

Resistance to multi organ damage after hemorrhagic shock induced ischemia/ reperfusion in arctic ground squirrels

Bogren LK, Olson JM, Carpluk J, Moore JT, Drew KL

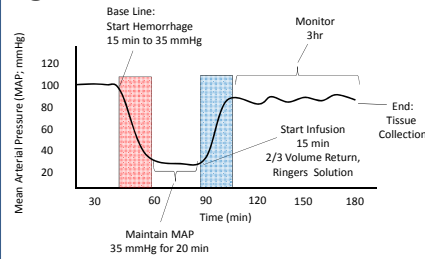
University of Alaska Fairbanks, Institute of Arctic Biology, Department of Chemistry and Biochemistry

INTRODUCTION

- Worldwide hemorrhagic shock is the number one cause of death in trauma patients, the majority of those die from multi organ dysfunction syndrome [1, 2].
 - During hemorrhagic shock (HS), the body undergoes global ischemia as blood pressure drops below the threshold at which tissues can be adequately perfused.
 - Resistance to ischemic injury is a characteristic of hibernating mammals, including ground squirrels.
 - There is debate on if this resistance is dependent on hibernation season or if it is an intrinsic plasticity of the organism.
- QUESTION: Are AGS protected from HS-induced ischemia reperfusion (I/R) injury on the whole organism and tissue-specific levels and if any protection is dependent upon their hibernation season.**

METHODS

Figure 1: HS isobaric procedure



Prior to hemorrhage (baseline), immediately after resuscitation and 3 hours after resuscitation, blood was sampled and analyzed physiological parameters (Table 1). Blood sampled during hemorrhage and 3hr after resuscitation was also analyzed for markers of organ damage and levels of inflammatory cytokines as indicators of systemic inflammation. After 3 hours, organs were collected and fixed for histological analysis (Table 1). The primary endpoints were: 1) plasma markers for organ damage, 2) histopathological damage to tissues, 3) circulating and tissue levels of inflammatory cytokines. In addition, a group of animals were monitored for 72 hours after reperfusion to assess survival rates and any physiological impairments. During hemorrhage, infusion, and monitoring period, animals' body and head temperatures were maintained between 36.5-37.5 °C. Sham animals were instrumented and monitored in the same way as HS animals, except blood was not withdrawn for hemorrhage. AGS were tested during their summer (euthermic) and winter (interbout arousal) season. All AGS were housed at 2°C, 4L:20D conditions, year round. Interbout arousal AGS were aroused 20 hours prior to experiment. All animals were fasted 20 hours prior to experiment.

Table 1: Components of analysis

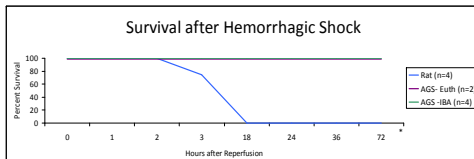
Analysis	Parameters
Physiological	HR, MABP, temperature (head, core, limb), pO ₂ , O ₂ Sat, pCO ₂ , pH, bicarb, base excess, blood glucose, blood lactate, complete blood count (CBC)
Blood chemistries	blood urea nitrogen (BUN), creatinine, bilirubin, creatine phosphokinase (CPK), lactate dehydrogenase (LDH), aspartate aminotransferase (AST), and alanine aminotransferase (ALT)
Histopathology	heart, lungs, intestine, kidney, spleen, and liver
Cytokine Immunocytochemistry	IL-1 alpha, IL-1 beta, IL-2, IL-4, IL-6, IL-10, IL-12, TNF-alpha, INF-gamma, GM-CSF (Luminex-EDTA plasma)

Table 2: AGS seasonal group parameters

Summer/Euthermic	Winter/IBA (prior to induced arousal)
Tb > 35°C	Tb < 5°C
RR > 80 rpm	RR < 5 rpm
Alert and responsive	Wood shavings on back undisturbed
No spontaneous torpor ≥ 4 wk	> 3 prior torpor bouts

RESULTS

Figure 2: Rats do not survive 18 hrs after HS while AGS survive a minimum of 72 hrs afterward



*AGS were euthanized at the 72hr timepoint

Figure 3: AGS recover to normal behavior by 18 hrs after HS

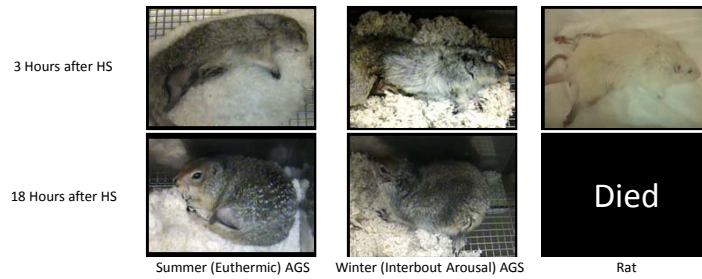
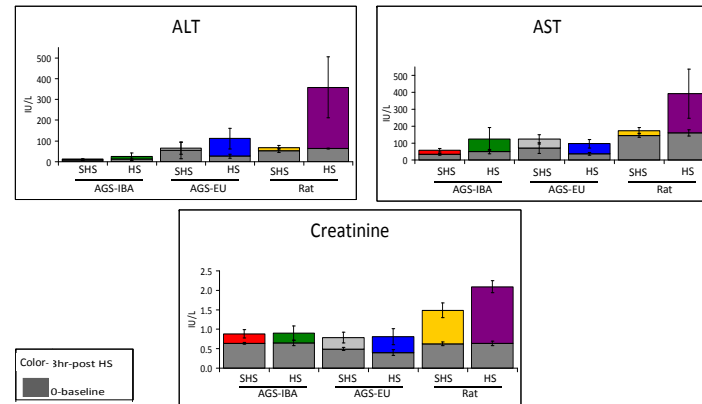
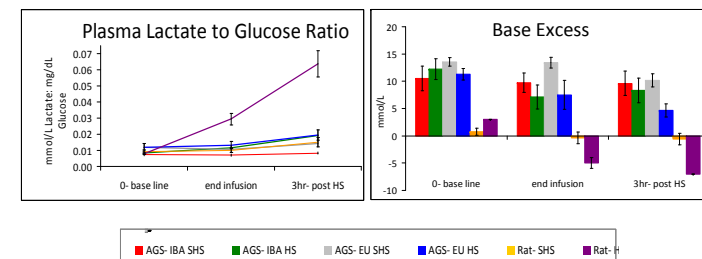


Figure 4: AGS do not show early indicators of organ damage in the kidney (creatinine) or liver (ALT, AST) regardless of hibernation season



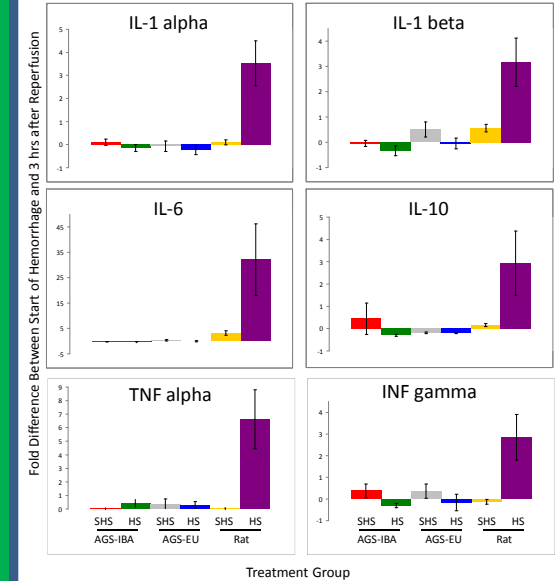
Data are expressed as Mean ± SEM. N ≥ 6 for all groups. SHS- Sham Hemorrhagic Shock, HS- Hemorrhagic Shock, IBA- Interbout Arousal, EU- Euthermic.

Figure 5: Metabolic shift indicated by an increase in lactate production and negative base excess occurs in rats during and after HS but not in AGS



Data are expressed as Mean ± SEM. N ≥ 6 for all groups. SHS- Sham Hemorrhagic Shock, HS- Hemorrhagic Shock, IBA- Interbout Arousal, EU- Euthermic.

Figure 6: Circulating plasma cytokine levels do not increase in AGS after HS



Data are expressed as Mean ± SEM. N ≥ 6 for all groups. SHS- Sham Hemorrhagic Shock, HS- Hemorrhagic Shock, IBA- Interbout Arousal, EU- Euthermic.

CONCLUSIONS

Independent of hibernation season, AGS were resistant to HS induced I/R injury on the whole organism and tissue specific level.

- Both euthermic and interbout arousal AGS
 - can survive without apparent physiological deficit for 3 days after HS at euthermic body temperatures when blood loss corresponds to an MAP of 35 mmHg (~30% total blood volume)
- Maintain a high base excess during and after HS
- Do not show blood serum markers for organ damage
- Do not have a systemic inflammatory cytokine response after HS I/R injury

Acknowledgments: We would like to thank UAF Veterinary Services, VDX Veterinary Diagnostics, and Fairbanks Memorial Hospital for assistance with sample analysis.

This research is funded in part by USAMRMC W81XWH09-2-0134, Eagle-I Core Bucks award, and the University of Alaska Graduate School Dissertation Completion Fellowship.

Literature Cited:
 1. Hietbrink, F., et al., *Trauma: the role of the innate immune system.* World J Emerg Surg. 2006. 1: p. 15.; 2. Angele, M.K., C.P. Schneider, and I.H. Chaudry, *Bench-to-bedside review: latest results in hemorrhagic shock.* Crit Care, 2008. 12(4): p. 218;