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1	Increased carboxylate production in high-rate activated A-sludge by
2	forward osmosis thickening
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22 Abstract

23 Domestic wastewater represents a considerable feedstock for organics but the high dilution makes their recovery typically unsuccessful. Here we investigated three routes to 10-fold concentrate the organics 24 25 using Forward Osmosis (FO) (Draw solution (DS) 2.2 M MgCl₂): directly on domestic wastewater, A-26 sludge, or secondary sludge, with the end goal of increasing volatile fatty acid (VFA) yield from 27 subsequent 9-day fermentation tests. Forward osmosis concentrated the total COD by a factor of 8.2 \pm 1.2, 10.1 \pm 2.4 and 4.8 \pm 0.2 with respect to the raw streams of wastewater, secondary sludge and A-28 sludge. The soluble fraction of the COD was concentrated up to 3.5 times in the A-sludge and 2.1 times 29 in the secondary sludge; the result of a combined effect of the chemical action of Mg²⁺ (diffused from 30 31 the DS) on sludge disaggregation and cell lysis, and the physical action of recirculation and air-scouring 32 of the A-sludge in the FO-unit.

The FO-concentrated A-sludge produced 445 \pm 22 mg COD-VFA g⁻¹ COD_{fed}, which was 4.4 times higher than for the untreated A-sludge. No VFA were produced from untreated secondary sludge, but after FO-concentration 71 \pm 5 mg COD-VFA g⁻¹ COD_{fed} could be reached. Due to the low organics in wastewater even after FO-concentration (1.08 \pm 0.08 g COD L⁻¹), no notable VFA production occurred. The combination of A-stage technology and membrane technology for dewatering and COD concentration could be a key advancement to increase VFA production from domestic wastewater, whereby at least 45% of the COD can be recovered as valuable VFA.

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42 Keywords: Carboxylate fermentation; Forward osmosis (FO); AB system; Volatile fatty acids (VFA);
43 Waste activated sludge (WAS).

44 1. Introduction

The global domestic wastewater production is estimated at 330 billion m^3 annually [1]. Significant 45 46 research efforts are focused on the recovery of water and nutrients [2] from this waste stream and particularly in the conversion of the organic fraction into energy as biogas. Recently, an alternative route 47 48 was developed whereby these organics are converted into volatile fatty acids (VFA), which are building 49 blocks for a multitude of valuable products such as biopolymers [3], medium or long chain fatty acids 50 [4] and biofuels [5]. However, the generally low organic content of domestic wastewater (< 600 mg L⁻ 51 ¹) hampers efficient recovery, and is one of the main limitations in developing feasible bioproduction platforms [6]. 52

An evident means to concentrate organics is to work via sludge, generated from the wastewater as substrate and separated via settling. The conventional activated sludge (CAS) process is typically performed with high aeration energy, and at solid retention time (SRT) of 8 - 20 days, resulting in a very efficient organic carbon removal, but yielding a sludge with poor digestion efficiency due to the high sludge age and high sludge oxidation [7, 8].

An attractive alternative approach is the "Adsorption-Bioxidation" (AB) process [9]. The AB system is a two-stage treatment system where the first stage (A-stage) is typically operated at low hydraulic retention time (HRT 15 - 30 min) and low SRT (between 3 hours and 1 day). The soluble and colloidal organic matter is removed in the A-stage through bioflocculation into microbial A-sludge. The A-sludge is also separated in a clarifier, while the effluent is then taken to the subsequent B-stage to ensure polishing of the wastewater [2, 9]. A-sludge is usually digested to biogas with high digestion efficiency (higher than that of CAS) due to the remarkably low sludge age and high biodegradability [9, 10].

However, due to the poor settling performance of the A-sludge, the COD content is still too diluted to ensure high VFA production [6, 11]. A-stage is operated at 2 to 10 kg BOD kg⁻¹ VSS d⁻¹ [9] while optimum settling is obtained between 0.3 and 0.6 kg BOD kg⁻¹ VSS d⁻¹ [12]. CAS systems are typically operated at 0.25 kg BOD kg⁻¹ VSS d⁻¹ [13], achieving good sludge settling but with a relatively high water content. Thus, a further concentration of both A-sludge and secondary sludge is needed to ensure sufficiently high VFA production for extraction (> 5 g L^{-1}) and further valorization into high value products [14, 15].

72 Forward osmosis (FO) is a potentially energy-efficient and low-cost technique for concentration of 73 challenging feed streams such as waste activated sludge (WAS) [16, 17] and domestic wastewater [18]. 74 FO is based on the spontaneous process of osmosis, in which water is transported through a semi-75 permeable membrane from a feed solution (FS) of relatively low solute concentration (low osmotic pressure) to a draw solution (DS) of relatively high solute concentration (high osmotic pressure). The 76 membrane allows for water transport but displays a high rejection of solutes [19]. Draw solutes, such as 77 MgCl₂, need to have a low molecular weight, a high diffusivity, produce high osmotic pressures, are 78 79 well-retained by the FO membrane, are non-toxic and cheap [20]. Concentrating any stream using FO 80 causes an increase in its dissolved solutes content, due to a build-up of rejected solutes from the feed itself as well as the diffusion of solutes from the draw solution into the feed. The latter is referred to as 81 82 reverse salt diffusion (RSD).

83 Here, we applied FO to concentrate the organics of domestic wastewater, high-rate activated A-sludge 84 and secondary sludge to undergo a subsequent fermentation for VFA production purposes. FO 85 dewatering fluxes of the different streams were compared. MgCl₂ was chosen as draw solute because of 86 the high osmotic pressure generated by these solutions combined with the lower RSD compared to monovalent salts such as NaCl [19]. VFA yields and composition of the concentrated streams were 87 measured and compared to the non-concentrated streams to assess the effect of concentration by FO. 88 The effect of salt leakage during FO was investigated by carrying out fermentation experiments with 89 and without addition of MgCl₂. 90

91 2 Materials and Methods

92 2.1 Substrate collection

A-sludge was collected from the return flux to the A-tank in Nieuwveer WWTP (Breda, The Netherlands). Domestic secondary sludge and domestic wastewater were collected from Destelbergen WWTP (Destelbergen, Belgium). Characteristics of the substrates are summarized in Table 1. Mixed liquors as collected were used as FO feed. Part of a second batch of A-sludge (7.76 ± 0.16 gCOD L⁻¹, 4.41 ± 0.14 gVSS L⁻¹) was stored at 4°C and gravity-concentrated (removing the liquid volume above the settled sludge – resulting in a concentration of 3.8 times) to carry out fermentation tests to study the effects of RSD.

100 2.2 Aerobic and Anaerobic Forward osmosis (FO) experiments

101 A schematic overview of the setup is given in Figure S1 (Supplementary information (SI)) and a 102 description of the set-up, cross-flow velocity, flow measurements and analysis are provided in the 103 methodology section of SI.

The FO membranes used in this study (supported in a membrane module, Figure S1), were commercial thin film composite (TFC) membranes produced by Hydration Technology Innovations (HTI) (Albany, Oregon, USA) [21]. The membrane orientation in this study was AL-FS (active layer facing the feed solution). For each test, a new membrane coupon of 14 cm x 4 cm (56 cm²) was used. 10-fold concentration of the FO feeds was done by recirculating a fixed volume of feed in batch mode. Experiments were carried out in triplicate.

110 The DS was maintained at 2.2 M MgCl₂ by recirculating it over a bed of solid MgCl₂ when necessary 111 using a PID control based on conductivity measurements. Aeration (0.6 L min⁻¹) was applied to the FS 112 just before entering into the membrane cell to increase scouring and to reduce membrane fouling.

An anaerobic FO set-up was built to investigate the potential effect of microbial respiration during FO treatment. In order to keep the feed compartment anaerobic and to mitigate concentration polarization and fouling, an N_2/CO_2 (9:1 ratio) gas mixture was recirculated in the feed compartment (0.6 L min⁻¹).

116 2.3 Fermentation experimental procedure

117 Fermentation batch tests were carried out in triplicate with fermentative inoculum as described in SI.
118 The 3 feed substrates were fermented before and after FO-concentration. A-sludge was also fermented
119 after anaerobic FO-concentration. To avoid overloading of the fermentation tests by organics, different
120 substrate quantities were tested according each feed concentration as defined in Table S1.

To assess the reasons for changes in VFA production before and after FO-concentration, extra experiments were carried out: the effect of sludge concentration, aeration during FO-concentration, RSD or a combination of these effects was evaluated towards changes in soluble COD in the substrate and VFA production. To this extent, the raw A-sludge and the gravity-concentrated A-sludge (obtained removing the liquid volume above the sludge settled over night at 4 °C) were subjected to three different types of treatments:

- i) Addition of 7.1 g L⁻¹ of MgCl₂ (same concentration as in the FO-concentrate A-sludge
 (Table 2));
- 129 ii) Addition of 28.4 g L⁻¹ of MgCl₂ (salt overdose to evaluate the effect of build-up salinity
 130 and conductivity in acidogenesis)
- 131 iii) Air bubbling (flow 0.6 L min⁻¹, the same used during the FO);
- iv) A combination of conditions i) or ii) with condition iii).

All conditions were applied for 37.3 hours (same time needed to FO-concentrate the A-sludge 10 times
(Table 2)). Fermentation tests were performed in each of the pre-treated substrates as indicated in Table
S1. The potential impact of RSD on cell lysis was determined by flow cytometry with live/dead staining
as described in SI.

137 All the analyses carried out during fermentation tests were performed as described in the SI.

138 2.4. Statistical analysis

139 Statistical significance between intact cell densities, cell proportions and VFA concentrations for the 140 different experiments were determined by generalized mixed model regression of the cell densities with the experimental condition (e.g., GC-ASM, etc.) as a categorical predictor. A random intercept effect was incorporated for each replicate analysis. All models were first checked for normal distributed residuals (Shapiro Wilks test and QQ-plots) and homogenous variance (Levene's test). Parameters were estimated by the maximum likelihood method. Posthoc analysis was performed in case of a significant treatment effect (p < 0.05) by Tukey's all pair comparison method. Pair-wise comparisons were considered significant for p < 0.01.

147 3. Results and Discussion

- 148 3.1 Concentration of sewage and sludge by FO
- 149 3.1.1 Dewatering performance/flux behavior in FO

The goal of the FO treatment was to concentrate the different substrates and produce a concentrate rich
in organic carbon for fermentative VFA production. To this end, A-sludge, secondary waste activated
sludge (WAS) and domestic wastewater were concentrated 10-fold by FO in batch mode.

153 During FO concentration, the water flux across the membrane decreases over time due to fouling and increased salinity build-up on the concentrate side. The flux profile obtained for the different feed 154 solutions is shown in Figure 1. Repeated experiments for the respective feed solutions (n = 3) yielded 155 156 similar flux behavior (Figure S2). Despite the relatively similar conductivities of the different substrates, 157 the initial fluxes, defined as the average flux during the first 2 hours of FO concentration, were $11.4 \pm$ 158 0.2 LMH (L m² h⁻¹) for secondary sludge, 8.0 ± 0.7 LMH for A-sludge and 9.6 ± 0.4 LMH for 159 wastewater, respectively. We hypothesize that the different initial fluxes between the different streams 160 can be explained by the feed characteristics in terms of different solids (total solids (TS), volatile solids 161 (VS) and suspended solids (SS)) and organic content (soluble COD (sCOD) and total COD (tCOD)) (Table 1). Secondary sludge yielded the highest initial flux, approaching the DI water baseline. This was 162 consistent with an earlier study from Cornelissen et al. [16] that found similar flux values for secondary 163 activated sludge and DI water during 7 to 8 hour experiments (5.1 - 6.2 LMH for the DI water and 5.8 164 LMH for the activated sludge, respectively by using a TFC FO-membrane and 0.5 M NaCl draw 165 solution). Secondary sludge had a low content of soluble or colloidal VS (calculated as difference 166 167 between VS and VSS) compared to A-sludge. A-sludge contained the highest amounts of TS, VS and

SS of all feeds. Moreover, the sCOD content of A-sludge, and the sCOD/tCOD ratio $(1.25 \pm 0.02 \text{ g L}^{-1})$ 168 and 15 %, respectively) were higher than that of secondary sludge (0.22 \pm 0.01 g L⁻¹ and 5 %, 169 respectively) (Table 1). Thus, we hypothesize that a combination of a fast formation of a dense fouling-170 layer on the membrane, combined with cake-enhanced concentration polarization caused the initial 171 172 lower flux during A-sludge FO-dewatering. The latter is due to the dominance of small, soluble/colloidal 173 organic compounds which potentially impart extra osmotic pressure in A-sludge, caused the initial lower flux of A-sludge during FO-dewatering. In the case of secondary sludge, the sludge flocs could aid in 174 175 scouring the membrane surface and in sequestering the soluble organics, thus leading to less fouling (or potentially a cake with less hydraulic resistance). Wastewater had the lowest VS/TS ratio (38 %) but the 176 177 highest sCOD fraction of tCOD (25 %) (Table 1). Similar conditions were found by Lutchmiah et al. 178 [18] who reported a 20% reduction of the initial dewatering flux from DI water to wastewater. It is likely that the dominance of small, soluble organic compounds in wastewater caused the rapid formation of a 179 gel layer on the membrane and a higher osmotic pressure in the proximity of the membrane, resulting 180 181 in a lower initial flux compared with that of secondary sludge.

182 For both A-sludge and secondary sludge (containing higher solids concentration), the initial flux 183 remained stable for the first 5 hours of filtration, after which the flux declined linearly for all repetitions (Figure S2). Flux decline rate for the secondary sludge $(0.302 \pm 0.005 \text{ LMH h}^{-1})$ was twice that of the 184 A-sludge (0.114 \pm 0.029 LMH h⁻¹), which might be related to the higher initial flux of the secondary 185 sludge. It was visually observed that the secondary sludge thickened to a highly viscous mixture by the 186 187 end of the FO concentration. This caused the formation of a thick cake on the membrane, which inhibited further sludge dewatering depicted by a profound flux decline in Figure 1. The A-sludge did not become 188 189 viscous and the final flux decline was less pronounced, but FO-dewatering lasted longer than that for 190 the secondary sludge to reach the same 10-fold concentration (28.1, 29.7 and 37.3 hours for secondary 191 sludge, wastewater and A-sludge, respectively (Table 2)) due to the lower average flux. Consistently 192 with what reported in literature [18], only limited flux decline occurred during wastewater concentration. 193 Most likely, the relatively low fouling propensity of the wastewater was related to the low total 194 concentration of organics in the wastewater (Table 1) and the low concentration of suspended solids.

The concentration ratio of the organics during FO was determined as FO-concentrated COD to feed 196 197 COD. As a 10-fold volume reduction was reached, the same theoretical organics concentration ratio was expected, assuming that no organics are transported through or adsorbed onto the membrane, converted 198 into biomass or mineralized. The COD concentration ratio was only 8.2 ± 1.2 for the domestic 199 200 wastewater and 4.8 ± 0.2 for the A-sludge, but 10.1 ± 2.4 for the secondary sludge. This indicates a 201 COD loss of 18 % for the domestic wastewater, 52 % for the A-sludge and no loss for the secondary sludge. The high organic loss for the A-sludge might be due to biological degradation, given its high 202 203 biodegradability, low sludge age and high initial organic content [9, 22, 23] and the constant aeration applied for membrane scouring. Its dewatering also took the longest, allowing more time for organic 204 205 loss though biological activity. Tests to demonstrate this are reported in the next section. Secondary 206 sludge contains less biodegradable organics as most of the SS and organics from sewages are removed during upstream primary treatment [7] and due to the extent oxidation and high SRT of the secondary 207 treatment (CAS) itself. Hence, no detectable COD loss could be observed during secondary sludge FO 208 209 concentration. Wastewater showed a modest COD loss. The organic matter in wastewater is considered 210 relatively easy to degrade, as it is soluble and has not yet been incorporated in microbial biomass, however, wastewater typically does not contain an enriched population of organisms able to degrade 211 212 organics, which are present in sludge [7, 9, 24].

213 FO concentration in general caused an increase in the dissolved compounds content of the feed streams, 214 which is caused both by concentrating the initial feed solutes in a smaller volume and by diffusion of draw solutes into the feed. This can be seen in Table 2, where both conductivity and Mg²⁺ content of the 215 feed streams are shown. The conductivity of the feed streams increased from about 1.5 to about 30 mS 216 cm⁻¹ in the case of wastewater and A-sludge, and to about 20 mS cm⁻¹ in the case of secondary sludge. 217 The measured conductivity of the concentrates does not relate well to the MgCl₂ concentration measured 218 219 on filtered samples by ion chromatography (IC). Apparently, the different types of flocs in the different types of sludge appeared to be specifically adsorbing different amounts of Mg^{2+} . This became clear by 220 221 freezing and thawing concentrated secondary sludge, which caused the conductivity to increase from

222 18.2 to 51.3 mS cm⁻¹ after thawing, indicating that $MgCl_2$ was sequestered by EPS or intracellular 223 uptake, which was subsequently released by lysis of the sludge structure when freezing-thawing. As a 224 result, no clear correlations can be seen between flux decline and measured Mg^{2+} concentrations.

225 3.2 Identifying organic loses during FO concentration

226 During FO concentration of A-sludge, half of the incoming COD was lost, almost twice as much as in 227 domestic wastewater. To elucidate if this loss was caused by bacterial respiration, 10-fold FO concentration under anaerobic conditions was targeted with the A-sludge. Considerably lower FO flux 228 in anaerobic conditions caused that the concentration factor of 10 was not reached. The initial flux was 229 about 7.5 LMH (Figure S3), similar to the initial flux during aerobic A-sludge concentration (8.0 LMH 230 (Figure 1)). Flux declined steadily down to 0.5 - 1 LMH (after 30 hours) and then remained constant for 231 232 the rest of the FO treatment (Figure S3). After 95.4 hours, a concentration factor of only 3.9 was reached, 233 and the test was aborted in order to limit the salinity build-up in the feed due to RSD. In contrast to aerobic FO experiments, the COD content increased from 5.5 ± 0.5 to 21.4 ± 1.7 g L⁻¹, equaling a $3.9 \pm$ 234 0.1 -fold increase, equal to the concentration factor obtained. Chen et al. [25] studied the treatment of 235 236 low-strength wastewater using an anaerobic osmotic membrane bioreactors, reporting a flux decline to 237 3.5 LMH. The stable flux obtained during anaerobic FO concentration in this study is lower of that 238 obtained in literature [29], due to a higher initial COD of the A-sludge (8.40 gCOD L⁻¹ and 0.46 gCOD L⁻¹, respectively). 239

240 The large extent of flux decline, starting already after few hours of anaerobic FO-concentration (Figure 241 S3), was most likely due to a major formation of gel-like substances (and deposition of these substances 242 on the membranes surface) by the high concentration of organics retained in the feed compartment, which prevented dewatering. For concentration of A-sludge, especially under anaerobic conditions, it is 243 thus clear that a continuous scouring of the membrane, using alternative techniques in combination with 244 245 N_2 :CO₂ (9:1) gas bubbling, is needed to prevent flux decline. However, a continuous flow of oxygen free gases would increase the operational cost of the system, and just using air (as in the aerobic FO-246 dewatering) will result in COD-loss. Alternative scouring systems, such as (osmotic) backwashing, 247 248 relaxation or periodic air bubbling should be considered.

249 3.3 Effect of FO-concentration on fermentation

250 *3.3.1 Fermentation yields of the different streams*

High initial organic matter content is needed to obtain high VFA production [6]. Fermentation batch tests were performed on all the streams (domestic wastewater, secondary sludge and A-sludge) as well as their FO-concentrates to assess if the application of an FO system after a CAS, an A-stage or directly to the domestic wastewater could improve VFA production and yields. During fermentation tests, maximum VFA production was obtained between day 4 and 7 followed by a decrease in product concentrations due to conversion of VFA to methane.

No notable VFA production occurred for domestic wastewater, either before or after the FO 257 concentration. To maximize the available organics for VFA production from the domestic wastewater, 258 259 the organic load during fermentation was increased by 6 times compared to the initial test (Table S1), but again no notable VFA production occurred (Figure 2). The tCOD after FO-concentration of the 260 domestic wastewater was 1.08 ± 0.08 g L⁻¹ and the sCOD was 0.24 ± 0.01 g L⁻¹. This concentration was 261 insufficient to sustain the growth of the microorganisms and, to a lesser extent, allow substantial 262 263 production of VFA or CH₄. Higher concentrations still might be needed in order to produce VFA from 264 FO-concentrated domestic wastewater, however this would further increase contact time in the FO, leading to more draw solutes leaking and increased conductivity. Such highly conductive feedstocks 265 could be used for VFA accumulation as conductivity above 30 mS cm⁻¹ negatively affects 266 267 methanogenesis while acetogenic bacteria still produce VFA at high salt concentrations De Vrieze et al. [26]. However, an economic assessment would be required as draw solute leakage comes at a cost. It is 268 unknown whether other factors, such as accumulation of toxic compounds, could also play a role in the 269 observed lack of VFA production. 270

Fermentation of the FO-concentrated secondary sludge led to a production of 71 ± 5 mg COD-VFA g⁻¹COD_{fed} after 7 days, corresponding to 7 % of COD converted into VFA. No notable VFA production occurred for the raw secondary sludge. The digestion efficiency of the secondary sludge and the VFA yields produced during fermentation are low compared to substrates typically used in fermentation. This is mainly due to the high sludge age and the extent of oxidation [6, 8] of the secondary sludge, evenwhen the concentration is increased in the FO treatment.

For the FO-concentrated A-sludge, a VFA production of 346 ± 30 mg COD-VFA g⁻¹COD_{fed} was reached 277 already after 1 day of fermentation (Figure S4), with a production up to 445 \pm 22 mg COD-VFA g⁻ 278 ¹COD_{fed} after 4 days (Figure 2, Figure S4). For the raw A-sludge, the VFA production after 4 days of 279 fermentation was only $102 \pm 3 \text{ mg COD-VFA g}^{-1}\text{COD}_{\text{fed}}$, which is 4.4 times lower (p < 0.01) than that 280 for the FO-concentrated A-sludge (Figure 2). This shows that FO-concentration increases fermentability 281 of A-sludge as higher VFA is produced compared to the same quantity of organics fed. Fermentation 282 was also performed with the anaerobic FO-concentrated A-sludge. VFA production already reached its 283 284 maximum after 2 days of fermentation with $444 \pm 8 \text{ mg COD-VFA g}^{-1}\text{COD}_{\text{fed}}$ (Figure 2) (significantly higher than A-sludge itself with p < 0.01). This fast start-up of VFA formation might be attributed to 285 hydrolysis of the sludge already starting during the 95.4 hours of anaerobic FO treatment. 286

287 *3.3.2 Factors affecting improvement in fermentation capacity*

FO-concentration of the two sludge types increased their overall COD as well as the sCOD concentration (Table 3). For secondary sludge the increase corresponded to 10.1 ± 1.0 % sCOD/tCOD while for Asludge, this rose up to 30.4 ± 1.9 % sCOD/tCOD. Two main reasons why FO-concentration increases the sCOD fraction might be a sort of physical pretreatment by the recirculation through the peristaltic pump (for 37.3/95.4 hours), as well as the shear force of the continuous air bubbling in the feed compartment. Because of this improvement on sCOD fraction and overall increased organics content, specific VFA production improves after FO-concentration of sludge.

During FO, water molecules follow the osmotic gradient from the FS to the DS; likewise the ions 295 296 contained in the DS can migrate to the FS. One of the hypotheses we put forward in this manuscript, is 297 that the draw solute used for the dewatering might also have an impact on fermentation through improved hydrolysis. A high concentration of cations such as Mg²⁺ (Table 2) facilitates the formation 298 299 of single-cells which are more sensitive to lysis by spontaneous disaggregation from the floc structure [27] (hypothesis further evaluated in the next sections). Furthermore, several ions can also interact with 300 301 the negative charges of the EPS (extracellular polymeric substances) surrounding the bacterial cells, 302 breaking the bounds among these polymers and favoring cell lysis and release of organic matter [28].

Finally, an increased concentration of the sludge can enhance the hydrolysis due to overload shock [29].
The increased organic loading can also lead to the accumulation of carboxylates, which are crucial in
inhibiting methanogenesis, increasing the overall VFA accumulation [30].

306 *3.3.3 Fermentation product speciation linked to stream composition*

307 A different VFA speciation was found for the different streams and also for the same stream before and 308 after the FO-concentration (Figure 3). In case of the A-sludge before FO, acetate was the most abundant VFA formed after 4 days of fermentation $(60 \pm 3\%)$ besides $26 \pm 3\%$ propionate and $9 \pm 0\%$ isovalerate. 309 In the FO-concentrated A-sludge, the proportion of acetate dropped to 24 ± 4 % after 4 days of 310 fermentation, with propionate increasing to 43 ± 3 %. Moreover, a larger proportion of longer chain 311 312 fatty acids was produced after 4 days of fermentation for the FO-concentrated A-sludge compared to the non-concentrated sludge, $4 \pm 0\%$ isobutyrate, $7 \pm 0\%$ butyrate, $8 \pm 0\%$ isovalerate and $8 \pm 0\%$ valerate 313 which shows an impact of higher loading. 314

Similar VFA speciation was found for the anaerobically FO-concentrated A-sludge, with 31 \pm 0 % 315 acetate and 35 ± 0 % propionate present after 2 days of fermentation. The proportion of longer chain 316 317 fatty acids also increased with 6 ± 0 % isobutyrate, 10 ± 0 % butyrate, 11 ± 0 % isovalerate and 5 ± 0 % 318 valerate, for anaerobic FO-concentration of A-sludge, confirming again that the application of FO may lead to the accumulation of longer chain fatty acids again due to increased loading. For the FO-319 concentrated secondary sludge, acetate and propionate were the main VFA formed, at 46 ± 3 % and 18 320 321 \pm 0 % respectively, which is similar to the raw A-sludge with approximate sCOD fractions of that of secondary sludge (14.9 ± 0.7 % and 10.1 ± 1.0 % sCOD/tCOD, respectively (Table 3)). FO-concentrated 322 secondary sludge fermentation also led to the production of longer chain fatty acids, at 6 ± 0 % 323 isobutyrate, 7 ± 0 % butyrate and 13 ± 0 % valerate after 4 days of fermentation (Figure 3). 324

Production of CH₄ at day 4 was relatively low for the FO-concentrated streams with 7 ± 2 mg COD-CH₄ g⁻¹COD_{fed} (9% initial COD) for secondary sludge and 29 ± 3 mg COD-CH₄ g⁻¹COD_{fed} (5% of initial COD) for the A-sludge, while 6 ± 1 mg COD-CH₄ g⁻¹COD_{fed} (6% of initial COD) was obtained with the raw A-sludge (Figure 3). The use of an FO-system to concentrate A-sludge and secondary sludge thus seems to promote the production of more VFA with a higher proportion of longer chain fatty acids 330 (C_4-C_5) , but not improving the conversion of VFA into methane. The increased organic loading together 331 with the increased conductivity reduces the acetoclastic activity of methanogens and thus leads to higher 332 VFA production and accumulation [28, 30, 31].

333 3.4 Effect of FO operational conditions on fermentation

334 FO-concentration of both secondary sludge and A-sludge increased VFA yields, from 0 to 7 % and 10 335 to 45 %, respectively. To understand whether the main reason for this was 1) the increased organics concentration due to dewatering, 2) the enhanced hydrolysis of the organics by RSD, 3) enhanced 336 breakdown of the cells due to the air scouring, or 4) any combination of these phenomena, 9-days 337 fermentation batch tests were performed as described in section 2.3. The effect of the increase in 338 organics concentration was investigated by gravity-concentrating the sludge; the potential enhanced 339 340 hydrolysis by RSD was investigated by adding a 7.1 g L⁻¹ MgCl₂ (same concentration in the A-sludge after aerobic FO-concentration) or 28.4 g L⁻¹ MgCl₂ (concentration needed in order to reach the same 341 342 conductivity after anaerobic FO-concentration of the A-sludge); and the potential breakdown of cells 343 due to air scouring was investigated by aerating at 0.6 L min⁻¹. All treatments were carried out for 37.3 344 hours, same time needed to reach a 10-fold by FO-concentration of A-sludge. Results are depicted in 345 Figure 4.

For the non-concentrated A-sludge, addition of MgCl₂ increased the sCOD/tCOD ratio from 19.0 % to 25.0 % (with low salt) and to 27.5 % (with high salt), respectively (Table 3). However, production of VFA staid constant at 265 \pm 3 mg COD–VFA g COD_{fed} with low salt addition and only 177 \pm 5 mg COD–VFA g COD_{fed} were produced by increasing the MgCl₂ up to 28.4 g L⁻¹ (Figure 4). For all the other conditions tried VFA production was lower than that of untreated A-sludge. Thus, for the nonconcentrated A-sludge, neither MgCl₂ addition nor aeration nor the combination of these could explain the increased VFA production resulting from the FO-concentration.

Gravity concentration of the A-sludge did not improve the VFA yield. Only $196 \pm 13 \text{ mg COD -VFA g}$ COD_{fed} (corresponding to 20 % yield) were produced by fermenting gravity-concentrated (GC) Asludge, significantly lower (p < 0.01) than the yield obtained from non-concentrated A-sludge (around 26 %). This might be caused as gravity-concentrating the A-sludge 3.8 times was carried by removing 357 the supernatant rich in sCOD after settling (Table 3), but consequently the sCOD/tCOD ratio was 358 reduced from 19.0 to 13.1 % (Table 3). This is due to the fact that after gravity concentration the 359 wastewater rich in biodegradable organics (sCOD/tCOD 26.7 % (Table 3)) was removed and only the 360 sludge settled at the bottom, which is more difficult to hydrolyze, was used for the fermentation. Adding 361 MgCl₂ to the GC-A-sludge increased the sCOD/tCOD ratio somewhat from 13.1 % to 16.1 % (Table 3) resulting in a VFA yield of 248 \pm 7 mg COD-VFA g COD_{fed} (25 %), significantly higher (p < 0.01) 362 than that of GC-A-sludge (20 %) (Figure 4). This indicates that bringing the concentrated sludge in 363 contact with a relatively high concentration of salt (7.1 g L⁻¹) causes some disaggregation of the sludge 364 flocs and enhances the hydrolysis of the cells as reported by Chen et al. [28]. Again, although raising 365 366 the MgCl₂ up to 28.4 g L⁻¹ increased the sCOD/tCOD ratio from 13.1 % to 14.2 %, only 158 ± 1 mg COD -VFA g COD_{fed} were produced (significantly lower than that of GC-A-sludge with p < 0.01). High 367 conductivity ($< 40 \text{ mS cm}^{-1}$ (Table S2)) might be affecting batch fermentation. 368

Aerating the GC-A-sludge for 37.3 hours decreased the sCOD/tCOD ratio (from 13.1 % to 10.8 %) due to COD uptake for biomass growth. Combination of aeration and salt addition (7.1 g L⁻¹ MgCl₂) resulted in increased sCOD/tCOD ratio to 12.6 %. Combining GC-A-sludge with aeration produced 197 \pm 2 mg COD -VFA g COD_{fed}, and with aeration and 7.1 g L⁻¹ MgCl₂ production was 210 \pm 1 mg COD -VFA g COD_{fed} (Figure 4). Further increase of the MgCl₂ concentration, yet again, resulted in a significantly lower (p < 0.01) VFA production (132 \pm 8 mg COD -VFA g COD_{fed}).

375 In all conditions tested, VFA production yield from GC-A-sludge was lower than for the FO-376 concentrated A-sludge (between 16 and 25 %) (Figure 4), suggesting that none of these treatments nor the combination of them - alone - could explain the increased VFA production obtained after FO-377 378 concentration. The sCOD/tCOD ratio of the different treatments on GC-A-sludge varied between 10.4 379 % and 16.1%, while recirculating A-sludge throughout the FO unit increased the ratio up to 30.4% for 380 the aerated system and 52.5 % for the anaerobic system (Table 3) (experiments run for 37.3 and 95.4 381 hours, respectively (Table 2)). Therefore, although the combination of sludge concentration, aeration and salts leakage to the FS increased sCOD/tCOD, the main cause of VFA production enhancement was 382 probably due to the physical action of the peristaltic pump and the scouring of the bubbles that enter the 383

feed compartment. This leads to disaggregation of the sludge flocs promoting cell lysis. This hypothesis
was further investigated in the next section.

386 3.5 Effect of FO-concentration on cell lysis

387 The number of intact planktonic cells and permeabilized cells, thus damaged by lysis, in the GC-A-388 sludge and the FO-concentrated A-sludge was determined with a live/dead staining coupled to flow-389 cytometry. These measurements were performed to confirm the hypothesis that FO facilitated breakage of sludge flocs and cells (Figure 5) leading to enhanced fermentability. Intact and permeabilized cells 390 391 are defined as bacterial cells with a cellular membrane that is either impenetrable (intact) or penetrable (permeabilized) by the nucleic acid binding compound propidium iodide (PI) [32]. (Table S3). Although 392 393 by gravity concentrating the A-sludge a 3.8-fold concentration could be reached, the concentration of total intact cells only increased (significantly) from 4.11 x $10^{+9} \pm 2.12$ x 10^{+7} to 9.41 x $10^{+9} \pm 5.66$ x 10^{+7} 394 cells mL⁻¹, corresponding to a 2.3-fold increase. Consequently, the intact cells/total cells ratio 395 significantly decreased from 68.2 ± 0.4 % to 56.1 ± 2.2 % (Figure 5) after gravity concentration 396 397 indicating that part of the intact cells (both sludge flocs and single cells) were present in the discarded 398 supernatant. The changes in intact cells/ total cells ratio followed the same behavior for both A-sludge and GC-A-sludge under different treatments (Figure 5). Addition of 7.1 g L⁻¹ MgCl₂ resulted in a 399 400 significant decrease of the number of intact cells/ total cells ratio to 54.0 ± 3.7 % for the A-sludge and down to 29.5 ± 0.8 % for the GC-A-sludge, respectively. These differences confirmed that Mg²⁺ and Cl⁺ 401 ions have a lytic effect on the sludge flocs and cells, which simultaneously disaggregates the flocs and 402 403 releases sCOD (Table 3). This was even more pronounced upon adding higher concentration of MgCl₂ (28.4 g L⁻¹), the intact cells/ total cells ratio significantly decreased to 29.5 ± 0.8 % for the A-sludge and 404 405 down to 15.9 ± 0.4 % for the GC-A-sludge. Aeration resulted in an increase of the intact cells/ total cells 406 ratio up to 69.0 ± 1.2 % for the A-sludge and up to 61.8 ± 1.4 % for the GC-A-sludge (Figure 5). This 407 was most likely due to biomass respiration and thus growth, with consequent reduction of sCOD. 408 Combining aeration and salt addition (7.1 g L^{-1}) the intact cells/ total cells ratio significantly increased up to 75.7 ± 2.9 % for the A-sludge and to 65.9 ± 5.6 % for the GC-A-sludge when compared to the 409 mere aeration. In contrast, by increasing the concentration of the MgCl₂ to 28.4 g L⁻¹ with active aeration, 410 the intact cells/ total cells ratio significantly decreased to 51.6 ± 1.9 % for the A-sludge and down to 411

 35.2 ± 0.8 % for the GC-A-sludge. This supports the initial hypothesis that the lytic action of MgCl₂ 412 413 increased the sCOD which was taken up to enhance biomass growth when aerating, only when 414 conductivity values (at low salt addition) did not affect the microbial performance. The intact cells/total 415 cells ratio for the aerobically FO-concentrated A-sludge was rather high (68.2 \pm 11.9 %). For the 416 membrane-compromised fraction of FO-concentrated A-sludge (both anaerobically and aerobically 417 concentrated), permeabilised cell population overlapped with the background noise and was therefore questionable. Thus the absolute value of intact cells has to be considered over the intact cells/total cells 418 419 ratio. 10-fold and a 3.4-fold concentrations could be reached by FO-concentrating the A-sludge aerobic and anaerobically, respectively, while GC-concentration resulted in a 3.8-fold concentration of the A-420 421 sludge. Nonetheless, FO concentration resulted in an significantly lower (p < 0.01) intact cells content of 3.18 x $10^{+9} \pm 8.39$ x 10^{+7} , compared to that of GC-A-sludge and GC-A-sludge treated with a 422 combination of salt addition and aeration (8.86 x $10^{+9} \pm 9.53$ x 10^{+8} and 6.96 x $10^{+9} \pm 3.76$ x 10^{+8} , 423 424 respectively).

425 These findings are thus consistent with the hypothesis set above, that FO-concentration leads to a 426 combined effect of the chemical action of the salt on the sludge flocs and cells, and the physical action of the recirculation and the scouring by bubbles of the A-sludge in the FO-cell. The more the A-sludge 427 was recirculated throughout the FO unit the more the flocs were broken down and their constituent cells 428 lysed (Figure 5). The number of intact cells found in the anaerobically FO-concentrated A-sludge was 429 $2.13 \times 10^{+8} \pm 2.74 \times 10^{+7}$. This was significantly lower than all of the values reported for the gravity 430 concentrated sludge and the aerobically FO-concentrated sludge, confirming the mechanical shear effect 431 of FO on the breakage of the sludge, which ultimately resulted in increased sCOD and increased VFA 432 433 production yield.

434 **3.6 Economical assessment**

Replacing thickening with a more compact FO-concentration will lead to higher VFA conversion
efficiency (from 20 % to 45 %) but also to additional cost. To assess the economic viability of the
proposed technology, three scenarios were considered, comparing the use of either 2.2 M MgCl₂ or
seawater (0.5 M NaCl) as DS in the FO, to the scenario with a settling (thickening) based AB-system.
The costs for the technologies and their references are listed in Table S4 and a description of the

assumptions made to calculate capital expenses (CAPEX) and operational expenses (OPEX) is givenin section S4 (SI).

442 Table 4 then gathers the scenarios, where it can be seen that the CAPEX for the 2.2 M MgCl₂ FO-443 system is $0.79 \notin \text{m}^3$ sludge and for the seawater $1.06 \notin \text{m}^3$ sludge (due to the lower driving force and 444 higher membrane requirement). For the OPEX only the specific energy consumption (SEC) needed for 445 reverse osmosis (RO) reconcentration of the DS was considered (section S4, SI) and was as such calculated only for the scenario with 2.2 M MgCl₂ ($0.67 \in m^{-3}$ sludge) – it was assumed that when 446 447 using seawater as DS, the seawater is discharged back into the ocean. RO drinking water production from this seawater might be possible, and might lead to a lower total cost (as the income of the 448 449 drinking water needs to be taken into account). Hence, the total costs (OPEX + CAPEX) for FOconcentration of A-sludge amounted to 1.46 € m⁻³sludge and 1.06 € m⁻³sludge for 2.2 M MgCl₂ and 450 sea-water as DS, respectively. For the gravity concentration of A-sludge the CAPEX was 0.08 € m⁻ 451 ³sludge and the OPEX 0.06 € m⁻³sludge and thus the total costs (OPEX + CAPEX) were 0.14 € m⁻ 452 453 ³sludge (Table 4).

454 Profits obtained from the VFA production of FO-concentrated A-sludge (for both DS considered) and
455 the GC-A-sludge were calculated considering a conversion efficiency of 45 % for the former and 20 %

for the latter (sections 3.3.1 and 3.4) and amounted to $2.36 \notin m^{-3}$ sludge and $1.05 \notin m^{-3}$ sludge,

457 respectively. The net profits were comparable for the GC-A-sludge and for the A-sludge FO-

458 concentrated with 2.2 M MgCl₂, at 0.91 € m⁻³sludge (0.12 € kg⁻¹sludge) and 0.90 € m⁻³sludge (0.12 €

459 kg⁻¹sludge), respectively (Table 4). However, these calculations are based on current generation of FO

460 membranes in the market. Considerable research is currently aiming at novel FO-membranes with

461 increased water flux (LMH) during FO, reducing the CAPEX and therefore increasing the total profit.

462 In fact, several of these membranes are already close to commercialization [33]. In addition, it has to

463 be stated that, the use of FO system over conventional thickening for the sludge concentration has the

464 advantage of a smaller footprint at WWTP level.

465 When sea-water is used as DS, the total profit amounts to $1.30 \notin \text{m}^3$ sludge (0.17 $\notin \text{kg}^1$ sludge), higher

than that of the other scenarios considered but still underestimated. Indeed, for this scenario the same

467 conversion efficiency obtained for 2.2 M MgCl₂ was considered (45 %), but as in sea-water the

468 concentration of NaCl is 4.4 times lower, longer time would be needed to arrive to the same 10-fold
469 concentration. This would result in higher sCOD/tCOD of the FS compared to that reached with 2.2 M
470 MgCl₂ and therefore it would allow higher VFA production. On the other hand, the effect of Na⁺ on
471 cell lysis and sCOD increase and its effect on the subsequent fermentation should be further
472 investigated.

473 3.7 Strategies to further increase VFA production after A-sludge FO-concentration

FO-concentration of A-sludge increased the VFA conversion efficiency from 10 % to 45 % (4.4-fold), which is higher than previously reported [11]. It has been described that high salinity and high conductivity (above 30 mS cm⁻¹) negatively affect methanogenesis while acetogenic bacteria can acclimatize to high salt concentration and produce VFA [34]. FO-concentration of A-sludge could be a strategy to increase VFA production (sections 3.4 and 3.5) and, inhibit methanogens due to RSD, thus allowing VFA accumulation in a continuous system.

Conversion efficiency up to 70 % has been reported during A-sludge digestion [10] suggesting that the 480 481 potential for VFA production from A-sludge should be higher. Methodologies such as the use of acid 482 or alkali pretreatment [35, 36] and/or thermal pretreatment [6] and the application of alkaline pH to the 483 fermentation are reported to increase VFA production from sludge [37, 38]. Particularly, alkaline 484 conditions increase both hydrolysis and acidification rates and also increase solubilization of carbohydrates and proteins, main components of the sludge, resulting in higher VFA production [39]. 485 Therefore, the combination of FO-concentration of the sludge with pretreatments or with fermentation 486 487 at alkaline pH might promote VFA production and should be further investigated.

488

489 **4.** Conclusions

Forward Osmosis enabled 10-fold dewatering of domestic wastewater, secondary activated sludge and high-rate activated A-sludge, yet 18% and 52% of the COD were lost for the domestic wastewater and A-sludge, respectively. Aeration, used to scour the fouling layer on the membrane, promoted COD consumption, therefore other techniques to mitigate membrane fouling need to be investigated. Forward osmosis increased the soluble COD (sCOD) fraction in the sludge enhancing VFA production by 7 % for secondary sludge and by 35 % for A-sludge (from 10 % to 45 %) but did not enable VFA production for the domestic wastewater due to the limited sCOD. This proves
the importance of an intermediate step to convert the sCOD into microbial sludge before the FOconcentration. Increased VFA production for FO-concentrated streams is due to both chemical
action of the ions leaking from the draw solution and the physical effect caused by recirculation of
the stream and air scouring. Both effects together result in disaggregation of the sludge flocs and
lysis of the cells, increasing sCOD and VFA production.

502

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515 **5. References**

- 516 [1] J. Mateo-Sagasta, L. Raschid-Sally, A. Thebo, Global Wastewater and Sludge Production,
- 517 Treatment and Use, in: Wastewater, Springer, 2015, pp. 15-38.
- 518 [2] W. Verstraete, P. Van de Caveye, V. Diamantis, Maximum use of resources present in domestic
- ⁵¹⁹ "used water", Bioresource Technology, 100 (2009) 5537-5545.
- 520 [3] M.G.E. Albuquerque, V. Martino, E. Pollet, L. Avérous, M.A.M. Reis, Mixed culture
- 521 polyhydroxyalkanoate (PHA) production from volatile fatty acid (VFA)-rich streams: Effect of
- substrate composition and feeding regime on PHA productivity, composition and properties, Journal
- 523 of Biotechnology, 151 (2011) 66-76.
- 524 [4] M.T. Agler, B.A. Wrenn, S.H. Zinder, L.T. Angenent, Waste to bioproduct conversion with
- undefined mixed cultures: the carboxylate platform, Trends in Biotechnology, 29 (2011) 70-78.

- 526 [5] H. Ge, P.D. Jensen, D.J. Batstone, Pre-treatment mechanisms during thermophilic-mesophilic
- 527 temperature phased anaerobic digestion of primary sludge, Water Research, 44 (2010) 123-130.
- 528 [6] W.S. Lee, A.S.M. Chua, H.K. Yeoh, G.C. Ngoh, A review of the production and applications of
- 529 waste-derived volatile fatty acids, Chemical Engineering Journal, 235 (2014) 83-99.
- 530 [7] M. Von Sperling, Wastewater characteristics, treatment and disposal, IWA publishing, 2007.
- 531 [8] D. Bolzonella, P. Pavan, P. Battistoni, F. Cecchi, Mesophilic anaerobic digestion of waste
- 532 activated sludge: influence of the solid retention time in the wastewater treatment process, Process
- 533 Biochemistry, 40 (2005) 1453-1460.
- [9] B. Boehnke, B. Diering, S.W. Zuckut, Cost-effective wastewater treatment process for removal of
- organics and nutrients, Water Eng. Manage., 144 (1997) 30-&.
- 536 [10] J. De Vrieze, L. De Lathouwer, W. Verstraete, N. Boon, High-rate iron-rich activated sludge as
- stabilizing agent for the anaerobic digestion of kitchen waste, Water Research, 47 (2013) 3732-3741.
- 538 [11] C. Cagnetta, M. Coma, S.E. Vlaeminck, K. Rabaey, Production of carboxylates from high rate
- activated sludge through fermentation, Bioresource Technology, (2016).
- 540 [12] R. Ramalho, Introduction to wastewater treatment processes, Elsevier, 2012.
- 541 [13] PaDEP, Module 17: The Activated Sludge Process Part III. Pennsylvania Department of
- 542 Environmental Protection., (2014).
- 543 [14] S.J. Andersen, T. Hennebel, S. Gildemyn, M. Coma, J. Desloover, J. Berton, J. Tsukamoto, C.
- 544 Stevens, K. Rabaey, Electrolytic Membrane Extraction Enables Production of Fine Chemicals from
- 545 Biorefinery Sidestreams, Environmental Science & Technology, 48 (2014) 7135-7142.
- 546 [15] M. Agler, C. Spirito, J. Usack, J. Werner, L. Angenent, Development of a highly specific and
- productive process for n-caproic acid production: applying lessons from methanogenic microbiomes,(2013).
- 549 [16] E. Cornelissen, D. Harmsen, K. De Korte, C. Ruiken, J.-J. Qin, H. Oo, L. Wessels, Membrane
- 550 fouling and process performance of forward osmosis membranes on activated sludge, Journal of
- 551 Membrane Science, 319 (2008) 158-168.
- 552 [17] H. Zhu, L. Zhang, X. Wen, X. Huang, Feasibility of applying forward osmosis to the
- simultaneous thickening, digestion, and direct dewatering of waste activated sludge, Bioresource
 Technology, 113 (2012) 207-213.
- 555 [18] K. Lutchmiah, E.R. Cornelissen, D.J. Harmsen, J.W. Post, K. Lampi, H. Ramaekers, L.C.
- 556 Rietveld, K. Roest, Water recovery from sewage using forward osmosis, Water Science &
- 557 Technology, 64 (2011) 1443-1449.
- 558 [19] T.Y. Cath, A.E. Childress, M. Elimelech, Forward osmosis: Principles, applications, and recent
- developments, Journal of Membrane Science, 281 (2006) 70-87.
- 560 [20] A. Achilli, T.Y. Cath, A.E. Childress, Selection of inorganic-based draw solutions for forward
- osmosis applications, Journal of Membrane Science, 364 (2010) 233-241.

- 562 [21] J. Ren, J.R. McCutcheon, A new commercial thin film composite membrane for forward osmosis,
- 563 Desalination, 343 (2014) 187-193.
- [22] M. De Graaff, K. Roest, Inventarisatie van AB-systemen optimale procescondities in de A-trap,
 Nieuwegein, KWR, (2012).
- 566 [23] F.A. Meerburg, N. Boon, T. Van Winckel, J.A.R. Vercamer, I. Nopens, S.E. Vlaeminck, Toward
- 567 energy-neutral wastewater treatment: A high-rate contact stabilization process to maximally recover
- sewage organics, Bioresource Technology, 179 (2015) 373-381.
- 569 [24] Metcalf&Eddy, Wastewater Engineering: Treatment and Reuse. 4th edition, McGraw-Hill, New570 York, 2003.
- 571 [25] L. Chen, Y. Gu, C. Cao, J. Zhang, J.-W. Ng, C. Tang, Performance of a submerged anaerobic
- 572 membrane bioreactor with forward osmosis membrane for low-strength wastewater treatment, Water
 573 Research, 50 (2014) 114-123.
- 574 [26] J. De Vrieze, M. Coma, M. Debeuckelaere, P. Van der Meeren, K. Rabaey, High salinity in
- 575 molasses wastewaters shifts anaerobic digestion to carboxylate production, Water Research, 98 (2016)
- **576** 293-301.
- 577 [27] J.E. Schmidt, B.K. Ahring, Effects of magnesium on thermophilic acetate-degrading granules in
- upflow anaerobic sludge blanket (UASB) reactors, Enzyme and Microbial Technology, 15 (1993) 304310.
- 580 [28] Y. Chen, J.J. Cheng, K.S. Creamer, Inhibition of anaerobic digestion process: A review,
- 581 Bioresource Technology, 99 (2008) 4044-4064.
- 582 [29] E. Doğan, G.N. Demirer, Volatile fatty acid production from organic fraction of municipal solid
- waste through anaerobic acidogenic digestion, Environmental Engineering Science, 26 (2009) 14431450.
- [30] I. Siegert, C. Banks, The effect of volatile fatty acid additions on the anaerobic digestion of
- cellulose and glucose in batch reactors, Process Biochemistry, 40 (2005) 3412-3418.
- 587 [31] J. De Vrieze, T. Hennebel, N. Boon, W. Verstraete, Methanosarcina: The rediscovered
- 588 methanogen for heavy duty biomethanation, Bioresource Technology, 112 (2012) 1-9.
- [32] S. Van Nevel, S. Koetzsch, H.-U. Weilenmann, N. Boon, F. Hammes, Routine bacterial analysis
 with automated flow cytometry, Journal of Microbiological Methods, 94 (2013) 73-76.
- 591 [33] G. Blandin, A.R.D. Verliefde, J. Comas, I. Rodriguez-Roda, P. Le-Clech, Efficiently Combining
- 592 Water Reuse and Desalination through Forward Osmosis—Reverse Osmosis (FO-RO) Hybrids: A
- 593 Critical Review, Membranes.
- [34] J. De Vrieze, M. Coma, M. Debeuckelaere, P. Van der Meeren, K. Rabaey, High salinity in
- 595 molasses wastewaters shifts anaerobic digestion to carboxylate production, Water Research,
- 596 (Submitted).

- 597 [35] H. Carrère, C. Dumas, A. Battimelli, D.J. Batstone, J.P. Delgenès, J.P. Steyer, I. Ferrer,
- 598 Pretreatment methods to improve sludge anaerobic degradability: A review, Journal of Hazardous
- 599 Materials, 183 (2010) 1-15.
- 600 [36] D.C. Devlin, S.R.R. Esteves, R.M. Dinsdale, A.J. Guwy, The effect of acid pretreatment on the
- anaerobic digestion and dewatering of waste activated sludge, Bioresource Technology, 102 (2011)4076-4082.
- 603 [37] H. Yuan, Y. Chen, H. Zhang, S. Jiang, Q. Zhou, G. Gu, Improved Bioproduction of Short-Chain
- 604 Fatty Acids (SCFAs) from Excess Sludge under Alkaline Conditions, Environmental Science &
- 605 Technology, 40 (2006) 2025-2029.
- 606 [38] Y. Chen, S. Jiang, H. Yuan, Q. Zhou, G. Gu, Hydrolysis and acidification of waste activated
- sludge at different pHs, Water Research, 41 (2007) 683-689.
- 608 [39] C. Mengmeng, C. Hong, Z. Qingliang, S.N. Shirley, R. Jie, Optimal production of
- 609 polyhydroxyalkanoates (PHA) in activated sludge fed by volatile fatty acids (VFAs) generated from
- alkaline excess sludge fermentation, Bioresource Technology, 100 (2009) 1399-1405.

611

612

Table 1. Characteristics of the streams. Analysis were carried out in triplicate.

	Domestic wastewater	Secondary sludge	A-sludge
pH	7.42	7.26	7.15
Conductivity (mS cm ⁻¹)	1.4	1.3	1.8
Total solids, TS (g L ⁻¹)	0.26 ± 0.04	4.89 ± 0.83	6.73 ± 0.59
Volatile solids, VS (g L ⁻¹)	0.10 ± 0.03	2.83 ± 0.48	5.06 ± 0.62
Volatile solids, VS/TS (%)	38	58	78
Total suspended solids, TSS (g L ⁻¹)	<0.01	4.01 ± 0.04	5.43 ± 0.52
Volatile suspended solids, VSS (g L ⁻¹)	<0.01	2.51 ± 0.05	4.22 ± 0.48
Volatile suspended solids, VSS/TSS (%)	-	63	78
Kjeldahl nitrogen, TKN (mg N L ⁻¹)	9 ± 0	974 ± 15	476 ± 40
Total ammonia nitrogen, TAN (mg N L ⁻¹)	7 ± 5	56 ± 7	577 ± 20
Total chemical oxygen demand, tCOD (g L ⁻¹)	0.132 ± 0.016	4.482 ± 0.979	8.398 ± 0.362
Soluble chemical oxygen demand, sCOD (g L ⁻¹)	0.033 ± 0.003	0.215 ± 0.007	1.252 ± 0.023
Soluble chemical oxygen demand, sCOD/tCOD (%)	25	5	15
Fe (mg L ⁻¹)	6 ± 0	141 ± 1	271 ± 7
P (mg L ⁻¹)	463 ± 38	223 ± 1	200 ± 4
S (mg L ⁻¹)	3111 ± 189	85 ± 9	56 ± 5
K (mg L ⁻¹)	437 ± 29	47 ± 0	40 ± 2
Na (mg L ⁻¹)	45 ± 2	85 ± 3	54 ± 2
Ca (mg L ⁻¹)	387 ± 3	201 ± 7	233 ± 4
Mg (mg L ⁻¹)	1658 ± 242	37 ± 0	41 ± 1

Table 2: Operational parameters changes during FO-concentration (FO) and anaerobic FO-concentration (AnaFO). t⁰ stands for initial values.

	Time FO		Volume/concentration		COD concentration		Conductivity			Mg ²⁺ concentration	
	(hrs)		ratio		ratio		(mS cm ⁻¹)			(g L ⁻¹)	
	FO	AnaFO	FO	AnaFO	FO	AnaFO	t ⁰	FO	AnaFO	FO	AnaFO
Domestic wastewater	28.1	-	10	-	8.2 ± 1.2	-	1.4	31.6	-	20.5 ± 0.6	-
Secondary sludge	29.7	-	10	-	10.1 ± 2.4	-	1.3	18.2	-	11.7 ± 0.2	-
A-sludge	37.3	95.4	10	4	4.8 ± 0.2	3.9 ± 0.1	1.8	29.6	41.7	7.1 ± 0.1	8.7 ± 0.3

Table 3: Total and soluble COD and its ratio (COD soluble/COD total) for inputs and outputs of the concentration tests. Acronyms stand for raw A-sludge (AS), secondary sludge (WAS), domestic wastewater (WW), FO-concentrated streams (FO-AS, FO-WAS, FO-WW), anaerobically FO-concentrated A-sludge (AnaFO-AS), gravity concentrated A-sludge (GC-AS), addition of 7.1 g L⁻¹ of MgCl₂ (ASM/ GC-ASM), aeration at 0.6 L min⁻¹ (ASA/ GC-ASA), a combination of salt addition and aeration (ASAM/ GC-ASAM) and the wastewater discharged after A-sludge gravity concentration (GC-WW). * estimated soluble COD according to typical COD ratios for A-sludge.

		COD total	COD soluble	COD ratio
		(g L ⁻¹)	(g L ⁻¹)	(%)
	AS	8.40 ± 0.36	1.25 ± 0.02	14.9
is.	FO-AS	40.49 ± 1.09	12.30 ± 0.70	30.4
son	AS	5.46 ± 0.53	0.81 *	14.9 *
Osr test	AnaFO-AS	21.40 ± 1.65	11.23 ± 0.23	52.5
(O)	WAS	4.48 ± 0.98	0.22 ± 0.01	4.8
rwa (F	FO-WAS	48.53 ± 0.27	4.92 ± 0.50	10.1
Fo	WW	0.13 ± 0.02	0.03 ± 0.00	25.2
	FO-WW	1.08 ± 0.08	0.24 ± 0.01	21.7
	AS	7.76 ± 0.16	1.47 ± 0.04	19.0
ests	ASML	7.65 ± 0.16	1.91 ± 0.00	25.0
C) t	ASMH	7.78 ± 0.01	2.14 ± 0.07	27.5
<u>(</u>	ASA	6.84 ± 0.31	1.12 ± 0.03	16.4
tion	ASAML	6.50 ± 0.20	1.24 ± 0.02	19.0
ltra	ASAMH	7.05 ± 1.10	1.28 ± 0.06	18.1
cen	GC-AS	29.42 ± 0.19	3.84 ± 0.01	13.1
con	GC-ASML	29.14 ± 0.67	4.69 ± 0.01	16.1
ity	GC-ASMH	28.14 ± 0.01	4.01 ± 0.01	14.2
rav	GC-ASA	28.12 ± 0.15	3.04 ± 0.04	10.8
6	GC-ASAML	29.25 ± 0.49	3.68 ± 0.08	12.6
	GC-ASAMH	28.18 ± 0.72	2.93 ± 0.09	10.4
	GC-WW	3.24 ± 0.04	0.86 ± 0.01	26.7

Table 4: Cost analysis for the Forward Osmosis (FO) concentration and of A-sludge (average concentration of 7.5 g COD L⁻¹) by using either 2.2 M MgCl₂ or sea-water (NaCl 0.5 M) as draw solution. For the former Reverse Osmosis (RO) was considered for the reconcentration of the DS and relative costs were included. Profit gained for the VFA production of either FO-concentrated or A-sludge concentrated by thickening are calculated considering a conversion efficiency (mg COD-VFA g⁻¹ COD_{fed}) of 45 % for the FO-concentrated A-sludge and of 20 % for the thickened A-sludge. Costs and profits are reported as ratio of \in for m³ or kg of raw A-sludge (CAPEX capital expenses, OPEX operational expenses, SEC Specific Energy Consumption). Total profits (balance) are calculated as difference between the net for the VFA for the technology and the total costs (OPEX+CAPEX) of that technology.

		Forward Osmosis						Thickening
		2.2 M MgCl ₂			Sea-water (0.5 M NaCl)			
		FO	RO	Total	FO	RO	Total	
CAPEX	€ m ⁻³	0.53	0.26	0.79	1.06	-	1.06	0.08
OPEX								
SEC	€ m ⁻³	-	0.67	0.67	-	-	-	-
PE + Electricity	€ m ⁻³	-	-	-	-	-	-	0.06
	03	0.53	0.02	1.46	1.07		1.07	0.14
Total costs	€ m ^{-s}	0.53	0.93	1.40	1.00	-	1.06	0.14
(OPEX+CAPEX)	€ kg ⁻¹	0.07	0.12	0.19	0.14	-	0.14	0.02
	€ m ⁻³		2.36			2.36		1.05
VFA net	€ kg ⁻¹		0.32			0.32		0.14
	€ m ⁻³		0.90			1.30		0.91
Balance	€ kg ⁻¹		0.12			0.17		0.12



Figure 1. FO dewatering flux in $L m^2 h^{-1}$ for the domestic wastewater, secondary sludge or A-sludge (feed solution) when in contact with a solution of MgCl₂ 2.2 M (draw solution) through a semipermeable TFC membrane. The streams were concentrated/dewatered with a factor 10.



Figure 2. Specific VFA production for raw and FO-concentrated streams on their peak-production day. The test was carried out at 35 °C and pH 7. (n = 3).



Figure 3. VFA composition in mg COD $g^{-1}COD_{fed}$ on the peak production day. The test was carried out at 35 °C and pH 7 (n = 3).



Figure 4. Specific VFA production on the peak-production day for A-sludge and gravity concentrated A-sludge (GC-A-sludge) with different treatments and A-sludge concentrated aerobically or anaerobically with forward osmosis (FO-A-sludge). The fermentation was performed at 35 $^{\circ}$ C and pH 7 (n = 3).



Figure 5. A. total intact cells/ permiabilized cells ratio (%) (n = 3) and B. total intact cells concentration for A-sludge and gravity concentrated A-sludge (GC-A-sludge) with different treatments and A-sludge concentrated aerobically or anaerobically with forward osmosis (FO-A-sludge) (n = 3).