Divergent Mechanisms of Acoustic Mate Recognition Between Closely-Related Field Cricket Species (*Teleogryllus spp.*)

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1 ABSTACT

2 Effective recognition of conspecific mating signals reduces the risk of maladaptive hybridisation. Dissecting the signal recognition algorithms that underlie preferences is a 3 4 useful approach for testing whether closely related taxa evaluate the same or different signal 5 features to achieve mate recognition. Such data provide information about potential 6 constraints and targets of selection during evolutionary divergence. Using a series of mate choice trials, we tested whether closely-related, but genetically and phenotypically divergent, 7 8 field cricket species (Teleogryllus oceanicus and Teleogryllus commodus) use shared or 9 distinct recognition algorithms when evaluating acoustic male calling songs. These species 10 overlap in sympatry, show premating isolation based on female discrimination of male calling songs, yet are capable of producing hybrid offspring. Unexpectedly, female selectivity 11 12 for features of male song differed between the two species. We found that the two species use a combination of shared and unique signal filtering mechanisms, and we characterised how 13 14 information about male carrier frequency, pulse rate and temporal patterning is integrated to achieve song recognition in each species. These results illustrate how comparatively few, 15 16 simple modifications in key components of signal recognition algorithms can lead to striking 17 interspecific discrimination among closely related taxa, despite apparent signal complexity. The finding that some steps during signal recognition and filtering are shared between the 18 19 species, while others differ, can help to identify behavioural traits targeted by selection 20 during evolutionary divergence.

Keywords: acoustic communication, divergence, female preference, mate recognition, reproductive isolation, sexual selection, speciation, *Teleogryllus*

22 INTRODUCTION

23

24 The decision-making processes that animals use to evaluate and select among potential mates 25 can have an important influence on the evolutionary outcome of sexual selection (Bateson, 26 1983). For example, mismatches between populations in sexually-selected traits and 27 preferences can generate reproductive isolation and promote speciation (West-Eberhard, 1983; Greenfield, 2002; Coyne & Orr, 2004; Mendelson & Shaw, 2005; Safran et al., 2013; 28 29 Shaw & Mendelson, 2013). Understanding how individuals recognise different male signals 30 is therefore a fundamental goal of sexual selection research (Bateson, 1983; Andersson, 1994; 31 Ritchie, 2007; Chenoweth & McGuigan, 2010), and theoretical models of sexual selection in 32 systems with female choice have predicted a key role for female responsiveness, preference 33 and discrimination of such signals (Lande, 1981; Bateson, 1983; Mead & Arnold, 2004; 34 Andersson & Simmons, 2006). Understanding the mechanistic bases of mating preferences 35 and decision-making behaviours can help to answer questions about their function and 36 evolution. For example, work on the genetic basis of mate choice in drosophilid fruit flies has 37 illustrated an evolutionary link between ecological and mating traits (Chung et al., 2014), 38 studies of the zebra finch Taenopygia guttata have clarified neural architecture that might 39 control species difference in song preferences (MacDougall-Shackleton, Hulse, & Ball, 40 1998), and characterising perceptual tuning in the acoustically-signalling anuran 41 Physalaemus pustulosus has shown how pre-existing sensory biases can facilitate evolution 42 via sexual selection (Ryan et al., 1990). 43 One way to study the neurophysiological mechanisms underlying mate recognition is 44 to treat the decision-making process as a computational algorithm, or series of operations 45 used to evaluate incoming signals and transform that evaluation into a behavioural action 46 such as a mating response (Ronacher, Hennig, & Clemens, 2015). Filters are integral

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47 components of such signal processing algorithms, and in animals, signal filters represent 48 traits of the organism that exclude irrelevant information contained in incoming signals to 49 focus reception upon important signal features. In acoustically-signalling organisms, for 50 example, species can differ in the physical or mechanical properties of structures used to 51 receive sounds, such as tympana, providing peripheral filtering of signals, and the central 52 nervous system can also filter incoming signals depending on the configuration of neural 53 pathways (Greenfield, 2002).

54 By designing tests that manipulate male signal components and assess female 55 responses, it is possible to gain insight into which signal features females attend, which are 56 filtered out, how different signal features might be traded off against one another during 57 assessment, and which ones are possible targets of sexual selection (Kostarakos, Hartbauer, 58 & Römer, 2008; Hedwig, 2006; Hennig, 2009; Henni, Heller, & Clemens, 2014). Much work 59 examining signal recognition algorithms underlying female choice has focused on evaluations 60 that females make among potential mating partners of the same species, and this has taken the 61 form of measuring female preference functions (Wagner, 1998). However, it is less clear 62 whether closely related taxa that risk coming into contact and producing low fitness hybrids 63 use the same, different, or more complex algorithms when faced with the challenges of mate recognition. For instance, closely related species in the treefrog genus Hyla have been found 64 65 to distinguish conspecific from heterospecific calls using different sets of temporal call 66 features, reflecting divergence in signal recognition algorithms (Schul & Bush 2002). In addition to clarifying similarities and differences in the neural mechanisms underlying mate 67 68 recognition in related species, such data can inform likely targets of sexual selection and 69 constraints during the evolution of reproductive isolation and reinforcement (Coyne & Orr, 2004). 70

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71 We tested whether the algorithms and filters underlying mate recognition differ 72 between two closely related field cricket species, *Teleogryllus oceanicus* and *T. commodus*, 73 which are a classic system in the study of acoustic signalling and reproductive isolation (e.g. 74 Hoy & Paul, 1973; Hoy, 1974). These crickets are firmly established as separate species, and 75 both attract mates using long-range acoustic signals that are clearly distinguishable at the 76 phenotypic level (Otte & Alexander, 1983). Both species inhabit coastal regions of Australia, 77 with T. oceanicus in the north and T. commodus in the south, and their distributions overlap 78 for several hundred kilometres along the central eastern seaboard (Fig. 1a) (Otte & 79 Alexander, 1983). The species readily hybridise in the lab, though hybrid females are almost 80 always infertile, providing an unusual, reciprocal exception to Haldane's Rule (Hogan & 81 Fontana, 1973; Moran, Ritchie, & Bailey, in press). Despite their known ability to interbreed, 82 hybridisation is thought to be rare or absent in the wild (Hill, Loftus-Hills, & Gartside, 1972, 83 though see Otte & Alexander 1983).

84 Long-range male advertisement songs of Australian Teleogryllus are unusual owing to 85 a patterning complexity not normally observed in grylline crickets: the songs consist of two stereotyped elements, or phonemes: a higher-amplitude pulse train we refer as the "chirp" 86 87 followed by a series of shorter, lower-amplitude pulses we refer to as "trills" (Figs. 1b, c). Both species also produce a similarly-structured, short-range courtship song which functions 88 89 to release female mounting behaviour (Balakrishnan & Pollack, 1996), but here we focus on 90 the long-range attraction signal given its known contribution to premating isolation (Hill, 91 Loftus-Hills, & Gartside, 1972, Bailey & MacLeod 2014). Both species exhibit this two-part 92 calling song pattern, although a distinguishing feature between them is that in *T. oceanicus*, 93 the lower-amplitude trills following the initial chirp are comprised of paired pulses (with 94 occasional triplets or, less frequently, higher pulse number trills), whereas the lower amplitude trills of T. commodus are comprised of a smaller number of longer-duration trill-95

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96 like elements composed of a greater number of pulses (Fig. 1b, c). Average carrier 97 frequencies are also higher for T. oceanicus (ca. 5 kHz) than for T. commodus (ca. 4 kHz) (Bailey & Macleod, 2014). This system therefore provided an opportunity to test whether 98 99 recognition algorithms underlying female mate choice for conspecific vs. heterospecific songs rely on differential filtering of the same acoustical traits of male calling song, or 100 101 whether females have diverged in the traits that their signal filters target. Put another way, are 102 females of both species selective for the same or different song features when exerting 103 preference?

104 Previous work illustrated the importance of pattern recognition for conspecific female phonotaxis in *T. commodus*, and suggested that a different balance of peripheral versus 105 106 central nervous processing contributes to conspecific song recognition in each species 107 (Hennig & Weber, 1997). After validating this result, we developed tests to examine the 108 overall selectivity for con- and heterospecific song patterns and test the contributions of 109 carrier frequency, pulse rate during chirps and trills, and trill pattern composition to song 110 recognition and selectivity. We expected that both species use a combination of spectral and temporal filters (Hennig & Weber, 1997), but given that frequency differences may not 111 112 definitively identify T. commodus and T. oceanicus, temporal patterns of song envelopes 113 were expected to play an important role. Two main findings provide insight into divergence 114 of mate recognition algorithms. First, closely related taxa do not necessarily employ the same 115 filter types to differentiate individuals of the other taxon, and second, the strong divergence in mate recognition that this causes can reflect relatively few, minor shifts in the way signals are 116 117 processed by the nervous system.

118

119 **METHODS**

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123	We used laboratory-reared adults from two allopatric populations located near
124	Townsville, QLD (T. oceanicus) and Moss Vale, NSW (T. commodus). Otte & Alexander
125	(1983) reported a single recording of <i>T. commodus</i> calling song during a field survey near
126	Townsville. However, that specimen's reported carrier frequency was consistent with T.
127	oceanicus (4.6 kHz compared to an average of 3.65 kHz for T. commodus reported by Otte &
128	Alexander (1983)), and we observed no <i>T. commodus</i> in the field or among the laboratory-
129	reared offspring of field-caught individuals from Townsville (Moran & Bailey, 2013). We
130	therefore considered the populations used in this study to be allopatric. Prior to testing, the
131	populations had been reared separately in common-garden conditions in the lab for at least
132	one generation to mitigate maternal effects that could reflect field conditions. Stock crickets
133	were kept in 16L translucent plastic containers at ca. 25 °C on a photo-reversed 12h:12h
134	light:dark cycle. They were fed Supa Rabbit Excel Junior and Dwarf Rabbit nuggets ad
135	libitum, and provisioned with cardboard egg cartons and moistened cotton pads. Sexually
136	mature adult females (7 days or older) were tested.
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138	Female Phonotaxis Tests
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140	Female phonotaxis responses were tested used a trackball system and a series of
141	artificially-constructed song playbacks. Setup of the trackball and its operation followed
142	Dahmen (1980) and Hedwig & Poulet (2004). The general protocol we used for phonotaxis
143	assays has been described in detail elsewhere (e.g. Blankers, Hennig & Gray, 2015; Hennig,
144	Blankers & Gray, 2016), so here we summarise the approach and highlight key differences in

our experiments. 145

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146 Females were suspended in a walking position over a hollow Styrofoam ball (100 mm 147 diameter, weighing 1.2 to 1.8 g) positioned within a 50 x 50 x 50 cm box lined with acoustic 148 foam. The ball floated on an airstream and its movements were recorded from the bottom by 149 an optical sensor (Agilent ADNS-2051), or by two laterally-focused sensors (ADNS-5050, Avago Technologies) positioned perpendicular to one another. Each channel had a sampling 150 151 rate of 10 kHz and signal was processed through an A/D-board (PCI-6221, National Instruments, Texas) with Labview v.7.1 or v.9 software. This enabled us to record 152 153 longitudinal and lateral movements of the trackball when crickets responded during 154 playbacks.

155 Playbacks with the required carrier frequencies and pulse characteristics (1 ms rise 156 and fall) were constructed using LabView 7.0 and transmitted as described in Hennig, 157 Blankers and Gray (2016). Briefly, songs were played back at 80 dB through two Piezo Horntweeter PH8 loudspeakers 25 cm away and 45° to the left and right of the trackball's 158 159 upper surface. Speakers were calibrated by playing a 1s tone matching the required carrier 160 frequency and assessing with a Bruel and Kjaer sound level meter and a condenser microphone on a fast reading relative to 2×10^{-5} Pa (Bruel and Kjaer 2231 and 4133, 161 respectively). Test sessions were run at 25 ± 1 °C, and for each, we performed one 45 s silent 162 control at the beginning, one 45 s continuous tone control at the end, a positive control at the 163 164 beginning and a positive control at the end (Fig. 2a, b), plus the 8 focal test signals in 165 randomized order. Parameter values for test signals are provided in the figure captions for each species. Signal presentations were separated by 10 s silent intervals. Silent and tone 166 167 controls allowed us to monitor and adjust for female motivation and selectivity. Positive 168 controls represented the most attractive combination of song elements for each species (Fig. 2d: Positive controls for T. oceanicus: 5.0 kHz and TP1: chirp duration: 275 ms, pulse rate 169 during chirp: 16 pulses per second, pps, pulse duty cycle 0.6; trill duration 960 ms composed 170

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171	of double pulses at pulse periods of 40 ms and 80 ms. T. commodus 4.0 kHz and TP3: chirp
172	duration: 320 ms, pulse rate during chirp 18 pulses per second, pps, pulse duty cycle 0.65;
173	trill duration 700 ms at a pulse rate of 35 pps followed by a pause of 200 ms). Here we
174	consider female selectivity as the degree to which females discriminate trait values to which
175	they respond most strongly (cf. 'preference window' in Butlin (1993), 'discrimination' in
176	Bailey (2008), and 'tolerance' in Fowler-Finn & Rodríguez (2011)).
177	

- 1//
- 178 Phonotaxis Response Scores
- 179

We calculated phonotaxis scores (*PS*) of 9-32 females for each species, for each 45 second test pattern, using females' longitudinal forward (*X*) and lateral sideward deviations (*Y*) towards the playback. Both *X* and *Y* were normalised to the attractive controls, and female response relative to the two speakers was averaged to obtain a robust measure of response strength. The *PS* was calculated using the formula:

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$$PS = \left[\frac{\left(\frac{X_T}{\bar{X}_{CP1,2}}\right) + \left(\frac{|Y_T|}{\bar{Y}_{CP1,2}}\right)}{2}\right] \times [sgn(Y_T)]$$

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where X_T and Y_T represent the forward (*X*) and lateral (*Y*) walking components during the test, and $\overline{X}_{CP1,2}$ and $\overline{Y}_{CP1,2}$ represent forward (*X*) and lateral (*Y*) walking components averaged over positive controls at the beginning (*CP1*) and end (*CP2*) of a test session. Multiplication by the sign of the lateral walking component, $sgn(Y_T)$ (equivalent to turns away from the active speaker), ensured that the overall *PS* could obtain negative values. Negative scores and scores larger than 1 could thus be obtained, although *PS* typically ranged between 0 and 1. For example, *PS* < 0 could result if females turned away from the active

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194 speaker, and PS > 1 could result if during a test, females exhibited a turning response stronger than that which they exhibited during the control stimulus. In some of the presented 195 196 data, responses of females were high but did not reach scores of 1.0 (e.g. Figs. 3c, 4a and 5a, 197 b). This reduction was most likely due to suboptimal combinations of the large number of 198 parameters that describe the song patterns of these species. If female PS to the initial positive 199 control of a test session fell below 0.5, the session was aborted. Females were also excluded from further analysis if their final positive control PS was less than 50% of their initial 200 201 positive control PS, or if they were highly responsive during silent and tone controls, 202 although the latter occurred infrequently (Fig. 2a, b). 203 Statistical comparisons of the turning responses to test patterns were performed using 204 paired t-tests. Statistical significance was assessed at $\alpha = 0.05$. Unless otherwise specified, 205 means and standard errors of the data are presented, and sample sizes (n) for each test series 206 are given in the figure captions. Degrees of freedom (df) were calculated as df = 2(n-2). R v. 2.15.2 was used in construction of the map in Fig. 1 (R Core Team 2012; Becker & 207 208 Wilks 2013a,b).

209

210 **RESULTS**

211

212 Interspecific Variation in Female Selectivity

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Females of both species were tested for their ability to discriminate con- and

215 heterospecific song types. As illustrated in Fig. 1, song structure is distinct between these

species (see also: Otte & Alexander, 1983; Hennig & Weber, 1997; and Table S1 in Bailey &

217 Macleod, 2014). For this test, song patterns were constructed that exhibited an initial chirp

218 section typical for the respective species, plus a trill part that mimicked the song pattern with 219 respect to pulse rate (TP1 for T. oceanicus, TP3 for T. commodus in Fig. 2d). Additionally, 220 females were tested with patterns representing a fusion of otherwise separated trill pulses to 221 longer blocks of sound (TP2, TP4 in Fig. 2d). The latter two test patterns were expected to be 222 indicative of potential differences in selectivity for the trill part between both species. Each 223 test pattern was presented at the con- and heterospecific carrier frequency (4.0 and 5.0 kHz in 224 Fig. 2c). T. commodus females were highly selective for carrier frequency and temporal 225 patterning elements, whereas T. oceanicus females were less selective for temporal pattern 226 features (Fig. 2c). For instance, T. oceanicus accepted all test patterns, provided they were 227 broadcast at 5.0 kHz. T. oceanicus responses were attenuated at 4.0 kHz. In contrast, females 228 of T. commodus only responded if both the carrier frequency and the temporal pattern 229 corresponded to the conspecific song. This distinction illustrates that T. commodus females 230 only showed strong responses to song models with the lower species-specific 4 kHz carrier 231 frequency when they were presented with an appropriate species-specific pulse pattern, 232 whereas T. oceanicus females responded strongly to species-specific 5 kHz frequency 233 playbacks regardless of the pulse pattern presented. Females of T. commodus were therefore 234 more selective for the temporal pattern than females of *T. oceanicus* (Fig. 2). 235 Components of female selectivity 236 237 238 In a further series of tests, females of both species were exposed to test patterns 239 designed to dissect the contribution to the selectivity observed before of carrier frequency, 240 pulse rates in chirp and trill, and trill composition (Fig. 2c). As predicted, responses to carrier 241 frequency were differently tuned in the two species. T. commodus females showed a peak

response to calling songs at 4.0 kHz, whereas *T. oceanicus* females preferred songs 4.5 kHz

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243 or higher in frequency (Fig. 3a). Female responses for pulse rate during chirps were broadly 244 similar, with only a small difference in the most preferred pulse rate (T. oceanicus: 12 pps, T. 245 commodus 16 -18 pps, Fig. 3b). However, T. commodus showed selectivity for a specific 246 pulse rate of 32 pulses per second during the trill portion of the calling song, whereas T. oceanicus females only responded if the pulse rate during the trill part was the same as during 247 248 the chirp part, that is at 12 pps (Fig. 3c, c.f. T. oceanicus in Fig. 3b). T. commodus thus 249 exhibits different pulse rate selectivity for the two song phonemes, requiring two pulse rate 250 filters, whereas the most preferred pulse rate (12 pps) is the same for each phoneme in T. 251 oceanicus, for which a single pulse rate filter suffices. The addition of a separate filter for 252 pulse rate selectivity suggests higher sensitivity to temporal pattern properties of calling song 253 for T. commodus females than for T. oceanicus. Indeed, the preferred pulse rate of 12 pps by 254 T. oceanicus in Fig. 3c indicated that females did not require a trill part for recognition and 255 that the pulse rate of the chirp part alone sufficed.

The contribution of the trill composition in terms of pulses per trill and trill duration indicated broadly similar responses in both species (Fig. 4). *T. oceanicus* females accepted trills built from two or more pulses, whereas *T. commodus* accepted trills built from three pulses or more (Fig. 4). Longer trills were accepted by both species equally readily, although only females of *T. commodus* appeared to be selective for a particular pulse rate during this part (Fig. 3c).

To examine whether *T. oceanicus* simply ignored features of the trill part or whether they exhibited selectivity to other temporal cues, females were tested with patterns that varied the pulse duty cycle. Such patterns exhibit different amounts of sound energy independent of a particular pulse rate as illustrated in Fig. 5c, as the duty cycle is calculated from the pulse duration divided by the pulse period. *T. oceanicus* females exhibited a strong selectivity for

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267 all patterns with a pulse duty cycle higher than 0.5, which corresponded to patterns with high 268 sound energy as they contained pulses longer than the pauses in between (Fig. 5).

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DISCUSSION 270

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272 The origin and maintenance of mating barriers is a fundamental requirement for 273 speciation to occur in situations where diverging populations could hybridise, or when 274 secondary contact occurs between closely related taxa (Coyne & Orr, 2004). Divergence in 275 signalling and mate recognition traits facilitates the establishment of such barriers. While 276 changes in signalling traits and mate recognition at the phenotypic level have been well-277 characterised in a number of systems, less is known about whether the underlying 278 physiological mechanisms that control such mate recognition are shared or not in such taxa. 279 Because signals are typically multi-component and complex, divergence could occur as a result of changes in the same filtering mechanism in different species, such that different 280 281 values of the same signal trait are preferred, or by establishment of new filters such that divergent taxa are tuned to different signal traits. We found a mixture of both scenarios in T. 282 283 *oceanicus* and *T. commodus*, which we can illustrate by separating the different filter 284 components of the processing algorithm much like a flow diagram (Fig. 6).

285 Our dissection of mate recognition algorithms in *Teleogryllus* showed that females of both species attended to frequency differences and showed sharply tuned filters that almost 286 287 perfectly match the documented differences in carrier frequency of conspecific male calling 288 songs, consistent with prior reports (Hennig & Weber, 1997; Bailey & Macleod, 2014). The 289 majority of known examples of acoustic species recognition in insects, and particularly 290 crickets and other ensiferan insects, focus on temporal patterning of male advertisement 291 songs (e.g. Ritchie, 1991; Mendelson & Shaw, 2005; Meckenhäuser, Hennig, & Nawrot

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292 2013; Kostarakos & Hedwig, 2015), and a longstanding assumption about the evolution of 293 cricket calling songs is that there is unlikely to be significant variation in carrier frequency 294 among closely related taxa, due to the mechanical constraints imposed by physical features of 295 male forewings used in song production (Alexander, 1962). For example, neural recordings 296 of responses to courtship song in a gryllid from the western hemisphere, Gryllus assimilis, 297 indicate the importance of temporal song patterning compared to carrier frequency, with 298 female auditory neurons exhibiting a broad frequency response spectrum ranging from 3.5 299 kHz to 14.5 kHz (Vedenina & Pollack, 2012), and early perceptual models for discrimination 300 of acoustic signals in T. oceanicus suggested that the main frequency-based distinction this 301 species makes is of a categorical nature, between low frequency and ultrasound (Wyttenbach, 302 May & Hoy, 1998).

303 Nevertheless, our results confirm that both T. oceanicus and T. commodus share 304 frequency filters, with the result that females of both species filter incoming male signals as a 305 function of those signals' dominant carrier frequency. Selectivity for frequency indicated that 306 peak female responses were only approximately 1kHz apart. However, this selectivity 307 matches observed differences in frequency differences of males, both from these populations 308 (Moran & Bailey, 2013) and from other populations of the same species (Bailey & Macleod, 309 2014). Such a shift in the frequency filter does not necessarily require evolutionary change in 310 complex neural architecture or physiological processes, and could be underpinned by simple 311 size scaling differences that have arisen during the evolutionary history of these two species. For example, a meta-analysis of 58 species of tettigoniids, an ensiferan group in which males 312 sing using a forewing file and scraper mechanism, uncovered significant overall covariance 313 314 between body size and carrier frequency (Montealegre-Z, 2009). Teleogryllus commodus are larger than *T. oceanicus* on average, and if male forewing structures and tympanal hearing 315 316 organs scaled with body size in a correlated manner during divergence, corresponding

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frequency filters in females of each species could be selectively tuned to the dominant carrierfrequency produced by conspecific males.

319 Both cricket species appear to share another filter, by which the pulse rate of the chirp 320 portion of the song is evaluated. Pulse rate selectivity can arise from only a small network of 321 neurons, in which the property of a rebound oscillation plays a crucial role (Weber & 322 Thorson, 1989; Pollack, 2000; Clemens & Hennig, 2013; Schöneich, Kostarakos, & Hedwig, 323 2015). Notably, the preference functions for this song component were very similar in the 324 two species (Fig. 3b, 6). This similarity is consistent with previous reports suggesting that 325 pulse rate during the chirp is under stabilising selection in both species (Hennig & Weber, 1997). In contrast with the chirp filter, the species differ in selectivity of the trill portion of 326 327 the song. T. oceanicus females appear to be unselective towards the trill pattern (Figs. 2, 3 328 and 4), but they preferred trill patterns with longer pulses and shorter pauses (Fig. 5). Taken 329 together, this is indicative of duty cycle selectivity favouring patterns with higher energy. The 330 particular timing of pulses as given by a pulse rate did not appear relevant, which contrasted 331 distinctly with T. commodus females (Fig. 3c). Thus, female selectivity for pulse rate within 332 the trill portion of calling song highlights a key difference between the species: T. commodus 333 females are more highly selective of trill patterning, focusing on temporal aspects of trill pulses such as pulse rate, whereas T. oceanicus females attend to the pulse duty cycle of the 334 335 trill irrespective of the patterning (Fig. 2, Fig. 6). T. commodus appears to be the rarer species 336 in sympatry (Moran & Bailey, 2013), and it enters a diapause in more southern populations 337 (Otte & Alexander, 1983). Both scenarios might favour enhanced female selectivity in T. 338 *commodus* females: rarity would increase the chances of maladaptive hybridization, and 339 introgression of genes that reduce or eliminate the tendency to enter diapause would be 340 detrimental to T. commodus females.

341

The integration of similar signal recognition algorithms based on frequency filters

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342 with a different mechanism based on discrimination of pulse rate during the trill portion of 343 the song contrasts with recent findings in several gryllids producing either short, chirp-like 344 phonemes (Hennig, Blankers, & Gray, 2016) or long, trill-like phonemes (Blankers, Hennig, 345 & Gray, 2015). The latter species show identical computational algorithms for evaluating 346 acoustic signals based on pulse pattern and chirp/trill features. (Blankers, Hennig, & Gray, 347 2015; Hennig, Blankers, & Gray, 2016). Nevertheless, these species differ in their preference 348 for a particular pulse rate or chirp/trill duty cycle. Some gryllid species show a transition 349 from a pulse rate filter to a pulse duty cycle filter, consistent with what we have observed in 350 Teleogryllus (Fig. 5) (Hennig, Blankers, & Gray, 2016). Our behavioural experiments cannot resolve how the algorithmic flow of information during phonotaxis or particular filter 351 352 component is implemented in terms of physiological or neural activity. Nevertheless, 353 physiological recordings from sensory cells in the tympanic ear (Imaizumi & Pollack, 1999) 354 and brain neurons sensitive to pulse rate (Schöneich, Kostarakos & Hedwig, 2015) support 355 the proposed sequential processing steps and filter properties illustrated in Fig. 6. 356 There are several illustrative differences between song pattern recognition in the 357 gryllids mentioned above versus *Teleogryllus*, which suggest a more general, taxonomically-358 widespread pattern underlying the evolution of signals and signal recognition during 359 diversification. For example, most gryllids produce a series of pulses grouped into chirps or 360 trills, which are separated by variable durations of silence (Blankers, Hennig, & Gray, 2015; 361 Hennig, Blankers, & Gray, 2016). In contrast, the Teleogryllus species we studied produce 362 calling songs with a greater number of phonemes, as in the chirp and trill part (Fig. 1), 363 although *Teleogryllus* species with simpler song patterns are known (Rothbart & Hennig, 364 2012). The tendency toward additional song pattern elements, or phonemes, can be even greater in other ensiferan taxa; certain species of the Tettigoniid genus Amblycorypha 365 366 produce some of the most complex acoustic signals of any insect, with varied arrangements

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of up to four phonemes (Walker & Dew, 1972). A tempting prediction is that the signal
recognition filters required to process complex incoming signals will be correspondingly
complex, and may therefore provide a larger target for selection or drift to modify (Fig. 6)
(Hebets & Papaj, 2005).

Despite the phenotypic differences in song recognition and apparently larger number 371 372 of filters required for mate recognition in *Teleogryllus* (Fig. 6), the filters themselves are in principle similar or even identical to those described in other crickets. This observation 373 374 suggests that the apparently derived situation in *Teleogryllus* builds on existing schemes of 375 pattern recognition. Two important transitions are worth highlighting: first the duplication of a pulse rate filter, and second, the transformation of a pulse rate filter to a duty cycle filter 376 377 (Fig. 6), the latter of which appears complicated at first but can be simply achieved by small 378 changes of the filter template used for song recognition (Hennig, Heller, & Clemens, 2014). 379 These observations also suggest that recognition of a complex song pattern such as the trill 380 portion of T. oceanicus calling song (Fig. 1) does not necessarily evolve because of a more 381 complex filter, but may arise in response to a relatively simple duty cycle filter (Figs. 5, 6). 382 The combined effects of multiple, simple filters thus provide a parsimonious explanation for 383 the multitude of different ways in which species-identifying signals can diverge alongside recognition mechanisms for those signals. In T. oceanicus and T. commodus, divergence in 384 385 signal recognition appears to have arisen from a combination of different filters applied to the 386 same signal features, plus the modification of filters to target distinct signal features. Changes 387 in decision algorithms must ultimately reflect measurable physical changes in the structure or 388 neural connections within the organism, and our results are consistent with the idea that such divergence will follow an evolutionary "path of least resistance": apparent signal recognition 389 390 complexity can arise from few, basic decision algorithms.

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393

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541 **Figure captions**:

543 Figure 1. Cricket ranges and signals. (a) Approximate Australian distribution of T. oceanicus 544 (light grey), T. commodus (dark grey), and region of sympatry (stripes)., based on Otte & Alexander (1983) and Moran & Bailey (2013). Locations of populations used in this study 545 546 are indicated with arrows. Our field and laboratory observations are consistent with these 547 being pure-species, allopatric populations (see main text for details). Male calling song 548 diagrams are based on Bailey & Macleod (2014) and illustrate song features of interest for (b) 549 T. oceanicus and (c) T. commodus. Different authors have historically used different 550 terminology to describe elements of *T. oceanicus* calling song. Those employed in the present 551 study are indicated with larger font, while alternative terms for the same song features are 552 indicated with smaller font in parentheses to ease comparison with prior work. 553 554 Figure 2. Female selectivity for male calling song models that varied in carrier frequency and 555 temporal patterning. Phonotaxis scores are shown for *T. oceanicus* females (black bars, n = 15) and T. commodus females (open bars, n = 13). (a,b) Female response to positive 556 557 (attractive stimuli) and negative controls (unattractive stimuli) during a test session (CP1, 2 positive controls at the beginning and end of a test session, CS: silent control, CT tone 558 559 control). (c,d) Females were presented with test patterns (shown in (d)) similar to a T. 560 oceanicus (TP1,2) or T. commodus (TP3,4) calling song. Each test pattern was presented at 561 4.0 and 5.0 kHz, corresponding to the carrier frequency of the song of both species. Responses in (c) marked with '#' were not significantly different from the positive controls in 562 (a) and (b), and the response marked with '*' was significantly (p<0.05, t-test) different from 563 564 the negative controls in (a) and (b). Means and standard errors are presented in (a)-(c). 565

566 Figure 3. Preferences (means and standard errors) for calling song features exerted by 567 females of each species: (a) carrier frequency (*T. oceanicus* n = 15; *T. commodus* n = 13), (b) pulse rate in the chirp portion of the song, holding trill pulse rate constant (T. oceanicus n 568 569 = 9; T. commodus n = 10), and (c) pulse rate in the trill portion of the song, holding chirp pulse rate constant (*T. oceanicus* n = 12; *T. commodus* n = 41). Pulse rates are given in pulses 570 571 per second. Response levels higher than 0.7 were not significantly different from the positive 572 controls, response levels below 0.3 were not significantly different from the negative controls 573 (c.f. Fig. 2A, B). Test patterns in (a) corresponded to conspecific songs as in Fig. 2D (TP1 for 574 T. oceanicus and TP3 for T. commodus). Test patterns in (b) corresponded to continuous 575 pulse trains with variable pulse rate for T. oceanicus and variable pulse rate during the part 576 with a continuous pulse train during the trill for T. commodus as for TP3 in Fig. 2D. Test 577 patterns in (c) had a constant chirp part as TP1 and TP3 in Fig. 2D and a continuous trill part 578 with variable pulse rate as TP3 in Fig. 2D. Typical trait values for the calling song signal of 579 both species are available from Bentley & Hoy (1972), Hill, Loftus-Hills & Gartside (1972), 580 and Hennig & Weber (1997). (For T. commodus//T. oceanicus, respectively: carrier frequency: 3.5-3.8//4.5-4.9, pulse rate within chirp: 19-20//15-16, pulse rate within trill: 31.5-581

582 583 31.6//24-26).

Figure 4. Preferences for overall trill composition. (a) Phonotaxis scores (means and standard errors) for *T. oceanicus* (n = 11) and *T. commodus* females (n = 23). (b) Diagram of test patterns in which the number of pulses was varied during the trill portion, thereby changing the trill duration. Pulse periods were set to 40 ms, and pulse periods between groups of pulses were set to 80 ms. Phonotaxis scores higher than 0.3 were significantly different from the negative controls (p< 0.05, c.f. Fig. 2a,b).

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591 **Figure 5**. (a) Selectivity for temporal cues during the trill part containing more sound energy 592 by females of T. oceanicus. Numbers refer to test patterns in (c). The open circle at the center 593 refers to phonotaxis score to the positive control pattern. Diameter of circles indicates 594 strength of phonotaxis score which was 1.0 for the positive control. (b) Females exhibit 595 selectivity for the pulse duty cycle in the trill portion of the calling song (means and standard 596 errors are presented). The curves correspond to transects through (a) from upper left to lower 597 right at different pulse periods, as indicated. (c) Selected test patterns, as indicated in (a), with 598 a constant chirp part (TP1 in Fig. 2D) and a varied trill section. Numbers to the right refer to 599 the pulse duty cycle (pdc) of each pattern. Response levels higher than 0.3 were significantly 600 different from the negative controls (p < 0.05, c.f. Fig. 2a,b).

601

602 Figure 6. Flow diagram describing differential processing for processing for signal 603 recognition in *Teleogryllus* species. (Top): representative song signals for each species. (First 604 recognition level): sensitivity to carrier frequency given by the frequency response of the 605 tympanic ear and sensory cells depicted as tuning curves. (Middle level): processing of the 606 pulse pattern within the phonemes of chirp and trill depicted by sensory templates for pulse 607 rate and integration of sound energy for duty cycle evaluation. (Bottom level): integration of 608 processing across time scales of both phonemes of chirp and trill. Common filters for carrier 609 frequency of the song are differently tuned in the two species, leading to quantitative 610 differences in female responses (grey boxes: brown lines indicate preferences for lower carrier frequencies by T. commodus, blue lines for higher carrier frequencies by T. 611 612 *oceanicus*). Both species also share similar filters for the pulse rate during the chirp portion 613 (greyboxes: black rectangles indicate sound pulses, brown lines (T. commodus) and blue lines (T. oceanicus) indicate sensory templates with rebound properties that will respond best to 614 615 the given pulse rate in the chirp pattern (Schöneich, Kostarakos, & Hedwig, 2015)). A

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616 qualitative difference is a more selective pulse rate filter in T. commodus for pulse rates during the trill part of a song, while T. oceanicus remain largely unselective for the trill 617 618 pattern provided sound energy remains high (i.e. sensitivity for high duty cycle, yellow 619 boxes: filters for trill pulse rate are symbolised by a rebound oscillation of the sensory 620 template, filters for pulse duty cycle by an integration). Separate streams of information about 621 chirp and trill features are finally integrated similarly for song recognition and discrimination 622 in both species. In aggregate, while females of both species might employ similar algorithms to process incoming signals on the basis of carrier frequency and chirp pulse rate (grey 623 624 boxes), they show divergent filter properties for the trill part (yellow boxes), for which T. 625 commodus females are more selective.







Fig. 2



628

629 **Figure 2**





632 Figure 3

Fig. 4



634 Figure 4

Fig. 5







637

638 Figure 6