

## DARWIN REVIEW

# Wheat

P. R. Shewry\*

Department of Plant Sciences, Rothamsted Research, Harpenden, Hertfordshire AL5 2JQ, UK

Received 8 December 2008; Revised 11 February 2009; Accepted 13 February 2009

## Abstract

**Wheat is the dominant crop in temperate countries being used for human food and livestock feed. Its success depends partly on its adaptability and high yield potential but also on the gluten protein fraction which confers the viscoelastic properties that allow dough to be processed into bread, pasta, noodles, and other food products. Wheat also contributes essential amino acids, minerals, and vitamins, and beneficial phytochemicals and dietary fibre components to the human diet, and these are particularly enriched in whole-grain products. However, wheat products are also known or suggested to be responsible for a number of adverse reactions in humans, including intolerances (notably coeliac disease) and allergies (respiratory and food). Current and future concerns include sustaining wheat production and quality with reduced inputs of agrochemicals and developing lines with enhanced quality for specific end-uses, notably for biofuels and human nutrition.**

**Key words:** Allergy, bread, crop evolution, dietary fibre, flour, gluten proteins, grain, intolerance, nutrition, processing, wheat.

## Introduction

Wheat is counted among the ‘big three’ cereal crops, with over 600 million tonnes being harvested annually. For example, in 2007, the total world harvest was about 607 m tonnes compared with 652 m tonnes of rice and 785 m tonnes of maize (<http://faostat.fao.org/>). However, wheat is unrivalled in its range of cultivation, from 67° N in Scandinavia and Russia to 45° S in Argentina, including elevated regions in the tropics and sub-tropics (Feldman, 1995). It is also unrivalled in its range of diversity and the extent to which it has become embedded in the culture and even the religion of diverse societies.

Most readers will be aware of the significance of bread in the Judaeo-Christian tradition including the use of matzo (hard flat bread) at the Jewish Passover and of bread to represent the ‘host’ at the Christian Eucharist (Holy Communion). The latter may be a thin unleavened wafer, similar to the Jewish matzo, in the Roman Catholic Church and some Protestant denominations, or leavened in other Protestant denominations and the Eastern Orthodox Church. But how many readers are aware that bread is treated as sacred in everyday life in the largely Muslim communities of Central Asia, such as Uzbekistan and Kyrgyzstan? In this culture, the leavened round breads

(nan) are stamped before baking and must be treated with respect, including being kept upright and never left on the ground or thrown away in public. These customs almost certainly originate from earlier indigenous religions in the Middle East in which wheat played a similar role and was sometimes equated with the sun and its god.

Although such cultural and religious traditions are fascinating and will certainly reward further study, they are essentially outside the scope of this article which will examine why wheat has developed and continues to be so successful as a crop and food source.

## Origin and evolution of wheat

The first cultivation of wheat occurred about 10 000 years ago, as part of the ‘Neolithic Revolution’, which saw a transition from hunting and gathering of food to settled agriculture. These earliest cultivated forms were diploid (genome AA) (einkorn) and tetraploid (genome AABB) (emmer) wheats and their genetic relationships indicate that they originated from the south-eastern part of Turkey (Heun *et al.*, 1997; Nesbitt, 1998; Dubcovsky and Dvorak, 2007). Cultivation spread to the Near East by about 9000

\* To whom correspondence should be addressed: E-mail: peter.shewry@bbsrc.ac.uk

years ago when hexaploid bread wheat made its first appearance (Feldman, 2001).

The earliest cultivated forms of wheat were essentially landraces selected by farmers from wild populations, presumably because of their superior yield and other characteristics, an early and clearly non-scientific form of plant breeding! However, domestication was also associated with the selection of genetic traits that separated them from their wild relatives. This domestication syndrome has been discussed in detail by others, but two traits are of sufficient importance to mention here. The first is the loss of shattering of the spike at maturity, which results in seed loss at harvesting. This is clearly an important trait for ensuring seed dispersal in natural populations and the non-shattering trait is determined by mutations at the *Br* (*brittle rachis*) locus (Nalam *et al.*, 2006).

The second important trait is the change from hulled forms, in which the glumes adhere tightly to the grain, to free-threshing naked forms. The free forms arose by a dominant mutant at the *Q* locus which modified the effects of recessive mutations at the *Tg* (*tenacious glume*) locus (Jantasuriyarat *et al.*, 2004; Simons *et al.*, 2006; Dubkovsky and Dvorak, 2007).

Cultivated forms of diploid, tetraploid, and hexaploid wheat all have a tough rachis apart from the spelt form of bread wheat. Similarly, the early domesticated forms of einkorn, emmer, and spelt are all hulled, whereas modern forms of tetraploid and hexaploid wheat are free-threshing.

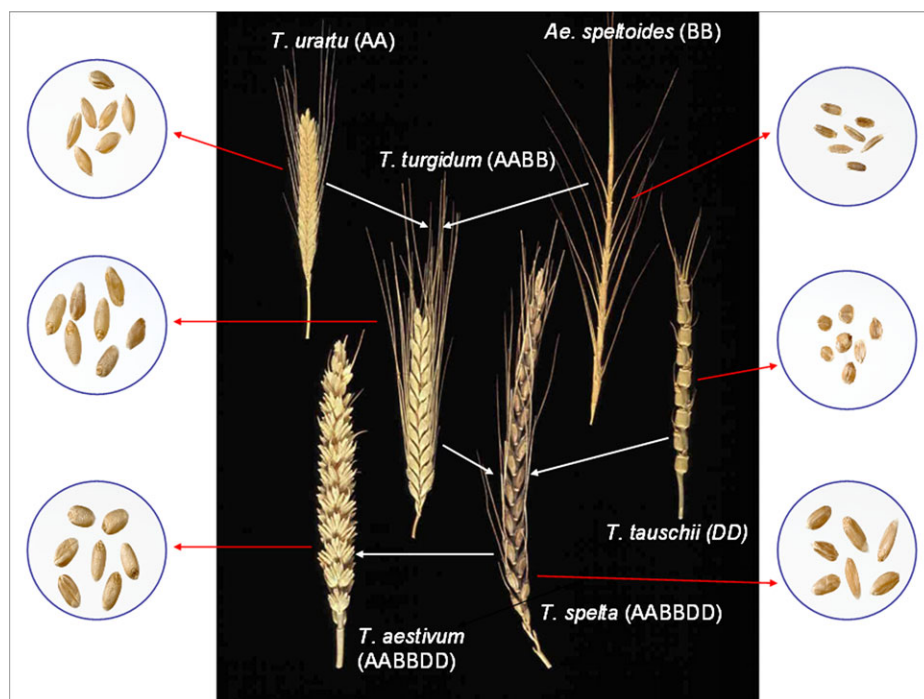
Whereas einkorn and emmer clearly developed from the domestication of natural populations, bread wheat has only existed in cultivation, having arisen by hybridization of cultivated emmer with the unrelated wild grass *Triticum*

*tauschii* (also called *Aegilops tauschii* and *Ae. squarosa*). This hybridization probably occurred several times independently with the novel hexaploid (genome AABBDD) being selected by farmers for its superior properties. The evolution of modern wheats is illustrated in Fig. 1 which also shows examples of spikes and grain.

The genetic changes during domestication mean that modern wheats are unable to survive wild in competition with better adapted species. This was elegantly demonstrated by John Bennet Lawes in the 1880s when he decided to allow part of the famous long-term Broadbalk experiment at Rothamsted to return to its natural state (Dyke, 1993). He therefore left part of the wheat crop unharvested in 1882 and monitored the growth in successive years. After a good crop in 1883 the weeds dominated and in 1885 the few remaining wheat plants (which were spindly with small ears) were collected and photographed.

The A genomes of tetraploid and hexaploid wheats are clearly related to the A genomes of wild and cultivated einkorn, while the D genome of hexaploid wheat is clearly derived from that of *T. tauschii*. In fact, the formation of hexaploid wheat occurred so recently that little divergence has occurred between the D genomes present in the hexaploid and diploid species. By contrast, the B genome of tetraploid and hexaploid wheats is probably derived from the S genome present in the Sitopsis section of *Aegilops*, with *Ae. speltoides* being the closest extant species. The S genome of *Ae. speltoides* is also closest to the G genome of *T. timopheevi*, a tetraploid species with the A and G genomes (Feldman, 2001).

The spread of wheat from its site of origin across the world has been elegantly described by Feldman (2001) and



**Fig. 1.** The evolutionary and genome relationships between cultivated bread and durum wheats and related wild diploid grasses, showing examples of spikes and grain. Modified from Snape and Pánková (2006), and reproduced by kind permission of Wiley-Blackwell.

is only summarized here. The main route into Europe was via Anatolia to Greece (8000 BP) and then both northwards through the Balkans to the Danube (7000 BP) and across to Italy, France and Spain (7000 BP), finally reaching the UK and Scandinavia by about 5000 BP. Similarly, wheat spread via Iran into central Asia reaching China by about 3000 BP and to Africa, initially via Egypt. It was introduced by the Spaniards to Mexico in 1529 and to Australia in 1788.

## Cultivated wheats today

Currently, about 95% of the wheat grown worldwide is hexaploid bread wheat, with most of the remaining 5% being tetraploid durum wheat. The latter is more adapted to the dry Mediterranean climate than bread wheat and is often called pasta wheat to reflect its major end-use. However, it may also be used to bake bread and is used to make regional foods such as couscous and bulgar in North Africa. Small amounts of other wheat species (einkorn, emmer, spelt) are still grown in some regions including Spain, Turkey, the Balkans, and the Indian subcontinent. In Italy, these hulled wheats are together called faro (Szabó and Hammer, 1996) while spelt continues to be grown in Europe, particularly in Alpine areas (Fossati and Ingold, 2001).

The recent interest in spelt and other ancient wheats (including kamut, a tetraploid wheat of uncertain taxonomy, related to durum wheat) as healthy alternatives to bread wheat (Abdel-Aal *et al.*, 1998) may also lead to wider growth for high value niche markets in the future.

## Why has wheat been so successful?

Despite its relatively recent origin, bread wheat shows sufficient genetic diversity to allow the development of over 25 000 types (Feldman *et al.*, 1995) which are adapted to a wide range of temperate environments. Provided sufficient water and mineral nutrients are available and effective control of pests and pathogens is ensured, yields can exceed 10 tonnes ha<sup>-1</sup>, comparing well with other temperate crops. However, deficiencies in water and nutrients and the effects of pests and pathogens cause the global average yield to be low, at about 2.8 tonnes ha<sup>-1</sup>. Wheat is also readily harvested using mechanical combine harvesters or traditional methods and can be stored effectively indefinitely before consumption, provided the water content is below about 15% dry weight and pests are controlled.

There is no doubt that the adaptability and high yields of wheat have contributed to its success, but these alone are not sufficient to account for its current dominance over much of the temperate world. The key characteristic which has given it an advantage over other temperate crops is the unique properties of doughs formed from wheat flours, which allow it to be processed into a range of breads and other baked products (including cakes and biscuits), pasta and noodles, and other processed foods. These properties depend on the structures and interactions of the grain storage proteins, which together form the 'gluten' protein fraction.

## Wheat gluten proteins and processing properties

Transcriptomic studies have shown that over 30 000 genes are expressed in the developing wheat grain (Wan *et al.*, 2008) while proteomic analysis of mature grain has revealed the presence of about 1125 individual components (Skylas *et al.*, 2000). However, many of these components are present in small amounts and have little or no impact on the utilization of the grain, with one protein fraction being dominant in terms of amount and impact. This fraction is the prolamin storage proteins, which correspond to the gluten proteins. The precise number of individual gluten protein components has not been determined, but 2D gel analyses suggest that about 100 is a reasonable estimate. Together they have been estimated to account for about 80% of the total grain protein in European wheats (Seilmeier *et al.*, 1991).

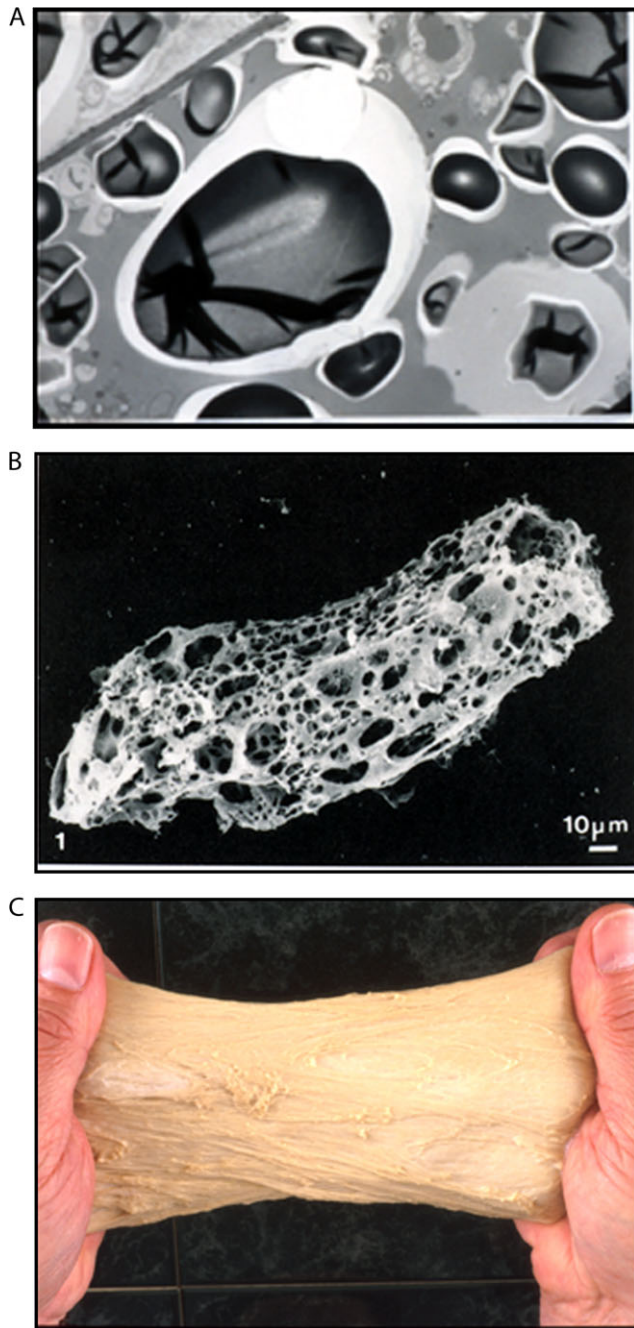
Gluten was one of the earliest protein fractions to be described by chemists, being first described by Beccari in 1728 (see translation by Bailey, 1941). It is traditionally prepared by gently washing wheat dough in water or dilute salt solution, leaving a cohesive mass which comprises about 80% protein, the remainder being mainly starch granules which are trapped in the protein matrix.

The ability to prepare the gluten proteins in an essentially pure state by such a simple procedure depends on their unusual properties. Firstly, they are insoluble in water or dilute salt solutions but are soluble in alcohol/water mixtures (as discussed below) and were hence defined as 'prolamins' by TB Osborne in his classic studies of plant proteins carried out at the end of the 19th century and the start of the 20th century (Osborne, 1924). Secondly, the individual gluten proteins are associated by strong covalent and non-covalent forces which allow the whole fraction to be isolated as a cohesive mass.

### What is the origin of gluten?

In common with other seed storage proteins, the gluten proteins are secretory proteins, being synthesized on the rough endoplasmic reticulum and co-translationally transported into the lumen of the ER. Once within the ER lumen, cereal seed storage proteins may follow two routes: a Golgi-dependent route leading to deposition within protein bodies of vacuolar origin or a Golgi-independent route in which protein deposits formed within the ER lumen may ultimately fuse with protein bodies of vacuolar origin (see Kumamaru *et al.*, 2007, for a review).

Work carried out by Galili and colleagues (Levanany *et al.*, 1992; Galili *et al.*, 1995; Galili, 1997) indicated that wheat gluten proteins may follow both routes, and this has recently been confirmed using epitope tags and specific antibodies to follow individual proteins and groups of proteins in cells of developing grain (Tosi *et al.*, 2009). It is also clear that the protein deposits fuse to form a continuous matrix as the cells of the starchy endosperm dry and die during the later stages of grain maturation (Fig. 2A). Thus



**Fig. 2.** The origin of wheat gluten. (A) Transmission electron microscopy of the developing starchy endosperm cells at 46 d after anthesis shows that the individual protein bodies have fused to form a continuous proteinaceous matrix. Taken from Shewry *et al.*, 1995, (*Biotechnology* **13**, 1185–1190) and provided by Dr M Parker (IFR, Norwich, UK). (B) Digestion of a flour particle with amylases to remove starch reveals a continuous proteinaceous network. Taken from Amend and Beauvais (1995) and reproduced by kind permission of Getreidetechnologie. (C) After kneading, dough can be washed to recover the gluten network as a cohesive mass which is stretched in the photograph to demonstrate its viscoelastic properties.

a proteinaceous network is present in each endosperm cell (Fig. 2B) and these networks are brought together when flour is mixed with water to form a continuous network in the dough. Washing the dough to remove non-gluten components therefore allows the network to be recovered as the cohesive mass which is called gluten (Fig. 2C).

#### *The biochemical and molecular basis for gluten functionality*

Humankind has been aware for many centuries that wheat dough has unusual properties which are shared to a limited extent by doughs made from rye flour but not by those from other cereal flours. These properties, which are usually described as ‘viscoelasticity’, are particularly important in making leavened bread, as they allow the entrapment of carbon dioxide released during leavening. However, they also underpin a range of other uses including making unleavened breads, cakes, and biscuits, pasta (from durum wheat), and noodles (from bread wheat). They are also exploited in the food industry where gluten proteins may be used as a binder in processed foods.

The volume of research carried out on wheat gluten is vast, with a simple search of the Web of Science database showing almost 20 000 papers since 1945. This volume not only reflects the commercial importance of wheat processing, but also the complexity of the system which remains incompletely understood. They include studies at the genetic, biochemical, biophysical, and functional (ie processing) levels.

Genetic studies have exploited the extensive polymorphism which exists between the gluten protein fractions present in different genotypes to establish genetic linkages between either groups of gluten proteins, or allelic forms of these, and aspects of processing quality. Similarly, studies at the biochemical and biophysical levels have demonstrated a relationship between dough strength and the ability of the gluten proteins to form polymeric complexes (called glutenins). Combining results from these two approaches highlighted the importance of a specific group of gluten proteins, called the high molecular weight (HMW) subunits of glutenin.

Cultivars of bread wheat express between three and five HMW subunit genes, with the encoded proteins accounting for up to about 12% of the total grain protein (Seilmeier *et al.*, 1991; Halford *et al.*, 1992). The HMW subunits are only present in high molecular mass polymers and allelic variation in both the number of expressed genes and the properties of the encoded proteins results in effects on the amount and size of the polymers and hence dough strength (reviewed by Payne, 1987; Shewry *et al.*, 2003b). These glutenin polymers are known to be stabilized by inter-chain disulphide bonds, but it is apparent that non-covalent hydrogen bonds are also important in stabilizing the interactions between glutenin polymers and monomeric gluten proteins (called gliadins) (Belton, 2005). Hence, the individual gliadins and glutenin polymers can be separated using solvents which disrupt hydrogen bonding (such as

urea) but reducing agents (such as 2-mercaptoethanol or dithiothreitol) are required to break down the glutenin polymers to release the individual subunits.

Although the HMW subunits are the main determinants of glutenin elasticity relationships between other gluten proteins and functional properties have also been reported (reviewed by Shewry *et al.*, 2003a).

The relationship between the HMW subunits and dough strength was first established over 25 years ago (Payne *et al.*, 1979) and allelic forms associated with good processing quality have been selected by plant breeders for over two decades, using simple SDS-PAGE separations. The established relationships between the number of expressed HMW subunit genes, the total amount of HMW subunit protein and dough strength have also resulted in the HMW subunit genes being an attractive target for genetic transformation, in order to increase their gene copy number and hence dough strength.

The first studies of this type were reported over 10 years ago (Altpeter *et al.*, 1996; Blechl and Anderson, 1996; Barro *et al.*, 1997) and many studies have since been reported (reviewed by Shewry and Jones, 2005; Jones *et al.*, 2009). It is perhaps not surprising that the results have been 'mixed', but some conclusions can be drawn. Firstly, expression of an additional HMW subunit gene can lead to increased dough strength, even when a modern good quality wheat cultivar is used as the recipient (see Field *et al.*, 2008; Rakszegi *et al.*, 2008, as recent examples, and reviews of earlier work cited above). However, the effect depends on the precise HMW subunit gene which is used and on the expression level, with the transgenes resulting in over-strong (ie too elastic) gluten properties in some studies. Thus, although transgenesis is a realistic strategy to increase dough strength in wheat, it is also necessary to have an understanding of the underlying mechanisms in order to optimize the experimental design.

## Wheat in nutrition and health

Wheat is widely consumed by humans, in the countries of primary production (which number over 100 in the FAO production statistics for 2004) and in other countries where wheat cannot be grown. For example, imported wheat is used to meet consumer demands for bread and other food products in the humid tropics, particularly those with a culinary tradition dating back to colonial occupation. Statistics are not available for the total volume of wheat which is consumed directly by humans as opposed to feeding livestock, although figures for the UK indicate about one-third of the total production (approximately 5.7 m tonnes per annum are milled with home production being 15–16 m tonnes). Globally there is no doubt that the number of people who rely on wheat for a substantial part of their diet amounts to several billions.

The high content of starch, about 60–70% of the whole grain and 65–75% of white flour, means that wheat is often considered to be little more than a source of calories, and this

is certainly true for animal feed production, with high-yielding, low-protein feed varieties being supplemented by other protein-rich crops (notably soybeans and oilseed residues).

However, despite its relatively low protein content (usually 8–15%) wheat still provides as much protein for human and livestock nutrition as the total soybean crop, estimated at about 60 m tonnes per annum (calculated by Shewry, 2000). Therefore, the nutritional importance of wheat proteins should not be underestimated, particularly in less developed countries where bread, noodles and other products (eg bulgar, couscous) may provide a substantial proportion of the diet.

## Protein content

Although wheat breeders routinely select for protein content in their breeding programmes (high protein for breadmaking and low protein for feed and other uses), the current range of variation in this parameter in commercial cultivars is limited. For example, Snape *et al.* (1993) estimated that typical UK breadmaking and feed wheats differed in their protein content by about 2% dry weight (eg from about 12–14% protein) when grown under the same conditions, which is significantly less than the 2-fold differences which can result from high and low levels of nitrogen fertilizer application. This limited variation in conventional wheat lines has led to searches for 'high protein genes' in more exotic germplasm.

Early studies of the USDA World Wheat Collection showed approximately 3-fold variation in protein content (from 7–22%), with about one-third of this being under genetic control (Vogel *et al.*, 1978). However, the strong environmental impact on protein content (accounting for two-thirds of the variation) underpins the difficulty of breeding for this trait. Nevertheless, some success has been achieved by incorporating sources of variation from exotic bread wheat lines or related wild species.

The former include Atlas 50 and Atlas 66, derived from the South American line Frandoso, and Nap Hal from India. These lines appear to have different 'high protein genes' and both were extensively used in breeding programmes in Nebraska with the Atlas 66 gene being successfully incorporated into the commercial variety Lancota (Johnson *et al.*, 1985). Frandoso and related Brazilian lines have also been successfully exploited in other breeding programmes in the USA (Busch and Rauch, 2001). The Kansas variety, Plainsman V, also contained a high protein gene(s) from a related *Aegilops* species (Finney, 1978).

The most widely studied source of 'high protein' is wild emmer (tetraploid *Tr. turgidum* var. *dicoccoides*) wheats from Israel. One accession, FA15-3, accumulates over 40% of protein when grown with sufficient nitrogen (Avivi, 1978). The gene in this line was mapped to a locus on chromosome 6B (called *Gpc-B1*), which accounted for about 70% of the variation in protein content in crosses (Chee *et al.*, 2001; Distelfeld *et al.*, 2004, 2006). More recent studies have shown that the gene *Gpc-B1* encodes a transcription factor which accelerates senescence in the

vegetative parts of the plant, resulting in increased mobilization and transfer to the grain of both nitrogen and minerals (notably iron and zinc) (Uauy *et al.*, 2006). However, it remains to be shown whether this gene can be incorporated into high-yielding and commercially viable lines.

### Protein composition

Of the 20 amino acids commonly present in proteins, 10 can be considered to be essential in that they cannot be synthesized by animals and must be provided in the diet. Furthermore, if only one of these is limiting the others will be broken down and excreted. There has been much debate about which amino acids are essential and the amounts that are required, with the most recent values for adult humans being shown in Table 1. This table includes a combined value for the two aromatic amino acids, tyrosine and phenylalanine, which are biosynthetically related, and both single and combined values for the two sulphur-containing amino acids: methionine, which is truly essential, and cysteine which can be synthesized from methionine. Comparison with the values for whole wheat grain and flour shows that only lysine is deficient, with some essential amino acids being present in considerably higher amounts than the requirements. However, the lysine content of wheat also varies significantly with the values shown in Table 1 being typical of grain of high protein content and the proportion increasing to over 30 mg g<sup>-1</sup> protein in low protein grain (Mossé and Huet, 1990). This decrease in the relative lysine content of high protein grain results from proportional increases in the lysine-poor gluten proteins when excess N is available (for example, when fertilizer is applied to increase grain yield and protein content) and also accounts for the lower lysine content of the white flour (the gluten proteins being located in the starchy endosperm tissue).

**Table 1.** Recommended levels of essential amino acids for adult humans compared with those in wheat grain and flour (expressed as mg g<sup>-1</sup> protein)

Amino acid protein	FAO/WHO/UNU <sup>a</sup>	Wheat	
		Grain <sup>b</sup>	Flour <sup>b</sup>
Histidine	15	23	22
Isoleucine	30	37	36
Leucine	59	68	67
Lysine	45	28	22
Methionine+cysteine	22	35	38
methionine	16	12	13
cysteine	6	23	25
Phenylalanine+tyrosine	38	64	63
Threonine	23	29	26
Tryptophan	6	11	11
Valine	39	44	41
Total indispensable amino acids	277	339	326

<sup>a</sup> FAO/WHO/UNU (2007).

<sup>b</sup> Calculated from literature values as described in Shewry (2007).

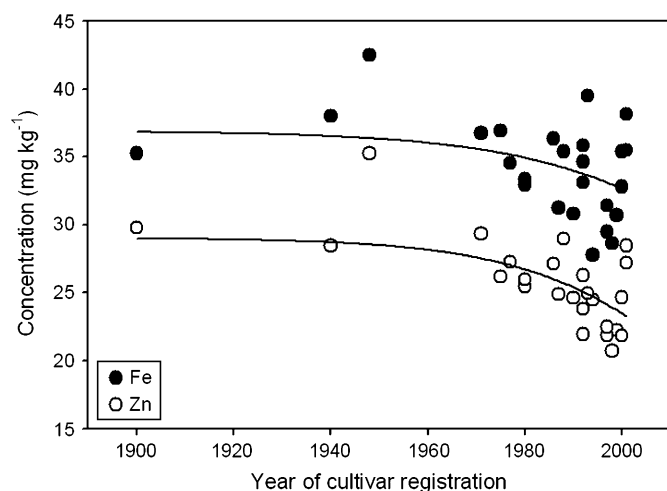
The amino acid requirements for infants and children vary depending on their growth rate, being particularly high in the first year of life. Similarly, higher levels of essential amino acids are required for rapidly growing livestock such as pigs and poultry.

### Wheat as a source of minerals

Iron deficiency is the most widespread nutrient deficiency in the world, estimated to affect over 2 billion people (Stoltzfus and Dreyfuss, 1998). Although many of these people live in less developed countries, it is also a significant problem in the developed world. Zinc deficiency is also widespread, particularly in Sub-Saharan Africa and South Asia, and has been estimated to account for 800 000 child deaths a year (Micronutrient Initiative, 2006), in addition to non-lethal effects on children and adults. Wheat and other cereals are significant sources of both of these minerals, contributing 44% of the daily intake of iron (15% in bread) and 25% of the daily intake of zinc (11% in bread) in the UK (Henderson *et al.*, 2007). There has therefore been considerable concern over the suggestion that the mineral content of modern wheat varieties is lower than that of older varieties.

This was initially suggested by Garvin *et al.* (2006) who grew 14 red winter wheat cultivars bred between 1873 and 2000 in replicate field experiments and determined their mineral contents. Plants were grown at two locations in Kansas and significant negative correlations were found between grain yield, variety release date, and the concentrations of zinc in material from both of these sites and of iron in materials from only one site. Similar trends were reported by Fan *et al.* (2008a, b) who took a different approach. Rather than carrying out direct comparisons of varieties in field trials, they analysed grain grown on the Rothamsted Broadbalk long-term wheat experiment. This experiment was established in 1843 and uses a single variety which is replaced by a more modern variety at regular intervals. Analysis of archived grain showed significant decreases in the contents of minerals (Zn, Fe, Cu, Mg) since semi-dwarf cultivars were introduced in 1968. A similar difference was observed between the cultivars Brimstone (semi-dwarf) and Squareheads Master (long straw) which were grown side by side in 1988–1990, the concentrations of Zn, Cu, Fe, and Mg being 18–29% lower in Brimstone. A more recent comparison of 25 lines grown also showed a decline in the concentrations of Fe and Zn since semi-dwarf wheats were introduced (Zhao *et al.*, 2009) (Fig. 3). Although the decrease in the mineral content of modern wheats is partly due to dilution, resulting from increased yield (which was negatively correlated with mineral content), it has been suggested that short-strawed varieties may be intrinsically less efficient at partitioning minerals to the grain compared with the translocation of photosynthate.

Such genetic differences in mineral content are clearly relevant to international efforts to increase the mineral content of wheat to improve health in less developed countries. Thus, increasing iron, zinc, and vitamin A



**Fig. 3.** The relationship between the iron content of wholemeal flours from 25 wheat cultivars grown on six trial sites/seasons and their release dates. Taken from Zhao *et al.* (2009) and reproduced by kind permission of Elsevier.

contents are a major focus of the HarvestPlus initiative of the Consultative Group on International Agricultural Research (CGIAR) which is using conventional plant breeding (Ortiz-Monasterio *et al.*, 2007) while other laboratories are using genetic engineering approaches (reviewed by Brinch-Pedersen *et al.*, 2007).

These initiatives are focusing not only on contents of minerals but also on their bioavailability. Iron is predominantly located in the aleurone and as complexes with phytate (*myo*-inositolphosphate 1,2,3,4,5,6-hexa-kisphosphate). These complexes are largely insoluble, restricting mineral availability to humans and livestock. The use of transgenesis to express phytase in the developing grain can result in increased mineral availability, particularly when a heat-stable form of the enzyme is used to allow hydrolysis to occur during food processing (reviewed by Brinch-Pedersen *et al.*, 2007).

Guttieri *et al.* (2004) also reported an EMS-induced low phytate mutant of wheat. This mutation resulted in 43% less phytic acid in the aleurone, but has not so far been incorporated into commercial cultivars. However, previous experience with low phytic acid mutants of maize, barley, and soy bean has shown that they may also have significant effects on yield and germination rates (reviewed by Brinch-Pedersen *et al.*, 2007).

#### *Wheat as a source of selenium*

Selenium is an essential micronutrient for mammals (but not plants), being present as selenocysteine in a number of enzymes. However, it is also toxic when present in excess (above about  $600 \mu\text{g d}^{-1}$ ; Yang and Xia, 1995). Cereals are major dietary sources of selenium in many parts of the world, including China (FAO/WHO, 2001), Russia (Golubkina and Alfthan, 1999), and the UK (MAFF, 1997). However, the content of selenium in wheat varies widely

from about  $10 \mu\text{g kg}^{-1}$  to over  $2000 \mu\text{g kg}^{-1}$  (FAO/WHO, 2001; Combs, 2001).

The concentration of selenium in wheat is largely determined by the availability of the element in the soil. Consequently, wheat produced in Western Europe may contain only one-tenth of the selenium that is present in wheat grown in North America. Thus, a survey of 452 grain samples grown in the UK in 1982 and 1992 showed a mean value of  $27 \mu\text{g Se kg}^{-1}$  fresh weight (Adams *et al.*, 2002) compared with  $370 \mu\text{g SE kg}^{-1}$  fresh weight for 290 samples from the USA (Wolnik *et al.*, 1983).

Because the import of wheat from North America into Europe has declined over the last 25 years, the intake of selenium in the diet has also decreased, which has resulted in concern in some European countries. One response to this is to apply selenium to the crops in fertilizer (called biofortification), which is practised in Finland (Eurola *et al.*, 1991).

Unlike iron, selenium is not concentrated in the aleurone, being present wherever sulphur is present. The concentration of selenium in grain from the Broadbalk continuous wheat experiment also appeared to be determined principally by the sulphur availability in the soil (which competes to prevent selenium uptake), with no evidence of decreased levels over time (Fan *et al.*, 2008b). However, sulphur fertilizer is often applied to wheat to improve the grain quality (Zhao *et al.*, 1997) and this could clearly have negative impacts on selenium in grain.

The reader is referred to a recent review article by Hawkesford and Zhao (2007) for a detailed review of selenium in wheat.

#### *Wholegrain wheat and health*

The consumption of white flour and bread have historically been associated with prosperity and the development of sophisticated roller mills in Austro-Hungary during the second part of the 19th century allowed the production of higher volumes of whiter flour than it was possible to produce by traditional milling based on grinding between stones and sieving (see Jones, 2007, for a fascinating account of the history of roller milling). However, the increased consumption of bread made from highly refined white flour was not accepted universally, leading to what we would today recognize as a movement to increase the consumption of wholegrain products.

In 1880, May Yates founded the Bread Reform League in London to promote a return to wholemeal bread, particularly to improve the nutrition of the children of the poor, and suggested in 1909 that an official minimum standard of 80% flour extraction rate should be adopted. This was called 'Standard Bread'. Although we now appreciate the nutritional advantages of wholegrain products, this was not supported by the science of the time and clearly conflicted with the tastes of consumers as well as the economics of bread production. Nevertheless, the League continued to campaign and received scientific support in 1911 when Gowland Hopkins agreed that 'Standard Bread' may contain 'unrecognized food

substances' which were vital for health: these were subsequently called vitamins (Burnett, 2005).

By contrast, Thomas Allinson (1858–1918) had a much greater impact by marketing and vigorously promoting his own range of wholemeal products. He can therefore be regarded as the father of the wholegrain movement and remains a household name to this day in the UK (Pepper, 1992).

We now know that wholegrain wheat products contain a range of components with established or proposed health benefits which are concentrated or solely located in the bran. Hence they are either present in lower amounts or absent from white flour which is derived almost exclusively from starchy endosperm cells. They vary widely in their concentrations. For example, lignans, a group of polyphenols with phytoestrogen activity, are present at levels up to about  $10 \mu\text{g g}^{-1}$  in wholemeal wheat and twice this level in bran (Nagy-Scholz and Ercsey, 2009), while total phenolic acids in wholemeal range up to almost  $1200 \mu\text{g g}^{-1}$  (Li *et al.*, 2008).

The most detailed study of wheat phytochemicals which has so far been reported was carried out as part of the EU Framework 6 HEALTHGRAIN programme (Poutanen *et al.*, 2008; Ward *et al.*, 2008). This study determined a range of phytochemicals in 150 wheat lines grown on a single site in one year, meaning that the levels of the components may have been influenced by environmental as well as genetic effects. The lines were selected to represent a broad range of dates and places of origin. The choice of phytochemicals focused on those which have putative health benefits and for which cereals are recognized dietary sources. For example, cereals are considered to account for about 22% of the daily intake of folate (vitamin B12) in the UK (Goldberg, 2003) and 36% and 43% of the daily intake in Finnish women and men, respectively (Findiet Study Group, 2003). In the HEALTHGRAIN study the contents of folates in wholemeal varied from 364 to 774  $\text{ng g}^{-1}$  dry weight in 130 winter wheats and from 323 to 741  $\text{ng g}^{-1}$  dry weight in 20 spring wheats, with the content in the former being positively correlated with bran yield and negatively correlated with seed weight (indicating concentration in the bran) (Piironen *et al.*, 2008).

The quantitatively major group of phytochemicals in the wheat grain is phenolic acids, derivatives of either hydroxybenzoic acid or hydroxycinnamic acid. Epidemiological studies indicate that phenolic acids have a number of health benefits which may relate to their antioxidant activity; the total antioxidant activities of grain extracts and their phenolic acid contents being highly correlated (Drankham *et al.*, 2003; Beta *et al.*, 2005; Wende *et al.*, 2005).

Cereals are also significant sources of tocopherols (which include vitamin E) ( $27.6\text{--}79.7 \mu\text{g g}^{-1}$  in the HEALTHGRAIN study) (Lampi *et al.*, 2008) and sterols ( $670\text{--}959 \mu\text{g g}^{-1}$ ) (Nurmi *et al.*, 2008).

The HEALTHGRAIN study also determined the levels of dietary fibre. In wheat, this mainly derives from cell wall polymers: arabinoxylans (approximately 70%) with lower amounts of (1-3)(1-4) $\beta$ -D-glucans (approximately 20%) and other components. The arabinoxylans also occur in soluble

and insoluble forms, with the latter being rich in bound phenolic acids which form oxidative cross-links. These bound phenolic acids account, on average, for 77% of the total phenolic acid fraction and are predominantly ferulic acid. Soluble fibre is considered to have health benefits (Moore *et al.*, 1998; Lewis and Heaton, 1999) which are not shared by insoluble fibre and these may therefore be reduced by the phenolic acid cross-linking. However, insoluble fibre may also have benefits in delivering phenolic antioxidants into the colon: these benefits may include reduction in colo-rectal cancer (Vitaglione *et al.*, 2008).

The HEALTHGRAIN study showed wide variation in the contents of total and water-extractable arabinoxylans in both white flour and bran fractions (Gebruers *et al.*, 2008) (Fig. 4). Similarly, Ordaz-Ortiz *et al.* (2005) showed variation from 0.26% to 0.75% dry weight in the content of water-extractable arabinoxylan in 20 French wheat lines and from 1.66% to 2.87% dry weight in total arabinoxylans. A high proportion of the variation in water-extractable arabinoxylans is also heritable (Martinant *et al.*, 1999).

It is clear from these and other studies that there is sufficient genetically determined variation in the phytochemical and fibre contents of wheat to be exploited in breeding for varieties with increased nutritional benefits.

## Adverse reactions to wheat

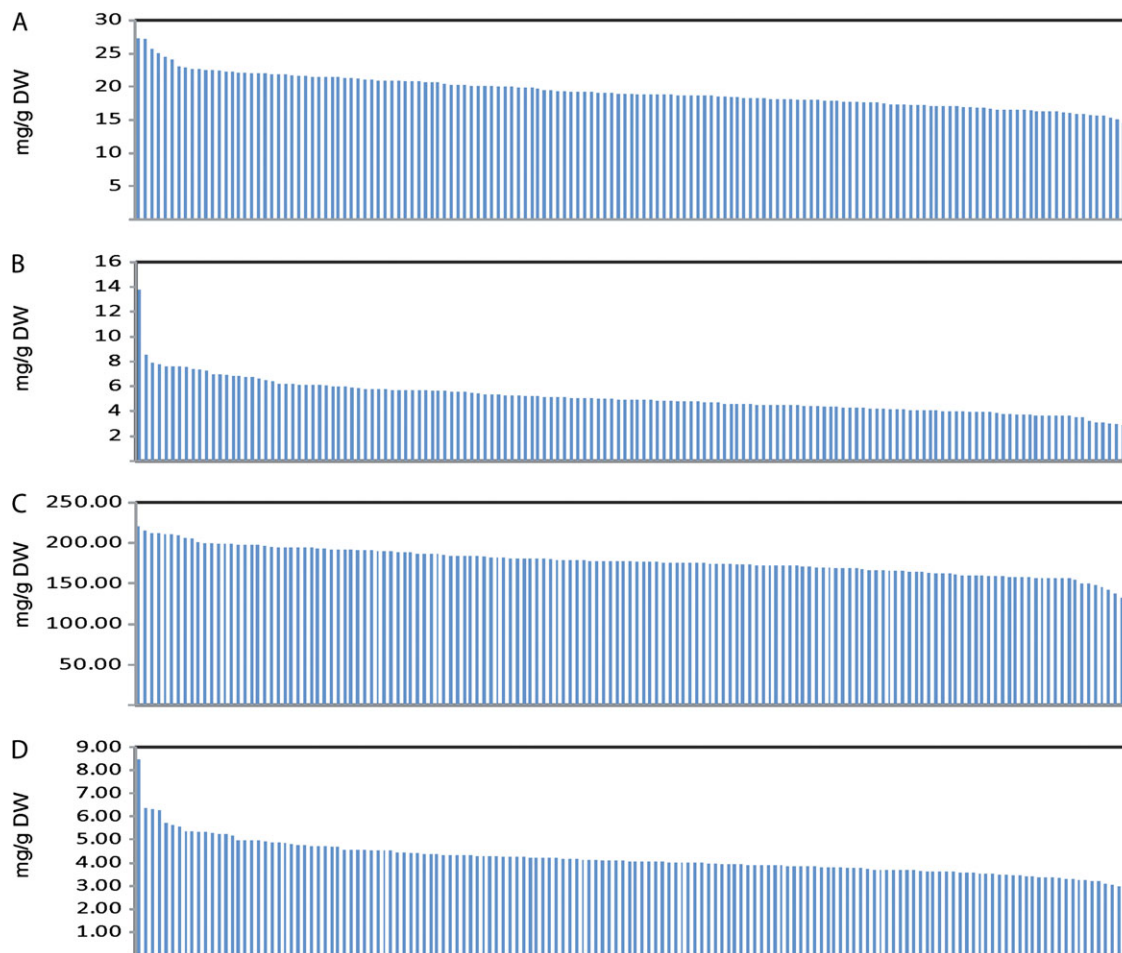
### Allergy to wheat

Both respiratory and food allergies to wheat have been reported.

Respiratory allergy (bakers' asthma) has been known since Roman times (when slaves handling flour and dough were required to wear masks) and is currently one of the most important forms of occupational allergy. For example, it is the second most widespread occupational allergy in the UK and has been reported to affect over 8% of apprentice bakers in Poland after only 2 years exposure (Walusiak *et al.*, 2004). A wide range of wheat grain proteins have been shown to react with immunoglobulin (Ig)E in sera of patients with bakers' asthma, including gliadins, glutenins, serpins (serine proteinase inhibitors), thioredoxin, agglutinin, and a number of enzymes ( $\alpha$ - and  $\beta$ -amylases, peroxidase, acyl CoA oxidase, glyceraldehyde-3-phosphate dehydrogenase and triosephosphate isomerase) (reviewed by Tatham and Shewry, 2008). However, it is clear that the predominant wheat proteins responsible for bakers' asthma are a class of  $\alpha$ -amylase inhibitors, also known as CM proteins due to their solubility in chloroform:methanol mixtures (Salcedo *et al.*, 2004). Furthermore, their activity has been demonstrated by a range of approaches including skin pricks and RAST (radioallergosorbent test) as well as immunoblotting, ELISA, and screening expression libraries with IgE fractions.

The CM proteins comprise monomeric, dimeric, and tetrameric forms, with subunit masses ranging between about 10 000 and 16 000. They differ in their spectrum of





**Fig. 4.** Contents of arabinoxylan (AX) fibre in flour and bran of 150 wheat cultivars grown on a single site as part of the EU FP6 HEALTHGRAIN project. (A) Total AX in flour ( $\text{mg g}^{-1}$ ); (B) water-extractable AX in flour (%); (C) total AX in bran ( $\text{mg g}^{-1}$ ), and (D) water-extractable AX in bran. Prepared from data reported by Gebruers *et al.* (2008) with permission of the authors.

activity but all inhibit mammalian and insect  $\alpha$ -amylases (including those in some pest organisms) rather than endogenous wheat enzymes. Hence, they are considered to be protective rather than regulatory in function. Eleven individual subunits have been shown to play a role in bakers' asthma (using one or more of the assays listed above) but they differ in their activity, with a glycosylated form of CM16 being particularly active.

Wheat is listed among the 'big eight' food allergens which together account for about 90% of all allergic responses. However, the incidence of true (ie IgE-mediated) food allergy is, in fact, fairly infrequent in adults, although it may affect up to 1% of children (Poole *et al.*, 2006). A number of wheat proteins have been reported to be responsible for allergic responses to the ingestion of wheat products but only one syndrome has been studied in detail. Wheat-dependent exercise-induced anaphylaxis (WDEIA) is a well-defined syndrome in which the ingestion of a product containing wheat followed by physical exercise can result in an anaphylactic response. Work carried out by several groups has clearly established that this condition is associ-

ated with a group of  $\omega$ -gliadins (called  $\omega$ 5-gliadins) which are encoded by genes on chromosome 1B (Palosuo *et al.*, 2001; Morita *et al.*, 2003; Battais *et al.*, 2005). Mutational analysis has also identified immunodominant epitopes in the  $\omega$ 5-gliadins: short glutamine-rich and proline-rich sequences present in the repetitive domains of the proteins (Matsuo *et al.*, 2004, 2005; Battais *et al.*, 2005). However, a number of other proteins have also been shown to react with IgE from patients with WDEIA, including gliadins, glutenin subunits, and related proteins from barley and rye (reviewed by Tatham and Shewry, 2008).

Other allergic responses to wheat proteins include atopic dermatitis, urticaria, and anaphylaxis. Not surprisingly, these symptoms have been associated with a number of wheat proteins, most notably gluten proteins but also CM proteins, enzymes, and lipid transfer protein (LTP) (reviewed by Tatham and Shewry, 2008).

Comparison of the proteins identified as responsible for the respiratory and food allergy shows significant overlap in their functions (most being storage or protective) and identities (notably gluten proteins and CM proteins).

*Intolerance to wheat*

Dietary intolerance to wheat is almost certainly more widespread than allergy, notably coeliac disease (CD) which is estimated to affect 1% of the population of Western Europe (Feighery, 1999), and dermatitis herpetiformis which has an incidence between about 2-fold and 5-fold lower than CD (Fry, 1992).

CD is a chronic inflammation of the bowel which leads to malabsorption of nutrients. Like bakers' asthma, CD has been known since classical times but it was only defined in detail in 1887 and its relationship to wheat established by Dicke in the late 1940s (Losowsky, 2008).

A series of elegant studies carried out over the past decade, particularly by Sollid, Koning and co-workers, have established that CD results from an autoimmune response which is triggered by the binding of gluten peptides to T cells of the immune system in some (but not all) individuals with the human leucocyte antigens (HLAs) DQ2 or DQ8, expressed by specialized antigen-presenting cells. The presented peptides are then recognized by specific CD4<sup>+</sup> T cells which release inflammatory cytokines which lead to the flattening of the intestinal epithelium. It has also been demonstrated that tissue transglutaminase enzyme present in the epithelium of the intestine plays an important role, generating toxic peptides by deamidation of glutamine residues to give glutamate.

The HLA-DQ2 antigen is present in about 95% of coeliac patients (Karell *et al.*, 2003) and detailed studies have identified the peptide sequences which are recognized by intestinal T cell lines, using either peptide fractions produced from gluten proteins or synthetic peptides. This has led to the definition of two overlapping immunodominant epitopes corresponding to residues 57–68 ( $\alpha$ -9) and 62–75 ( $\alpha$ -2) of A gliadin (a form of  $\alpha$ -gliadin) (Arentz-Hansen *et al.*, 2000, 2002; Anderson *et al.*, 2000; Ellis *et al.*, 2003). Related epitopes were similarly defined in  $\gamma$ -gliadins, corresponding to residues 60–79, 102–113, 115–123, and 228–236 (Sjöström *et al.*, 1998; Arentz-Hansen *et al.*, 2002; Vader *et al.*, 2002a). Furthermore, Vader *et al.* (2002b) showed that the spacing between glutamine and proline residues determined the specificity of glutamine deamidation and hence peptide activation, and developed algorithms to predict the presence of novel T cell stimulatory peptides in gluten proteins and in related proteins from other cereals.

Less work has been carried out on the determinants of the HLA-DQ8 associated coeliac disease, which affects only about 6% of patients without HLA-DQ2 and 10% of patients with HLA-DQ2 (Karell *et al.*, 2003). This has again allowed immunodominant epitopes to be identified in gliadins and glutenin subunits (van der Wal *et al.*, 1998, 1999; Mazzarella *et al.*, 2003; Tollefsen *et al.*, 2006) although detailed structural studies indicate that the HLA-DQ2 and HLA-DQ8-mediated forms of the disease may differ in their molecular mechanisms (Henderson *et al.*, 2007).

The possibility of producing wheat which lacks the coeliac toxic peptides has been discussed for many years

but interest in the strategy tended to decline as it became clear that most, if not all, gluten proteins are toxic to at least some susceptible individuals, rather than only the  $\alpha$ -gliadins as initially thought. However, Spaenij-Dekking *et al.* (2005) and van Herpen *et al.* (2006) have shown that it is possible to identify natural forms of gliadin which have few or no coeliac toxic epitopes, raising the possibility of selecting for less toxic lines of wheat by classical plant breeding. RNA interference (RNAi) technology has also been used to silence the  $\alpha$ -gliadin (Becker *et al.*, 2006; Wieser *et al.*, 2006) and  $\gamma$ -gliadin (Gil-Humanes *et al.*, 2008) gene families, although some effects on grain-processing properties were observed.

The combination of these two approaches may therefore allow the production of less toxic, if not non-toxic, wheat for coeliac patients without significant loss of the processing properties conferred by the gluten proteins.

Dermatitis herpetiformis is a skin eruption resulting from ingestion of gluten, and is associated with the deposition of IgA antibodies in dermal papillae. These include IgA antibodies to epidermal transglutaminase which is considered to be an important autoantigen in disease development (Hull *et al.*, 2008).

*Other medical conditions related to gluten proteins*

There are many reports of the association of wheat, and particularly wheat proteins, with medical conditions, ranging from improbable reports in the popular press to scientific studies in the medical literature. Not surprisingly, they include autoimmune diseases such as rheumatoid arthritis which may be more prevalent in coeliac patients and relatives (Neuhausen *et al.*, 2008). It is perhaps easier to envisage mechanisms for relationships between such diseases which have a common immunological basis (Hvatum *et al.*, 2006) than to explain a well-established association between wheat, coeliac disease, and schizophrenia (Singh and Roy, 1975; Kalaydian *et al.*, 2006). Other reported associations include ones with sporadic idiopathic ataxia (gluten ataxia) (Hadjivassiliou *et al.*, 2003), migraines (Grant, 1979), acute psychoses (Rix *et al.*, 1985), and a range of neurological illnesses (Hadjivassiliou *et al.*, 2002). An association with autism has also been reported (Lucarelli *et al.*, 1995) with some physicians recommending a gluten-free, casein-free diet (Elder, 2008).

Some of these effects may be mediated via the immune system but effects which are not immune-mediated are notoriously difficult to define and diagnose. However, they could result from the release within the body of bioactive peptides, derived particularly from gluten protein. Thus, gluten has been reported to be a source of a range of such peptides including opioid peptides (exorphins) (Takahashi *et al.*, 2000; Yoshikawa *et al.*, 2003) and an inhibitor of angiotensin I-converting enzyme (Motoi and Kodama, 2003) (see also reviews by Dziuba *et al.*, 1999; Yamamoto *et al.*, 2003). However, these activities were demonstrated *in vitro* and their *in vivo* significance has not been established.

## The future for wheat

There is little doubt that wheat will retain its dominant position in UK and European agriculture due to its adaptability and consumer acceptance. However, it may also need to adapt to face changing requirements from farmers, food processors, governments, and consumers.

### Reducing inputs

Currently grown wheat cultivars require high inputs of nitrogen fertilizer and agrochemicals to achieve high yields combined with the protein content required for breadmaking. For example, UK farmers currently apply 250–300 kg N ha<sup>-1</sup> in order to achieve the 13% protein content required for the Chorleywood Breadmaking Process, which is the major process used for breadmaking in the UK. Since a 10 tonnes ha<sup>-1</sup> crop containing 13% protein equates to about 230 kg N ha<sup>-1</sup>, this means that 50–70 kg N ha<sup>-1</sup> may be lost. As fertilizer N currently costs about £1 kg<sup>-1</sup> this represents a significant financial loss as well as a loss of the energy required for fertilizer production and may also have environmental consequences.

A number of projects worldwide are therefore focusing on understanding the processes that determine the efficiency of uptake, assimilation, and utilization of nitrogen in order to improve the efficiency of nitrogen recovery in the grain (reviewed by Foulkes *et al.*, 2009).

Reducing the nitrogen requirement of wheat does not only relate to the grain protein content, as an adequate supply of nitrogen is also essential for high wheat yields in order to build a canopy and fix carbon dioxide by photosynthesis. Furthermore, a substantial proportion of this nitrogen is remobilized and redistributed to the developing grain during canopy senescence (Dalling, 1985). Hawkesford and colleagues at Rothamsted Research have targeted this process in order to develop a strategy for improving the recovery of N in the grain, using a combination of biochemical analysis and metabolite and transcript profiling to identify differences in metabolites and gene expression which are associated with efficient mobilization and redistribution (Howarth *et al.*, 2008). Some of the genes identified in these and similar studies are suitable candidates for manipulation to increase the proportion of the total nitrogen recovered in the grain.

### Stability of quality

The increases in temperature and carbon dioxide concentration associated with climate change are expected to have effects on crop development and yield, although the magnitude of these is difficult to predict due to interactions with other factors which may also be affected, notably water availability and populations of pests and pathogens (Coakley *et al.*, 1999; Semenov, 2008). Similarly, although it is generally accepted that higher growth temperatures result in greater dough strength, the precise effects are not clearly understood (see review by Dupont and Altenbach, 2003) with heat stress (ie above 30–33 °C) actually resulting in

dough weakening and reduced quality (Blumenthal *et al.*, 1993). A recent review of the effects of CO<sub>2</sub> concentration on grain quality also failed to draw clear conclusions (Högy and Fangmeier, 2008).

Of more immediate interest to wheat breeders and grain-utilizing industries are year-to-year fluctuations in growth conditions, and the frequency and magnitude of such fluctuations are also predicted to increase in the future (Porter and Semenov, 2005). Although some cultivars are generally considered to be more consistent in quality than others, this is largely anecdotal with no detailed scientific comparisons.

Given the recent advances in ‘omics’ technologies it should now be possible to dissect the effects of G×E on grain development and quality, and to establish markers suitable for use in plant breeding. However, this will require substantial resources and a multi-disciplinary approach: by growing mapped populations and lines in multi-site/multi-year trials and determining aspects of composition and quality from gene expression profiling to pilot scale breadmaking.

Wan *et al.* (2009) have reported the application of this approach using a limited set of seven doubled haploid lines to identify a number of transcripts whose expression profile was associated with quality traits independently of environmental conditions. Millar *et al.* (2008) also reported a larger study in which three doubled haploid populations of wheat were used to map novel QTLs (quantitative trait loci) for breadmaking and pastry making which were stable over two years field trials, but did not relate quality traits to gene expression profiles.

### Biofuels

Wheat is an attractive option as a ‘first generation biofuel’ as the high content of starch is readily converted into sugars (saccharification) which can then be fermented into ethanol. Murphy and Power (2008) recently reported that the gross energy recovered in ethanol using wheat was 66 GJ ha<sup>-1</sup> a<sup>-1</sup>, but that this only corresponds to 50% energy conversion and that the net energy production is as low as 25 GJ ha<sup>-1</sup> a<sup>-1</sup>. The same authors also calculated that the net energy production could be increased to 72 GJ ha<sup>-1</sup> a<sup>-1</sup> if the straw was combusted and the residue after distillation, called stillage or distillers grain and solubles (DGS), was converted to biogas (biomethane).

A major concern about using wheat grain for biofuel production is the high energy requirement for crop production, including that required to produce nitrogenous fertilizer. It is therefore necessary to develop new crop management strategies to reduce inputs (Loyce *et al.*, 2002) as well as exploiting wheats with low N input requirements combined with high starch contents (Kindred *et al.*, 2008).

The second major concern is, of course, the impact on international grain prices which may exacerbate problems of grain supply to less affluent populations.

### New benefits to consumers

The increasing awareness of the important role of wheat-based products in a healthy diet has been discussed above,

focusing on the identification and exploitation of natural variation in bioactive components. However, in some cases the natural variation in a trait may be limited in extent or difficult to exploit and, in this case, other approaches may be required. Currently, the most important target of this type of approach is resistant starch.

Most of the starch consumed in the human diet, including wheat starch, is readily digested in the small intestine, resulting in a rapid increase in blood glucose which may contribute to the development of type 2 diabetes and obesity (Sobal, 2007). However, a fraction of the starch may resist digestion and pass through the small intestine to the colon, where it is fermented to short chain fatty acids, notably butyrate, which may have health benefits including reduction of colo-rectal cancer (as discussed by Topping, 2007).

Although the proportion of resistant starch (RS) depends on a number of factors including the effects of food processing, the most widely studied form is high amylose starch. In most species, amylose accounts for 20–30% of starch and amylopectin for 70–80%. However, mutant lines have been identified in a number of species in which the proportion of amylose is increased up to about 40% (e.g. Glacier barley; Yoshimoto *et al.*, 2000).

Selection for high amylose mutants is relatively easy in a diploid species such as barley, but more painstaking approaches are required in hexaploid wheat as mutations in homoeologous genes on all three genomes may be required to have a significant effect on the phenotype. This has been demonstrated very elegantly by Yamamori *et al.* (2000) who combined mutations in the gene encoding the starch synthase II enzyme (also called starch granule protein 1) that catalyses the synthesis of amylopectin. The resulting triple mutant line contained about 37% amylose.

However, the complexity of starch biosynthesis means that similar high amylose phenotypes can result from changes in other enzymes, with a notable example being the use of RNA interference technology to down-regulate the gene encoding starch-branching enzyme IIa (Regina *et al.*, 2006). The resulting transgenic lines had up to 80% amylose and increased RS as measured in rat feeding trials. This study demonstrates the power of GM technology, although it remains to be shown that lines with such high levels of amylose will have acceptable yields and properties for milling and processing.

It also remains to be shown that consumers will be prepared to eat bread and other foods produced from GM wheat. The wheat grain and its products have been treated with reverence by humans for millennia and GM wheat may just be regarded as a step too far, even in countries in which other GM crops are currently accepted.

## Acknowledgements

I wish to thank all of my colleagues and collaborators who have contributed to the work discussed in this article, Professor John Snape and the John Innes Institute for providing Fig. 1 and Dr Jane Ward (Rothamsted) for

preparing Fig. 4. Rothamsted Research receives grant-aided support from the Biotechnology and Biological Sciences Research Council of the UK.

## References

- Abdel-Aal E-SM, Sosulski FW, Hucl P.** 1998. Origins, characteristics and potentials of ancient wheats. *Cereal Foods World* **43**, 708–715.
- Adams ML, Lombi E, Zhao FJ, McGrath SP.** 2002. Evidence of low selenium concentrations in UK bread-making wheat grain. *Journal of the Science of Food and Agriculture* **82**, 1160–1165.
- Altpeter F, Vasil V, Srivastava V, Vasil IK.** 1996. Integration and expression of the high-molecular-weight glutenin subunit 1Ax1 gene into wheat. *Nature Biotechnology* **14**, 1155–1159.
- Amend T, Beauvais F.** 1995. Der mechanismus der Teigbildung: Vorstoß in den molekularen Strukturbereich. *Getreide Mehl Und Brot* **49**, 359–362.
- Anderson RP, Degano P, Godkin AJ, Jewell DP, Hill AVS.** 2000. *In vivo* antigen challenge in celiac disease identifies a single transglutaminase-modified peptide as the dominant A-gliadin T-cell epitope. *Nature Medicine* **6**, 337–342.
- Arentz-Hansen H, Körner R, Molberg Ø, et al.** 2000. The intestinal T cell response to  $\alpha$ -gliadin in adult celiac disease is focused on a single deamidated glutamine targeted by tissue transglutaminase. *Journal of Experimental Medicine* **191**, 603–612.
- Arentz-Hansen H, McAdam SN, Molberg Ø, Fleckenstein B, Lundin KEA, Jørgensen TJD, Jung G, Roepstorff P, Sollid LM.** 2002. Celiac lesion T cells recognise epitopes that cluster in regions of gliadins rich in proline residues. *Gastroenterology* **123**, 803–809.
- Avivi L.** 1978. High grain protein content in wild tetraploid wheat *Triticum dicoccoides* Korn. In: *Fifth international wheat genetics symposium*, New Delhi, India, 23–28 February 1978. 372–380.
- Bailey CH.** 1941. A translation of Beccari's lecture 'Concerning Grain' (1728). *Cereal Chemistry* **18**, 555–561.
- Barro F, Rooke L, Békés F, Gras P, Tatham AS, Fido R, Lazzeri P, Shewry PR, Barcelo P.** 1997. Transformation of wheat with HMW subunit genes results in improved functional properties. *Nature Biotechnology* **15**, 1295–1299.
- Battais F, Mothes T, Moneret-Vautrin DA, Pineau F, Kanny G, Popineau Y, Bodinier M, Denery-Papini S.** 2005. Identification of IgE-binding epitopes on gliadins for patients with food allergy to wheat. *Allergy* **60**, 815–821.
- Becker D, Folck A, Knies P, Lörz H, Wieser H.** 2006. Silencing the  $\alpha$ -gliadins in hexaploid bread wheat. In: Lookhart GL, Ng PKW, eds. *Gluten proteins*. St Paul, MN, USA: AACC, 86–89.
- Belton PS.** 2005. New approaches to study the molecular basis of the mechanical properties of gluten. *Journal of Cereal Science* **41**, 203–211.
- Beta T, Man S, Dexter JE, Sapirstein HD.** 2005. Phenolic content and antioxidant activity of pearled wheat and roller-milled fractions. *Cereal Chemistry* **82**, 390–393.
- Blechl AE, Anderson OD.** 1996. Expression of a novel high-molecular-weight glutenin subunit gene in transgenic wheat. *Nature Biotechnology* **14**, 875–879.

- Blumenthal CS, Barlow EWR, Wrigley CW.** 1993. Growth environment and wheat quality: the effects of heat stress on dough properties and gluten proteins. *Journal of Cereal Science* **18**, 3–21.
- Brinch-Pedersen H, Borg S, Tauris B, Holm PB.** 2007. Molecular genetic approaches to increasing mineral availability and vitamin content of cereals. *Journal of Cereal Science* **46**, 308–326.
- Burnett J.** 2005. Brown is best. *History Today* **55**, 52–54.
- Busch RH, Rauch T.** 2001. US hard red spring wheat pool. In: Bonjean AP, Angus WJ, eds. *The world wheat book: a history of wheat breeding*. Paris, France: Groupe Limagrain, 431–443.
- Chee PW, Elias EM, Anderson JA, Kianian SF.** 2001. Evaluation of high grain protein concentration QTL from *Triticum turgidum* L. var. *dicoccoides* in an adapted durum wheat background. *Crop Science* **41**, 295–301.
- Coakley SM, Scherm H, Chakraborty S.** 1999. Climate change and plant disease management. *Annual Review of Phytopathology* **37**, 399–426.
- Combs GF.** 2001. Selenium in global food systems. *British Journal of Nutrition* **85**, 517–547.
- Dalling MJ.** 1985. The physiological basis of nitrogen redistribution during grain-filling in cereals. In: Harper JE, Schrader LE, Howell RW, eds. *Exploitation of physiological and genetic variability to enhance crop productivity*. Rockville, MD: American Society of Plant Physiologists, 55–71.
- Distelfeld A, Uauy C, Olmos S, Schlatter AR, Dubcovsky J, Fahima T.** 2004. Microlinearity between a 2-cM region encompassing the grain protein content locus *Gpc-6B1* on wheat chromosome 6B and a 350-kb region on rice chromosome 2. *Functional and Integrative Genomics* **4**, 59–66.
- Distelfeld A, Uauy C, Fahima T, Dubcovsky J.** 2006. Physical map of the wheat high-grain protein content gene *Gpc-B1* and development of a high-throughput molecular marker. *New Phytologist* **169**, 753–763.
- Drankham K, Carter J, Madl R, Klopfenstein C, Padula F, Lu Y, Warren T, Schmitz N, Takemoto DJ.** 2003. Antitumor activity of wheats with high orthophenolic content. *Nutrition and Cancer* **47**, 188–194.
- Dubcovsky J, Dvorak J.** 2007. Genome plasticity a key factor in the success of polyploidy wheat under domestication. *Science* **316**, 1862–1866.
- Dupont FM, Altenbach SB.** 2003. Molecular and biochemical impacts of environmental factors on wheat grain development and protein synthesis. *Journal of Cereal Science* **38**, 133–146.
- Dziuba J, Minkiewicz P, Nałęcz D, Iwaniak A.** 1999. Database of biologically active peptide sequences. *Nahrung* **43**, 190–195.
- Dyke GV.** 1993. *John Lawes of Rothamsted. Pioneer of science farming and industry*. Harpenden, UK: Hoos Press.
- Elder JH.** 2008. The gluten-free, casein-free diet in autism: an overview with clinical implications. *Nutrition in Clinical Practice* **23**, 583–588.
- Eurola MH, Ekholm PI, Ylinen ME, Koivistoinen PE, Varo PT.** 1991. Selenium in Finnish foods after beginning the use of selenate supplemented fertilizers. *Journal of the Science of Food and Agriculture* **56**, 57–70.
- Ellis HJ, Pollock EL, Engel W, Fraser JS, Rosen-Bronson S, Wieser H, Ciclitira PJ.** 2003. Investigation of the putative immunodominant T cell epitopes in coeliac disease. *Gut* **52**, 212–217.
- Fan M-S, Zhao F-J, Fairweather-Tait SJ, Poulton PR, Dunham SJ, McGrath SP.** 2008a. Evidence of decreasing mineral density in wheat grain over the last 160 years. *Journal of Trace Elements in Medicine and Biology* doi:10.1016/j.tem.2008.07.002.
- Fan M-S, Zhao F-J, Poulton PR, McGrath SP.** 2008b. Historical changes in the concentrations of selenium in soil and wheat grain from the Broadbalk experiment over the last 160 years. *Science of the Total Environment* **389**, 532–538.
- FAO/WHO.** 2001. Human vitamin and mineral requirements. In: *Report of a joint FAO/WHO expert consultation*. Bangkok, Thailand: Food and Nutrition Division, FAO, Rome, Italy.
- FAO/WHO/UNU.** 2007. *Protein and amino acid requirements in human nutrition*. Geneva, Switzerland: WHO Press.
- Feighery C.** 1999. Coeliac disease. *British Medical Journal* **29**, 236–239.
- Feldman M.** 1995. Wheats. In: Smartt J, Simmonds NW, eds. *Evolution of crop plants*. Harlow, UK: Longman Scientific and Technical, 185–192.
- Feldman M.** 2001. Origin of cultivated wheat. In: Bonjean AP, Angus WJ, eds. *The world wheat book: a history of wheat breeding*. Paris, France: Lavoisier Publishing, 3–56.
- Field JM, Bhandari D, Bonet A, Underwood C, Darlington H, Shewry PR.** 2008. Introgression of transgenes into a commercial cultivar confirms differential effects of HMW subunits 1Ax1 and 1Dx5 on gluten properties. *Journal of Cereal Science* **48**, 457–463.
- Findiet Study Group.** 2003. *National FINDIET 2002 study*, Vol. B3. Helsinki, Finland: National Public Health Institute.
- Finney KF.** 1978. Genetically high protein hard winter wheat. *The Bakers Digest* **June**, 32–35.
- Fossati D, Ingold M.** 2001. Mountain wheat pool. In: Bonjean AP, Angus WJ, eds. *The world wheat book: a history of wheat breeding*. Paris, France: Lavoisier Publishing, 311–332.
- Foulkes MJ, Hawkesford MJ, Barraclough PB, Holdsworth M, Kerr S, Kightly S, Shewry PR.** 2009. Identifying traits to improve the nitrogen economy of wheat: recent advances and future prospects. *Field Crops Research* (in press).
- Fry L.** 1992. Dermatitis herpetiformis. In: Marsh MN, ed. *Coeliac disease*. Oxford, UK: Blackwell Scientific Publications, 81–104.
- Galili G.** 1997. The prolamin storage proteins of wheat and its relatives. In: Larkins BA, Vasil IK, eds. *Cellular and molecular biology of plant seed development*. Dordrecht, The Netherlands: Kluwer, 221–256.
- Galili G, Altschuler Y, Levanony H, Giorini-Silfen S, Shimoni Y, Shani N, Karchi H.** 1995. Assembly and transport of wheat storage proteins. *Journal of Plant Physiology* **145**, 626–631.
- Garvin DF, Welch RM, Finley JW.** 2006. Historical shifts in the seed mineral micronutrient concentration of US hard red winter wheat germplasm. *Journal of the Science of Food and Agriculture* **86**, 2213–2220.

- Gebruers K, Domez E, Boros D, Fraš A, Dynkowska W, Bedó Z, Rakszegi M, Delcour JA, Courtin CM.** 2008. Variation in the content of dietary fiber and components thereof in wheats in the HEALTHGRAIN diversity screen. *Journal of Agricultural and Food Chemistry* **56**, 9740–9749.
- Gil-Humanes J, Pistón F, Hernando A, Alvarez JB, Shewry PR, Barro F.** 2008. Silencing of  $\gamma$ -gliadins by RNA interference (RNAi) in bread wheat. *Journal of Cereal Science* **48**, 565–568.
- Goldberg G. (ed).** 2003. *Plants: diet and health*. Report of a British Nutrition Foundation Task Force, Oxford, UK: Blackwell Science.
- Golubkina NA, Alfthan GV.** 1999. The human selenium status in 27 regions of Russia. *Journal of Trace Elements in Medicine and Biology* **13**, 15–20.
- Grant EC.** 1979. Food allergies and migraine. *Lancet* **1**, 66–69.
- Guttieri M, Bowen D, Dorsch JA, Raboy V, Souza E.** 2004. Identification and characterization of a low phytic acid wheat. *Crop Science* **44**, 418–424.
- Hadjivassiliou M, Grünewald RA, Davies-Jones GAB.** 2002. Gluten sensitivity as a neurological illness. *Journal of Neurology, Neurosurgery and Psychiatry* **72**, 560–563.
- Hadjivassiliou M, Grünewald RA, Sharrack B, Sanders D, Lobo A, Williamson C, Woodroffe N, Wood N, Davies-Jones A.** 2003. Gluten ataxia in perspective: epidemiology, genetic susceptibility and clinical characteristics. *Brain* **126**, 685–691.
- Halford NG, Field JM, Blair H, Urwin P, Moore K, Robert L, Thompson R, Flavell RB, Tatham AS, Shewry PR.** 1992. Analysis of HMW glutenin subunits encoded by chromosome 1A of bread wheat (*Triticum aestivum* L.) indicates quantitative effects on grain quality. *Theoretical and Applied Genetics* **83**, 373–378.
- Hawkesford MJ, Zhao F-J.** 2007. Strategies for increasing the selenium content of wheat. *Journal of Cereal Science* **46**, 282–292.
- Henderson KN, Tye-Din JA, Reid HH, et al.** 2007. A structural and immunological basis for the role of human leukocyte antigen DQ8 in celiac disease. *Immunity* **27**, 1–12.
- Heun M, Schäfer-Pregl R, Klawan D, Castagna R, Accerbi M, Borghi B, Salamini F.** 1997. Site of einkorn wheat domestication identified by DNA fingerprinting. *Science* **278**, 1312–1314.
- Högy P, Fangmeier A.** 2008. Effects of elevated atmospheric CO<sub>2</sub> on grain quality of wheat. *Journal of Cereal Science* **48**, 580–591.
- Howarth JR, Parmar S, Jones J, et al.** 2008. Co-ordinated expression of amino acid metabolism in response to N and S deficiency during wheat grain filling. *Journal of Experimental Botany* **59**, 3675–3689.
- Hull CM, Liddle M, Hansen N, Meyer LJ, Schmidt L, Taylor T, Jaskowski TD, Hill HR, Zone JJ.** 2008. Elevation of IgA anti-epidermal transglutaminase antibodies in dermatitis herpetiformis. *British Journal of Dermatology* **159**, 120–124.
- Hvatum M, Kanerud L, Hällgren R, Brandtzaeg P.** 2006. The gut–joint axis: cross-reactive food antibodies in rheumatoid arthritis. *Gut* **55**, 1240–1247.
- Jantasuriyarat C, Vales MI, Watson CJW, Riera-Lizarazu O.** 2004. Identification and mapping of genetic loci affecting the free-threshing habit and spike compactness in wheat (*Triticum aestivum* L.). *Theoretical and Applied Genetics* **108**, 261–273.
- Johnson VA, Mattern PJ, Peterson CJ, Kuhr SL.** 1985. Improvement of wheat protein by traditional breeding and genetic techniques. *Cereal Chemistry* **62**, 350–355.
- Jones G.** 2007. *The millers: a story of technological endeavour and industrial success, 1870–2001*. Lancaster, UK: Carnegie Publishing Ltd.
- Jones HD, Sparks CA, Shewry PR.** 2009. Transgenic manipulation of wheat quality. In: Khan K, Shewry PR, eds. *Wheat: chemistry and technology*, 4th edn. St Paul, MN, US: AACC (in press).
- Kalaydiian AE, Eaton W, Cascella N, Fasano A.** 2006. The gluten connection: the association between schizophrenia and celiac disease. *Acta Psychiatr Scandinavia* **113**, 82–90.
- Karell K, Louka AS, Moodie SJ, Ascher H, Clot F, Greco L, Ciclitira PJ, Sollid LM, Partanen J, the members of the European Genetics Cluster on Celiac Disease.** 2003. HLA types in celiac disease patients not carrying the *DQA1* \*05-*DQB1* \*02 (DQ2) heterodimer: results from the European Genetics Cluster on Celiac Disease. *Human Immunology* **64**, 469–477.
- Kindred DR, Verhoeven TMO, Weightman RM, Swanston JS, Agu RC, Brosnan JM, Sylvester-Bradley R.** 2008. Effects of variety and fertilizer nitrogen on alcohol yield, grain yield, starch and protein content, and protein composition of winter wheat. *Journal of Cereal Science* **48**, 46–57.
- Kumamaru T, Ogawa M, Satoh H, Okita TW.** 2007. Protein body biogenesis in cereal endosperms. *Plant Cell Monographs* doi 10.1007/7089\_2007\_115.
- Lampi A-M, Nurmi T, Ollilainen V, Piironen V.** 2008. Tocopherols and tocotrienols in wheat genotypes in the HEALTHGRAIN diversity screen. *Journal of Agricultural and Food Chemistry* **56**, 9716–9721.
- Levanany H, Rubin R, Altschuler Y, Galili G.** 1992. Evidence for a novel route of wheat storage proteins to vacuoles. *Journal of Cell Biology* **119**, 1117–1128.
- Lewis SJ, Heaton KW.** 1999. The metabolic consequences of slow colonic transit. *American Journal of Gastroenterology* **94**, 2010–2016.
- Li L, Shewry PR, Ward JL.** 2008. Phenolic acids in wheat varieties in the HEALTHGRAIN diversity screen. *Journal of Agricultural and Food Chemistry* **56**, 9732–9739.
- Losowsky MS.** 2008. A history of coeliac disease. *Digestive Diseases* **26**, 112–120.
- Loyce C, Relier JP, Meynard JM.** 2002. Management planning for winter wheat with multiple objectives (2): ethanol-wheat production. *Agricultural Systems* **72**, 33–37.
- Lucarelli S, Frediani T, Zingoni AM, Ferruzzi F, Giardini O, Quintieri F, Barbato M, D'Eufemia P, Cardi E.** 1995. Food allergy and infantile autism. *Panminerva Medicine* **37**, 137–141.
- Martinant JP, Billot A, Bouguennec A, Charmet G, Saulnier L, Branlard G.** 1999. Genetic and environmental variations in water-extractable arabinoxylans content and flour extract viscosity. *Journal of Cereal Science* **30**, 45–48.
- Matsuo H, Morita E, Tatham AS, Morimoto K, Horikawa T, Osuna H, Ikezawa Z, Kaneko S, Kohno K, Dekio S.** 2004. Identification of the IgE-binding epitope in  $\omega$ -5 gliadin, a major allergen in wheat-dependent exercise-induced anaphylaxis. *Journal of Biological Chemistry* **279**, 12135–12140.

- Matsuo H, Kohno K, Nihara H, Morita E.** 2005. Specific IgE determination to epitope peptides of omega-5 and high molecular weight glutenin subunit is a useful tool for diagnosis of wheat-dependent exercise-induced anaphylaxis. *Journal of Immunology* **175**, 8116–8122.
- Mazzarella G, Maglio M, Paparo F, et al.** 2003. An immunodominant DQ8 restricted gliadin peptide activates small intestinal immune response in *in vitro* cultured mucosa from HLA-DQ8 positive but not HLA-DQ8 negative coeliac patients. *Gut* **52**, 57–62.
- Micronutrient Initiative.** 2006. *Controlling vitamin and mineral deficiencies in India: meeting the goal*. New Delhi, India: Micronutrient Initiative.
- Millar SJ, Snape J, Ward J, Shewry PR, Belton P, Boniface K, Summers R.** 2008. *Investigating wheat functionality through breeding and end use*. CCFRA Report on LINK Project FQS 23.
- Ministry of Agriculture, Fisheries and Food.** 1997. *United Kingdom dietary intake of selenium*. London, UK: MAFF Food Surveillance Information Sheet No. 126, HMSO.
- Moore MA, Beom Park C, Tsuda H.** 1998. Soluble and insoluble fiber influences on cancer development. *Critical Reviews in Oncology/Hematology* **27**, 229–242.
- Morita E, Matsuo H, Mihara S, Morimoto K, Savage AWJ, Tatham AS.** 2003. Fast  $\omega$ -5 gliadin is a major allergen in wheat-dependent exercise-induced anaphylaxis. *Journal of Dermatological Science* **33**, 99–104.
- Mossé J, Huet J-C.** 1990. Amino acid composition and nutritional score for ten cereals and six legumes or oilseeds: causes and ranges of variations according to species and to seed nitrogen content. *Sciences des Alimentations* **10**, 151–173.
- Motoi H, Kodama T.** 2003. Isolation and characterization of angiotensin 1-converting enzyme inhibitory peptides from wheat gliadin hydrolysate. *Nahrung* **47**, 354–358.
- Murphy JD, Power NM.** 2008. How can we improve the energy balance of ethanol production from wheat? *Fuel* **87**, 1799–1806.
- Nagy-Scholz E, Ercsey K.** 2009. Lignan analysis of cereal samples by GC/MS method. In: Shewry PR, Ward J, eds. *The HEALTHGRAIN methods book*. St Paul, MN, USA: AACCC (in press).
- Nalam VJ, Vales MI, Watson CJW, Kianian SF, Riera-Lizarazu O.** 2006. Map-based analysis of genes affecting the brittle rachis character in tetraploid wheat (*Triticum turgidum* L.). *Theoretical and Applied Genetics* **112**, 373–381.
- Nesbitt M.** 1998. Where was einkorn wheat domesticated? *Trends in Plant Science* **3**, 1360–1385.
- Neuhausen SL, Steele L, Ryan S, Mousavi M, Pinto M, Osann KE, Flodman P, Zone JJ.** 2008. Co-occurrence of celiac disease and other autoimmune diseases in celiacs and their first-degree relatives. *Journal of Autoimmunity* **31**, 160–165.
- Nurmi T, Nyström L, Edelmann M, Lampi A-M, Piironen V.** 2008. Phytosterols in wheat genotypes in the HEALTHGRAIN diversity screen. *Journal of Agricultural and Food Chemistry* **56**, 9710–9715.
- Ordaz-Ortiz JJ, Devaux M-F, Saulnier L.** 2005. Classification of wheat varieties based on structural features of arabinoxylans as revealed by endoxylanase treatment of flour and grain. *Journal of Agricultural and Food Chemistry* **53**, 8349–8356.
- Ortiz-Monasterio JI, Palacios-Rojas N, Meng E, Pixley K, Trethowan R, Peña RJ.** 2007. Enhancing the mineral and vitamin content of wheat and Maite through plant breeding. *Journal of Cereal Science* **46**, 293–307.
- Osborne TB.** 1924. *The vegetable proteins*, 2nd edn. London, UK: Longmans Green & Co.
- Palosuo K, Varjonen E, Kekki OM, Klemola T, Kalkkinen N, Alenius H, Reunala T.** 2001. Wheat omega-five gliadin is a major allergen in children with immediate allergy to ingested wheat. *Journal of Allergy and Clinical Immunology* **108**, 634–638.
- Payne PI.** 1987. Genetics of wheat storage proteins and the effect of allelic variation on breadmaking quality. *Annual Review of Plant Physiology* **38**, 141–153.
- Payne PI, Corfield KG, Blackman JA.** 1979. Identification of a high molecular weight subunit of glutenin whose presence correlates with breadmaking quality in wheats of related pedigree. *Theoretical and Applied Genetics* **55**, 153–159.
- Pepper S.** 1992. Allinson's staff of life. *History Today* **42**, 30–35.
- Piironen V, Edelmann M, Kariluoto S, Bedó Z.** 2008. Folate in wheat genotypes in the HEALTHGRAIN diversity screen. *Journal of Agricultural and Food Chemistry* **56**, 9726–9731.
- Poole JA, Barriga K, Leung DYM, Hoffman M, Eisenbarth GS, Rewers M, Norris JM.** 2006. Timing of initial exposure to cereal grains and the risk of wheat allergy. *Pediatrics* **117**, 2175–2182.
- Porter JR, Semenov MA.** 2005. Crop responses to climatic variation. *Philosophical Transactions of the Royal Society B* **360**, 2021–2035.
- Poutanen K, Shepherd R, Shewry PR, Delcour JA, Björck I, Kamp JW.** 2008. Beyond whole grain: the European HEALTHGRAIN project aims at healthier cereal foods. *Cereal Foods World* **53**, 32–35.
- Rakszegi M, Pastori G, Jones HD, Békés F, Butow B, Láng L, Bedó Z, Shewry PR.** 2008. Technological quality of field-grown transgenic lines of commercial wheat cultivars expressing the 1Ax1 HMW glutenin subunit gene. *Journal of Cereal Science* **47**, 310–321.
- Regina A, Bird A, Topping D, Bowden S, Freeman J, Barsby T, Kosar-Hashemi B, Li Z, Rahman S, Morell M.** 2006. High-amylose wheat generated by RNA interference improves indices of large-bowel health in rats. *Proceedings of the National Academy of Sciences, USA* **103**, 3546–3551.
- Rix KJ, Ditchfield J, Freed DL, Goldberg DP, Hillier VF.** 1985. Food antibodies in acute psychoses. *Psychological Medicine* **15**, 347–354.
- Salcedo G, Sánchez-Monge G, Garcia-Casado G, Armentia A, Gomez L, Barber D.** 2004. The cereal  $\alpha$ -amylase/trypsin inhibitor family associated with bakers' asthma and food allergy. In: Mills ENC, Shewry PR, eds. *Plant food allergens*. Oxford, UK: Blackwell Science Ltd, 70–86.
- Seilmeier W, Belitz H-D, Wieser H.** 1991. Separation and quantitative determination of high-molecular-weight subunits of glutenin from different wheat varieties and genetic variants of the variety Sicco. *Zeitschrift für Lebensmittel-Untersuchung und Forschung* **192**, 124–129.
- Semenov MA.** 2008. Impacts of climate change on wheat in England and Wales. *Journal of the Royal Society Interface* doi:10.1098/rsif.2008.0285.

- Shewry PR.** 2000. Seed proteins. In: Black M, Bewley JD, eds. *Seed technology and its biological basis*. Sheffield, UK: Sheffield Academic Press, 42–84.
- Shewry PR.** 2007. Improving the protein content and composition of cereal grain. *Journal of Cereal Science* **46**, 239–250.
- Shewry PR, Jones HD.** 2005. Transgenic wheat: where do we stand after the first 12 years? *Annals of Applied Biology* **147**, 1–14.
- Shewry PR, Tatham AS, Barro F, Barcelo P, Lazzeri P.** 1995. Biotechnology of breadmaking: unraveling and manipulating the multi-protein gluten complex. *Biotechnology* **13**, 1185–1190.
- Shewry PR, Halford NG, Lafiandra D.** 2003a. The genetics of wheat gluten proteins. In: Hall JC, Dunlap JC, Friedman T, eds. *Advances in genetics*, Vol. 49. Academic Press, 111–184.
- Shewry PR, Halford NG, Tatham AS, Popineau Y, Lafiandra D, Belton PS.** 2003b. The high molecular weight subunits of wheat glutenin and their role in determining wheat processing properties. *Advances in Food and Nutrition Research* **45**, 221–302.
- Simons KJ, Fellers JP, Trick HN, Zhang Z, Tai Y-S, Gill BS, Faris JD.** 2006. Molecular characterization of the major wheat domestication gene Q. *Genetics* **172**, 547–555.
- Singh MM, Roy SR.** 1975. Wheat gluten as a pathogenic factor in schizophrenia. *Science* **191**, 401–402.
- Sjöström H, Lunkin KEA, Molberg Ø, et al.** 1998. Identification of a gliadin T cell epitope in coeliac disease: general importance of gliadin deamidation for intestinal T cell recognition. *Scandinavian Journal of Immunology* **48**, 111–115.
- Skylas DJ, Mackintosh JA, Cordwell SJ, Basseal DJ, Walsh BJ, Harry J, Blumenthal C, Copeland L, Wrigley CW, Rathmell W.** 2000. Proteome approach to the characterization of protein composition in the developing and mature wheat grain endosperm. *Journal of Cereal Science* **32**, 169–188.
- Snape J, Hyne V, Aitken K.** 1993. Targeting genes in wheat using marker mediated approaches. In: *Proceedings of the 8th international wheat genetics symposium*, Beijing, 749–759.
- Snape J, Pánková K.** 2006. *Triticum aestivum* (wheat). In: Marquart L, Jacobs DR Jr, McIntosh GH, Poutanen K, Reicks M, eds. *Encyclopedia of life sciences*. John Wiley & Sons Ltd.
- Sobal J.** 2007. Using a model of the food and nutrition system for examining whole grain foods from agriculture to health. In: *Whole grains and health*. Oxford, UK: Blackwell Publishing, 17–25.
- Spaenij-Dekking L, Kooy-Winkelaar Y, Van Veelen P, Drijfhout JW, Jonker H, Van Soest L, Smulders MJM, Bosch D, Gilissen LJWJ, Koning F.** 2005. Natural variation in toxicity of wheat: potential for selection of non-toxic varieties for celiac disease patients. *Gastroenterology* **129**, 797–806.
- Stoltzfus RJ, Dreyfuss ML.** 1998. *Guidelines for the use of iron supplements to prevent and treat iron deficiency anaemia*. Washington DC, USA: ILSI Press.
- Szabó AT, Hammer K.** 1996. Notes on the taxonomy of farro: *Triticum monococcum*, *T. dicoccon* and *T. spelta*. In: *Hulled wheats*. Proceedings of the 1st International Workshop on Hulled Wheats, 21–22 July 1995, Tuscany, Italy. IPGRI, Italy, 2–40.
- Takahashi M, Fukunaga H, Fukudome S-I, Yoshikawa M.** 2000. Behavioural and pharmacological studies on gluten exorphin A5, a newly isolated bioactive food protein fragment, in mice. *Japanese Journal of Pharmacology* **84**, 259–265.
- Tatham AS, Shewry PR.** 2008. Allergy to wheat and related cereals. *Clinical and Experimental Allergy* **38**, 1712–1726.
- Tollefsen S, Arentz-Hansen H, Fleckenstein B, Molberg Ø, Ráki M, Kwok WW, Jung G, Lundin KEA, Sollid LM.** 2006. HLA-DQ2 and -DQ8 signatures of gluten T cell epitopes in celiac disease. *Journal of Clinical Investigation* **116**, 2226–2236.
- Topping D.** 2007. Cereal complex carbohydrates and their contribution to human health. *Journal of Cereal Science* **46**, 220–229.
- Tosi P, Parker M, Gritsch C, Carzaniga R, Martin B, Shewry PR.** 2009. Trafficking of storage proteins in developing grain of wheat. *Journal of Experimental Botany* **60**, 619–627.
- Uauy C, Distelfeld A, Fahima T, Blechl A, Dubcovsky J.** 2006. A NAC gene regulating senescence improves grain protein, zinc and iron content in wheat. *Science* **314**, 1298–1301.
- Vader LW, Kooy YMC, van Veelen P, de Ru A, Harris D, Benckhuijsen W, Peña S, Mearin ML, Drijfhout JW, Koning F.** 2002a. The gluten response in children with celiac disease is directed toward multiple gliadin and glutenin peptides. *Gastroenterology* **122**, 1729–1737.
- Vader LW, de Ru A, van der Wal Y, Kooy YMC, Benckhuijsen W, Mearin ML, Drijfhout JW, van Veelen P, Koning F.** 2002b. Specificity of tissue transglutaminase explains cereal toxicity in celiac disease. *Journal of Experimental Medicine* **195**, 643–649.
- Van de Wal Y, Kooy YMC, Van Veelen PA, et al.** 1998. Small intestinal T cells of celiac disease patients recognize a natural pepsin fragment of gliadin. *Proceedings of the National Academy of Sciences, USA* **95**, 10050–10054.
- Van de Wal Y, Kooy YMC, van Veelen P, Vader W, August SA, Drijfhout JW, Peña SA, Koning F.** 1999. Glutenin is involved in the gluten-driven mucosal T cell response. *European Journal of Immunology* **29**, 3133–3139.
- Van Herpen TWJM, Goryunova SV, van der Schoot J, et al.** 2006. Alpha-gliadin genes from the A, B, and D genomes of wheat contain different sets of celiac disease epitopes. *BMC Genomics* **7**, doi:10.1186/1471-2164-7-1.
- Vitaglione P, Napolitano A, Fogliano V.** 2008. Cereal dietary fibre: a natural functional ingredient to deliver phenolic compounds into the gut. *Trends in Food Science and Technology* **19**, 451–463.
- Vogel KP, Johnson VA, Mattern PJ.** 1978. Protein and lysine contents of endosperm and bran of the parents and progenies of crosses of common wheat. *Crop Science* **18**, 751–754.
- Walusiak J, Hanke W, Górski P, Palczynski C.** 2004. Respiratory allergy in apprentice bakers: do occupational allergies follow the allergic march? *Allergy* **59**, 442–450.
- Wan Y, Poole RL, Huttly AK, et al.** 2008. Transcriptome analysis of grain development in hexaploid wheat. *BMC Genomics* **9**, 121.
- Wan Y, Underwood C, Toole G, et al.** 2009. A novel transcriptomic approach to identify candidate genes for grain quality traits in wheat. *Plant Biotechnology Journal* (in press).
- Ward JL, Poutanen K, Gebruers K, et al.** 2008. The HEALTH-GRAIN cereal diversity screen: concept, results and prospects. *Journal of Agricultural and Food Chemistry* **56**, 9699–9709.



- Wende L, Fang S, Shancheng S, Corke H, Beta T.** 2005. Free radical scavenging properties and phenolic content of Chinese black-grained wheat. *Journal of Agricultural and Food Chemistry* **53**, 8533–8536.
- Wieser H, Koehler P, Folck A, Becker D.** 2006. Characterization of wheat with strongly reduced  $\alpha$ -gliadin content. In: Lookhart GL, Ng PKW, eds. *Gluten proteins*. St Paul, MN, USA: AACC, 13–16.
- Wolnik KA, Fricke FL, Capar SG, Braude GL, Meyer MW, Satzger RD, Kuennen RW.** 1983. Elements in major raw agricultural crops in the United States. 2. Other elements in lettuce, peanuts, potatoes, soybeans, sweet corn, and wheat. *Journal of Agricultural and Food Chemistry* **31**, 1244–1249.
- Yamamori M, Fujita S, Hayakawa K, Matsuki J, Yasui T.** 2000. Genetic elimination of a starch granule protein SGP-1 of wheat generates an altered starch with apparent high amylose. *Theoretical and Applied Genetics* **101**, 21–29.
- Yamamoto N, Ejiri M, Mizuno S.** 2003. Biogenic peptides and their potential uses. *Current Pharmaceutical Design* **9**, 1345–1355.
- Yang GQ, Xia YM.** 1995. Studies on human dietary requirements and a safe: range of dietary intakes of selenium and their application in the prevention of related endemic diseases. *Biomedical and Environmental Sciences* **8**, 187–201.
- Yoshikawa M, Takahashi M, Yang S.** 2003. Delta opioid peptides derived from plant proteins. *Current Pharmaceutical Design* **9**, 1325–1330.
- Yoshimoto Y, Tashiro J, Takenouchi T, Takeda Y.** 2000. Molecular structure and some physicochemical properties of high-amylose barley starches. *Cereal Chemistry* **77**, 279–285.
- Zhao FJ, Withers PJA, Evans EJ, Monaghan J, Salmon SE, Shewry PR, McGrath SP.** 1997. Sulphur nutrition: an important factor for the quality of wheat and rapeseed. *Soil Science and Plant Nutrition* **43**, 1137–1142.
- Zhao FJ, Su YH, Dunham SJ, Rakszegi M, Bedo Z, McGrath SP, Shewry PR.** 2009. Variation in mineral micronutrient concentrations in grain of wheat lines of diverse origin. *Journal of Cereal Science* (in press).