

Comparison of endotoxin levels in cow's milk samples derived from farms and shops

Sándor Sipka¹, Andrea Béres², Lóránd Bertók³, Tamara Varga² and Geza Bruckner⁴

Innate Immunity
2015, Vol. 21(5) 531–536
© The Author(s) 2014
Reprints and permissions:
sagepub.co.uk/journalsPermissions.nav
DOI: 10.1177/1753425914557100
ini.sagepub.com



Abstract

The observations on the protective effect of bacterial endotoxin in farm-derived cow's milk on childhood asthma and allergy are contradictory. The aim of this study was to determine the endotoxin levels in 'farm-derived whole raw' and 'processed shop' sources of cow's milk, and to test how the temperature and storing conditions might alter their endotoxin concentrations. Milk was collected from farms and shops. The level of endotoxin was measured by micro (gel-clot) *Limulus* amoebocyte lysate test expressed as EU/ml. The concentration ranges of endotoxin were much higher and more widely scattered in the samples of whole raw farm milk than in the processed shop milk. Cold storage or heating increased the endotoxin concentrations in all samples of farm milk, but not in the processed shop milk. These results show that elevated levels of endotoxin in raw farm milk samples can occur from the cowshed or be formed during storage. In processed shop milk, storage does not cause any changes in the amount of endotoxin. Therefore, it is consistent that the handling and storage of raw milk alters the endotoxin concentrations, which may explain previous contradictory findings regarding the beneficial modulating effects on innate immunity toward allergy prevention in early childhood.

Keywords

Allergy prevention, cow's milk, endotoxin, temperature dependence

Date received: 5 May 2014; revised: 2 September 2014; accepted: 17 September 2014

Introduction

The GABRIEL Advanced Studies and an earlier study show the protective effects of Alpine farm environments on childhood asthma and allergy,^{1,2} and suggest that the daily consumption of farm milk is one of the protective factors. Increased endotoxin concentrations were found in the house dust of farms compared with urban environments, suggesting a possible protective effect against the development of atopy.^{3–6} Other microbial products also have been associated with a reduced risk of allergy and asthma.^{7,8} In the PASTURE study, endotoxin concentrations were found to be significantly higher in milk samples from non-farming families compared with farming families, but no significant difference was detected in the endotoxin levels of shop milk and farm milk samples.⁹ After publication of this article, a new series of analysis (GABRIELA study) concluded that the beneficial, asthma and allergy protecting constituents of milk

could be related to the whey fraction of unprocessed cow's milk,^{10,11} and not to the microbial contamination of milk samples. Recently, increased regulatory T cell numbers were found to be associated with farm milk exposure and lower atopic sensitization and asthma in childhood.¹²

The aim of our study was to compare the endotoxin concentration in samples of 'farm-derived whole raw'

¹Division of Clinical Immunology, Institute of Internal Medicine, University of Debrecen, Debrecen, Hungary

²Pharmaceutical Control and Development Laboratory, Budapest, Hungary

³National Research Institute of Radiobiology and Radio Hygiene, Budapest, Hungary

⁴Division of Clinical Nutrition, University of Kentucky, KY, USA

Corresponding author:

Sándor Sipka, Division of Clinical Immunology, University of Debrecen, Nagyerdei krt 98, 4032 Debrecen, Hungary.
Email: sipka@iibel.dote.hu

and 'processed shop' sources of cow's milk under strict pre-analytical conditions. Further, the effects of storing, freezing, heating and boiling were tested to determine if they affected endotoxin concentrations.

Methods

The measurement of endotoxin concentration in various cow's milk samples

Cow's milk samples: (a) whole raw milk collected from cowsheds and farm tanks fat level $>3.5\%$ ($n=15$); (b) processed shop milk, fat level $2.8\text{--}1.5\%$ ($n=8$). All processed milk samples were previously homogenized and heat-treated by their manufacturers using a combination of three methods: (1) pasteurization; (2) extended shelf life (ESL) technique; (3) micro-filtration. All milk samples were collected and maintained at 4°C until the special heat/cold treatments and the measurements were carried out; all samples were collected in endotoxin-free glass bottles. Endotoxin levels measured within 24 h represented the 'fresh' values. In all samples, just prior to heat or cold treatments the endotoxin concentrations were again determined (basal concentrations; basal I, II, III). The changes in the endotoxin concentrations caused by the treatments were compared to these basal concentrations.

The temperature treatments for the 500 ml milk samples were as follows: (1) freezing at -16°C for 72 h; (2) cold storage (cooling) at 4°C for 72 h (3) heating at 98°C for 10 min in a water bath; (4) boiling for 10 min at 100°C . Measurement of endotoxin in milk samples took place by gel-clot micro *Limulus* amoebocyte lysate (LAL) (Pyrotell; Associates of Cape Cod Inc., East Falmouth, MA, USA.) using LAL Reconstitution Buffer (Pyrosol; Associates of Cape Cod Inc.) and concentration standards (Control Standard Endotoxin; Associates of Cape Cod Inc.) at pH 7.2–7.8, always in the presence of both positive and negative controls. The range of pH in the various milk samples without LAL buffer was 6.1–6.7. The validity of bacterial endotoxin micro-LAL test was pH 6.0–8.0. All samples reached room temperature before they were treated or diluted by endotoxin-free LAL water (PCDL, Debrecen, Hungary). The dilutions were prepared after the samples and LAL water reached 38°C . Thus, there was no temperature and difference between the milk and water diluted milk samples throughout the measurements carried out in comparable homogeneous solutions. The minimum dilution was 1:10, the maximum dilution was 1:100,000. At first, a 10-fold dilution series was prepared, followed by a two-fold dilution series. The amount of endotoxin was expressed in endotoxin units EU/ml. The losses in the volumes during heating and boiling were corrected in concentration calculations. The detection limit of the assay was 0.03 EU/ml.

Statistical analysis

For statistical analysis the non-parametric Mann-Whitney test was used in the two sources of milk. A P -value <0.05 was regarded as significant. In addition, 'median', 'minimum' and 'maximum' values also were calculated. In Figure 1, the thick lines in the box-plots show medians, whereas the boxes reflect the 50 percentiles. The calculations were carried out using the statistical software SPSS 20.0 (IBM, Armonk, NY, USA).

Results

Box-plots of endotoxin concentrations (EU/ml) in the samples of whole raw farm-derived and processed shop milks

The peak concentration of endotoxin was much higher in the whole raw farm milk (6144 EU/ml) than in the processed shop milk samples (240 EU/ml); however, the difference was not significant between the two groups ($P=0.538$) owing to the large variance, whereas the median value of farm milk was 60.0 and the median value of shop milk was 102.5 EU/ml in this series of samples.

Table 1 denotes data for all of the milk sources regarding their 'fresh' endotoxin values measured within 24 h, differences in the fat contents, milking times, places of collection, the forms of processing and the number of days guaranteed for use (expiration days).

Both of the treatments at high temperatures (98°C and 100°C) and the prolonged cold storage time (72 h) of farm milk at 4°C (cooling) resulted in large increases in the concentrations of endotoxin (2.5–50.0-fold and 2.5–223.5 fold) compared with the 'basal concentrations'; the largest increases occurred in whole raw farm milk samples. However, freezing at -16°C prevented the increase of endotoxin level during the storage for 72 h. In the processed shop milk, neither the increased storage time (72 h) nor the cooling (4°C), freezing (-16°C), heating (98°C) or boiling (100°C) caused any remarkable increases (changes) in the concentrations of endotoxin (Table 2). The complete treatment procedure, including storage and temperature (cooling, freezing, heating and boiling) treatments could only be carried out *simultaneously* in three of the whole raw farm-derived milk samples.

Discussion

The data indicate that by using controlled individual sample collections, storage and treatment for various sources of cow's milk, marked differences occur in the concentrations of endotoxin. In contrast to the processed shop milk, whole raw farm milk can contain much higher concentrations of endotoxin likely derived from contamination in the sheds. It appears that longer storage time, even at 4°C , and heating boiling

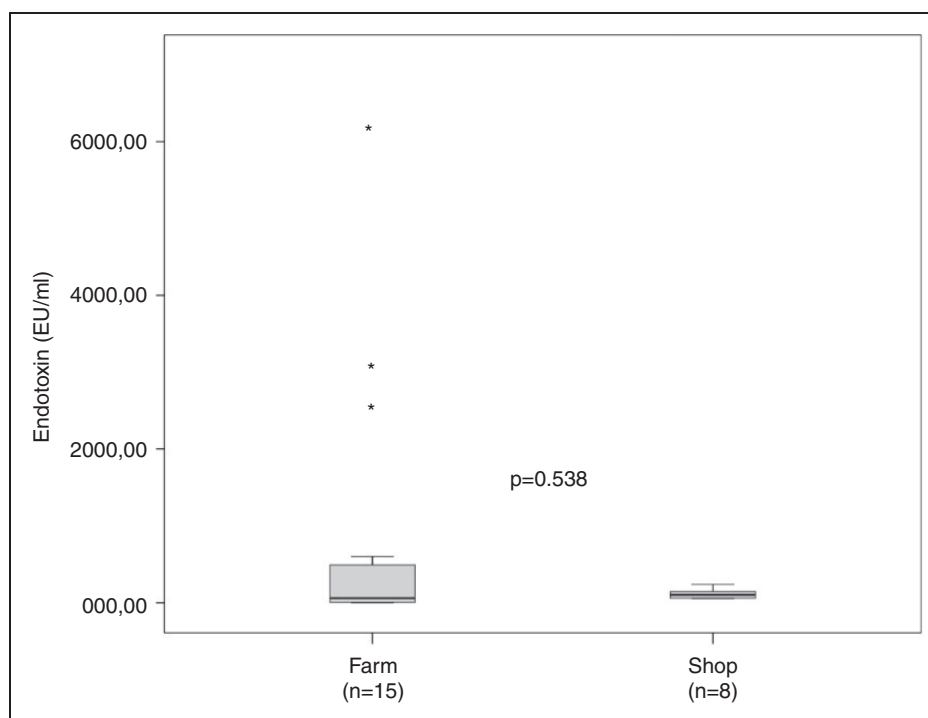


Figure 1. Box-plots of endotoxin concentrations (EU/ml) in the samples of whole raw farm-derived and processed shop milks. The thick lines in the box-plots show medians, whereas the boxes reflect the 50 percentiles of data.

Table 1. The 'fresh' endotoxin levels, fat percentages, origin, forms of processing and types of various cow's milk samples derived from farms and shops stored at 4°C for 24 h.

Whole raw farm milks (fresh)					Processed shop milks (fresh)				
Origin	Endotoxin EU/ml	Milking	Fat (%)	Source	Origin	Endotoxin EU/ml	Fat (%)	Processing	Days guaranteed
1.A	3	E	>3.5	Tank	1.E	60	2.8	Homogenized pasteurized	3–10
2.A	3	E	>3.5	Tank	2.E	60	2.8	Homogenized heat-treated	14–21
3.A	6	E	>3.5	Tank	3.F	120	2.8	with ESL technology	14–21
4.A	6	E	>3.5	Tank	4.F	240	1.5		14–21
5.A	6	E	>3.5	Tank	5.G	120	2.8	Homogenized pasteurized	14–21
6.A	1.5	E	>3.5	Tank	6.G	120	2.8	microfiltered	14–21
7.A	96	M	>3.5	Tank	7.G	60	1.5		14–21
8.B	384	M	>3.5	Tank	8.G	60	1.5		14–21
9.C	600	M	>3.5	Tank					
10.C	2400	E	>3.5	Shed					
11.C	6144	M	>3.5	Shed					
12.C	3000	E	>3.5	Shed					
13.D	60	M	>3.5	Tank					
14.D	120	E	>3.5	Tank					
15.D	12	E	>3.5	Tank					

1A–15D: places of farm milk purchase; 1E–8G: places of shop milk purchase; E: evening; M: morning.

Table 2. The effects of storing, cooling, freezing, heating and boiling on the results of the measurements of endotoxin levels in cow's milk samples derived from farms and shops compared to the basal concentrations (basal I, II, III) measured just before the treatments.

Type of milk	Milk sample	Endotoxin EU/ml						
		Basal I.	Refrigerated stored 4°C 72 h	Frozen stored -16°C 72 h	Basal II	Refrigerated and heat treated 98°C 10 min	Basal III	Refrigerated and boiled >100°C 10 min
Whole raw farm	4.A	12	n.t.	n.t.	n.t.	n.t.	n.t.	120 (10×)
	5.A	12	n.t.	n.t.	n.t.	n.t.	n.t.	600 (50×)
	6.A	3	n.t.	n.t.	n.t.	15 (5×)	n.t.	n.t.
	7.A	192	n.t.	n.t.	n.t.	192 (1×)	n.t.	n.t.
	8.B.	768	n.t.	n.t.	n.t.	6000 (7.8×)	n.t.	n.t.
	9.C	1200	3000 (2.5×)	2400 (2×)	n.t.	n.t.	n.t.	n.t.
	10.C	2400	6000 (2.5×)	2400 (1×)	n.t.	n.t.	n.t.	n.t.
	11.C	12,288	n.t.	n.t.	n.t.	15,000 (1.2×)	n.t.	n.t.
	12.C	6000	n.t.	n.t.	n.t.	n.t.	n.t.	60,000 (10×)
	13.D	120	3000 (25×)	240 (2×)	6000	60,000 (10×)	12,000	120,000 (10×)
	14.D	240	240 (1×)	240 (1×)	300	12,000 (40×)	2400	6000 (2.5×)
	15.D	24	2683 (223.5×)	24 (1×)	3000	12,000 (4×)	3000	12,000 (4×)
	Processed shop	2E	120	n.t.	n.t.	n.t.	n.t.	n.t.
3F		120	120 (1×)	120 (1×)	n.t.	n.t.	n.t.	n.t.
4F		240	268 (1.1×)	240 (1×)	n.t.	n.t.	n.t.	n.t.
5G		120	n.t.	n.t.	n.t.	120 (1×)	n.t.	120 (1×)
6G		120	120 (1×)	120 (1×)	n.t.	n.t.	n.t.	n.t.
7G		60	60 (1×)	60 (1×)	n.t.	n.t.	n.t.	n.t.
8G		60	n.t.	n.t.	n.t.	60 (1×)	n.t.	120 (2.0×)

n. t.: non-tested; × : fold of elevation; IA–I5D: places of farm milk purchase; IE–8G: places of shop milk purchase.

procedures increase the level of endotoxin, but only in the unprocessed farm milk. The change may be correlated with the milk fat content or that the shop milk has been processed. In milk with higher fat content, the higher concentration of endotoxin may be due to the farm milieu because even the house dust contains more endotoxin than in a non-farm environment;^{3–6} therefore, endotoxin may enter via the airways by inhalation,¹³ and thus may be transferred from the blood into the fat fraction of milk. The partly lipid character of endotoxin and its sequestration in lipid, protein and aqueous colloid components may contribute to the temperature sensitivity observed. The farm milk with higher fat content appeared to have higher endotoxin concentrations than the processed shop milks with lower fat levels. The farm milk samples stored at 4°C and/or heated showed increased endotoxin values compared with basal concentrations; this trend did not occur in the shop milk, possibly owing to the lower fat content or to the absence of a 'microbial load' (bacterial counts were not determined). The elevated endotoxin concentration in farm milk may explain the asthma and atopy protective effect of farm milk noted in the GABRIELA study.^{1,2} However, our results are only in a partial agreement with the PASTURE study, which showed a trend for shop milk having higher

endotoxin concentrations than the farm milk; this is contrary to our findings.⁹ Our data clearly show that the concentration of endotoxin in farm cow's milk samples depended on the temperature treatments (cooling, freezing, heating, boiling). It was of a special interest that during prolonged storage at 4°C great increases (223.5-fold) were noted, which could occur in raw farm milk. We speculate that the reason for these changes are due to increased destruction of the persisting Gram-negative bacteria at this temperature; however, a significant part of the raw farm milk samples stored and cooled at 4°C for 72 h had an increased endotoxin concentration. However, freezing at -16°C prevented the increase of endotoxin level during the 72-h storage.

Two important observations in the GABRIELA study were: (1) raw milk consumption was inversely associated with childhood asthma; (2) boiled farm milk did not show a protective effect.¹⁰ Our findings indicate that heating of the raw farm milk samples can increase endotoxin concentrations and therefore should have offered more and not less protection. An explanation for this phenomenon might be related to endotoxin being bound in milk by a great variety of proteins, some being required for its biological activity, for example LPS binding protein,^{14,15}

immunoglobulins, complement factors,^{16,17} lysozyme^{18,19} and high mobility group box 1 protein (HMBG1).^{20,21} However, a part of these proteins, for example certain complement factors, as well as lysozyme and HMBG1, are heat-labile molecules. Their biological activity and endotoxin binding capacity may be altered by heating and boiling thus altering their activity.

We speculate that the controversy related to endotoxin's role in asthma and allergy prevention in whole raw farmer cow's milk may be due to the protein/lipid/endotoxin interactions, which are heat and storage sensitive, and/or differences in the microbial load. The whey protein fraction of unprocessed bovine milk, which has been implicated in allergy prevention, may also be affected with regard to temperature sensitivity.¹¹

Of note, bile acids have been shown to block the intestinal absorption of endotoxin.^{22–24} This pathway plays a crucial role in the regulation of innate immunity in early childhood (< 4 yr) when the bile acid production is still low.²⁵ Thus, the endotoxin molecules derived either from fresh or cold stored (cooled at 4°C) raw farm milk can attain higher blood concentration in this age group than from processed shop milk resulting in a driving developing immune system toward Th1 responses causing allergy protection,^{4,26} as bacterial isolates from cowsheds also had strong allergy protective properties imparted to the lipopolysaccharide moiety.^{27,28}

The very recent article, on the increased T regulatory cell (Treg) numbers associated with farm milk exposure has two important observations:¹² (1) the stronger LPS reaction of children drinking farm milk (possibly stored at 4°C) but not living on the farm vs. farm children drinking fresh farm milk may be explained by our data that very often the endotoxin content (and biological effect) of cooled stored raw farm milk can contain higher LPS concentrations than that found in fresh raw milk; (2) IL-1 β induced by farm milk endotoxin from the monocytes can stimulate the expansion and differentiation of peripheral Treg (immunosuppressive) cells.^{14,29}

In conclusion, our data suggest that elevated concentrations of endotoxin can occur rather often in farm-derived (especially cold-stored, cooled) cow's milk and its potential protective effect on asthma and allergy prevention cannot be neglected. However, both the concentration and the bioactivity of endotoxin, found in farm milk can be influenced by storage time and temperature treatment, which may alter protein/lipid/endotoxin interactions.³⁰ In addition, the consumption of raw farm milk might have all the risks and health hazards associated with the unpasteurized, unprocessed state.

Funding

This study was supported by the "Hungarian Research Fund" (OTKA No. 71883), and the "Foundation for Immunology XXI" from Debrecen.

Conflict of interest

The authors do not have any potential conflicts of interest to declare.

Acknowledgements

We acknowledge the laboratory work of all members of the Department of Toxicology in the Pharmaceutical Control and Development Laboratory in Budapest, but especially the contribution of Dr. Éva Kiss. We are grateful for the very useful help given by Professor Dr. Erika von Mutius (München) during the preparation of the manuscript.

References

1. Illi S, Depner M, Genuneit J, et al. Protection from childhood asthma and allergy in Alpine farm environments- the GABRIEL Advanced Studies. *J Allergy Clin Immunol* 2012; 129: 1470–1477.
2. Riedler J, Eder W, Oberfeld G, et al. Austrian children living on a farm have less hay fever, asthma and allergic sensitization. *Clin Exp Allergy* 2000; 30: 194–200.
3. von Mutius E, Braun-Fahrlander C, Schierl R, et al. Exposure to endotoxin or other bacterial components might protect against the development of atopy. *Clin Exp Allergy* 2000; 30: 1230–1234.
4. Gereda JE, Leung DY, Thatayatikom A, et al. Relation between house-dust endotoxin exposure, type 1 T cell development, and allergen sensitisation in infants at high risk of asthma. *Lancet* 2000; 355: 1680–1683.
5. Gehring U, Bischof W, Fahlbusch B, et al. House dust endotoxin and allergic sensitization in children. *Am J Respir Crit Care Med* 2002; 166: 939–944.
6. Braun-Fahrlander C, Riedler J, Herz U, et al. Environmental exposure to endotoxin and its relation to asthma in school-age children. *N Engl J Med* 2002; 347: 869–877.
7. Heederik D and von Mutius E. Does diversity of environmental exposure matter for the occurrence of allergy and asthma? *J Allergy Clin Immunol* 2012; 130: 44–50.
8. Wlasiuk G and Vercelli D. The farm effect, or: when, what and how a farming environment protects from asthma and allergic disease. *Curr Opin Allergy Clin Immunol* 2012; 12: 461–466.
9. Gehring U, Spithoven J, Schmid S, et al. Endotoxin levels in cow's milk samples from farming and non-farming families. The PASTURE study. *Environment International* 2008; 34: 1132–1136.
10. Loss G, Apprich S, Waser M, et al. GABRIELA study group: The protective effect of farm milk consumption on childhood asthma and atopy: the GABRIELA study. *J Allergy Clin Immunol* 2011; 128: 766–773.
11. von Mutius E. Maternal farm exposure/ingestion of unpasteurized cow's milk and allergic disease. *Curr Opin Gastroenterol* 2012; 28: 570–576.
12. Lluís A, Depner M, Gaugler B, et al. Increased regulatory T cell numbers are associated with farm milk exposure and lower atopic sensitization and asthma in childhood. *J Allergy Clin Immunol* 2014; 133: 551–559.
13. Alexis NE, Eldridge MW and Peden DB. Effect of inhaled endotoxin on airway and circulating inflammatory cell phagocytosis and CD11b expression in atopic asthmatic subjects. *J Allergy Clin Immunol* 2003; 112: 353–361.
14. Chaby R. Lipopolysaccharide binding molecules: transporters, blockers and sensors. *Cell Mol Life Sci* 2004; 61: 1697–1713.
15. Wheeler TT, Smolenski GA, Harris DP, et al. Host-defence related proteins in cow's milk. *Animal* 2012; 6: 415–422.
16. Korhonen H, Marnila P and Gill HS. Milk immunoglobulins and complement factors. *Br J Nutr* 2000; 84((Suppl.); 1): S75–S80.

17. Gewurz H, Mergenhagen SE, Nowotny A, et al. Interactions of the complement system with native and chemically modified endotoxins. *J Bacteriol* 1968; 95: 397–405.
18. Kuczynska B, Puppel K, Golebiewski M, et al. Differences in whey protein content between cow's milk collected in late pasture and early indoor feeding season from conventional and organic farms in Poland. *J Sci Food Agric* 2012; 92: 2899–2904.
19. Kudou M, Shiraki K, Fujiwara S, et al. Prevention of thermal interaction and aggregation of lysozyme by polyamines. *Eur J Biochem* 2003; 270: 4547–4554.
20. Furukawa Y, Hayashi T, Mizuta M, et al. Increased concentration of high mobility group box 1 protein in milk is related to the severity of bovine mastitis. *Vet Res Commun* 2011; 35: 47–54.
21. Youn JH, Oh YJ, Kim ES, et al. High mobility group box 1 protein binding into lipopolysaccharide facilitates transfer of lipopolysaccharide to CD14 and enhances lipopolysaccharide mediated TNF alpha production in human monocytes. *J Immunol* 2008; 180: 5067–5074.
22. Bertók L. Role of bile in detoxification of lipopolysaccharide. In: Schlessinger D (ed.) *Microbiology*. Washington, DC: American Society for Microbiology, 1980, pp.91–93.
23. Kocsár LT, Bertók L and Várteresz V. Effect of bile acids on the intestinal absorption of endotoxin in rats. *J Bacteriol* 1969; 100: 220–223.
24. Parlesak A, Schaeckeler S, Moser L, et al. Conjugated primary bile salts reduce permeability of endotoxin through intestinal epithelial cells and synergize with phosphatidylcholine in suppression of inflammatory cytokine production. *Crit Care Med* 2007; 35: 2367–2374.
25. Huang CT, Rodriguez JT, Woodward WE, et al. Comparison of pattern of fecal bile acid and neutral sterol between children and adults. *Am J Clin Nutr* 1976; 29: 1196–1203.
26. Roponen M, Hyvarinen A, Hirvarinen A, et al. Change in IFN-gamma producing capacity in early life and exposure to environmental microbes. *J Allergy Clin Immunol* 2005; 116: 1048–1052.
27. Debarry J, Garn H, Hanuszkiewicz A, et al. *Acinetobacter lwoffii* and *Lactococcus lactis* strains isolated from farm cowsheds possess strong allergy protective properties. *J Allergy Clin Immunol* 2007; 119: 1514–1521.
28. Debarry J, Hanuszkiewicz A, Stein K, et al. The allergy-protective properties of *Acinetobacter lwoffii* F78 are imparted by its lipopolysaccharide. *Allergy* 2010; 65: 690–697.
29. Brinster C and Shevach EM. Costimulatory effects of IL-1 on the expansion/differentiation of CD4⁺CD25⁺Foxp3⁺ and CD4⁺CD25⁺Fox3⁻ T cells. *J Leukoc Biol* 2008; 84: 480–487.
30. Chang JC, Chen CH, Fang LJ, et al. Influence of prolonged storage process, pasteurization, and heat treatment on biologically-active human milk proteins. *Pediatr Neonatol* 2013; 54: 360–366.