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Identifying New Antiepileptic Drugs Through Genomics-Based Drug Repurposing

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Abstract

Currently available antiepileptic drugs (AEDs) fail to control seizures in 30% of patients.

Genomics-based drug repurposing (GBR) offers the potential of savings in the time and cost of developing new AEDs. In the current study, we used published data and software to identify the transcriptomic signature of chronic temporal lobe epilepsy and the drugs that reverse it. After filtering out compounds based on exclusion criteria, such as toxicity, 36 drugs were retained. 11 of the 36 drugs identified (>30%) have published evidence of antiepileptic efficacy (for example, curcumin) or antiepileptogenic affect (for example, atorvastatin) in recognised rodent models or patients. By objectively annotating all ~20,000 compounds in the LINCS database as either having published evidence of antiepileptic efficacy or lacking such evidence, we demonstrated that our set of repurposable drugs is ~6-fold more enriched with drugs having published evidence of antiepileptic efficacy in animal models than expected by chance (p-value <0.006). Further, we showed that another of our GBR-identified drugs, the commonly-used well-tolerated antihyperglycemic sitagliptin, produces a dose-dependent reduction in seizures in a mouse model of pharmacoresistant epilepsy. In conclusion, GBR successfully identifies compounds with antiepileptic efficacy in animal models and, hence, it is an appealing methodology for the discovery of potential AEDs.

Introduction

Epilepsy is amongst the most common neurological disorders: 65 million people have epilepsy worldwide (1). Currently available AEDs (AEDs) have been identified using traditional drug-discovery approaches (2): serendipity, phenotypic screening, hypothesis-driven target-orientated design, and structural modification of existing molecules. These compounds act primarily upon a small number of targets involved mainly in modulating neuronal excitability (2). Currently available AEDs have a number of significant shortcomings. They fail to control seizures in 30% of patients (3), and possess no antiepileptogenic or disease-modifying activity (4, 5). In addition, they do not exhibit any meaningful efficacy against significant epilepsy comorbidities (6), such as cognitive impairment and psychiatric conditions. Over 100 years of utilising traditional drug discovery approaches for the development of an increasingly long list of approved AEDs, now approaching 30, has failed to overcome these challenges. A need exists, therefore, for newer more effective AED-discovery approaches.

In recent years, there has been an increasing interest in exploiting genomics-based drug repurposing (GBR) to aid and accelerate the drug discovery process. GBR is based on the following precepts (7). Every biological state and, hence, every disease state can be described by a gene expression signature. Treatments that restore gene expression patterns to their norm are associated with the successful restoration of the disease phenotype (8). The methodology can be summarised as follows (9). The first step is to generate a signature of differential gene expression for the disease through a genome-wide gene expression analysis comparing normal tissue with the disease tissue of interest. Then, using the gene expression signature of disease, databases of drug-induced gene expression signatures, such as Connectivity Map (cMap) and Library of

Integrated Network-based Cellular Signatures (LINCS) are queried. These databases contain drug-induced signatures of differential gene expression for thousands of compounds. Each signature is generated through genome-wide gene expression analysis comparing cells before and after drug exposure. If disease- and drug-induced signatures are sufficiently negatively correlated (i.e. the genes upregulated in the disease-induced signature are downregulated in the drug-induced signature and *vice versa*) then the effect of the drug on transcription is opposite to the effect of the disease. Hence, the drug might revert the disease signature of differential gene expression and the disease phenotype itself. GBR has successively produced a number of therapeutic leads for different diseases (8-11).

GBR holds much promise for drug discovery in epilepsy. It is known that hundreds of genes influence susceptibility to epilepsy (12) and are altered in the epileptic focus (13, 14). By targeting these dysregulated genes, and restoring global gene expression patterns to their healthy norm, repurposed drugs could ameliorate the diverse pathological changes that create and promote the hyperexcitable neuronal network which generates seizures. Hence, this approach offers the potential of efficacy against epilepsy, epileptogenesis, and the comorbidities that share genes, biological pathways and pathophysiological mechanisms with epilepsy.

Results

The transcriptomic signature of epilepsy and the drugs that reverse it

The transcriptomic signature of disease was extracted from a published RNA sequencing analysis of a mouse model of chronic temporal lobe epilepsy (TLE) (15). The genes constituting the transcriptomic signature of TLE used in the current study are listed in Table S1.

Drugs that reverse the epileptic transcriptomic signature were identified using the Library of Integrated Network-based Cellular Signatures (LINCS) data and software. 123 compounds (Table S2) met the inclusion criterion (LINCS mean connectivity score threshold of -85). After filtering out compounds (Table S2) based on the exclusion criteria of toxicity, parenteral route of administration, lack of animal or human dosage data, or BBB-impermeability, 36 compounds remained (Table S3). We found no explicit published evidence of BBB-impermeability for any of the drugs; most have never undergone an *in vitro* or *in vivo* assessment of BBB-permeability. The known primary mode of action of each of the 36 compounds is listed in Table S3.

Dissimilar drugs modulate the same biological pathways

We used the Drug-Set Enrichment Analysis (DSEA) tool to demonstrate that the identified compounds, although pharmacologically diverse, are alike in modulating biological pathways. DSEA showed that the unselected sample of five GBR-identified drugs with varying modes of action shared 134 (out of 669) Reactome pathways at a significance threshold of $p < 0.05$ (Table S4). The median number of significant pathways shared by 1000 randomly-selected five-compound drug-sets was 31, the mean was 33.2, and the range was 10-98 (Table S5 and Figure S1). Hence, none of the 1000 random drug-sets shared ≥ 134 pathways, demonstrating that the number of pathways shared by GBR-identified drugs is statistically significant (permutation-based p -value < 0.001).

Repurposable drugs are better than conventional AEDs at reversing the pathway dysregulation underlying epilepsy

The Reactome pathways most significantly dysregulated in TLE were identified by performing Gene-Set Enrichment Analysis (<http://www.broadinstitute.org/gsea/>) separately on the up- and down-regulated genes constituting the transcriptomic signature of TLE identified above. The hundred most up-regulated and the hundred most down-regulated pathways were noted; the degree of enrichment was statistically significant ($FDR < 0.05$) for all of these 200 pathways. 29 pathways were both up- and down-regulated; these pathways were excluded. It should be noted that in the Reactome database, pathways are assembled into a hierarchy of biological processes. The top hierarchy of biological processes represent very broad categories, such as ‘Developmental Biology’ and ‘Neuronal System’, within which subcategories can be differently dysregulated; this was the case with the 29 pathways both up- and down-regulated in the current analysis. Of the 142 (out of 200) pathways remaining, one was not present in the DSEA database (as the database excludes the very smallest and largest pathways) leaving 141 pathways to be considered in the present analysis.

We hypothesized that repurposable drugs are better than conventional AEDs at rectifying the dysregulated pathways identified above. In turn, each repurposable drug present in the in the DSEA database was compared with the set of conventional AEDs present in the DSEA database. Table S6 and Figure 1 show the percentage of dysregulated pathways corrected most effectively by each drug. In each comparison, the repurposable drug was the best performing compound for the highest proportion of pathways. The fraction of dysregulated pathways for which the repurposable drug surpassed conventional AEDs was statistically significant (binomial test $FDR < 0.05$) for all comparisons.

The set of identified compounds is enriched with AEDs

At the time of conducting this analysis, there were 19,767 unique compounds within the LINCS database. Using the strategy detailed in Methods of objective computational data retrieval and parsing, followed by careful manual filtering based on pre-defined exclusion criteria, 203 drugs (1%) were identified as possessing published evidence of antiepileptic efficacy in animal models.

Of the 123 drugs included in our transcriptionally-repurposed drug-set, seven (5.7%) possessed published evidence of antiepileptic efficacy, which represents an approximately six-fold enrichment.

LINCS database compounds were randomly divided into 159 sets of 123 drugs each. The median number of AEDs within the random drug-sets was 1, the mean was 1.2, and the range was 0-4. None of the random drug-sets included seven or more drugs with published evidence of antiepileptic efficacy, confirming that the enrichment of AEDs within the transcriptionally-repurposed drug-set is statistically significant, with a permutation-based p-value of <0.006 ($=1/159$ random drug-sets). This enrichment was also statistically significant according to the hypergeometric distribution ($p<0.0003$).

Further details about the data used for completing this enrichment analysis, including evidence used for considering or discounting drugs as antiepileptic, can be found in the supplementary material.

In addition to above unsupervised computational approach for parsing information from Medline abstracts, which was applied to all 123 drugs identified by the LINCS computational algorithm, a more detailed manual literature-review was performed in order to identify additional evidence of antiepileptic efficacy for the 36 compounds that were selected after application of exclusion criteria; results for this have been summarised in Table 1.

In vivo validation

Sitagliptin, one of the drugs identified using our GBR pipeline, was chosen for *in vivo* validation as it is a commonly-used well-tolerated licensed medication. Levetiracetam, an AED which is well characterised and effective in this model (16, 17), was included in the drug screening to verify adequate execution of the screening process. Sitagliptin produced a dose-dependent reduction in seizure scores (Figure S2) in the 6 Hz psychomotor seizure mouse model (18) of pharmacoresistant epilepsy (16, 19). The mean (\pm standard error) seizure scores for the vehicle, sitagliptin 125 mg/kg, sitagliptin 250 mg/kg and sitagliptin 500 mg/kg groups were 0.93 ± 0.12 , 0.73 ± 0.18 , 0.60 ± 0.14 , and 0.47 ± 0.13 , respectively. In keeping with published reports, the AED levetiracetam (16, 17) also produced a reduction in seizures scores. There was a statistically significant difference in seizure scores across the groups (Kruskal–Wallis p-value <0.05). The difference between control and treatment group seizure scores was statistically significant (two-sided Mann-Whitney U test FDR <0.05) for the highest dose of sitagliptin evaluated (500 mg/kg) and for levetiracetam. Individual seizure scores are shown in Table S9.

Discussion

In this study, we have applied GBR to epilepsy, demonstrated that this identifies a set of compounds significantly enriched with drugs possessing efficacy in animal models of epilepsy, and presented *in vivo* validation of a GBR-identified drug.

Our GBR strategy—identifying compounds using the LINCS algorithm and excluding those unsuited for oral administration—identified 36 drugs. Using DSEA, we showed that our sample of GBR-identified compounds exert a similar modulatory effect on 134 Reactome pathways; significantly greater than expected by chance (permutation-based p-value <0.001). This observation is useful for two reasons. Firstly, the GBR-identified compounds appear to have little in common with each other: they are pharmacologically diverse with different molecular targets. Dissimilar drugs with disparate modes of action would seem unlikely to induce the same antiepileptic phenotype. Our analysis shows that, in spite of their different modes of action, GBR-identified compounds modulate the same biological pathways, enabling them to induce the same therapeutic effect. Secondly, the DSEA algorithm is based on gene expression data from cMap, not LINCS. Hence, it is encouraging to see that an analysis using a paradigm and a database of drug-induced gene expression signatures different from that used to perform GBR corroborates the pathway-level functional similarity of our GBR-identified compounds.

It is interesting to note the lack of conventional licensed AEDs amongst our GBR-identified compounds, which suggests that the GBR-identified compounds are more effective than some conventional AEDs at reversing the transcriptomic signature of TLE. In keeping with this, we showed, using DSEA-based analysis, that each repurposable drug in our sample is better than all tested conventional AEDs in rectifying the highest number of dysregulated biological pathways

underlying TLE. Again, it is encouraging to see that an analysis using a paradigm and a database of drug-induced gene expression signatures different from that used to perform GBR corroborates the superiority of the tested GBR-identified compounds over the tested conventional AEDs at correcting the TLE transcriptomic signature. Whether this mechanistic difference translates into a more effective, broader or disease-modifying clinical affect should be investigated in future functional studies. However, our bioinformatic, literature-based and *in vivo* analysis hints at this possibility, as discussed below.

A number of the GBR-identified drugs have published evidence of antiepileptic or antiepileptogenic efficacy in animal models. Almost a third of the final 36 compounds identified have functional evidence of efficacy in animal models of epilepsy. We demonstrated that the GBR-identified drugs are significantly enriched (permutation-based p-value <0.006) with drugs having published evidence of antiepileptic efficacy in animal models by using a novel objective methodology, based upon the auto-parsing of relevant information from Medline abstracts. We demonstrated this enrichment within all 123 compounds identified by the LINCS algorithm, rather than to just the 36 compounds that remained after filtering, in order to avoid the possibility of perceived bias in the manual selection of the 36 compounds.

As mentioned above, one motivation for seeking transcriptome-rectifying therapies is the anticipation of antiepileptogenic efficacy. This expectation is supported our findings: almost half of the compounds with published data from animal models have evidence of an antiepileptogenic effect (Table 1). An example is the drug lestaurtinib. Using the rat hypoxia-induced epilepsy model, researchers showed that administration of lestaurtinib at the time of the epileptogenic

insult attenuates susceptibility to seizures later in life (20). Within our list also is the first drug shown to be potentially antiepileptogenic in man: atorvastatin (21). After studies demonstrating the antiepileptogenic effects of atorvastatin in rodent models (22, 23), Guo *et al.* have recently shown that administration of statins to patients with early-onset poststroke seizures is associated with reduced risk of poststroke epilepsy (24). In the study by Guo and colleagues, most patients received atorvastatin; there were not enough data to conduct appropriate subgroup analyses because too few patients were given simvastatin or rosuvastatin.

It is clear that epilepsy and its comorbidities, such as anxiety, depression and cognitive impairment, share genes, biological pathways and pathophysiological mechanisms (25-29). Hence, another motivation for seeking transcriptome-rectifying antiepileptic therapies is the anticipation of efficacy against epilepsy comorbidities. Again, this expectation is supported by our findings: a number of compounds with published data from animal models of epilepsy have evidence of an effect against comorbidities in the models. For example, atorvastatin ameliorates anxiety, depression and cognitive impairment (23, 30), resveratrol is effective against cognitive impairment and depression (31), and mepacrine ameliorates cognitive impairment (32) in rodent models of epilepsy.

We wished to show that, in addition to the compounds with published evidence of antiepileptic and antiepileptogenic efficacy found to be enriched within our drug-set, other GBR-identified compounds yet unstudied in animal models of epilepsy also have therapeutic efficacy. We demonstrated that the commonly-used well-tolerated licenced antihyperglycaemic agent sitagliptin produces a dose-dependent reduction in seizure scores in a mouse of model of

pharmacoresistant epilepsy—the 6 Hz psychomotor seizures model. Seizures in this model are resistant to many established AEDs, for example carbamazepine, ethosuximide, lamotrigine, phenytoin and topiramate (33), although they are sensitive to valproic acid, levetiracetam and phenobarbital. Sitagliptin at twice the equivalent of the maximum dose used in the current study has been administered to rodents for repeated doses without adverse effect (34, 35), and sitagliptin equal to the maximum dose used in the current study been taken in man without adverse effects (36), although it is higher than the dose currently licensed for diabetes. Sitagliptin, on its own, does not cause hypoglycaemia (37). Prolonged courses of sitagliptin at different doses should be analysed in future studies to determine if they produce greater antiepileptic efficacy. At the same time, it is important to note that sitagliptin screening is presented here only as validation of our GBR approach and as a proof-of-principle. It is possible that there are more efficacious AEDs within our GBR-identified drug-set; the other GBR-identified compounds should be screened in animal models of epilepsy in order to identify the ones that are most promising. In a recent study, intracerebroventricular injection of sitagliptin reduced the average electroencephalographic power spectrum induced by heperthermia; any affect on febrile or non-febrile seiures was not reported (38).

Limitations and need for future investigations should be noted. The transcriptomic signature of disease used in this study was extracted from a mouse model of chronic TLE. It is possible that some of the transcriptomic changes in this signature are species-specific. It should be noted, however, that there is significant conservation of genomic networks in hippocampal tissue from this mouse model and people with epilepsy (39). Transcriptomic profiles from studies comparing human epileptic and normal postmortem hippocampi are known to include postmortem-specific

changes (40), which would compromise the quality of the disease signature. Another potential limitation is that the efficacy of a drug in rodent models does not necessarily translate to effectiveness in man. At the same time, it is important to note that drug screening in rodent models of epilepsy has been successful and crucially contributed to the development of numerous clinically effective AEDs (41). Finally, some of the compounds identified in this analysis are promising leads, rather than repurposing candidates, as they are not currently in routine clinical use.

In future work, prolonged courses of sitagliptin and other GBR-identified compounds should be analysed in animal models of epilepsy in order to identify the drugs that are most promising. To confirm the antiepileptogenic or disease-modifying potential of the identified compounds, specifically designed studies that include a drug washout phase should be performed (41). With further supportive evidence from rodent models of epilepsy, drugs like sitagliptin and atorvastatin, for which there is extensive clinical experience of safe use in other conditions, would be promising repurposing candidates to take into clinical trials.

In conclusion, this is the first study to show that GBR in epilepsy identifies a coherent set of compounds significantly enriched with drugs possessing efficacy in animal models of epilepsy and, hence, GBR is an appealing methodology for the discovery of potential AEDs.

Materials and Methods

Experimental design

This study has three main parts. The first part is a bioinformatic analysis to identify, using the LINCS software and database of drug-induced transcriptomic signatures, compounds that revert the transcriptomic signature of epilepsy extracted from a published RNA-sequencing study of a rodent model of the disease. In the second part, the success of the first part of the study is assessed by determining, using a permutation-based strategy, if the identified drugs are enriched with compounds with published evidence of efficacy in rodent models of epilepsy. In the third part, we perform *in vivo* screening of one of the identified drugs, sitagliptin, in a rodent model of refractory epilepsy. The sample size for the *in vivo* study was dictated by resource limitations, powering the study to 0.7 with an α error of 0.05 for the highest dose of sitagliptin screened.

Identifying the transcriptomic signature of epilepsy and the drugs that reverse it

A transcriptomic signature of chronic temporal lobe epilepsy (TLE) was generated as follows. Differentially expressed genes were extracted from the results of a recently published high-throughput RNA sequencing analysis comparing hippocampal tissue from normal mice and from mice with pilocarpine-induced chronic epilepsy (15). The mouse genes were mapped to human homologues listed in the Mouse Genome Database (42). Human homologues were mapped to Affymetrix U133A microarray probe identifiers listed in the NetAffx database (43). Probes corresponding to the most differentially expressed mouse genes (FDR <0.05 & fold change ≥ 2) were retained.

Drugs that reverse the epileptic transcriptomic signature were identified using the Library of Integrated Network-based Cellular Signatures (LINCS) data and software (<http://apps.lincscloud.org/query>). LINCS is successor to the highly successful Connectivity Map

(44), with a vastly expanded number of drugs profiled in multiple human cell lines, producing more than a million transcriptomic drug signatures.

The LINCS software quantitates the (dis)similarity between the disease and each of the drug signatures. For each drug signature, the software generates a ‘connectivity score’ ranging from +100 to -100. A high positive connectivity score indicates that the corresponding compound’s transcriptomic signature is like that of the disease signature, while a high negative connectivity score indicates that the compound’s transcriptomic signature is opposite that of the disease. As each drug has been profiled in multiple human cells lines, there are multiple transcriptomic signatures for each drug, and multiple connectivity scores for each drug-disease combination. For each drug, the mean of the four highest connectivity scores across different cell lines is the default statistic for summarising the relationship of the drug-disease pair, in the LINCS algorithm. The cell lines utilised for calculating the mean connectivity score are not pre-selected by tissue of origin; any tissue-specific noise is thought to be drowned out by the powerful directionality of the induced signature (45). The mean connectivity score threshold for drug selection may be set at different points: a threshold score of -90 has been proposed and used in published literature (46). In the current study, a mean connectivity score threshold of -85 was chosen in order to generate a longer list of drugs for evaluation.

Drugs identified using the above process were then filtered based on the following exclusion criteria:

1. Drugs included in the National Institute for Occupational Safety and Health List of Antineoplastic and Other Hazardous Drugs (47), or known to be toxic, according to published literature
2. Drugs requiring parenteral administration
3. Drugs with no published data to indicate effective *in vivo* doses in humans or animals
4. Drugs reported to be BBB-impermeable in published literature

For the drugs that remained after the above filtering process, the primary known mode of action was identified through literature review.

Do dissimilar drugs modulate the same biological pathways?

The drugs identified above appeared to have little in common with each other: they were pharmacologically diverse with different molecular targets. Dissimilar drugs with disparate modes of action would appear unlikely to induce the same antiepileptic phenotype. We hypothesized that the identified drugs, in spite of their different modes of action, modulate the same biological pathways, enabling them to induce the same therapeutic effect. To test this hypothesis, Drug-Set Enrichment Analysis (DSEA), which is based upon concepts similar to those first introduced by Hegde and colleagues (48), was utilized. DSEA (49) is designed to search the transcriptional responses of different drugs for shared pathways whose genes are upregulated (or downregulated) by the drugs in the set. If drugs in a drug-set tend to modulate the same pathway more than the other drugs in the database, this pathway will be detected by DSEA. A set of drugs of interest is tested against a database of pathways. Each pathway in the database is stored as a ranked list of drugs, sorted from the one most up-regulating the pathway to the one most down-regulating it. Given a query-set of drugs, DSEA checks for each pathway

whether the drugs tend to be significantly ranked at the top (or the bottom) of the list, by applying a Kolmogorov-Smirnov (KS) test. A KS p-value can thus be computed for each pathway. P-values assess how much the ranks of the chosen drugs are consistently small (up-regulation) or large (down-regulation) for each pathway. The final output of DSEA is a list of pathways ranked by the p-value.

The current DSEA database is based upon the cMap 2.0 dataset containing transcriptional profiles for 1309 drugs. Of the compounds identified through our drug repurposing pipeline, five (camptothecin, mepacrine, quinidine, resveratrol and scriptaid) are found in the cMap 2.0 and, hence, DSEA database. This subset is an unselected sample of five different drugs with widely varying modes of action and, hence, a suitable collection with which to test our hypothesis that GBR-identified drugs, though seemingly dissimilar, modulate the same pathways.

The five drugs were analysed using the DSEA webserver (<http://dsea.tigem.it>). The number of significant Reactome pathways ($p < 0.05$) shared by these five drugs was counted. In order to determine if the number of pathways shared by the GBR-identified drugs is statistically significant, 1000 randomly-selected five-compound drug-sets were also analysed through the webserver, and the number of shared pathways was counted for each random set.

Are repurposable drugs better than conventional AEDs at reversing the pathway dysregulation underlying epilepsy?

We wished to determine if repurposable drugs were more effective than conventional AEDs at correcting the pathway dysregulation underlying TLE. The DSEA dataset was again used for this

analysis. As stated above, each pathway in the DSEA database is stored as a ranked list of drugs, sorted from the one most up-regulating the pathway to the one most down-regulating it. Hence, for any pathway up- or down-regulated in disease, the DSEA dataset can be utilized for identifying the compound most effective at reversing the dysregulation.

As mentioned above, of the compounds identified through our drug repurposing pipeline, five (camptothecin, mepacrine, quinidine, resveratrol and scriptaid) are found in the DSEA database. The following eight conventional AEDs are in the DSEA database: acetazolamide, carbamazepine, ethosuximide, gabapentin, primidone, topiramate, valproic acid and vigabatrin. This set of drugs is an unselected sample of different compounds with widely varying modes of action and, hence, a suitable collection with which to test our hypothesis that repurposable drugs are better than conventional AEDs at reversing the pathway dysregulation underlying TLE.

The Reactome pathways most significantly dysregulated in TLE were identified by performing Gene-Set Enrichment Analysis (<http://www.broadinstitute.org/gsea/>) separately on the up- and down-regulated genes constituting the transcriptomic signature of TLE identified above. The hundred most up-regulated and the hundred most down-regulated pathways were noted; the degree of enrichment was statistically significant ($FDR < 0.05$) for all of these 200 pathways

The following strategy was adopted:

1. in turn, compare each repurposable drug with the set of conventional AEDs,
2. for each dysregulated pathway, determine if the repurposable drug is better than conventional AEDs at rectifying the dysregulation,

3. determine if the repurposable drug outperforms conventional AEDs for more pathways than expected by chance alone.

The exact binomial test (R 3.2.4) was used to determine if the fraction of dysregulated pathways for which the repurposable drug surpasses conventional AEDs is statistically significant.

Is the set of identified compounds enriched with AEDs?

We hypothesized that the set of compounds identified through GBR is enriched with drugs of known antiepileptic efficacy. In order to test this hypothesis, the following strategy was devised:

1. Each of the compounds in the LINCS database was annotated as either having published evidence of antiepileptic efficacy or lacking published evidence of antiepileptic efficacy.
2. Based on the above, we determined the number of compounds with evidence of antiepileptic efficacy amongst our set of repurposable drugs.
3. To determine if compounds with antiepileptic efficacy are overrepresented in our set of repurposable drugs, the LINCS compounds were divided into random sets of the same size as the repurposed set, and the number of drugs with antiepileptic efficacy was determined for each random set.

For added robustness, we also used the hypergeometric distribution (R 3.3.2) to determine the statistical significance of the enrichment of compounds with antiepileptic efficacy within our set of repurposable drugs.

For the above analysis, we included all 123 drugs identified by the LINCS computational algorithm, rather than the 36 that remained after application of exclusion criteria, as the latter

manual filtering steps might inadvertently advantage our chosen drug set. Annotation of the LINCS compounds according to evidence of antiepileptic efficacy was not limited to licensed AEDs, but included all drugs with evidence of efficacy in recognised animal models of epilepsy and seizures. In order to ensure that this process is objective, an unsupervised computational approach was adopted for parsing information from Medline abstracts about antiepileptic efficacy of the LINCS compounds. As there are ~20,000 drugs in the LINCS database, a solely manual literature search would, in any case, be impracticable. A clear predefined search strategy and vocabulary of search terms was used for this data collection. Abstracts were downloaded from Medline and then automatically parsed (auto-parsed) using Unix command line text manipulation tools. Methodological details are provided in the supplementary material, including search terms and synonyms used for the Medline search and auto-parsing, and the Unix commands utilised. In order to maximise the relevance of the auto-parsed data, we sought sentence-level concurrence of search terms, which increases specificity and precision of data retrieval (50). A Medline abstract sentence was extracted if the following search terms concurred within it: the name of a compound from the LINCS database, suppression (or a synonym thereof), seizures (or a synonym thereof), and the name of a recognised rodent model of epilepsy or seizures.

The results of the auto-parsing were manually reviewed to exclude any irrelevant results. Exclusion was based on clear objective pre-defined criteria: drugs were excluded if the information parsed (1) did not clearly indicate evidence of independent clinical antiepileptic efficacy, or (2) indicated promotion, rather than inhibition, of seizures by the drug.

It is important to note that these data are not presented as a comprehensive list of all drugs with published evidence of antiepileptic efficacy, but rather as a tool for objectively testing the overrepresentation of such drugs within the repurposable drug-set.

In addition to above unsupervised computational approach for parsing information from Medline abstracts, which was applied to all 123 drugs identified by the LINCS computational algorithm, a more detailed manual literature-review was performed in order to identify additional evidence of antiepileptic efficacy for the 36 compounds that were selected after application of exclusion criteria.

In vivo validation

For *in vivo* validation, we wished to choose a drug that: (1) was previously unstudied in animal models of epilepsy, and (2) had Medicines and Healthcare products Regulatory Agency (MHRA) or European Medicines Agency (EMA) approval and extensive data of routine safe clinical use in humans, as such a drug would encounter the fewest research and regulatory barriers on the repurposing pathway and will be more readily adopted by the clinical community. After excluding drugs that already have independent published evidence of antiepileptic efficacy (Table 1), sitagliptin (an antihyperglycaemic) was the highest ranked MHRA/EMA-approved drug in routine clinical use in this country. Hence, sitagliptin was chosen for *in vivo* validation. The 6 Hz psychomotor seizure mouse model (18), a mouse model of pharmaco-resistant epilepsy (16, 19), was used for validating the efficacy of this drug. Sitagliptin was evaluated at 3 doses—125, 250 and 500 mg/kg—chosen because they exert a therapeutic effect without producing toxicity in rodent models of brain diseases (34, 51-53). Levetiracetam (100 mg/kg), an AED

which is well characterised and effective in this model (16, 17), was included in the drug screening to verify adequate execution of the screening process. Physiological saline was used as the vehicle control substance. Hence, five groups of mice were studied: sitagliptin 125 mg/kg, sitagliptin 250 mg/kg, sitagliptin 500 mg/kg, levetiracetam 100 mg/kg, and saline. There were fifteen male NMRI mice (Janvier Labs, France), 22 - 31 g body weight range, in each group. The animal experiments were performed by the Porsolt Research Laboratory (France).

Compounds were administered intraperitoneally (i.p.) 60 minutes before application of the current. Mice were administered a rectangular current (44 mA, rectangular pulse: 0.2 ms pulse width, 3 s duration, 6 Hz) via corneal electrodes connected to a constant current shock generator (Ugo Basile: type 7801). Seizure scores were recorded for each animal. Seizure scores were based on the occurrence and severity of forelimb clonus: absent (0), mild (1) or strong (2). Presence or absence of Straub tail was also recorded, but it was noted that Straub tail was observed in some animals in the absence of forelimb seizures and, hence, this feature was not used in the evaluation. Each test was performed blind.

Statistical analysis

For the bioinformatic analyses, permutation-based approaches used to determine statistical significance have been described in the relevant sections above. A two-sided exact binomial test (R 3.2.4) was used to determine if the fraction of dysregulated pathways for which the repurposable drug surpasses conventional AEDs is statistically significant. For the *in vivo* analysis, seizure scores across all groups were compared using the Kruskal–Wallis test; threshold of statistical significance was $p < 0.05$. For post hoc analysis, seizure scores for each of the drug

treated groups were compared with the vehicle control group using a two-sided Mann-Whitney U test, with Benjamini and Hochberg false discovery rate (FDR) correction for multiple testing; threshold of statistical significance was $FDR < 0.05$. All statistical calculations were performed using R 3.2.4. G*Power version 3.1 (54) was used to calculate the power of the two-tailed Mann-Whitney U test ($\alpha = 0.05$).

Supplementary Materials

Supplementary Methods

Supplementary Results

Supplementary tables:

Table S1. Genes constituting the transcriptomic signature of epilepsy used in the current study

Table S2. Compounds meeting the LINCS mean connectivity score inclusion threshold of -85

Table S3. Compounds retained after applying the exclusion criteria

Table S4. DSEA output for the set of five GBR-identified drugs

Table S5. Number of Reactome pathways shared by 1000 randomly-selected five-drugs sets at a significance threshold of $p < 0.05$

Table S6. The percentage of dysregulated pathways corrected most effectively by each drug in the comparison.

Table S7. Medline search terms used

Table S8. Search terms and synonyms used for auto-parsing

Table S9. 6 Hz psychomotor seizures model seizure scores

Supplementary data: Included drugs

Supplementary data: Excluded drugs

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Conflicts of interest

None.

References and Notes:

1. A. K. Ngugi, C. Bottomley, I. Kleinschmidt, J. W. Sander, C. R. Newton, Estimation of the burden of active and life-time epilepsy: a meta-analytic approach. *Epilepsia* **51**, 883 (May, 2010).
2. M. J. Brodie, Antiepileptic drug therapy the story so far. *Seizure : the journal of the British Epilepsy Association* **19**, 650 (Dec, 2010).
3. S. D. Shorvon, The epidemiology and treatment of chronic and refractory epilepsy. *Epilepsia* **37 Suppl 2**, S1 (1996).
4. W. Loscher, C. Brandt, Prevention or modification of epileptogenesis after brain insults: experimental approaches and translational research. *Pharmacological reviews* **62**, 668 (Dec, 2010).
5. A. Pitkanen *et al.*, Issues related to development of antiepileptogenic therapies. *Epilepsia* **54 Suppl 4**, 35 (Aug, 2013).
6. G. L. Holmes, J. L. Noebels, The Epilepsy Spectrum: Targeting Future Research Challenges. *Cold Spring Harbor perspectives in medicine* **6**, (2016).
7. F. Iorio, T. Rittman, H. Ge, M. Menden, J. Saez-Rodriguez, Transcriptional data: a new gateway to drug repositioning? *Drug discovery today* **18**, 350 (Apr, 2013).
8. A. Wagner *et al.*, Drugs that reverse disease transcriptomic signatures are more effective in a mouse model of dyslipidemia. *Molecular systems biology* **11**, 791 (Jan, 2015).
9. M. Sirota *et al.*, Discovery and preclinical validation of drug indications using compendia of public gene expression data. *Science translational medicine* **3**, 96ra77 (Aug 17, 2011).
10. J. T. Dudley *et al.*, Computational repositioning of the anticonvulsant topiramate for inflammatory bowel disease. *Science translational medicine* **3**, 96ra76 (Aug 17, 2011).
11. N. S. Jahchan *et al.*, A drug repositioning approach identifies tricyclic antidepressants as inhibitors of small cell lung cancer and other neuroendocrine tumors. *Cancer discovery* **3**, 1364 (Dec, 2013).
12. D. Speed *et al.*, Describing the genetic architecture of epilepsy through heritability analysis. *Brain : a journal of neurology* **137**, 2680 (Oct, 2014).

13. N. Mirza *et al.*, Identifying the biological pathways underlying human focal epilepsy: from complexity to coherence to centrality. *Human molecular genetics* **24**, 4306 (Aug 1, 2015).
14. N. Mirza, O. Vasieva, A. G. Marson, M. Pirmohamed, Exploring the genomic basis of pharmacoresistance in epilepsy: an integrative analysis of large-scale gene expression profiling studies on brain tissue from epilepsy surgery. *Human molecular genetics* **20**, 4381 (Nov 15, 2011).
15. K. F. Hansen, K. Sakamoto, C. Pelz, S. Impey, K. Obrietan, Profiling status epilepticus-induced changes in hippocampal RNA expression using high-throughput RNA sequencing. *Scientific reports* **4**, 6930 (2014).
16. M. E. Barton, B. D. Klein, H. H. Wolf, H. S. White, Pharmacological characterization of the 6 Hz psychomotor seizure model of partial epilepsy. *Epilepsy research* **47**, 217 (Dec, 2001).
17. N. M. Rowley, H. S. White, Comparative anticonvulsant efficacy in the corneal kindled mouse model of partial epilepsy: Correlation with other seizure and epilepsy models. *Epilepsy research* **92**, 163 (Dec, 2010).
18. W. C. Brown, D. O. Schiffman, E. A. Swinyard, L. S. Goodman, Comparative assay of an antiepileptic drugs by psychomotor seizure test and minimal electroshock threshold test. *The Journal of pharmacology and experimental therapeutics* **107**, 273 (Mar, 1953).
19. W. Loscher, Critical review of current animal models of seizures and epilepsy used in the discovery and development of new antiepileptic drugs. *Seizure : the journal of the British Epilepsy Association* **20**, 359 (Jun, 2011).
20. M. Obeid, E. C. Rosenberg, P. M. Klein, F. E. Jensen, Lestaurtinib (CEP-701) attenuates "second hit" kainic acid-induced seizures following early life hypoxic seizures. *Epilepsy research* **108**, 806 (May, 2014).
21. A. Siniscalchi, S. Mintzer, Statins for poststroke seizures: the first antiepileptogenic agent? *Neurology* **85**, 661 (Aug 25, 2015).
22. N. Sehar, N. B. Agarwal, D. Vohora, S. Raisuddin, Atorvastatin prevents development of kindling by modulating hippocampal levels of dopamine, glutamate, and GABA in mice. *Epilepsy & behavior : E&B* **42**, 48 (Jan, 2015).
23. R. Citraro *et al.*, Protective effects of some statins on epileptogenesis and depressive-like behavior in WAG/Rij rats, a genetic animal model of absence epilepsy. *Epilepsia* **55**, 1284 (Aug, 2014).
24. J. Guo *et al.*, Statin treatment reduces the risk of poststroke seizures. *Neurology* **85**, 701 (Aug 25, 2015).
25. L. Kandratavicius, J. E. Hallak, C. G. Carlotti, Jr., J. A. Assirati, Jr., J. P. Leite, Hippocampal expression of heat shock proteins in mesial temporal lobe epilepsy with psychiatric comorbidities and their relation to seizure outcome. *Epilepsia* **55**, 1834 (Nov, 2014).
26. C. E. Stafstrom, Epilepsy comorbidities: how can animal models help? *Advances in experimental medicine and biology* **813**, 273 (2014).
27. M. S. Hester, S. C. Danzer, Hippocampal granule cell pathology in epilepsy - a possible structural basis for comorbidities of epilepsy? *Epilepsy & behavior : E&B* **38**, 105 (Sep, 2014).
28. A. Gaitatzis, S. M. Sisodiya, J. W. Sander, The somatic comorbidity of epilepsy: a weighty but often unrecognized burden. *Epilepsia* **53**, 1282 (Aug, 2012).
29. A. Mazarati, R. Sankar, Common Mechanisms Underlying Epileptogenesis and the Comorbidities of Epilepsy. *Cold Spring Harbor perspectives in medicine* **6**, (2016).
30. G. Uzum, K. Akgun-Dar, U. Aksu, The effects of atorvastatin on memory deficit and seizure susceptibility in pentylenetetrazole-kindled rats. *Epilepsy & behavior : E&B* **19**, 284 (Nov, 2010).
31. K. M. Choudhary, A. Mishra, V. V. Poroikov, R. K. Goel, Ameliorative effect of Curcumin on seizure severity, depression like behavior, learning and memory deficit in post-pentylenetetrazole-kindled mice. *European journal of pharmacology* **704**, 33 (Mar 15, 2013).
32. M. Ahmad, G. M. Abu-Taweel, A. E. Aboshaiqah, J. S. Ajarem, The effects of quinacrine, proglumide, and pentoxifylline on seizure activity, cognitive deficit, and oxidative stress in rat lithium-pilocarpine model of status epilepticus. *Oxidative medicine and cellular longevity* **2014**, 630509 (2014).
33. M. Smith, K. S. Wilcox, H. S. White, Discovery of antiepileptic drugs. *Neurotherapeutics : the journal of the American Society for Experimental NeuroTherapeutics* **4**, 12 (Jan, 2007).
34. S. Lim *et al.*, Effect of a dipeptidyl peptidase-IV inhibitor, des-fluoro-sitagliptin, on neointimal formation after balloon injury in rats. *PloS one* **7**, e35007 (2012).
35. F. Briand, Q. Thieblemont, R. Burcelin, T. Sulpice, Sitagliptin promotes macrophage-to-faeces reverse cholesterol transport through reduced intestinal cholesterol absorption in obese insulin resistant CETP-apoB100 transgenic mice. *Diabetes, obesity & metabolism* **14**, 662 (Jul, 2012).
36. M. A. Darracq, J. M. Toy, T. Chen, C. Mo, F. L. Cantrell, A retrospective review of isolated gliptin-exposure cases reported to a state poison control system. *Clin Toxicol (Phila)* **52**, 226 (Mar, 2014).

37. P. Aschner *et al.*, Effect of the dipeptidyl peptidase-4 inhibitor sitagliptin as monotherapy on glycemic control in patients with type 2 diabetes. *Diabetes care* **29**, 2632 (Dec, 2006).
38. Z. Wang *et al.*, Transcriptome analysis of the hippocampus in novel rat model of febrile seizures. *PloS one* **9**, e95237 (2014).
39. M. R. Johnson *et al.*, Systems genetics identifies Sestrin 3 as a regulator of a proconvulsant gene network in human epileptic hippocampus. *Nature communications* **6**, 6031 (2015).
40. P. Roncon *et al.*, MicroRNA profiles in hippocampal granule cells and plasma of rats with pilocarpine-induced epilepsy--comparison with human epileptic samples. *Scientific reports* **5**, 14143 (2015).
41. W. Loscher, H. Klitgaard, R. E. Twyman, D. Schmidt, New avenues for anti-epileptic drug discovery and development. *Nature reviews. Drug discovery* **12**, 757 (Oct, 2013).
42. J. T. Eppig, J. A. Blake, C. J. Bult, J. A. Kadin, J. E. Richardson, The Mouse Genome Database (MGD): facilitating mouse as a model for human biology and disease. *Nucleic acids research* **43**, D726 (Jan, 2015).
43. G. Liu *et al.*, NetAffx: Affymetrix probesets and annotations. *Nucleic acids research* **31**, 82 (Jan 1, 2003).
44. J. Lamb *et al.*, The Connectivity Map: using gene-expression signatures to connect small molecules, genes, and disease. *Science* **313**, 1929 (Sep 29, 2006).
45. Y. A. Lussier, J. L. Chen, The emergence of genome-based drug repositioning. *Science translational medicine* **3**, 96ps35 (Aug 17, 2011).
46. J. C. Siavelis, M. M. Bourdakou, E. I. Athanasiadis, G. M. Spyrou, K. S. Nikita, Bioinformatics methods in drug repurposing for Alzheimer's disease. *Briefings in bioinformatics* **17**, 322 (Mar, 2016).
47. K. Traynor, NIOSH revamps hazardous drugs update. *American journal of health-system pharmacy : AJHP : official journal of the American Society of Health-System Pharmacists* **71**, 2099 (Dec 15, 2014).
48. R. N. Hegde *et al.*, Unravelling druggable signalling networks that control F508del-CFTR proteostasis. *Elife* **4**, (Dec 23, 2015).
49. F. Napolitano, F. Sirci, D. Carrella, D. di Bernardo, Drug-set enrichment analysis: a novel tool to investigate drug mode of action. *Bioinformatics* **32**, 235 (Jan 15, 2016).
50. M. S. Siadaty, J. Shu, W. A. Knaus, Relemed: sentence-level search engine with relevance score for the MEDLINE database of biomedical articles. *BMC medical informatics and decision making* **7**, 1 (2007).
51. A. E. El-Sahar, M. M. Safar, H. F. Zaki, A. S. Attia, A. A. Ain-Shoka, Sitagliptin attenuates transient cerebral ischemia/reperfusion injury in diabetic rats: implication of the oxidative-inflammatory-apoptotic pathway. *Life sciences* **126**, 81 (Apr 1, 2015).
52. H. D. Theiss *et al.*, Antidiabetic gliptins in combination with G-CSF enhances myocardial function and survival after acute myocardial infarction. *International journal of cardiology* **168**, 3359 (Oct 9, 2013).
53. C. Brenner *et al.*, Short-term inhibition of DPP-4 enhances endothelial regeneration after acute arterial injury via enhanced recruitment of circulating progenitor cells. *International journal of cardiology* **177**, 266 (Nov 15, 2014).
54. F. Faul, E. Erdfelder, A. G. Lang, A. Buchner, G*Power 3: a flexible statistical power analysis program for the social, behavioral, and biomedical sciences. *Behavior research methods* **39**, 175 (May, 2007).
55. L. A. Etherington *et al.*, Astrocytic adenosine kinase regulates basal synaptic adenosine levels and seizure activity but not activity-dependent adenosine release in the hippocampus. *Neuropharmacology* **56**, 429 (Feb, 2009).
56. G. Zhang, P. H. Franklin, T. F. Murray, Manipulation of endogenous adenosine in the rat prepiriform cortex modulates seizure susceptibility. *The Journal of pharmacology and experimental therapeutics* **264**, 1415 (Mar, 1993).
57. N. Gouder, L. Scheurer, J. M. Fritschy, D. Boison, Overexpression of adenosine kinase in epileptic hippocampus contributes to epileptogenesis. *The Journal of neuroscience : the official journal of the Society for Neuroscience* **24**, 692 (Jan 21, 2004).
58. B. G. Ugarkar *et al.*, Adenosine kinase inhibitors. 1. Synthesis, enzyme inhibition, and antiseizure activity of 5-iodotubercidin analogues. *Journal of medicinal chemistry* **43**, 2883 (Jul 27, 2000).
59. J. B. Wiesner *et al.*, Adenosine kinase inhibitors as a novel approach to anticonvulsant therapy. *The Journal of pharmacology and experimental therapeutics* **289**, 1669 (Jun, 1999).
60. R. M. Kaminski, H. Marini, P. I. Ortinski, S. Vicini, M. A. Rogawski, The pheromone androstenol (5 alpha-androst-16-en-3 alpha-ol) is a neurosteroid positive modulator of GABAA receptors. *The Journal of pharmacology and experimental therapeutics* **317**, 694 (May, 2006).
61. E. Russo *et al.*, Pharmacodynamic potentiation of antiepileptic drugs' effects by some HMG-CoA reductase inhibitors against audiogenic seizures in DBA/2 mice. *Pharmacological research : the official journal of the Italian Pharmacological Society* **70**, 1 (Apr, 2013).

62. H. Shafaroodi *et al.*, The involvement of nitric oxide in the anti-seizure effect of acute atorvastatin treatment in mice. *Neurological research* **34**, 847 (Nov, 2012).
63. J. K. Lee, J. S. Won, A. K. Singh, I. Singh, Statin inhibits kainic acid-induced seizure and associated inflammation and hippocampal cell death. *Neuroscience letters* **440**, 260 (Aug 8, 2008).
64. K. Moazzami, S. Emamzadeh-Fard, M. Shabani, Anticonvulsive effect of atorvastatin on pentylenetetrazole-induced seizures in mice: the role of nitric oxide pathway. *Fundamental & clinical pharmacology* **27**, 387 (Aug, 2013).
65. L. Moezi *et al.*, Chronic administration of atorvastatin induced anti-convulsant effects in mice: the role of nitric oxide. *Epilepsy & behavior : E&B* **23**, 399 (Apr, 2012).
66. V. R. Funck *et al.*, Differential effects of atorvastatin treatment and withdrawal on pentylenetetrazol-induced seizures. *Epilepsia* **52**, 2094 (Nov, 2011).
67. T. C. Piermartiri *et al.*, Atorvastatin prevents hippocampal cell death due to quinolinic acid-induced seizures in mice by increasing Akt phosphorylation and glutamate uptake. *Neurotoxicity research* **16**, 106 (Aug, 2009).
68. J. Song, L. Parker, L. Hormozi, M. A. Tanouye, DNA topoisomerase I inhibitors ameliorate seizure-like behaviors and paralysis in a Drosophila model of epilepsy. *Neuroscience* **156**, 722 (Oct 15, 2008).
69. A. M. Orellana-Paucar *et al.*, Anticonvulsant activity of bisabolene sesquiterpenoids of *Curcuma longa* in zebrafish and mouse seizure models. *Epilepsy & behavior : E&B* **24**, 14 (May, 2012).
70. P. Du *et al.*, Curcumin inhibits amygdaloid kindled seizures in rats. *Chinese medical journal* **122**, 1435 (Jun 20, 2009).
71. N. B. Agarwal *et al.*, Liposomal formulation of curcumin attenuates seizures in different experimental models of epilepsy in mice. *Fundamental & clinical pharmacology* **27**, 169 (Apr, 2013).
72. N. Bharal, K. Sahaya, S. Jain, P. K. Mediratta, K. K. Sharma, Curcumin has anticonvulsant activity on increasing current electroshock seizures in mice. *Phytotherapy research : PTR* **22**, 1660 (Dec, 2008).
73. A. Jyoti, P. Sethi, D. Sharma, Curcumin protects against electrobehavioral progression of seizures in the iron-induced experimental model of epileptogenesis. *Epilepsy & behavior : E&B* **14**, 300 (Feb, 2009).
74. Y. K. Gupta, S. Briyal, M. Sharma, Protective effect of curcumin against kainic acid induced seizures and oxidative stress in rats. *Indian journal of physiology and pharmacology* **53**, 39 (Jan-Mar, 2009).
75. E. Tamaddonfard, A. Erfanparast, N. Hamzeh-Gooshchi, S. Yousofizadeh, Effect of curcumin, the active constituent of turmeric, on penicillin-induced epileptiform activity in rats. *Avicenna journal of phytomedicine* **2**, 196 (Fall, 2012).
76. L. Saha, A. Chakrabarti, S. Kumari, A. Bhatia, D. Banerjee, Antiapoptotic and neuroprotective role of Curcumin in Pentylenetetrazole (PTZ) induced kindling model in rat. *Indian journal of experimental biology* **54**, 133 (Feb, 2016).
77. W. Zhu, J. Su, J. Liu, C. Jiang, The involvement of neuronal nitric oxide synthase in the anti-epileptic action of curcumin on pentylenetetrazol-kindled rats. *Bio-medical materials and engineering* **26 Suppl 1**, S841 (2015).
78. K. K. Akula, S. K. Kulkarni, Effect of curcumin against pentylenetetrazol-induced seizure threshold in mice: possible involvement of adenosine A1 receptors. *Phytotherapy research : PTR* **28**, 714 (May, 2014).
79. J. Mehla, K. H. Reeta, P. Gupta, Y. K. Gupta, Protective effect of curcumin against seizures and cognitive impairment in a pentylenetetrazole-kindled epileptic rat model. *Life sciences* **87**, 596 (Nov 20, 2010).
80. N. A. Noor, H. S. Aboul Ezz, A. R. Faraag, Y. A. Khadrawy, Evaluation of the antiepileptic effect of curcumin and *Nigella sativa* oil in the pilocarpine model of epilepsy in comparison with valproate. *Epilepsy & behavior : E&B* **24**, 199 (Jun, 2012).
81. H. S. Ezz, Y. A. Khadrawy, N. A. Noor, The neuroprotective effect of curcumin and *Nigella sativa* oil against oxidative stress in the pilocarpine model of epilepsy: a comparison with valproate. *Neurochemical research* **36**, 2195 (Nov, 2011).
82. M. Ahmad, Protective effects of curcumin against lithium-pilocarpine induced status epilepticus, cognitive dysfunction and oxidative stress in young rats. *Saudi journal of biological sciences* **20**, 155 (Apr, 2013).
83. N. B. Agarwal, S. Jain, N. K. Agarwal, P. K. Mediratta, K. K. Sharma, Modulation of pentylenetetrazole-induced kindling and oxidative stress by curcumin in mice. *Phytomedicine : international journal of phytotherapy and phytopharmacology* **18**, 756 (Jun 15, 2011).
84. Z. Jiang *et al.*, Protection against cognitive impairment and modification of epileptogenesis with curcumin in a post-status epilepticus model of temporal lobe epilepsy. *Neuroscience* **310**, 362 (Dec 3, 2015).

85. J. Auta *et al.*, Participation of mitochondrial diazepam binding inhibitor receptors in the anticonflict, antineophobic and anticonvulsant action of 2-aryl-3-indoleacetamide and imidazopyridine derivatives. *The Journal of pharmacology and experimental therapeutics* **265**, 649 (May, 1993).
86. E. Romeo *et al.*, 2-Aryl-3-indoleacetamides (FGIN-1): a new class of potent and specific ligands for the mitochondrial DBI receptor (MDR). *The Journal of pharmacology and experimental therapeutics* **262**, 971 (Sep, 1992).
87. M. Tsuda, T. Suzuki, M. Misawa, Subsensitivity to mitochondrial diazepam binding inhibitor receptor agonist FGIN-1-27-induced antiseizure effect in diazepam-withdrawn mice. *Life sciences* **62**, PL213 (1998).
88. M. Steriade, E. Stoica, On the anticonvulsive effect of quinidine. I. Experimental investigation of somatomotor, vegetative and bioelectrical aspects of convulsive seizures elicited by electroshocks. *Epilepsia* **1**, 264 (Apr, 1960).
89. M. Steriade, E. Stoica, On the anticonvulsive effect of quinidine. II. Experimental investigations of focal electrical after-discharge and penicillin epilepsy. *Epilepsia* **1**, 275 (Apr, 1960).
90. Y. K. Gupta, G. Chaudhary, K. Sinha, A. K. Srivastava, Protective effect of resveratrol against intracortical FeCl₃-induced model of posttraumatic seizures in rats. *Methods and findings in experimental and clinical pharmacology* **23**, 241 (Jun, 2001).
91. U. Hoda, N. B. Agarwal, D. Vohora, S. Parvez, S. Raisuddin, Resveratrol suppressed seizures by attenuating IL-1beta, IL1-Ra, IL-6, and TNF-alpha in the hippocampus and cortex of kindled mice. *Nutritional neuroscience*, 1 (Jun 2, 2016).
92. Y. K. Gupta, S. Briyal, G. Chaudhary, Protective effect of trans-resveratrol against kainic acid-induced seizures and oxidative stress in rats. *Pharmacology, biochemistry, and behavior* **71**, 245 (Jan-Feb, 2002).
93. Z. Wu *et al.*, Protective effect of resveratrol against kainate-induced temporal lobe epilepsy in rats. *Neurochemical research* **34**, 1393 (Aug, 2009).
94. Y. K. Gupta, G. Chaudhary, A. K. Srivastava, Protective effect of resveratrol against pentylenetetrazole-induced seizures and its modulation by an adenosinergic system. *Pharmacology* **65**, 170 (Jul, 2002).
95. L. Saha, A. Chakrabarti, Understanding the anti-kindling role and its mechanism of Resveratrol in Pentylenetetrazole induced-kindling in a rat model. *Pharmacology, biochemistry, and behavior* **120**, 57 (May, 2014).

Legends to figures:

Fig. 1. Percentage of dysregulated pathways corrected most effectively by each drug in the comparison. Asterisk indicates that the fraction of dysregulated pathways for which the repurposable drug surpassed conventional AEDs was statistically significant (binomial test FDR<0.05)

Tables:

Table 1. Drugs, from amongst the 36 GBR-identified compounds, that have evidence of antiepileptic or antiepileptogenic efficacy in animal models or humans. Please note that in the study by Guo and colleagues (24) most patients received atorvastatin; there were not enough

data to conduct appropriate subgroup analyses because too few patients were given simvastatin or rosuvastatin.

Drug	Mechanism of action	Summarised evidence of antiepileptic and antiepileptogenic efficacy
5-iodotubercidin	Adenosine kinase inhibitor	Suppression of epileptiform activity evoked by brief, high-frequency stimulation in rat hippocampal slices (55). Antiepileptic effect in rodent bicuculline (56), kainic acid (57), maximal electric shock (58, 59) models.
Androstenol	Neurosteroid, GABA _A receptor modulator	Antiepileptic effect in the rodent 6 Hz electroshock and pentylenetetrazol (60) models
Atorvastatin	HMG-CoA reductase inhibitor	Antiepileptic effect in the rodent DBA/2 audiogenic seizures (61), increasing current electroshock (22, 62), kainic acid (63), pentylenetetrazole (30, 62, 64-66), and quinolinic acid (67) models. Antiepileptogenic effect in the rodent pentylenetetrazole (22) and WAG/Rij (23) models. Association with reduced risk of poststroke epilepsy in patients (24).
Camptothecin	Inhibitor of DNA topoisomerase I	Antiepileptic effect in a <i>Drosophila</i> model of epilepsy (68)
Curcumin	Antioxidant	Antiepileptic effect in the zebrafish model (69), and in rodent amygdaloid kindling (70), increasing current

		electroshock (71, 72), iron-induced epileptogenesis (73), kainic acid (74), penicillin-induced epileptiform activity (75), pentylenetetrazole (31, 71, 76-79), and pilocarpine (80, 81) models. Anticonvulsant in pilocarpine- (82) and pentylenetetrazol-induced status epilepticus (71). Protects against pentylenetetrazole-induced kindling (83), and exerts a favourable disease-modifying effect in the kainic acid model (84).
FGIN-1-27	Mitochondrial benzodiazepine receptor agonist	Antiepileptic effect in the rodent isoniazid (85, 86), metrazol (85) and pentylenetetrazole (87) model.
Lestaurtinib	Tyrosine kinase inhibitor	Antiepileptogenic effect in rats: administration of lestaurtinib within 12 hours of the first neonatal seizure attenuates increased susceptibility to seizures in later life (20).
Mepacrine	Multiple actions, including inhibition of PLA ₂	Antiepileptic effect in the rodent lithium-pilocarpine (32) model.
Quinidine	Sodium channel blocker	Antiepileptic effect in electroshock (88) and penicillin-induced (89) seizures in cats.
Resveratrol	Produced naturally by	Antiepileptic effect in the rodent FeCl ₃ (90), increasing current electroshock (91), kainic acid (92, 93), and

	several plants; principal mode of action unknown	pentylentetrazole (94) models. Antiepileptogenic effect in the rodent pentylentetrazole model (91, 95).
Sitagliptin	Dipeptidyl peptidase-4 inhibitor	Antiepileptic effect: sitagliptin produces a dose-dependent reduction in seizures in a mouse-model of drug-resistant epilepsy (current study)