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8 **Effects of photoperiod and temperature on the development**

9 **of *Bonagota cranaodes***

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23 **Effects of photoperiod and temperature on the development**  
24 **of *Bonagota cranaodes***

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35 **Abstract.** The Brazilian apple leafroller, *Bonagota cranaodes* (Meyrick) (Lepidoptera:  
36 Tortricidae) was reared in the laboratory under a long-day (LD 14 : 10 h) and a short-  
37 day (LD 7 : 17 h) photoperiod at 22°C, and under two different temperatures (10-13°C  
38 and 21-22°C). The development time from larval to adult eclosion did not differ  
39 between the two photoperiods, but between the two temperature regimes. However, the  
40 larvae did not enter diapause, even at short day conditions and low temperatures. The  
41 number of adults obtained did not differ with temperature and light conditions. Field  
42 captures with pheromone traps showed that Brazilian apple leafroller occurs in apple  
43 orchards throughout the year and that population densities were lower in winter. Control  
44 measures should accordingly be taken during off-season.

45 **Key words.** Diapause, field trapping test, insect control, monitoring, sex pheromone.

## 46 **Introduction**

47 Insects in temperate *climate* zones are challenged to endure harsh temperature regimes  
48 and the absence of food resources during winter. They survive such unfavorable  
49 conditions in diapause. Some univoltine species undergo an obligatory, genetically  
50 fixed diapause. In other univoltine and all multivoltine species, the diapause is induced  
51 by external cues which indicate the end of the summer, such as decreasing day length or  
52 temperatures (Beck, 1980).

53 Control of orchard insects in temperate climate zones, such as Oriental fruit moth  
54 *Grapholita molesta* (Lepidoptera: Tortricidae) and codling moth, aims at the non-  
55 diapausing life stages. Pheromone-based methods are obviously restricted to the flight  
56 period of adult moths, but even insecticide sprays can hardly be used to control  
57 overwintering larvae, which are protected by a hibernaculum and which are hidden in  
58 the soil or under tree bark.

59 In orchards in tropical and subtropical climate zones, control measures against native  
60 insects are not necessarily restricted to the periods when trees are in leaf, as native  
61 species may have access to native host plants providing food resources throughout the  
62 year.

63 We are currently developing a pheromone-based control method for Brazilian apple  
64 leafroller *Bonagota cranaodes* (Coracini *et al.*, 2001, 2003), which is an important pest  
65 of apple in Southern Brazil and Uruguay (Lorenzato, 1984). One important part of this  
66 program is to time the use of pheromones. We have therefore monitored the occurrence  
67 of adult moths in fruit orchards throughout the year and we have investigated whether  
68 *B. cranaodes* undergoes diapause.

## 69 **Material and Methods**

### 70 *Insect rearing*

71 *B. cranaodes* were obtained from a laboratory rearing at Embrapa, Vacaria, Brazil,  
72 where the insects are reared on a semiartificial agar-based diet (Mani *et al.*, 1978).  
73 Insects were reared from first instar larvae until adults under four conditions, involving  
74 two photoperiods, LD 7 : 17 h and LD 14 : 10 h, and two temperatures, 10-13°C and 21-  
75 22°C.

### 76 *Development under different photoperiods*

77 Two containers (1.5 L of diet) infested with 500 first instar larvae (first generation)  
78 were kept under a LD 7 : 17 h photoperiod. The adults eclosing from these containers  
79 were counted daily, and the adults were transferred for mating and oviposition to cages  
80 which were kept in the same room. Four days after the last moth had emerged, the diet  
81 container was checked for remaining larvae and pupae. These were transferred to plastic

82 Petri dishes (9 x 3.5 cm) containing moistened filter paper. Eclosed adults were  
83 counted daily.

84 The larvae hatching from the oviposition cages (second generation) were placed in  
85 batches of 500 into containers with 1.5 L of agar diet. One of these container was kept  
86 under a LD 7 : 17 h photoperiod and the other one under a LD 14 : 10 h photoperiod,  
87 both at a constant temperature of 22°C. Adults were counted after eclosion, and the diet  
88 was checked for dead larvae.

#### 89 *Development under different temperatures*

90 In this experiment 500 newly hatched larvae (first generation) were placed in groups  
91 of 25 larvae each into small plastic recipients with 75 g of agar diet. The recipients were  
92 kept inside two climatic chambers, one with the temperature of 10-13°C and the other  
93 one with the temperature of 21-22°C, both at a constant photoperiod of LD 7 : 17 h. It  
94 was used the same procedure for counting dead larvae/pupae and adults eclosion as  
95 described above. The adults were transferred for mating and oviposition to cages which  
96 were kept in climatic chamber.

97 The larvae hatching from the oviposition cages (second generation) were placed in  
98 groups of 25 larvae each into small plastic recipients (75 g of agar diet), and kept inside  
99 the same climatic chamber as the first generation. It was counted dead larvae/pupae and  
100 adults eclosion.

#### 101 *Field trapping tests*

102 Trap tests were done at Rubi Apple Orchard, Vacaria-RS, Brazil, from January to  
103 December, 2004. Tetra traps (Arn *et al.*, 1979) were baited with 10 µg of the optimized  
104 four-component sex pheromone blend (Coracini *et al.*, 2001), formulated on red rubber

105 septa (Merck ABS, Dietikon, Switzerland). Chemical and isomeric purity of the  
106 compounds was >99.5%.

107 The traps ( $n = 10$ ) were placed at ca. 1.7 m in apple trees. Traps were 5 m apart, and  
108 were arranged in random order in a line along tree rows. Traps were inspected once a  
109 week.

#### 110 *Statistical analysis*

111 Prior to statistical analysis, data were checked for ANOVA assumptions and, if  
112 needed, transformed to avoid heterogeneity of variances. The number of days required  
113 for *B. cranaodes* adults to emerge and the number of adults obtained under different  
114 photoperiods, different temperatures, and different generations were compared using  
115 Fisher's test. Significance level was set to 0.05.

116

## 116 **Results**

### 117 *Effects of daylength on B. cranaodes development*

118 The development time from first instar larvae to eclosion of adult was very similar  
119 under the long- and short-day photoperiods. This shows that exposure to a short  
120 daylength did not induce *B. cranaodes* to enter diapause. The mean development time in  
121 these experiments ranged from 52 to 59 days, which compares to a development time of  
122 53.4 days in the continuous lab-rearing under a LD 14 : 10 h photoperiod ( $n = 12$ ).  
123 There was also no difference between the number of adults emerging under the two  
124 photoperiods (Table 1).

125 Observations of mating and oviposition behavior under long and short photoperiod  
126 did not indicate a difference between the treatments. Matings occurred within the first  
127 hour after onset of the dark period, and female oviposition behaviour was the same,  
128 under both photoperiods.

129 The most important mortality factor was migration of larvae out of the diet boxes.  
130 More larvae escaped during the second generation (Table 1).

### 131 *Effects of temperature on B. cranaodes development*

132 Larval development time from hatching until adult depended on temperature, but a  
133 similar number of adults emerged for both temperatures. There was no difference  
134 between the number of adults emerged for both generations and both temperatures ( $P <$   
135  $0.02$ ) (Table 2). However, it was needed about 43 days to obtain the first adult at 21 -  
136  $22^{\circ}\text{C}$ , and 160 days at 10 -  $13^{\circ}\text{C}$  ( $P < 0.02$ ). The results showed that low temperature did  
137 not induce *B. cranaodes* to enter diapause.



138 As during the previous experiment, the most important mortality factor was  
139 migration of larvae out of the rearing recipients.

#### 140 *Field trapping tests*

141 Captures in pheromone traps show that *B. cranaodes* adults were present in the apple  
142 orchard all year around, even during the winter (Fig. 1). Rather high captures of *B.*  
143 *cranaodes* were recorded during the end of the peak growing season from February to  
144 April, when multiple insecticide sprays were applied to control *B. cranaodes* and *G.*  
145 *molesta* infestations.

146 The control level recommended for *B. cranaodes* is when weekly pheromone trap  
147 captures surpass 30 males/trap. However, in fall and winter, the grower sprayed  
148 insecticide when detected any increase on the adult population (June, July, and August)  
149 (Fig. 1). From September on started the frequent insecticide use due to the occurrence of  
150 *B. cranaodes*, *G. molesta*, and *Anastrepha fraterculus* (Diptera: Tephritidae).

151 This field test also showed that 10 µg lures baited with the optimized 4-component  
152 pheromone remained attractive over six months.

#### 153 **Discussion**

154 According to our findings, short daylength and low temperature do not induce  
155 diapause in Brazilian apple leafroller *B. cranaodes*.

156 Diapause is the basic means by which insects and related arthropods in temperate  
157 zones cope with unfavorable environmental conditions (Tauber *et al.*, 1986). Diapause  
158 induction, maintenance, termination, and postdiapause development and growth are  
159 mainly regulated by abiotic factors such as photoperiod, temperature, and moisture.  
160 Several studies have illustrated the influence of photoperiod and temperature on  
161 diapause maintenance and termination (Boyne *et al.*, 1985; Ishirara & Shimada 1995).

162 Photoperiod is the major diapause-inducing environmental stimulus in most species. So  
163 far, it has been shown in a few species only that diapause induction is mediated by  
164 temperature (Tauber *et al.*, 1986; Danks, 1987). Photoperiod has been shown to effect  
165 the growth rate in other lepidopteran species, with larval growth being slower under  
166 shorter photoperiods (Danilevskii *et al.*, 1970; Goettel & Philogène, 1978). Beck (1980)  
167 suggests that these growth responses are correlated with the photoperiodic effect of  
168 diapause induction. In *B. cranaodes*, duration of larval development was the same for  
169 long and short-day conditions (Table 1).

170 Although the ecology of insect diapause has been extensively studied in insects, most  
171 of the available data concerns insects from temperate climate zones, where insects are  
172 subject to marked seasonal changes in photoperiod, temperature and availability of food  
173 resources. Diapause is usually induced by decreasing day length (Chippendale &  
174 Reddy, 1973; Goettel & Philogène, 1978). The situation is quite different in the Tropics,  
175 since there are only minor seasonal changes in daylength (Tanzubil *et al.*, 2000). Under  
176 such conditions, the key environmental factors influencing diapause are rainfall,  
177 temperature and food in conjunction with photoperiod (Adkisson *et al.*, 1963; Scheltes,  
178 1978; Denlinger, 1986; Kfir, 1993). In many insect species from temperate climate  
179 zones, larval exposure to low temperatures is not necessary for diapause development.  
180 However, low temperatures that might have occurred during the larval development  
181 could have an impact on diapause development. Many of the photoperiodic responses  
182 are also temperature-dependent, with temperature affecting circadian entrainment,  
183 photoperiodic summation and aspects of general physiology involved in diapause  
184 induction (Veerman & Vaz Nunes, 1980). This was observed for example for the  
185 tortricidae species *Adoxophyes orana*, *Choristoneura fumiferana*, and *Endopiza viteana*  
186 (Han & Bauce, 1996; Tobin *et al.*, 2002; Milonas & Savopoulou-Soultani, 2004) and

187 for the noctuidae specie *Sesamia nonagrinoidea* (Fantinou *et al.*, 2003). For *B.*  
188 *cranaodes*, the interaction between short day and low temperature did not lead to  
189 diapause (Table 2). Under these conditions, *B. cranaodes* larvae slowed down the  
190 growth and development. It may be that the low temperature provides a shorter period  
191 suitable for feeding, which in turn reduces metabolic functions and retards the larval  
192 development.

193 Our field tests corroborate the results of the laboratory tests and confirm that *B.*  
194 *cranaodes* does not diapause. The adults were present all year around, despite the lower  
195 temperature and shorter day regime during winter. This highlights the potential of  
196 pheromone-based methods for control of *B. cranaodes* during off-season. Population  
197 densities are lowest during off-season and attempts should then be made to further  
198 reduce population densities before onset of the new apple growing period. Therefore,  
199 the use of mating disruption method for *B. cranaodes* is under development in Brazil.

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207 **References**

- 208 Adkisson, P.L., Bell, R.A. & Wellso, S.G. (1963) Environmental factors controlling the  
209 induction of diapause in the pink bollworm, *Pectinophora gossypiella*. *Journal of*  
210 *Insect Physiology*, **9**, 299-230.
- 211 Arn, H., Rauscher, S. & Schmid, A. (1979) Sex attractant formulations and traps for the  
212 grape moth *Eupoecilia ambiguella* Hb. *Mitteilungen der Schweizerischen*  
213 *Entomologischen Gesellschaft*. **52**, 49-55.
- 214 Beck, S.D. (1980) *Insect photoperiodism*. 2<sup>nd</sup> edition. Academic Press, New York.
- 215 Boyne, J.V., Rock, G.C. & Nelson, L.A. (1985) Diapause in *Platynota idaeusalis*  
216 (Lepidoptera: Tortricidae): effects of temperature, photoperiod, and time of  
217 inoculation in the field on diapause termination. *Environmental Entomology*, **14**,  
218 790-796.
- 219 Chippendale, G.M. & Reddy, A.S. (1973) Temperature and photoperiod regulation of  
220 diapause of the southwestern corn borer *Diatraea grandiosella*. *Journal of Insect*  
221 *Physiology*, **19**, 1397-1408.
- 222 Coracini, M.D.A., Bengtsson, M., Reckziegel, A., Löfqvist, J., Francke, W., Vilela,  
223 E.F., Eiras, A.E., Kovaleski, A. & Witzgall, P. (2001) Identification of a four-  
224 component sex pheromone blend in *Bonagota cranaodes* (Lepidoptera: Tortricidae).  
225 *Journal of Economic Entomology*, **94**, 911-914.
- 226 Coracini M.D.A., Bengtsson, M., Reckziegel, A., Eiras, A.E., Vilela, E.F., Anderson,  
227 P., Francke, W., Löfqvist, J. & Witzgall, P. (2003) Behavioral effects of minor sex  
228 pheromone components in Brazilian apple leafroller *Bonagota cranoades*  
229 (Lepidoptera: Tortricidae). *Journal of Applied Entomology*, **127**, 1-8.

- 230 Danilevskii, A.S., Goryshin, N.I. & Tyshchenko, V.P. (1970) Biological rhythms in  
231 terrestrial arthropods. *Annual Review of Entomology*, **15**, 201-244.
- 232 Danks, H.V. (1987) Insect dormancy: an ecological perspective. *Biological Survey of*  
233 *Canada*, Ottawa.
- 234 Denlinger, D.L. (1986) Dormancy in tropical insects. *Annual Review of Entomology*, **31**,  
235 239-264.
- 236 Fantinou, A.A., Kourti, A.T. & Saitanis, C.J. (2003) Photoperiodic and temperature  
237 effects on the intensity of larval diapause in *Sesamia nonagrinoidea*. *Physiological*  
238 *Entomology*, **28**, 82-87.
- 239 Goettel, M.S. & Philogène, B.J.R. (1978) Effects of photoperiod and temperature on the  
240 development of a univoltine population of the banded woollybear, *Pyrrharctia (Isia)*  
241 *isabella*. *Journal of Insect Physiology*, **24**, 523-527.
- 242 Han, E-N. & Bause, E. (1996) Diapause development of spruce budworm larvae,  
243 *Choristoneura fumiferana* (Clem.) (Lepidoptera: Tortricidae), at temperatures  
244 favouring post-diapause development. *The Canadian Entomologist*, **128**, 167-169.
- 245 Ishirara, M. & Shimada, M. (1995) Photoperiodic induction of larval diapause and  
246 temperature-dependent termination in a wild multivoltine bruchid, *Kytorhinus*  
247 *sharpianus*. *Entomologia Experimentalis et Applicata*, **75**, 127-134.
- 248 Kfir, R. 1993. Diapause termination in *Chilo partellus* in the laboratory. *Annals of*  
249 *Applied Biology*, **123**, 1-7.
- 250 Lorenzato, D. (1984) Ensaio laboratorial de controle da "traça da maçã" *Phtheochroa*  
251 *cranaodes* Meyrick, 1937 com *Bacillus thuringiensis* Berliner e inseticidas químicos.  
252 *Agronomia Sulriograndense*, **20**, 157-163.

- 253 Mani, E., Riggenbach, W. & Mendik, M. (1978) Zucht des Apfelwicklers (*Laspeyresia*  
254 *pomonella*) (L.) auf künstlichem Nährboden, 1968-78. *Mitteilungen der*  
255 *Schweizerischen Entomologischen Gesellschaft*, **51**, 315-326.
- 256 Milonas, G.P. & Savopoulou-Soultani, M. (2004) Diapause termination in  
257 overwintering larvae of a Greek strain of *Adoxophyes orana* (Lepidoptera:  
258 Tortricidae). *Environmental Entomology*, **33**, 513-519.
- 259 Scheltes, P. (1978) The condition of the host plant during aestivation-diapause of the  
260 stalk borers *Chilo partellus* and *Chilo orichalcociliella* in Kenya. *Entomologia*  
261 *Experimentalis et Applicata*, **24**, 479-488.
- 262 Tanzubil, P.B., McCaffery, A.R. & Mensah, G.W.K. (2000) Diapause termination in the  
263 millet stem borer, *Coniesa ignefusalis* (Lepidoptera: Pyralidae) in Ghana as affected  
264 by photoperiod and moisture. *Bulletin of Entomological Research*, **90**, 89-95.
- 265 Tauber, M.J., Tauber, C.A. & Masaki, S. (1986) *Seasonal adaptations of insects*.  
266 Oxford University Press, New York.
- 267 Tobin, P.C., Nagarkatti, S. & Saunders, M.C. (2002) Diapause maintenance and  
268 termination in grape berry moth (Lepidoptera: Tortricidae). *Environmental*  
269 *Entomology*, **31**, 708-713.
- 270 Veerman, A. & Vaz Nunes, M. (1980) Circadian rhythmicity participates in the  
271 photoperiodic determination of diapause in spider mites. *Nature*, **287**, 140-141.  
272

**Table 1.** Development of Brazilian apple leafroller *B. cranaodes* larvae under two different photoperiods.

Treatm.	Generation	Photoperiod (L/D)	N° larvae used <sup>1</sup>	N° dead insects	N° adults emerged	Development time (days) <sup>2</sup>
dark	1 <sup>st</sup>	7/17 h	500	33	326a	51.9a
dark	1 <sup>st</sup>	7/17 h	500	52	325a	52.3a
dark	2 <sup>nd</sup>	7 /17 h	500	51	249a	58.9a
light	2 <sup>nd</sup>	10/14 h	500	43	228a	51.8a

<sup>1</sup> All treatments began with recém-emerged larvae.

<sup>2</sup> Mean value for growth period from larvae to adult.

Within the same column and same generation, numbers followed by the same letter are not significantly different (Fisher test,  $P > 0.05$ ).

**Table 2.** Development of Brazilian apple leafroller *B. cranaodes* larvae under two different temperatures.

Temp. (°C)	Generation	Photoperiod (L/D)	N° larvae used <sup>1</sup>	N° dead insects	N° adults emerged	Development time (days) <sup>2</sup>
10-13	1 <sup>st</sup>	7/17 h	500	45	237a	167.1a
10-13	2 <sup>nd</sup>	7/17 h	500	47	241a	155.6a
21-22	1 <sup>st</sup>	7 /17 h	500	34	273a	45.3a
21-22	2 <sup>nd</sup>	7 /17 h	500	41	257a	42.8a

<sup>1</sup> All treatments began with recém-emerged larvae.

<sup>2</sup> Mean value for growth period from larvae to adult.

Within the same column and same temperature, numbers followed by the same letter are not significantly different from each other (Fisher test,  $P > 0.05$ ).



**Fig. 1.** Weekly mean air temperature and trap catch of Brazilian apple leafroller *B. cranaodes* males in pheromone traps at Schio Orchard, Vacaria-RS, Brazil, from January to December 2004.

