

McSorley, S. T., Tham, A., Jones, I., Talwar, D. and McMillan, D. C. (2019) Regression correction equation to adjust serum iron and ferritin concentrations based on C-reactive protein and albumin in patients receiving primary and secondary care. *Journal of Nutrition*, 149(5), pp. 877-883. (doi: 10.1093/jn/nxz008)

The material cannot be used for any other purpose without further permission of the publisher and is for private use only.

There may be differences between this version and the published version. You are advised to consult the publisher's version if you wish to cite from it.

http://eprints.gla.ac.uk/178066/

Deposited on 17 January 2019

Enlighten – Research publications by members of the University of Glasgow
http://eprints.gla.ac.uk

Original article:

Regression correction equation to adjust serum iron and ferritin concentrations based on C-reactive protein and albumin in patients receiving primary and secondary care.

Stephen T McSorley¹, Alexander Tham¹, Iain Jones², Dinesh Talwar², Donald C McMillan¹

- 1. Academic Unit of Surgery, School of Medicine, University of Glasgow, Glasgow, UK
- 2. The Scottish Trace Elements and Micronutrients Reference Laboratory, Department of Clinical Biochemistry, Glasgow Royal Infirmary, Glasgow G4 0SF.

Corresponding author:

Mr Stephen McSorley, Clinical Research Fellow

Level 2, New Lister Building, Glasgow Royal Infirmary, Glasgow, UK

Postcode: G31 2ER Tel: 0141 211 8675 Email: s.mcsorley@doctors.org.uk

Conflict of interest/Disclosure: none Financial support: none

Running title: Adjusting ferritin by CRP and albumin

Abbreviations: AGP α-1-acid glycoprotein, Alb_{obs} observed albumin, Alb_{ref} Albumin reference value, ASA American Society of Anesthesiology, BRINDA Biomarkers Reflecting Inflammation and Nutritional Determinants of Anemia, CRP C-reactive protein, CRP_{obs} observed CRP, CRP_{ref} CRP reference value, EQA External Quality Assessment, Fer_{adj} adjusted ferritin, Fer_{obs} observed ferritin, Hb haemoglobin, IL interleukin, IQC Internal Quality Control, Ir_{adj} adjusted serum iron, Ir_{obs} observed serum iron, IQR interquartile range, MCV mean corpuscular volume, mGPS modified Glasgow Prognostic Score, nCRT neoadjuvant chemoradiotherapy, NEQAS National External Quality Assessment Services, NHS National Health Service, sTFR soluble transferrin receptor, TNF Tumor Necrosis Factor, TNM Tumor nodes metastasis stage, UICC Union for International Cancer Control, WHO World Health Organization,

Abstract

- 2 Background:
- 3 Systemic inflammation, even at low levels, is associated with significant derangement of
- 4 measures of iron status, making diagnosis of iron deficiency difficult. The objective of the
- 5 present study was to create linear regression correction equations to adjust serum ferritin and
- 6 iron using the acute phase proteins C-reactive protein (CRP) and albumin.

7

1

- 8 Methods:
- 9 Data from a cohort (1) of patients (n=7,226) in primary and secondary care who had serum
- ferritin, iron, CRP and albumin measured at the same time point were examined. Linear
- regression coefficients calculated for CRP and albumin with serum iron and ferritin as the
- 12 outcome variables. Patients were categorised as iron deficient (ferritin <15μg/L, or serum
- iron <10 µmol/L). The equation was then applied to a cohort (2) of patients with colorectal
- cancer who had ferritin and iron measured preoperatively (n=356).

- 16 Results:
- In cohort 1 there was a significant difference in the proportions of patients with serum ferritin
- $<15\mu g/L$ and serum iron $<10\mu mol/L$ respectively when the unadjusted (7% and 55%),
- adjusted using CRP alone (13% and 26%), adjusted using albumin alone (11% and 37%), and
- using both CRP and albumin (24% and 15%) values were compared (both p<0.001). In
- 21 cohort 2 there was a significant difference in the proportions of patients with serum ferritin
- $<15\mu g/L$ and serum iron $<10\mu mol/L$ respectively when the unadjusted (28% and 66%),

23	adjusted using CRP alone (39% and 57%), adjusted using albumin alone (39% and 59%), and
24	using both CRP and albumin (46% and 44%) values were compared (p<0.001 and p<0.004).
25	
26	
27	Conclusions:
28	In both cohorts the greatest increase in the proportion of patients meeting definitions for iron
29	deficiency was found when adjustment was made for both CRP and albumin together. Even
30	low levels of inflammation had a significant effect on serum iron and ferritin values.
31	
32	
33	Keywords
34	Systemic inflammation, ferritin, iron deficiency, C-reactive protein, albumin, colorectal
35	cancer

Introduction

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

36

Iron deficiency remains a significant burden of disease and in both developing and developed countries [1]. Total serum iron can be used to assess iron status, however, in clinical practice, ferritin is a more robust measure of iron status. Currently the World Health Organization (WHO) define adults with ferritin <15 µg/L as iron deficient [2]. However, ferritin is a positive acute phase reactant and therefore serum concentrations rise in the presence of systemic inflammation, leading to the possibility that patients may be truly iron deficient despite a serum ferritin $> 15 \mu g/L$. Compounding matters, inflammation is associated with lower serum total iron and transferrin saturation, due to the action of hepcidin, itself driven by circulating IL 6 [3]. This often leads to a difficult to interpret picture of serum measures of iron status which may mask functional iron deficiency [4], thereby making it difficult to assess the need for iron replacement, and to monitor the effectiveness of iron supplementation. Therefore, attempts have been made to correct measures of iron status for the presence systemic inflammation, using exclusion criteria [5], correction factors [6], and novel markers of iron status [7]. Recently, the Biomarkers Reflecting Inflammation and Nutritional Determinants of Anemia (BRINDA) groups have used the acute phase markers C-reactive protein (CRP) and α-1-acid glycoprotein (AGP) to create linear correction regression equations to adjust serum ferritin by inflammation [8]. They argue that the significant effects of even low level inflammation make this the best method of adjustment [9]. However, as AGP is not widely clinical available at present, and compared to albumin is more expensive [10].

The effect on serum iron and ferritin of the combination of CRP and the widely clinically available negative acute phase protein albumin has also been quantified, having been shown to stratify serum iron and ferritin [11]. CRP is an opsonin and member of the pentraxin family synthesised by hepatocytes in response to stimulation by the proinflammatory cytokine IL 6, whilst the plasma transport protein albumin is redistributed due to changes in vascular permeability, and its synthesis is supressed by TNF and IL 1 [12]. To date, no regression correction equation to adjust serum iron or ferritin by these markers of systemic inflammation has been created.

Therefore, the aim of the present study was to derive linear correction regression equations to adjust serum iron and ferritin by CRP and albumin, and then to apply this to a cohort of patients with colorectal cancer.

Patients and Methods

72

73

71

Patients:

74 The present study included two cohorts of patients. The first (n=7,226), in which the regression correction equation was formulated, has been described previously [11]. 75 Briefly, details of all requests for serum iron or ferritin were obtained from the laboratory 76 information systems of 4 Glasgow hospitals respectively for the period 1st August 2006 to 77 31st July 2011. These requests were made from secondary inpatient, secondary outpatient 78 and primary care healthcare providers. Iron and ferritin results were matched to CRP and 79 albumin results obtained on the same calendar day using electronic laboratory patient 80 identifiers. Any requests which were not accompanied by a CRP and albumin request were 81 not included in the study. Where an individual had repeat measurements, only the first was 82 included in the study to prevent the need for repeated measures analysis in the subgroup of 83 included patients. 84 The second (n=356) were patients with colorectal cancer undergoing potentially curative 85 treatment at a single surgical centre in Glasgow, between 1st January 2008 and 30th June 2017 86 who had a serum iron or ferritin measured in the preoperative period. Patients who had not 87 had either a serum iron or ferritin request were not included in the study. 88 Ferritin was analysed using 2 step chemiluminescent microparticle immunoassay within the 89 90 routine haematology laboratories. Serum total iron (chemically using ferene), serum CRP (by the immunoturbidimetric method) and albumin (chemically using Bromocresol purple) were 91 92 measured using an automated analyser (Architect, Abbot Diagnosis, Maidenhead, UK) in the 93 routine biochemistry laboratories. All sites used the same analytic materials and automated

94 platforms. There were no sustained concerns regarding IQC performance requiring investigation into the performance of the assays. The A, B and C scores were within the EQA 95 (NEQAS) targets during the study period. 96 The audit was conducted with the intent of developing local guidelines and to aid in the 97 interpretation of serum measurements of iron status and was approved by the health board 98 (NHS Greater Glasgow and Clyde). 99 100 101 Methods: The formulation of the regression correction equation was based on a modification of that 102 used by the BRINDA group to correct ferritin for inflammation using CRP and AGP in 103 asymptomatic populations [8, 13]. The equation used linear regression to adjust ferritin 104 concentrations by the CRP and albumin concentrations on a continuous scale. 105 106 The observed ferritin (Fer_{obs}, normal range 41-400 µg/L), serum iron (Ir_{obs}, normal range 10-30 μmol/L), CRP (CRP_{obs.} normal range <10 mg/L) and albumin (Alb_{obs.} normal range 35-107 50g/L) values were ln transformed to approximate a normal distribution. Bivariate linear 108 regression coefficients were calculated with ferritin as the outcome variable and CRP (β1) 109 and albumin (β 2) as each of the explanatory variables. Multivariate linear regression was 110 then used to calculate the variance inflation factor (VIF) as a measure of collinearity, with a 111 value <5 allowing both variables to be included in the final equation. 112 Internal reference values for CRP (CRP_{ref}) and albumin (Alb_{ref}) were derived similarly to that 113 used in the BRINDA methodology to prevent overcorrection for very low levels of 114 inflammation. In CRP_{ref} was calculated as the lowest decile of ln CRP_{obs} in the cohort as 115

CRP is a positive acute phase reactant, and ln Albref was calculated as the highest decile of ln

Alb_{obs} in the cohort as albumin is a negative acute phase reactant. These reference values

116

- were subtracted from the observed CRP and albumin values within the equation, and the equation only applied to those patients with observed CRP above the internal reference cut
- off, and / or observed albumin below the internal reference cut off.
- 121 The final ferritin correction equation is as follows:
- ln Fer_{adj} = ln Fer_{obs} $(\beta 1(ln CRP_{obs} ln CRP_{ref})) (\beta 2(ln Alb_{obs} ln Alb_{ref}))$
- The final serum iron correction equation is as follows:
- The calculated adjusted ferritin (ln Fer_{adi}) and serum iron (ln Ir_{obs}) was then back transformed
- to obtain an adjusted estimate for ferritin (Fer_{adj}) and serum iron (Ir_{adj}) respectively.
- 128 Statistical analysis:

- In both cohorts. Continuous data were presented as medians and interquartile range (IQR).
- The median unadjusted serum iron and ferritin were compared to the median serum iron and
- ferritin using adjustment by CRP alone, albumin alone, and both CRP and albumin using the
- related samples Friedmann's two way analysis of variance by ranks. The proportion of
- patients defined as iron deficient (ferritin <15µg/L, or serum iron <10µmol/L) was calculated
- for unadjusted serum iron and ferritin and using adjustment for both CRP and albumin and
- compared using McNemar's test.
- In the second cohort patients were classified as having anemia using haemoglobin (Hb) based
- on WHO guidelines for males; <130 g/L and females; <120 g/L [13]. Furthermore, anemic
- patients were classified as having microcytic anemia with mean corpuscular volume (MCV)
- 480 f/L, or normocytic anemia with MCV 80-100 f/L.

- All analyses were performed using SPSS software (SPSS version 24, Chicago, Illinois,
- USA). Two sided P values <0.05 were considered statistically significant.

142 Results

143

144 Internal CRP and albumin reference values:

- The ln CRP_{ref} was calculated as lowest decile of ln CRP_{obs} which was 0.262 (back
- transformed corresponding $CRP_{obs} = 1.3 \text{mg/L}$). The ln Alb_{ref} was calculated as the highest
- decile of ln Alb_{obs} which was 3.71 (back transformed corresponding Alb_{obs} = 40.9 g/L).

148

- 149 Ferritin correction regression equation:
- Bivariate linear regression with ln Fer_{obs} as outcome variable and ln CRP_{obs} as the
- explanatory variable resulted in a regression coefficient (β1) of 0.331 (95% CI: 0.313, 0.348)
- and an intercept of 4.112 (95% CI: 4.057, 4.167). Bivariate linear regression with ln Fer_{obs} as
- outcome variable and ln Alb_{obs} as the explanatory variable resulted in a regression coefficient
- 154 (β2) of -2.087 (95% CI: -2.212, -1.962) and an intercept of 12.157 (95% CI: 11.728, 12.586).
- At multivariate analysis there was no evidence of significant collinearity between ln CRP_{obs}
- 156 and $\ln \text{Alb}_{\text{obs}}$ (VIF = 1.5).
- The final series of regression correction equation was therefore as follows:
- ln Fer_{adj} = ln Fer_{obs} (0.331(ln CRP_{obs} 0.262)) (-2.087(ln Alb_{obs} 3.71))
- 159 IF $\ln \text{CRP}_{\text{obs}} > 0.262 \text{ AND } \ln \text{Alb}_{\text{obs}} < 3.71$
- 160 $\operatorname{Fer}_{\operatorname{adj}} = \operatorname{EXP} (\operatorname{ln} \operatorname{Fer}_{\operatorname{adj}})$

161

- 163 Serum iron correction regression equation:
- Bivariate linear regression with ln Ir_{obs} as outcome variable and ln CRP_{obs} as the explanatory
- variable resulted in a regression coefficient (β1) of -0.261 (95% CI: -0.270, -0.251), and an
- intercept of 2.747 (95% CI: 2.716, 2.778). Bivariate linear regression with ln Ir_{obs} as outcome
- variable and ln Alb_{obs} as the explanatory variable resulted in a regression coefficient (β 2) of
- 1.176 (95% CI: 1.100, 1.252), and an intercept of -1.949 (95% CI: -2.211, -1.687). At
- multivariate analysis there was no evidence of significant collinearity between ln CRP_{obs} and
- 170 $\ln \text{Alb}_{\text{obs}} \text{ (VIF} = 1.5).$
- 171 The final series of regression correction equation was therefore as follows:
- ln Ir_{adj} = ln Ir_{obs} (-0.261(ln CRP_{obs} 0.262)) (1.176(ln Alb_{obs} 3.71))
- 173 IF ln CRP_{obs} >0.262 AND ln Alb_{obs} <3.71
- 174 $Ir_{adj} = EXP (ln Ir_{adi})$

- 176 Ferritin and serum iron adjustment in primary and secondary care patients:
- The primary and secondary care cohort included 7,226 patients of whom the majority 3,958
- 178 (55%) were female and 3,260 (45%) were male, with a median age of 68 years (range: 16-
- 179 103 y). Within the cohort the median CRP_{obs} was 15.0 mg/L (range: 0.15-476 mg/L), and
- 180 Alb_{obs} 32 g/L (range: 7-76 g/L).
- There was a significant decrease (Table 1) in the median serum ferritin (P < 0.001) and a
- significant increase in the proportions of patients with serum ferritin <15 μ g/L (P<0.001)
- when the unadjusted, adjusted using CRP alone, adjusted using albumin alone, and using both
- 184 CRP and albumin values were compared.

There was a significant increase (Table 1) in the median serum iron (P<0.001) and a significant decrease in the proportions of patients with serum iron <10 μ mol/L (P<0.001) when the unadjusted, adjusted using CRP alone, adjusted using albumin alone, and using both CRP and albumin values were compared.

Concordance between serum ferritin and serum iron definitions of iron deficiency following adjustment:

The proportion of patients in the primary and secondary care cohort found to not meet definitions of iron deficiency with both serum ferritin and serum iron (i.e. negative result agreement / true negative) increased from 43% using the unadjusted values to 66% when adjustment was made by both CRP and albumin (Table 2). The proportion of patients found to meet definitions of iron deficiency with both serum ferritin and serum iron (i.e. positive result agreement / true positive) remained unchanged at 5% following the same adjustment.

Ferritin and serum iron adjustment in patients with colorectal cancer:

There were 356 patients with UICC stage I-IV colorectal cancer who underwent surgery with curative intent at a single centre and had a preoperative serum ferritin or measured, of which 67 had serum iron measured, 307 had CRP measured, and 236 had both CRP and albumin measured (Table 2). Within the cohort there were 161 females (45%) and 195 males (55%), with a median age of 71 years (range: 27-90 y), and the majority having colonic (*n*=259, 73%) and node negative (*n*=209, 59%) disease.

There was a significant decrease (Table 3) in the median serum ferritin (P<0.001) and a significant increase in the proportions of patients with serum ferritin <15 μ g/L (P<0.001)

when the unadjusted, adjusted using CRP alone, adjusted using albumin alone, and using both CRP and albumin values were compared.

There was a significant increase (Table 3) in the median serum iron (P<0.001) and a significant decrease in the proportions of patients with serum iron <10 μ mol/L (P<0.001) when the unadjusted, adjusted using CRP alone, adjusted using albumin alone, and using both CRP and albumin values were compared.

Association between adjusted iron deficiency and anaemia in patients with colorectal cancer: Of the cohort of patients with colorectal cancer, 354 had serum ferritin, Hb and MCV measured allowing them to be grouped as having no anemia (n=174, 50%), microcytic anemia (n=41, 11%) or normocytic anemia (n=139, 39%) (Table 4). There was a significant association between the anemia groups and age (P<0.001), American Society of Anesthesiology (ASA) grade (P<0.001), tumor site (P<0.001), TNM stage (P=0.010), tumor differentiation (P<0.001), and modified Glasgow Prognostic Score (mGPS) (P<0.004). The proportion of patients found to have serum ferritin <15 μ g/L was significantly different amongst the no anemia, microcytic anemia and normocytic anemia groups respectively whether unadjusted (P<0.001), adjusted by CRP alone (P<0.001), adjusted by albumin alone (P<0.001) or adjusted by both CRP and albumin (P<0.001) values were considered. The proportion of patients found to have serum iron <10 μ mol/L was significantly different amongst the no anaemia, microcytic anemia and normocytic anemia groups using the unadjusted values only (P=0.007)

Discussion

7	2	\cap
_	2	υ

229

231 The results of the present study report linear regression based correction equations to adjust observed serum iron and ferritin concentrations based on CRP and albumin in a cohort of 232 primary and secondary care patients, and patients with colorectal cancer. 233 The BRINDA Working Group have previously reported the use of a linear regression based 234 correction equation for ferritin adjusted by CRP and AGP in populations across multiple 235 countries [8]. Unlike serum albumin, AGP is used to define the "phase" of inflammation, 236 rather than its magnitude making estimation both dependent on their concentrations and the 237 phase in which the patient is found [15]. In addition, unlike albumin, AGP is not at present 238 widely used in clinical practice, especially in the developing countries from which many of 239 the BRINDA cohorts were derived [13]. 240 Although prior studies have suggested the use of correction factors based cut off values of 241 acute phase proteins [6], or changing the ferritin concentration threshold for the diagnosis of 242 iron deficiency to <30 µg/L to account for inflammation [14], the BRINDA group show that 243 serum ferritin concentrations are significantly affected even at low levels of inflammation, 244 below those cut off values used previously [9]. The same is true for serum total iron [11], 245 which in addition is highly variable from individual to individual, within individuals, and is 246 also significantly affected by dietary intake [3]. 247 These findings are in keeping with the results of the present study and the previously reported 248 results from this cohort in which patients were grouped by CRP and albumin [11]. In it, 13% 249 of patients with CRP <10mg/L and albumin >35g/L (therefore thought to be minimally 250 inflamed), were still found to have ferritin <15 µg/L. Although the use of such groupings to 251 account for inflammatory status is useful, the BRINDA group suggest that regression based 252

correction, as in the present paper, provides greater accuracy. Particularly, as with CRP and AGP in the BRINDA papers, because even low and "sub-clinical" levels of systemic inflammation can significantly alter serum measures of iron status [16]. This study differs from our previously reported work by using continuous regression equation adjustment rather than grouping in attempt to account for the aforementioned "sub-clinical" inflammation. Indeed, in the present study the CRP and albumin reference values used to prevent overcorrection for low level inflammation were 1.3 mg/L and 40.9 g/L respectively, much different from "minimally inflamed" group threhsolds used previously. The greatest difference in the proportion of iron deficient patients was found when adjustment was made for both CRP and albumin together. It may be that this relates to the different half-lives of CRP and albumin, with CRP peaking around 48 to 72 hours following an inflammatory insult and then ending to decrease rapidly, with albumin taking much longer to return to reference range concentrations [17]. It could be considered in this context that albumin is perhaps a longer term index of inflammation and CRP a short term marker, hence the additive effect with regard to ferritin and iron correction. The present study reports the application of CRP and albumin based regression correction to a small cohort of patients with colorectal cancer. A high proportion of patients with normocytic anaemia (57%) and without anaemia (29%) were found to be apparently iron deficient. This may represent patients who have functional iron deficiency related to the presence of a host systemic inflammatory response to cancer. At present there is little data to suggest whether iron replacement therapy will be of use in those patients with inflammation who meet definitions of iron deficiency following mathematical adjustment compared to those who do not. This is indeed an area requiring prospective investigation, and hopefully the use of such correction strategies might allow patient stratification in such studies.

253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

The present study has a number of limitations. The lack of clinical data regarding reasons for the investigation of iron status in the first cohort means that there were no data relating to the presence of underlying autoimmune disease or liver disease or markers of synthetic liver function associated with treated systemic inflammation or altered iron homeostasis. Within the second cohort, the selection of patients for whom ferritin had been requested may introduce selection bias, although the prevalence of anaemia, and its subtypes is in keeping with recently published reports [18].

In summary the present study reports a regression correction equation to adjust serum iron and ferritin concentrations based on CRP and albumin in a cohort of patients in primary and secondary care, and in a cohort of patients with colorectal cancer. In both cohorts the greatest difference in the proportion of iron deficient patients was found when adjustment was made for both CRP and albumin together. Even very low levels of inflammation were associated with significant perturbment of ferritin. Although the presented regression equations do not produce perfect agreement amongst different measures of iron status, their use did improve concordance and suggest that with ongoing refinement further similar techniques may prove useful in the absence of novel markers of iron status.

Disclosure

None of the authors have any conflicts of interest.

Contributions

SM, IJ, DM and DT designed research. AT and IJ conducted research. AT, SM and IJ analyzed data. SM, IJ, DM and DT wrote the paper, SM had primary responsibility for final content. All authors read and approved the final manuscript.

Acknowledgements

Special thanks are due to our retired colleagues Dr Denis O'Reilly and Dr Andrew Duncan who provided advice during the planning of the study.

References

- 1. McLean E, Cogswell M, Egli I, Wojdyla D, de Benoist B. Worldwide prevalence of anaemia, WHO Vitamin and Mineral Nutrition Information System, 1993-2005. Public Health Nutr 2009;12:444-454
- 2. World Health Organization/Centers for Disease Control and Prevention. Assessing the iron status of populations. Geneva (Switzerland): WHO;2007
- 3. Kelly AU, McSorley ST, Patel P, Talwar D. Interpreting iron studies. BMJ;2017:357:j2513.
- 4. Thomas DW, Hinchcliffe RF, Briggs C, Macdougall IC, Littlewood T, Cavill I. British Committee for Standards in Haematology. Guidelines for laboratory diagnosis of functional iron deficiency. Br J Haematol 2013;161:639-648
- 5. World Health Organization. Serum ferritin concentrations for the assessment of iron status and iron deficiency in populations. Vitamin and Mineral Nutrition Information System (WHO/NMH/NHD/MNM/112) Geneva (Switzerland) WHO;2011
- 6. Thurnham DI, Northrop-Clewes CA, Knowles J. The use of adjustment factors to address the impact of inflammation on vitamin A and iron status in humans. J Nutr 2015;145:1137S-1143S
- 7. Rohner F, Namaste SML, Larson LM, Addo OY, Mei Z, Suchdev PS, Sakr Ashour FA, Rawat R, Raiten DJ, et al. Adjusting soluble transferrin reception concentrations for inflammation: Biomarkers Reflecting Inflammation and Nutritional Determinants of Anemia (BRINDA) project. Am J Clin Nutr 2017;106(Suppl):383S-389S

- 8. Namaste SM, Aaron GJ, Varadhan R, Peerson JM, Suchdev PS, BRINDA working group.

 Methodologic approach for the Biomarkers Reflecting Inflammation and Nutritional

 Determinants of Anemia (BRINDA) project. Am J Clin Nutr 2017;106(S1):333S-347S
- 9. Suchdev PS, Young MF, Williams AM, Addo Y, Namaste SL, Aaron GJ, Neufeld L, Raiten DJ, Flores-Ayala R, BRINDA Working Group. Reply to ST McSorley et al. Am J Clin Nutr 2018;108:(1):202-203
- 10. McSorley ST, Talwar D, McMillan DC. Comment on the Biomarkers Reflecting Inflammation and Nutritional Determinants of Anemia (BRINDA) project. Am J Clin Nutr 2018;108(1):204-205
- 11. McSorley ST, Jones I, McMillan DC, Talwar D. Quantitative data on the magnitude of the systemic inflammatory response and its relationship with serum measures of iron status. Trans Res 2016;176:119-126
- 12. Perlmutter DH, Dinarello CA, Punsal PI, Colten HR. Cachectin/tumor necrosis factor regulates hepatic acute-phase gene expression. J Clin Invest. 1986;78(5):1349-54.
- 13. Namaste SM, Rohner F, Huang J, Bhushan NL, Flores-Ayala R, Kupka R, Mei Z, Rawat R, Williams AM, Raiten DJ et al. Adjusting ferritin concentrations for inflammation:

 Biomarkers Reflecting Inflammation and Nutritional Determinants of Anemia (BRINDA)

 project. Am J Clin Nutr 2017 doi: 10.3945/ajcn.116.141762 [Epub ahead of print]
- 14. World Health Organization. Haemoglobin concentrations for the diagnosis of anaemia and assessment of severity. Vitamin and Mineral Nutritional Information System. Geneva, World Health Organization, 2011 (WHO/NMH/NHD/MNM/11.1)
- 15. Thurnham DI, Northrop-Clewes CA. Inflammation and biomarkers of micronutrient status. Curr Opin Clin Nutr Metab Care 2016;19:458–63

- 16. Suchdev PS, Young MF, Williams AM, Addo Y, Namaste SML, Aaron GJ, Neufeld L, Raiten DJ, Flores-Ayala R on behalf of the BRINDA Working Group. Reply to McSorley et al. Am J Clin Nutr 2018;108:202-203
- 17. Gabay C, Kushner I. Mechanisms of disease: acute-phase proteins and other systemic responses to inflammation. N Engl J Med 1999;340:448-454
- 18. Vayrynen JP, Tuomisto A, Vayrynen SA, Klintrup K, Karhu T, Makela J, Herzig KH, Karttunen TJ, Makinen MJ. Preoperative anemia in colorectal cancer: relationships with tumor characteristics, systemic inflammation, and survival. Sci Rep 2018;8:1126

Tables and Footnotes

Table 1 Estimated prevalence of iron deficiency; serum ferritin $<15\mu g/L$ or serum iron <10 μ mol/L, based on regression correction using serum C-reactive protein (CRP) and albumin in an unselected cohort of primary and secondary care patients.¹

Iron measure	Unadjusted	CRP only	Albumin only	CRP and albumin	P
Serum Ferritin					
n	7226	7219	6406	6406	-
Ferritin, μg/L,	162 (57-385)	69 (28-161)	89 (35-203)	38 (15-92)	$< 0.001^2$
Ferritin $<$ 15 µg/L, n (%)	479 (7)	939 (13)	679 (11)	1557 (24)	<0.001 ³
Serum Iron					
n	7182	7182	6371	6371	-
Iron, μmol/L,	8 (4-15)	15 (10-23)	12 (7-19)	20 (13-32)	$< 0.001^2$
Iron <10 μmol/L, <i>n</i> (%)	4027 (55)	1877 (26)	2705 (37)	925 (15)	< 0.0013

¹Continuous values are presented as median and IQR, ²Medians of all four adjustment methods compared using related samples Friedmann's two way analysis of variance by ranks, ³Proportions of "Unadjusted" and "CRP and albumin adjusted" methods compared using McNemar's test. CRP, C-reactive protein; IQR, interquartile range.

Table 2 Concordance between serum ferritin and serum iron definitions of iron deficiency before and after adjustment by serum C-reactive protein (CRP) and albumin:

Unadjusted			
		Iron <10μmol/L	ı
		No, <i>n</i> (%)	Yes, <i>n</i> (%)
Ferritin <15µg/L	No, n(%)	3059 (43)	3644 (51)
	Yes, n(%)	96 (1)	383 (5)
Adjusted by CRP an albumin	d		
		Iron <10μmol/L	1
		No, <i>n</i> (%)	Yes, <i>n</i> (%)
Ferritin <15µg/L	No, n(%)	4253 (66)	630 (10)
, -	Yes, n(%)	1241 (19)	294 (5)

CRP C-reactive protein

Table 3 Estimated prevalence of iron deficiency; serum ferritin $<15\mu g/L$, or serum iron $<10\mu mol/L$; based on regression correction using serum C-reactive protein (CRP) and albumin in patients undergoing surgery for colorectal cancer.¹

Iron measure	Unadjusted	CRP only	Albumin only	CRP and albumin	P
Serum Ferritin					_
n	356	307	236	236	-
Ferritin, µg/L	38 (13-93)	24 (8-66)	24 (9-70)	18 (6-52)	$< 0.001^2$
Ferritin <15 μ g/L, n (%)	100 (28)	120 (39)	91 (39)	107 (46)	< 0.0013
Serum Iron					
n	67	58	46	46	-
Iron, μmol/L	5 (3-13)	8 (3-19)	6 (4-17)	11 (5-26)	$< 0.001^2$
Iron <10 μmol/L, <i>n</i> (%)	44 (66)	33 (57)	27 (59)	20 (44)	0.004^{3}

¹Continuous values are presented as median and IQR. ²Medians of all four adjustment methods compared using related samples Friedmann's two way analysis of variance by ranks, ³Proportions of "Unadjusted" and "CRP and albumin adjusted" methods compared using McNemar's test. CRP, C-reactive protein; IQR, interquartile range.

Table 4 Association between anemia¹ and estimated prevalence of iron deficiency (ferritin $<15\mu g/L$) based on regression correction using serum C-reactive protein (CRP) and albumin in patients undergoing surgery for colorectal cancer.

Anemia ¹		None	Microcytic ²	Normocytic ³	P
n		174 (50)	41 (11)	139 (39)	-
Demographic					
Sex, <i>n</i> (%)					0.398^{4}
	Male	89 (51)	24 (59)	81 (58)	
	Female	85 (49)	17 (41)	58 (42)	
Age, <i>n</i> (%)					< 0.0015
	<65	66 (38)	16 (39)	24 (17)	
	65-70	73 (42)	12 (29)	51 (37)	
	>70	35 (20)	13 (32)	64 (46)	
ASA, <i>n</i> (%)					< 0.0015
	1	42 (28)	6 (16)	12 (11)	
	2	63 (43)	17 (45)	50 (45)	
	3	39 (26)	14 (37)	45 (40)	
	4	4 (3)	1 (2)	5 (5)	
nCRT, <i>n</i> (%)					0.739^4
	Yes	8 (5)	2 (5)	9 (7)	
	No	165 (95)	38 (95)	127 (93)	
Pathological					
Tumor site, n (%)					< 0.0014
	Colon	106 (61)	39 (95)	113 (82)	
	Rectum	67 (39)	2 (5)	25 (18)	
TNM stage, n (%)					0.010^{5}
	0/1	44 (26)	1 (3)	15 (11)	
	2	66 (38)	18 (45)	64 (47)	
	3	60 (34)	18 (45)	51 (38)	
	4	3(2)	3 (7)	6 (4)	
Differentiation, n (%)			, ,	. ,	< 0.0014
, , ,	Well / Mod	166 (97)	29 (74)	118 (88)	
	Poor	5 (3)	10 (26)	16 (12)	
Venous invasion, n (%)		. ,	,	,	1.000^4
, , ,	Present	111 (64)	25 (64)	88 (64)	
	Absent	62 (36)	14 (36)	49 (36)	
Systemic inflammation		· /	,	,	
mGPS					0.004^{5}
	0	114 (80)	25 (76)	76 (72)	
	1	18 (13)	3 (9)	5 (5)	
	2	10 (7)	5 (15)	24 (23)	
Serum Ferritin		()	,	,	
Unadjusted, n (%)					< 0.0014
(n=354)					
,	$<15\mu g/L$	26 (15)	28 (68)	46 (33)	
	≥15μg/L	148 (85)	13 (32)	93 (67)	
CRP only, $n (\%)(n=305)$		- 10 (00)	()	7 (0 1)	< 0.0014
= - 3, - ()()	$<15\mu g/L$	33 (22)	31 (80)	56 (48)	
	≥15μg/L	117 (78)	8 (21)	60 (52)	
Albumin only, n (%)	=10 MB/ Z	11, (,0)	0 (21)	00 (02)	< 0.0014
(n=234)					0.001
(· ·)	$<15\mu g/L$	25 (21)	23 (85)	43 (50)	
	≥15µg/L	96 (79)	4 (15)	43 (50)	
CRP and albumin, n	ro =	()	. (-0)	- (50)	< 0.0014
(%)(n=230)					0.001
(70)(11 230)	<15µg/L	34 (29)	24 (89)	48 (57)	
	$\geq 15 \mu g/L$ $\geq 15 \mu g/L$	84 (71)	3 (11)	37 (44)	
	_15µg/L	07 (71)	J (11)	31 (77)	

Serum Iron Unadjusted, n (%) (n =66)					0.007^{4}
,	<10µmol/L	5 (36)	12 (92)	27 (69)	
	≥10μmol/L	9 (64)	1(8)	12 (31)	
CRP only, n (%) (n =57)	·	. ,	. ,	, ,	0.129^{4}
	<10µmol/L	5 (36)	8 (73)	20 (63)	
	≥10µmol/L	9 (64)	3 (27)	12 (37)	
Albumin only, <i>n</i> (%) (<i>n</i> =45)	·				0.055^4
	<10µmol/L	5 (36)	6 (86)	16 (67)	
	≥10µmol/L	9 (64)	1 (14)	8 (33)	
CRP and albumin, <i>n</i> (%) (<i>n</i> =45)	·		` ,	, ,	0.173^4
	$<10\mu mol/L$	4 (29)	5 (71)	11 (46)	
	≥10μmol/L	10 (71)	2 (29)	13 (54)	

¹ Males = Hb <130g/L, females <120g/L, ²Anemia and MCV <80fL, ³Anemia and MCV 80-100fL. ⁴Chi square test, ⁵Chi square test for linear association. ASA, American Society of Anesthesiology; mGPS, modified Glasgow Prognostic Score; MCV, mean corpuscular volume; nCRT, neoadjuvant chemoradiotherapy; TNM, tumor nodes metastasis stage.



