutic effects. *J Clin Psychopharmacol* 2000; 20: 175–180
- 610 Søndergaard Khinchi M, Nielsen KA, Dahl M *et al.* Lamotrigine therapeutic thresholds. *Seizure* 2008; 17: 391–395
- 611 Soni SD. Fluspirilene in the treatment of non-hospitalized schizophrenic patients. *Curr Med Res Opin* 1977; 4: 645–649
- 612 Sparshatt A, Taylor D, Patel MX *et al.* A systematic review of aripiprazole-dose, plasma concentration, receptor occupancy, and response: implications for therapeutic drug monitoring. *J Clin Psychiatry* 2010; 71: 1447–1456
- 613 Sparshatt A, Taylor D, Patel MX *et al.* Amisulpride – dose, plasma concentration, occupancy and response: implications for therapeutic drug monitoring. *Acta Psychiatr Scand* 2009; 120: 416–428
- 614 Spencer TJ, Biederman J, Ciccone PE *et al.* PET study examining pharmacokinetics, detection and likeability, and dopamine transporter receptor occupancy of short- and long-acting oral methylphenidate. *Am J Psychiatry* 2006; 163: 387–395
- 615 Spencer TJ, Bonab AA, Dougherty DD *et al.* A PET study examining pharmacokinetics and dopamine transporter occupancy of two long-acting formulations of methylphenidate in adults. *Int J Mol Med* 2010; 25: 261–265
- 616 Spigset O, Hägg S, Stegmayr B *et al.* Citalopram pharmacokinetics in patients with chronic renal failure and the effect of haemodialysis. *Eur J Clin Pharmacol* 2000; 59: 699–703
- 617 Spina E, Avenoso A, Facciola G *et al.* Relationship between plasma risperidone and 9-hydroxyrisperidone concentrations and clinical response in patients with schizophrenia. *Psychopharmacology (Berl)* 2001; 153: 238–243
- 618 Spina E, Birgersson C, von Bahr Ö *et al.* Phenotypic consistency in hydroxylation of desmethylmipramine and debrisoquine in healthy subjects and in human liver microsomes. *Clin Pharmacol Ther* 1984; 36: 677–682
- 619 Sproule BA, Hardy BG, Shulman KI. Differential pharmacokinetics of lithium in elderly patients. *Drugs Aging* 2000; 16: 165–177
- 620 Stassen HH, Angheliescu IG, Angst J *et al.* Predicting response to psychopharmacological treatment. Survey of recent results. *Pharmacopsychiatry* 2011; 44: 263–272
- 621 Stassen HH, Angst J, Hell D *et al.* Is there a common resilience mechanism underlying antidepressant drug response? Evidence from 2848 patients. *J Clin Psychiatry* 2007; 68: 1195–1205
- 622 Stead AH, Moffat AC. A collection of therapeutic, toxic and fatal blood drug concentrations in man. *Hum Exp Toxicol* 1983; 3: 437–464
- 623 Steimer W. Pharmacogenetics and Psychoactive Drug Therapy: Ready for the Patient? *Ther Drug Monit* 2010; 32: 381–386
- 624 Steimer W, Potter JM. Pharmacogenetic screening and therapeutic drugs. *Clin Chim Acta* 2002; 315: 137–155
- 625 Steimer W, Zöpf K, von Amelnunx S *et al.* Amitriptyline or not, that is the question: pharmacogenetic testing of CYP2D6 and CYP2C19 identifies patients with low or high risk for side effects in amitriptyline therapy. *Clin Chem* 2005; 51: 376–385
- 626 Stieffenhof V, Hiemke C. Pharmacogenetics, therapeutic drug monitoring and non compliance. *Ther Umsch* 2010; 67: 309–315
- 627 Stieffenhof V, Saglam H, Schmidtman I *et al.* Clozapine plasma level monitoring for prediction of rehospitalization schizophrenic outpatients. *Pharmacopsychiatry* 2011; 44: 55–59
- 628 Stock B, Spittler G. Metabolism of antiparkinson drugs. An example of competitive hydroxylation. *Arzneimittelforschung* 1979; 29: 610–615
- 629 Störmer E, Brockmüller J, Roots I *et al.* Cytochrome P-450 enzymes and FMO3 contribute to the disposition of the antipsychotic drug in vitro. *Psychopharmacology (Berl)* 2000; 151: 312–320
- 630 Störmer E, von Moltke LL, Shader RI *et al.* Metabolism of the antidepressant mirtazapine in vitro: contribution of cytochromes P-450 1A2, 2D6, and 3A4. *Drug Metab Dispos* 2000; 28: 1168–1175
- 631 Suhara T, Takano A, Sudo Y *et al.* High levels of serotonin transporter occupancy with low-dose clomipramine in comparative occupancy study with fluvoxamine using positron emission tomography. *Arch Gen Psychiatry* 2003; 60: 386–391
- 632 Sutfin TA, Perini GI, Molnar G *et al.* Multiple-dose pharmacokinetics of imipramine and its major active and conjugated metabolites in depressed patients. *J Clin Psychopharmacol* 1988; 8: 48–53
- 633 Suzuki A, Otani K, Ishida M *et al.* Increased plasma concentrations of bromperidol and its reduced metabolite with levomepromazine, but not with thioridazine. *Ther Drug Monit* 1997; 19: 261–264
- 634 Suzuki Y, Fukui N, Sawamura K *et al.* Concentration-response relationship for fluvoxamine using remission as an endpoint: a receiver operating characteristics curve analysis in major depression. *J Clin Psychopharmacol* 2008; 28: 325–328
- 635 Svrbely JR, Speicher CE. The importance of request and report forms in the interpretation of therapeutic drug monitoring data. *Ther Drug Monit* 1980; 2: 211–216
- 636 Sweet RA, Pollock BG, Kirshner M *et al.* Pharmacokinetics of single- and multiple-dose bupropion in elderly patients with depression. *J Clin Pharmacol* 1995; 35: 876–884
- 637 Sweet RA, Pollock BG, Mulsant BH *et al.* Pharmacologic profile of perphenazine's metabolites. *J Clin Psychopharmacol* 2000; 20: 181–187
- 638 Szegei A, Jansen WT, van Willigenburg AP *et al.* Early improvement in the first 2 weeks as a predictor of treatment outcome in patients with major depressive disorder: a meta-analysis including 6562 patients. *J Clin Psychiatry* 2009; 70: 344–353
- 639 Takano A, Suhara T, Ichimiya T *et al.* Time course of in vivo 5-HTT transporter occupancy by fluvoxamine. *J Clin Psychopharmacol* 2006; 26: 188–191
- 640 Takano A, Suzuki K, Kosaka J *et al.* A dose-finding study of duloxetine based on serotonin transporter occupancy. *Psychopharmacology* 2006; 185: 395–399
- 641 Tan L, Yu JT, Sun YP *et al.* The influence of cytochrome oxidase CYP2A6, CYP2B6, and CYP2C9 polymorphisms on the plasma concentrations of valproic acid in epileptic patients. *Clin Neurol Neurosurg* 2010; 112: 320–323
- 642 Tanaka O, Kondo T, Otani K *et al.* Single oral dose kinetics of zotepine and its relationship to prolactin response and side effects. *Ther Drug Monit* 1998; 20: 117–119
- 643 Tanaka E, Kurata N, Yasuhara H. How useful is the “cocktail approach” for evaluating human hepatic drug metabolizing capacity using cytochrome P450 phenotyping probes in vivo? *J Clin Pharm Ther* 2003; 28: 157–165
- 644 Tasker TCG, Kaye CM, Zussman BD *et al.* Paroxetine plasma levels: lack of correlation with efficacy or adverse events. *Acta Psychiatr Scand* 1989; 80 (350): 152–155
- 645 Tateishi T, Watanabe M, Kumai T *et al.* CYP3A is responsible for N-dealkylation of haloperidol and bromperidol and oxidation of their reduced forms by human liver microsomes. *Life Sci* 2000; 67: 2913–2920
- 646 Taylor D. Antidepressant drugs and cardiovascular pathology: a clinical overview of effectiveness and safety. *Acta Psychiatr Scand* 2008; 118: 434–442
- 647 Taylor D. Psychopharmacology and adverse effects of antipsychotic long-acting injections: a review. *Br J Psychiatry Suppl* 2009; 52: S13–S19
- 648 Thanacoody RH, Daly AK, Reilly JG *et al.* Factors affecting drug concentrations and QT interval during thioridazine therapy. *Clin Pharmacol Ther* 2007; 82: 555–565
- 649 The Scottish Schizophrenia Research Group. The Scottish first episode Schizophrenia study II. Treatment: pimozide versus flupenthixol. *Br J Psychiatry* 1987; 150: 334–338
- 650 Thieme D, Rolf B, Sachs H *et al.* Correlation of inter-individual variations of amitriptyline metabolism examined in hairs with CYP2C19 and CYP2D6 polymorphisms. *Int J Legal Med* 2008; 122: 149–155

- 651 Timmer CJ, Sitsen JM, Delbressine LP. Clinical pharmacokinetics of mirtazapine. *Clin Pharmacokinet* 2000; 38: 461–474
- 652 Tiseo PJ, Rogers SL, Friedhoff LT. Pharmacokinetic and pharmacodynamic profile of donepezil HCl following evening administration. *Br J Clin Pharmacol* 1998; 46 (Suppl 1): 13–18
- 653 Titier K, Canal M, Déridet E et al. Determination of myocardium to plasma concentration ratios of five antipsychotic drugs: comparison with their ability to induce arrhythmia and sudden death in clinical practice. *Toxicol Appl Pharmacol* 2004; 199: 52–60
- 654 Toennes SW, Maurer HH. Microsoft Excel in pharmacokinetics – an easy way to solve kinetic problems in clinical toxicology, legal medicine or doping control. In: Sachs H, Bernhard W, Jeger A (eds.). Proceedings of the 34th International TIAFT Meeting, Interlaken. 11–15 August 1996. Leipzig: Molina, 1997; 201–204
- 655 Tokairin T, Fukasawa T, Yasui-Furukori N et al. Inhibition of the metabolism of brotizolam by erythromycin in humans: in vivo evidence for the involvement of CYP3A4 in brotizolam metabolism. *Br J Clin Pharmacol* 2005; 60: 172–175
- 656 Tokunaga H, Kudo K, Imamura T et al. Plasma concentrations of antipsychotic drugs in psychiatric inpatients. *Nippon Hoigaku Zasshi* 1997; 51: 417–422
- 657 Tompson DJ, Vearer D. Steady-state pharmacokinetic properties of a 24-hour prolonged-release formulation of ropinirole: results of two randomized studies in patients with Parkinson's disease. *Clin Ther* 2007; 29: 2654–2666
- 658 Topiwala A, Fazel S. The pharmacological management of violence in schizophrenia: a structured review. *Expert Rev Neurother* 2011; 11: 53–63
- 659 Tornio A, Neuvonen PJ, Backman JT. The CYP2C8 inhibitor gemfibrozil does not increase the plasma concentrations of zopiclone. *Eur J Clin Pharmacol* 2006; 62: 645–651
- 660 Touw DJ, Neef C, Thomson AH et al. Cost-effectiveness of therapeutic drug monitoring: a systematic review. *Ther Drug Monit* 2005; 27: 10–17
- 661 Trivedi MH, Rush AJ, Gaynes BN et al. Maximizing the adequacy of medication treatment in controlled trials and clinical practice: STAR(*)D measurement-based care. *Neuropsychopharmacol* 2007; 32: 2479–2489
- 662 Tsai MH, Lin KM, Hsiao MC et al. Genetic polymorphisms of cytochrome P450 enzymes influence metabolism of the antidepressant escitalopram and treatment response. *Pharmacogenomics* 2010; 11: 537–546
- 663 Tuerck D, Wang Y, Maboudian M et al. Similar bioavailability of dexamethylphenidate extended (bimodal) release, dexamethylphenidate immediate release and racemic methylphenidate extended (bimodal) release formulations in man. *Int J Clin Pharmacol Ther* 2007; 45: 662–668
- 664 Turbott J, Norman TR, Burrows GD et al. Pharmacokinetics of nortriptyline in elderly volunteers. *Commun Psychopharmacol* 1980; 4: 225–231
- 665 Turpeinen M, Koivuviita N, Tolonen A et al. Effect of renal impairment on the pharmacokinetics of bupropion and its metabolites. *Br J Clin Pharmacol* 2007; 64: 165–173
- 666 Uchida H, Mamo DC, Mulsant BH et al. Increased antipsychotic sensitivity in elderly patients: evidence and mechanisms. *J Clin Psychiatry* 2009; 70: 397–405
- 667 Uhr M, Steckler T, Yassouridis A et al. Penetration of amitriptyline, but not of fluoxetine, into brain is enhanced in mice with blood-brain barrier deficiency due to mdr1a P-glycoprotein gene disruption. *Neuropsychopharmacology* 2000; 22: 380–387
- 668 Uhr M, Tontsch A, Namendorf C et al. Polymorphisms in the drug transporter gene ABCB1 predict antidepressant treatment response in depression. *Neuron* 2008; 57: 203–239
- 669 Ujii Y, Fukasawa T, Yasui-Furukori N et al. Rifampicin markedly decreases plasma concentration and hypnotic effect of brotizolam. *Ther Drug Monit* 2006; 28: 299–302
- 670 Ulrich S, Baumann B, Wolf R et al. Therapeutic drug monitoring of clozapine and relapse – a retrospective study of routine clinical data. *Int J Clin Pharmacol Ther* 2003; 41: 3–13
- 671 Ulrich S, Hiemke C, Laux G et al. TDM group of the Arbeitsgemeinschaft Neuropsychopharmakologie und Pharmakopsychiatrie (AGNP). Value and actuality of the prescription information for therapeutic drug monitoring of psychopharmaceuticals: a comparison with the medico-scientific evidence. *Pharmacopsychiatry* 2007; 40: 121–127
- 672 Ulrich S, Läuter J. Comprehensive survey of the relationship between serum concentration and therapeutic effect of amitriptyline in depression. *Clin Pharmacokinet* 2002; 41: 853–876
- 673 Ulrich S, Sandmann U, Genz A. Serum concentrations of haloperidol pyridinium metabolites and the relationship with tardive dyskinesia and parkinsonism: a cross-section study in psychiatric patients. *Pharmacopsychiatry* 2005; 38: 171–177
- 674 Ulrich S, Würthmann C, Brosz M et al. The relationship between serum concentration and therapeutic effect of haloperidol in patients with acute schizophrenia. *Clin Pharmacokinet* 1998; 34: 227–263
- 675 Valdes R Jr, Payne DA, Linder MW (eds.). Laboratory medicine practice guidelines and recommendations for laboratory analysis and application of pharmacogenetics to clinical practice. Washington, DC: National Academy of Clinical Biochemistry, 2010
- 676 Van der Weide J, Steijns LS, van Weelden MJ. The effect of smoking and cytochrome P450 CYP1A2 genetic polymorphism on clozapine clearance and dose requirement. *Pharmacogenetics* 2003; 13: 169–172
- 677 Van der Weide J, van Baalen-Benedek EH, Kootstra-Ros JE. Metabolic ratios of psychotropics as indication of cytochrome P450 2D6/2C19 genotype. *Ther Drug Monit* 2005; 27: 478–483
- 678 Van der Zwaag C, McGee M, McEvoy JP et al. Response of patients with treatment-refractory schizophrenia to clozapine within three serum level ranges. *Am J Psychiatry* 1996; 153: 1579–1584
- 679 Van Gorp F, Whyte IM, Isbister GK. Clinical and ECG effects of Escitalopram overdose. *Ann Emerg Med* 2009; 54: 4–408
- 680 Van Putten T, Marder SR, Wirshing WC et al. Neuroleptic plasma levels. *Schizophr Bull* 1991; 17: 197–216
- 681 Varsaldi F, Miglio G, Scordo MG et al. Impact of the CYP2D6 polymorphism on steady-state plasma concentrations and clinical outcome of donepezil in Alzheimer's disease patients. *Eur J Clin Pharmacol* 2006; 62: 721–726
- 682 Vasudev K, Das S, Goswami U et al. Pharmacokinetics of valproic acid in patients with bipolar disorder. *J Psychopharmacol* 2001; 15: 187–190
- 683 Vasudev K, Goswami U, Kohli K. Carbamazepine and valproate monotherapy: feasibility, relative safety and efficacy, and therapeutic drug monitoring in manic disorder. *Psychopharmacology (Berl)* 2000; 150: 15–23
- 684 Veeffkind AH, Haffmans PMJ, Hoencamp E. Venlafaxine serum levels and CYP2D6 genotype. *Ther Drug Monit* 2000; 22: 202–208
- 685 Velligan DI, Lam YW, Glahn DC et al. Defining and assessing adherence to oral antipsychotics: a review of the literature. *Schizophr Bull* 2006; 32: 724–742
- 686 Venkatakrishnan K, Culm KE, Ehrenberg BL et al. Kinetics and dynamics of intravenous adinazolam, N-desmethyl adinazolam, and alprazolam in healthy volunteers. *J Clin Pharmacol* 2005; 45: 529–537
- 687 Venkatakrishnan K, von Moltke LL, Greenblatt DJ. Nortriptyline E-10-hydroxylation in vitro is mediated by human CYP2D6 (high affinity) and CYP3A4 (low affinity): implications for interactions with enzyme-inducing drugs. *J Clin Pharmacol* 1999; 39: 567–577
- 688 Vernaleken I, Fellows C, Janouschek H et al. Striatal and extrastriatal D2/D3-receptor-binding properties of ziprasidone: a positron emission tomography study with [18F]fallypride and [11C]raclopride (D2/D3-receptor occupancy of ziprasidone). *J Clin Psychopharmacol* 2008; 28: 608–617
- 689 Vernaleken I, Janouschek H, Raptis M et al. Dopamine D2/3 receptor occupancy by quetiapine in striatal and extrastriatal areas. *Int J Neuropsychopharmacol* 2010; 13: 951–960
- 690 Vernaleken I, Siessmeier T, Buchholz HG et al. High striatal occupancy of D2-like dopamine receptors by amisulpride in the brain of patients with schizophrenia. *Int J Neuropsychopharmacol* 2004; 7: 421–430
- 691 Ververs FF, Voorbij HA, Zwarts P et al. Effect of cytochrome P450 2D6 genotype on maternal paroxetine plasma concentrations during pregnancy. *Clin Pharmacokinet* 2009; 48: 677–683
- 692 Viala A, Ba B, Durand A et al. Comparative study of the pharmacokinetics of zuclopenthixol decanoate and fluphenazine decanoate. *Psychopharmacology (Berl)* 1988; 94: 293–297
- 693 Vine W, Bowers LD. Cyclosporine: structure, pharmacokinetics, and therapeutic drug monitoring. *Crit Rev Clin Lab Sci* 1987; 25: 275–311
- 694 Viola MS, Bercellini MA, Saiton P et al. Pharmacokinetic variability of oxcarbazepine in epileptic patients. *Medicina (B Aires)* 2000; 60: 914–918
- 695 Vogel F, Gansmüller R, Leiblein T et al. The use of ziprasidone in clinical practice: Analysis of pharmacokinetic and pharmacodynamic aspects from data of a drug monitoring survey. *Eur Psychiatry* 2009; 24: 143–148
- 696 Voineskos AN, Wilson AA, Boovariwala A et al. Serotonin transporter occupancy of high-dose selective serotonin reuptake inhibitors during major depressive disorder measured with [11C]DASB positron emission tomography. *Psychopharmacology (Berl)* 2007; 193: 539–545

- 697 Von Moltke LL, Greenblatt DJ, Giancarlo GM et al. Escitalopram (S-citalopram) and its metabolites in vitro: cytochromes mediating biotransformation, inhibitory effects, and comparison to R-citalopram. *Drug Metab Dispos* 2001; 29: 1102–1109
- 698 Von Moltke LL, Greenblatt DJ, Granda BW et al. Zolpidem metabolism in vitro: responsible cytochromes, chemical inhibitors, and in vivo correlations. *Br J Clin Pharmacol* 1999; 48: 89–97
- 699 Vormfelde SV, Bitsch A, Meineke I et al. Non-response to maprotiline caused by ultra-rapid metabolism that is different from CYP2D6? *Eur J Clin Pharmacol* 1997; 52: 387–390
- 700 Vuille F, Amey M, Baumann P. Use of plasma level monitoring of antidepressants in clinical practice. Towards an analysis of clinical utility. *Pharmacopsychiatry* 1991; 24: 190–195
- 701 Waade RB, Christensen H, Rudberg I et al. Influence of comedication on serum concentrations of aripiprazole and dehydroaripiprazole. *Ther Drug Monit* 2009; 31: 233–238
- 702 Waldschmitt C, Vogel F, Maurer C et al. Measurement of duloxetine in blood using high-performance liquid chromatography with spectrophotometric detection and column switching. *Ther Drug Monit* 2007; 29: 767–772
- 703 Waldschmitt C, Vogel F, Pfuhlmann B et al. Duloxetine serum concentrations and clinical effects. Data from a therapeutic drug monitoring (TDM) survey *Pharmacopsychiatry* 2009; 42: 189–193
- 704 Wan J, Xia H, He N et al. The elimination of diazepam in Chinese subjects is dependent on the mephenytoin oxidation phenotype. *Br J Clin Pharmacol* 1996; 42: 471–474
- 705 Wang JH, Liu ZQ, Wang W et al. Pharmacokinetics of sertraline in relation to genetic polymorphism of CYP2C19. *Clin Pharmacol Ther* 2001; 70: 42–47
- 706 Ward E, Musa MN, Bailey LG. Clinical pharmacokinetics of lithium. *J Clin Pharmacol* 1994; 34: 280–285
- 707 Weber J, McCormack PL. Asenapine CNS. *Drugs* 2009; 23: 781–792
- 708 Weiden PJ, Kozma C, Grogg A et al. Partial compliance and risk of rehospitalization among California Medicaid patients with schizophrenia. *Psychiatr Serv* 2004; 55: 886–891
- 709 Weigmann H, Bierbrauer J, Härter S et al. Automated determination of clozapine and major metabolites in serum and urine. *Ther Drug Monit* 1997; 19: 480–488
- 710 Weigmann H, Härter S, Hiemke C. Automated determination of clomipramine and its major metabolites in human and rat serum by high-performance liquid chromatography with on-line column-switching. *J Chromatogr B Biomed Sci Appl* 1998; 710: 227–233
- 711 Weigmann H, Härter S, Maehrlin S et al. Simultaneous determination of olanzapine, clozapine and demethylated metabolites in serum by on-line column-switching high-performance liquid chromatography. *J Chromatogr B Biomed Sci Appl* 2001; 759: 63–71
- 712 Weiss U, Marksteiner J, Kemmler G et al. Effects of age and sex on olanzapine plasma concentrations. *J Clin Psychopharmacol* 2005; 25: 570–574
- 713 Wen B, Ma L, Zhu M. Bioactivation of the tricyclic antidepressant amitriptyline and its metabolite nortriptyline to arene oxide intermediates in human liver microsomes and recombinant P450s. *Chem Biol Interact* 2008; 173: 59–67
- 714 Wen B, Zhou M. Metabolic activation of the phenothiazine antipsychotics chlorpromazine and thioridazine to electrophilic iminoquinone species in human liver microsomes and recombinant P450s. *Chem Biol Interact* 2009; 181: 220–226
- 715 White NC, Litovitz T, Clancy C. Suicidal antidepressant overdoses: a comparative analysis by antidepressant type. *J Med Toxicol* 2008; 4: 238–250
- 716 Wienkers LC, Allievi C, Hauer MJ et al. Cytochrome P-450-mediated metabolism of the individual enantiomers of the antidepressant agent bupropion in human liver microsomes. *Drug Metab Dispos* 1999; 27: 1334–1340
- 717 Wiesel FA, Alfredsson G, Ehrnebo M et al. The pharmacokinetics of intravenous and oral sulpiride in healthy human subjects. *Eur J Clin Pharmacol* 1980; 17: 385–391
- 718 Wille SM, Cooreman SG, Neels HM et al. Relevant issues in the monitoring and the toxicology of antidepressants. *Crit Rev Clin Lab Sci* 2008; 45: 25–89
- 719 Willmore LJ, Abelson MB, Ben-Menachem E et al. Vigabatrin: 2008 update. *Epilepsia* 2009; 50: 163–173
- 720 Wilson JF. Survey of reference ranges and clinical measurements for psychoactive drugs in serum. *Ther Drug Monit* 2003; 25: 243–247
- 721 Wilting I, Heerdink ER, Mersch PP et al. Association between lithium serum level, mood state, and patient-reported adverse drug reactions during long-term lithium treatment: a naturalistic follow-up study. *Bipolar Disord* 2009; 11: 434–440
- 722 Wincor MZ, Munjack DJ, Palmer R. Alprazolam levels and response in panic disorder: preliminary results. *J Clin Psychopharmacol* 1991; 11: 48–51
- 723 Winter HR, Earley WR, Hamer-Maansson JE et al. Steady-state pharmacokinetic, safety, and tolerability profiles of quetiapine, Norquetiapine, and other quetiapine metabolites in pediatric and adult patients with psychotic disorders. *J Child Adolesc Psychopharmacol* 2008; 18: 81–98
- 724 Wójcikowski J, Boksa J, Daniel WA. Main contribution of the cytochrome P450 isoenzyme 1A2 (CYP1A2) to N-demethylation and 5-sulfoxidation of the phenothiazine neuroleptic chlorpromazine in human liver – a comparison with other phenothiazines. *Biochem Pharmacol* 2010; 80: 1252–1259
- 725 Wójcikowski J, Daniel WA. Perazine at therapeutic drug concentrations inhibits human cytochrome P450 isoenzyme 1A2 (CYP1A2) and caffeine metabolism – an in vitro study. *Pharmacol Rep* 2009; 61: 851–858
- 726 Wójcikowski J, Pichard-Garcia L, Maurel P et al. Contribution of human cytochrome p-450 isoforms to the metabolism of the simplest phenothiazine neuroleptic promazine. *Br J Pharmacol* 2003; 138: 1465–1474
- 727 Wolff K, Hay AW, Rasitrick D et al. Steady-state pharmacokinetics of methadone in opioid addicts. *Eur J Clin Pharmacol* 1993; 44: 189–194
- 728 Wong SL, Granneman GR. Modeling of sertindole pharmacokinetic dispositions in healthy volunteers in short term dose-escalation studies. *J Pharmacol Sci* 1998; 87: 1629–1631
- 729 Wong SL, Menacherry S, Mulford D et al. Pharmacokinetics of sertindole and dehydrosertindole in volunteers with normal or impaired renal function. *Eur J Clin Pharmacol* 1997; 52: 223–227
- 730 Wright CE, Sisson TL, Ichhpurani AK et al. Steady-state pharmacokinetic properties of pramipexole in healthy volunteers. *J Clin Pharmacol* 1997; 37: 520–525
- 731 Wynalda MA, Wienkers LC. Assessment of potential interactions between dopamine receptor agonists and various human cytochrome P450 enzymes using a simple in vitro inhibition screen. *Drug Metab Dispos* 1997; 25: 1211–1214
- 732 Xiang Q, Zhao X, Zhou Y et al. Effect of CYP2D6, CYP3A5, and MDR1 genetic polymorphisms on the pharmacokinetics of risperidone and its active moiety. *J Clin Pharmacol* 2010; 50: 659–666
- 733 Xu P, Li HD, Zhang BK et al. Pharmacokinetics and tolerability of modafinil tablets in Chinese subjects. *J Clin Pharm Ther* 2008; 33: 429–437
- 734 Yao C, Raoufina A, Gold M et al. Steady-state pharmacokinetics of galantamine are not affected by addition of memantine in healthy subjects. *J Clin Pharmacol* 2005; 45: 519–528
- 735 Yasui-Furukori N, Kondo T, Ishida M et al. The characteristics of side-effects of bromperidol in schizophrenic patients. *Psychiatry Clin Neurosci* 2002; 56: 103–106
- 736 Yasui-Furukori N, Saito M, Nakagami T et al. Association between multidrug resistance 1 (MDR1) gene polymorphisms and therapeutic response to bromperidol in schizophrenic patients: a preliminary study. *Prog Neuropsychopharmacol Biol Psychiatry* 2006; 30: 286–291
- 737 Yasui-Furukori N, Saito M, Nakagami T et al. Clinical response to risperidone in relation to plasma drug concentrations in acutely exacerbated schizophrenic patients. *J Psychopharmacol* 2010; 24: 987–994
- 738 Yeung PK, Hubbard JW, Korschinski ED et al. Pharmacokinetics of chlorpromazine and key metabolites. *Eur J Clin Pharmacol* 1993; 45: 563–569
- 739 Yin OQ, Wing YK, Cheung Y et al. Phenotype-genotype relationship and clinical effects of citalopram in Chinese patients. *J Clin Psychopharmacol* 2006; 26: 367–372
- 740 Yu DK, Dimmitt DC, Lanman RC et al. Pharmacokinetics of dothiepin in humans: a single dose dose-proportionality study. *J Pharm Sci* 1986; 75: 582–585
- 741 Zernig G, Hiemke C, Havemann-Reinecke U et al. Empfehlungen für die gutachterliche Bewertung von Medikamentenspiegeln in der Psychiatrie im gerichtsanhängigen Schadensfall. *Psychopharmakotherapie* 2009; 16: 57–64
- 742 Zernig G, Lechner T, Kramer-Reinstadler K et al. What the clinician still has to be reminded of. *Ther Drug Monit* 2004; 26: 582
- 743 Zernig G, Ng K, Hiemke C et al. Therapeutic drug monitoring-based clozapine dosing recommendations. *Ther Drug Monit* 2007; 29: 130–131

- 744 Zhao Q, Iyer GR, Verhaeghe T *et al*. Pharmacokinetics and safety of galantamine in subjects with hepatic impairment and healthy volunteers. *J Clin Pharmacol* 2002; 42: 428–436
- 745 Zhou SF. Polymorphism of human cytochrome P450 2D6 and its clinical significance: Part I. *Clin Pharmacokinet* 2009; 48: 689–723
- 746 Zhou SF. Polymorphism of human cytochrome P450 2D6 and its clinical significance: Part II. *Clin Pharmacokinet* 2009; 48: 761–804
- 747 Zhou SF, Liu JP, Chowbay B. Polymorphism of human cytochrome P450 enzymes and its clinical impact. *Drug Metab Rev* 2009; 41: 89–295
- 748 Zimmerman NP, Hickie IB, McGorry PD. Guidelines for youth depression: time to incorporate new perspectives. *Med J Aust* 2010; 193: 557
- 749 Zullino DF, Delessert D, Eap CB *et al*. Tobacco and cannabis smoking cessation can lead to intoxication with clozapine or olanzapine. *Int Clin Psychopharmacol* 2002; 17: 141–143