**[JOURNAL NAME HERE] | www.rsc.org/[JOURNAL] Communication**

# **Inhibition of Dimethylarginine Dimethylamino Hydrolase (DDAH) and Arginine Deiminase (ADI) by Pentafluorophenyl (PFP) sulfonates**

 $\mathbf{P}$ atrick Vallance, $^{*b}$  Hannah D. Bush,  $^a$  B. James, Mok,  $^a$  Ramon Hurtado-Guerrero,  $^b$  Herpreet Gill,  $^b$ Sharon Rossiter,  $^b$  Jonathan D. Wilden,  $^a$  Stephen Caddick,  $^{a*}$ 

<sup>5</sup>*Receipt/Acceptance Data* **[DO NOT ALTER/DELETE THIS TEXT]** *Publication data* **[DO NOT ALTER/DELETE THIS TEXT] DOI: 10.1039/b000000x [DO NOT ALTER/DELETE THIS TEXT]** 

10 **shown to have significant inhibitory activity against the bacterial A range of pentafluorophenyl (PFP) sulfonate esters derived from the reaction of PFP vinyl sulfonate and various nitrones are enzymes DDAH and ADI.**

Nitric oxide (NO) is an important mediator of intracellular signaling and has attracted interest as a target for therapeutic intervention, as it is widely acknowledged that there are a variety

- $15$  of disease states for which NO is implicated.<sup>1</sup> One of the most significant problems associated with the design of inhibitors, is to target the pathological excess NO production without disrupting essential NO-mediated processes, often by seeking selectivity for a particular NOS isoform. One method for potentially
- 20 circumventing these problems is the indirect modulation of NO levels by inhibition of the enzyme DDAH which is responsible for controlling levels of  $N^G$ -methyl-L-arginine (MMA) and  $N^G$ ,  $N<sup>G</sup>$ , dimethyl-L-arginine (ADMA) which are endogeneous inhibitors of NOS. $<sup>2</sup>$ </sup>
- 25 30 conditions. 5a Inhibition of bacterial  $DDAH<sup>4</sup>$  is also of interest as it offers opportunities for the development of new anti-bacterial agents. The structurally related enzyme arginine deiminase (ADI) is also a possible antibacterial/antiprotozoal target, as various pathogenic organisms utilize ADI to generate ATP under anaerobic

35 (Cys) (Figure 1). Both enzymes are known to catalyze the Recently high-resolution structures of a bacterial  $DDAH<sup>6</sup>$  and ADI<sup>5</sup> have been disclosed and it has been shown in both enzymes that the active site comprises a catalytic triad containing an acidic residue (Glu / Asp), a basic residue (His) and a cysteine residue conversion of the substrate(s) MMA and ADMA to citrulline as shown in figure 1.



40

**Figure 1***:* Conversion of ADMA ( $R = Me$ ) and / or MMA ( $R = H$ ) to citrulline are catalysed by DDAH and ADI.

45 the possible development of covalent inhibitors of DDAH and In a recent communication Knipp *et al.* described the cysteine modification of DDAH by HcyNO and proposed this as a lead for  $ADI<sup>7</sup>$  In that work it was proposed that it should be possible to rationally design covalent inhibitors of DDAH based on those

50 ADI. Whilst the development of small molecule inhibitors of 55 the use of pentafluorophenyl (PFP) sulfonates as an findings. That work has prompted us to disclose our own studies, which identify novel small molecule inhibitors of DDAH and both DDAH and ADI is appealing, it is notable that there is only one known inhibitor of bacterial DDAH, which has modest affinity and is an arginine homologue, ${}^{8}$  and there are no known inhibitors of ADI. Herein we disclose our preliminary studies on unprecedented new class of enzyme inhibitors. The biological activity of the PFP-sulfonate group is completely unexplored and is highlighted here by the development of inhibitors of DDAH and ADI

60 As had previously been noted by one of us in the disclosure of the crystal structure of DDAH, the active site resembles that of a cysteine protease with a catalytic triad (Fig 1). $5$ 

65 speculate that it may be possible to generate inhibitors of DDAH 70 potential inhibitor. In order to test this speculative hypothesis we The work of Roush *et al.* on the use of sulfonates and sulfonamides as inhibitors of cysteine proteases,<sup>9</sup> stimulated us to and ADI based on sulfonates and sulfonamides or closely related structures. This would offer an opportunity to develop molecular scaffolds, which would be markedly different in their structure to arginine mimetics, which may be a more obvious class of decided to evaluate a diverse collection of heterocyclic PFPsulfonates as potential inhibitors of DDAH and ADI.

 $75$  sulfonate with nitrones (Scheme 1).<sup>10</sup> Our previously disclosed synthetic approach to such species was based on the 1,3-dipolar cycloaddition reaction of a PFP-



Scheme 1. Synthetic approach to heterocyclic PFP-sulfonates via 1,3dipolar cycloaddition

An initial screen of a variety of PFP-sulfonates and related so structures at relatively high concentrations (500 µm, data not shown) provided some encouraging inhibition of both pseudomonas enzymes DDAH and ADI. At 50 µm a smaller selection of PFP sulfonates retained significant activity and their molecular structures are shown in Figure 2.<br>As can be seen from Table 1 the majority of these compounds

85 <sup>†</sup> Electronic Supplementary Information (ESI) available: [details of any <sub>90</sub> and it can be seen that compounds 2, 3, 4, 6 and 9 exhibit were found to have activity against both DDAH and ADI at 50 µm concentration. These preliminary data indicate that there is greater inhibition of DDAH compared with ADI.  $IC_{50}$  values were determined for a small selection of the most active species

This journal © Royal Society of Chemistry **and Contact Contact** 

supplementary information available should be included here]. See http://dx.doi.org/10.1039/b000000x/

significant activity against DDAH and compounds **3** and **6** also show significant activity against ADI.



**Figure 2:** Molecular structures of PFP sulfonates exhibiting significant 95 inhibition of PaDDAH and PaADI

These data suggest that there is potential for considerable optimization to give a new series of inhibitors based on the PFPisoxazolidine structural motif.

100 substrate) **Table 1**. Activity of PFP-sulfonates against PaDDAH and PaADI (1mM



In order to assess the nature of the inhibition we carried out experiments on reversibility and time-dependence (Figure 3).



**Figure 3.** (a) Effect of increasing substrate concentration on inhibition of PaDDAH by compound **6**, 75µM. (b) Time-dependence of PaDDAH inhibition by compound **3**, 10µM.

115 120 exhibited Inhibition by compound **6** can be at least partially reversed by addition of increased amounts of substrate (Figure 3a), suggesting competitive inhibition. The large excess of substrate required to reverse inhibition may indicate that the inhibitor is more tightly bound than the substrate. In a separate experiment, compound **3** a constant level of DDAH inhibition over an 80-minute period (Figure 3b). Covalent

125 inhibitors would show a time-dependent increase in inhibition as the enzyme becomes progressively irreversibly bound, therefore the evidence suggests that our inhibitors are not acting by a covalent mechanism. This is consistent with the observed reversibility of inhibition.<sup>11</sup>

130 Irrespective of the detailed mechanism underlying the inhibition, 135 study has also demonstrated that the PFP-sulfonate motif may 140 In summary we have described new inhibitors of the enzymes DDAH and ADI. From these experiments it would appear that these PFP-sulfonates may be reversible inhibitors of DDAH. this is the first time that non-substrate-like inhibitors for DDAH have been identified and these are the most potent inhibitors of bacterial DDAH currently known. Moreover these results identify the first small molecules to inhibit the enzyme ADI. The present play an important role in future studies directed toward identification of small molecule enzyme inhibitors and / or ligands for proteins. The simplicity with which diverse arrays of PFP-derivatives can be prepared may facilitate further smallmolecule discovery activities. The further development of this work to identify details of the molecular interaction of these PFPsulfonate derivatives with DDAH and ADI are underway.

### **Acknowledgements**

145 EPSRC, BBSRC and the British Heart Foundation (BHF). We are The authors gratefully acknowledge financial support from grateful to EPSRC Mass Spectrometry Service in Swansea for provision of mass spectral facilities. Further support of our programme from GSK, CEM Microwave Technology (UK), Novartis, AstraZeneca is gratefully acknowledged.

### 150 Stephen Caddick, \*<sup>*a*</sup> Hannah D. Bush, <sup>*a*</sup> B. James, Mok, <sup>*a*</sup> Ramon **Hurtado-Guerrero,** *<sup>b</sup>*  **Herpreet Gill,** *<sup>b</sup>*  **Sharon Rossiter,** *<sup>b</sup>*  **Jonathan D. Wilden,** *<sup>a</sup>*  **Patrick Vallance,\****<sup>b</sup>*

*a Department of Chemistry, University College London, 20, Gordon Street, London, WC1H 0AJ, UK. E-mail: s.caddick@ucl.ac.uk. <sup>b</sup>*

155  *Department of Medicine, The Rayne Building, 5 University Street London WC1E 6JJ, UK. E-mail: patrick.vallance@ucl.ac.uk.*

#### **Notes and references**

- 160 1. (a) J. M. Fukuto; G. Chaudhuri *Annu. Rev. Pharmacol. Toxicol.* **1995**, *35*, 165-194. (b) S. Moncada; A. Higgs *FASEB J.* **1995**, *9*, 1319-1330. (c) A. J. Hobbs; A. Higgs; S. Moncada *Annu. Rev. Pharmacol. Toxicol.* **1999**, *39*, 191-220.
	- 2. (a) P. Vallance; J. M. Leiper *Nat. Rev. Drug Discov.* **2002**, *1*, 939- 950.
- 165 3. T. Ogawa; M. Kimoto; K. Sasaoka *J. Biol. Chem.* **1989**, *264*, 10205- 10209.
	- 4. J. Santa Maria; P. Vallance; I. G. Charles; J. M. Leiper *Mol. Microbiol.* **1999**, *33*, 1278-1279.
- 170 5. (a) A. Galkin; L. Kulakova; E. Sarikaya; K. Lim; A. Howard; O. Herzberg *J. Biol. Chem.* **2004**, *279*, 14001-14008. (b) K. Das; G. H. Butler; V. Kwiatkowski; A. D. Clark Jr.; P. Yadav; E. Arnold *Structure* **2004**, *12*, 657-667.
- 6. J. Murray-Rust; J. M. Leiper; M. McAlister; J. Phelan; S. Tilley; J. Santa Maria; P. Vallance; N. McDonald *Nat. Struct. Biol.* **2001**, *8*, 679-683.
- 175 7. 7. M. Knipp; O. Braun; M. Vašák *J. Am. Chem. Soc.* **2005**, *127*, 2372- 2373.
- 180 8. R. J. MacAllister; H. Parry; M. Kimoto; T. Ogawa; R. J. Russell; H. Hodson; G. St. J. Whitley, G; P. Vallance B*rit. J. Pharmacol.* **1996**, *119*, 1533-1540. S. Rossiter, C. L. Smith; M. Malaki; M. Nandi; H. Gill; J. M. Leiper; P. Vallance, D. L. Selwood *J. Med. Chem*. **2005**, *48*, 4670-4678
- 9. (a). W. R. Roush; S. L. Gwaltney; J. M. Cheng; K. A. Scheidt; J. H. McKerrow; E. Hansell *J. Am. Chem. Soc* **1998**, *120*, 10994-10995. (b) W. R. Roush; J. M. Cheng; B. Knapp-Red *Bioorg. Med. Chem.*  <sup>185</sup>*Lett.* **2001**, *11*, 2759-2762. (c) B. R. Shenai; A. Alvarez-Hernandez,

## **[JOURNAL NAME HERE] | www.rsc.org/[JOURNAL] Communication**

P. Y. Chong; C. D. Emal; R. J. Neitz; W. R. Roush; P. J. Rosenthal *Antimicrob. Agents Chemother.* **2003**, *47*, 154-160.

- 10. S. Caddick; H. D. Bush *Org. Lett.* **2003**, *5*, 2489-2492.
- 11. J. D. Winkler; C. M. Sung; M. Chabot-Fletcher; D. E. Griswold, D; L. A. Marshall; F. H. Chilton; W. Bondinell; R. J. Mayer *Mol. Pharmacol* **1998**, *53*, 322-329. 190