

Attenuated levels of pro-inflammatory markers in diabetic retinopathy patients undergoing treatment with antihyperglycemic and antihypertensive drugs

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OBJECTIVE: This study aimed to assess the circulating levels of activated nuclear factor kappa B p65 and monocyte chemoattractant protein-1 in diabetic retinopathy patients who were taking antihyperglycemic and antihypertensive drugs.

METHODS: In total, 235 healthy controls and 371 Type 2 diabetic patients [171 without retinopathy (DNR) and 200 patients with retinopathy (diabetic retinopathy)] were recruited for this study. Plasma and the nuclear fraction of peripheral blood mononuclear cells were isolated for the quantification of the monocyte chemoattractant protein-1 and nuclear factor kappa B p65 levels, respectively.

RESULTS: Non-medicated diabetic retinopathy patients had significantly higher levels of activated nuclear factor kappa B p65 and plasma monocyte chemoattractant protein-1 than DNR patients. Diabetic retinopathy patients who were taking antihyperglycemic and antihypertensive drugs showed significant reductions in both the nuclear factor kappa B p65 and monocyte chemoattractant protein-1 levels compared with the non-medicated patients.

CONCLUSION: This study demonstrated the significant attenuation of both the nuclear factor kappa B p65 and circulating monocyte chemoattractant protein-1 levels in diabetic retinopathy patients taking antihyperglycemic and antihypertensive drugs.

KEYWORDS: Antihyperglycemic; Antihypertensive; Diabetic Retinopathy; Monocyte chemoattractant protein-1; Nuclear Factor kappa B.

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INTRODUCTION

The increasing prevalence of diabetes mellitus (DM) worldwide will inevitably be accompanied by an increased development of irreversible DM complications, including diabetic retinopathy (DR). DR is the most common complication among patients with type 2 DM (1), and it is the leading cause of preventable vision impairment among working-age adults (2). More than 50% of type 2 DM patients are likely to experience DR within 20 years after diagnosis (3). DR is characterized by microvascular lesions such as microaneurysms, basement membrane thickening,

loss of pericytes leading to blood barrier dysfunction and pre-retinal neovascularization (4).

The presence of oxidative stress and the generation of reactive oxygen species and advanced glycation end products are known to contribute to the development of DR (4). The role of inflammation in the development of DR is supported by increasing evidence that has shown the involvement of various cytokines and inflammatory cells in the pathogenesis of DR (5). Prolonged hyperglycemia in a person with type 2 DM creates a hypoxic state that triggers abnormal local leukocyte-endothelial interactions, which lead to retinal microvascular damage (6). Nuclear factor kappa B (NF-κB) is a "redox-sensitive" nuclear transcription factor present in many cell types that mainly regulates immune and inflammatory responses as well as apoptosis by controlling the expression of numerous genes coding for pro-inflammatory cytokines, chemokines, inflammatory enzymes and adhesion molecules (7) that are essential for leukocyte migration and adherence to vascular endothelial cells. Increased NF-κB expression had been demonstrated in

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many inflammatory diseases (7). Monocyte chemoattractant protein-1 (MCP-1) is a chemokine regulated by NF- κ B that is mainly expressed in smooth muscle cells, macrophages, endothelial cells and adipocytes (8). Its main function is to recruit circulating monocytes into the subendothelial cell layer of the blood vessel wall, and it has been reported to be involved in the pathogenesis of atherosclerosis, cardiovascular disease, obesity and insulin resistance (8).

The 1998 United Kingdom Prospective Diabetes Study provided the first evidence that early intervention with oral glucose-lowering and antihypertensive drugs reduced the incidence of diabetic complications and improved the survival of type 2 DM patients (9-10). However, it is uncertain whether this beneficial effect was derived directly from tight glycemic and blood pressure control or through other means of action. The recent large-scale Diabetic Retinopathy Candesartan control trial reported that antihypertensive drugs (angiotensin-converting enzyme inhibitors and angiotensin II receptor antagonists) delayed the progression of DR (11-12). Antihyperglycemic and antihypertensive drugs are believed to possess anti-inflammatory and endothelial-dysfunction modulating effects independent of their blood glucose- and blood pressure-lowering properties. Thus, this study aimed to assess the circulating levels of activated peripheral blood mononuclear cell (PBMC) NF- κ B p65 and plasma MCP-1 in DR patients treated with antihyperglycemic and antihypertensive drugs.

METHODS

Study population

The study subjects were recruited from the diabetes and ophthalmology clinics at the University Malaya Medical Center, Malaysia, between September 2009 and October 2011. DM had been previously diagnosed according to the World Health Organization criteria. A total of 371 unrelated Type 2 DM patients [171 patients without retinopathy (DNR) and 200 patients with retinopathy (DR)] (210 men, 161 women) aged 58.2 ± 9.7 years (mean \pm SD; range, 35 to 78 years) were recruited for this study. Detailed medical histories and socio-demographic data for each patient were noted. The patients were carefully selected by excluding any patients with a previous history of inflammatory disease who had received anti-inflammatory drug treatment or antioxidant supplements. Type 1 DM patients and type 2 DM patients with complications other than retinopathy were also excluded from the study. The non-retinopathy controls were recruited from among blood-donor volunteers. They consisted of 235 unrelated healthy subjects (134 men, 101 women) aged 57.1 ± 4.1 years (mean \pm SD; range, 45 to 65 years). Both the DNR patients and healthy subjects were confirmed to be free from any diabetic complications including retinopathy by attending doctors and ophthalmologists. Written informed consent was obtained from each subject prior to sample collection.

All of the DR patients underwent a complete eye examination that included dilated retinal examination and seven-field stereoscopic Diabetic Retinopathy Study retinal photography (13). The color fundus photographs were graded for DR severity in a blinded fashion by two independent ophthalmologists at the University of Malaya Eye Research Center, Malaysia. The modified Early Treatment of Diabetic Retinopathy Study Airlie House classification of DR was used to grade the retinopathy into

non-proliferative diabetic retinopathy (NPDR) and proliferative diabetic retinopathy (PDR) (14). Among the DR patients, 26 had mild NPDR, 85 had moderate NPDR, 14 had severe NPDR and 75 had PDR. The study was performed in adherence to the principles of the 1983 Declaration of Helsinki and approved by the Medical Ethics Review Committee of the University Malaya Medical Center, Malaysia (IRB reference number: 744.12).

Sample collection and preparation

Six milliliters of blood was drawn from patients and control subjects. Three ml of the freshly collected blood was sent for routine blood examination at the Clinical Diagnostic Laboratory of the University Malaya Medical Center. The collected whole blood samples in EDTA tubes were centrifuged for 15 minutes at 1000 xg. The plasma was extracted and stored at -80°C for the MCP-1 enzyme-linked immunosorbent assay. Subsequently, the cell sediments of the EDTA blood tubes were reconstituted with isotonic phosphate-buffered saline solution. The peripheral blood mononuclear cells (PBMCs) were then isolated as previously described (15) using a density gradient centrifugation method. PBMCs (approximately 4 million cells) were subjected to nuclear extraction, and the extracted nuclear fraction was used for the NF- κ B p65 transcription factor assay. Analysis of all of the samples was performed within 1 month after collection.

PBMC nuclear extraction

The nuclear fraction of the isolated PBMCs was extracted using a commercially available nuclear extraction kit (Cayman Chemical Company, MI, United States) according to the manufacturer's protocol. In brief, the extracted PBMCs were collected in ice-cold PBS in the presence of phosphatase inhibitors to prevent the events controlled by dephosphorylation. The pelleted cells were then resuspended in ice-cold hypotonic buffer, causing cell swelling and increased membrane fragility. The addition of detergent (10% Nonidet P-40) ruptured the cell membranes and released the cytoplasmic fraction while maintaining the integrity of the nuclear membranes. After separation of the cytoplasmic fraction from the nuclei by a brief centrifugation, the pelleted nuclei were lysed in ice-cold extraction buffer containing a mixture of protease and phosphatase inhibitors. The nuclear extract was then isolated by microcentrifugation (14,000 xg for 10 minutes at 4°C) and stored at -80°C .

Measurement of the NF- κ B p65 levels in PBMCs

The levels of activated NF- κ B p65 in the nuclei of PBMCs were measured using a transcription factor assay kit (Cayman Chemical Company, MI, United States). The kit utilized a specific double-stranded DNA sequence containing an NF- κ B response element to specifically bind the activated NF- κ B p65 in the nuclear extract. NF- κ B p65 was detected by the addition of a specific primary antibody directed against NF- κ B p65. A secondary antibody conjugated to horseradish peroxidase was added to provide a sensitive colorimetric readout at 450 nm. The inter-assay coefficient of variation was 8%. The nuclear protein concentration was determined using a Bradford assay (16), and the activated NF- κ B p65 level was expressed as arbitrary units per milligram of protein (AU/mg of nuclear protein).



Measurement of the plasma MCP-1 levels

The plasma MCP-1 levels were quantitatively measured with a sandwich enzyme-linked immunosorbent assay standard kit (Raybiotech® Inc., GA, United States) according to the manufacturer's protocol. The plate was coated with a specific monoclonal antibody directed against human MCP-1, and a polyclonal antibody conjugated to horseradish peroxidase was used for sensitive colorimetric detection at 450 nm. The inter-assay coefficient of variation was 7.7%. The mean minimal detectable level of MCP-1 was typically less than 2 pg/ml. The results were expressed as pg/ml.

Statistical analysis

The continuous variables were checked for normality prior to statistical analysis. A chi-squared test with one degree of freedom (for dichotomous variables) and an unpaired t-test (for continuous variables) were used to evaluate the differences between the groups. Comparison of subgroups was performed with one-way analysis of variance (ANOVA) and Tukey's post-hoc test. Associations between parameters were determined by Pearson's correlation coefficient (*r*) with Bonferroni correction. A logistic regression model was used to estimate the odds ratio (OR) and 95% confidence interval (CI) for each risk factor for DR among the type 2 DM patients. Statistical significance was set at *p*<0.05. All of the data were

analyzed using GraphPad Prism® for Windows® version 5.02 (GraphPad® Software Inc., CA, United States).

RESULTS

The general clinical parameters for the healthy controls and the DNR and DR patients are listed in Table 1. Both the DNR and DR patients showed significantly (*p*<0.05) higher levels of glycosylated hemoglobin (HbA_{1c}), total cholesterol, high-density lipoprotein (HDL-C) and low-density lipoprotein (LDL-C), higher systolic blood pressures (SBP), a higher prevalence of hypertension, a lower HDL/LDL ratio and lower diastolic blood pressures (DBP) compared to the healthy controls. When the two patient groups were compared, the DR patients had significantly (*p*<0.05) higher levels of HbA_{1c} and total cholesterol, a longer duration of DM and more subjects who received insulin treatment. No significant differences (*p*>0.05) in gender, age, body mass index (BMI), triglyceride levels, alanine aminotransferase (ALT) levels or aspartate aminotransferase (AST) levels were observed.

In this study, DR patients who were free from the influence of antihyperglycemic and antihypertensive medications had significantly higher levels of NF-κB p65 and plasma MCP-1 than the healthy controls (Figure 1). In addition, significantly higher levels of NF-κB p65 were

Table 1 - General clinical parameters of healthy controls and DNR and DR patients.

Demographics	Ctrl (n = 235)	DNR (n = 171)	DR (n = 200)
Age (years)	57.1 ± 4.1	59.2 ± 9.6	57.2 ± 9.8
Gender (male/female)	134/101	100/71	110/90
Race (Malay/Chinese/Indian)	106/90/39	63/28/80 ^a	70/47/83 ^a
BMI (kg/m ²)	25.6 ± 4.8 (n = 100)	27.2 ± 4.4	26.3 ± 5.0
HbA _{1c} (%)	5.6 ± 0.4 (n = 100)	7.9 ± 1.8 ^a	8.9 ± 2.1 ^{a,b}
SBP (mmHg)	124.0 ± 8.0 (n = 100)	136.5 ± 19.5 ^a	139.3 ± 22.4 ^a
DBP (mmHg)	83.0 ± 7.0 (n = 100)	79.0 ± 10.5 ^a	78.4 ± 13.1 ^a
Total cholesterol (mmol/l)	3.8 ± 0.6 (n = 100)	4.5 ± 1.0 ^a	4.8 ± 1.5 ^{a,b}
Triglycerides (mmol/l)	1.8 ± 1.3 (n = 100)	1.6 ± 0.7	1.7 ± 1.0
HDL-C (mmol/l)	1.0 ± 0.3 (n = 100)	1.2 ± 0.3 ^a	1.2 ± 0.3 ^a
LDL-C (mmol/l)	2.1 ± 0.5 (n = 100)	2.5 ± 0.9 ^a	2.8 ± 1.2 ^a
HDL-C/LDL-C ratio	0.6 ± 0.2 (n = 100)	0.5 ± 0.2 ^a	0.5 ± 0.2 ^a
ALT (IU/l)	30-65 ^c	37.8 ± 17.5	36.8 ± 24.6
AST (IU/l)	15-37 ^c	22.0 ± 14.0	22.8 ± 16.4
Diabetes duration (years)	-	10.4 ± 7.9	15.7 ± 9.1 ^b
Retinopathy duration (years)	-	-	5.0 ± 3.6
Current smoker (yes/no)	43/192	29/142	13/187 ^{a,b}
Alcohol intake (yes/no)	70/165	24/147 ^a	16/184 ^a
Hypertension (yes/no)	0/235	104/67 ^a	119/81 ^a
Antihyperglycemic treatment duration (years)	-	9.5 ± 5.5 ^a (n = 107)	11.5 ± 7.5 ^a (n = 130)
Antihyperglycemic medication (yes/no)	0/235	107/64 ^a	130/70 ^a
Insulin (yes/no)	0/235	34/137 ^a	98/102 ^{a,b}
Oral medication (yes/no)	0/235	119/81 ^a	81/119 ^{a,b}
Antihypertensive treatment duration (years)	-	7.0 ± 3.5 ^a (n = 104)	8.5 ± 4.0 ^a (n = 119)
Antihypertensive medication (yes/no)	0/235	104/67 ^a	119/81 ^a
ACEI & ARA (yes/no)	0/235	67/104 ^a	83/117 ^a
CCB & Diuretics (yes/no)	0/235	37/134 ^a	36/164 ^a

The data are expressed as the mean ± SD unless otherwise indicated; dichotomous variables are given in absolute numbers. ACEI, angiotensin-converting enzyme inhibitors; ALT, alanine aminotransferase; ARA, angiotensin II receptor antagonists; AST, aspartate aminotransferase; BMI, body mass index; CCB, calcium channel blockers; Ctrl, healthy controls; DBP, diastolic blood pressure; DNR, diabetic non-retinopathy; DR, diabetic retinopathy; HbA_{1c}, glycosylated hemoglobin; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; and SBP, systolic blood pressure. ^a*p*<0.05 versus healthy control; ^b*p*<0.05 versus DNR; ^cnormal value range provided.

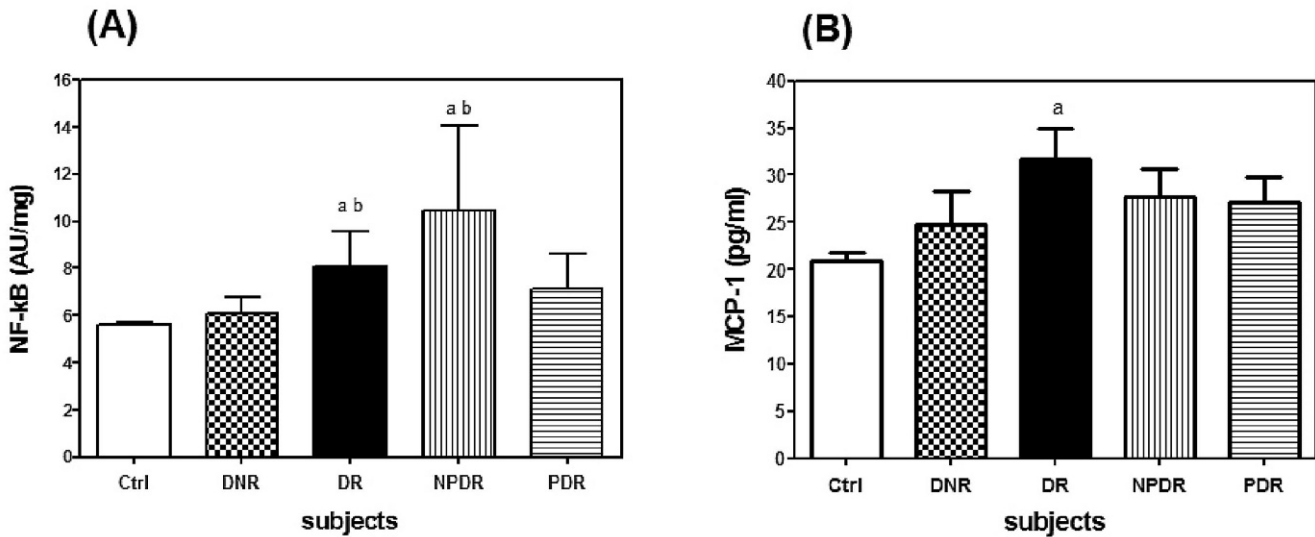


Figure 1 - Comparison of the levels of (A) NF-κB p65 and (B) plasma MCP-1 in Ctrl (n = 235), DNR (n = 50), DR (n = 70), NPDR (n = 40) and PDR (n = 30) patients who were not taking antihyperglycemic and antihypertensive medications. The data are expressed as the mean ± SD. Ctrl, healthy controls; DNR, diabetic non-retinopathy; DR, diabetic retinopathy; NPDR, non-proliferative DR; PDR, proliferative DR. ^ap < 0.05 versus Ctrl; ^bp < 0.05 versus DNR.

found in the DR and NPDR patients compared with the DNR patients. Both the DNR and DR patients who were taking antihyperglycemic and antihypertensive medications showed lower levels of NF-κB p65 and plasma MCP-1 compared with the non-medicated patients (Table 2). Further comparisons between the medicated and non-medicated patients (NPDR and PDR) showed similar trends in the results.

The correlation results observed in this study (Table 3) were weak (r < 0.5) but significant (p < 0.05). The NF-κB p65 level was positively correlated with the plasma MCP-1 level in healthy controls and DNR patients as well as with the diabetes duration in both DNR and DR patients. In addition, positive correlations were also observed between the NF-κB p65 and HbA_{1c} levels as well as the triglyceride level in the DNR patients. Both the SBP and DM duration in the DNR patients were positively correlated with the plasma MCP-1 level, but an inverse correlation was found between the plasma MCP-1 and HDL-C levels. In DR patients, the

plasma MCP-1 level was positively correlated with the HbA_{1c} level, DM duration and retinopathy duration.

The risk factors for DR were investigated among the type 2 DM patients (DNR and DR) using a logistic regression model (Table 4). High levels of HbA_{1c} (OR = 1.20, p < 0.05), plasma MCP-1 (OR = 1.04, p < 0.05) or activated NF-κB p65 (OR = 1.08, p < 0.05) and long diabetes duration (OR = 1.08, p < 0.05) were risk factors for DR after adjusting for age, gender and other metabolic factors. Antihyperglycemic (OR = 0.81 for oral and OR = 0.63 for insulin, p < 0.05 for both) and antihypertensive (OR = 0.79, p < 0.05) medications were protective against the development of DR by type 2 DM patients.

DISCUSSION

DR is a chronic, low-grade inflammatory disease caused by the presence of microscopic signs of inflammation in the retina such as vasodilatation, fluid exudation, leukocyte

Table 2 - Comparison of the NF-κB p65 and plasma MCP-1 levels in DNR and DR patients with different clinical factors.

Parameters/groups	Antihyperglycemic medication		Antihypertensive medication	
	yes	no	yes	no
NF-κB p65 (AU/mg)				
DNR	5.0 ± 1.6 ^a (107)	6.1 ± 3.2 (50)	4.9 ± 1.3 ^a (104)	6.1 ± 3.2 (50)
DR	6.1 ± 2.8 ^b (130)	8.1 ± 5.8 (70)	6.2 ± 3.3 ^b (119)	8.1 ± 5.8 (70)
NPDR	6.0 ± 2.9 ^c (85)	10.4 ± 8.1 (40)	5.2 ± 1.6 ^c (74)	10.4 ± 8.1 (40)
PDR	5.6 ± 1.9 ^d (45)	7.1 ± 4.5 (30)	5.2 ± 1.4 ^d (45)	7.1 ± 4.5 (30)
MCP-1 (pg/ml)				
DNR	13.4 ± 11.1 ^a (107)	24.7 ± 19.8 (50)	12.1 ± 9.0 ^a (104)	24.7 ± 19.8 (50)
DR	18.7 ± 14.4 ^b (130)	30.5 ± 18.8 (70)	16.8 ± 12.1 ^b (119)	30.5 ± 18.8 (70)
NPDR	20.8 ± 18.5 ^c (85)	28.3 ± 19.8 (40)	20.4 ± 15.6 ^c (74)	28.3 ± 19.8 (40)
PDR	15.6 ± 10.4 ^d (45)	27.1 ± 17.3 (30)	18.6 ± 13.1 ^d (45)	27.1 ± 17.3 (30)

The data are expressed as the mean ± SD; the number in the parentheses indicates the number of subjects. DNR, diabetic non-retinopathy; DR, diabetic retinopathy; MCP-1, monocyte chemoattractant-1; NF-κB p65, nuclear factor kappa B p65; NPDR, non-proliferative diabetic retinopathy; and PDR, proliferative diabetic retinopathy. ^ap < 0.05 versus DNR with no antihyperglycemic and antihypertensive medication; ^bp < 0.05 versus DR with no antihyperglycemic and antihypertensive medication; ^cp < 0.05 versus NPDR with no antihyperglycemic and antihypertensive medication; ^dp < 0.05 versus PDR with no antihyperglycemic and antihypertensive medication.

**Table 3** - Pearson correlations between NF-κB, MCP-1 and several clinical parameters of different clinical groups.

Biochemical parameters	Pearson r	p-value
Ctrl		
NF-κB p65/MCP-1	0.19	<0.01
DNR		
NF-κB p65/MCP-1	0.18	<0.05
NF-κB p65/HbA _{1c}	0.18	<0.05
NF-κB p65/diabetes duration	0.20	<0.05
NF-κB p65/triglycerides	0.19	<0.05
MCP-1/HDL-C	-0.17	<0.05
MCP-1/diabetes duration	0.16	<0.05
MCP-1/SBP	0.17	<0.05
DR		
NF-κB p65/diabetes duration	0.17	<0.05
MCP-1/HbA _{1c}	0.25	<0.01
MCP-1/diabetes duration	0.26	<0.001
MCP-1/retinopathy duration	0.16	<0.05

Ctrl, healthy controls; DNR, diabetic non-retinopathy; DR, diabetic retinopathy; HbA_{1c}, glycated hemoglobin; HDL-C, high-density lipoprotein cholesterol; MCP-1, monocyte chemoattractant protein-1; NF-κB p65, nuclear factor kappa B p65; and SBP, systolic blood pressure.

migration and altered retinal blood flow (5). Previous reports have shown that the local expression of MCP-1 and NF-κB in the retina as well as their levels in the vitreous

fluids are increased in PDR patients (17-18). This study aimed to assess the circulating levels of activated NF-κB p65 in PBMCs and plasma MCP-1 in type 2 DM patients with retinopathy who were taking antihyperglycemic and anti-hypertensive drugs. The basal levels of plasma MCP-1 and NF-κB p65 in the PBMCs of healthy controls in this study were comparable to previously observed values (19-20). NF-κB and MCP-1 could be involved in the pathogenesis of DR due to the findings of significantly elevated levels of these markers in non-medicated DR patients, and the plasma MCP-1 level was positively correlated with retinopathy duration.

In addition, both DM duration and HbA_{1c} were found to be positively correlated with both of the inflammatory markers in DR patients. This confirms that prolonged hyperglycemia triggers inflammation, possibly by causing abnormal local leukocyte-endothelial interaction in DR patients. Inflammatory chemokines are also potential angiogenic factors (21). Therefore, MCP-1 may act with vascular endothelial growth factor (5) to promote angiogenesis in the retina and cause neovascularization in PDR. The positive correlation between the NF-κB p65 and plasma MCP-1 levels in the healthy controls and DNR patients further supports the regulatory role of NF-κB on MCP-1. Inflammatory chemokines, including MCP-1, also activate NF-κB, leading to the generation of reactive oxygen species and creating a vicious cycle (7). This study also showed that

Table 4 - Risk assessment for DR in type 2 DM patients.

Characteristics	Unadjusted model		Adjusted model ^a	
	OR (95% CI)	p-value	OR (95% CI)	p-value
Age (years)	0.96 (0.93-1.00)	0.052		
Gender Female	1.00 (reference)			
Male	0.87 (0.58-1.31)	0.511		
BMI (kg/m ²)	0.96 (0.92-1.01)	0.099		
SBP (mmHg)	1.01 (1.00-1.02)	0.274		
DBP (mmHg)	1.00 (0.98-1.02)	0.736		
Diabetes duration (years)	1.08 (1.05-1.11)	0.000	1.08 (1.04-1.13)	0.000
HbA _{1c} (%)	1.34 (1.18-1.53)	0.000	1.20 (1.00-1.44)	0.046
Total cholesterol (mmol/l)	1.25 (1.04-1.49)	0.018	1.46 (0.76-2.79)	0.251
Triglycerides (mmol/l)	1.25 (0.96-1.63)	0.100		
HDL-C (mmol/l)	1.02 (0.53-1.95)	0.952		
LDL-C (mmol/l)	1.25 (1.01-1.54)	0.045	0.68 (0.33-1.42)	0.308
ALT (IU/l)	1.00 (0.99-1.01)	0.998		
AST (IU/l)	1.00 (0.99-1.02)	0.615		
Current Smokers No	1.00 (reference)			
Yes	0.51 (0.30-0.87)	0.013	0.40 (0.15-1.03)	0.057
Alcohol intake No	1.00 (reference)			
Yes	0.58 (0.31-1.07)	0.081		
Hypertension No	1.00 (reference)			
Yes	0.79 (0.49-1.28)	0.347		
Insulin antihyperglycemic medication				
No	1.00 (reference)			
Yes	0.74 (0.37-1.52)	0.000	0.63 (0.16-4.66)	0.018
Oral antihyperglycemic medication				
No	1.00 (reference)			
Yes	0.67 (0.40-1.12)	0.003	0.81 (0.50-1.78)	0.033
Antihypertensive medication				
No	1.00 (reference)			
Yes	0.56 (0.37-0.86)	0.008	0.79 (0.40-1.58)	0.040
NF-κB p65 (AU/mg)	1.11 (1.01-1.18)	0.005	1.08 (0.98-1.19)	0.030
MCP-1 (pg/ml)	1.05 (1.03-1.06)	0.000	1.04 (1.02-1.07)	0.002

Type 2 DM patients (DNR and DR) (n = 371) were included in the analysis model. ALT, alanine aminotransferase; AST, aspartate aminotransferase; BMI, body mass index; CI, confidence interval; DBP, diastolic blood pressure; DR, diabetic retinopathy; HbA_{1c}, glycated hemoglobin; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; MCP-1, monocyte chemoattractant protein-1; NF-κB p65, nuclear factor kappa B p65; OR, odds ratio; and SBP, systolic blood pressure. ^a Adjusted for age, gender and metabolic risk factors (BMI, HbA_{1c} and DM duration).



NF- κ B p65 was positively correlated with the triglyceride levels and that the plasma MCP-1 levels were inversely associated with the HDL-C levels in DNR patients. The above observations are in agreement with a previous report suggesting the possible involvement of MCP-1 in obesity-related health complications (8).

The 1998 United Kingdom Prospective Diabetes Study, involving 3,867 type 2 DM patients, demonstrated that early intervention with oral glucose-lowering and antihypertensive drugs reduced the incidence of diabetic complications and improved the survival of these patients (9-10). However, there was no evidence to show whether this beneficial effect was derived directly from tight glycemic and blood pressure control or through other means of action. In this study, approximately 60% of the DM patients received antihyperglycemic and antihypertensive medications. The patient groups (DNR, DR, NPDR and PDR) who were taking either antihyperglycemic (insulin, biguanides, sulfonylureas and thiazolidinediones) or antihypertensive drugs (angiotensin-converting enzyme inhibitors, calcium channel blockers and angiotensin II receptor antagonists) had significantly lower levels of NF- κ B p65 and plasma MCP-1 compared with the non-medicated group. These findings may explain the beneficial effects of the antihyperglycemic and antihypertensive drugs on diabetic microvascular complications that were observed in the United Kingdom Prospective Diabetes Study. However, this study did not investigate the local effect of these medications on the expression of NF- κ B and MCP-1 in the retina of DR patients. A previous report showed that a reduction of local retinal inflammation and the NF- κ B and MCP-1 expression levels was found in cultured murine endothelial cells treated with antihypertensive drugs (22). Thus, we speculate that the vitreous levels of NF- κ B and MCP-1 in the DR patients are concurrently reduced with the circulating levels of these pro-inflammatory markers.

This transversal study could not identify the actual mechanism by which antihypertensive drugs ameliorate DR. A local renin-angiotensin system exists in retinal glial cells (23), and it is upregulated in DR (24). Angiotensin II is a well-known vasoconstrictor, and it may induce vascular endothelial growth factors that lead to retinal neovascularization (24), a common microscopic sign in PDR. We speculate that the beneficial effect of antihypertensive treatments on DR could partly be due to blocking of the renin-angiotensin system. In addition, a cross-interaction between the advanced glycation end product (AGE)-receptor for AGE (RAGE) and the renin-angiotensin system has been proposed in the pathogenesis of DR (25). Previously, we have shown that RAGE is a pertinent factor leading to DR, and its genetic variants are associated with the development of DR (26). Antihypertensive drugs may reduce the AGE-RAGE interaction by attenuating the NF- κ B that controls the cellular expression of RAGE. It should be noted that NF- κ B is not the only regulator of diabetes-induced inflammation in DR. The presence of hypoxia in DR could activate another transcription factor, hypoxia inducible factor 1 (27), which was not investigated in this study.

In conclusion, this study showed significant attenuation of both the PBMC NF- κ B p65 and circulating MCP-1 levels in DR patients who were taking antihyperglycemic and antihypertensive drugs. Nevertheless, a prospective case-control study is required to substantiate our findings.

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■ AUTHOR CONTRIBUTIONS

Ng ZX, Kuppasamy UR and Chua KH carried out the experiments, data analysis and manuscript preparation. Tajunisah I, Pendek R, Ng ZX, Kuppasamy UR and Chua KH were involved in experimental design, sample collection, and grant and ethics applications.

■ REFERENCES

1. Uthra S, Raman R, Mukesh BN, Rajkumar SA, Padmaja KR, Paul PG, et al. Association of vegf gene polymorphisms with diabetic retinopathy in a south indian cohort. *Ophthalmol Genet.* 2008;29(1):11-5, <http://dx.doi.org/10.1080/13816810701663527>.
2. Liew G, Klein R, Wong TY. The role of genetics in susceptibility to diabetic retinopathy. *Int Ophthalmol Clin.* 2009;49(2):35-52, <http://dx.doi.org/10.1097/IIO.0b013e31819fd5d7>.
3. Lamoureux EL, Tai ES, Thumboo J, Kawasaki R, Saw SM, Mitchell P, et al. Impact of diabetic retinopathy on vision-specific function. *Ophthalmology.* 2010;117(4):757-65, <http://dx.doi.org/10.1016/j.ophtha.2009.09.035>.
4. Cai J, Boulton M. The pathogenesis of diabetic retinopathy: Old concepts and new questions. *Eye (Lond).* 2002;16(3):242-60, <http://dx.doi.org/10.1038/sj.eye.6700133>.
5. Ozturk BT, Bozkurt B, Kerimoglu H, Okka M, Kamis U, Gunduz K. Effect of serum cytokines and vegf levels on diabetic retinopathy and macular thickness. *Mol Vis.* 2009;15(12):1906-14.
6. Cheung N, Mitchell P, Wong TY. Diabetic retinopathy. *Lancet.* 2010;376(9735):124-36, [http://dx.doi.org/10.1016/S0140-6736\(09\)62124-3](http://dx.doi.org/10.1016/S0140-6736(09)62124-3).
7. Ho E, Bray TM. Antioxidants, nfkappab activation, and diabetogenesis. *Proc Soc Exp Biol Med.* 1999;222(3):205-13, <http://dx.doi.org/10.1046/j.1525-1373.1999.d01-137.x>.
8. Christiansen T, Richelsen B, Bruun JM. Monocyte chemoattractant protein-1 is produced in isolated adipocytes, associated with adiposity and reduced after weight loss in morbid obese subjects. *Int J Obes (Lond).* 2005;29(1):146-50, <http://dx.doi.org/10.1038/sj.ijo.0802839>.
9. Effect of intensive blood-glucose control with metformin on complications in overweight patients with type 2 diabetes (ukpds 34). *Uk prospective diabetes study (ukpds) group. Lancet.* 1998;352(9131):854-65.
10. Efficacy of atenolol and captopril in reducing risk of macrovascular and microvascular complications in type 2 diabetes: Ukpds 39. *Uk prospective diabetes study group. BMJ.* 1998;317(7160):713-20.
11. Chaturvedi N, Porta M, Klein R, Orchard T, Fuller J, Parving HH, et al. Effect of candesartan on prevention (direct-prevent 1) and progression (direct-protect 1) of retinopathy in type 1 diabetes: Randomised, placebo-controlled trials. *Lancet.* 2008;372(9647):1394-1402, [http://dx.doi.org/10.1016/S0140-6736\(08\)61412-9](http://dx.doi.org/10.1016/S0140-6736(08)61412-9).
12. Sjolie AK, Klein R, Porta M, Orchard T, Fuller J, Parving HH, et al. Effect of candesartan on progression and regression of retinopathy in type 2 diabetes (direct-protect 2): A randomised placebo-controlled trial. *Lancet.* 2008;372(9647):1385-93, [http://dx.doi.org/10.1016/S0140-6736\(08\)61411-7](http://dx.doi.org/10.1016/S0140-6736(08)61411-7).
13. Diabetic retinopathy study. Report number 6. Design, methods, and baseline results. Report number 7. A modification of the airle house classification of diabetic retinopathy. Prepared by the diabetic retinopathy. *Invest Ophthalmol Vis Sci.* 1981;21(1 Pt 2):1-226.
14. Grading diabetic retinopathy from stereoscopic color fundus photographs—an extension of the modified airle house classification. *Etdrs report number 10. Early treatment diabetic retinopathy study research group. Ophthalmology.* 1991;98(5 Suppl):786-806.
15. Boyum A. Separation of leukocytes from blood and bone marrow. *Introduction. Scand J Clin Lab Invest Suppl.* 1968;97:7.
16. Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem.* 1976;72:248-54, [http://dx.doi.org/10.1016/0003-2697\(76\)90527-3](http://dx.doi.org/10.1016/0003-2697(76)90527-3).
17. Meleth AD, Agron E, Chan CC, Reed GF, Arora K, Byrnes G, et al. Serum inflammatory markers in diabetic retinopathy. *Invest Ophthalmol Vis Sci.* 2005;46(11):4295-301, <http://dx.doi.org/10.1167/iovs.04-1057>.
18. Harada C, Okumura A, Namekata K, Nakamura K, Mitamura Y, Ohguro H, et al. Role of monocyte chemoattractant protein-1 and nuclear factor kappa b in the pathogenesis of proliferative diabetic retinopathy. *Diabetes Res Clin Pract.* 2006;74(3):249-56, <http://dx.doi.org/10.1016/j.diabetes.2006.04.017>.



19. Hashimoto S, Nakayama T, Gon Y, Hata N, Koura T, Maruoka S, et al. Correlation of plasma monocyte chemoattractant protein-1 (mcp-1) and monocyte inflammatory protein-1 α (mip-1 α) levels with disease activity and clinical course of sarcoidosis. *Clin Exp Immunol*. 1998;111(3):604-10, <http://dx.doi.org/10.1046/j.1365-2249.1998.00519.x>.
20. Hofmann MA, Schiekofer S, Isermann B, Kanitz M, Henkels M, Joswig M, et al. Peripheral blood mononuclear cells isolated from patients with diabetic nephropathy show increased activation of the oxidative-stress sensitive transcription factor nf-kappab. *Diabetologia*. 1999;42(2):222-32, <http://dx.doi.org/10.1007/s001250051142>.
21. Wakabayashi Y, Usui Y, Okunuki Y, Kezuka T, Takeuchi M, Iwasaki T, et al. Increases of vitreous monocyte chemotactic protein 1 and interleukin 8 levels in patients with concurrent hypertension and diabetic retinopathy. *Retina*. 2011;31(9):1951-57, <http://dx.doi.org/10.1097/IAE.0b013e31820d3cee>.
22. Nagai N, Izumi-Nagai K, Oike Y, Koto T, Satofuka S, Ozawa Y, et al. Suppression of diabetes-induced retinal inflammation by blocking the angiotensin ii type 1 receptor or its downstream nuclear factor-kappab pathway. *Invest Ophthalmol Vis Sci*. 2007;48:4342-50, <http://dx.doi.org/10.1167/iovs.06-1473>.
23. Sarlos S, Wilkinson-Berka JL. The renin-angiotensin system and the developing retinal vasculature. *Invest Ophthalmol Vis Sci*. 2005;46(9):1069-77, <http://dx.doi.org/10.1167/iovs.04-0885>.
24. Fletcher EL, Phipps JA, Ward MM, Vessey KA, Wilkinson-Berka JL. The renin-angiotensin system in retinal health and disease: Its influence on neurons, glia and the vasculature. *Prog Retin Eye Res*. 2010;29(4):284-311, <http://dx.doi.org/10.1016/j.preteyeres.2010.03.003>.
25. Yamagishi S, Nakamura K, Matsui T, Ueda S, Fukami K, Okuda S. Agents that block advanced glycation end product (age)-rage (receptor for ages)-oxidative stress system: A novel therapeutic strategy for diabetic vascular complications. *Expert Opin Investig Drugs*. 2008;17(7):983-96, <http://dx.doi.org/10.1517/13543784.17.7.983>.
26. Ng ZX, Kuppusamy UR, Tajunisah I, Fong KC, Koay AC, Chua KH. 2245g/a polymorphism of the receptor for advanced glycation end-products (rage) gene is associated with diabetic retinopathy in the malaysian population. *Br J Ophthalmol*. 2012;96(2):289-92, <http://dx.doi.org/10.1136/bjophthalmol-2011-300658>.
27. Treins C, Giorgetti-Peraldi S, Murdaca J, Monthouel-Kartmann MN, Van Obberghen E. Regulation of hypoxia-inducible factor (hif)-1 activity and expression of hif hydroxylases in response to insulin-like growth factor. *Mol Endocrinol*. 2005;19(5):1304-17, <http://dx.doi.org/10.1210/me.2004-0239>.