

UV-B exposure, ROS and stress: inseparable companions or loosely linked associates?

Éva Hideg¹, Marcel A.K. Jansen² and Åke Strid³

¹Institute of Biology, University of Pécs, Ifjuság u. 6. H-7624 Pécs, Hungary

²School of Biological, Earth and Environmental Sciences, University College Cork, North Mall, Cork, Ireland

³School of Science & Technology, Örebro Life Science Center, Örebro University, SE-70182 Örebro, Sweden

All authors contributed equally to this paper

Corresponding author: Jansen, M.A.K. (M.Jansen@ucc.ie).

1 **Ultraviolet-B (UV-B) radiation has long been perceived as a stressor. However, a**
2 **conceptual U-turn has taken place, and UV-B damage is now considered rare. We**
3 **question whether UV-stress and UV-B-induced reactive oxygen species (ROS) are still**
4 **relevant concepts, and if ROS-mediated signaling contributes to UV-B acclimation.**
5 **Measurements of antioxidants and of antioxidant genes show that both low and high**
6 **UV-B doses alter ROS metabolism. Yet, there is no evidence that ROS control gene**
7 **expression under low UV-B. Instead, expression of antioxidant genes is linked to the**
8 **UV RESISTANCE LOCUS 8 pathway. We hypothesize that low UV-B doses cause**
9 **'eustress' (good stress) and that stimuli-specific signaling pathways pre-dispose**
10 **plants to a state of low alert that includes activation of antioxidant defenses.**

11

12

13

14

15 **Keywords:** UV-B, stress, ROS, antioxidant, acclimation, signaling

16

1 **Evaluating consequences of UV-B exposure**

2 In the late 1980s, awareness of stratospheric ozone layer depletion triggered concerns
3 about the potentially harmful effects of increased ultraviolet-B (UV-B) radiation. Many
4 studies have since shown that UV-B causes damage to DNA, proteins and membranes,
5 impedes photosynthetic activities, and impedes plant growth. Oxidative stress has been
6 flagged as a key factor in such UV-B stress (e.g. [1]). Oxidative pressure [i.e. imbalances
7 between the production of reactive oxygen species (ROS) and anti-oxidant scavenging
8 capacity], has been linked to non-specific damage to DNA, proteins and lipids [2,3].
9 However, ROS, DNA damage and membrane degradation products also play a role in
10 mediating UV-B protection. ROS and antioxidants orchestrate stress defense responses by
11 adjusting gene expression, proteolysis, and thioredoxin dynamics [2,4]. Such ROS-mediated
12 signaling is a tightly regulated process that links actual stress conditions with stress
13 acclimation [5].

14 Notwithstanding the damaging potential of UV-B photons, it has become
15 increasingly clear that under realistic UV-B exposure conditions (see glossary), UV-B does
16 not substantially impede plant growth [6,7], and that 'the balance of current research
17 suggests that UV-damage is probably the exception rather than the rule' [8]. Indeed, in a
18 recent large scale study of the responses of perennial ryegrass (*Lolium perenne*) no
19 significant effect of ambient UV-B on aboveground biomass was discernable along a
20 latitudinal gradient (27-68°N) across Europe [9]. However, lack of stress does not mean a
21 lack of biological impact. On the contrary, there is overwhelming evidence that UV-B is an
22 environmental regulator, controlling gene expression, cellular and metabolic activities, and
23 growth and development [10]. Regulatory UV-B effects can be observed under low UV-B

1 fluences [11] and it has been proposed that such low UV-B effects are, at least partially,
2 mediated by the UV-B-specific UV RESISTANCE LOCUS 8 (UVR8) photoreceptor and
3 signaling pathway [12–16].

4 The lack of UV-B-mediated stress observed in many studies [6] has triggered debate
5 about the relationships between UV-B exposure, ROS and plant stress (Figure 1). In this
6 Review, we question whether UV-B-induced ROS and UV-dependent stress are still relevant
7 concepts, or if they are artifacts of particularly harsh UV exposure conditions. We examine
8 the role played by generic ROS signaling under low UV-B conditions, particularly in
9 comparison with the stimuli-specific UVR8 response pathway. Our analysis shows that low
10 UV-B doses induce considerable alterations in antioxidant status, but that there is no direct
11 evidence that these changes are mediated by ROS.

12 **Is UV-B radiation a stressor?**

13 To address the question of whether UV-B radiation is a stressor, it is necessary to define
14 stress [17]. The term ‘plant stress’ is commonly used by authors in a very broad sense,
15 whereby almost every environmentally induced change in metabolic activity, growth, and
16 developmental pattern can be referred to as stress or stress response [18]. ‘Plant stress’
17 can refer to destructive or constructive effects on plants, or for example a selecting factor
18 driving adaptive evolution. In order to differentiate between these various aspects of
19 stress, a general plant stress concept with unifying terminology has been developed [18-
20 21]. This concept is based on analogy with the field of mechanics where a material can be
21 exposed to a ‘stress’ (a force) which results in a ‘strain’ (bending). In plant sciences, the
22 terms of ‘stress factor’ or ‘stressor’ are used to describe this imposed, external factor.
23 Exposure of plants to a stressor can cause reversible, elastic eustress (strain or bending in

1 mechanics) and, once exposure exceeds a tolerance-limit, irreversible plastic distress (in
2 mechanics: a strain resulting in rupturing) [17,20]. Eustress is an activating, stimulating
3 stress which is a positive element in plant development, and is also referred to as 'good
4 stress' or "constructive stress" [18-21]. When a plant experiences a mild, elastic eustress,
5 metabolism is adjusted, and the plant acclimates to the new environment. For example, a
6 mild water deficit, above the permanent wilting point, can induce plant hardening and
7 increased water-use efficiency [20]. In contrast, distress is a severe stress that has a
8 predominantly negative effect on the plant and its development, and is also referred to as
9 "destructive stress" [18-21]. Distress occurs if the environment becomes too unfavorable
10 for a particular plant [22]. For example, a severe water deficit below the permanent wilting
11 point will cause severe cellular damage, and impede growth [20]. The onset of distress
12 does, however, not always occur under the same stressor exposure conditions, as plants
13 can increase elastic and plastic stress resistance through genetic adaptation and/or
14 physiological acclimation. The plant stress concept generates the terminology to dissect
15 plant stress responses, and this makes the concept particularly suitable to describe plant
16 responses to environmental factors that cause a mixture of eu- and distress, such as for
17 example UV-B radiation, low and high temperatures, wind and/or touch, and drought.

18 UV-B radiation has been amply demonstrated to induce specific changes in gene
19 expression [23–28], increased accumulation of UV-screening pigments [29] and altered
20 phytochemical content [30]. Many of these responses have been linked to increased UV-B
21 tolerance, and can be induced by below ambient, chronic UV-doses which do not cause
22 substantial damage [6,8,26]. These responses can therefore be defined as eustress.
23 However, whereas productivity may not be directly affected by UV-radiation under

1 eustress conditions, regulatory changes in photosynthate allocation and morphology [31],
2 may still cause subtle decreases in biomass accumulation [6]. In contrast, macroscopic
3 damage, accumulation of damaged DNA and inactivation of the photosynthetic machinery
4 are consistent with distress. The balance between eustress and distress does not simply
5 depend on UV-dose and/or the spectral quality, but will also depend on, for example,
6 background intensity of photosynthetically active radiation (PAR), plant acclimation state
7 and genotype. Many early UV-B studies showed extensive distress [32,33], and this was
8 typically associated with unrealistic experimental conditions, including high levels of UV-B
9 and/or low levels of accompanying PAR. A review of the UV-exposure protocols used in
10 these early studies concluded that there was little evidence to support a general
11 impediment of photosynthesis by ambient UV-B [34]. This conclusion has been widely
12 accepted, and is a key message of the 2011 United Nations Environment Programme
13 assessment, which reported the minimal effects of realistic UV-B on biomass accumulation
14 [6].

15 **UV-B radiation as a stressor under unfavorable environmental conditions**

16 Realistic field-based studies have shown that ambient UV-B can decrease photosynthetic
17 activity under certain circumstances. For example, in the harsh Arctic environment,
18 ambient levels of UV-B decrease photosynthetic performance of Arctic willow (*Salix*
19 *arctica*) [35]. Several studies have demonstrated that other environmental factors can also
20 influence the effect of UV-B on plants, which may explain the inconclusive results of many
21 field studies. For example, water supply has been shown to influence the effect of
22 supplemental ($1.2 \text{ kJ m}^{-2} \text{ d}^{-1}$ UV above ambient) UV-B on the growth and photosynthetic
23 electron flow of several Arctic bryophytes [36]. A study of photosynthetic soil organisms

1 (cyanobacteria, lichens and mosses) under desert conditions showed that the effects of UV-
2 B radiation were influenced by precipitation: for example, UV-B stress increased when the
3 precipitation frequency was increased [37]. Similarly, the sensitivity of clover (*Trifolium*
4 *repens*) exposed to $13.3 \text{ kJ m}^{-2} \text{ d}^{-1}$ UV-B has been shown to depend on both water
5 availability and genotype [38]. However, not all studies show a link between water supply
6 and UV-susceptibility. For example, UV-B ($24 \text{ kJ m}^{-2} \text{ d}^{-1}$) had no impact on photosynthesis
7 in drought-stressed, green-house-grown olive (*Olea europea*), rosemary (*Rosmarinus*
8 *officinalis*), and lavender (*Lavandula stoechas*) [39]. Nutrient supply has also been shown to
9 influence the effect of UV-B. For example, ambient UV-B (~ 9 or $\sim 15 \text{ kJ m}^{-2} \text{ d}^{-1}$) decreased
10 the photosynthetic activities of maize (*Zea mays*) that received low levels of nutrients, but
11 did not affect well-fertilized plants [40]. UV-B ($7.2 \text{ kJ m}^{-2} \text{ day}^{-1}$ UV above ambient)
12 decreased the photosynthetic rates of radish (*Raphanus sativus*) grown on super-optimal
13 nutrient levels, but not that of plants grown under optimal conditions [41]. Thus, plants
14 that are exposed to unfavorable environmental conditions appear to be more susceptible to
15 UV-mediated distress.

16 It is overly simplistic to conclude that any plant exposed to a stressor will be
17 susceptible to UV-mediated distress. On the contrary, the literature contains numerous
18 examples of cross-tolerances between UV-B and other environmental stressors. For
19 example, the severity of drought stress has been shown to decrease when pea (*Pisum*
20 *sativum*) [42] or tobacco (*Nicotiana tabacum*, Petit Havanna SR1) [43] were grown under
21 supplemental UV-B (32 and $\sim 13.2 \text{ kJ m}^{-2} \text{ d}^{-1}$, respectively). Similarly, UV radiation
22 diminishes drought stress in Stone pine (*Pinus pinea*) during the hot, dry Mediterranean
23 summer [44]. In tobacco, increased drought tolerance is associated with the induction of

1 antioxidant defenses [43]. Furthermore, in cucumber (*Cucumis sativus*), antioxidant
2 defenses are synergistically upregulated by a combination of drought and UV-B [45]. Thus,
3 exposure to multiple stressors can either result in aggravated distress or in increased
4 cross-tolerance; the factors that determine the direction of this interaction have
5 considerable ecological and agronomical relevance.

6 **ROS in UV-B-exposed plants**

7 Generally, UV-B has no significant effects on photosynthesis, and just subtle effects on plant
8 growth and development [6], implying that widespread, oxidative damage is rare under
9 realistic UV-B levels. This does not necessarily mean that ROS formation and metabolism
10 are unimportant. It is plausible that ROS play a role in eustress (i.e. UV-B acclimation and
11 the readjustment of metabolism). ROS-mediated signaling is a complex process affected by
12 individual ROS species, ROS-producing enzymes, and the oxidation–reduction states of
13 various antioxidants [4]. The concept of a cellular redox state has been envisaged as the
14 sum of all reducing and oxidizing redox active molecules in the cell; it is not just a control
15 point for stress responses, but also plays a far broader regulatory role in cellular regulation
16 [22].

17 In UV-B-exposed plants, increased levels of ROS may be formed as a result of
18 disruption of metabolic activities [1,46] or owing to increased activity of membrane-
19 localized NADPH-oxidase [47]. Visualization of production and fate of UV-induced ROS,
20 under *in vivo* conditions, contributes to our understanding of the role of these species.
21 However, this is technically not straightforward because of the reactivity of ROS. Target
22 identification may appear easier, particularly in the case of high ROS concentrations.
23 However, cascades of secondary oxidations can hide the identity of the primary ROS target

1 and, therefore, obscure mechanistic aspects of ROS activity [48]. Tools have been
2 developed to visualize ROS directly or indirectly, ranging from ROS-specific reporter
3 molecules to rather indirect indicators of ROS involvement, such as fingerprinting methods,
4 and are overviewed below. Unfortunately, plant scientists cannot use the full range of ROS-
5 visualizing tools that are successfully used in the medical or physical sciences. For example,
6 inhibition of ROS production by excluding oxygen is not an option for plant physiologists.
7 Similarly, direct identification of H₂O₂ based on its UV absorption is hampered by the
8 abundance of UV-absorbing molecules in plants.

9 **Direct ROS measurements**

10 Owing to its physical characteristics, singlet oxygen (¹O₂) is the only ROS that can be
11 detected without the use of a reporter. The monomolar infrared (1270 nm) photoemission
12 of ¹O₂ has been used to demonstrate the presence of this ROS in illuminated, isolated
13 reaction centers of photosystem II [49]. So far, singlet oxygen has not been detected in
14 intact leaves by this method. Singlet oxygen as well as other ROS can be visualized using
15 colorimetric, electron paramagnetic resonance (EPR) or fluorescent ROS reporter
16 molecules. Externally supplied reporter molecules compete with natural ROS targets and
17 undergo a discernible physical change, such as a change in color, fluorescence or EPR
18 absorption upon oxidation [50]. The presence of ¹O₂ and superoxide radicals has been
19 demonstrated in spinach (*Spinacia oleracea*) leaves using selective fluorescent probes, but
20 only in response to high, damaging UV doses [51]. Similarly, ROS have been detected in
21 broad bean (*Vicia faba*) leaves [46] and isolated rice (*Oryza sativa*) thylakoids [1] treated
22 with high intensity UV-B by using EPR spin trapping reporters. Thus, there is direct
23 evidence for increased ROS production under conditions typically associated with distress.

1 **Antioxidants and oxidized targets**

2 Oxidized, endogenous target molecules can also be used as ROS reporter molecules. For
3 example, accumulation malondialdehyde (MDA) [43,52] or of DNA thymine dimers [53],
4 products of ROS-mediated oxidation of polyunsaturated membrane lipids and of DNA,
5 respectively, imply the presence of ROS. MDA has been reported in the leaves of rice
6 cultivars treated with UV-B ($13 \text{ kJ m}^{-2} \text{ day}^{-1}$) [54]. Absence of MDA in plants exposed to
7 low UV-B doses may imply lack of oxidative stress. However, this is not necessarily the case
8 given that MDA may undergo secondary reactions and/or catabolism [55].

9 Because of the balance between pro-oxidants and antioxidants, changes in the
10 oxidation–reduction state of antioxidants provide a further tool for deducing changes in
11 ROS concentrations. A short period of exposure to 0.46 kJ m^{-2} UV causes a fourfold increase
12 in the level of oxidized dehydroascorbate radical in broad bean (*Vicia faba*) leaves,
13 reflecting UV-induced oxidative pressure [56]. However, changes in the redox state of the
14 ascorbate–dehydroascorbate redox pair cannot simply be equated to oxidative pressure
15 because of concomitant re-reduction reactions by glutathione and, ultimately, NADP(H). In
16 pea, acute exposure to 1.4 W m^{-2} UV-B has been shown to result in the ratio of reduced
17 glutathione to oxidized glutathione (GSH:GSSG) decreasing to just 6-10% of control values
18 [57], again indicating UV-induced oxidative pressure. Furthermore, it is not just the
19 Halliwell–Asada antioxidant system that needs to be considered, any molecule with radical
20 scavenging capacity can provide information about ROS [58]. Plants contain large numbers
21 of non-enzymatic antioxidants, including phenolics, carotenoids, cytochromes, tocopherols
22 and tocotrienols, polyamines and proteins that carry redox active S-groups, creating a
23 dynamic network of redox interactions [22]. Using the oxidation–reduction state of

1 extracted antioxidants to evaluate ROS involvement in UV-B responses is an indirect tool,
2 but this is still an attractive choice owing to the sensitivity of the method.

3 When plants are exposed to low, chronic UV-B conditions, another effect of UV-B
4 exposure becomes clear: pool sizes of antioxidants such as ascorbate, GSH, xanthophylls
5 and α -tocopherol are increased (compare with [21]), indicating greater anti-oxidative
6 defenses. For example, exposure of spinach to low, chronic UV-B (2 weeks exposure to 1 kJ
7 $\text{m}^{-2} \text{day}^{-1}$) resulted in a 2.7-fold increase in ascorbate levels [59], whereas α -tocopherol
8 levels increased about eightfold in spinach and lettuce (*Lactuca sativa*) that were exposed
9 to UV-B for one week [60]. Exposure to 1.4 W m^{-2} UV-B resulted in a 4.5-fold increase in
10 total GSH levels in pea [57]. It has been argued that the functional role of the well-
11 documented UV-B-mediated accumulation of phenylpropanoids and flavonoids is primarily
12 to increase ROS scavenging activity [29,61]. Flavonoid accumulation occurs under both low
13 and high UV-B conditions. In particular, the UV-induced increase in the
14 quercetin:kaempferol-ratio [62] represents an increase in ROS scavenging activity, rather
15 than an increase in UV absorbance. Thus, there is considerable evidence for changes in
16 antioxidant metabolism under conditions of both distress and eustress.

17 **Activation of antioxidant pathways**

18 A common strategy for studying ROS metabolism is to quantify the activity of the enzyme
19 components of the antioxidant system as proxies for oxidative pressure [63,64]. Measured
20 enzymes typically include Cu- or Zn-superoxide dismutases (SODs), ascorbate peroxidase,
21 dehydroascorbate reductase, glutathione peroxidase, glutathione reductase and catalase,
22 and their activities are mostly measured following exposure to high doses of UV-B.
23 However, interpretation of data is complicated owing to differences in antioxidant

1 responses between species, between genotypes of the same species [65–67] and between
2 leaves of different age, and/or developmental stage [52,68]. Nevertheless, there is some
3 consensus. Elevated SOD, catalase, glutathione reductase and glutathione peroxidase
4 activities were found in many UV-B exposure studies (compare with [69]). In winter wheat
5 (*Triticum aestivum*), the antioxidant system was up-regulated by UV-B (4.2 or 10.3 kJ m⁻² d⁻¹)
6 under optimal temperatures; however, under low (10°C during daytime and 5°C at night)
7 temperatures, UV-B decreased photosynthetic yield [70], which again emphasizes that
8 distress is most likely to occur when plants are exposed to multiple unfavorable factors.
9 UV-B (0.18 W m⁻²) also induced the production of the pyridoxine biosynthesis enzyme
10 PDX1, and increased the levels of the antioxidant pyridoxine in *Arabidopsis* (*Arabidopsis*
11 *thaliana*) [71]. However, despite the publication of numerous papers on UV-B-induced
12 antioxidant pathways, there is still considerable uncertainty regarding to what extent
13 enzyme components of antioxidant pathways are up-regulated under eustress conditions.

14 **UV-B-dependent expression of oxidative defense genes**

15 The problem with the aforementioned biochemical approaches is that they are either
16 relatively insensitive (reporter molecules), or indirect (changes in oxidation state,
17 reduction state or the total pool size of antioxidants). Molecular approaches can potentially
18 avoid some of these pitfalls by yielding information on expression of antioxidant pathways.
19 Nine *Arabidopsis* DNA array studies on UV acclimation performed by five different
20 laboratories have been published in journals or are searchable in Genevestigator
21 (<https://genevestigator.com/gv/>) [23–28,72–75]. These studies used a range of daily UV-B
22 doses (from 0.093 to 7.0 W m⁻²), spectra, durations of UV-B exposure (from 15 minutes to
23 12 days) and PAR background levels (from low 25 μmol m⁻² s⁻¹ to ambient glass house

1 conditions that include UV-A). In a study using particularly low levels of UV-B (0.093–0.137
2 W m⁻²), expression of glutathione reductase and the pyridoxine biosynthetic protein
3 PDX1.3 were found to increase. Glutathione reductase reduced glutathione with the help of
4 NADPH and is therefore a key component of the ascorbate–glutathione antioxidant system
5 [23]. Glutathione peroxidase, and several glutathione transferases and glutaredoxins were
6 shown to be upregulated following exposure to short periods of relatively high intensity
7 UV-B [24,25]. Glutaredoxin expression was decreased in plants exposed to chronic (12 day;
8 0.564 kJ m⁻² day⁻¹) UV-B, possibly reflecting a down-regulation following an initial up-
9 regulation of expression [26]. Thus, there is considerable evidence for altered expression of
10 glutathione-related genes across a range of UV doses and exposure times, complementing
11 measurements of altered GSH:GSSG ratios and pool size [57], and implying that alterations
12 in ROS metabolism are a feature of all UV-B exposure conditions.

13 PDX gene products are strong antioxidants that neutralize singlet oxygen, hydroxyl
14 radicals, and superoxide [71,75,76]. The *PDX1.3* gene is up-regulated following exposure to
15 short periods of low- [23] or high-intensity UV-B [24,25]. However, *PDX1.3* has not been
16 found to be differentially expressed in plants exposed to chronic (12 day) UV-B, suggesting
17 that PDX antioxidant activities are components of the fast, initial response to UV-B.

18 Numerous genes encoding enzymes involved in phenol metabolism such as flavonol
19 synthase, caffeoyl-CoA *O*-methyltransferase, and 4-coumarate-CoA ligase 3 are upregulated
20 in *Arabidopsis* following exposure to short periods of low level UV-B [23]. Short exposures
21 to high UV-B levels induce expression of isoflavone reductase, phenylalanine ammonia
22 lyase, cinnamoyl-CoA reductase, caffeoyl-CoA *O*-methyltransferase, leucoanthocyanidin
23 dioxygenase [24] and flavanone 3-hydroxylase, chalcone synthase, flavonol synthase,

1 chalcone isomerase, dihydroflavonol reductase, cinnamoyl-CoA reductase in *Arabidopsis*
2 [25]. Thus, the altered expression of genes involved in the biosynthesis of phenols is a
3 shared feature of plants exposed to low and high UV-B doses. Given the well-documented
4 accumulation of phenolic metabolites in UV-B-exposed plants, and given the important role
5 of phenolics as antioxidants [29], it is concluded that alterations in ROS metabolism occur
6 across all UV-B-exposure conditions.

7 **ROS and regulation of gene expression**

8 ROS are both stress-inducing compounds and signaling molecules that control, among
9 others, gene expression. Therefore, analyzing regulation of UV-B-dependent gene
10 expression can shed light on the potential role of ROS in UV-acclimation. We have reviewed
11 the expression of genes encoding proteins involved in 'traditional' antioxidative pathways,
12 such as SOD, ascorbate and glutathione metabolic enzymes, as well as isoprenoid, phenolic,
13 and pyridoxine biosynthetic genes, in published microarray data. Fourteen genes have
14 been reported to be up-regulated at least twofold in different studies reported by at least
15 two separate laboratories (Table 1). The protein products of five of these genes are
16 involved in glutathione metabolism, seven in phenylpropanoid metabolism (cinnamates
17 and flavonoids) and one in pyridoxine and one in isoprene biosynthesis (solanesyl
18 diphosphate). Studies using mutants [25,27] have shown that each of these genes needed
19 the UV-B photoreceptor UVR8 [12–14] for expression (Table 1), and that most of them
20 were also dependent on the downstream regulatory proteins CONSTITUTIVELY
21 PHOTOMORPHOGENIC 1 (COP1) and ELONGATED HYPOCOTYL 5 (HY5) [27,28,72]. Thus,
22 the genes belong to the UV-B-specific, 'low UV dose' route of gene expression [10,11] and,

1 therefore, support the concept that even low doses of UV-B can cause changes in
2 antioxidant metabolism.

3 A pertinent question is whether ROS control the expression of the same 14 genes. To
4 answer this, we compared gene expression under UV-B with that under oxidative stress
5 conditions involving various types of ROS (O_3 , O_2^- , H_2O_2 , 1O_2) (Table 1) [77–101]. Stressors
6 such as ozone [77–85,86–89], methyl viologen and high light [89,90,99] increased the
7 expression of several genes involved in antioxidative metabolism; however, overlap with
8 UV-B-induced genes is more or less non-existent. Similarly, expression of genes encoding
9 several antioxidative proteins was increased in the singlet oxygen scavenging-deficient
10 *Arabidopsis flu* mutant [94–96]. However, overlap with UV-B-induced genes was limited.
11 Thus, plants express different enzyme systems and/or different isoenzymes when exposed
12 to UV-B compared with general oxidative stress conditions. There are two notable
13 exceptions to this: (i) the GRX480 glutaredoxin gene (At1g28480) was induced during most
14 of the conditions examined; (ii) norflurazon treatment, inhibiting carotenoid biosynthesis
15 [102] and, thus, leading to singlet oxygen formation in the chloroplast [103,104], resulted
16 in induction of five out of the fourteen UV-B-regulated genes, which infers some overlap in
17 action.

18 Expression of genes linked to eustress and antioxidative protection is not controlled
19 by ROS, but rather through the UVR8 pathway. We therefore hypothesize that low,
20 ecologically relevant doses of UV-B cause eustress, pre-disposing the plant to a state of 'low
21 alert' in case conditions worsen, including activation of genes involved in generic
22 antioxidant defense. This is in contrast to the situation under high-UV-B, distress
23 conditions (Figure 2). For example, similarities in gene expression have been noted

1 between plants exposed to artificially generated ROS and plants exposed to high levels of
2 UV-B [105]. Furthermore, the UV-B-mediated expression of several genes can be modified
3 by treating plants with effectors of ROS metabolism, including free-radical scavengers. It
4 was concluded that ROS mediate responses to high UV-B levels [105].

5 **Conclusion**

6 High levels of UV-B can cause distress in plants. Distressed plants produce elevated levels
7 of ROS. Thus, under these conditions, UV-B exposure, ROS and stress are closely linked.
8 Distress can also occur when plants are simultaneously exposed to ambient UV-B and
9 unfavorable environmental conditions. By contrast, under low, chronic UV conditions,
10 distress is a rare event, prompting the question: do ROS play a role in the cellular and
11 organismal acclimation responses under these conditions? Both low and high levels of UV-
12 B radiation can change antioxidant metabolism (i.e. change the size and/or oxidation-
13 reduction state of the ascorbate, glutathione, and tocopherol pools, and induce
14 accumulation of flavonols and related phenolics, which are strong cellular antioxidants).
15 UV-B also affects expression of genes that impact on the cellular redox state (i.e. genes
16 whose products are involved in glutathione, pyridoxine and phenolic metabolism). We
17 conclude that changes in ROS and antioxidant metabolism are an intrinsic part of both
18 eustress and distress. Nevertheless, low UV-B-induced changes in antioxidant metabolism
19 do not appear to be linked to control of gene expression. Instead, UV-B-specific perception
20 and signaling pathways involving UVR8, COP1 and HY5 [10] comprise the main regulatory
21 pathway under low UV-conditions, activating antioxidant defenses before potential
22 oxidative pressure. ROS-mediated signaling appears to be restricted to high UV-B distress
23 conditions. This conclusion triggers two important questions for future research. Firstly,

1 there is a need to elucidate the precise combination of environmental conditions, and
2 physiological acclimation states where either eustress or distress will occur. Secondly, an
3 important follow-up question is how plants 'balance' generic ROS-specific signaling
4 pathways with stimuli-specific systems such as the UV-B photoreceptor-mediated
5 responses. Understanding this balancing act should give us an insight into the fundamental
6 issues underlying one of the most important plant characteristics, the capability to
7 acclimate to variable environmental conditions.

8 **Acknowledgements**

9 We acknowledge support by COST Action FA0906, UV4Growth. Å.S. received financial
10 support from the Faculty of Business, Science and Technology at Örebro University. É.H.
11 and M.A.K.J. were supported by joint grants from Science Foundation Ireland (SFI project
12 11/RFP.1/EOB/3303) and Hungarian Scientific Research Fund (OTKA NN-85349). We
13 gratefully acknowledge stimulating discussions with Prof. E. Rosenqvist, University of
14 Copenhagen, Denmark.

References

1. Lidon, F. J. C. *et al.* (2012) Decay of the Chloroplast Pool of Ascorbate Switches on the Oxidative Burst in UV-B-Irradiated Rice. *J. Agron. Crop Sci.* 198, 130–144
2. Pitzschke, A. *et al.* (2006). Reactive oxygen species signaling in plants. *Antioxid. Redox Signalling* 8, 1757–1764
3. Gill, S.S. and Tuteja, N. (2010) Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiol. Biochem.* 48, 909–930
4. De Tullio, M.C., (2010) Antioxidants and redox regulation: Changing notions in a changing world *Plant Physiol. Biochem.* 48, 289-291
5. Noctor, G., (2006) Metabolic signalling in defence and stress: the central roles of soluble redox couples. *Plant Cell Environ.* 29, 409-425
6. Ballaré, C.L. *et al.* (2011) Effects of solar ultraviolet radiation on terrestrial ecosystems. Patterns, mechanisms, and interactions with climate change. *Photochem. Photobiol. Sci.* 10, 226–241
7. Li, F.-R. *et al.* (2010) A meta-analysis of the responses of woody and herbaceous plants to elevated ultraviolet-B radiation. *Acta Oecol* 36, 1-9
8. Paul, N. and Gwynn-Jones, D. (2003) Ecological roles of solar UV radiation: towards an integrated approach. *Trends Ecol Evol* 18, 48-55
9. Comont, D. *et al.* (2012) UV responses of *Lolium perenne* raised along a latitudinal gradient across Europe: a filtration study. *Physiol Plant* 145, 604-618

10. Jenkins, G.I. (2009), Signal Transduction in Responses to UV-B Radiation. *Annu. Rev. Plant Biol.* 60, 407-431
11. Brosché, M. and Strid, Å. (2003) Molecular events following perception of ultraviolet-B radiation by plants: UV-B induced signal transduction pathways and changes in gene expression. *Physiol. Plant* 117, 1-10
12. Rizzini, L. *et al.* (2011) Perception of UV-B by the Arabidopsis UVR8 Protein. *Science* 332, 103–106
13. Wu, M. *et al.* (2011) Computational evidence for the role of *Arabidopsis thaliana* UVR8 as UV-B photoreceptor, and identification of its chromophore amino acids. *J. Chem Inf. Model.* 51, 1287-1295
14. Christie, J.M. *et al.* (2012) Plant UVR8 Photoreceptor Senses UV-B by Tryptophan-Mediated Disruption of Cross-Dimer Salt Bridges. *Science* 335, 1492-1496
15. Heijde, M. and Ulm, R. (2012) UV-B photoreceptor-mediated signaling in plants. *Trends Plant Sci.* 17, 230–237
16. Wu, D. *et al.* (2012) Structural basis of ultraviolet-B perception by UVR8. *Nature* 484, 214–219
17. Levitt, J. (1980) Responses of plants to environmental stresses. In: *Chilling, Freezing, and High Temperature Stresses* (Vol. I), pp. 101–107. Academic Press, New York
18. Lichtenthaler, H.K. (1996) Vegetation Stress: an Introduction to the Stress Concept in Plants. *J Plant Physiol.* 148, 4-14
19. Gaspar, T. *et al.* (2002) Concepts in plant stress physiology. Application to plant tissue cultures. *Plant Growth Regul* 37, 263–285

20. Kranner, I. *et al.* (2010) Tansley review; What is stress? Concepts, definitions and applications in seed science. *New Phytol.* 188, 655-673
21. Jansen, M.A.K. *et al.* (2008) Plant stress and human health; do human consumers benefit from UV-B acclimated crops? *Plant Sci.* 175, 449-458
22. Potters, G. *et al.* (2010) The cellular redox state in plant stress biology - a charging concept. *Plant Physiol. Biochem.* 48, 292-300
23. Brosché, M. *et al.* (2002) Gene regulation by low level UV-B radiation: identification by DNA array analysis. *Photochem. Photobiol. Sci.* 1, 656-664
24. Ulm, R. *et al.* (2004) Genome-wide analysis of gene expression reveals function of the bZIP transcription factor HY5 in the response of Arabidopsis. *Proc. Natl. Acad. Sci. U. S. A.* 101, 1397-1402
25. Brown, B. A. *et al.* (2005) A UV-B-specific signaling component orchestrates plant UV protection. *Proc. Natl. Acad. Sci. U.S.A.* 102, 18225-18230
26. Hectors, K. *et al.* (2007) *Arabidopsis thaliana* plants acclimated to low dose rates of ultraviolet B radiation show specific changes in morphology and gene expression in the absence of stress symptoms. *New Phytol.* 175, 255-270
27. Brown, B.A. and Jenkins, G.I. (2008) UV-B signaling pathways with different fluence-rate response profiles are distinguished in mature Arabidopsis leaf tissue by requirement for UVR8, HY5, and HYH. *Plant Physiol.* 146, 576-588
28. Favory, J.J. *et al.* (2009) Interaction of COP1 and UVR8 regulates UV-B induced photomorphogenesis and stress acclimation in Arabidopsis. *EMBO J.* 28, 591-601
29. Agati, G., and Tattini, M. (2010) Multiple functional roles of flavonoids in photoprotection. *New Phytol.* 186, 786-793

30. Schreiner, M. *et al.* (2012) UV-B induced secondary plant metabolites - potential benefits for plant and human health, *Crit. Rev. Plant Sci.* 31, 229–240
31. Jansen, M.A.K. *et al.* (2012) UV-B induced morphogenesis: Four players or a quartet? *Plant Signal Behav* 7, 1185-1187
32. Caldwell, M.M. and Flint, S.D. (1994) Stratospheric ozone reduction, solar UV-B radiation and terrestrial ecosystems *Clim. Change* 28, 375-394
33. Caldwell, M.M. *et al.* (1998) Effects of increased solar ultraviolet radiation on terrestrial ecosystems. *J. Photochem. Photobiol. B* 46, 40–52
34. Fiscus, E.L. and Booker, F.L. (1995) Is increased UV-B a threat to crop photosynthesis and productivity? *Photosynth. Res.* 43, 81-92
35. Albert, K.R. *et al.* (2011) Ambient UV-B radiation reduces PSII performance and net photosynthesis in high Arctic *Salix arctica*. *Environ. Exp. Bot.* 72, 439-447
36. Arróniz-Crespo, M. *et al.* (2011) Impacts of long-term enhanced UV-B radiation on bryophytes in two sub-Arctic heathland sites of contrasting water availability *Ann. Bot.* 108, 557-565
37. Belnap, J. *et al.* (2008) Global change and biological soil crusts: effects of ultraviolet augmentation under altered precipitation regimes and nitrogen additions. *Glob. Change Biol.* 14, 670-686
38. Hofmann, R.W. *et al.* (2003) Sensitivity of white clover to UV-B radiation depends on water availability, plant productivity and duration of stress. *Glob Change Biol* 9, 473-477

39. Nogués, S. and Baker, N.R. (2000) Effects of drought on photosynthesis in Mediterranean plants grown under enhanced UV-B radiation. *J. Exp. Bot.* 51, 1309–1317
40. Lau, T.S.L. *et al.* (2006) Ambient levels of UV-B in Hawaii combined with nutrient deficiency decrease photosynthesis in near-isogenic maize lines varying in leaf flavonoids: Flavonoids decrease photoinhibition in plants exposed to UV-B *Photosynthetica* 44, 394-403
41. Singh, S. *et al.* (2011) Modification in growth, biomass and yield of radish under supplemental UV-B at different NPK levels. *Ecotoxicol. Environ. Saf.* 74, 897–903
42. Nogués, S. *et al.* (1998) Ultraviolet-B radiation effects on water relations, leaf development, and photosynthesis in droughted pea plants, *Plant Physiol.* 117, 173–181
43. Hideg, É. *et al.* (2003) Detoxification function of aldose/aldehyde reductase during drought and UV-B (280-320 nm) stresses. *Plant Cell Environ.* 26, 513-522
44. Manetas, Y. *et al.* (1997) Beneficial effects of enhanced UV-B radiation under field conditions: improvement of needle water relations and survival capacity of *Pinus pinea* L. seedlings during the dry Mediterranean summer. *Plant Ecol.* 128, 101-108
45. Kubis, J. and Rybus-Zajac, M. (2008) Drought and excess UV-B irradiation differentially alter the antioxidant system in cucumber leaves. *Acta Biol. Cracov., Ser. Bot.* 50/2, 35-41
46. Hideg, É. And Vass, I. (1996) UV-B induced free radical production in plant leaves and isolated thylakoid membranes. *Plant Sci.* 115, 251-260

47. Kalbina, I. and Strid, Å. (2006) The role of NADPH oxidase and MAP kinase phosphatase in UV-B-dependent gene expression in *Arabidopsis*. *Plant Cell Environ.* 29, 1783-1793
48. Elstner, E.F. and Osswald, W. (1994) Mechanisms of oxygen activation during plant stress. *Proc. - R. Soc. Edinburgh, Sect. B: Biol.* 102, 131-154
49. Macpherson, A.N. *et al.* (1993) Direct detection of singlet oxygen from isolated Photosystem II reaction centres. *Biochim Biophys Acta* 1143, 301-309
50. Wardman, P. (2007) Fluorescent and luminescent probes for measurement of oxidative and nitrosative species in cells and tissues: Progress, pitfalls, and prospects. *Free Radical Biol. Med.* 43, 995–1022
51. Barta, C. *et al.* (2004) Differences in the ROS generating efficacy of various ultraviolet wavelengths in detached spinach leaves. *Funct. Plant Biol.* 31, 23-28
52. Lidon, F.C. and Ramalho, J.C. (2011) Impact of UV-B irradiation on photosynthetic performance and chloroplast membrane components in *Oryza sativa* L. *J. Photochem. Photobiol. B: Biol.* 104, 457-466
53. Schmitz-Hoerner, R. and Weissenböck, G. (2003) Contribution of phenolic compounds to the UV-B screening capacity of developing barley primary leaves in relation to DNA damage and repair under elevated UV-B levels. *Phytochem.* 64, 243-255
54. Dai, Q. *et al.* (1997) Response of oxidative stress defense systems in rice (*Oryza sativa*) leaves with supplemental UV-B radiation. *Physiol. Plant.* 101, 301–308

55. Janero, D.R. (2003) Malondialdehyde and thiobarbituric acid-reactivity as diagnostic indices of lipid peroxidation and peroxidative tissue injury. *Free Radical Biol. Med.* 9, 515-540
56. Hideg, É. *et al.* (1997) Increased levels of monodehydroascorbate radical in UV-B irradiated broad bean leaves. *Plant Cell Physiol.* 38, 684-690
57. Kalbin, G. *et al.* (1997) Ultraviolet-B-radiation induced changes in nicotinamide and glutathione metabolism and gene expression in plants, *Eur. J. Biochem.* 249, 465-472
58. Jayaraj, J. and Punja, Z.K. (2008) Transgenic carrot plants accumulating ketocarotenoids show tolerance to UV and oxidative stresses. *Plant Physiol. Biochem.* 46, 875-883
59. Heuberger, H. *et al.* (2004) Precision stressing by UV-B radiation to improve quality of spinach under protected cultivation, *Acta Hort.* 659, 201-206
60. Anonymous (2003) Ultraviolet lighting for vegetables: enhancing the vitamin C and vitamin E content. FFTC Practical Technology, Report PT2003-31, 1-2
61. Fini, A. *et al.* (2011) Stress-induced flavonoid biosynthesis and the antioxidant machinery of plants. *Plant Signaling Behav.* 6, 709-711
62. Ryan, K.G. *et al.* (1998) UVB radiation induced increase in quercetin:kaempferol ratio in wild-type and transgenic lines of Petunia. *Photochem. Photobiol.* 68, 323-330
63. Costa, H. *et al.* (2002) Effect of UV-B radiation on antioxidant defense system in sunflower cotyledons. *Plant Sci.* 162, 939-945

64. Santos, I. *et al.* (2004) Biochemical and ultrastructural changes in leaves of potato plants grown under supplementary UV-B radiation. *Plant Sci.* 167, 925-935
65. Hideg, É. *et al.* (2006) A comparison of UV-B induced stress responses in three barley cultivars. *Funct. Plant Biol.* 33, 77-90
66. Agrawal, S.B. and Rathore, D. (2007) Changes in oxidative stress defense system in wheat (*Triticum aestivum* L.) and mung bean (*Vigna radiata* L.) cultivars grown with and without mineral nutrients and irradiated by supplemental ultraviolet-B. *Environ. Exp. Bot.* 59, 21-33
67. Xu, C. *et al.* (2008) Impact of solar ultraviolet-B radiation on the antioxidant defense system in soybean lines differing in flavonoid contents. *Environ. Exp. Bot.* 63, 39-48
68. Majer, P. and Hideg, É. (2012) Developmental stage is an important factor that determines the antioxidant responses of young and old grapevine leaves under UV irradiation in a green-house. *Plant Physiol. Biochem.* 50, 15-23
69. Agrawal, S.B. *et al.* (2009) Ultraviolet-B induced changes in gene expression and antioxidants in plants. In: *Advances in Botanical Research* (Vol. 52), (Jacquot, J.P., ed.), pp 47-86, Academic Press
70. Yang, S.H. *et al.* (2007) The effects of UV-B radiation on photosynthesis in relation to Photosystem II photochemistry, thermal dissipation and antioxidant defenses in winter wheat (*Triticum aestivum* L.) seedlings at different growth temperatures. *Funct. Plant Biol.* 34, 907-917
71. Ristilä, M. *et al.* (2011) The role of the pyridoxine (vitamin B₆) biosynthesis enzyme PDX1 in ultraviolet-B radiation responses in plants. *Plant Physiol. Biochem.* 49, 284-292

72. Oravecz, A. *et al.* (2006) CONSTITUTIVELY PHOTOMORPHOGENIC 1 is required for the UV-B response in *Arabidopsis*. *Plant Cell* 18, 1975-1990
73. Ulm, R. *et al.* (2006) Transcription profiling of *Arabidopsis* response to UV-B. E-MEXP-550. (<http://www.ebi.ac.uk/arrayexpress/experiments/E-MEXP-550>), Accessed April 29, 2012.
74. Kretsch, T. *et al.* (2007) AtGenExpress:Light treatments. GSE5617. (<http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE5617>), Accessed April 29, 2012.
75. Raschke, M. *et al.* (2011) Enhanced levels of vitamin B₆ increase aerial organ size and positively affect stress tolerance in *Arabidopsis*. *Plant J.* 66, 414-432
76. Wu, M. *et al.* (2011) Theoretical Study of Pyridoxine (Vitamin B₆) Photolysis. *J. Phys. Chem. A.* 115, 13556–13563
77. Overmeyer, K. *et al.* (2000) Ozone-sensitive *Arabidopsis rcd1* mutant reveals opposite roles for ethylene and jasmonate signaling pathways in regulating superoxide-dependent cell death. *Plant Cell* 12, 1849-1862
78. Tamaoki, M. *et al.* (2003) Transcriptome analysis of O₃-exposed *Arabidopsis* reveals that multiple signal pathways act mutually antagonistically to induce gene expression. *Plant Mol. Biol.* 53, 443-456
79. Ludwikow, A. *et al.*, (2004) Ozone-induced oxidative stress response in *Arabidopsis*: transcription profiling by microarray approach. *Cell Mol. Biol. Lett.* 9, 829-842
80. Mittler, R. *et al.* (2004) Reactive oxygen gene network of plants. *Trends Plant Sci.* 9, 491-498

81. Overmeyer, K. *et al.* (2005) Ozone-induced programmed cell death in the *Arabidopsis radical-induced cell death1* mutant. *Plant Physiol.* 137, 1092-1104
82. Mahalingam, R. *et al.* (2005) Temporal evolution of the *Arabidopsis* oxidative stress response. *Plant Mol. Biol.* 57, 709-730
83. Mahalingam, R. *et al.* (2006) Analysis of oxidative signaling induced by ozone in *Arabidopsis thaliana*. *Plant Cell Environ.* 29, 1357-1371
84. Tosti, N. *et al.* (2006) Gene expression profiles of O₃-treated *Arabidopsis* plants. *Plant Cell Environ.* 29, 1686-1702
85. Gadjev, I. *et al.* (2006) Transcriptomic footprints disclose specificity of reactive oxygen species signaling in *Arabidopsis*. *Plant Physiol.* 141, 436-445
86. Ludwików, A. *et al.* (2009) Gene expression profiling of ozone-treated *Arabidopsis abi1td* insertional mutant: protein phosphatase 2C ABI1 modulates biosynthesis ratio of ABA and ethylene. *Planta* 230, 1003-1017
87. Blomster, T. *et al.* (2011) Apoplastic reactive oxygen species transiently decrease auxin signaling and cause stress-induced morphogenic response in *Arabidopsis*. *Plant Physiol.* 157, 1866-1883
88. Short, E. *et al.* (2008) Functional Genomics of Ozone Stress in *Arabidopsis*, GSE5722. (<http://www.ncbi.nlm.nih.gov/projects/geo/query/acc.cgi?acc=GSE5722>), Accessed April 29, 2012
89. Ditzer, A. *et al.* (2004) AtGenExpress: Oxidative stress time course, ME00340. http://arabidopsis.org/servlets/TairObject?type=expression_set&id=1007966941. Accessed April 29, 2012

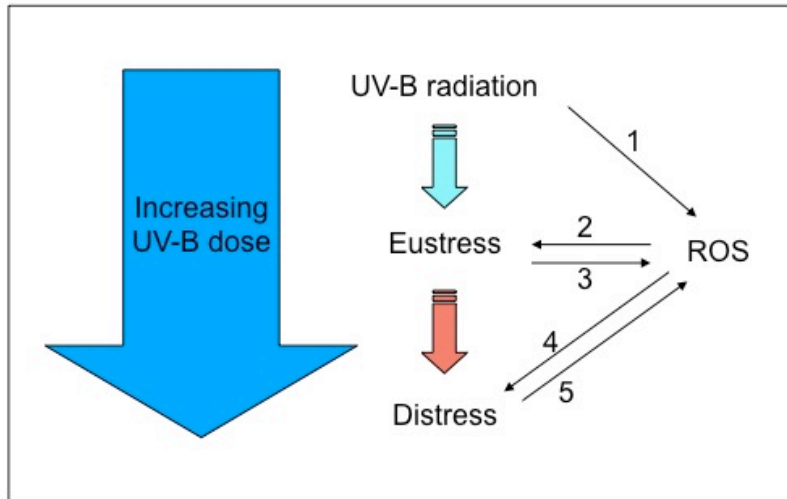
90. Scarpeci, T.E. *et al.* (2008) Generation of superoxide anion in chloroplasts of *Arabidopsis thaliana* during active photosynthesis: a focus on rapidly induced genes. *Plant Mol. Biol.* 66, 361-378
91. Lee, K.P. *et al.* (2007) EXECUTER1- and EXECUTER2-dependent transfer of stress-related signals from the plastid to the nucleus of *Arabidopsis thaliana*. *Proc. Natl. Acad. Sci. U.S.A.* 104, 10270-10275
92. Laloi, C. *et al.* (2008) Cross-talk between singlet oxygen- and hydrogen peroxide-dependent signaling of stress responses in *Arabidopsis thaliana*. *Proc. Natl. Acad. Sci. U.S.A.* 104, 672-677
93. Mittler, R. *et al.* (2006) Hydrogen peroxide stress and Zat12 over-expression in *Arabidopsis*, GSE5530.
(<http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE5530>), Accessed April 29, 2012
94. McCormac, A. *et al.* (2007) Seedling transcriptome affected by norflurazon-induced photobleaching of chloroplasts, GSE 5726.
(<http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE5726>), Accessed April 29, 2012
95. Koussevitzky, S. *et al.* (2008) Signals from chloroplasts converge to regulate nuclear gene expression. *Science* 316, 715-719
96. Op den Camp, R.G.L. *et al.* (2003) Rapid Induction of Distinct Stress Responses after the Release of Singlet Oxygen in *Arabidopsis*. *Plant Cell* 15, 2320-2332
97. Daveltova, S. *et al.* (2003) Cytosolic ascorbate peroxidase 1 is a central component of the reactive oxygen gene network of *Arabidopsis*. *Plant Cell* 17, 268-281

98. Gechev, T.S. *et al.* (2005) Hydrogen peroxide-induced cell death in Arabidopsis: transcriptional and mutant analysis reveals a role of an oxoglutarate-dependent dioxygenase gene in the cell death process. *IUBMB Life* 57, 181-188
99. Shirras, A.D. *et al.* (2005) Transcript profiling by array of Arabidopsis after exposure to ozone, E-MEXP-342. <http://www.ebi.ac.uk/arrayexpress/experiments/E-MEXP-342>. Accessed April 29, 2012
100. Rizhsky, L. *et al.* (2003) The water-water cycle is essential for chloroplast protection in the absence of stress. *J. Biol. Chem.* 278, 38921-38925
101. Vanderauwera, S. *et al.* (2005) Genome-wide analysis of hydrogen peroxide-regulated gene expression in Arabidopsis reveals a high light-induced transcriptional cluster involved in anthocyanin biosynthesis. *Plant Physiol.* 139, 806-821
102. Sandmann, G. *et al.* (1991) Phytoene Desaturase, the Essential Target for Bleaching Herbicides. *Weed Sci.* 39, 474-479
103. Sandmann, G. *et al.* (1980) The inhibitory mode of action of the pyridazinone herbicide norflurazon on a cell-free carotenogenic enzyme system. *Pestic. Biochem. Physiol.* 14, 185-191
104. Beat, B. *et al.* (2007) Role of singlet oxygen in chloroplast to nucleus retrograde signaling in *Chlamydomonas reinhardtii*. *FEBS Lett.* 581, 5555-5560
105. Mackerness, S.A.H. *et al.* (2001) Early signaling components in ultraviolet-B responses: distinct roles for different reactive oxygen species and nitric oxide. *FEBS Lett.* 489, 237-242

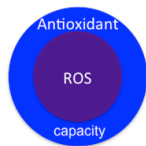
Legends

Figure 1. UV-B radiation has been well documented to induce eustress, leading to UV-acclimation. UV-B-induced distress appears to be a relatively rare phenomenon under natural light conditions. There is some evidence that UV-B exposure can directly induce ROS (1), although it is not clear to what extent this happens under realistic UV-B conditions. ROS that are formed may contribute to eustress and the UV acclimation response (2) or cause oxidative damage (4). Conversely, further ROS can be produced as part of the UV response of the plant; either as part of UV-acclimation by induction of NAPH oxidase activity (3) or as a result of metabolic disruption (distress) of, for example, photosynthetic electron transfer reactions (5).

Figure 2. ROS levels and antioxidant capacities under physiologically relevant UV-B levels (eustress) and under high UV-B conditions (distress). Under physiologically relevant UV-B levels, the ROS scavenging capacity, regulated by the UV-B-specific signaling pathway containing the UVR8 UV-B photoreceptor and the COP1 and HY5 signaling components, is sufficient to deal with the oxidative pressure inflicted by UV-B. Under high UV-B conditions, the UV-B levels are high enough to lead to a massive development of ROS, over-riding the antioxidant capacity regulated by non-specific stress pathways and contributing to both signaling and gene expression.

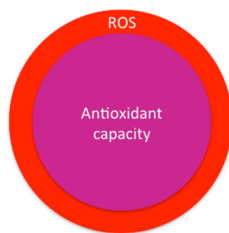


Ambient UV-B levels
 UV-B-specific gene expression regulated by UVR8
 Low ROS levels
 Sufficient ROS scavenging capacity



Eustress

High UV-B levels
 Gene expression regulated by ROS
 High ROS levels
 Insufficient ROS scavenging capacity



Distress