



## Screening of a PDE-focused library identifies imidazoles with *in vitro* and *in vivo* antischistosomal activity

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### ABSTRACT

We report the evaluation of 265 compounds from a PDE-focused library for their antischistosomal activity, assessed *in vitro* using *Schistosoma mansoni*. Of the tested compounds, 171 (64%) displayed selective *in vitro* activity, with 16 causing worm hypermotility/spastic contractions and 41 inducing various degrees of worm killing at 100  $\mu$ M, with the surviving worms displaying sluggish movement, worm unpairing and complete absence of eggs. The compounds that did not affect worm viability ( $n = 72$ ) induced a complete cessation of ovipositing. 82% of the compounds had an impact on male worms whereas female worms were barely affected. *In vivo* evaluation in *S. mansoni*-infected mice with the *in vitro* 'hit' NPD-0274 at 20 mg/kg/day orally for 5 days resulted in worm burden reductions of 29% and intestinal tissue egg load reduction of 35% at 10 days post-treatment. Combination of praziquantel (PZQ) at 10 mg/kg/day for 5 days with NPD-0274 or NPD-0298 resulted in significantly higher worm killing than PZQ alone, as well as a reduction in intestinal tissue egg load, disappearance of immature eggs and an increase in the number of dead eggs.

### 1. Introduction

This study phenotypically evaluated the antischistosomal activity potential of 265 compounds from a cyclic nucleotide phosphodiesterase (PDE) focused library constructed by the PDE4NPD consortium ([www.PDE4NPD.eu](http://www.PDE4NPD.eu)), which focused on exploring PDEs as potential drug targets for several parasite species, including *S. mansoni*. Several PDE inhibitors are currently in use as therapeutic agents for various human conditions, acting by targeting specific PDE isoenzymes and thereby the breakdown of cyclic nucleotides (cAMP, cGMP) and thus prolonging their biological effects (Ghosh et al., 2009). Pharmacologically, PDEs gained great interest as drug targets for a large variety of clinical conditions including their antiparasitic potential (Shakur et al., 2011). The *in vitro* screening campaign used *S. mansoni* worm killing as the

primary parameter with *in vivo* follow-up evaluation of selected actives in a mouse model.

Schistosomiasis has among the highest morbidity of the world's neglected tropical diseases, causing an estimated 4,026,000 disability-adjusted life years (DALYs). In socio-economic terms, public health importance and prevalence in the developing world, it ranks second only to malaria. More than 780 million people are at risk and approximately 261 million are infected in 78 countries of which 85% reside in sub-Saharan Africa (WHO, 2017). No vaccine is yet available and almost all control initiatives rely on praziquantel (PZQ), which has been considered the drug of choice since the 1970s as it is highly effective after a single oral dose against all *Schistosoma* species that are pathogenic to humans. In Egypt, as in most African countries, control measures rely on campaigns of mass PZQ administration targeting high-

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**Table 1**

Compound prioritization of antischistosomal potency based principally on killing of mature *S. mansoni* worms *in vitro* and their toxicity to MRC-5 human lung fibroblasts.

	Serial No	PDE inhibitors	Worm killing at different concentrations (% of total; male + female)					EC <sub>50</sub> (μM)	Uncoupling & complete absence of eggs			Reduction in number of eggs (%)			MRC-5 EC <sub>50</sub> (μM)
			100 μM	50 μM	25 μM	10 μM	5 μM		25 μM	10 μM	5 μM	25 μM	10 μM	5 μM	
Killing from 50% to 100% (Class I)	1	NPD-0274 <sup>a</sup>	56	47	13	0	0	72	No	No	No	79	50	33	> 64
	2	NPD-0029	77	27	0	0	0	69	No	No	No	40	37	35	30
	3	NPD-0356	50	39	14	0	0	89	No	No	No	60	10	0	60
	4	NPD-0048	63	25	19	0	0	79	No	No	No	42	50	33	9
	5	NPD-1012	62	0	0	0	0	99	No	No	No	67	32	38	ND
	6	NPD-1246	100	53	0	0	0	50	Yes	No	No	100	20	10	> 64
	7	NPD-1253	50	37	0	0	0	91	Yes	No	No	100	21	29	35
	8	NPD-2904	50	0	0	0	0	100	Yes	Yes	Yes	100	100	100	> 64
	9	NPD-1014	56	37	0	0	0	81	No	No	No	0	0	0	ND
	10	NPD-1211	52	30	0	0	0	91	Yes	No	No	100	13	8	> 64
	11	NPD-1085	50	7	0	0	0	100	No	No	No	25	17	8	> 64
	12	NPD-1013	56	0	0	0	0	99	No	No	No	25	0	0	ND
Egg insult (Class II)	13	NPD-0298 <sup>a</sup>	16	0	0	0	0	> 100	Yes	Yes	No	100	100	77	> 64
	14	NPD-0264 <sup>a</sup>	6	0	0	0	0	> 100	No	No	No	79	75	71	20

All compounds listed displayed 100% reduction in egg numbers with uncoupling at 100 μM and 50 μM.

<sup>a</sup> Compounds selected for *in vivo* testing. Yes: uncoupling and complete absence of eggs. No: no uncoupling and incomplete absence of eggs. ND, not determined.

**Table 2**

Percentage of male and female worm killing at different concentrations of test compounds.

Serial No	PDE inhibitors	% Worm killing												
		Male worms						Female worms						
		100 μM	50 μM	25 μM	10 μM	5 μM	EC <sub>50</sub> male (μM)	100 μM	50 μM	25 μM	10 μM	5 μM	EC <sub>50</sub> female (μM)	
Killing from 50% to 100% (Class I)	1	NPD-0274*	100	88	25	0	0	32	0	0	0	0	0	NE
	2	NPD-0029	91	33	0	0	0	59	0	0	0	0	0	NE
	3	NPD-0356	88	71	29	0	0	36	0	0	0	0	0	NE
	4	NPD-0048	100	50	33	0	0	41	14	0	0	0	0	> 100
	5	NPD-1012	60	50	0	0	0	68	0	0	0	0	0	NE
	6	NPD-1246	100	50	0	0	0	50	100	57	0	0	0	50
	7	NPD-1253	100	66	0	0	0	49	0	0	0	0	0	NE
	8	NPD-2904	73	0	0	0	0	97	0	0	0	0	0	NE
	9	NPD-1014	100	67	0	0	0	49	0	0	0	0	0	NE
	10	NPD-1211	88	56	0	0	0	48	56	13	0	0	0	93
	11	NPD-1085	78	13	0	0	0	76	0	0	0	0	0	NE
	12	NPD-1013	70	0	0	0	0	98	0	0	0	0	0	NE
Egg insult (Class II)	13	NPD-0298*	33	0	0	0	0	> 100	0	0	0	0	0	NE
	14	NPD-0264*	12	0	0	0	0	> 100	0	0	0	0	0	NE

Controls: PZQ was 100% effective against both male and female worms at all concentrations tested (100, 50, 25, 10 and 5 μM), and DMSO (negative control, solvent; 0.125%–2%) had no effect on worm viability.

NE, no effect.

risk groups (WHO, 2012). PZQ possesses positive features with respect to safety, efficacy, cost and ease of distribution (Cioli et al., 2014) but its efficacy is dependent on the age of the infection, the sex of the worms and their paired or unpaired status (Pica-Mattocchia and Cioli, 2004). Immature worms (between 1 and 5 weeks after infection) are much less sensitive to PZQ and hence such infections require re-treatments. Moreover, findings denoting a possible threat of resistance development were reported in mice under laboratory conditions (Fallon and Doenhoff, 1994; Ismail et al., 1994; Sabra and Botros, 2008; Liang et al., 2011) and during the intra-molluscan phase (Couto et al., 2011). At the clinical level, reduced susceptibility to PZQ in *S. mansoni* field isolates has been reported (Gryseels et al., 2001; William et al., 2001; Cioli et al., 2004; Doenhoff et al., 2008; Melman et al., 2009; Mwangi et al., 2014) but goes largely unnoticed in the mass-administration campaigns, in part because of the insufficient efforts to monitor PZQ resistance. For the above reasons, the search for new antischistosomal agents represents an overriding priority. To date, a large number of compounds have been identified as potential antischistosomal agents but none has yet represented a suitable alternative to PZQ.

## 2. Materials and methods

### 2.1. Drugs and compounds

Praziquantel (PZQ) tablets (Distocide<sup>®</sup>) were obtained from Egyptian International Pharmaceutical Industries Company (EIPICO). The experimental compounds were prepared at University of Amsterdam (VUA), University of Antwerp (UA) and Centro de Investigaciones Biológicas (CIB–CSIC) and have a purity ≥ 95% by HPLC. Synthesis of the imidazoles tested *in vivo* (NPD-0274, NPD-0298 and NPD-0264) was performed according to described procedures (García et al., 2017).

### 2.2. Cytotoxicity on human lung fibroblasts

MRC-5 human lung fibroblasts were cultured in MEM medium supplemented with 20 mM L-glutamine, 16.5 mM NaHCO<sub>3</sub> and 5% fetal calf serum (FCS). Assays were performed at 37 °C and 5% CO<sub>2</sub> in 96-well tissue culture plates with confluent monolayers. After 7 days of incubation, cell viability was assessed after addition of resazurin and

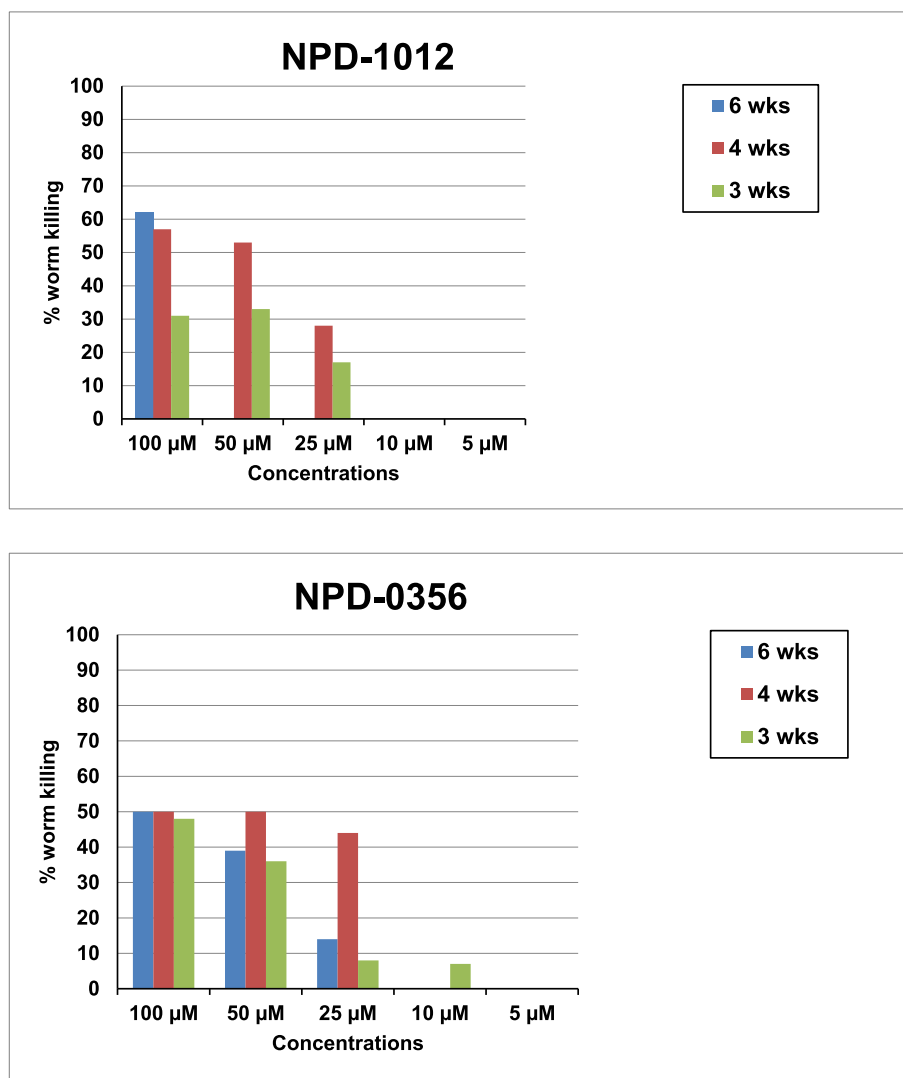


Fig. 1. Effect of PDE inhibitors NPD-1012 and NPD-0356 on worm killing of different *S. mansoni* maturity stages.

fluorescence reading.

### 2.3. In vitro studies

#### 2.3.1. Preparation of compounds

5 mM stock solutions of PZQ and test compounds were prepared in DMSO. At the day of experiment, 100 μM, 50 μM, 25 μM, 10 μM and 5 μM concentrations were freshly prepared in RPMI-1640 medium. All compounds were initially tested at 100 and 50 μM; those showing worm killing were further tested at 25, 10 and 5 μM.

#### 2.3.2. Worm killing

Six to eight worms (obtained from Schistosome Biology Supply Center (SBSC) of the Theodor Bilharz Research Institute (TBRI)), including a minimum of one worm couple, were placed in each well of a 12-well tissue culture plate containing 2 ml of fresh RPMI-1640 medium supplemented with glutamine, 20% newborn calf serum and antibiotics (streptomycin, penicillin (2 ml/100 ml) and gentamicin (200 μl/100 ml)), and the indicated concentration of the test compounds (Pica-Mattoccia and Cioli, 2004; Botros et al., 2005). The plates were incubated overnight at 37 °C and 5% CO<sub>2</sub>. Worms were examined by phase-contrast microscopy, 24 h after the start of the incubation, washed thrice with sterile saline, fresh medium was added and the incubation was continued. After 48 h, worm motility was observed and

72 h later, medium was changed again. After 96 h (end of observation period) worms were microscopically examined for motility and appearance. Each concentration was tested in duplicate. The final recording of percent worm mortality was determined as the number of dead worms [contracted and opaque] divided by the total number of worms × 100. Negative controls using pure medium without test compound or medium with DMSO (2%), and positive control media containing parallel concentrations of PZQ were tested in parallel. Immature, early mature and mature *S. mansoni* worms were 3 weeks, 4 weeks and 6–7 weeks old respectively.

#### 2.3.3. Worm ovipositing

Compounds showing worm killing, sluggish movement and/or unpairing were tested for inhibitory effect on ovipositing using 12-well tissue culture plates with each well containing at least one worm couple. Each concentration was tested in duplicate and eggs were counted a first time after 72 h, upon which the eggs were discarded and the medium changed. After 96 h (end of observation period), the newly deposited eggs were counted and the total number was calculated for each concentration tested.

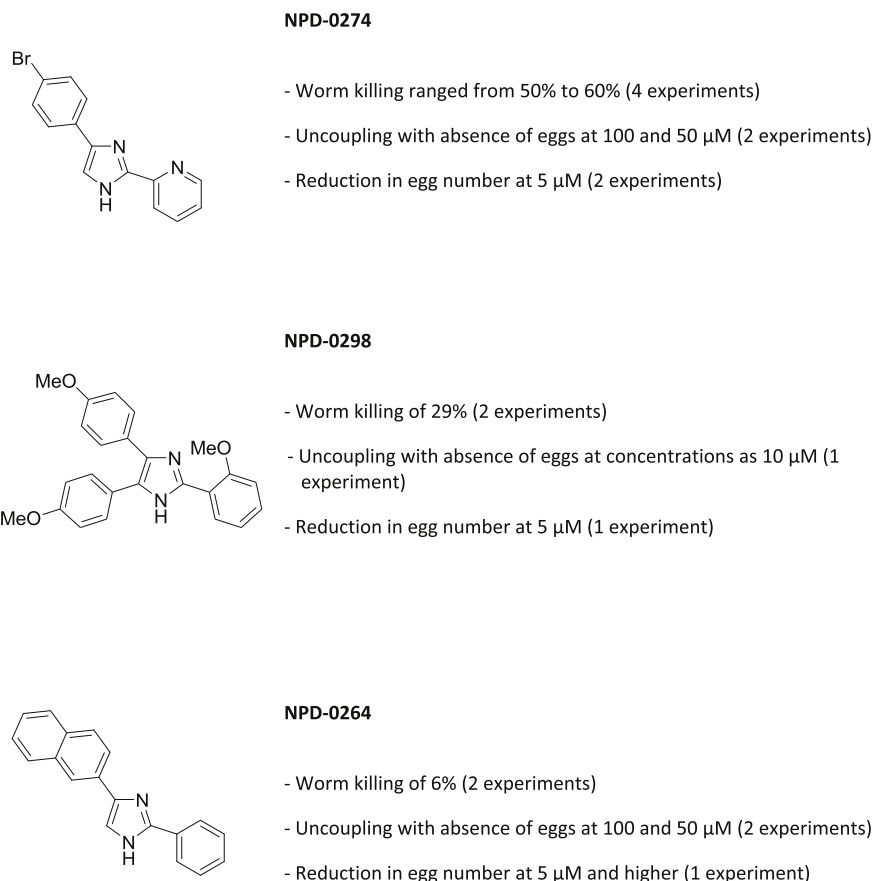


Fig. 2. *In vitro* findings for selected compounds for *in vivo* experiments.

## 2.4. *In vivo* antischistosomal activity

### 2.4.1. Experimental infection of animals

Male Swiss albino mice (CD-1) obtained from SBSC and weighing 18–20 g were housed under environmentally controlled room temperature of 20–22 °C, 12 h light/dark cycle and 50–60% humidity with food and water *ad libitum* throughout the acclimatization and experimental periods. Mice were infected with *S. mansoni* cercariae (provided by SBSC) using body immersion (Liang et al., 1987) by exposure to  $80 \pm 10$  cercariae/mouse. All the animal experiments were conducted in accordance with the Guide for Care and Use of Laboratory Animals and were approved by the Institutional Review Board of TBRI.

### 2.4.2. Test compounds and experimental design

NPD-0274, NPD-0298, NPD-0264 and PZQ were freshly suspended in 2% Cremophore-EL (Sigma-Aldrich, St Louis, MO, USA). Infected mice were divided into 8 groups: the first three groups were treated with NPD-0274, NPD-0298 and NPD-0264, respectively, at 20 mg/kg/day for 5 days. Group 4 was treated with PZQ at 10 mg/kg/day for 5 days. Groups 5, 6 and 7 were treated with each of the test compounds combined with PZQ, each at 10 mg/kg/day for 5 days starting from the 7<sup>th</sup> week post-infection. Group 8 was the vehicle control. To minimize first-pass elimination, the CYP450 inhibitor aminobenzotriazole (ABT) was administered at 100 mg/kg/day for 5 days 2 h prior to each compound administration. All drug administrations were performed orally.

### 2.4.3. Parasitological criteria for cure

Ten days post-treatment, all mice were sacrificed and perfused, and the number of worms recovered (worm burden) was quantified and sexed (Duvall and De Witt, 1967). The number of eggs per gram of liver or intestinal tissue was counted (Cheever, 1968). The percentage of egg developmental stages (oogram pattern) was studied (Pellegriano et al.,

1962), in which eggs at different stages of maturity (from I to IV) were identified and counted. Mature eggs and dead eggs (granular, dark, and semi-transparent) were also counted in three fragments of intestine and the mean number of each stage was calculated.

## 2.5. Statistical analysis

The percentage reduction of worm or egg burden in each treated group was calculated. The 50% effective concentration ( $EC_{50}$ ) or dose ( $ED_{50}$ ) was calculated using Prism (GraphPad; Version 5.0) software using a variable slope for the sigmoidal curve with an upper limit of 100%. Results are expressed as mean  $\pm$  SEM. A two-tailed, unpaired Student's *t*-test was used to detect the significance of difference between the means of different groups. Results are considered significant if *P* value is  $< 0.05$ .

## 3. Results

### 3.1. *In vitro* activity against *S. mansoni*

265 compounds from a PDE-targeting library were initially screened for worm killing at 100  $\mu\text{M}$ . Antischistosomal effects were recorded in 171 compounds (64%) as worm killing, sluggish worm movement, unpairing, and absence or reduction in egg numbers; 16 resulted in worm hypermotility/spastic contraction. Based on these *in vitro* findings, the compounds were categorized into five classes: Class-I and –II showed worm killing while the other three classes did not show worm killing but impacted ovipositing, resulting in either a reduction or the complete absence of eggs. Class-I compounds (12/171) showed worm killing of 50–100% with  $EC_{50}$  values in the range of 50–100  $\mu\text{M}$  (Table 1). Class-I consisted of known PDE-like scaffolds, including phthalazinones (4  $\times$ ), imidazoles (2  $\times$ ) and a quinazoline (Table S1).

## NPD-0274

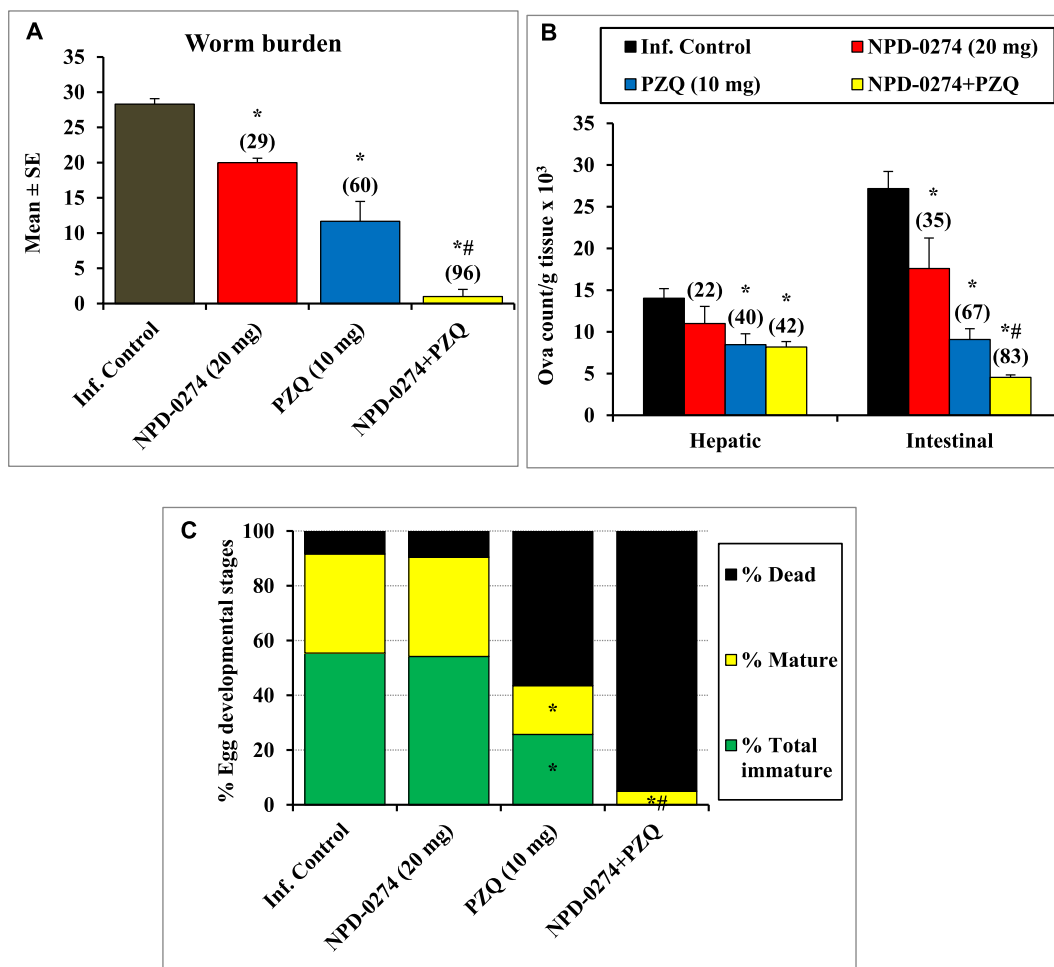


Fig. 3. (A) Worm burden, (B) tissue egg load and (C) oogram pattern of NPD-0274 sacrificed 10 days post end of treatment when used alone at 20 mg/kg/day or in combination with PZQ at 10 mg/kg/day for 5 days. \*significantly different from infected control at  $P < 0.05$ , #significantly different from PZQ group at  $P < 0.05$ . Numbers above columns and between parentheses represent % change from infected control group. ABT (100 mg/kg orally) was administered 2 h prior to administration of NPD-0274 and PZQ, whether alone or in combination. Error bars represent SEM.

These 12 hits were further investigated for dose-related worm pairing, and reduction or complete absence of eggs. The Class-II compounds NPD-0298 and NPD-0264, displaying strong effects on ovipositing and slight effects on worm killing, were also included in the *in vitro* follow-up study (Table 2).

All 14 compounds displayed 100% uncoupling of worm pairs with complete absence of eggs at both 100  $\mu$ M and 50  $\mu$ M, and several even impacted on egg numbers at 5  $\mu$ M, a concentration with no effect on worm viability at all. NPD-0298 and NPD-0264, which only had a slight effect on viability at 100  $\mu$ M, strongly reduced ovipositing down to 5  $\mu$ M. For NPD-0298 this may be mostly explained through worm uncoupling; for NPD-0264, however, there was no uncoupling below 50  $\mu$ M while egg production was still severely affected. A similar pattern was observed with NPD-0274 and some other compounds, indicating a specific effect on egg depositing rather than on viability or coupling (Table 1).

The above screening was conducted on mixed groups of males and females where it was noted that the male worms were generally much more severely affected than the female worms. We therefore re-analyzed worm killing for the hits in Table 1, this time performing parallel incubations with male and female populations. Out of the 14 compounds (Table 1), 11 displayed no activity on females even at 100  $\mu$ M. However, NPD-1246 had 100% worm killing on both sexes while NPD-

1211 displayed moderate effects at 100  $\mu$ M (Table 2).

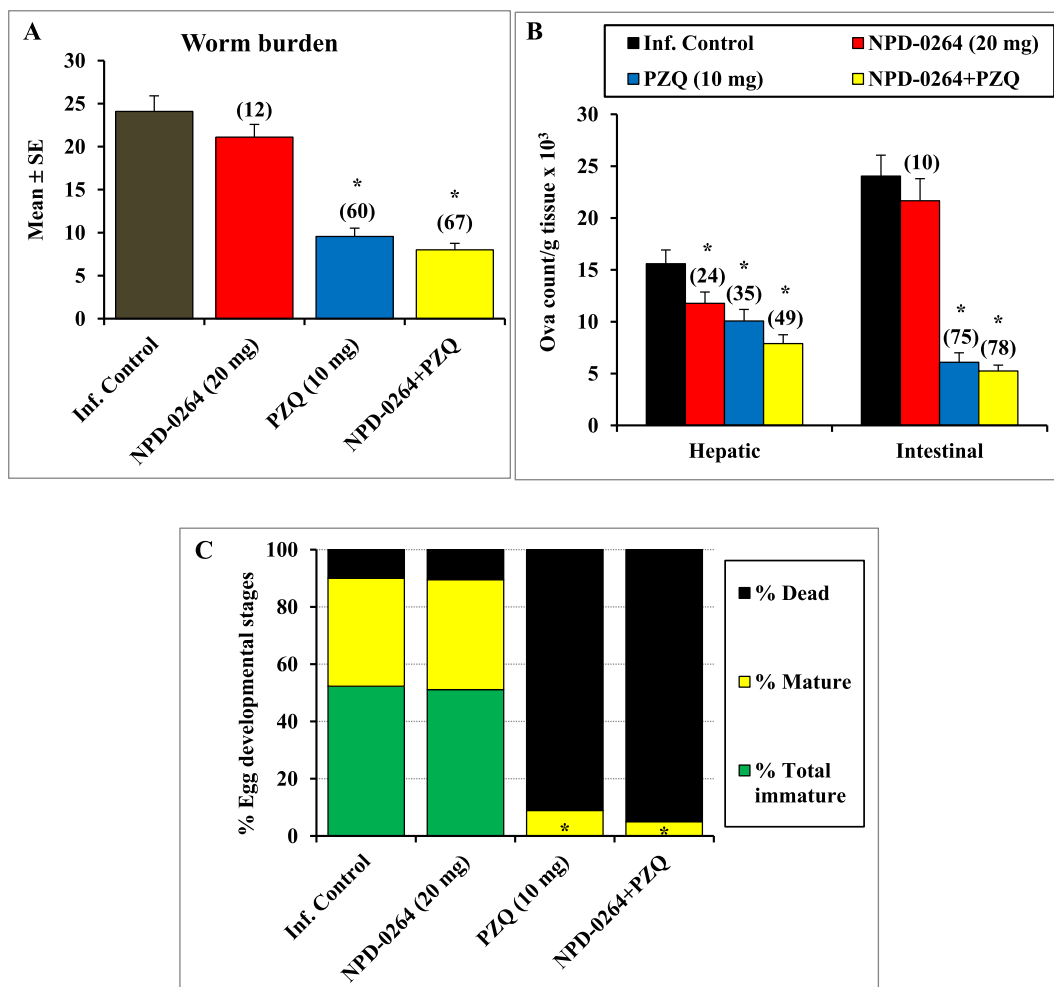
One of the major shortcomings of PZQ is its limited efficacy against immature worms, which prompted us to look at the effect of the Class-I compounds NPD-1012 and NPD-0356 against 3 week-old immature and 4 week-old early mature worms in comparison to the effects on mature 6 week-old parasites (Table 1). More prominent effects on early mature and to a lesser extent on immature worms were noted at concentrations of 50  $\mu$ M and 25  $\mu$ M compared to parallel findings in mature worms (Fig. 1).

### 3.2. *In vivo* activity in the *S. mansoni* mouse model

Guided by the *in vitro* findings, three *in vitro* active 2, 4-arylimidazole compounds, NPD-0274, NPD-0264 and NPD-0298 (Fig. 2), were tested in *S. mansoni*-infected mice. Because these imidazole derivatives had been reported to be metabolically unstable (Sebastián-Pérez et al., 2018), concomitant dosing with the CYP450 inhibitor aminobenzotriazole (ABT) (Watanabe et al., 2016) was carried out with each experimental treatment (NPD, PZQ or combination of both). In a separate experiment, ABT itself had no significant effect on worm burden compared to untreated control mice ( $31.3 \pm 1.3$  ( $n = 3$ ) vs  $29.0 \pm 1.2$  ( $n = 6$ ),  $p = 0.28$ ).

Treatment of *S. mansoni*-infected mice with the Class-I compound

## NPD-0264



**Fig. 4.** (A) Worm burden, (B) tissue egg load and (C) oogram pattern of NPD-0264 sacrificed 10 days post end of treatment when used alone at 20 mg/kg/day or in combination with PZQ at 10 mg/kg/day for 5 days. \*significantly different from infected control at  $P < 0.05$ . Numbers above columns and between parentheses represent percentage change from infected control group. ABT (100 mg/kg orally) was administered 2 h prior to administration of NPD-0264 and PZQ, whether alone or in combination. Error bars represent SEM.

NPD-0274 at 20 mg/kg/day for 5 days significantly decreased the worm load and the intestinal tissue egg burden by 29% ( $P < 0.05$ ) and 35% ( $P < 0.05$ ), respectively, as compared to the untreated controls (Fig. 3A and B). Combination treatment with NPD-0274 and PZQ in reduced doses of 10 mg/kg/day each for 5 days revealed significantly higher worm killing (96% vs 60%;  $P < 0.05$ ), and higher intestinal tissue egg burden reduction (83% vs 67%;  $P < 0.05$ ) compared to PZQ-treated alone (Fig. 3A and B). Interestingly, the maturation of eggs did not change upon treatment with NPD-0274 alone (almost half the eggs were immature or mature), but after the combination treatment only very few mature eggs remained and the rest were dead (Fig. 3C).

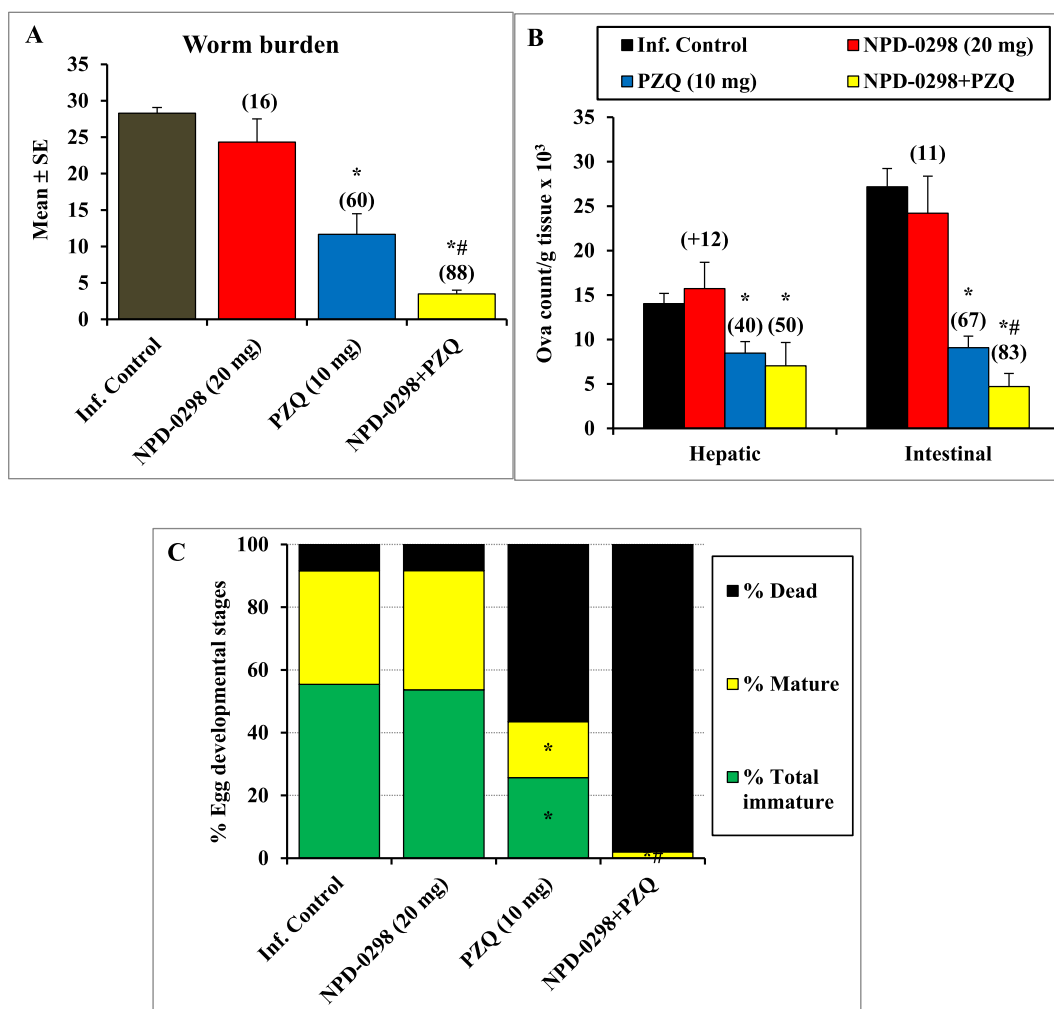
With monotherapy of Class-I compound NPD-0247 yielding only moderate relief of worm and egg burdens, it was decided also to evaluate the Class-II compounds NPD-0264 and NPD-0298, particularly since *in vitro* NPD-0298 had impacted greatly on oviposition and coupling (Table 1). Infected mice were given the same treatment dose and schedule as the Class-I compounds. NPD-0264 did not reveal significant antischistosomal activity, either alone or when combined with PZQ (Fig. 4). NPD-0298 alone also did not reveal significant antischistosomal activity. However, when co-administered with PZQ, worm killing was significantly enhanced compared to PZQ alone (88% vs 60%;  $P < 0.05$ ), intestinal tissue egg load was significantly reduced (83% vs 67%) and the percentage of dead eggs was markedly increased

with complete absence of immature eggs (Fig. 5).

#### 4. Discussion

From a PDE-focused library, 265 compounds were phenotypically screened for antischistosomal efficacy and 171 (64%) displayed various degrees of antischistosomal action revealed by worm mortality, motor activity alterations (sluggish worm movement or spastic contractions), reduced ovipositing and unpairing. The 171 'hits' were sorted into five classes based on their primary *in vitro* profile at 100  $\mu$ M and 50  $\mu$ M. At 100  $\mu$ M, 12 (7%) displayed 50–100% killing (Class-I), and 29 (17%) displayed  $< 50\%$  worm killing but with worm unpairing, sluggish movement of survivors and absence of eggs (Class-II). The reason why the percentage of worm killing barely exceeded 50% was due to the fact that almost all compounds mainly affected males with little or no impact on the females, whereas PZQ was not sex-specific over the dose range tested. As such, the  $EC_{50}$  values for males were substantially lower than those for the mixed populations (Tables 1 and 2) with 32  $\mu$ M as the lowest  $EC_{50}$  value, for NPD-0274. Disparities in *in vitro* drug susceptibility between male and female *S. mansoni* worms have indeed been previously reported. Higher susceptibility of male over female schistosomes has previously been reported for PZQ, oxamniquine, ginger extract, and some essential oils (Mikhail et al., 1978; Pica-

## NPD-0298



**Fig. 5.** (A) Worm burden, (B) tissue egg load and (C) oogram pattern of NPD-0298 sacrificed 10 days post end of treatment when used alone at 20 mg/kg/day or in combination with PZQ at 10 mg/kg/day for 5 days. \*significantly different from infected control at  $P < 0.05$ , #significantly different from PZQ group at  $P < 0.05$ . Numbers above columns and between parentheses represent percentage change from infected control group. ABT (100 mg/kg orally) was administered 2 h prior to administration of NPD-298 and PZQ, whether alone or in combination. Error bars represent SEM.

Mattoccia and Cioli., 2004; Mostafa et al., 2011; Tonuci et al., 2011). Absolute killing of male worms only was also reported by Fernandes et al. (2013) when the antischistosomal activity of different synthetic preparations of N-alkylated diamines, amino alcohols, and glycosylated amino alcohols were examined *in vitro*. Preferential killing of female schistosomes has also been reported for some compounds, including 2-(butylamino)-1-phenyl-1-ethanethiosulfuric acid, amino alkanethiosulfuric acids and artesunate (de Araújo et al., 2007; de Oliveira Penido et al., 2008; Mitsui et al., 2009).

Many of the 171 test compounds, including those in Class-I and Class-II, also induced motor activity alterations, which were usually manifested as sluggish movement of the male worms. Spastic worm contractions were observed with 16 compounds including only 1 from Class-I (NPD-1014, a pyridazinone), but these contractions did not result in death during the *in vitro* observations. This phenotype has recently been linked to inhibition of SmPDE4A (Long et al., 2017) and the fact that few compounds in our PDE-focused library induced spastic contractions may indicate that this particular PDE was not the main target of most of the compounds. Indeed, the imidazole series that includes the hits NPD-0274, NPD-0264 and NPD-0298 is associated with inhibition of the PDE10 family and has potential utility in Parkinson's disease (García et al., 2017). Motor activity alterations are considered

important indicators of schistosomicidal activity, disturbing not only the whole worm's muscle function and movement but also the muscles of the suckers essential to attach to the host vessels and the tight pairing of male and female worms (Ribeiro and Patocka, 2013; Patocka et al., 2014).

Possibly the most important observation was the high impact on ovipositing at very low, sub-lethal concentrations. Complete absence of eggs is obviously expected in the presence of 100% worm killing or separated worms (Magalhães et al., 2012). Reduction or absence of eggs can also be expected when severe motor activity alterations are recorded as these disturb the muscle lining of the reproductive excretory organs (Ribeiro and Patocka, 2013; Patocka et al., 2014). Complete absence of eggs was recorded not only for Class-I and -II compounds, but also after incubation with compounds displaying no schistosomicidal activity, both in the presence of intact couples and after unpairing (Class-III [14% of compounds] and Class-IV [28% of compounds]). However, in some cases, cessation of ovipositing was observed without any signs of motor disturbances in living, intact couples, which may point to specific insult of the reproductive organs consistent with the strong sex-specificity of the compounds.

Class-I compounds belong to several chemical families (Table S1). Of particular interest were those bearing a phthalazinone or an

imidazole scaffold. The phthalazinones are related to the well-known *T. brucei* PDEB1 and B2 inhibitors NPD-001 and NPD-008 (De Koning et al., 2012; Veerman et al., 2016; Blaazer et al., 2017), while the imidazoles are chemically related to previously described human PDE10A inhibitors (García et al., 2017). As the imidazole scaffold yielded several of the most active compounds and, as mammalian PDE inhibitors, may be among the most likely inhibitors of helminth PDEs, three imidazole derivatives were selected for evaluation in *S. mansoni*-infected mice: one from Class-I (NPD-0274) and two from Class-II (NPD-0264 and NPD-0298) (Fig. 2). The selection took into account factors such as cytotoxicity and observed adverse effects in mice at fixed dose regimens. Particular significance was given to the % reduction of egg production, since the eggs cause the clinical pathology of schistosomiasis, as well as disease transmission. If ovipositing could be irreversibly affected, the killing of the worm becomes of secondary importance.

NPD-0274 showed a modest but significant reduction in worm burden *in vivo*. In line with the *in vitro* effect on ovipositing rather than worm viability, the other two imidazoles (NPD-0264, NPD-0298) showed no significant worm killing *in vivo*. When administered at reduced doses with PZQ, NPD-0274 and NPD-0298 significantly enhanced worm killing (96% and 88% respectively, compared to 60% killing for PZQ alone). The same combinations also showed a higher reduction in intestinal egg load (both 83%, vs 67% for PZQ); of the eggs that were still present, the vast majority was dead and none were immature (*i.e.* viable and recently deposited). An increase of > 50% of mature eggs with absence of one or more of the immature stages is definite proof that a drug possesses antischistosomal activity (Pellegrino et al., 1962). Combination therapy of PZQ and oxamniquine, omeprazole or mefloquine in experimental schistosomiasis demonstrated increased worm lethality (Botros et al., 1989; El-Lakkany et al., 2011; Keiser et al., 2011; Almeida et al., 2015). Information as to whether drug combinations provide increased therapeutic efficacy over monotherapy is scarce, although in the studies conducted to date no curative advantages over single treatment were reported (Kamel et al., 2000; Utzinger et al., 2003; Cui and Su, 2009).

In conclusion, this study enabled successful selection of several antischistosomal compounds, based on *in vitro* findings such as worm killing, unpairing, alterations of motor activity (sluggish worm movement, spastic contractions) and ovipositing. Guided by these criteria, three imidazole derivatives were administered alone and in combination with PZQ to *S. mansoni* infected mice, providing enhanced therapeutic efficacy in combination with PZQ. When administered alone *in vitro*, there was a clear reduction to complete absence of eggs with no recovery over 7 days in the absence of drug, suggesting long-term disabling of ovipositing. The near-complete eradication of viable eggs with the PZQ/NPD-0274 and PZQ/NPD-0298 combinations is particularly promising as it would greatly diminish or eliminate pathology and above all transmission. It is the rapid reinfection caused by the failure to break the transmission cycle that remains the most important factor in the persistently high infection rates in endemic areas.

These imidazoles are structurally related to a family of human PDE10A inhibitors and have also shown phenotypic activities in trypanosomatids (Sebastián-Pérez et al., 2018; de Araújo et al., 2019). Further developments will focus on additional SAR, on validating the SmpDEs (particularly of the PDE10 family) as the targets for these compounds, on transcriptomics to explain the observed sex differences in drug susceptibility, and on pharmacokinetic studies to rationally improve dosage regimens. The fact that these imidazoles are at least equipotent against immature and early mature worms gives hope that the most severe limitation of PZQ can be overcome.

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## Appendix A. Supplementary data

Classification of compounds from Class I–V in chemical classes (Tables S1–S5).

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ijpdr.2019.01.001>.

## References

- Almeida, G.T., Lage, R.C., Anderson, L., Venancio, T.M., Nakaya, H.I., Miyasato, P.A., Rofatto, H.K., Zerlotini, A., Nakano, E., Oliveira, G., Verjovski-Almeida, S., 2015. Synergy of omeprazole and praziquantel *in vitro* treatment against *Schistosoma mansoni* adult worms. *PLoS Neglected Trop. Dis.* 9, e0004086.
- Blaazer, A.R., Singh, A.K., Edink, E., Orrling, K.M., Veerman, J., Van den Bergh, T., Jansen, C., Balasubramaniam, E., Mooij, W.J., De Heuvel, E., Tagoe, D.N.A., Munday, J.C., Tenor, H., Matheussen, A., Wijtmans, M., Siderius, M., De Graaf, C., Maes, L., De Koning, H.P., Bailey, D., Sterk, G.J., De Esch, L.J.P., Brown, D.G., Leurs, R., 2017. Targeting a subpocket in *Trypanosoma brucei* B1 assists structure-based drug discovery for African sleeping sickness. *J. Med. Chem.* 6, 3870–3888.
- Botros, S., Pica-Mattoccia, L., William, S., El-Lakkany, N., Cioli, D., 2005. Effect of praziquantel on the immature stages of *Schistosoma haematobium*. *Int. J. Parasitol.* 35, 1453–1457.
- Botros, S., Soliman, A., el-Gawhary, N., Selim, M., Guirguis, N., 1989. Effect of combined low dose praziquantel and oxamniquine on different stages of Schistosome maturity. *Trans. R. Soc. Trop. Med. Hyg.* 83, 86–89.
- Cheever, A.W., 1968. Conditions affecting the accuracy of potassium hydroxide digestion techniques for counting *Schistosoma mansoni* eggs in tissues. *Bull. World Health Organ.* 39, 328–331.
- Cioli, D., Botros, S.S., Wheatcroft-Francklow, K., Mbaye, A., Southgate, V., Tchuente, L.A., Pica-Mattoccia, L., Troiani, A.R., El-Din, S.H., Sabra, A.N., Albin, J., Engels, D., Doenhoff, M.J., 2004. Determination of ED<sub>50</sub> values for praziquantel in praziquantel-resistant and -susceptible *Schistosoma mansoni* isolates. *Int. J. Parasitol.* 34, 979–987.
- Cioli, D., Pica-Mattoccia, L., Basso, A., Guidi, A., 2014. Schistosomiasis control: praziquantel forever? *Mol. Biochem. Parasitol.* 195, 23–29.
- Couto, F.F., Coelho, P.M., Araujo, N., Kusel, J.R., Katz, N., Jannotti-Passos, L.K., Mattos, A.C., 2011. *Schistosoma mansoni*: a method for inducing resistance to praziquantel using infected *Biomphalaria glabrata* snails. *Mem. Inst. Oswaldo Cruz* 106, 153–157.
- Cui, L., Su, X.Z., 2009. Discovery, mechanisms of action and combination therapy of artemisinin. *Expert Rev. Anti Infect. Ther.* 7, 999–1013.
- de Araújo, J.S., Garcia-Rubia, A., Sebastián-Pérez, V., Kalejaiye, T.D., Bernardino da Silva, P., Fonseca-Berzal, C.R., Maes, L., De Koning, H.P., Soeiro, M.N.C., Gil, C., 2019. Imidazole derivatives as promising agents for the treatment of Chagas disease. *Antimicrob. Agents Chemother.* (in press).
- de Araújo, S.C., de Mattos, A.C.A., Teixeira, H.F., Coelho, P.M.Z., Nelson, D.L., de Oliveira, M.C., 2007. Improvement of *in vitro* efficacy of a novel schistosomicidal drug by incorporation into nanoemulsions. *Int. J. Pharm.* 337, 307–315.
- De Koning, H.P., Gould, M.K., Sterk, G.J., Tenor, H., Kunz, S., Lugimbuehl, E., Seebeck, T., 2012. Pharmacological validation of *Trypanosoma brucei* phosphodiesterases as novel drug targets. *J. Infect. Dis.* 206, 229–237.
- de Oliveira Penido, M.L., Zech Coelho, P.M., de Mello, R.T., Piló-Veloso, D., de Oliveira, M.C., Kusel, J.R., Nelson, D.L., 2008. Antischistosomal activity of aminoalkanethiols, aminoalkanethiosulfuric acids and the corresponding disulfides. *Acta Trop.* 108, 249–255.
- Doenhoff, M.J., Cioli, D., Utzinger, J., 2008. Praziquantel: mechanisms of action, resistance and new derivatives for schistosomiasis. *Curr. Opin. Infect. Dis.* 21, 659–667.
- Duvall, R.H., De Witt, W.B., 1967. Technique for recovering adult schistosomes from laboratory animals. *Am. J. Trop. Med. Hyg.* 16, 438–486.
- El-Lakkany, N.M., el-Din, S.H., Sabra, A.N., Hammam, O.A., 2011. Pharmacodynamics of mefloquine and praziquantel combination therapy in mice harbouring juvenile and adult *Schistosoma mansoni*. *Mem. Inst. Oswaldo Cruz* 106, 814–822.
- Fallon, P.G., Doenhoff, M.J., 1994. Drug-resistant schistosomiasis: resistance to praziquantel and oxamniquine induced in *Schistosoma mansoni* in mice is drug specific. *Am. J. Trop. Med. Hyg.* 51, 83–88.
- Fernandes, F.S., Rezende Júnior, C.O., Fernandes, T.S., da Silveira, L.S., Rezende, C.A.M., de Almeida, M.V., de Paula, R.G., Rodrigues, V., Da Silva Filho, A.A., Couri, M.R., 2013. Anthelmintic effects of alkylated diamines and amino alcohols against *Schistosoma mansoni*. *BioMed Res. Int.* 2013, 783490.
- García, A.M., Salado, I.G., Perez, D.I., Brea, J., Morales-García, J.A., González-García, A., Cadavid, M.I., Loza, M.I., Luque, F.J., Perez-Castillo, A., Martínez, A., Gil, C., 2017. Pharmacological tools based on imidazole scaffold proved the utility of PDE10A inhibitors for Parkinson's disease. *Future Med. Chem.* 9, 731–748.
- Ghosh, R., Sawant, O., Ganpathy, P., Pitre, S., Kadam, V.J., 2009. Phosphodiesterase inhibitors: their role and implications. *Int. J. Pharm. Tech. Res.* 1, 1148–1160.
- Gryseels, B., Mbaye, A., De Vlas, S.J., Stelma, F.F., Guisse, F., Van Lieshout, L., Faye, D., Diop, M., Ly, A., Tchuente, L.A., Engels, D., Polman, K., 2001. Are poor



- responses to praziquantel for the treatment of *Schistosoma mansoni* infections in Senegal due to resistance? An overview of the evidence. *Trop. Med. Int. Health* 6, 864–873.
- Ismail, M.M., Taha, S.A., Farghaly, A.M., el-Azomy, A.S., 1994. Laboratory induced resistance to praziquantel in experimental schistosomiasis. *J. Egypt. Soc. Parasitol.* 24, 685–695.
- Kamel, G., Metwally, A., Guirguis, F., Nessim, N.G., Noseir, M., 2000. Effect of a combination of the new antischistosomal drug Ro 15-5458 and praziquantel on different strains of *Schistosoma mansoni* infected mice. *Arzneimittelforschung* 50, 391–394.
- Keiser, J., Manneck, T., Vargas, M., 2011. Interactions of mefloquine with praziquantel in the *Schistosoma mansoni* mouse model and *in vitro*. *J. Antimicrob. Chemother.* 66, 1791–1797.
- Liang, Y.S., John, B.I., Boyd, D.A., 1987. Laboratory cultivation of schistosome vector snails and maintenance of schistosome life cycles. *Proc. 1st Sino-Am. Symp.* 1, 34–48.
- Liang, Y.S., Li, H.J., Dai, J.R., Wang, W., Qu, G.L., Tao, Y.H., Xing, Y.X., Li, Y.Z., Qian, K., Wei, J.Y., 2011. Studies on resistance of *Schistosoma* to praziquantel XIII resistance of *Schistosoma japonicum* to praziquantel is experimentally induced in laboratory. *Zhongguo Xue Xi Chong Bing Fang Zhi Za Zhi* 23, 605–610.
- Long, T., Rojo-Arreola, L., Shi, D., El-Sakkary, N., Jarnagin, K., Rock, F., Meewan, M., Rascón Jr., A.A., Lin, L., Cunningham, K.A., Lemieux, G.A., Podust, L., Abagyan, R., Ashrafi, K., McKerrow, J.H., Caffrey, C.R., 2017. Phenotypic, chemical and functional characterization of cyclic nucleotide phosphodiesterase 4 (PDE4) as a potential anthelmintic drug target. *PLoS Neglected Trop. Dis.* 11, e0005680.
- Magalhães, L.G., de Souza, J.M., Wakabayashi, K.A., da S. Laurentiz, R., Vinhólis, A.H., Rezende, K.C., Esperandim, V.R., Ferreira, D.S., Crotti, A.E., Cunha, W.R., e Silva, M.L., 2012. *In vitro* efficacy of the essential oil of *Piper cubeba* L. (Piperaceae) against *Schistosoma mansoni*. *Parasitol. Res.* 110, 1747–1754.
- Melman, S.D., Steinauer, M.L., Cunningham, C., Kubatko, L.S., Mwangi, I.N., Wynn, N.B., Mutuku, M.W., Karanja, D.M.S., Colley, D.G., Black, C.L., Secor, W.E., Mkoji, G.M., Loker, E.S., 2009. Reduced susceptibility to praziquantel among naturally occurring Kenyan isolates of *Schistosoma mansoni*. *PLoS Neglected Trop. Dis.* 3, e504.
- Mikhail, E.G., Tadros, M.B., Mahran, S.G., Gaber, A.A., Guiguis, Z.I., 1978. Therapeutic effect of oxamniquine in experimental schistosomiasis. *J. Egypt. Soc. Parasitol.* 8, 109–120.
- Mitsui, Y., Miura, M., Aoki, Y., 2009. *In vitro* effects of artesunate on the survival of worm pairs and egg production of *Schistosoma mansoni*. *J. Helminthol.* 83, 7–11.
- Mostafa, O.M., Eid, R.A., Adly, M.A., 2011. Antischistosomal activity of ginger (*Zingiber officinale*) against *Schistosoma mansoni* harbored in C57 mice. *Parasitol. Res.* 109, 395–403.
- Mwangi, I.N., Sanchez, M.C., Mkoji, G.M., Agola, L.E., Runo, S.M., Cupit, P.M., Cunningham, C., 2014. Praziquantel sensitivity of Kenyan *Schistosoma mansoni* isolates and the generation of a laboratory strain with reduced susceptibility to the drug. *Int. J. Parasitol. Drugs Drug Resist.* 4, 296–300.
- Patocka, N., Sharma, N., Rashid, M., Ribeiro, P., 2014. Serotonin signaling in *Schistosoma mansoni*: a serotonin-activated G protein-coupled receptor controls parasite movement. *PLoS Pathog.* 10, e1003878.
- Pellegrino, J., Oliveira, C.A., Faria, J., Cunha, A.S., 1962. New approach to the screening of drugs in experimental schistosomiasis *mansoni* in mice. *Am. J. Trop. Med. Hyg.* 11, 201–215.
- Pica-Mattoccia, L., Cioli, D., 2004. Sex- and stage-related sensitivity of *Schistosoma mansoni* to *in vivo* and *in vitro* praziquantel treatment. *Int. J. Parasitol.* 34, 527–533.
- Ribeiro, P., Patocka, N., 2013. Neurotransmitter transporters in schistosomes: structure, function and prospects for drug discovery. *Parasitol. Int.* 62, 629–638.
- Sabra, A.N., Botros, S.S., 2008. Response of *Schistosoma mansoni* isolates having different drug sensitivity to praziquantel over several life cycle passages with and without therapeutic pressure. *J. Parasitol.* 94, 537–541.
- Sebastián-Pérez, V., Hendrickx, S., Munday, J.C., Kalejaiye, T., Martínez, A., Campillo, N.E., de Koning, H., Caljon, G., Maes, L., Gil, C., 2018. Cyclic nucleotide specific phosphodiesterases as potential drug targets for anti-Leishmania therapy. *Antimicrob. Agents Chemother.* 62, e00603–e00618.
- Shakur, Y., de Koning, H.P., Ke, H., Kambayashi, J., Seebeck, T., 2011. Therapeutic potential of phosphodiesterase inhibitors in parasitic diseases. *Handb. Exp. Pharmacol.* 204, 487–510.
- Tonuci, L.R.S., de Melo, N.I., Dias, H.J., Wakabayashi, K.A.L., Aguiar, G.P., Aguiar, D.P., Mantovani, A.L.L., Ramos, R.C., Groppo, M., Rodrigues, V., Veneziani, R.C.S., Cunha, W.R., da Silva Filho, A.A., Magalhães, L.G., Crotti, A.E.M., 2011. *In vitro* schistocidal effects of the essential oil of *Tagetes erecta*. *Rev. Bras. Farmacogn.* 22, 88–93.
- Utzinger, J., Keiser, J., Xiao, S.H., Tanner, M.H., Singer, H.B., 2003. Combination chemotherapy of schistosomiasis in laboratory studies and clinical trials. *Antimicrob. Agents Chemother.* 47, 1487–1495.
- Veerman, J., van den Bergh, T., Orrling, K.M., Jansen, C., Cos, P., Maes, L., Chatelain, E., Ioset, J.R., Edink, E.E., Tenor, H., Seebeck, T., de Esch, I., Leurs, R., Sterk, G.J., 2016. Synthesis and evaluation of analogs of the phenylpyridazinone NPD-001 as potent trypanosomal TbrPDEB1 phosphodiesterase inhibitors and *in vitro* trypanocidals. *Bioorg. Med. Chem.* 24, 1573–1581.
- Watanabe, A., Mayumi, K., Nishimura, K., Osaki, H., 2016. *In vivo* use of the CYP inhibitor 1-aminobenzotriazole to increase long-term exposure in mice. *Biopharm Drug Dispos.* 37, 373–378.
- WHO, 2012. Schistosomiasis Progress Report 2001–2011 and Strategic Plan 2012–2020. World Health Organization, Geneva.
- WHO, 2017. *Global Health Estimates*. <http://www.who.int/healthinfo>.
- William, S., Sabra, A., Ramzy, F., Mousa, M., Demerdash, Z., Bennett, J.L., Day, T.A., Botros, S., 2001. Stability and reproductive fitness of *Schistosoma mansoni* isolates with decreased sensitivity to praziquantel. *Int. J. Parasitol.* 31, 1093–1100.