



Citation for published version:

Johannsen, NM, Sparks, LM, Zhang, Z, Earnest, C, Smith, SR, Church, TS & Ravussin, E 2013, 'Determinants of the changes in glycaemic control with exercise training in type 2 diabetes: A randomized trial', PLoS ONE, vol. 8, no. 6, e62973. <https://doi.org/10.1371/journal.pone.0062973>

DOI:

[10.1371/journal.pone.0062973](https://doi.org/10.1371/journal.pone.0062973)

Publication date:

2013

Document Version

Publisher's PDF, also known as Version of record

[Link to publication](#)

Publisher Rights

CC BY

© 2013 Johannsen et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

University of Bath

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Determinants of the Changes in Glycemic Control with Exercise Training in Type 2 Diabetes: A Randomized Trial

Neil M. Johannsen^{1,2,3}, Lauren M. Sparks^{3,9}, Zhengyu Zhang¹, Conrad P. Earnest⁴, Steven R. Smith⁵, Timothy S. Church¹, Eric Ravussin^{1*}

1 Pennington Biomedical Research Center, Baton Rouge, Louisiana, United States of America, **2** Louisiana State University, Baton Rouge, Louisiana, United States of America, **3** Translational Research Institute for Metabolism and Diabetes, Orlando, Florida, United States of America, **4** University of Bath, Department of Health, Bath, United Kingdom, **5** Sanford-Burnham Medical Research Institute, Orlando, Florida, United States of America

Abstract

Aims: To assess the determinants of exercise training-induced improvements in glucose control (HbA_{1C}) including changes in serum total adiponectin and FFA concentrations, and skeletal muscle peroxisome proliferator-activated receptor- γ coactivator-1 α (PGC-1 α) protein content.

Methods: A sub-cohort (n = 35; 48% men; 74% Caucasian) from the HART-D study undertaking muscle biopsies before and after 9 months of aerobic (AT), resistance (RT), or combination training (ATRTR).

Results: Changes in HbA_{1C} were associated with changes in adiponectin (r = -0.45, P = 0.007). Participants diagnosed with type 2 diabetes for a longer duration had the largest increase in PGC-1 α (r = 0.44, P = 0.008). Statistical modeling examining changes in HbA_{1C} suggested that male sex (P = 0.05), non-Caucasian ethnicity (P = 0.02), duration of type 2 diabetes (r = 0.40; P < 0.002) and changes in FFA (r = 0.36; P < 0.004), adiponectin (r = -0.26; P < 0.03), and PGC-1 α (r = -0.28; P = 0.02) explain ~65% of the variability in the changes in HbA_{1C}.

Conclusions: Decreases in HbA_{1C} after 9 months of exercise were associated with shorter duration of diabetes, lowering of serum FFA concentrations, increasing serum adiponectin concentrations and increasing skeletal muscle PGC-1 α protein expression.

Trial Registration: ClinicalTrials.gov NCT00458133

Citation: Johannsen NM, Sparks LM, Zhang Z, Earnest CP, Smith SR, et al. (2013) Determinants of the Changes in Glycemic Control with Exercise Training in Type 2 Diabetes: A Randomized Trial. PLoS ONE 8(6): e62973. doi:10.1371/journal.pone.0062973

Editor: Hugo ten Cate, Maastricht University Medical Center, The Netherlands

Received: September 18, 2012; **Accepted:** March 24, 2013; **Published:** June 21, 2013

Copyright: © 2013 Johannsen et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was partially funded by an institutional grant to ER and by a NORC Center Grant (#2P30DK072476; PI-ER). Cost of analyses was partially supported by an unrestricted research funding grant from The Coca-Cola Company. Further funding was contributed by Dr. Donna Ryan from the Pennington Biomedical Research Center Clinical and Translational Research Fund. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: A commercial funding source is acknowledged in the manuscript: The Coca-Cola Company. The authors have no declarations for this funding source relating to employment, consultancy, patents, products in development or marketed products and the funding source does not alter their adherence to the PLOS ONE policies on sharing data and materials, as detailed in the online guide for authors.

* E-mail: eric.ravussin@pbrcc.edu

⁹ These authors contributed equally to this work.

Introduction

Type 2 diabetes is as much a disease of disordered lipid metabolism as a disease of abnormal glucose metabolism [1]. Failure to balance skeletal muscle lipid uptake (free fatty acids, FFA) and storage in intracellular triacylglycerol with oxidation in mitochondria is implicated in impaired insulin action [2]. Individuals with type 2 diabetes have a reduced plasma adiponectin concentrations [3], reduced number of mitochondria [4], and lower skeletal muscle gene/protein expressions of peroxisome proliferator-activated receptor- γ coactivator-1 α (PGC-1 α), a key regulator of mitochondrial biogenesis [5] and oxidative metabolism [6]. While increased PGC-1 α has been observed with acute exercise in humans [7], studies examining the effects of chronic exercise training on adiponectin and PGC-1 α

and their relationship to the change in glycemic control (HbA_{1C}) in individuals with type 2 diabetes are absent.

We recently demonstrated in the Health Benefits of Aerobic and Resistance Training in individuals with type 2 Diabetes (HART-D) study that 9 months of combined aerobic (AT) and resistance (RT) training significantly reduced HbA_{1C} levels in individuals with type 2 diabetes [8]. The HART-D study also included AT only and RT only training groups that, together with the combined AT and RT training group, provide a unique opportunity to examine the influence of factors known to influence glycemic control, specifically serum FFA and adiponectin, skeletal muscle PGC-1 α protein content, and anthropometric/demographic measures. We therefore hypothesized that the change in HbA_{1C} after exercise would be independently associated with serum FFA and adiponectin and the activation of skeletal muscle

mitochondrial biogenesis pathways as determined by PGC-1 α protein content.

Methods

Seventy-eight participants from the previously reported HART-D study volunteered for and were enrolled into an ancillary study that included the collection of muscle samples at baseline and after 9 months of intervention (Figure 1). Detailed methods as well as inclusion/exclusion criteria are provided in the main outcomes paper [8]. Briefly, 262 sedentary individuals with type 2 diabetes (HbA_{1C} 6.5% to 11%, inclusive) aged 30 to 75 years were enrolled into the study. Volunteers were excluded if body mass index was ≥ 48 kg/m², blood pressure was $\geq 160/100$ mmHg, fasting triglycerides were ≥ 500 mg/dL, urine protein was >100 mg/dL, or serum creatinine was >1.5 mg/dL. In addition, volunteers were excluded if they used an insulin pump or had a history of stroke, advanced neuropathy or retinopathy, or any other serious medical condition that prevented them from adhering to the protocol or exercising safely. Of the original 78 participants, 23 were lost to follow-up due to non-compliance ($<80\%$ attendance) with the study protocol leaving a total of 55 participants with both baseline and follow-up data. An additional 10 participants randomized to the control group were excluded from these analyses. An additional 10 participants' samples did not have protein extracted. The final group of 35 participants (17 males and 18 females) aged 57.0 ± 7.7 y are included in the present analysis. The follow-up bloods draw and muscle biopsy for this ancillary project occurred approximately 1 week after their last exercise bout. The study was approved by the Pennington Biomedical Research Center institutional review board and written informed consent was obtained prior to study screening. The protocol for this trial and supporting CONSORT checklist are available as supporting information; see Checklist S1 and Protocol S1.

Intervention

HART-D participants were randomized to 9 months of aerobic training (AT), resistance training (RT), a combination of both (ATRT), or a non-exercise control group. The AT group exercised

3–4 times per week at 50–80% of VO_{2peak} for a total energy expenditure of 12 kcal·kg⁻¹·wk⁻¹. The RT group completed 3 days per week of 2 sets of 4 arm exercises, 3 sets of 3 leg exercises, and 2 sets of back extension and abdominal crunch. The ATRT group completed 3 sessions of AT (10 kcal·kg⁻¹·wk⁻¹) and 2 sessions of RT (1 set each of the RT exercises) per week in conjunction with the Federal Physical Activity Guidelines [9]. Both exercise modalities were progressive in nature whereby the treadmill speed and grade was increased to maintain the appropriate intensity, reported in metabolic equivalent tasks (METs) [10] and the weight lifted on each RT exercise was increased when a participant could lift 12 repetitions on 2 consecutive sessions.

VO_{2max} and body composition

Maximal cardiorespiratory fitness (VO_{2max}) was assessed at baseline and after 9 months of intervention on a treadmill (Trackmaster 425, Newton, KS) with respiratory gases analyzed using a True Max 2400 Metabolic Cart (Parvomedics, Salt Lake City, UT). Peak oxygen uptake (VO_{2peak}) was expressed relative to fat-free mass (mL·kg FFM⁻¹·min⁻¹). Body composition was measured by DXA (QDR 4500A, Hologic, Inc. Waltham, MA).

Blood analyses

Blood was collected by venipuncture in the morning after a 10-hour fast at baseline and after the intervention. Blood samples were placed into pre-chilled red-top tubes (serum FFA and adiponectin) or EDTA collection tubes (plasma HbA_{1C}) and transported to the laboratory on ice (FFA and HbA_{1C}). The blood samples for FFA and HbA_{1C} were cold centrifuged and the media was immediately analyzed for FFA and HbA_{1C} (Beckman Coulter DXC600 Pro; Beckman Coulter Inc., Brea, CA). Blood collected for total serum adiponectin (Linco, St. Charles, MO) concentrations were transferred to the laboratory at room temperature, centrifuged, and serum was stored at -80°C until run in a single batch at the end of the trial. The intra-assay variation for HbA_{1C}, FFA, and adiponectin was $\pm 2.84\%$, $\pm 0.75\%$, and $\pm 6.21\%$, respectively.

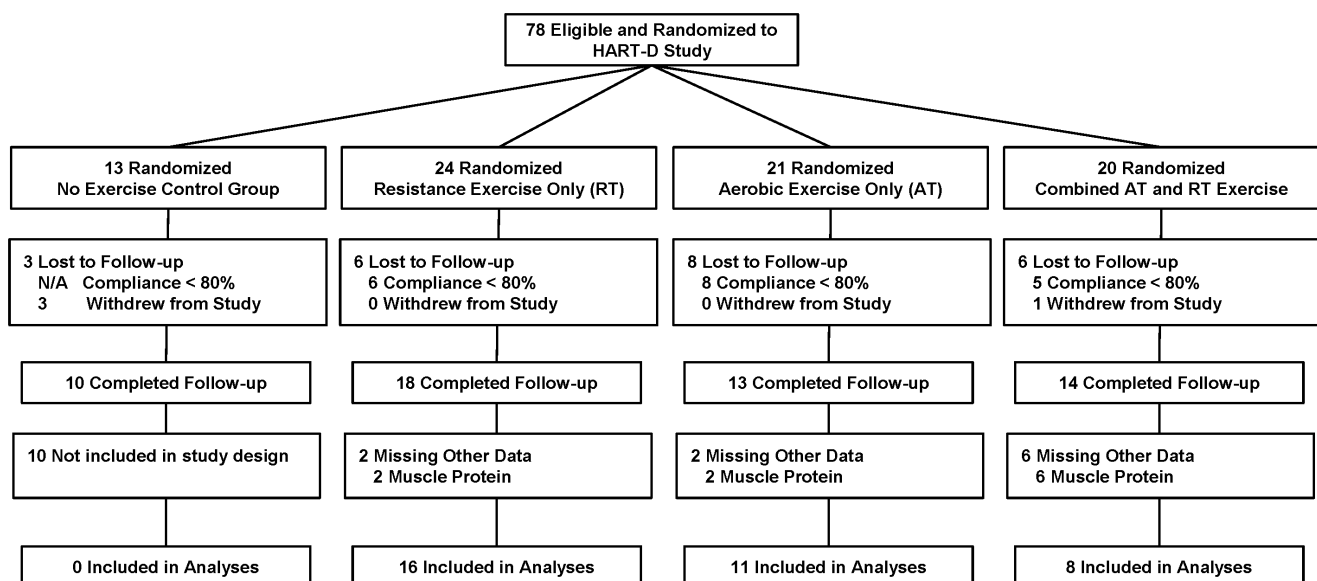


Figure 1. Consort diagram.

doi:10.1371/journal.pone.0062973.g001

Muscle analysis

Muscle samples were obtained under local anaesthesia from the *vastus lateralis* using the Bergstrom technique and immediately snap frozen. Samples were homogenized by Kontes Duall tissue grinders in RIPA buffer with a cocktail of protease and phosphatase inhibitors (Sigma, St. Louis, MO). Total protein content was quantified by BCA assay (Pierce, Rockford, IL) and 25 μ L was added to each gel for determination of PGC-1 α protein content. PGC-1 α protein content quantified relative to GAPDH (Abcam, Cambridge, MA) was determined by Western blot using an Odyssey 9120 Imaging System (LI-COR, Lincoln, NE) after probing with goat anti-mouse IgG Alexa Fluor 680 (Invitrogen, Carlsbad, CA). No data are available for the intra-assay variability in PGC-1 α or GAPDH protein content as replicates were not run to preserve the remaining muscle tissue.

Randomization and blinding

After signing the informed consent for the ancillary study, volunteers had baseline blood drawn and a muscle biopsy performed. Volunteers were later randomized to an intervention group according to the randomization schema within the main HART-D trial resulting in unequal group randomization. The nature of this intervention study prevents blinding of the exercise intervention personnel. However, every effort was made to maintain blinding of the assessment staff from the participant's intervention group.

Statistical analysis

Analysis of variance (ANOVA) was used to assess baseline group differences (JMP 9.0.2, SAS Institute, Inc., Cary, NC) and dichotomous variables were examined using a χ^2 test. All exercise groups were collapsed into a single group for the remaining analyses since no effect of exercise group was observed in the final model ($P=0.15$), and the addition of exercise group to the final model did not influence the other factors in the model. Linear regression (Pearson r) was used to determine the relationships among change (post-pre) scores. Baseline-adjusted changes in anthropometric data, HbA_{1C}, PGC-1 α protein content, and fasting adiponectin and free-fatty acids were determined using analysis of covariance (ANCOVA) with group differences analyzed using Student-t post-hoc tests. In addition, ANCOVA was used to examine the relationships between changes in HbA_{1C} and changes in independent determinants (semi-partial correlation coefficients (r)). To determine the relationship between changes in HbA_{1C} and changes in FFA, adiponectin, and PGC1 α content, we first adjusted for baseline HbA_{1C}, sex, ethnicity, and duration of diabetes; all variables that were adjusted for in the main outcomes paper [8]. Furthermore, we aimed to determine whether weight, body composition and/or fitness (VO_{2peak}) further explained the change in HbA_{1C} or removed any of the existing parameters from the model. Statistical significance was set at $P<0.05$ and data are reported as mean (95%CI) unless noted otherwise.

Results

The average compliance for all exercise groups was $95.0\pm 6.0\%$ (mean \pm SD; range = 80.6% to 100.0%). From Month 2 to Month 9, estimated average METS in AT (AT = 5.7 ± 1.0 to 7.2 ± 1.0 METS) and ATRT = 5.0 ± 0.7 to 5.8 ± 1.0 METS) and total weight lifted during RT (RT = $56,422\pm 17,316$ to $73,602\pm 25,631$ lbs) and ATRT = 17881 ± 6345 to $23,909\pm 9,532$ lbs) increased similarly to those reported in the main outcomes paper [8].

Baseline data for age, sex, diabetes duration, HbA_{1C} body composition, and VO_{2peak} (ml/kg FFM) in this subset of HART-D

participants were similar to the main study for participants with an overall compliance $\geq 80\%$ (all $P>0.18$). However, the percent of Caucasian participants was higher in this subset compared to the main study (74.3% vs. 56.7%, respectively, $P=0.045$). Baseline data and baseline-adjusted treatment effects are presented in Table 1. Age was different across groups at baseline ($P=0.02$). After training, change in VO_{2peak} was not significantly different between intervention groups in this cohort. Body weight was lower after AT compared with RT ($P<0.05$) due to a tendency for RT to increase FFM ($P=0.05$). The baseline-adjusted change in HbA_{1C} in this cohort was independent of treatment group ($P=0.29$). The change in HbA_{1C} adjusted for baseline HbA_{1C}, age, ethnicity and type 2 diabetes duration, were similar to those in the larger cohort [8] albeit, not significant in this subset (exercise group effect, $P=0.60$; -0.15% (-0.58% , 0.27%), -0.34% (-0.84% , 0.15%), and -0.49% (-1.03% , 0.05%) for RT, AT, and ATRT, respectively). PGC-1 α response tended to differ by intervention group ($P=0.08$; Table 1).

Change in fasting serum adiponectin was inversely associated with the change in HbA_{1C} ($r=-0.45$; $P=0.007$). Change in HbA_{1C} was not related to change in fasting serum FFA ($r=0.25$; $P=0.15$) or change in PGC-1 α protein content ($r=0.05$; $P=0.80$). Participants with longer durations of type 2 diabetes had the largest increase in PGC-1 α ($r=0.44$; $P=0.008$). Last, change in VO_{2peak} was inversely related with age ($r=-0.36$; $P<0.04$). No other significant relationships exist.

In the process of building our statistical model, we examined the effect of change in serum FFA and adiponectin, and PGC-1 α protein content, separately, on changes in HbA_{1C} after adjusting for baseline HbA_{1C}, sex, and ethnic group. The changes in serum FFA ($r=0.33$; $P<0.02$) and adiponectin ($r=-0.30$; $P<0.03$) were associated with the changes in HbA_{1C}; however, change in PGC-1 α protein content did not reach statistical significance ($r=0.17$; $P=0.24$).

After adjusting for baseline HbA_{1C}, changes in HbA_{1C} were related to type 2 diabetes duration ($r=0.40$, $P<0.002$; Figure 2A), changes in fasting serum FFA ($r=0.36$, $P<0.004$; Figure 2B) and, adiponectin ($r=-0.26$, $P<0.03$; Figure 2C) levels, and skeletal muscle PGC-1 α protein ($r=-0.28$, $P=0.02$; Figure 2D) independent of sex (-0.09% (-0.36% , 0.17%) vs. -0.55% (-0.92% , -0.17%); women vs. men, respectively; $P=0.05$) and ethnicity (-0.00% (-0.23% , 0.23%) vs. -0.64% (-1.09% , -0.19%), Caucasian vs. non-Caucasian, respectively; $P=0.02$). Together, these factors explained $\sim 65\%$ of the variance in the change in HbA_{1C}. The regression equation for the model is presented below. Changes in fitness (VO_{2peak}) and body composition did not influence this model.

$$\begin{aligned} \Delta HbA_{1C} = & -0.48(\text{baseline HbA}_{1C}) - 0.23(\text{sex}) \\ & + 0.32(\text{ethnic group}) + 0.07(\text{type 2 diabetes duration}) \\ & + 1.54(\Delta FFA) - 0.06(\Delta \text{adiponectin}) \\ & - 9.29(\Delta PGC - 1\alpha) + 2.52 \end{aligned}$$

Conclusions

The novel finding of the present investigation was that the changes in HbA_{1C} after 9 months of exercise were independently associated with the duration of type 2 diabetes and changes in serum FFA and negatively associated with changes in serum adiponectin and skeletal muscle PGC-1 α .

Table 1. Baseline participant characteristics and baseline adjusted changes in anthropometric data, HbA_{1C}, PGC-1 α protein content, and fasting adiponectin and free-fatty acids after 9 months of exercise.

	RT		AT		ATRT		P-value	
	Baseline	Change	Baseline	Change	Baseline	Change	Baseline	Change
n	16		11		8			
Age, y	60.9 (7.6)		53.8 (6.6)		53.6 (6.4)		0.02 ^a	
Sex, male %	8 (50.0)		6 (54.6)		3 (37.5)		0.75	
Ethnicity, Caucasian %	12 (75.0)		9 (81.8)		5 (62.5)		0.64	
Type 2 Diabetes Duration, y	9.5 (6.9)		6.0 (4.8)		5.9 (3.4)		0.20	
Body Weight, kg	98.5 (15.1)	0.5 (−0.7, 1.6)	90.5 (11.0)	−1.6 (−3.0, −0.2)	100.5 (22.3)	−1.3 (−2.9, 0.3)	0.32	<0.05 ^a
Fat Mass, %	37.4 (8.5)	−0.9 (−1.8, 0.0)	31.7 (8.8)	−0.3 (−1.4, 0.8)	38.4 (6.6)	−0.9 (−2.1, 0.4)	0.14	0.67
FFM, kg	61.6 (11.5)	1.3 (0.4, 2.2)	61.6 (9.3)	−0.3 (−1.4, 0.8)	62.3 (15.6)	−0.1 (−1.4, 1.2)	0.99	0.05
VO _{2peak} , mL·kg FFM ^{−1} ·min ^{−1}	31.3 (4.6)	−0.1 (−2.1, 1.9)	34.7 (6.6)	2.0 (−0.5, 4.5)	30.3 (3.3)	0.6 (−2.2, 3.5)	0.13	0.43
HbA _{1C} , %	7.1 (1.1)	0.0 (−0.3, 0.4)	7.4 (1.4)	−0.2 (−0.7, 0.2)	6.9 (0.7)	−0.4 (−0.9, 0.1)	0.61	0.29
FFA, mmol/L	0.56 (0.21)	−0.00 (−0.11, 0.11)	0.66 (0.22)	0.04 (−0.09, 0.17)	0.54 (0.24)	−0.07 (−0.22, 0.08)	0.47	0.56
Adiponectin, μ g/mL	8.1 (4.2)	−0.9 (−2.9, 1.0)	6.7 (2.9)	0.3 (−2.0, 2.6)	6.4 (3.9)	−1.8 (−4.5, 0.9)	0.47	0.47
PGC-1 α /GAPDH, au	0.04 (0.04)	0.02 (0.00, 0.03)	0.05 (0.04)	0.00 (−0.01, 0.02)	0.06 (0.04)	−0.01 (−0.03, 0.01)	0.53	0.08

All baseline data are means (SD), n (%), and baseline-adjusted change scores are mean (95%CI). Abbreviations: RT, resistance training; AT, aerobic training; ATRT, combination of aerobic and resistance training; VO₂ volume of oxygen consumed in mL per kg fat-free mass (FFM) per minute; HbA_{1C}, glycated hemoglobin A_{1C}; PGC-1 α /GAPDH, peroxisome proliferator-activated receptor- γ coactivator-1 α adjusted for glyceraldehyde 3-phosphate dehydrogenase protein content; FFA, free fatty acid. ^aRT group significantly greater than greater than AT. Blank cells are not applicable.
doi:10.1371/journal.pone.0062973.t001

The change in free-fatty acid concentration was a major determinant of the change in glycemic control. Free-fatty acid accumulation in the circulation may be the result of an imbalance between skeletal muscle uptake and oxidation and defects in adipose tissue insulin signaling [2]. Lipid oversupply to skeletal muscle may cause insulin resistance through accumulation of incompletely oxidized lipid species [11]. In the present investigation, improved balance between muscle lipid storage and oxidation (less lipid species storage), as suggested by a decrease in serum FFA, may be an important contributor to the improvement in HbA_{1C}.

We observed a strong inverse relationship between the changes in adiponectin and the changes in HbA_{1C}. Civitarese et al. recently uncovered a pathway by which adiponectin increases skeletal muscle PGC-1 α , mitochondrial number and oxidative capacity [12]. Adiponectin may also have beneficial effects, independent of PGC-1 α , by enhancing ceramidase activity, thus reducing the amount of insulin desensitizing ceramides [13]. Our results demonstrate a clinically relevant situation in which the change in serum adiponectin was associated with an improvement in HbA_{1C}.

A recent review called for more effective and sophisticated exercise prescriptions for the improvement of glucose control through increasing intramuscular PGC-1 α [14]. Studies demonstrate increased PGC-1 α with aerobic training [7], while others show no effect or improvements with resistance training [15,16]. We tested different exercise modalities and only resistance training had a tendency to increase PGC-1 α content. Additionally, the change in PGC-1 α was greater in those with longer duration of type 2 diabetes. In our final model, a greater increase in PGC-1 α and a shorter duration of type 2 diabetes were independently associated with improved glycemic control. The main results from

the HART-D trial together with this ancillary study suggest that: 1) a program of combined aerobic and resistance training has the greatest effect on HbA_{1C}; 2) resistance training may potentiate a greater change in PGC-1 α , and therefore, HbA_{1C}; and 3) individuals who start an exercise program soon after diagnosis may see a larger effect on HbA_{1C} levels.

Our investigation is limited by a small sample size with almost 50% of the participants assigned to the resistance training group. However, we had adequate power to detect independent effects of substrate (FFA), hormone (adiponectin) and muscle (PGC-1 α) changes on glycemic control after collapsing the exercise groups. Drug changes were not controlled during the intervention period resulting in a limitation whereby a tendency to reduce diabetes medications after the combination intervention reduces the effect on change in HbA_{1C} levels. Furthermore, dietary assessments were taken at baseline and follow-up by food frequency questionnaire thus limiting our ability to determine the impact of dietary modifications such as changes in caloric intake and dietary composition. Finally, research suggests that GAPDH gene transcription may be sensitive to treatments that may influence PGC-1 α protein content, including exercise-induced changes in insulin sensitivity [17]. However, analysis of the crude and GAPDH adjusted PGC-1 α data yielded similar results providing evidence that the effects noted in this manuscript were the result of the exercise intervention on PGC-1 α specifically and not secondary changes in GAPDH. The primary strengths are a well-controlled exercise training study with high adherence rates and a wide range of changes in the dependent and independent variables.

In summary, exercise in individuals with type 2 diabetes should be initiated soon after diagnosis and include training programs aimed at improving plasma substrate availability, endocrine

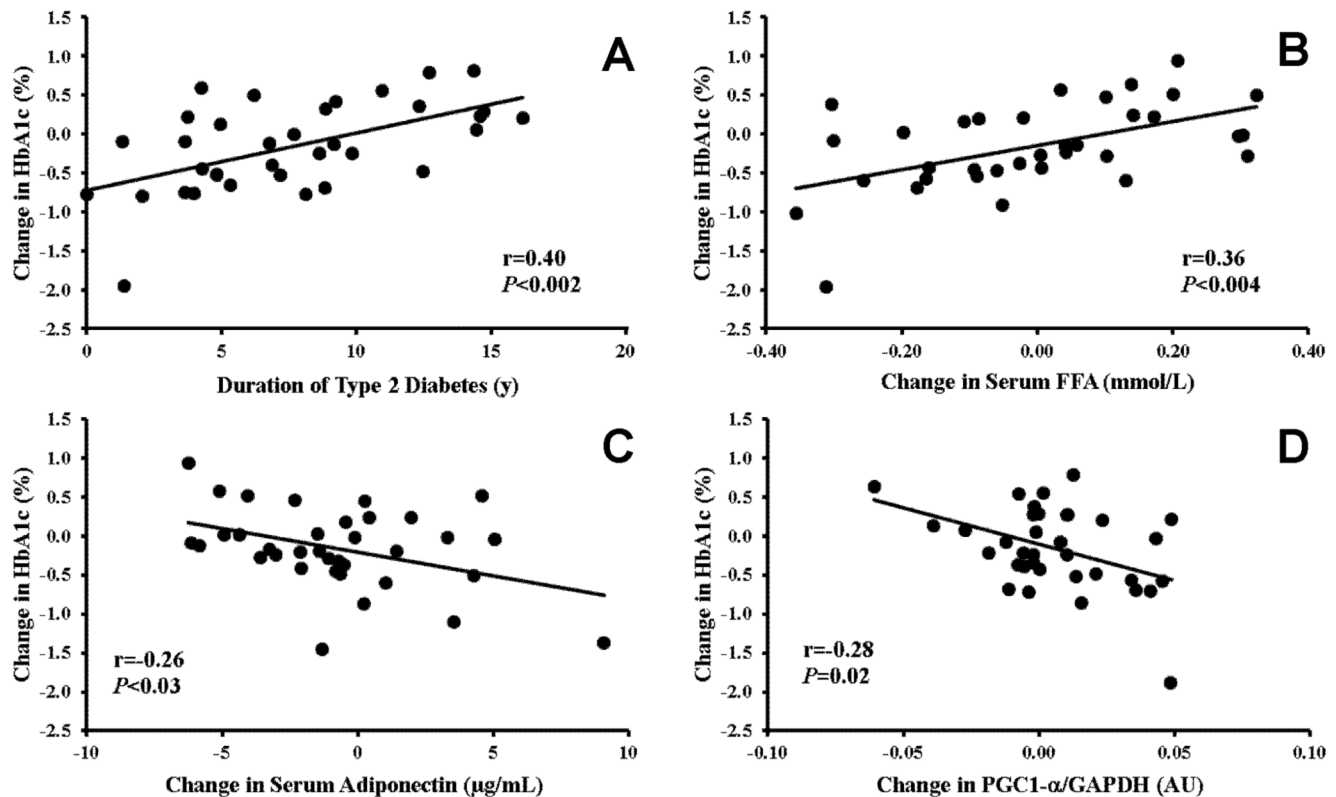


Figure 2. Independent effects (semipartial correlation coefficients (r)) of type 2 diabetes duration (A) and change in serum FFA (B), serum adiponectin (C), and intramuscular peroxisome proliferator-activated receptor- γ coactivator-1 α (PGC-1 α) protein content (D) on change in HbA_{1c} (%) after 9 months of exercise after adjusting for baseline HbA_{1c}, sex, and ethnicity. Change in HbA_{1c} is reported as residuals (A–D) from the regression model (Δ HbA_{1c} = $-.48(\text{baseline HbA}_{1c}) - 0.23(\text{sex}) + 0.32(\text{ethnic group}) + 0.07(\text{type 2 diabetes duration}) + 1.54(\Delta\text{FFA}) - 0.06(\Delta\text{adiponectin}) - 9.29(\Delta\text{PGC-1}\alpha) + 2.52$). Units for the prediction equation are similar to those in each panel; sex, 1 = men and 0 = women; ethnic group, 1 = Caucasian and 0 = Non-Caucasian. doi:10.1371/journal.pone.0062973.g002

function, and skeletal muscle factors shown to improve glycemic outcomes.

Supporting Information

Checklist S1 CONSORT Checklist.
(DOC)

Protocol S1 Trial Protocol.
(PDF)

Acknowledgments

The authors would like to acknowledge the volunteers from the ancillary HART-D study for their participation.

Author Contributions

Conceived and designed the experiments: CPE SRS ER. Performed the experiments: CPE ZZ. Analyzed the data: NMJ LMS ZZ. Contributed

References

- McGarry JD (1992) What if Minkowski had been ageusic? An alternative angle on diabetes. *Science* 258: 766–770.
- Ravussin E, Smith SR (2002) Increased fat intake, impaired fat oxidation, and failure of fat cell proliferation result in ectopic fat storage, insulin resistance, and type 2 diabetes mellitus. *Ann N Y Acad Sci* 967: 363–378.
- Hotta K, Funahashi T, Arita Y, Takahashi M, Matsuda M, et al. (2000) Plasma concentrations of a novel, adipose-specific protein, adiponectin, in type 2 diabetic patients. *Arterioscler Thromb Vasc Biol* 20: 1595–1599.
- Ritov VB, Menshikova EV, He J, Ferrell RE, Goodpaster BH, et al. (2005) Deficiency of subsarcolemmal mitochondria in obesity and type 2 diabetes. *Diabetes* 54: 8–14.
- Mootha VK, Lindgren CM, Eriksson KF, Subramanian A, Sihag S, et al. (2003) PGC-1 α -responsive genes involved in oxidative phosphorylation are coordinately downregulated in human diabetes. *Nat Genet* 34: 267–273.

6. Puigserver P, Spiegelman BM (2003) Peroxisome proliferator-activated receptor-gamma coactivator 1 alpha (PGC-1 alpha): transcriptional coactivator and metabolic regulator. *Endocr Rev* 24: 78–90.
7. Pilegaard H, Saltin B, Neufer PD (2003) Exercise induces transient transcriptional activation of the PGC-1alpha gene in human skeletal muscle. *J Physiol* 546: 851–858.
8. Church TS, Blair SN, Cocroham S, Johansen N, Johnson W, et al. (2010) Effects of aerobic and resistance training on hemoglobin A1c levels in patients with type 2 diabetes: a randomized controlled trial. *JAMA* 304: 2253–2262.
9. Services USDoHaH (2008) 2008 Physical Activity Guidelines for Americans. Rockville, MD: U.S. Department of Health and Human Services, Office of Disease Prevention and Health Promotion.
10. (2006) American College of Sports Medicine. ACSM's Guidelines for Exercise Testing and Prescription. Philadelphia, PA: Lippincott Williams & Williams.
11. Koves TR, Ussher JR, Noland RC, Slentz D, Mosedale M, et al. (2008) Mitochondrial overload and incomplete fatty acid oxidation contribute to skeletal muscle insulin resistance. *Cell Metab* 7: 45–56.
12. Civitarese AE, Ukropcova B, Carling S, Hulver M, DeFronzo RA, et al. (2006) Role of adiponectin in human skeletal muscle bioenergetics. *Cell Metab* 4: 75–87.
13. Holland WL, Miller RA, Wang ZV, Sun K, Barth BM, et al. (2011) Receptor-mediated activation of ceramidase activity initiates the pleiotropic actions of adiponectin. *Nat Med* 17: 55–63.
14. Benton CR, Wright DC, Bonen A (2008) PGC-1alpha-mediated regulation of gene expression and metabolism: implications for nutrition and exercise prescriptions. *Appl Physiol Nutr Metab* 33: 843–862.
15. Layne AS, Nasrallah S, South MA, Howell ME, McCurry MP, et al. (2011) Impaired muscle AMPK activation in the metabolic syndrome may attenuate improved insulin action after exercise training. *J Clin Endocrinol Metab* 96: 1815–1826.
16. Heilbronn LK, Gan SK, Turner N, Campbell LV, Chisholm DJ (2007) Markers of mitochondrial biogenesis and metabolism are lower in overweight and obese insulin-resistant subjects. *J Clin Endocrinol Metab* 92: 1467–1473.
17. Alexander-Bridges M, Dugast I, Ercolani L, Kong XF, Gierle L, et al. (1992) Multiple insulin-responsive elements regulate transcription of the GAPDH gene. *Adv Enzyme Regul* 32: 149–159.