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1 Adaptive significance of functional germination traits in crop wild relatives of

2 Brassica

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12 Abstract

Functional germination traits contribute to both niche competitiveness and crop yield 13 14 outcomes. However, there is little understanding of the adaptive significance of the 15 germination thermal- and hydro-parameters in crop wild relatives (CWRs), yet these species are anticipated to be the source of adaptive traits for future agriculture. Seeds of 16 10 lots of Brassica species, sub-species and inbred lines from across Europe, North 17 Africa and the Middle East were subjected to a range of temperature and water potential 18 19 conditions. The germination progress curves recorded were analysed using repeated probit analysis and the functional trait parameters (thermal- and hydro thresholds and 20 21 times) determined. Relationships between these seed parameters (and the physical trait, 22 seed mass) and the seed source environment were investigated.

23 The Brassica genus was found to have diverse seed germination phenotypes, with thermal (θ_T) and hydro times (θ_H) differing by 3 to 7-fold, base temperatures (T_b) by c. 9 24 25 °C and base water potentials ($\Psi_{\rm b}$) by -1.5 MPa. Crop seed lots of *Brassica oleracea* had shorter $\theta_{\rm H}$ for germination and higher values of $\Psi_{\rm b}$, but longer $\theta_{\rm T}$ for germination than 26 27 their CWR. For the CWRs, the mean monthly precipitation or the precipitation of the predicted month of germination of the seed collection site, was linearly correlated with 28 29 T_b , θ_T , and Ψ_b . This increases the predictability of identifying valuable brassica germplasm for crop development through regeneration trait screening. 30

In conclusion, the selection of the crop *B. oleracea* for fastest germination under

- 32 irrigated conditions likely limits the capacity of the current genepool to cope with
- 33 erratic periods of drought predicted in future European climates.

Keywords: Seed functional traits; thermal time; hydro time; vigor; precipitation; seed
mass.

36 **1. Introduction**

Global climate change is predicted to increase environmental temperatures, but may 37 have other consequences such as earlier frosts and changes to the frequency of flooding 38 and drought (IPCC, 2013), all of which may affect the outcome of germination and 39 seedling emergence. In particular, warming temperatures and fluctuations in 40 precipitation are known to increase the risk to yield in the most economically important 41 42 crops, such as wheat, rice, maize and barley (Lobell and Field, 2007; Porter and 43 Semenov, 2005). Moreover, crops bred for predictable and more uniform germination and harvest may contribute to loss of variability in responses to environmental 44 45 conditions (Gepts, 2010). In contrast, crop wild relatives (CWRs) represent a genepool that is potentially adapted to a large range of habitats and, therefore, likely better 46 47 equipped to survive the challenges associated with climate change (Dempewolf et al., 2014). In addition to possessing high genetic diversity, CWR species may also have 48 49 greater plasticity in traits enabling them to cope better with rapid environmental change 50 (Nicotra et al., 2010).

Global plant variability can be explained by just six adult plant-functional traits, only
one of which relates to the regenerative phase, specifically diaspore mass (Díaz et al.,
2016). This seed physical trait has been shown to be predictive of a light requirement
for germination, persistence in the soil and dispersal (Baskin and Baskin, 2014).
However, increasing importance is being attributed to germination functional traits such
as germination rate, thermal time and hydro time, determined by the quantification of
the physiological process of germination as a descriptor of regeneration potential.

Germination rate is an important and widely used descriptor of seed vigour (Pollock and
Roos 1972). These traits are now recognised as critical components of the multiple
environmentally regulated factors that define the ecological niche for population growth
(Dürr et al., 2015; Fernández-Pascual et al., 2015; Grubb, 1977; Poschlod et al., 2013)

Recent characterisation of germination functional traits (base temperature, T_b; and 62 63 thermal time, θ_T) in CWRs of grapevine have identified populations at greater risk of 64 climate change on the basis of insufficient cold to remove seed dormancy (Orru et al., 2012). In consideration of future proofing agriculture in France, 36 cover crops (across 65 six plant families including the *Brassicaceae*) have been characterised for germination 66 functional traits (Tribouillois et al., 2016). Four Brassicas (B. juncea, B. rapa, B. napus, 67 B. carinata) had T_b of c. 7°C and three species had similar base water potentials, Ψ_b , for 68 69 germination (-0.9 to -1 MPa), the exception being B. rapa (-2.2 MPa) (Tribouillois et 70 al., 2016). These findings lend weight to the argument that a species geographical origin 71 defines its ability to germinate (Cochrane et al., 2014; Dürr et al., 2015).

The convergence of functional traits in species occupying similar environments can be 72 73 considered an adaptation to environmental filtering (Keddy, 1992). Precipitation and 74 temperature are the main environmental factors that have an impact on plant traits, e.g., 75 leaf mass, yield, height and flowering time; and seed traits, such as seed mass, 76 germination and dormancy, show similar ecological associations (Ackerly et al., 2000; 77 Baskin and Baskin, 1998; Dornbos and Mullen, 1991; Franks et al., 2007; Menzel et al., 2006; Peñuelas et al., 2004; Porter, 2005). Seed germination (i.e., radicle emergence) 78 79 and seedling establishment are thought to be the most sensitive stage of a plant to 80 environmental changes (Fay and Schultz, 2009; Kimball et al., 2010; Lloret et al., 81 2004). Consequently, the effects of both precipitation and temperature on germination

rate (Céspedes et al., 2012; Clauss and Venable, 2000; Levine et al., 2008; Meyer et al.,
1990), final germination (Alexander and Wulff, 1985; Gareca et al., 2012; Gutterman,
2000b) and seedling establishment (Cochrane et al., 2015; Jump et al., 2008; Lloret et
al., 2004; Smith et al., 2000) have been widely explored.

86 Crop domestication has resulted in larger seed size (Preece et al., 2017) and seed mass

is also known to be responsive to environmental fluctuations (Donohue et al., 2005;

Nicotra et al., 2010; Roach and Wulff, 1987). In particular, seed filling is influenced by

89 environmental conditions and mean seed mass of a population has been positively

90 correlated with annual rainfall (Harel et al., 2011) or with mean annual temperature

91 (Murray et al., 2004). Whilst seed size is thought to affect germination rate (Norden et

al., 2009), seed mass is rarely co-analysed with seed physiological traits of germination

base temperature (T_b) or base water potential (Ψ_b) . In this context, seed mass and T_b of

94 tree seeds of *Aesculus hippocastanum* from across Europe had no correlation (Daws et

al., 2004), and a negative correlation was found between seed mass and Ψ_b of

96 neotropical forest species (Daws et al., 2008).

97 Our aim was to investigate how the originating environment of *Brassica* CWRs impact

98 on their germination functional traits, testing the assumption that current *Brasssica*

99 crops may have been selected for fast germination under managed, irrigated conditions.

100 We used seven geo-referenced seed collections (from across Europe, North Africa and

the Middle East) and three commerically available crop seed lots and research

102 genotypes. The genus *Brassica* possesses a wide diversity in morphology, genetics

103 (Arias et al., 2014), environment stress tolerance and is economically important around

the world for its edible roots crops, vegetables and oilseeds (Tsunoda et al., 1980). We

selected wild taxa from locations with variable mean monthly precipitation (1 - 94 mm)

- and temperature (5 25 °C). We show an intimate relationship between *Brassica* seed
- 107 collection site and germination functional traits.

108 **2. Material and Methods**

109 2.1 Seed material

110	Studies were performed on seven seed lots of non-dormant wild brassica: three species
111	(Brassica rapa, B. nigra and B. tournefortii) and two sub-species (Brassica rapa subsp.
112	campestris and B. rapa subsp. sylvestris). (Table 1A). These CWR seed lots were
113	obtained from the Millennium Seed Bank, Royal Botanic Gardens, Kew, UK. Seed lots
114	used were from across Europe, North Africa and the Middle East. Selection was based
115	on an environmental cline typical of <i>Brassica</i> species: annual mean temperature from 5
116	to 26 °C; and a mean monthly precipitation from 1 mm to 94 mm, in seed lots with
117	sufficient quantity of seeds. The values were calculated as the mean monthly
118	precipitation and annual mean temperature, i.e. the sum of all 12-monthly means,
119	divided by 12. Seeds were handled in a consistent way on receipt at the seed bank:
120	equilibration to 15 % RH and 15 $^{\circ}$ C, hermetical sealing in containers and then storage at
121	-20 °C (FAO/IPGRI, 1994). On withdrawal from the bank, seeds were re-equilibrated to
122	15 °C in a room operating at c. 15 % RH.

123 A commercial seed lot of *B. oleracea* (cabbage) was obtained from B&T World Seeds

124 (France) (Table 1B) and seeds of two research genotypes derived from *B. oleracea*

125 crops (A12DHd from kale and AGSL101from kale and broccoli) were provided by the

- 126 University of Warwick. Both crop research genotypes have the same genetic
- background (A12) but SL101 has introgressions from at least two QTL (RABA1 and
- SOG1) from a faster germinating genotype (GD33DH, broccoli) that confers higher

- vigour (Morris et al., 2016). For simplicity, henceforth A12DHd and AGSL101 will be
- 130 called RG_L (low vigour research genotype) and RG_H (high vigour research
- 131 genotype). The commercial genotype will be called CG. Both crop research genotypes
- 132 were grown under the same controlled conditions at 18/22 °C (night/day) in a
- 133 glasshouse to produce seeds. On receipt at RBG Kew, all seed lots were equilibrated to
- 134 15 % RH and 15 °C and stored under these conditions for a few weeks until use.

А.			Annua	al mean temp	perature	
Species	Collection site	Mean monthly precipitation (mm)	Min T (°C)	Mean T (°C)	Max T (°C)	
B. rapa (CWR)	Chur	94.17	4.8	9.2	13.7	
/	(Switzerland)				perature Max T (°C) 13.7 13.8 15.4 11.5 20.4 27.1 34.0	
B. nigra (CWR)	Dorset	68.83	6.2	9.9	13.8	
	(England)					
B. rapa (CWR)	Memsault	64.08	6.5	10.9	15.4	
/	(France)					
<i>B. rapa</i> subsp.	Göle-Kars	40.84	-1.6	4.8	11.5	
campestris (CWR)	(Turkey)					
<i>B. rapa</i> subsp.	Ait Marghad	26.8	3.8	12.1	20.4	
sylvestris (CWR)	(Morocco)					
B. tournefortii	Natrun –	4.83	14.3	20.7	27.1	
(CWR)	Alamin (Egypt)					
<i>B. rapa</i> subsp.	S. Oran	0.83	17.5	25.7	34.0	
sylvestris (CWR)	(Algeria)					

Growth conditions

	Provider	Genotype	Vigour	Day (°C)	Night (°C)
<i>B. oleracea</i> (crop)	B&T World Seeds (France)	CG (Commercial seed lot)	Unknown	Unknown	Unknown
<i>B. oleracea</i> (crop research genotype)	University of Warwick (UK)	RG_H (AGSL101)	High	22	18
<i>B. oleracea</i> (crop research genotype)	University of Warwick (UK)	RH_L (A12DHd)	Low	22	18

Table 1. Details of the 10 *Brassica* seed lots used. (A) The environment information
was obtained by combining seed collection geo-locations (GIS coordinates) for the *Brassica* crop wild relatives (CWRs) and WorldClim data, as historical annual mean
temperature (T) and mean monthly precipitation since 1960. (B) Plant growth and seed
production conditions are shown for the research genotype seed lots, but are unknown
for the commercially purchased *B. oleracea* crop seed lot.
2.2 Seed mass and germination

- 142 To characterise the seed mass of the *Brassica* seed lots, 100 seeds were individually
- 143 weighed to 0.0001 mg accuracy. The distribution of seed mass at 0.5 mg intervals was
- analysed and the mean and coefficient of variation (CV) calculated.
- 145 For germination, seeds were sown at constant temperatures from 5 to 45 $^{\circ}$ C, at 5 $^{\circ}$ C
- intervals. Seeds were also sown at a range of water potentials (0, -0.3, -0.5, -0.8 and -1.0

MPa) using water and increasing concentrations of polyethylene glycol 8000 (PEG, 147 148 Fisher Scientific, UK) solution, but only at constant temperatures of 25 and 30 °C. Due to limited seed availability, *B. nigra* seeds were sown under constant temperatures 149 150 without water stress. Either three replicates of 25 seeds, for the CWRs, or four replicates of 25 seeds for the crops, were used in each treatment. Seeds were sown onto two layers 151 of germination test paper (90 mm diameter; Fisher Scientific, UK) in Petri dishes. The 152 153 volume of PEG solution used (7 mL) was calculated following the equation of 154 Hardegree and Emmerich (1990) to account for the weight of the dry germination paper. The water potential of the solutions was confirmed with an osmometer (Camlab, UK). 155 156 For consistency, the same volume (7 mL) of solution was used for all PEG and water treatments. The vented Petri dishes were placed in sealable plastic bags and incubated at 157 the relevant temperature with a 12 hour photoperiod (radiometric flux density of 50-100 158 159 W/m^2). Germination (radicle emergence of 2 mm) was recorded every hour during the 160 first 24 hours and then every two-three hours until no more germination was observed. 161 Non-germinated seeds that were still firm after PEG treatment were briefly washed with 162 distilled water to remove the PEG and transferred to a new Petri dish with germination test paper and 7 mL of water. After 15 days in this recovery assessment, if the seeds did 163 164 not germinate, the seed coat was removed and the viability was tested with 1 % 165 triphenyl tetrazolium chloride (TZ) in the darkness at 30 °C (ISTA, 2003) for 18 hours. Stained red embryos were considered to be viable. 166

167 2.3 Data analysis

Germination was quantified as final germination percentage and germination rate (GR).
Germination progress over time was represented as a sigmoidal curve (fitted using the
Boltzmann distribution) from which the GR was estimated for all treatments in each

population. The GR at the 50th percentile (t50) is the reciprocal of time the population 171 172 needs to reach 50 % germination of viable seeds. The seeds were germinated at a range of temperatures and the GR describes a positive (sub-optimal range of temperatures) or 173 174 negative (supra-optimal range of temperatures) regression line when plotted against temperature. The intercept of both lines is the optimal temperature (T_0) where the GR is 175 176 estimated to be at a maximum. Base temperature (T_b) and ceiling temperature (T_c) are 177 the intercept of the lines when GR is estimated to be zero, and germination does not 178 proceed to completion when seeds are below T_b and above T_c (García-Huidobro et al., 1982). Thermal time (θ_T) was estimated by repeated probit analysis in GenStat 12.1 179 180 software (VSN International Ltd, 2009). This method consisted of varying T_b until the best fit was obtained (minimum residual) (Bradford, 1995; Ellis et al., 1986) based on 181 182 the following two equations:

183 Sub-optimal θ_T is Probit(g)= K + (log $\theta_T(g)$)/ $\sigma_{\theta T}$ (Bradford, 1995; Ellis et al., 1986) (1)

184 Supra-optimal θ_{T} is Probit(g)= K + $\theta_{Tsupra}(g)/\sigma_{\theta T}$ (Covell et al., 1986; García-Huidobro 185 et al., 1982). (2)

186 In both equations, K is the intercept constant when θ_{Tsupra} is zero. θ_{T} is the heat units the 187 seed needs to accumulate for a given percentile (g) to complete germination in

188 chronological time and σ is the standard deviation of the seed population response.

189 The basis of estimating hydro-time is the positive regression line between the GR and

190 water potentials. The base water potential (Ψ_b) is calculated as the intercept of the

191 regression line of GR on water potential. Hydro time (θ_H) was also estimated by

192 repeated probit analysis. In this case, the method consisted of varying $\theta_{\rm H}$ until the best

193 fit was obtained (minimum residual) (Bradford, 1995) using equation 3:

where K is the intercept constant, Ψ_b is the base water potential (below this water potential no germination is predicted to occur) for a given percentile (g) and σ is the standard deviation.

Additionally, analysis of variance (ANOVA) and t-tests were used to compare the seed
mass and seed germination parameters between wild seed lots and between crop seed
lots.

201 2.4. Environmental parameters associated with seed lot provenance

202 The original, geo-referenced location of the CWR seed lots was used to extract climate 203 data from WordClim with an accuracy of one kilometre for historical data over the 204 period 1960 – 1990 (Hijmans et al., 2005). Associations between germination 205 parameters and the following environmental factors were assessed: annual mean temperature (minimum, maximum and median in °C), mean monthly precipitation 206 207 (mm), mean precipitation and temperature of the month of germination and the altitude. The month of germination was the first month after seed collection date that the 208 209 following assumptions were met: (1) the minimum rainfall required for germination is a 210 monthly mean of 15 mm (Freas and Kemp, 1983; Gutterman, 1993; Gutterman, 2000a) and (2) the temperature exceeded T_b but did not exceed T_c (Figure S1). The exception 211 212 was *B. rapa* subsp. *sylvestris* from Algeria where the maximum monthly precipitation 213 did not exceed 2 mm (Figure S1). For this species, the first period of significant rainfall was taken as the precipitation threshold. In addition, correlations between the 214 215 environmental factors and the seed germination parameters of the wild species, θ_T , θ_H

time, T_b and Ψ_b were tested with scatter matrix and linear correlations in Origin 9.0 software (OriginLab Corporation, 2013).

218 **3. Results**

The seeds of all *Brassica* seed lots had high viability. Seed germination of the CWRs
varied from 93 % (*B. rapa*) to 100 % (*B. tournefortii*) and seeds of the crop genotypes

- of *B. oleracea* reached 100 % germination.
- 222 3.1 Germination trait parameters

223 Seed germination of the 10 *Brassica* seed lots (CWRs and crops) showed a c. 9 °C

range in each of the cardinal temperatures: T_b from 1.7 to 10.5 °C; T_o from 25 to 35 °C;

and T_c from 36 to 45 °C (Figures 1A, C, E and G). Intra-specific variability in trait

parameters was observed in two wild seed lots of *B. rapa* from France and Switzerland

227 (Figure 1C) and two wild seed lots of *B. rapa* subsp. *sylvestris* from Morocco and

Algeria (Figure 1E), with significant differences (P < 0.05) in germination rate (GR)

and T_b (Table S1). Thermal time (θ_T) values also differed three-fold amongst CWRs

230 (from 214.7 °Ch for *B. rapa* subsp. *sylvestris* from Morocco to 775.5 °Ch for *B. rapa*

from Switzerland) and the crop research genotype *B. oleracea* RG_L had the longest θ_T

232 of 951.6 °Ch (Table S1).

In general, seeds of the crop genotypes *B. oleracea* (RG_L, RG_H and CG) were slower

to germinate compared with the CWRs in terms of $\theta_{\rm T}$ in the sub-optimal range of

temperatures (Table S1). The high vigour crop genotype, RG_H, had faster germination

(i.e. shorter θ_T) than the low vigour crop genotype, RG_L (P < 0.01); whilst the CG had

 $\theta_{\rm T}$ very similar to the CWR *B. rapa* subsp. *campestris* (i.e., c. 445 °Ch). In contrast, T_b

- for the *B. oleracea* seed lots did not differ (Figure 1G). However, T_c was higher in the
- crop genotype RG_H and CG than the genotype RG_L (40 °C and 41.5 °C vs 36 °C,
- 240 respectively, P < 0.001). In the supra-optimal temperatures, θ_{Tsupra} was shorter in CG,
- but the other two crop genotypes did not differ.
- 242 Seeds were also germinated under five water potentials. GR slowed and final
- 243 germination decreased with more negative water potentials, and Ψ_b values varied
- between -0.4 MPa and -2 MPa (Figure 1B, D, F and H). Both seed lots of the CWR B.
- 245 *rapa* (France, Switzerland) had the lowest Ψ_b of -1.59 MPa and -1.94 MPa respectively
- (Figure 1D). The hydro time ($\theta_{\rm H}$) values also differed among all CWRs from 9.0 MPah
- for *B. rapa* subsp. *campestris* to 72 MPah of *B. rapa* Switzerland (Table S1). At the end
- of the experiment, all non-germinated seeds were transferred to water and subsequently
- 249 germinated within 48 hours, except for *B. rapa* subsp. *sylvestris* (Algeria) which only
- achieved 40 % of germination after 15 days. However, a tetrazolium test showed that all
- the seeds of *B. rapa* subsp. *sylvestris* (Algeria) were viable.
- 252 Whilst the $\theta_{\rm H}$ of the crop seed lots was within the range of the CWRs, $\Psi_{\rm b}$ of the crop
- seed lots was higher (less negative) than the CWRs, except for *B. tournefortii* and *B.*
- 254 rapa subsp. campestris (Table S1). Within the crop seed lots, the high vigour genotype,
- 255 RG_H, had a larger $\theta_{\rm H}$ than RG_L (P < 0.05), with CG of *B. oleracea*, having the
- smallest θ_H (Table S1). In contrast, the Ψ_b was lower in both crop seed lots, RG_H and
- 257 CG, than in RG_L (-0.74 MPa in RG_H, -0.78 MPa in the CG and -0.40 MPa in
- 258 RG_L). Therefore, seeds of RG_H and CG could germinate over a wider range of water
- 259 potentials than RG_L (Figure 1H).



Figure 1: The germination rate (GR) was plotted against temperature (A, C, E and G)
to describe cardinal temperatures and the water potential thresholds (B, D, F and H) of
seven wild *Brassica* seed lots, two research genotypes of *B. oleracea* (RG_L and

263 RG_H) and the crop genotype CG. The seeds were germinated under a range of sub and

- supra optimal temperatures between 5 to 35 °C and on PEG solutions at 0, -0.3, -0.5, -
- 0.8 and -1.0 MPa at one temperature (between 20 and 25 °C). The regression lines were
- calculated from repeated probit analysis estimations and the error bars are the standard

- deviation (SD) of three replicates for each treatment in CWRs and four replicates forcrops.
- 269 Variation in seed mass of the CWRs was normally distributed, based on a normality test
- 270 (D'Agostino's test, P < 0.05) (Figure 2A). Two CWRs (*B. rapa* from France and *B.*
- 271 *nigra* from UK) had the highest variability (CV 36 %) and *B. tournefortii* had the
- smallest variability in seed mass (CV 18.3 %). The crop seed lot variability in mass was
- similar to that of the CWRs from CV 13% for CG to CV 30% for RG_L.
- Of the CWRs, mean seed mass of *B. rapa* from Switzerland was the largest (P < 0.05)
- (Figure 2A). In general, the crop seed lots had heavier seeds than the CWRs, with the
- seeds of RG_H heavier than those of RG_L and CG of *B. olereacea* (Figure 2B).



Figure 2. Seed mass of 100 individual seeds were shown as normal distribution for **A**)

278 CWRs, 1) *Brassica* rapa (Switzerland), 2) *B. nigra* (England), 3) *B. rapa* (France), 4) *B.*

- 279 *rapa* subsp. *campestris* (Turkey), 5) *B. rapa* subsp. sylvestris (Morocco), 6) *B.*
- 280 tournefortii (Egypt) and 7) B. rapa subsp. sylvestris (Algeria); and B) two research
- 281 genotypes of *B. oleracea*, RG_L (open stars) and RG_H (open circles) and one CG
- 282 (open squares). Each symbol is the mass of an individual seed.
- 283 3.2 Inter-relations between seed traits
- 284 The seed traits of all *Brassica* seed lots (including both crop and wild) were compiled
- and subjected to linear regression models. A negative relationship was found between
- 286 T_b and θ_T for all *Brassica* seed lots tested (P < 0.0001; r = -0.72, Figure 3A). Ψ_b and θ_H
- were also negatively correlated (P < 0.0001; r = -0.90 Figure 3B). In both cases the
- lower the base threshold (temperature or water potential) the longer the germination

- 289 process (thermal or hydro-time). A trend was found between the mean seed mass and
- 290 the θ_T and T_b (r = 0.53 and -0.61, respectively) so that heavier seeds need to accumulate
- 291 more θ_T to germinate and had a lower T_b , but it was not significant (P = 0.1 and 0.06,
- respectively). Furthermore, the CV of the seed mass was positively correlated with θ_T (P
- 293 < 0.05; r = 0.40), but not with T_b. Neither θ_H nor Ψ_b were correlated with mean seed
- 294 mass, however, θ_H was positively correlated with the CV of seed mass (r = 0.48; P <
- 295 0.01).



Figure 3. Correlations between seed traits of 10 *Brassica* seed lots (7 CWRs and 3

297 crops). A) Base temperature (T_b) and thermal time (θ_T); B) base water potential (Ψ_b) and

- hydro time (θ_H) and. The values of θ_T , T_b, θ_H and Ψ_b were calculated on the 50th
- 299 percentile. Each point represents a replicate of each population, three replicates for
- 300 CWRs and four replicates for crops. Asterisks indicating the significance at P <
- 0.001^{***} DF = 30 in A, 27 in B

302 3.3 Relations between environment and CWR seed traits

- 303 The dependencies of CWR functional seed traits on the collection site environment were
- assessed for the seven wild *Brassica* seed lots (Table S2). T_b was lower when the
- 305 monthly precipitation, and the precipitation of the predicted month of germination based
- 306 on the quantified responses, were higher (Figure 4A). Seed lots of species from wetter
- 307 environments also had slower germination (i.e., longer θ_T ; Figure 4B). The mean





Figure 4. Correlation of the seed germination traits of seven CWRs and the

environment of the seed collection site. Mean monthly precipitation (open symbols and

dashed line) and the precipitation of the month of germination (MoG) (solid symbols

and solid line) were plotted against the base temperature ($T_b A$), the thermal time (θ_T ,

B), the base water potential (Ψ_b , **C**) and the hydro time (θ_H , **D**). The values of θ_T , T_b , θ_H

and Ψ_b were calculated on the 50th percentile. Each point is one of the three replicates of

- each population. Asterisks indicating the significance at $P < 0.05^*$ and $P < 0.01^{**}$ DF =
- 322 5 in A and B, and 4 in C and D

323 **4. Discussion**

324 4.1. Comparative seed germination thresholds

The variation in Brassica germination thresholds (c. 9 °C) amongst 10 seed lots is wider 325 326 than that observed for other comparative studies based on taxonomy or habitat. For example, in cold and warm season grasses, T_b ranges by 2.6 °C to 5.9 °C, respectively 327 (Jordan and Haferkamp, 1989). Pasture species (e.g., clover, ryegrass and chicory) also 328 display a relatively narrow T_b range of 3.5 °C (Moot et al., 2000). As the local 329 environment is known to influence the expression of this trait, such variation in CWRs 330 331 of *Brassica* can be explained by the selection of taxa from seven countries with widely differing environments, having annual mean temperatures varying from 5 to 26 °C, 332 across a latitudinal span of c. 20°. In more than 50 species of Cactaceae, sampled across 333 an environmental envelope that covers 70° of latitude, seed T_b varies by c. 20 °C (Seal 334 et al., 2017). 335 The range of Ψ_b for the 50th percentile of germination was also wide for species in the 336

- Brassica genus (i.e., -1.54 MPa) from -0.40 to -1.94 MPa. This range is similar to that
- for other crop seeds from different families [such as, *Daucus carota* (Apiaceae),
- 339 Helianthus annuus (Asteraceae), Hordeum vulgare and Zea mays (Poaceae)], that have

a range extending to -2.1 MPa (Dürr et al., 2015). Interestingly, *B. rapa* has been estimated to have a Ψ_b as low as -2.2 MPa (Tribouillois et al., 2016). However, this study used data for only the 20th and 30th percentiles of germination as viability was poor. We also found an average Ψ_b for the same percentiles of germination to be -2.2 and -1.99 MPa for wild *B. rapa* from Switzerland and France, respectively. However, the 50th percentile of the population is a better descriptor of germination performance if seed viability is above 50 % (Soltani et al., 2016).

347 The diversity of germination performance in terms of the thermal and hydro traits 348 observed in the Brassica genus is due to both, their genetic variability (Arias et al., 349 2014; Arias and Pires, 2012) and the influence of the environment (phenotypic 350 plasticity) (Mousseau and Fox, 1998; Schmid and Dolt, 1994; Van Kleunen et al., 351 2007). In this study, we cannot definitively distinguish between genetic or phenotypic 352 variability in the CWRs. However, by studying two *Brassica* seed lots, each of two CWRs from different environments, along with different species from across a broad 353 environmental range, we were able to assess the influence on seed functional traits of 354 355 the collection site abiotic features. Moreover, studying two B. oleracea research crop 356 genotypes selected for differences in vigour (RG_L and RG_H), and grown under 357 identical glasshouse conditions, enabled us to consider the impact of high vigour alleles 358 in these research lines on germination traits.

359 4.2. Selecting for seed performance in Brassica

360 Seed quality is an essential trait for crop production and food security (Finch-Savage

and Bassel, 2015). As a consequence, the seed industry strives to produce seed lots with

- 362 enhanced performance, particularly vigour which is often assessed under non-optimal
- 363 conditions. Vigour is a property of the seed that determines performance in a wide range

364 of environments (ISTA, 2017). Amongst 10 Brassica seed lots assessed over many 365 temperature and water potential conditions we show strong correlations between T_b and $\theta_{\rm T}$ and $\Psi_{\rm b}$ and $\theta_{\rm H}$ (Figure 3) in addition to the correlations with the precipitation of the 366 367 climate of seed collection site of the CWRs (Figure 4). A relationship between Ψ_b and $\theta_{\rm H}$ is anticipated, as the hydro time model shows that germination rate is inversely 368 proportional to the difference between the actual water potential (Ψ) and the Ψ_b 369 (Bradford, 1995). Thus, seeds with lower Ψ_b (more negative water potentials) will 370 371 require more hydro time to germinate, i.e., have longer $\theta_{\rm H}$ (Bradford, 1995). On the other hand, the correlation between T_b and θ_T has been reported previously for a range 372 373 of different species (Dürr et al., 2015; Gardarin et al., 2011; Seal et al., 2017; Trudgill et 374 al., 2005; Trudgill et al., 2000). This might reflect ecological adaptation such that the seeds with a high threshold then proceed to germinate faster, i.e., shorter thermal times 375 376 (Gardarin et al., 2011; Trudgill et al., 2005). As there is intra- and inter-specific 377 variation in the thresholds for seed germination progress under a wide range of (thermal 378 and water potential) environments (Dürr et al., 2015), it is critical that these parameters 379 are determined for each seed lot so that vigour can be more accurately described.

In general, the CWRs had a lower Ψ_b , hence a wider window for water potential

tolerance than the crops. Whereas the seed selection of the crop *B. oleracea* for growth

under optimal, irrigated monoculture agricultural conditions may have led to a higher

383 Ψ_b that increases the risk of crop failure in a future climates of variable water

availability. The low Ψ_b are a greater ability to cope with variable water conditions

indicated in the CWRs may provide a means to improve crop genotypes in the future.

386 The CWRs tended to have similar $\theta_{\rm T}$ compared to the crops. Therefore, the selection and 387 breeding of *B. olereacea* has not resulted in particularly faster germination in terms of

388 thermal time, based on the seed lots characterised here. Even though crop genotype 389 RG_H was the product of the introgression of two high vigour alleles, the impact of this on thermal time was not beneficial compared with CWRs, but it was beneficial 390 391 compared with the low vigour genotype. RG_L had the longest θ_T compared with the 392 other crops and CWRs, i.e. germination is slower, which would increase the risk of 393 inclement drought or seedbed deterioration impacting on the more slowly emerging 394 seedlings. Such subtle differences in thermal and hydro-time characteristics amongst a 395 range of Brassica seed lots (Figure 3, Dürr et al., 2015; Tribouillois et al., 2016) tends to suggest a continuum of responses within the genus. 396

397 4.3. *Seed mass*

Crops were compared to CWRs to observe the effect of breeding and domestication on 398 399 the Brassica genus, one of the main effects of which can be greater seed mass (Doganlar et al., 2000). Research crop genotype RG_H had the heaviest seeds compared to the 400 401 other B. oleracea seed lots and the CWRs (Figure 2). Moreover, less population 402 variation in seed mass (CV) would be expected for crops and this was generally the 403 case. A trend was found where heavier seeds tended to germinate slower than lighter 404 seeds on a thermal basis (longer θ_T). This finding was contrary to that of Huang et al. (2016) who found that larger seeds germinated faster in wild desert species. Smaller 405 406 seeds germinating faster than larger seeds has been found in other species. For example, 407 Grime et al., 1981, who studied 400 species and reported a decrease of germination rate with increased seed weight. Norden et al. (2009) found a similar correlation between the 408 409 mean seed mass and the mean time to germination (MTG) in more than 1000 tropical forest trees. Kikuzawa and Koyama (1999) determined that small seeds had the capacity 410 411 to imbibe water faster than larger seeds, thus the germination process could start earlier.

There was also a tendency for heavier seeds to have lower T_b. Thus, small seeds should require less time to germinate because they need to accumulate less heat units above a higher T_b. Counterintuitively, it seems that breeding to increase seed mass in *Brassica* crops is likely to have unintended consequences on germination rate based on thermal time characteristics. However, further investigation using a higher number of *Brassica* accessions and/or species is needed to confirm these trends.

418 4.4. *Ecological correlates of seed performance*

In addition to temperature, precipitation controls the timing of germination in the wild. 419 420 One general assumption is that germination occurs only if the monthly precipitation is 421 >15 mm (Freas and Kemp, 1983; Gutterman, 1993; Gutterman, 2000a). On this basis, 422 we predicted in which month the seeds will germinate. However, that assumption was 423 not true for *B. rapa* subsp. sylvestris from Algeria, where the maximum monthly 424 precipitation was not above 2 mm (Figure S1). The behaviour of *B. rapa* subsp. 425 sylvestris from Algeria is similar to that of annual plants in an extreme desert climate (Gutterman, 1993) and also coincides with one of the fastest seed lots to germinate. 426 427 When water is regularly available, temperature becomes the major influence of 428 germination timing.

In *Brassica* CWRs, θ_T was positively correlated with precipitation, which is in agreement with the suggestion that seeds from drier environments might be adapted to germinate faster to avoid drought periods during seedling establishment (Evans and Etherington, 1990; Fenner and Thompson, 2004; Gardarin et al., 2011; Moles and Westoby, 2004). At the same time, T_b and Ψ_b were negatively correlated with precipitation which means that: 1) the T_b might be higher in drier environments; and 2) in drier environments the seeds might be adapted to germinate rapidly in response to

436 sporadic rainfall events that increase soil water potential. According to these findings, 437 the germination of wild *Brassica* seed lots, especially thermal time parameters (θ_{T} and 438 T_b) seem to be closely adapted to the precipitation of the maternal environment.

There are many reports of correlations between the mean seed mass and the maternal 439 440 environment (mainly precipitation) including species such as B. campestris (Sinniah et 441 al., 1998), 15 herbaceous species from UK (Evans and Etherington, 1990), annual species (e.g. *Glycine* genus) from a Mediterranean climate (Murray et al., 2004) and two 442 443 wild species of barley and oat (Volis, 2012). Pakeman et al. (2008) found the 444 temperature of several vegetation types across Europe was positively correlated with seed mass. In this study, the mean seed mass was only correlated with the precipitation 445 446 of the month of germination (Table S2; P < 0.05). This finding is in agreement with that on wheat, soya and walnut (Brocklehurst et al., 1978; Meckel et al., 1984; Stromberg 447 and Patten, 1990). Since the size of the seed and number of seeds is limited by the 448 449 availability of resources (Baskin and Baskin, 1998; Wulff, 1986), in this case, mother 450 plants from environments with low precipitation are likely to produce fewer and small 451 seeds.

452 **5. Conclusions**

Based on the thermal and hydro-time characteristics (thresholds and rates) *Brassica*CWRs appear better equipped to adapt to changes in the environment than the crop *B*.

455 *oleracea*. CWRs tended to have lower Ψ_b than crops with suggest more tolerance to low

456 water potentials. These adapted traits are somewhat predictable as the interspecific

457 variation in germination functional traits (T_b , Ψ_b , θ_T) correlate strongly to the climate at

458 the seed collection site, particularly to precipitation (the mean monthly and the mean of

459 the month of cumulative thermal time germination). Our findings reinforce the need to

460	characterise each seed lot, with the additional benefit of more precisely defining seed
461	vigour 'under a wide range of environments'.
462	
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471	Supplementary data
472	Supplementary data is associated with this manuscript.
473	
474	References
475	Ackerly, D.D. et al., 2000. The Evolution of Plant Ecophysiological Traits: Recent
476	Advances and Future Directions: New research addresses natural selection,
477	genetic constraints, and the adaptive evolution of plant ecophysiological traits.
478	AIBS Bulletin, 50, 979-995.
479	Alexander, H.M. and Wulff, R.D., 1985. Experimental ecological genetics in <i>Plantago</i> .
480	X. The effects of maternal temperature on seed and seedling characters in <i>P</i> .
481	lanceolata. J. Ecol. 73, 271-282.

482	Arias, T.	, Beilstein,	M.A.,	Tang, I	M., I	McKain,	M.R.	and Pires,	C.J.,	2014.
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- 483 Diversification times among *Brassica* (Brasicaceae) crops suggest hybrid
 484 formation after 20 million years of divergence. Am. J. Bot. 101, 86-91.
- Arias, T. and Pires, C.J., 2012. A fully resolved chloroplast phylogeny of the brassica
 crops and wild relatives (Brassicaceae: Brassiceae): novel clades and potential
- 487 taxonomics implications. Taxon 61, 980-988.
- Baskin, C.C. and Baskin, J.M., 1998. Seeds: ecology, biogeography and evolution of
 dormancy and germination. Academic Press, San Diego.
- 490 Bradford, K.J., 1995. Water Relations in Seed Germination. In: J.K.a.G. Galili (Editor),

491 Seed development and germination, New York, pp. 351-396.

492 Brocklehurst, P., Moss, J. and Williams, W., 1978. Effects of irradiance and water

493 supply on grain development in wheat. Ann. Appl. Bot. 90, 265-276.

Céspedes, B., Torres, I., Urbieta, I.R. and Moreno, J.M., 2012. Effects of changes in the
timing and duration of the wet season on the germination of the soil seed bank of

496 a seeder-dominated Mediterranean shrubland. Plant Ecolog. 213, 919-931.

- Clauss, M.J. and Venable, D.L., 2000. Seed germination in desert annuals: An empirical
 test of adaptive bet hedging. Am. Nat. 155, 168-186.
- Cochrane, A., Hoyle, G.L., Yates, C.J., Wood, J. and Nicotra, A.B., 2014. Predicting the
 impact of increasing temperatures on seed germination among populations of
 Western Australian Banksia (Proteaceae). Seed Sci. Res. 24, 195-205.
- Cochrane, J.A., Hoyle, G.L., Yates, C.J., Wood, J. and Nicotra, A.B., 2015. Climate
 warming delays and decreases seedling emergence in a Mediterranean
 ecosystem. Oikos 124, 150-160.
- 504 CC05ystem: Orkos 124, 150 100.
- Covell, S., Ellis, R.H., Roberts, E.H. and Summerfield, R.J., 1986. The influence of
 temperatures on seed germination rate in grain legumes. A comparion of

507	chickpea, lentil, soybean and cowpea at constant temperatures. Exp. Bot. 37,
508	705-715.

- Daws, M.I., Crabtree, L.M., Dalling, J.W., Mullins, C.E. and Burslem, D.F.R.P., 2008.
 Germination responses to water potential in netropical pioneers suggest largeseeded species take more risks. Ann. Bot. 102, 945-951.
 Daws, M.I. et al., 2004. Developmental heat sum influences recalcitrant seed traits in *Aesculus hippocastanum* across Europe. New. Phytol. 162, 157-166.
 Dempewolf, H. et al., 2014. Adapting agriculture to climate change: a global initiative
- to collect, conserve and use crop wild relatives. Agroecol. Sust. Food 38, 369-377.
- 517 Díaz, S. et al., 2016. The global spectrum of plant form and function. Nature 529, 167518 171.
- Doganlar, S., Frary, A. and Tanksley, S., 2000. The genetic basis of seed-weight
 variation: tomato as a model system. TAG 100, 1267-1273.
- Donohue, K. et al., 2005. Environmental and genetic influences on the germination of
 Arabidopsis thaliana in the field. Evolution 59, 740-757.
- Dornbos, D.L. and Mullen, R.E., 1991. Influence of stress during soybean seed fill on
 seed weight, germination and seedling growth rate. Can. J. Plant. Sci. 71, 373383.

526 Dürr, C., Dickie, J.B., Yang, X.Y. and Pritchard, H.W., 2015. Ranges of critical

- 527 temperature and water potential values for the germination of species
- worldwide: contribution to a seed trait database. Agr. Forest Meteorol. 200, 222232.
- Ellis , R.H., Covell, S., Roberts, E.H. and Summerfield, R.J., 1986. The influence of
 temperature on seed germination rate in grain legumes. II. Intraspecific variation

532	in chickpea (Cicer arietinum L.) at constant temperatures. J. Exp. Bot. 37, 1503-
533	1515.

- Evans, C.E. and Etherington, J.R., 1990. The effect of soil-water potential on seed-534 535 germination of some British plants. New Phytol. 115, 539-548. FAO/IPGRI, 1994. Genebank Standards, Food and Agriculture Organization of the 536 537 United Nations, Rome. International Plant Genetic Resources Institute, Rome. 538 Fay, P.A. and Schultz, M.J., 2009. Germination, survival and growth of grass and forb 539 seedlings: Effects of soil moisture variability. Acta Oecol. 35, 679-684. Fenner, M. and Thompson, K., 2004. The ecology of seeds, Cambridge University 540 541 Press. Fernández-Pascual, E., Jiménez-Alfaro, B., Hájek, M., Díaz, T. E. and Pritchard, H. W., 542 543 2015. Soil thermal buffer and regeneration niche may favour calcareous fen 544 resilience to climate change. Folia Geobot. 50, 293-301. 545 Finch-Savage, W.E. and Bassel, G.W., 2015. Seed vigour and crop establishment: 546 extending performance beyond adaptation. J. Exp. Bot. 67, 567-591. 547 Franks, S.J., Sim, S. and Weis, A.E., 2007. Rapid evolution of flowering time by an annual plant in response to climate fluctuation. PNAS 104, 1278-1282. 548 549 Freas, K.E. and Kemp, P.R., 1983. Some relationships between environmental 550 reliability and seed dormancy in desert annual plants. J. Ecol. 71, 211-217. García-Huidobro, J., Monteith, J.L. and Squire, G.R., 1982. Time temperature and 551 552 germination of pearl millet (Pennisetum typhoides S&H). I. Constant temperature. J. Exp. Bot. 33, 288-296. 553 Gardarin, A., Dürr, C. and Colbach, N., 2011. Prediction of germination rates of weed 554 555 species: Relationship between germination speed parameters and species traits.
- 556 Ecol. Modell. 222, 626-636.

557	Gareca, E.E., Vandelook, F., Fernandez, M., Hermy, M. and Honnay, O., 2012. Seed
558	germination, hydrothermal time models and the effects of global warming on a
559	threatened high Andean tree species. Seed Sci. Res. 22, 287-298.
560	Gepts, P., 2010. Crop domestication as a long-term selection experiment. Plant Breed.
561	Rev. 24, 1-44.
562	Grime, J., Mason, G., Curtis, A., Rodman, J. and Band, S., 1981. A comparative study
563	of germination characteristics in a local flora. J. Ecol. 69, 1017-1059.
564	Grubb, P.J., 1977. The maintenance of species- richness in plant communities: the
565	importance of the regeneration niche. Biol. Rev. 52, 107-145.
566	Gummerson, R.J., 1986. The effect of constant temperatures and osmotic potentials on
567	the germination of sugar beet. Exp. Bot. 37, 729-741.
568	Gutterman, Y., 1993. Seed germination in desert plants. Springer-Verlag, Berlin,
569	Germany.
570	Gutterman, Y., 2000a. Environmental factors and survival strategies of annual plant
571	species in the Negev Desert, Israel. Plant Species Biol. 15, 113-125.
572	Gutterman, Y., 2000b. Maternal effects on seeds during development. In: M. Fenner
573	(Editor), Seeds: the ecology of regeneration in plant communities. CAB
574	International, pp. 27-59.
575	Hardegree, S.P. and Emmerich, W.E., 1990. Effect of polyethylene glycol exclusion on
576	the water potential of solution-saturated filter paper. Plant Physiol. 92, 462-466.
577	Harel, D., Holzapfel, C. and Sternberg, M., 2011. Seed mass and dormancy of annual
578	plant populations and communities decreases with aridity and rainfall
579	predictability. Basic Appl. Ecol. 12, 674-684.

580	Hijmans, R.J., Cameron, S.E., Parra, J.L., Jones, P.G. and Jarvis, A., 2005. Very high
581	resolution interpolated climate surfaces for global land areas. Int. J. Clim. 25,
582	1965-1978.
583	Huang, Z., Liu, S., Bradford, K. J., Huxman, T. E., & Venable, D. L. (2016). The
584	contribution of germination functional traits to population dynamics of a desert
585	plant community. Ecol. 97, 250-261.
586	IPCC, 2013. Intergovernmental Panel on Climate Change. Climate Change, Cambridge
587	University.
588	ISTA, 2003. Working sheets on tetrazolium testing, I and II, Bassersdorf, Switzerland.
589	ISTA, 2017. International Rules for Seed Testing. International Seed Testing
590	Association, Bassersdorf, Switzerland.
591	Jordan, G.L. and Haferkamp, M.R., 1989. Temperature responses and calculated heat
592	units for germination of several range grasses and shrubs. J. Range Manage. 42,
593	41-45.
594	Jump, A.S. et al., 2008. Simulated climate change provokes rapid genetic change in the
595	Mediterranean shrub Fumana thymifolia. Global Change Biol. 14, 637-643.
596	Keddy, P.A., 1992. Assembly and response rules: two goals for predictive community
597	ecology. J. Veg. Sci. 3, 157-164.
598	Kikuzawa, K. and Koyama, H., 1999. Scaling of soil water absorption by seeds: an
599	experiment using seed analogues. Seed Sci. Res. 9, 171-178.
600	Kimball, S., Angert, A.L., Huxman, T.E. and Venable, D.L., 2010. Contemporary
601	climate change in the Sonoran Desert favors cold-adapted species. Global
602	Change Biol. 16, 1555-1565.
603	Levine, J.M., McEachern, A.K. and Cowan, C., 2008. Rainfall effects on rare annual
604	plants. J. Ecol. 96, 795-806.

- Lobell, D.B. and Field, C., 2007. Global scale climate-crop yield relationships and the
 impacts of recent warming. Environ. Res. Lett. 2, 014002
- 607 Lloret, F., Peñuelas, J. and Estiarte, M., 2004. Experimental evidence of reduced
- diversity of seedlings due to climate modification in a Mediterranean-typecommunity. Global Change Biol. 10, 248-258.
- Meckel, L., Egli, D., Phillips, R., Radcliffe, D. and Leggett, J., 1984. Effect of moisture
 stress on seed growth in soybeans. Agron. J. 76, 647-650.
- Menzel, A. et al., 2006. European phenological response to climate change matches the
 warming pattern. Global Change Biol. 12, 1969-1976.
- Meyer, S.E., Monsen, S.B. and McArthur, E.D., 1990. Germination response of
- *Artemisia-tridentata* (Asteraceae) to light and chill patterns of betweenpopulation variation. Bot. Gaz. 151, 176-183.
- Molesl, A.T. and Westoby, M., 2004. Seedling survival and seed size: a synthesis of the
 literature. J. Ecol. 92, 372-383.
- Moot, D.J., Scott, W.R., Roy, A.M. and Nicholls, A.C., 2000. Base temperature and
- thermal time requirements for germination and emergence of temperate pasturespecies. N. Z. J. Agric. Res. 43, 15-25.
- Morris, K., Barker, G.C., Walley, P.G., Lynn, J.R. and Finch- Savage, W.E., 2016.
- Trait to gene analysis reveals that allelic variation in three genes determines seedvigour. New Phytol. 212, 964-976.
- Mousseau, T. and Fox, C.W., 1998. The adaptative significance of maternal effects.
 Trends Ecol. Evol. 13, 403-407.
- Murray, B.R., Brown, A.H.D., Dickman, C.R. and Crowther, M.S., 2004. Geographical
 gradients in seed mass in relation to climate. J. Biogeogr. 31, 379-388.

629	Nicotra, A.B. et al., 2010. Plant phenotypic plasticity in a changing climate. Trends
630	Plant Sci. 15, 684-692.
631	Norden, N. et al., 2009. The relationship between seed mass and mean time to
632	germination for 1037 tree species across five tropical forests. Funct. Ecol. 23,
633	203-210.
634	Orru, M., Mattana, E., Pritchard, H.W. and Bacchetta, G., 2012. Thermal threshold as
635	predictors of seed dormancy release and germination timing: altitude-related
636	risks from climate warming for the wild grapevine Vitis vinifera subsp.
637	sylvestris. Ann. Bot. 110, 1651-1660.
638	Pakeman, R.J. et al., 2008. Impact of abundance weighting on the response of seed traits
639	to climate and land use. J. Ecol. 96, 355-366.
640	Peñuelas, J. et al., 2004. Complex spatiotemporal phenological shifts as a response to
641	rainfall changes. New Phytol. 161, 837-846.
642	Pollock, B.M. & Roos, E.E. (1972) Seed and seedling vigor. Seed Biology, I.
643	Importance, development and germination, pp. 314-387.
644	Porter, J.R., 2005. Rising temperatures are likely to reduce crop yields. Nature 436,
645	174-174.
646	Porter, J.R. and Semenov, M.A., 2005. Crop responses to climatic variation.
647	Philosophical Transactions of the Royal Society B: Biological Sciences, 360,
648	2021-2035.
649	Poschlod, P. et al., 2013. Seed ecology and assembly rules in plant communities.
650	Vegetation Ecology, Second Edition: 164-202.
651	Preece, C. et al., 2017. How did the domestication of Fertile Crescent grain crops
652	increase their yields? Funct. Ecol. 31, 387-397.

- Roach, D.A. and Wulff, R.D., 1987. Maternal effects in plants. Annu. Rev. Ecol. Evol.
 Syst. 18, 209-235.
- Schmid, B. and Dolt, C., 1994. Effects of maternal and paternal environment and
 genotype on offspring phenotype in *Solidago altissima* L. Evolution 48, 15251549.
- Seal, C.E. et al., 2017. Thermal buffering capacity of the germination phenotype across
 the environmental envelope of the Cactaceae. Global Change Biol. 23, 53095317.
- Sinniah, U.R., Ellis, R.H. and John, P., 1998. Irrigation and seed quality development in
 rapid-cycling Brassica: Seed germination and longevity. Ann. Bot. 82, 309-314.
- 663 Smith, S.E., Riley, E., Tiss, J.L. and Fendenheim, D.M., 2000. Geographical variation
- in predictive seedling emergence in a perennial desert grass. J. Ecol. 88, 139-149.
- Soltani, E., Ghaderi-Far, F., Baskin, C. C., & Baskin, J. M. (2016). Problems with using
 mean germination time to calculate rate of seed germination. Austral. J. Bot. 63,
 668 631-635.
- Stromberg, J.C. and Patten, D.T., 1990. Variation in seed size of a southwestern riparian
 tree, Arizona walnut (*Juglans major*). Am. Midland Nat. 124, 269-277.
- Tribouillois, H., Dürr, C., Demilly, D., Wagner, M.-H. and Justes, E., 2016.
- Determination of germination response to temperature and water potential for a
 wide range of cover crop species and related functional groups. PloS one 11,
 e0161185.
- Trudgill, D.L., Honek, A., Li, D. and Van Straalen, N.M., 2005. Thermal time- concepts
 and utility. Ann. Appl. Biol. 146, 1-14.

677	Trudgill, D.L., Squire, G.R. and Thompson, K., 2000. A thermal time basis for
678	comparing the germination requirements of some British herbaceous plants.
679	New Phytol. 145, 107-114.
680	Tsunoda, S., Hinata, K. and Gómez-Campo, C., 1980. Brassica crops and wild allies.
681	Biology and breeding, Japan Scientific Societies Press, Tokyo.
682	Van Kleunen, M., Lenssen, P.M., Fischer, M. and de Kroon, H., 2007. Selection on
683	phenotypic plasticity of morphological traits in response to flooding and
684	competition in the clonal shore plant Ranunculus reptans. J. Evol. Biol. 20,
685	2126-2137.
686	Volis, S., 2012. Demographic consequences of delayed germination in two annual
687	grasses from two locations of contrasting aridity. PPEES. 14, 335-340.
688	Wulff, R.D., 1986. Seed size variation in Desmodium paniculatum: I. Factors affecting

689 seed size. J. Ecol. 74, 87-97.