

## Does the physical disector method provide an accurate estimation of sensory neuron number in rat dorsal root ganglia?

Sibylle Delaloye<sup>b,1</sup>, Rudolf Kraftsik<sup>a,1</sup>, Thierry Kuntzer<sup>b</sup>, Ibtissam Barakat-Walter<sup>a,b,\*</sup>

<sup>a</sup> Department of Cell Biology and Morphology, University of Lausanne, 1005, Lausanne, Switzerland

<sup>b</sup> Laboratory of Neurology Research, University Hospital of Lausanne, 1011 Lausanne, Switzerland

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### ABSTRACT

The physical disector is a method of choice for estimating unbiased neuron numbers; nevertheless, calibration is needed to evaluate each counting method. The validity of this method can be assessed by comparing the estimated cell number with the true number determined by a direct counting method in serial sections. We reconstructed a 1/5 of rat lumbar dorsal root ganglia taken from two experimental conditions. From each ganglion, images of 200 adjacent semi-thin sections were used to reconstruct a volumetric dataset (stack of voxels). On these stacks the number of sensory neurons was estimated and counted respectively by physical disector and direct counting methods. Also, using the coordinates of nuclei from the direct counting, we simulate, by a Matlab program, disector pairs separated by increasing distances in a ganglion model. The comparison between the results of these approaches clearly demonstrates that the physical disector method provides a valid and reliable estimate of the number of sensory neurons only when the distance between the consecutive disector pairs is 60  $\mu\text{m}$  or smaller. In these conditions the size of error between the results of physical disector and direct counting does not exceed 6%. In contrast when the distance between two pairs is larger than 60  $\mu\text{m}$  (70–200  $\mu\text{m}$ ) the size of error increases rapidly to 27%.

We conclude that the physical disector method provides a reliable estimate of the number of rat sensory neurons only when the separating distance between the consecutive disector pairs is no larger than 60  $\mu\text{m}$ .

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### 1. Introduction

Because the total neuron number is an accepted parameter for evaluating neuronal survival or neuronal loss, several methods have been devised for determining the number of surviving neurons from histological sections (Tessler et al., 1985; Arvidsson et al., 1986; Liss et al., 1994; Ljungberg et al., 1999; Pu et al., 1999; McKay et al., 2002; Schmitz and Hof, 2005). Previously the main methods to estimate neuron number were based on two-dimensional microscopic images and involved assumptions about particle shape and distribution. The most commonly used method involved determining nucleolar number followed by the application of a correction factor to convert such counts to neuronal number (Abercrombie, 1946; Konigsmark, 1970). However, the shortcoming of these meth-

ods is that the assumptions are not completely met, and therefore, the methods are biased (de Groot and Bierman, 1986; Mouton, 2002; von Bartheld, 2002).

In 1984, a further counting method, called the physical disector method, was devised (Sterio, 1984), and this stereological method soon became the accepted way of determining the number of neurons. This method is particularly useful when objects to be counted are large relative to section thickness, and when their dimensions are unknown, or highly variable. Moreover, it can be used to count neurons without having to make any assumptions about their size, shape or orientation (West, 1999a, 2002; Guillery and August, 2002; Schmitz and Hof, 2005). This method based on a three-dimensional (3D) stereological probe, uses two sections, hence the name disector. Only the objects that are observable in the first section of the pair, but not observable in the second section are counted. This is a simple method for avoiding double counts, and its particular strength is that it is completely independent of the size and shape of the objects being counted. However, the use of this method requires that the investigator must be able to identify the sectional profiles that belong to the same object, and there must be at least one section in which it is possible to identify each object (West, 1993). This

\* Corresponding author at: Laboratoire de recherche neurologique, service de neurologie, CHUV, Rue du Bugnon 46, 1011 Lausanne, Switzerland.  
Tel.: +41 21 314 12 78; fax: +41 21 314 41 38.

E-mail address: [ibtissam.walter@unil.ch](mailto:ibtissam.walter@unil.ch) (I. Barakat-Walter).

<sup>1</sup> SD and RK have contributed equally to this work.

method has been widely applied to estimate the number of neurons in the nervous system. We also used the physical disector method to estimate the number of surviving axotomized sensory neurons in lumbar dorsal root ganglia (DRG), following adult rat sciatic nerve transection and local thyroid hormone (3,3',5-triiodo-L-thyronine, T<sub>3</sub>) administration (Schenker et al., 2003).

Although the physical disector method has important theoretical advantages, as summarized by several investigators (Howard and Reed, 1998; West, 1999b; Geuna, 2000), it is not a foolproof solution for every counting task (Saper, 1997). Therefore, each method should be validated by a calibration to reveal and eventually minimize the counting error (Coggeshall et al., 1990; Farel, 2002; von Bartheld, 2002). Obviously this is the first part of the experimental design, preceding the choice of the sample size for the experimental groups. The most precise method of validation is to compare the results of the method to be calibrated with the complete reconstruction of the organ (structure) of interest. The complete serial reconstruction gives the true number of neurons because there is no sampling or estimating (Pover and Coggeshall, 1991). Of course this calibration is rarely done because complete serial reconstruction requires intensive labour even using the modern powerful desktop computers and usually cannot be completed in a reasonable timeframe. What the investigator can do is to reconstruct a part of the organ or tissue and compare this to the results of other counting methods. With such an analysis it will be able to recognize, and eventually eliminate, most sources of bias.

In the present work, to test the accuracy of the physical disector method we compared the results of physical disector to those obtained by using direct counts derived from a voxel reconstruction of a part of a DRG. A stack of 200 adjacent sections from PBS-control and from T<sub>3</sub>-treated ganglia was used to reconstruct a volumetric dataset. On these stacks we estimated and counted the number of sensory neurons by physical disector and direct counting methods respectively. In addition, we devised a computer model of a ganglion in order to simulate the influence of separating distances

between the two consecutive disector pairs on the estimation of the number of neurons.

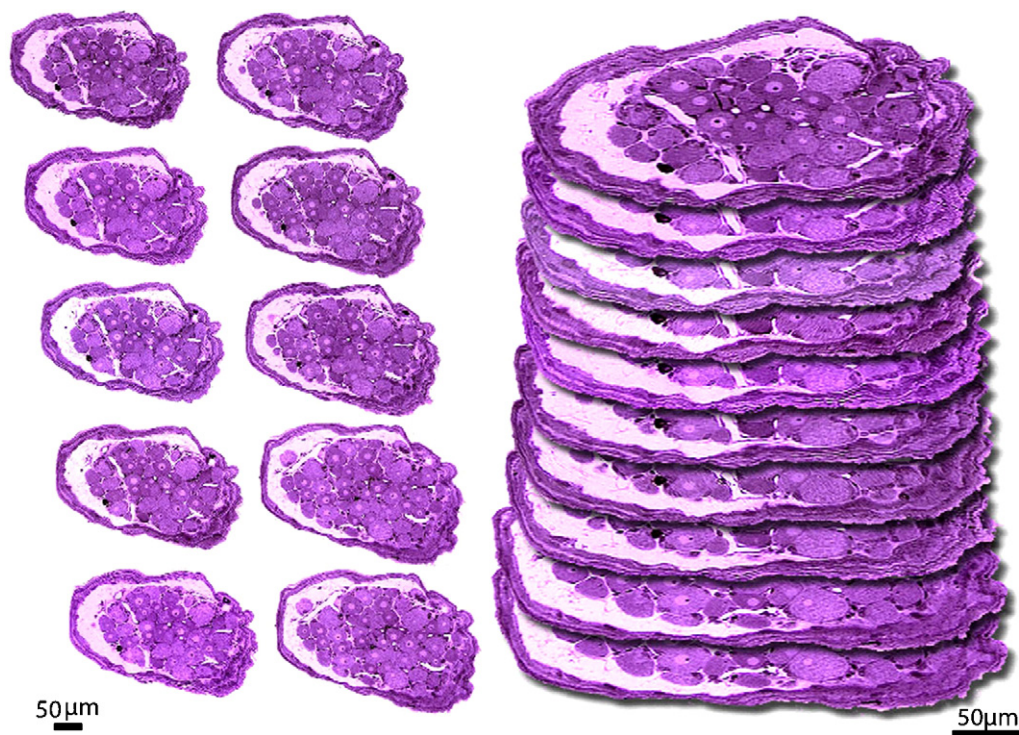
## 2. Materials and methods

### 2.1. Surgical procedures

All animal procedures were conducted according to the local guidelines for care and use of experimental animals. To reduce the number of animals used and to directly compare the results of this study with our previous results we used the same DRG sections described by Schenker et al. (2003). In brief, 10-week-old Wistar rats were anaesthetized. The skin of the right leg was cut followed by a longitudinal cut made through the biceps femoris muscle. The right sciatic nerve was transected near to the vertebral column, and a small piece (6 mm) of the nerve removed. The proximal and distal stumps of the transected nerve (approximately 1 mm) were secured into the ends of 10 mm of sterile silicone tubes with internal diameter of 1.5 mm and external diameter of 3 mm (Silclear, Medical Grade tubing, Midland, MI, U.S.A.). The transected nerve was held in place with three polyamide epineural sutures (8-0 Ethicon) at each end, leaving an 8 mm long gap between the nerve stumps. After placement, the tubes were filled with either a neutral pH sterile solution of 3,3',5-triiodo-L-thyronine, Sigma (1 mg of T<sub>3</sub> was dissolved in 1 ml 0.01 N NaOH, then the solution neutralized with 0.01 N HCl, final pH is 7.5), or with phosphate-buffered saline (PBS) as a control. In our experiments, the T<sub>3</sub>-treated rats, and PBS controls were taken from the same litter. The overlying muscle and skin of the thigh were sutured. The animals were allowed to survive for 45 days.

### 2.2. Processing for light microscopic and morphometric analysis

Forty-five days after surgery, rats were deeply anaesthetized, then transcardially perfused with 0.1 M phosphate-buffered saline



**Fig. 1.** Digital reconstruction of the ganglion from serial PBS-control DRG sections: the images of consecutive DRG sections were superimposed and matched one on the other using neuronal nuclear profiles as well as other landmarks such as the edge of the DRG section, blood vessels etc.

containing 0.1% heparin and 0.1% procaine, followed by a mixture of 3% paraformaldehyde and 1.5% glutaraldehyde in 0.1 M PBS at pH 7.4, as described in our previous work (Schenker et al., 2003). The axotomized transected lumbar dorsal root ganglia (L5) were removed from PBS-control and T<sub>3</sub>-treated rats, then post-fixed for 2 h at 4 °C in the respective fixative. After dehydration through increasing ethanol concentrations, the ganglia were oriented in the same direction along their long axis and embedded in Epon. Each entire ganglion was cut into serial longitudinal semi-thin sections of 1 μm thickness, comprising between 800 and 1000 sections. Then the sections were stained with 0.1% Toluidine blue for 10 s at 80 °C; this staining allows to visualize the nuclei and nucleolar profiles of the neuronal cells. Other ganglia were cut into serial cross semi-thin sections (1 μm thick) perpendicularly to the long axis of the dorsal root. In this case the number of the sections was 2400.

### 2.3. Digital reconstruction of the ganglion from serial images of DRG sections

About 200 adjacent sections (1 μm thick) were taken from one PBS-control and one T<sub>3</sub>-treated ganglion starting with the section in which the first neuronal profile was visible. Each histological section was scanned systematically and overlapping pictures of the microscope field were taken with a Sony 930XP video camera, at a magnification of 10× using the NeuroLucida (MicroBrightfield Inc., USA) program. The partial images were combined using the Adobe Photoshop (Adobe Systems Inc., USA) program, then these complete images of consecutive sections were best fit one on top of the other, based on their contours and nuclei profiles as well as other landmarks, (blood vessels, nucleoli, etc.) and using the transparency mode of the top layer (the overlay mode in Adobe Photoshop). The procedure was repeated throughout the whole stack to create a three-dimensional reconstruction (a volumetric dataset) of a part (1/5) of the ganglion. These aligned 3D stacks were used in the Imaris (Bitplane AG, Zurich, CH) program to obtain an interactive visualization of the reconstructed stack, and to control the quality of the 3D reconstruction.

Two methods were used either to determine or to estimate the total number of neurons in each 3D stack: direct counting and physical disector. To ensure that the counts were unbiased (blind), the observer did not know which treatment had been used for each set of images.

- (i) Direct counting method: this method is based on a direct identification and counting of nuclei from the digital images of the 3D image stacks using the confocal module of the NeuroLucida program (MicroBrightfield Inc.). In the PBS or T<sub>3</sub> DRG stacks, the nuclei, with identified nucleoli, were marked once, and the markers remained visible when the nucleus was followed in subsequent sections. New unmarked nuclei with nucleoli in the next sections were then added and followed until all the nuclei with nucleoli were identified. To facilitate the identification and marking of each nucleus found in the stack of 200 sections, each stack was divided into 5 smaller overlapping stacks. The nuclei were first followed and marked in each small stack. All the coordinates of the markers were saved in a file to be used in a computer model. Then, the total number of neurons in the whole PBS or T<sub>3</sub> stacks was counted using the NeuroExplorer (MicroBrightfield Inc.) program.
- (ii) Physical disector method: this method is based on sampling sections, called disector pairs. Twenty consecutive disector pairs were used from each stack. Since on the DRG sections the mean diameter of the nuclei (of large and small neurons) is around 20 μm, this permitted us to use sections separated by 8 μm as physical disector pairs. This design allows estimation

of the number of neurons by an unbiased approach. The estimation of the number of neurons was made using a protocol based on NeuroLucida software (MicroBrightfield Inc.) as described in our previous work (Schenker et al., 2003). To be sure that the random selection of the first section of the first pair gives an equal representation of all DRG regions being sampled, three independent estimations starting by the 3rd, the 5th or the 7th section of each stack were carried out. To investigate the role of the separating distance between the two consecutive pairs, we estimated the number of neurons by physical disector using different distances between the consecutive pairs (0, 10, 20, 30, 40 or 50 μm). We then compared neuron number obtained by physical disector and the true number of neurons determined by direct counting in the same DRG sections.

### 2.4. Creating a computer model by using the Matlab program

From the coordinates of markers recorded on the stack of 200 consecutive longitudinal DRG sections by the direct counting method, we created a computer model for the control and the T<sub>3</sub>-treated ganglia, using the Matlab (The MathWorks Inc., USA) program. Therefore, our computer model uses the three-dimensional coordinates of the markers of nuclei centers. The nuclei were simulated by software as spheres of 20 μm diameter, centred on the marker coordinates. The computer ganglion model is included in a rectangular parallelepiped of 200 μm depth, 1300 μm length and 800 μm width. In this ganglion model we simulated about 130 disector pairs of 10 μm thickness taken perpendicularly to the long axis of the reconstructed ganglion. We estimated the neuron number by the physical disector method by increasing the separating distance between two consecutive pairs from 0 to 200 μm and as a consequence, reducing the number of analysed pairs from 130 to 6. The results of these simulations were compared to the total number of neurons obtained by direct counting.

## 3. Results

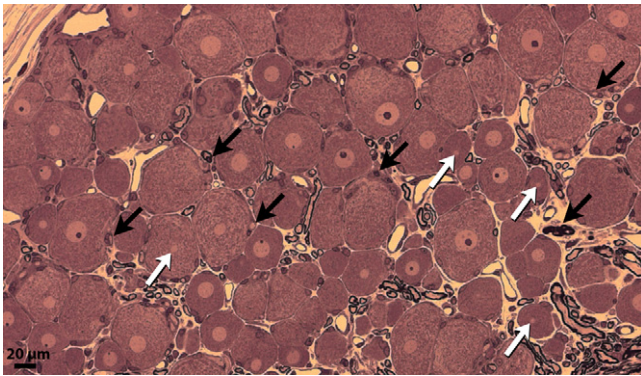
The major question addressed by the present paper is: does the physical disector method, used to estimate neuronal number, yield an accurate estimate when compared with direct counting? For this purpose the number of sensory neurons estimated by the physical disector method for various distances in defined stacks of DRG sections was compared with the true number of neurons determined by the direct counting method in the same DRG stacks.

### 3.1. Digital reconstruction of the ganglion from DRG serial sections

The images of 200 consecutive cross-sections taken either from one PBS-control or one T<sub>3</sub>-treated ganglion were used to create a 3D stack reconstruction (set of voxels) for each experimental condition. The images were superimposed one on the other using neuronal nuclear profiles as well as other landmarks such as the edge of the DRG section, blood vessels etc. (Fig. 1). Then the number of neurons was determined in each stack by direct counting and by the physical disector methods.

### 3.2. Determination of the true number of sensory neurons by direct counting method

The analysis of several sections by light microscopy showed that there are no detectable changes in the morphology of neurons between PBS-control (Fig. 1) and T<sub>3</sub>-treated DRG (Fig. 2). In both conditions sensory neurons can be easily identified and differentiated from non-neuronal cells (Schwann and satellite cells) on



**Fig. 2.** Micrograph of a histological section taken from  $T_3$ -treated dorsal root ganglion. The sensory neurons can easily be distinguished from satellite and Schwann cells (black arrows) by their pale round nuclei with one or two dark nucleoli. Even when the nuclei are cut at their ends they remain identifiable (white arrows).

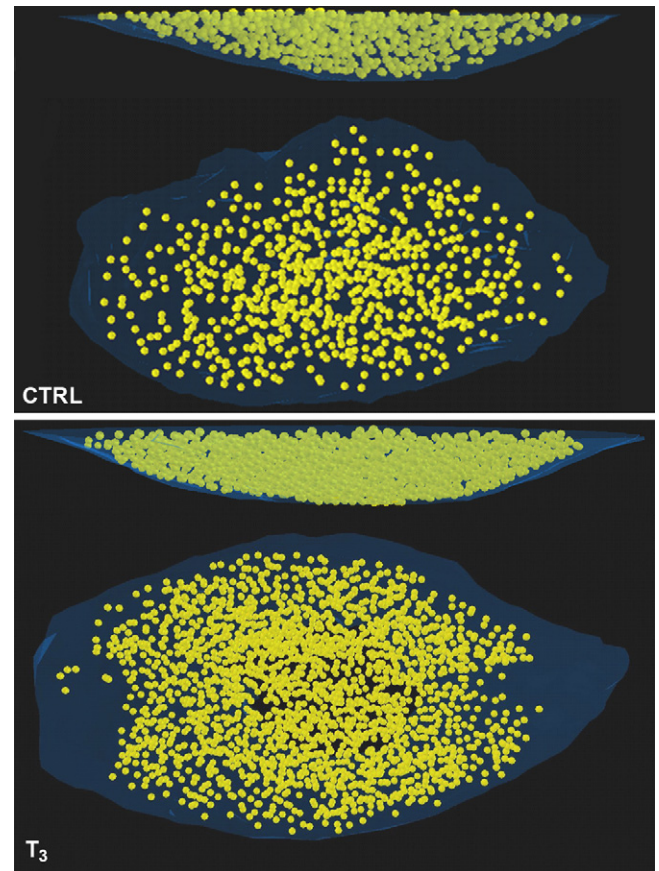
the basis of their large size and large nuclei bounded by a defined nuclear membrane, staining more lightly than the surrounding cytoplasm and containing at least one or more obvious nucleolar profiles which are darkly stained (Figs. 1 and 2).

The determination of the number of neurons in PBS-control and in  $T_3$ -treated stacks was based on the identification of the nuclei; each nucleus was identified, marked once at the level of the nucleolus and followed in the consecutive images of sections. On control or  $T_3$ -treated DRG sections, the diameter of the nucleus of sensory neurons varied from 4 to 24  $\mu\text{m}$ . The mean separating distance between the nuclei was around 24  $\mu\text{m}$  and varied from 12 to 50  $\mu\text{m}$ . Even when the nuclei are cut at their ends they remain identifiable without any difficulty (Fig. 2). All the nuclei identified in the PBS or in  $T_3$  DRG stacks were recorded. The total number of sensory neurons in each DRG stack corresponds to the total number of recorded nuclei. Each nucleus was visualized as a sphere using the Solid Views program (Microbrightfield Inc.). The lateral and top views of the total labelled nuclei showed a higher density in the  $T_3$ -treated stack (2057 nuclei) than in the PBS-control (1096 nuclei) stack (Fig. 3).

### 3.3. The reliability and validity of physical disector results based on direct counting

In a first step we used the images of the reconstructed sections to study the reliability of the disector method (see Section 2). The position within the ganglion of the horizontal sections constituting the first pair was selected randomly between the first 10th sections. To investigate the role of a random start on the evaluation of the neuron numbers, we estimated the number of sensory neurons beginning either with the 3rd, 5th or 7th section in the first 10 sections from each DRG stack. The results of the three estimations revealed that the choice of the first section does not change the estimated number of neurons. In fact, the number of estimated neurons corresponding to each disector pair is tightly similar (Fig. 4) in the three experimental counts either in PBS-control (Pearson  $R=0.98$ ;  $P<0.001$ ) or in  $T_3$ -treated (Pearson  $R=0.97$ ;  $P<0.001$ ) stacks.

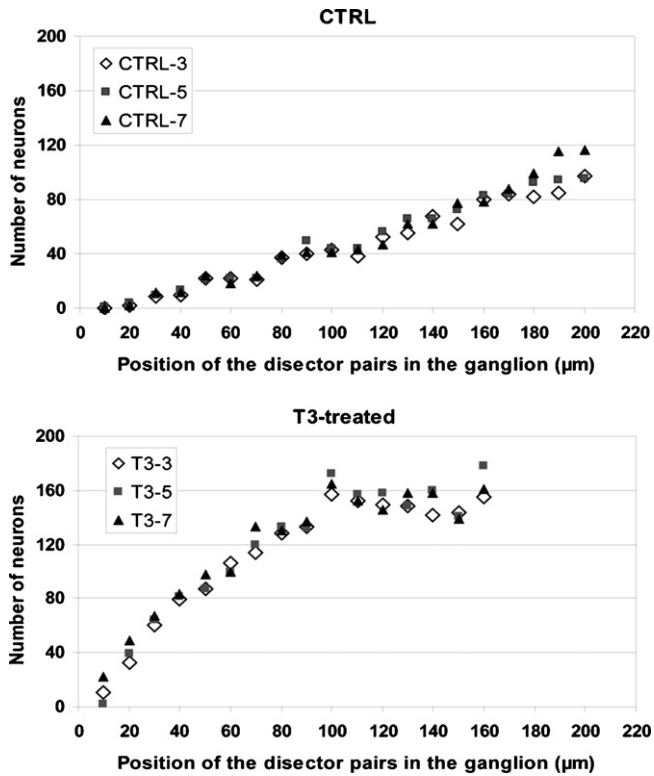
We also studied the role of distance on the number of estimated neurons by using increasing distances between the consecutive disector pairs (0, 10, 20, 30, 40 or 50  $\mu\text{m}$ ) and reducing the number of analysed pairs. The comparison between the number of neurons determined by direct counting and the number of neurons estimated by physical disector showed clearly that, in these conditions, the size of error does not exceed 6% (Fig. 5). Therefore, 50  $\mu\text{m}$  is an acceptable distance between two consecutive pairs.



**Fig. 3.** Lateral and top views of the total number of neuronal nuclei in 200 consecutive DRG sections taken from PBS-control (1096 nuclei) and  $T_3$ -treated (2057 nuclei) ganglia. Each nucleus is visualized as a sphere, using the Solid Views program (Microbrightfield Inc.). Note the higher density of nuclei in  $T_3$ -treated DRG stack compared with PBS-control stack.

### 3.4. Estimation of the number of neurons by simulation using a Matlab program and computer ganglion model

To study the influence of a large separating distance between the consecutive disector pairs on the estimated neuron number, we created a computer model of the ganglion based on the coordinates of nucleoli recorded in the direct counting of control and  $T_3$ -treated stacks. On this ganglion model we simulated 130 disector pairs perpendicularly to the long axis of the ganglia. Increasing the distances between the two consecutive pairs from 0 to 200  $\mu\text{m}$ , reduced the number of analysed pairs from 130 to 6 pairs. The comparison between the true number of neurons obtained by direct counting and the number of neurons estimated by physical disector in our simulation conditions confirmed that when the distance is relatively small (<60  $\mu\text{m}$ ), the variation and the size of error is acceptable and does not exceed 6%. In contrast when the distance between two pairs is larger than 60  $\mu\text{m}$  (70–200  $\mu\text{m}$ ) the size of error increases rapidly to 27% (Fig. 6). Moreover, when the separating distances between two consecutive disector pairs increase from 0 to 100  $\mu\text{m}$ , the total number of the tops (the nuclei present only in the top section of the disector pair) decreases in control PBS from 1096 to 99 and in  $T_3$ -treated DRG stacks from 2057 to 216 (Fig. 7). Therefore, the results of simulations on the ganglion model demonstrate that the reliability of the physical disector method is tightly related to the separating distance between the two consecutive pairs and to the number of analysed disector pairs.



**Fig. 4.** Three different estimations of the number of neurons in control and T<sub>3</sub>-treated DRG stacks beginning either with the 3rd, 5th or 7th section as the first section of the first disector pair. The three markers indicate the number of estimated neurons corresponding to each disector pair. Our results show that the numbers of estimated neurons corresponding to each disector pair (taken at regular interval through the ganglion) are highly correlated ( $R=0.98$ ,  $P<0.001$ ) in the three experimental conditions. Thus the random choice of the first section had no significant effect on the estimated number of neurons.

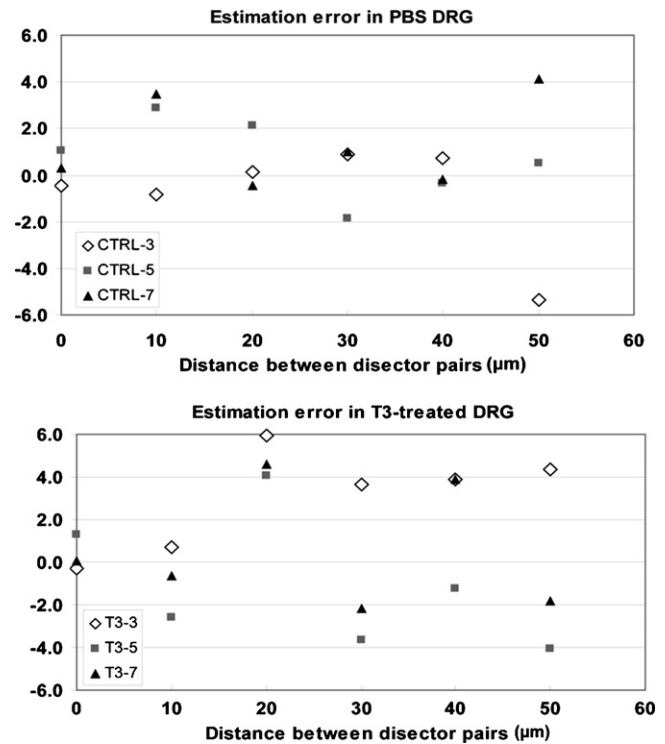
#### 4. Discussion

The result of the present study reveals that the physical disector method provides a valid and reliable estimate if the separating distance between two consecutive pairs is not larger than 60  $\mu\text{m}$  in DRG from the rat. This result is based on:

- (1) the reconstruction of a voxel-based, three-dimensional model from 200 semi-thin consecutive DRG sections (a volumetric dataset).
- (2) estimation and determination of the number of sensory neurons in each stack respectively by physical disector and direct counting methods.
- (3) simulation of 130 vertical disector pairs on a computer ganglion model, to study the effect of large distance separating the consecutive disector pairs on the number of estimated neurons.

##### 4.1. Neuron counts and stereological techniques

The counts of nerve cells, in sections prepared for light microscopy, allow the comparison of experimental and control conditions, and help to assess the loss of neurons following nerve lesion or neuropathy. If the counting is based on reliable methods, it provides crucial information; if not, it can be seriously misleading. The use of inappropriate stereological methods usually results in a high variation in the neurons counts (Guillery and August, 2002; Schmalbruch, 1987). At the present time, no method of counting can be certified as free from error, however, the emergence of a new generation of stereological techniques has provided a more pre-



**Fig. 5.** Size of error between the results of physical disector and direct counting. The comparison between the true number of neurons determined by direct counting and the number of neurons estimated by physical disector using various distances between the consecutive pairs shows that the size of error does not exceed 6% when the distances vary from 0 to 50  $\mu\text{m}$ .

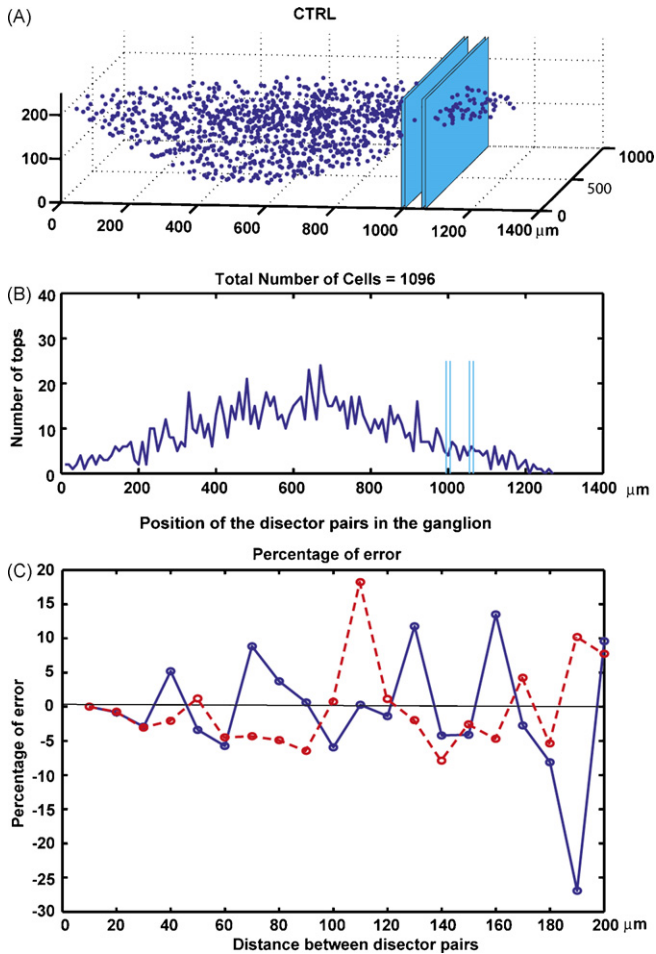
cise estimation of the number of neurons when they are correctly applied.

##### 4.2. The importance of the choice of methods and other parameters to estimate the number of neurons

One of the most important tasks in all quantitative approaches is choosing the method to determine the number of cells. Sources of systematic bias in morphometric data arise from faulty models or methods, assumptions and correction factors (Mouton, 2002). The most straightforward, accurate, and reliable means to determine neuron number is to follow each neuron through consecutive sections (Saper, 1997; von Bartheld, 2001). Even under the most favourable circumstances, this method is tedious and therefore it is inconceivable to use it in all situations.

The physical disector proposed in 1984 by Sterio (1984), Braendgaard and Gundersen (1986), Coggeshall (1992) has been shown to be useful to estimate the neuron number from histological sections (Gundersen et al., 1988; Saper, 1996; Coggeshall and Lekan, 1996). This method is based on sampling sections called disector pairs separated by a known distance (Tandrup, 1993, 2004; West, 2002). Since the physical disector method is a design-based sampling, several factors play a critical role in the reliability and validity of the results. The aspect of the organ which is used, the ability to identify the cells, the even or uneven distribution of the cells inside the organ, the number of disector pairs analysed, are all factors which may play an important role in the reliability of the results.

To have a good estimations using the physical disector method, the distance between the two sections from the same pair, should be proportionally linked to the diameter of the counted cells. The recommended protocol uses a separating distance roughly equiv-

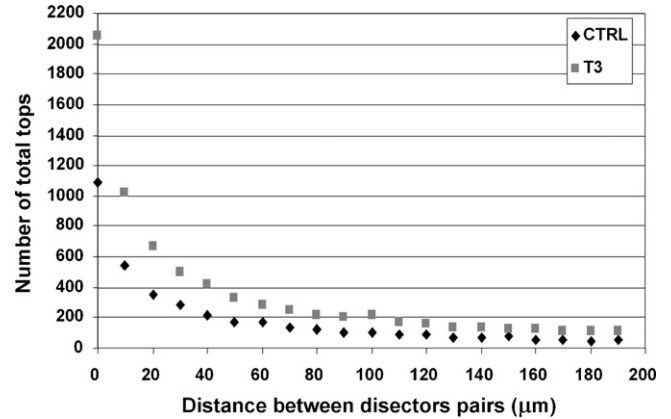


**Fig. 6.** Computer simulation of 130 vertical disector pairs taken perpendicularly to the long axis of the ganglion model. (A) 3D-view of the PBS-control ganglion model based on direct counting results with two simulated disector pairs. Our computer model uses the three-dimensional coordinates centers of the nuclei. The nuclei were simulated as spheres of 20  $\mu\text{m}$  diameter. The computer ganglion model is displayed in a rectangular parallelepiped of 200  $\mu\text{m}$  depth, 1300  $\mu\text{m}$  length and 800  $\mu\text{m}$  width. One disector pair is defined by two 1  $\mu\text{m}$  thick sections separated by 8  $\mu\text{m}$ . The separating distance between the two example disector pairs is around 50  $\mu\text{m}$ . (B) Diagram illustrating the number and the distribution of “tops” at each level of simulated physical disector pair on PBS-control ganglion. (C) Size and variation of error between the results of direct counting and the results of physical disector on the simulated models of ganglia using an increasing separation distance between the two consecutive pairs (from 0 to 200  $\mu\text{m}$ ). Our results show that the large distance between two consecutive pairs increases the variation and the size of error. Solid line: PBS-control, dashed line:  $T_3$ -treated DRG.

alent to half the diameter of nucleus. If the separating distance between the two sections is larger than this, the risk is that small cells hidden between the two sections, will not be counted. In contrast, if the distance is very small, only few tops may be counted (Coggeshall and Lekan, 1996; West, 2002).

The fact that sensory neurons in DRG can be easily identified facilitates the counting of neuron number (La Forte et al., 1991; Tandrup, 2004). Another advantage of DRG is that we can precisely match the two sections from the same disector pair by using the edge of the DRG section in addition to other landmarks such as the nuclear profiles and blood vessels etc.

However, the uneven distribution of sensory neurons inside the ganglion is well described, which is due to the presence of bundles of fibers intermingled with the neuron cell bodies. In addition, the histological sections taken from the middle of the ganglion contain a high number of neurons while the sections taken from



**Fig. 7.** Relation between the total number of “tops” and the increasing separating distance between the two consecutive disector pairs, derived from the computer ganglion model of PBS-control and  $T_3$ -treated DRG.

the two ends of the ganglion contain a small number of neurons (Coggeshall et al., 1994; Schenker et al., 2003; Tandrup, 2004). To give an equal opportunity of all the DRG regions to be sampled, the systematic random sampling must be applied (West, 1993, 2002; Geuna, 2000). In our present study we made three independent estimates by taking the 3rd, 5th or 7th section as the first section constituting the first pair. Our results revealed that the number of estimated neurons is tightly similar in the three experimental counts either in PBS-control or in  $T_3$ -treated DRG section stacks (Fig. 5). Therefore, our data confirmed that systematic random sampling gives equal probability to the different part of the ganglion to be sampled, and consequently produces reliable results.

Logically to reduce the impact of an uneven distribution of neurons within the ganglion on the estimated number of neurons, the number of analysed disector pairs must be increased and the separating distance between two consecutive pairs reduced. In this regard, several investigators had reported that the reliability of the physical disector method is increased when the number of disector pairs is increased, especially when the distribution of neurons is not uniform in the organ. Sampling only a few small regions within the ganglion could lead to substantial error of estimates (Popken and Farel, 1996, 1997; Farel, 2002). However, because the analysis of a large number of disector pairs is time consuming, some investigators examine only a small number of samples, and on the basis of the data obtained, they make statements about the total cell number. This process of extension from a small number of samples to the general number can induce a large error (Popken and Farel, 1996). However, to our knowledge no study was made to determine the separating distance between the disector pairs, which allows the physical disector method to give precise results in the rat DRG. In this study, the use of two approaches (direct counting and physical disector) revealed that 50  $\mu\text{m}$  is an acceptable separating distance between two consecutive pairs to allow the physical disector method to provide a valid result. In fact, the comparison between the neuron number estimated by physical disector and the true number of neurons determined by the direct counting method on the same DRG stack clearly showed that if the separating distances between the two consecutive disector pairs are 50  $\mu\text{m}$  or smaller (0–50  $\mu\text{m}$ ), the size of error between the results of the two methods does not exceed 6%, which seems a reasonable bias (von Bartheld, 2002).

Furthermore, the devised computer model based on the coordinates of the nuclei recorded in the direct count and the simulation of 130 disector pairs perpendicular to the long axis of the dorsal root allowed to test the effect of increasing distance (0–200  $\mu\text{m}$ )

between the disector pairs on the estimated number of neurons. Here again the results of this approach demonstrated that when the two consecutive disector pairs are separated by 50  $\mu\text{m}$ , the variation and the size of error remained small (less than 6%). In contrast, when the separating distance between consecutive pairs is larger than 60  $\mu\text{m}$  (70–200  $\mu\text{m}$ ), the variation and the size of error increased rapidly (27%) even when the number of analysed disector pairs is higher than 6 (18–6 pairs). The rationale for using the computer model of a part of the ganglion comes from the fact that the real complete serial reconstruction requires intensive labour and usually cannot be completed in a reasonable time. Thus the computer ganglion model allows to test the effect of some parameters on the estimated cell number and to confirm/or disprove the results of other methods without consuming much of time.

Therefore, on the basis of our data we conclude that to obtain accurate and reliable results by using the physical disector method we must analyse a large number of disector pairs separated by a distance which should not exceed 60  $\mu\text{m}$ . These recommendations are also valid when the cross semi-thin sections are cut perpendicularly to the long axis of the dorsal root. In fact, the estimation of neuron number by the physical disector method in a ganglion sectioned perpendicularly to the long axis and using 50  $\mu\text{m}$  as a separating distance between the consecutive pairs provided an estimated total neuron number similar to the mean number of neurons which we obtained in our previous work (Schenker et al., 2003). However, the disadvantage of using cross-sections is to cut a large number of sections which increases the histological workload several folds. Furthermore, in the latter case the obligation arises to analyse a large number of disector pairs (48 pairs) compared to 20 disector pairs when the DRG is sectioned parallel to the long axis. These remarks agree with observations reported by Tandrup (2004).

#### 4.3. Physical disector and direct counting methods confirmed that $T_3$ -treatment reduced the loss of axotomized sensory neurons

Since our interest is focused on the study of the effect of a local administration of  $T_3$  on the regeneration of transected adult rat sciatic nerve (Voinesco et al., 1998; Barakat-Walter, 1999; Schenker et al., 2002), we used the physical disector method to estimate the total number of surviving neurons in lumbar DRG in control and experimental animals. To this aim we analysed a large number of disector pairs separated by 50  $\mu\text{m}$  and we found that the mean number of neurons in PBS-control is  $8744 \pm 748$  and the mean number of neurons in  $T_3$ -treated ganglion is  $12046 \pm 930$  (Schenker et al., 2003). In this study using the same DRG sections, we found that the size of error between the results of direct counting and physical disector method does not exceed 6% (522 or 720 respectively). This small percentage of error cannot account for the difference between the neuron number in control and  $T_3$ -treated ganglia. Therefore, we can infer that  $T_3$ -treatment rescues a large number of axotomized sensory neurons from death.

In conclusion, the volumetric datasets reconstructed from 200 consecutive DRG semi-thin sections (stacks) on which the number of sensory neurons was estimated and determined by the physical disector and direct counting methods allowed us to demonstrate clearly that physical disector method provides valid and reliable results when the separating distance between the consecutive pairs is no larger than 60  $\mu\text{m}$ . Moreover, the simulation of 130 disector pairs on a computer ganglion model showed that when the separating distance between consecutive pairs is larger than 60  $\mu\text{m}$ , the variation and the size of error increased rapidly to 27%.

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