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REVIEW ARTICLE

Breakthroughs in modern cancer therapy and elusive cardiotoxicity: Critical research-practice gaps, challenges, and insights

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Abstract

To date, five cancer treatment modalities have been defined. The three traditional modalities of cancer treatment are surgery, radiotherapy, and conventional chemotherapy, and the two modern modalities include molecularly targeted therapy (the fourth modality) and immunotherapy (the fifth modality). The cardiotoxicity associated with conventional chemotherapy and radiotherapy is well known. Similar adverse cardiac events are resurging with the fourth modality. Aside from the conventional and newer targeted agents, even the most newly developed, immune-based therapeutic modalities of anticancer treatment (the fifth modality), e.g., immune checkpoint inhibitors and chimeric antigen receptor (CAR) T-cell therapy, have unfortunately led to potentially lethal cardiotoxicity in patients. Cardiac complications represent unresolved and potentially life-threatening conditions in cancer survivors, while effective clinical management remains quite challenging. As a consequence, morbidity and mortality related to cardiac complications now threaten to offset some favorable benefits of modern cancer treatments in cancer-related survival, regardless of the oncologic prognosis. This review focuses on identifying critical research-practice gaps, addressing real-world challenges and pinpointing real-time insights in general terms under the context of clinical cardiotoxicity induced by the fourth and fifth modalities of cancer treatment. The information ranges from basic science to clinical management in the field of cardio-oncology and crosses the interface between oncology

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and onco-pharmacology. The complexity of the ongoing clinical problem is addressed at different levels. A better understanding of these research-practice gaps may advance research initiatives on the development of mechanism-based diagnoses and treatments for the effective clinical management of cardiotoxicity.

KEYWORDS

cancer therapy-induced cardiotoxicity, chimeric antigen receptor (CAR) T-cell therapy, immune checkpoint inhibitors, molecularly targeted therapeutics, research-practice gaps

1 | INTRODUCTION

Currently available cancer treatments include the traditional surgery, radiotherapy, and conventional chemotherapy approaches and have been extended with two new modalities in recent decades: molecularly targeted therapy (MTT) and immunotherapy. Cardiac toxicities associated with conventional chemotherapy and radiotherapy are well known, and to a certain degree, the cellular and molecular mechanisms leading to cardiotoxicity have been discovered. Similar adverse cardiac events are encountered when applying MTTs, which represent a new generation of anticancer drugs.¹⁻¹¹ The MTTs that have received US FDA approval since their initiation in 2001 are listed in Tables 1 and 2, and 3. In addition, the recently developed immune-based therapeutic modalities, e.g., immune checkpoint inhibitors (ICIs)¹²⁻²⁹ and chimeric antigen receptor (CAR) T-cell therapy (CART),²⁹⁻⁴⁴ have also raised cardiovascular concerns in patients, including lethal cardiotoxicity (Table 3). Cardiotoxicity reflects functional, structural or a combination of both types of damage to the heart by various detrimental environmental insults, 45 e.g., conventional chemotherapy, radiotherapy, MTT, immunotherapy, and toxins. Cardiotoxicity manifests as electrophysiological disorder of the heart (cardiac dysfunction) and a spectrum of myocardial damage (cardiomyopathy), with heart failure as the most severe consequence. The clinical features are demonstrated by a wide range of cardiovascular manifestations or events, including bradycardia, tachycardia, cardiomyopathy, widened pulse pressure, hypotension, arrhythmias, decreased left ventricular ejection fraction (LVEF), troponinemia, QT prolongation, myocarditis, myocardial infarction, pericarditis, acute coronary syndromes, and congestive heart failure.^{20,46-49} The clinical scenarios derived from these novel anticancer approaches reflect new challenges to medical, pharmacological, and research communities because the cardiotoxicity from new clinical entities are clinically and mechanistically different from the cardiotoxicity resulting from traditional chemotherapeutic agents. Cardiotoxicity caused by these novel agents or modalities are diverse, and many specific mechanisms of individual compounds or modalities underlying this toxicity remain to be elucidated.⁵⁰ Some commonly conceptualized mechanisms underlie toxicity, including on- and off-target toxicity, production of toxic metabolites, harmful immune responses, unpredictable specificities of targeted tumor antigens, tumor lysis symptoms, cytokine release syndrome, T-cell receptor (TCR) mispairing, TCR cross-reactivity, and idiosyncratic mechanisms. 51-56

A primary goal of targeted therapy or more advanced immune-based modalities is to kill cancer cells more specifically than traditional treatment modalities while maintaining an acceptable level of side effects and quality of life. Unfortunately, the newer targeted agents or modalities exhibit a similar frequency and severity of toxicities as traditional cytotoxic agents do, albeit with differences in preference for the organs/tissues that are involved.⁵⁷ Many unanticipated short- and long-term adverse effects on multiple organs/tissues have shown the limitations of the new therapies,^{7,30,56-59} although MTTs have improved the overall survival of cancer patients over the past two decades,^{57,60,61} and advanced immunotherapies are promising to improve the outcomes of certain types of cancers (ICIs⁶²⁻⁷⁰ and CART⁷¹⁻⁷⁴). Cardiac complications associated with cancer treatment represent unresolved and potentially life-threatening conditions in cancer survivors. As a consequence, morbidity and mortality related to cardiac

TABLE 1 Approved molecularly targeted therapeutics (non-mAbs) in oncology and reported clinical cardiotoxicity/cardiovascular events from 2001 to May 2017

Generic/Trade Name	Approval Year	Approval Year Major Known Targets	Disease Indications	Cardiotoxicity/cardiovascular events
Gleevec (Imatinib mesylate)	2001,2002	ABL1-2, PDGFR, KIT	Ph+CML, GIST	SR1-13
Velcade (Bortezomib)	2003	Proteasome	Multiple myeloma	SR4,10,12,14-19
Iressa (Gefitinib)	2003	EGFR	NSCLC	SR20,21
Tarceva (Erlotinib)	2004	EGFR	NSCLC	SR22-26
Nexavar (sorafenib)	2005	KIT, PDGFR-B, RET, BRAF, VEGFR2/3, FLT-3	RCC, HCC, GIST	SR3,7,9,10,16,27-29
Sprycel (Dasatinib)	2006	ABL1, SRC	CML	SR3,4,7,8,10,12,16,30-32
Sutent (Sunitinib malate)	2006, 2011	VEGFR, PDGFR, KIT, FLT3, RET, CSF-1R	RCC, GIST, PNT	SR1-4,7-10,12,16,27,30,33-36
Zolinza (Vorinostat)	2006	HDAC	Cutaneous T-cell lymphoma	SR7,10,30,37
Tasigna (Nilotinib)	2007	ABL1, KIT, PDGF	Ph(+) CML	SR3,7,10,12,16,38
Torisel (Temsirolimus)	2007	mTOR	ARCC	SR10,16,39
Tykerb (Lapatinib)	2007	EGFR, ERBB2	Breast cancer	SR3,4,7,8,10,16,25,40-44
Istodax (Romidepsin)	2009	HDAC	Cutaneous T-cell lymphoma	SR7,10,45-50
Afinitor (Everolimus)	2009, 2011, 2012	mTOR	RCC, APNT, HER2(-) breast cancer	SR10,16,25,51
Votrient (Pazopanib)	2009, 2012	VEGFR1/2/3	RCC, soft tissue sarcoma	SR7,10,12,16,52
Xalkori (Crizotinib)	2011	MET, ALK	ALK(+) NSCLC	SR7,12,16,53-55
Vandetanib (Vandetanib)	2011	VEGFR, EGFR	Thyroid cancer	SR7,12,16,56,57
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Generic/Trade Name	Approval Year	Approval Year Major Known Targets	Disease Indications	Cardiotoxicity/cardiovascular events
Zelboraf (Vemurafenib)	2011	BRAF	BRAF(+) melanoma	SR12,28,58-63
JAKAFI (Ruxolitinib)	2011	JAK2	Myelofibrosis	SR64,65
Kyprolis (Carfilzomib)	2012	Proteasome	Multiple myeloma	SR12,16,66-70
Bosulif (Bosutinib)	2012	SRC, ABL1	Ph(+) chronic myelogenous leukemia	SR7,71
Stivarga (Regorafenib)	2012, 2013	VEGFR2, TIE2	Metastatic colorectal cancer, GIST	SR16,28,72
Cabometyx (Cabozantinib)	2012, 2016	RET, MET, VEGFR1/2/3, KIT, TRKB, FLT-3, AXL, TIE-2, TYRO3, MER	Metastatic medullary thyroid cancer, ARCC	SR7,16,73,74
Zaltrap (ziv-aflibercept)	2012	PIGF, VEGF-A	Metastatic colorectal cancer	SR7,10,16,75
Erivedge (Vismodegib)	2012	Hedgehog (Hh) signaling pathway	Basal cell carcinoma	Reports not found yet
Inlyta (Axitinib)	2012	VEGFR1/2/3	ARCC	SR16,76,77
Iclusig (Ponatinib)	2012	Abl, Src	CML, Ph(+) ALL	SR7,12,16,78
Gilotrif (Afatinib)	2013	EGFR, ERBB2, ERBB4	Metastatic NSCLC with EGFR mutations	SR43,79
Tafınlar (Dabrafenib)	2013	BRAF	Melanoma	SR7,80
Mekinist (Trametinib)	2013	MEK1, MEK2	Melanoma with BRAF V600E or V600K mutations	SR7,12,16,81-83
Imbruvica (ibrutinib)	2013, 2014	Bruton's kinase (BTK)	Mantle cell lymphoma, CLL	SR16,84
Zykadia (Ceritinib)	2014	ALK	ALK(+) metastatic NSCLC	SR12,16,85
Zydelig (Idelalisib)	2014	PI3K delta	CLL, follicular B-cell NHL, SLL	SR86
Lynparza (Olaparib)	2014	PARP	BRCA mutated advanced ovarian cancer	SR87
Beleodaq (Belinostat)	2014	HDAC	Relapsed or refractory peripheral T-cell lymphoma	SR88,89
Alecensa (Alectinib)	2015	ALK, RET	ALK-positive, metastatic NSCLC	SR90 (Continues)

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Generic/Trade Name	Approval Year	Approval Year Major Known Targets	Disease Indications	Cardiotoxicity/cardiovascular events
Cotellic (Cobimetinib)	2015	MAPK	BRAF V600E or V600K melanoma	SR91
Lenvima (Lenvatinib)	2015, 2016	VEGFR1/2/3, FGFR1/2/3, PDGFR α , KIT, RET	Thyroid cancer, ARCC	SR16,92
Tagrisso (Osimertinib)	2015	EGFR	EGFR T790M mutation positive NSCLC	SR93
Ibrance (Palbociclib)	2015	CDK4/6	ER(+), HER2(-) breast cancer	SR94,95
Odomzo (Sonidegib)	2015	Hedgehog pathway (Smoothened)	Locally advanced basal cell carcinoma	Reports not found yet
Farydak (Panobinostat)	2015	HDAC	Multiple myeloma	SR47,50,96,97
Ninlaro (Ixazomib)	2015	Beta 5 subunit of the 20S proteasome	Multiple myeloma	SR98
Lonsurf (Trifluridine and tipiracil)	2015	Nucleoside metabolic and thymidine Metastatic colorectal cancer phosphorylase	Metastatic colorectal cancer	SR99
Onivyde(Irinotecan liposome injection) 2015	2015	Topoisomerase	Metastatic pancreatic cancer	SR100
Venclexta (Venetoclax)	2016	BCL-2	CLL with 17p deletion	SR101
Rubraca (Rucaparib)	2016	PARP	Advanced ovarian cancer with BRCA mutation	Reports not found
Alunbrig (brigatinib)	2017	ALK	Advanced ALK-positive metastatic NSCLC	SR163, SR164
Rydapt (midostaurin)	2017	FLT3	${\sf FLT3}\ positive\ acute\ myeloid\ leukemia\ and\ mastocytosis$	SR165
Zejula (niraparib)	2017	Poly(ADP-ribose) polymerase (PARP) inhibitor	Recurrent epithelial ovarian, fallopian tube, etc	SR166
Kisqali (ribociclib)	2017	Cyclin-dependent kinase (CDK) 4 and 6	Breast cancer	SR168

Abbreviations: NSCLC = Non-small-cell lung carcinoma; RCC = Renal cell carcinoma; CML = chronic myelogenous leukemia; GISTs = Gastrointestinal stromal tumors; ARCC = Advanced lymphoblastic leukemia; HDAC = Histone deacetylase; PARP = Poly(ADP-ribose) polymerase; ALK = Anaplastic lymphoma kinase; CSF-1R = Colony stimulating factor receptor Type 1; FLT3 = Fms-like tyrosine kinase-3; PDGFR = Platelet-derived growth factor receptors; PI3K = Phosphoinositide-3 kinase; HCC = Hepatocellular cancer; MAPK = Mitogen-activated prorenal cell carcinoma; RCC = Renal cell carcinoma; PNT = Pancreatic neuroendocrine tumors; APNT = Advanced pancreatic neuroendocrine tumors; Ph(+) = Philadelphia (+); CLL = Chronic tein kinase; EGFR = Epidermal growth factor receptor; VEGFR = Vascular endothelial growth factor receptor; FGFR = Fibroblast growth factor receptors; PIFG = Placental Growth Factor.JAK2 = Janus kinase 2; TRKB = Tropomyosin receptor kinase B; TYRO3 = Tyrosine-protein kinase receptor; mTOR = Mammalian target of rapamycin; SLL = Small lymphocytic lymphoma; SR = Supplemental references.

Approved therapeutic monoclonal antibodies (non-ICIs) in oncology and the reported clinical cardiotoxicity/cardiovascular events from 2001 to May 2017 TABLE 2

Generic/Trade Names	FDA Approval	Antigens	Disease Indications	Cardiotoxicity/Cardiovascular Events
Campath (Alemtuzumab)	2001	CD52	B-CLL	SR102-105
Zevalin (Ibritumomab tiuxetan)	2002	CD20	NHL	SR14,102,106
Bexxar (Tositumomab)	2003	CD20	NHL, follicular NHL	SR107,108
Erbitux (Cetuximab)	2004	EGFR	EGFR(+) MBC	SR10,102,109
Avastin (Bevacizumab)	2004, 2009	VEGFR	Colorectal Cancer, RCC	SR4,7,9,10,16,25,27,40,102,103,110
Vectibix (Panitumumab)	2006	EGFR	Colorectal cancer	SR111
Rituxan (Rituximab)	2006, 2010, 2011, 2012	CD20	Follicular B-cell, CD20 (+) NHL, CLL	SR11,102,103,112,113
Arzerra (Ofatumumab)	2009	CD20	CLM	SR114
Herceptin (Trastuzumab)	2010, 2013	HER2	Gastric cancer, HER2(+) MBC	SR4,7-10,12,16,25,27,31,35,40,42,102,103,115-119
Xgeva (Denosumab)	2010, 2013	RANKL	Bone metastases from solid tumors, GCT	SR120
Perjeta (Pertuzumab)	2012	ERBB2	HER2+ MBC	SR7,16,25,43,102,121-123
Adcetris (Brentuximab vedotin)	2011	CD30	HL, ALCL	SR102,103,124
Kadcyla (Trastuzumab emtansine)	2013	HER2	HER2(+)MBC	SR16,43,102,103,125
Gazyva (Obinutuzumab)	2013	CD20	Chronic lymphocytic leukemia,	SR102,126,127
Cyramza (Ramucirumab)	2014	VEGFR	Gastric cancer	SR16,84,102,128
Darzalex (Daratumumab)	2015	CD38	Multiple myeloma	SR129
Empliciti (Elotuzumab)	2015	SLAMF7	Multiple myeloma	SR130
Portrazza (Necitumumab)	2015	EGFR	Metastatic SNSCLC	SR131
Unituxin (Dinutuximab)	2015	Glycolipid GD2	Neuroblastoma	SR132
Lartruvo (Olaratumab)	2016	$PDGFR_{lpha}$	Soft tissue sarcoma	SR133

Abbreviations: NHL = Non-Hodgkin lymphoma; HL = Hodgkin lymphoma; GCT = Giant cell tumor of bone; SLAMF7 = Signaling Lymphocytic Activation Molecule Family member 7; $CLM = Chronic Iymphocytic leukemia; RANKL = Receptor activator of nuclear factor kappa-B ligand; PDGFRa = Platelet-derived growth factor receptor alpha; IL-6 = Interleukin <math>\delta$; Non-ICIs = Non-immune checkpoint inhibitors; RCC = Renal cell carcinoma; B-CLL = B-cell chronic lymphocytic leukemia; PDGFR = Platelet-derived growth receptor; CLL = Chronic lymphocytic

leukemia; SNSCLC = Squamous non-small cell lung cancer; MBC = Metastatic breast cancer; ALCL = Anaplastic large cell lymphoma; SR = Supplemental references.

TABLE 3 Immune checkpoint inhibitors and CART in oncology and the reported clinical cardiotoxicity/cardiovascular events from 2011 to May 2017

Generic/Trade Name	Approval Year	Category	Antigens	Disease Indications	Cardiotoxicity/ Cardiovascular Events
Yervoy (Ipilimumab)	2011	ICI	CTLA-4	Metastatic melanoma	SR16,134-136
Opdivo (Nivolumab)	2014, 2015, 2016	ICI	PD-1	Metastatic melanoma, metastatic SNSCLC, ARCC	SR16,135-140
Keytruda (Pembrolizumab)	2014, 2015, 2016	ICI	PD-1	Head and neck SCC	SR16,135- 137,141-143
Tecentriq (Atezolizumab)	2016	ICI	PD-L1	Urothelial carcinoma and metastatic NSCLC	SR144
Bavencio (avelumab)	2017	ICI	PD-L1	Merkel cell carcinoma	SR160, SR161, SR167
Imfinzi (durvalumab)	2017	ICI	PD-L1	Advanced or metastatic urothelial carcinoma	SR162, SR167
CART	Not yet	CART	CD19, MAGE-A3	Hematopoietic malignancies, melanoma	SR145-159

Abbreviations: CTLA-4 = Cytotoxic T-lymphocyte-associated antigen 4; ICIs = Immune checkpoint inhibitors; ARCC = Advanced renal cell carcinoma; CHL = Classical Hodgkin lymphoma; SNSCLC = Metastatic squamous non-small cell lung cancer; SCC = Squamous cell carcinoma; NSCLC = Non-small cell lung cancer; CART = Chimeric antigen receptor T cell therapy; SR = Supplemental references.

complications now threaten to offset some of the favorable benefits of these novel cancer treatments in cancer-related survival and strongly impact the quality of life regardless of the oncologic prognosis. $^{75-77}$ Additionally, effective clinical management remains quite challenging because no evidence-based approaches are currently available for the effective monitoring and treatment of these patients. 78,79 This information is lacking because the risk of cardiovascular toxicity greatly differs from one treatment to another according to the targeted pathways or antigens and the presence of co-morbid conditions.^{75,80} Because the novel drugs or modalities are highly cost-intensive,^{81–84} cardiotoxicity may add additional costs without improving the outcome of a life-limiting illness with a generally unpredictable occurrence and uncertain cure. 8.54,85.86 Clinical cardiotoxicity challenges not only sustainable cancer survivorship but also precision oncology/medicine and affordable healthcare. Cost savings are a major incentive for the adoption of biosimilars, and some biosimilars have been developed and used in clinical oncology trials.^{87,88} Biosimilars refer to biologic products that demonstrate no clinically meaningful differences in terms of quality attributes, efficacy, safety, and immunogenicity compared with an existing licensed, originator biologic.⁸⁹ The incorporation of biosimilars into healthcare systems worldwide may result in a 30-45% cost savings.⁸⁷ A strategy to prevent clinical cardiotoxicity will have significant impacts on the overall prognosis and survival of cancer patients. This strategy is part of the field of cardiovascular safety, with multifaceted aspects at multiple levels involving improvement of preclinical models for the study of chemotherapy-induced cardiotoxicity, 90 appropriate preapproval investigations and monitoring during clinical practice. 91 In addition, it is important to realize that the approval decision following preapproval clinical trials does not necessarily represent a singular moment of clarity about the risks and benefits associated with a drug; thus, continuous post-marketing surveillance and vigilance for cardiotoxicity is required. 91 The research gaps in novel cancer treatment-related cardiotoxicity range from basic, translational, clinical, and epidemiologic sciences to cancer patients, medical professionals, research communities, pharmaceutical industries, regulatory bodies, and research funding agencies. In response to ongoing clinical challenges, "cardio-oncology" or "onco-cardiology" represents a new multidisciplinary discipline in recent decades 92,93 and is a newer frontier in clinical medicine leading to advances in clinical care, medical education, and dual subspecialist training programs. 94-96 The subspecialty harbors more questions than answers; thus, enormous research opportunities are embedded in the field. 76 The research is expected to address gaps from basic, translational, and clinical studies to epidemiological studies and to exploit the synergistic interactions among multiple disciplines. A joint effort of multiple disciplines, including cardiology, oncology, and clinical pharmacology is to lead advances in the field of cardio-oncology in hopes of ultimately eliminating cardiac diseases as a barrier to effective cancer therapy while providing affordable care to patients. This review focuses on the identification of the current gaps between research and clinical practice and addresses the challenges and pinpoints current insights of clinical cardiotoxicity induced by the fourth and fifth modalities of cancer treatments.

2 | FDA-APPROVED MTTS FOR ONCOLOGY

According to the latest reports, 97-100 approximately 76 targeted anticancer drugs received approval from the US FDA from 2001 to May 2017; these drugs include small molecule kinase inhibitors, other types of agents (e.g., proteasome and histone deacetylases targeting inhibitors) (Table 1), and therapeutic monoclonal antibodies (Tables 2 and 3). The clinical patterns of toxicity associated with some of these agents have been addressed or discussed in detail in other reviews.^{20,46-49} Many of the approved drugs are protein kinase inhibitors (PKIs), typically protein tyrosine kinase inhibitors (TKIs). 101-103 Only a few inhibitors affect serine/threonine kinases (e.g., Vemurafenib and Dabrafenib), 102 and one is a lipid kinase inhibitor (idelalisib). 104 This distribution is in line with the fact that tyrosine phosphorylation is a unique biochemical mechanism utilized by intra- and intercellular communication pathways, and these kinases regulate important fundamental cellular processes, including cell proliferation, differentiation, migration, and metabolism. 103 Protein kinases catalyze the transfer of the terminal phosphate of ATP to substrates that usually contain a serine, threonine, or tyrosine residue; this phosphate then serves as a ubiquitous mechanism for cellular signal transduction. 105 Thus, protein kinases are ATP-consuming enzymes with a high degree of conservation in their kinase domain, 8,105,106 especially for those within the family of protein tyrosine kinases (PTKs).^{2,8,101} The ATP-consuming feature is unfortunately associated with the ATP required for the vitality and viability of cardiomyocytes. Mitochondria are abundant in heart tissue, constituting approximately 45% of the myocardial volume, which is high in comparison with other tissues. 107 The abundance is due to the high-energy demand of the heart, which is satisfied during mitochondrial respiration; thus, more than 90% of the ATP generated by the mitochondria is utilized by cardiomyocytes for myocardial function and viability. 107

The protein kinase family includes more than 385 serine/threonine kinases and more than 90 PTKs. 102,103,105 More than 900 protein-encoding genes with kinase activity have been confirmed. 108-110 Out of 90 PTKs, 58 are receptors with an extracellular, transmembrane, and intracellular domain, and 32 are intracellular non-receptors. 102,111,112 Kinases comprise one of the largest classes of proteins encoded by the human genome, and their signaling molecules play an important role in regulating almost every aspect of cell function in many different tissue types and organs, ^{102,105,108,109} including the cell fate decisions of cardiac myocytes that lead to cardiac pathologies. ^{7,113} Thus, dysfunctional kinase activity disrupts the normal control of cellular phosphorylation signaling pathways and leads to numerous pathologies beyond cancer through pathway affiliation, 99 including immunological, neurological, metabolic, infectious diseases, diabetes, osteoporosis, and otology. 7,98,101,102,105,113-115 Consequently, the pharmacological utility of kinase-targeting has been expanded for kinase-targeted therapies in a broad array of indications beyond cancers. 98 The selectivity of drug-kinase interactions has become a common concern in clinical pharmacology, 116 and there is continuing debate surrounding target selection, mechanism of action, compound development prioritization, toxicity, and patient tailoring. 116,117 Kinase inhibitors also provide additional opportunities to investigate and $elucidate\ kinase\ functions\ under\ various\ circumstances,\ both\ physiologically\ and\ pathologically. ^{104}\ Intensive\ efforts$ in the exploration of disease targets have significantly extended the coverage of druggable targets in the human kinome from the tyrosine kinase family to several other families, such as calmodulin/calcium-regulated kinase, glycogen synthase kinase (GSK), cGMP-dependent protein kinase (PKG), cAMP-dependent protein kinase (PKA), CDClike kinase (CLK), and protein kinase C (PKC). 98,104 In addition, targeting the ubiquitin-proteasome pathway is an emerging concept in cancer therapy based on the hypothesis that many proteins in the proteasome are implicated in the regulation of important processes of carcinogenesis and cancer cell survival. 118 Several proteasome-targeting inhibitors (e.g., bortezomib, $^{20,47,119-125}$ carfilzomib $^{20,124,126-130}$) have been introduced in the clinic. A retrospective analysis of 3954 patients in phase 2/3 trials of bortezomib for the treatment of multiple myeloma reported that bortezomib-based treatment was associated with a low incidence of cardiac events. 131 Epigenetic cancer therapy using histone deacetylase inhibitors is also emerging, $^{132-134}$ as epigenetic gene silencing is a hallmark of cancer cells. Two important types of epigenetic changes are DNA methylation by DNA methyltransferases (DNMTs) and histone modification by histone deacetylases (HDACs). 133,134 DNMTs and HDACs have become attractive therapeutic targets, and several HDAC inhibitors e.g., Romidepsin, $^{47,57,135-140}$ Belinostat, 141,142 and Panobinostat, 137,140,143,144 have been used in the clinic. Unfortunately, clinical cardiotoxicity has been induced by these types of drugs. $^{47,57,135-144}$

3 | KINASE INHIBITORS AND CARDIAC CONCERNS

Based on kinship in structure and activity (different binding modes), kinase inhibitors can be grouped into two classes: irreversible and reversible. \$^{102,104,109,114,116,145}\$ Irreversible inhibitors form covalent bonds with cysteine or other nucleophilic residues in the ATP-binding pocket. \$^{105,116}\$ The advantage of an irreversible inhibitor is its ability to block the active site of the kinase, which might also block mutated versions of the kinase. \$^{101}\$ Theoretically, irreversible inhibitors may be superior to their reversible counterparts in several aspects. For example, they circumvent competition with high ATP concentrations in the cell and are less affected by changes in the ATP-binding affinity, and they have prolonged pharmacologic effects, high potency, and the ability to validate pharmacological specificity through mutation of the reactive cysteine residue. \$^{103,105}\$ However, progress in the implementation of irreversible kinase inhibitors in the clinic remains slow because of safety and toxicity concerns, \$^{109,146}\$ as covalently binding kinase inhibitors require the intrinsic reactivity of cysteine-reactive groups, which can lead to non-selective reactions with off-target proteins that increase toxicity. $^{101,147-150}$

Kinase inhibitors are further categorized into four main types based on their binding mode to a $target^{109,151}$: type I inhibitors constitute the majority of ATP-competitive inhibitors and recognize the active conformation of the kinase; type II inhibitors recognize the inactive conformation of the kinase; type III inhibitors bind next to the ATP-binding pocket; and type IV (irreversible inhibitors) do not bind to the ATP or peptide substrate binding sites 102,109,116,145,152 but characteristically form covalent bonds with their target enzyme. 109,145 Thus, type III and IV inhibitors are allosteric in nature. 109,114,150 So-called allosteric sites refer to locations outside of the ATP-binding pocket, and allosteric inhibitors block protein kinase catalytic activity while having no effect on ATP binding. 109 Allosteric inhibitors bind outside the ATP-binding site and thus induce a conformational shift in the target enzyme to block the kinase function. 150 Currently, the vast majority of PKIs in the clinic are type I inhibitors that target the ATP-binding site in its active conformation (reversible ATP-competitive inhibitors). 101,102,105,108,114,150,153 With high relevance in cardiac safety liabilities, kinases have one ATP-binding site, and thus, cardiotoxicity would theoretically be inevitable if a targeted kinase was functionally expressed in the heart (on-target toxicity) when ATP-competitive inhibitors are used. Even if a targeted kinase is not expressed in the heart, off-target toxicity is also a major issue for both ATP-competitive kinase inhibitors¹⁵⁴ and non-ATP-competitive kinase inhibitors (allosteric inhibitors). 101,147-150 Assuming an ATPcompetitive case, an energy system dysfunction or energy deprivation of cardiomyocytes becomes a high risk. This risk is in line with the fact that cardiac muscle tissue is highly energetic, and cardiac performance is typically reliant on aerobic metabolism as a source of ATP; impairments in this process can rapidly induce cardiac dysfunction. 155 Pharmacologically, a theoretical solution to this problem is the development of covalent-allosteric kinase inhibitors (type III and IV inhibitors), 156 in which an active site-directed moiety is tethered to another ligand that targets a location outside of the ATP-binding cleft. In principle, allosteric inhibitors should be superior to active-site-directed inhibitors due to the high degree of kinase selectivity because these sites are highly divergent across the kinome and are unique to a particular kinase. 101,114,116,151 In contrast, the high sequence similarity in the ATP-binding site among members of the kinase family often results in low selectivity and additional toxicities of these ATP-competitive inhibitors. Considering the superiority of allosteric inhibitors, the development of inhibitors that target sites other than the ATP cleft has been the reality in clinic. Trametinib is the only type III inhibitor that has been approved thus far, although several promising allosteric kinase inhibitors are currently in different stages of clinical trials. ^{104,109} Only a few are irreversible ones (afatinib and ibrutinib)¹⁰⁴ that have been approved by the FDA (Table 1). Currently, clinical cardiotoxicity induced by irreversible inhibitors, e.g., Trametinib, ^{20,57,124,157-159} Ibrutinib, ^{20,160,161} and Afatinib^{162,163} has been reported. Thus, novel mechanisms of action need to be further explored in type III and IV inhibitors. ¹⁰⁴ Despite the advantages of covalent-allosteric kinase inhibitors versus orthosteric inhibitors (types I and II), whether allosteric kinase inhibitors can radically overcome the non-selective problem (most importantly, considering clinical cardiotoxicity) while retaining their potency or effectiveness on cancer cells^{104,164} in the clinical endpoint remains unclear. The ultimate answer to this question, considering cardiac safety concerns and potency on cancer cells, will derive from the accumulated clinical data in the future because the predictive values of success in the preclinical model systems are limited to the use of homogenous cell lines and immune-compromised animals. ¹⁰¹

Because protein kinases play a key role in all aspects of cancer, kinase inhibitors represent the largest family of targeted agents that have entered or are entering the clinic. 101 However, an important concern is that only a very small percentage of kinases (approximately one-fifth of the human kinome) have been successfully targeted clinically, 104,165 Thus, the field remains largely unexplored in the research of numerous undiscovered kinases, their respective inhibitors, and their potential cardiotoxicity/toxicities. Moreover, approximately 10-20% of the kinases are classified as pseudokinases 109,166 because of the lack of one or several of the highly conserved motifs involved in nucleotide (nt) binding or catalytic activity of protein kinases. 110,165,166 Many pseudokinases in the kinome have evolved from active kinases by obtaining regulatory functions in which catalytic function is dispensable; however, a significant proportion of pseudokinases have retained their ATP-binding ability. 166 An important molecular property of pseudokinases is that they serve as allosteric regulators of signaling pathways. ¹⁰⁹ Pseudokinases also play an important regulatory role in cellular signaling, and the abnormal function of several human pseudokinases has been associated with human diseases, including cancers. 110,165-172 Pharmacological targeting of pseudokinases may be a possible alternative option in this regard. 165,173,174 For example, the receptor tyrosine kinase (RTK)-like orphan receptors (RORs) are RTK-like pseudokinases in this context. ¹⁷⁵ ROR1 is a type 1 transmembrane protein expressed on the plasma membrane and has an extracellular domain that is essential for ligand binding and signal transduction. 176,177 Interestingly, ROR1 is selectively overexpressed in many hematologic malignancies and a number of solid tumors, while it is without significant expression in normal adult tissues. 176,178,179 The unique expression profile of ROR1 makes it a promising candidate for novel drug targets, ¹⁷⁶ assuming that this expression pattern will maximize effectiveness and minimize off-target toxicities. A monoclonal antibody (cirmtuzumab) was developed by binding to ROR1 on tumor cells and inhibiting Wnt5a signaling, which is a pathway that is important for blocking tumor-cell proliferation, migration, and survival. 180 The antibody has been used in a phase I clinical trial for chronic lymphocytic leukemia. 175,178,179,181,182

4 | CAVEATS, CHALLENGES, AND INSIGHTS ON THE DIAGNOSIS AND THERAPY OF MTT-INDUCED CARDIOTOXICITY

4.1 | Caveats

An important caveat for clinicians is the fact that in many cases, the adverse cardiac events observed in the clinic were unobserved in preclinical safety evaluations or insufficiently addressed in clinical trials; these events may not even be mentioned on product labels when placed on the market. ^{2,8,58,154,183} Cardiotoxicity may become apparent only in routine clinical practice. ² Therefore, it is important to identify new models or techniques in both preclinical and clinical settings ¹⁵⁴ that can better predict adverse clinical outcomes with these agents. Some studies have shown that cancer treatment-related cardiotoxicity is the third leading cause of treatment-associated mortality in survivors of pediatric and adolescent cancers, with recurrence and second malignancies being the two leading causes. ¹⁸⁴ In adult patients, cardiotoxicity is agent-dependent, and the incidence can be as high as 50%, depending on the cardiac conditions. ¹⁸⁵ The incidence of treatment-induced heart damage in pediatric cancer survivors increases over time ^{186,187} and can be identified many years post-launch or even 30 years after treatment, as shown in adult survivors of childhood cancers. ^{186,188-192} A constant vigilance for cardiotoxicity is required ⁹¹ and is especially important for pediatric

cancer patients. These pediatric patients have a higher risk of developing treatment-associated cardiotoxicity/ toxicities subject to age-associated, tissue-specific sensitivity to cancer therapeutics 193 and can exhibit an insidious onset (occult cardiotoxicity). 194 Clinicians practice in a sea of uncertainty, with relatively few strategies leading to the safe prescription of anticancer drugs or use of therapeutic modalities. This uncertainty poses a threat to making life-altering decisions based on incomplete information in real-world practices. In addition, the so-called "breakthrough therapy designation" (applied to many novel anticancer therapies by the FDA since 2012) 195-208 refers to the allowance of particular drugs to have an accelerated approval timeframe in order to provide new treatment options for difficult diseases and conditions. Thus, the designation by no means guarantees an escape from cardiotoxicity of any investigative or post-marketing drugs in clinical endpoints. Breakthrough "magic bullet" therapies have been criticized by medical professionals who emphasize the importance of evidentiary requirements for meaningful clinical data. 209,210 Beyond cancer, treatment-related cardiotoxicity is a growing concern (not restricted to anticancer $agents), and almost all the rapeutic drug classes have unanticipated cardiotoxicities. ^{154,211} One \, major \, problem \, is \, chronellem \, in the rapeutic drug classes have unanticipated cardiotoxicities. ^{154,211} One \, major \, problem \, is \, chronellem \, in the rapeutic drug classes have unanticipated cardiotoxicities. ^{154,211} One \, major \, problem \, in the rapeutic drug classes have unanticipated cardiotoxicities. ^{154,211} One \, major \, problem \, in the rapeutic drug classes have unanticipated cardiotoxicities. ^{154,211} One \, major \, problem \, in the rapeutic drug classes have unanticipated cardiotoxicities. ^{154,211} One \, major \, problem \, in the rapeutic drug classes have unanticipated cardiotoxicities. ^{154,211} One \, major \, problem \, in the rapeutic drug classes have unanticipated cardiotoxicities. ^{154,211} One \, major \, problem \, in the rapeutic drug classes have unanticipated cardiotoxicities. ^{154,211} One \, major \, problem \, in the rapeutic drug classes have unanticipated cardiotoxicities. ^{154,211} One \, major \, problem \, in the rapeutic drug classes have unanticipated cardiotoxicities. ^{154,211} One \, major \, problem \, in the rapeutic drug classes have unanticipated cardiotoxicities. ^{154,211} One \, major \, problem \, in the rapeutic drug classes have a problem by the rapeutic drug classe$ ically administered drugs, such as neurologic/psychiatric agents and anticancer chemotherapeutic agents, because toxicity may become evident only after long-term accumulation of the drug or its metabolites (toxicology-metabolic activation) 148 in the heart. 135,136,193 Cancer has been transformed from a rapidly fatal disease into a chronic condition due to advancements in detection and treatment.^{212,213} As such, both short- and long-term cardiotoxic effects resulting from cancer therapy are becoming more evident.²¹³ The long-term adverse effects are typically seen in adult survivors of childhood cancer, but treatment-related cardiotoxicity can develop at any time after treatment initiation and can occur well into adulthood. 188-192 Given the concern of cardiac safety liabilities in relation to non-anticancer drugs, a particular caution should be taken when repurposing non-anticancer drugs for novel applications in oncology, 214,215

4.2 | Challenges, insights, and perspectives

4.2.1 | Current diagnostic modalities and biomarkers

Preventing cardiac damage is far more important than using a therapeutic intervention to counteract ongoing damage because in general, the heart is a terminally differentiated organ, and adult mammalian myocardium has very limited regeneration potential after injury.⁴⁵ Thus, prompt recognition of early signs of cardiotoxicity and initiation of appropriate management prior to major clinical manifestations (i.e., the irreversible phases) is paramount for the substantial recovery of cardiac function in the hope of preventing the development of irreversible cardiotoxicity.²¹⁶⁻²¹⁹ "Early detection" theoretically refers to the identification of early functional cardiotoxicity - at this stage, no morphological damage to cardiomyocytes is detectable, and the damage is usually reversible and clinically manifests as asymptomatic or subclinical cardiotoxicity, 185,220-222 Unlike functional cardiotoxicity, structural cardiotoxicity often appears in late stages and is usually irreversible, with detectable morphological and symptomatic damage. 185,220 For the detection of cardiotoxicity and the initiation of therapeutic measures in the early stages, a set of diagnostic and prognostic $methods\ have\ been\ suggested,\ including\ clinical,\ imaging,\ serological,\ and\ molecular\ investigations. \\ ^{198-200,205}\ Unformation for the control of the contr$ tunately, while 2D echocardiography or radionuclide angiography, ECG, and several blood markers that are commonly used in clinical practice allow the late diagnosis of cardiac dysfunction, these tests have a low sensitivity and insufficient predictive power in detecting subtle, incipient myocardial injury. 216,223,224 A cardiac biopsy-based approach is highly invasive and is not a suitable option for routinely diagnosing and monitoring cancer treatment-related cardiotoxicity. Noninvasive or minimally invasive approaches should be considered first. Furthermore, the currently used cardiotoxicity biomarkers provide few mechanistic insights into the underlying mechanisms that lead to the identification of actionable and mechanistic biomarkers--understanding these mechanisms is a fundamental effort to translational medicine. Circulating biochemical markers are alternative or synergistic tools for clinical diagnosis. However, circulating biochemical markers harbor potential problems related to sensitivity and specificity with respect to systemic disease because this type of biomarker could be significantly influenced by multiple microenvironmental factors both locally and systemically. Cancer is a systemic disease with local manifestations,²²⁵ and cancer cachexia of multiple organs, including cardiac cachexia,²²⁶ is a typical reflection of its systemic nature. An ideal circulating biomarker is defined by two important characteristics: disease specificity and a linear relationship (sensitivity) between the serum/plasma concentration and disease severity.²²⁷ For instance, cardiac troponin (cTn), B-type natriuretic peptide (BNP), and its N-terminal fragment (NT-proBNP) are the best-studied circulating biochemical markers of cardio-vascular diseases, including cardiotoxicity, and they have been used primarily in the clinic.^{228–230} However, cTn, BNP, and NT-proBNP are elevated not only in patients with acute and chronic cardiovascular diseases but also in patients with non-cardiovascular diseases,^{227,231} including untreated cancers.^{232–234} Therefore, these biomarker values may reflect more than the disease activity to which they are applied.^{228,235} Moreover, a new concern has been raised for the potential impact of the tumor itself on cardiovascular health; the underlying cancer pathophysiology may also affect cardiac biomarkers.²³³

4.2.2 | Application and development of advanced diagnostic modalities and biomarkers

Advanced non-nuclear molecular imaging modalities

The gap in relation to the early detection/diagnosis of cardiotoxicity should be filled by combined clinical and translational research. Advanced diagnostic imaging modalities are gaining momentum. Imaging methods have the advantage of in situ visualization of real-time cardiac performance with dynamic profiles to assess cardiac function. Currently, advanced, non-nuclear molecular imaging modalities in the clinic mainly include 3D echocardiography, cardiac magnetic resonance (CMR) imaging, cardiac computed tomography (CT), 3D speckle tracking echocardiography (STE), and cine images. Among these techniques, 3D echocardiography strain rate/deformation imaging, metabolic imaging, and myocardial systolic velocities seem more sensitive and should facilitate the identification of patients with more subtle measures of cardiac function. 236-240 Recently, CMR imaging 241,242 and cardiac CT243 have been explored for their utility in evaluating cancer treatment-induced cardiotoxicity. Cardiac CT has an excellent diagnostic performance for the detection of subclinical atherosclerosis, coronary atherosclerotic plaque, and obstructive coronary artery disease, 243 while CMR has added value in characterizing myocardial remodeling.²⁴¹ Conceptually, left ventricular (LV) remodeling is defined in response to myocardial injury or overload through chamber dilation and/or hypertrophy.²⁴⁴⁻²⁴⁶ Remodeling restores the contour of the chamber by removing poorly functioning areas to improve its efficiency.²⁴⁷ Classification of such states can be achieved by evaluating LV mass, LV volume, the ratio of LV mass/volume (M/V), and relative LV wall thickness (RWT).^{248–250} Thus, the spectrum of LV geometric adaptation can be measured using these geometric parameters. Ventricular hypertrophy (VH) is classified as concentric when the RWT is increased, while VH is classified as eccentric when the RWT is not increased; a third pattern, termed concentric remodeling, occurs when RWT but not ventricular mass is increased.^{244,248} Analysis of cardiac function is the major focus of echocardiography, and measuring LVEF has been the clinical standard; however, LVEF has shown its limitations for early detection of myocardial dysfunction. 251,252 In the past decade, STE has become a novel clinical tool for the analysis of regional and global myocardial function, and this tool has the potential to provide a more accurate assessment of overall and regional myocardial function.^{251,253} Cine images are short movies that show heart motion throughout the cardiac cycle, and the images can be very useful in studying cardiac and valvular function and the movement of blood through the heart.^{254,255} As the myocardium contracts and thickens throughout the cardiac cycle, any abnormality in this wall motion indicates a problem with the myocardium, such as ischemia or infarct.²⁵⁶ Cardiac cine images have already $been \, used \, to \, evaluate \, cardiotoxicity \, in \, breast \, cancer \, patients \, treated \, with \, trastuzum ab. ^{257} \, Although \, each \, non-nuclear \, cardiotoxicity \, in \, breast \, cancer \, patients \, treated \, with \, trastuzum ab. ^{257} \, Although \, each \, non-nuclear \, cardiotoxicity \, in \, breast \, cancer \, patients \, treated \, with \, trastuzum ab. ^{257} \, Although \, each \, non-nuclear \, cardiotoxicity \, in \, breast \, cancer \, patients \, treated \, with \, trastuzum ab. ^{257} \, Although \, each \, non-nuclear \, cardiotoxicity \, in \, breast \, cancer \, patients \, treated \, with \, trastuzum ab. ^{257} \, Although \, each \, non-nuclear \, cardiotoxicity \, in \, breast \, cancer \, patients \, cardiotoxicity \, in \, breast \, cancer \, patients \, cardiotoxicity \, cardi$ molecular imaging modality has their different strengths and weaknesses, a common feature of these modalities is the use of nonradioactive probes such as light or sound. In addition, these modalities are mainly used to detect changes in the anatomical and physiological features of cardiac events rather than function at the cellular and molecular level. In this regard, these diagnostic modalities have less value for understanding disease processes than do molecular imaging modalities (see the "Nuclear molecular imaging" section).

Nuclear molecular imaging

In contrast to traditional imaging modalities, molecular imaging in the myocardium has great potential to contribute to clinical cardiovascular medicine by improving the understanding of disease processes and therapeutic mechanisms

or effects, 258 enabling the visualization and interrogation of specific biologic targets and pathways that precede or underlie changes in morphology, physiology, and function of the heart prior to the manifestation of gross anatomical features or physiological consequences.²⁵⁸⁻²⁶¹ Thus, the potential for molecular imaging is much greater than the detection of changes in anatomical and physiological features using non-molecular imaging modalities, such as blood flow or contractile function. 262,263 In the preclinical setting, molecular imaging of the cardiovascular system uses multiple modalities, including optical, nuclear, MR, CT, ultrasound imaging, and fluorescence imaging, 258,263 However, clinical application in humans has been mainly restricted to the use of nuclear imaging techniques, 258,262 e.g., single photon emission computed tomography (SPECT) and positron emission tomography (PET). Nuclear molecular imaging characterizes specific disease processes (functions) in different individuals (personalized patient care) using the visualization, characterization, and measurement of biological processes at the molecular and cellular levels in humans and other living systems. Therefore, nuclear molecular imaging is different from routine X-rays, CT, MRI, or echocardiography, which largely show how the body appears structurally rather than how it functions. However, nuclear molecular imaging uses radioactive pharmaceuticals and traces their progress through the body to "view" how the body is functioning. The application of radioactive substances in the diagnosis and treatment of disease is a unique feature for nuclear imaging. Currently, the clinical application of molecular cardiac imaging based on nuclear imaging is largely limited to the imaging of metabolism and innervation.²⁵⁸ Activation of the cardiac sympathetic nervous system is a cardinal pathophysiological abnormality associated with the failing human heart. 264 Several SPECT and PET radiotracers can be used for imaging pre- and post-synaptic function. 265,266 Among those radiotracers, the norepinephrine analog¹²³ Imetaiodobenzylguanidine (¹²³ IMIBG) is FDA approved and the most commonly used agent. This agent can be used to investigate the activity of norepinephrine, which is the predominant neurotransmitter of the sympathetic nervous system.²⁶⁷⁻²⁶⁹ The discovery and application of agents such as ¹²³IMIBG are absolutely needed in clinical nuclear imaging to provide better prognostic risk stratification, which, in heart failure, may include innervation imaging (sympathetic imaging), because this imaging modality provides more accurate information than conventional markers and mechanistic insights that drive therapeutic decisions. 258,259,267 A reduced 123 IMIBGmyocardial uptake or higher washout rate can predict cardiac adverse events. 123,267 IMIBG imaging is more sensitive than decreased LVEF, which has been frequently used in the clinic,²⁶⁸ because a decrease in LVEF is a late manifestation of cardiotoxicity.²⁵² Moreover, with the imaging modality, it is also possible to predict whether LVEF recovery will occur.²⁶⁹ Indeed, seeking early predictors of cardiotoxicity is urgently needed so that treatment can be initiated earlier or that pre-emptive intervention can be used²⁷⁰ to prevent irreversible cardiac damage.

Photoacoustic imaging

Radiotracer-free molecular imaging modalities may serve as alternatives to nuclear molecular imaging. However, an important concern in the application of these imaging techniques in the clinic is the technical challenge of limited penetration depth in contrast to nuclear molecular imaging using radiotracers.^{258,262} To overcome these technical challenges, some advances are being made to provide the required signal amplification in order to enable molecular imaging with contrast agents, e.g., new nanoparticle contrast agents coupled with MRI and ultrasound.²⁶² However, the clinical safety of these contrast agents must be further evaluated in human studies.²⁶² To date, no single parameter or approach can accurately predict cardiotoxicity in the clinic. Multi-parameter test panels or modalities may be beneficial for preclinical and clinical investigations or applications because they enable the acquisition of complementary information from each panel/modality.²⁷¹ Each imaging modality has its intrinsic advantages and disadvantages for cardiovascular events,²⁶³ but none of the current clinically used modalities can simultaneously image both molecular events and anatomical and physiological features. Medical diagnosis and therapeutic options benefit greatly from imaging technologies that combine molecular and microscopic parameters with clinical observations; such a combination is provided by photoacoustic imaging (also known as optoacoustic imaging).²⁷²⁻²⁷⁶ Photoacoustic imaging is a label-free, non-ionizing, noninvasive, high-resolution optical imaging modality that uses optical absorption contrast and ultrasonic resolution. $^{277-282}$ This technology has a high scalability and allows imaging of biological structures, rang $ing from \, molecules, or ganelles, cells, and \, tissues \, to \, entire \, organs \, and \, even \, entire \, small \, animal \, bodies. \\ ^{273,275,280,283-286}$ The anatomical, functional, metabolic, and histologic properties of tissues or organs can be solely revealed by endogenous contrast (e.g., hemoglobin, lipids, melanin, and collagen), while exogenous contrast agents are only used to further increase the imaging contrast and specificity. ^{273,283} Various exogenous probes with high contrast have also been extensively developed, including inorganic and organic dyes, ²⁷⁴ magneto-optical and photochromic probes, ²⁸³ nanoparticles, ^{282,287} and genetically encoded probes, ²⁸¹ to achieve improved resolution and sensitivity while providing multi-parametric photoacoustic imaging. ²⁸³ This imaging modality has proven its clinical and preclinical value in functional, structural, and molecular aspects of diseases and has been used for physiologically and pathologically imaging various organs and tissues, including breast cancer, ^{288–292} neural tissues, ^{277,287,293–295} fingers, ²⁹⁶ sentinel lymph nodes, ^{277,292,297} the cardiovascular system, ^{298–302} the prostate, ^{303,304} skin, ³⁰⁵ cancer therapy, ^{273,274,306} muscle oxygenation, ³⁰⁷ metabolic status, ^{274,283} eyes, ^{308,309} plaque pathophysiology, ³¹⁰ tumor microenvironment (pH, enzymes, radical oxidation species (ROS), and metal ions, among others), ²⁷⁴ and biomaterial-tissue interactions to assess the functions of the engineered tissue/organ constructs. ²⁷⁶ Although photoacoustic imaging is conceptually different from some imaging modalities, it is complementary to many other imaging modalities. The major advantages include deep tissue penetration, good spatial and temporal resolution, a highly scalable nature, and dynamic imaging without ionizing radiation, ^{272,275,276} thus enabling the potential acceleration of its clinical translation and application.

Economically, ultrasound-based technologies are a valuable diagnostic tool for potential global adoption because of their affordability, availability, and portability.²⁷³ Currently, photoacoustic imaging has three major implementations with excellent scalability to meet the application at different levels^{283–286,311}: (1) photoacoustic microscopy (PAM), (2) photoacoustic computed tomography (PAT) or multispectral optoacoustic tomography (MSOT), and (3) photoacoustic endoscopy (PAE). PAT and MSOT are seen as a future alternative to conventional scanning methods, such as MRI and CT scan.^{311,312} These imaging modalities combine non-ionizing optical and ultrasonic waves via the photoacoustic effect, which provides in vivo multiscale functional, metabolic, and molecular imaging.³¹³ Photoacoustic imaging is starting to be used on patients, and the technology may revolutionize medical imaging in clinical practice.^{311–315} PAT/MSOT may be an optimal imaging modality for the detection of clinical and experimental cardiotoxicity. Thus far, PAT/MSOT has not been explored in clinical cardiac imaging to the best of our knowledge. However, experimental cardiac imaging using PAT/MSOT or PAM has been reported.^{298,302,316–320} The imaging modality may yield new insights into the cardiomyocytes and their life in vivo and further contribute to clinical imaging science and diagnosis.

Development of novel, non-invasive, and cost-effective biomarkers

Multiple diagnostic tools can assess cardiac abnormalities, and cardiac biomarkers may play a complementary role to cardiac imaging in monitoring patients for cardiotoxicity. However, biomarkers that could help identify the risk for cardiotoxicity at an earlier time point require further development. Identifying risks is the first and perhaps the most important step in the risk management process. If there is a failure to identify any particular risk, then other steps in the risk management process cannot be implemented for that risk. The concept of 'actionable' and/or mechanistic biomarkers refers to biomarkers that are embedded or rooted in disease pathogenesis (pathogenetic pathways). Thus, mechanistic/actionable biomarkers are more useful as predictive and/or pharmacodynamic (PD) biomarkers than as descriptive biomarkers that are byproducts of the disease process with limited value of the diagnostic and prognostic information. Actionable biomarkers in the diagnosis and management of diseases. Thus, circulating actionable and mechanistic/actionable biomarkers in the diagnosis and management of diseases.

An ideal biomarker in this context should meet the baseline criteria (superior tissue specificity, early time of release, a mechanistic association, and increased sensitivity with respect to bioanalysis).³²³ In addition, circulating biomarkers of cardiotoxicity should be able to distinguish myocardial from skeletal muscle or other tissue damage, such as cancer cachexia-induced damage of skeletal muscle. MicroRNAs (miRNAs) are small, non-coding RNAs of approximately 22 bp in length that post-transcriptionally regulate gene expression by binding and inhibiting particular mRNA targets.³²⁴ Due to the signature profiles of their tissue specificity and disease expression, miRNAs are being extensively explored or profiled for use as disease biomarkers, including drug-induced cardiotoxicity.^{224,324} Several circulating miRNAs (miR-1, miR-133a, miR-499, miR-208, and miR-423-5p) are promising cardiac injury biomarkers in



cardiovascular diseases.³²⁴ miRNAs exhibit two significant characteristics: (1) they are secreted from the producing cells, and (2) they can deliver gene-silencing signals between living cells in vitro and in vivo.³²⁵ The secretory mechanism of extracellular miRNAs has been explored, and the release of miRNAs into the circulation or body fluids is actively controlled through a ceramide-dependent machinery that is associated with the secretion of small membrane vesicles called exosomes or microvesicles as a versatile communication tool.³²⁶ Thus, the specific miRNA profile of cardiac exosomes or microvesicles can also be utilized as a novel diagnostic tool for chemotherapy-induced cardiotoxicity.

Regardless of the tremendous efforts made to discover novel biomarkers for clinical use, it remains unlikely that a single biomarker will be able to predict all drug-induced cardiotoxicities; rather, an optimal panel of biomarkers that reflects multiple aspects should be developed as a multiplex test for use in the laboratory and the clinic. ²¹¹ Further innovation is needed for the development of new noninvasive and cost-effective diagnostic biomarkers or modalities to manage the early detection of drug-induced myocardial injury. Compared to traditional approaches, an integrated mechanism-informed approach (advanced molecular and cellular cardiac imaging, ultrasensitive detection of subtle electrophysiological phenotypes, circulating actionable or mechanistic biomarkers) may allow a more sensitive and specific identification of in vivo cardiotoxicity and offer a better understanding of the fundamental mechanisms for individual compounds. Identifying the early signs of cardiotoxicity would certainly be beneficial to the management of oncologic patients and become essential for identifying patients who are at risk of irreversible heart disease and for monitoring treatment outcomes. Such approaches involve the assessment of cardiac function in the broadest sense.

4.3 | Novel therapeutic modalities in cancer versus cardiotoxicity

4.3.1 | Chronotherapeutic strategies using the concept of chronobiology

Biological processes and functions are organized in both space, as a physical anatomy, and time, as a biological time structure.³²⁷ The 24-hr cycle in physiological processes is known as the circadian rhythm in humans and many other living organisms. 328,329 Similarly, ultradian rhythm relates to hours, minutes, or seconds; infradian rhythms refer to those spanning days or months, and longer rhythms. 330 In general, the best-studied chronobiologic frequency is the circadian rhythm, and cell physiology and functions are regulated along the 24-hr time scale by the circadian timing system (CTS), which is composed of endogenous molecular clocks within each cell.³³¹ The suprachiasmatic nucleus in the brain acts as the central pacemaker (a central coordination system) for the human circadian system. $^{331-335}$ Chrono-the brain acts as the central pacemaker (a central coordination system) for the human circadian system. biology is the study of biological rhythms and the mechanisms of biological timekeeping, which is associated with the fields of medicine, pharmacology, and drug delivery.³²⁷ Chronotherapy addresses the use of circadian, ultradian, infradian, seasonal, or other rhythmic cycles in the application of therapy for various diseases³³⁰ and involves altering the timing of medication administration to improve the overall control of a disease and to minimize treatment side effects. This concept is emerging in the field of therapeutics.³³⁶ The initial idea in the development of chronotherapeutics was to synchronize the in vivo drug bioavailability with the rhythms of the diseases to optimize therapeutic outcomes and minimize side effects.³³⁰ Chronotherapeutics aim at improving treatment tolerability and efficacy through the adjustment of drug delivery to the CTS^{327,328} based on the endogenous biologic rhythms.³²⁹ With relevance to cancer, the CTS controls several critical molecular pathways of cancer processes and therapeutic effects over the 24 hours, including drug metabolism, cell cycle, apoptosis, and DNA damage repair mechanisms.³³⁷ The conventional concepts of drug delivery mostly consider constant release rates to maintain drug concentrations in the human body and achieve constant drug exposure at the sites of drug effects over time to optimize treatment efficacy, 338 regardless of the patient's physiological and biochemical conditions within the context of circadian rhythms.³²⁹ Drug delivery with such concepts (the homeostatic theory of drug delivery³³⁰) is being challenged with the advanced knowledge of the CTS; i.e., drug effects predictably vary not only as a function of dose but also as a function of administration timing. 328,337,339 If a drug release profile mimics the circadian rhythms of a living system, it may improve drug efficacy and reduce the toxicity of a specific drug administration schedule. 329 This hypothesis is in line with the main function of the CTS, which is to coordinate bodily and cellular functions, down to the main pathways that are responsible for drug pharmacokinetics (PK) and drug metabolism over 24 hours.^{328,340} For example, circadian timing can improve drug tolerability and/or efficacy up to several-fold in rodents and patients, regardless of administration routes.^{328,338} The tolerability of nearly 500 medications varies by up to five-fold according to circadian scheduling both in experimental models and patients.³⁴⁰ Thus, medication and treatments provided according to the body's circadian rhythms will result in better outcomes, and the relevance of timing may even exceed that of the dose.³⁴¹ Improved patient outcomes on circadian-based treatments (chronotherapy) have been demonstrated in randomized clinical trials, especially for cancer and inflammatory diseases.³⁴⁰

With advancements in the field of chronobiology, the circadian rhythms and their influence on biological systems have given rise to several mutually connected concepts beyond chronobiology, chronotherapy, and chronotherapeutics. 338,342 These concepts are extended to chronopharmacology, chronokinetics, chronodynamics, chronesthesy, chronotoxicology, and chronoprevention.^{327,338,343} Chronopharmacology is the study of the circadian dependencies in the PK and PD of drugs, PK-PD relationships, and their mechanisms. 327,328,342 Chronokinetics refers to dosing-time, i.e., rhythm-dependent differences in the absorption, distribution, metabolism, and elimination of medications.³²⁷ Chronodynamics refers to dosing-time, i.e., rhythm-dependent differences in the effects of medications. 327,344 Chronotoxicology is an aspect of chronodynamics that refers specifically to dosingtime, i.e., rhythm-dependent differences in the manifestation and severity of adverse effects and, thus, the intolerance of patients to medications. 327,340,345 Chronesthesy refers to rhythm-dependent differences in the sensitivity of target systems to medications that cannot be explained by corresponding administration-time differences in PK phenomena.^{327,340} The mechanisms of chronesthesy have yet to be elucidated^{327,340} but may represent rhythms in receptor number and conformation, second messenger dynamics, membrane permeability, or rate-limiting steps of metabolic pathways in drug-targeted tissues.³²⁷ Chronoprevention is the timing of medications or other interventions according to biological rhythm criteria as a means of averting disease or a decline in health status.³²⁷ The major difference of the goals between chronoprevention and chronotherapeutics is that chronoprevention focuses on the avoidance of disease, pathology, and other deleterious phenomena that compromise heath, while chronotherapeutics are the management or reversal of existing acute or chronic medical conditions.

4.3.2 Chronotherapeutic strategies in the context of precision oncology and cardiotoxicity/toxicity

Cancer is considered a chronotherapeutic disorder 332,346; thus, cancer can be a driver for system chronotherapeutics.³⁴⁰ The chronotherapy principle has been explored in the context of cancer therapies, and the results showed that circadian timing largely modifies the extent of toxicity of 40 anticancer drugs among agents in all pharmacological classes in rodents³²⁸ and humans.^{338,347} Doxorubicin (DOX), cisplatin, 5-fluorouracil (5-FU) and 5-fluoro-2'-deoxyuridine, irinotecan and oxaliplatin are representative anticancer agents that have been studied in the context of their circadian chronopharmacodynamics and chronotoxicologies. 327,348 The results of both laboratory animal and multicenter trials clearly show that the proper timing of cancer medications improves both patient tolerance to therapy and clinical outcomes.^{327,340} For instance, the cytotoxic effect of 5-FU was minimal for a circadian delivery peaking at 4 a.m. and maximal for a continuous infusion or a circadian pattern peaking at 4 p.m.^{328,349–351} The existence of a cancer-specific molecular clock can be used for the discovery and development of novel, therapeutic approaches to treat cancer.³⁵² A pharmacological modulation of clock-related proteins may be a suitable strategy for the identification of innovative anticancer approaches.³⁵³ To date, chronotherapy has been studied for some conventional anticancer agents, while data for MTT class and immunotherapies seem to be lacking. In line with the beneficial effects of chronotherapy, there is a critical and urgent need to prevent MTT and immunotherapy-induced cardiotoxicity of cancer patients who are also using chronopharmaceutics,³³⁸ which may potentially improve the safety, efficacy, and patient compliance of the new generation of anticancer drugs or modalities. Additionally, CTS robustness and phase varies among cancer patients, resulting in significant variabilities in response to chronotherapy³⁴⁰ and indicating that the development of personalized chronotherapeutics is highly expected through interdisciplinary systems approaches.³³⁷ Personalizing cancer chronotherapeutics requires an extensive molecular knowledge of anticancer chronopharmacology and chronotoxicity, both in healthy and tumor

tissues.³⁴⁰ Tumor tissues usually display a disrupted circadian organization relative to the circadian synchronization (molecular clock) in healthy tissues,^{327,348} although the molecular basis of this difference remains unclear.³⁴⁰ Moreover, circadian disruption has been observed in up to 50% of patients with metastatic cancer, and such disruption was associated with poor outcomes.³³⁷ Conceivably, the differential molecular clock between cancer and normal tissues can be exploited in treatment timing to specifically shield healthy cells while targeting cancer cells.^{340,354,355} Indeed, a combination of mathematical, statistical, technological, experimental, and clinical expertise is now shaping the development of dedicated delivery algorithms that enable treatment individualization (patient-tailored chronotherapies).³³⁰

The concept of chronotherapy offers further potential for improving current cancer treatments and optimizing the development of new anticancer or supportive agents. ³²⁸ Clock modulators or mediators are often considered natural candidates for a chronotherapeutic approach. ³⁵² One important mediator of circadian activity is the hormone melatonin (MEL), which peaks at the end of dark periods in both diurnal and nocturnal mammals. ³⁵⁶ Therefore, MEL has been used to mimic the dark period in humans as a means of treating sleep disorders and jetlag. ³⁵⁷. The determination of the circadian phase in individual patients through monitoring relevant CTS biomarkers could drive personalized chronotherapy and improve treatment effects, ³³⁷ with particular significance in cancer therapies. ³⁵⁸ CTS biomarkers mainly involve circulating MEL, cortisol, body temperature, and rest–activity rhythm. ³³¹ For example, DOX should be given mid-morning when circulating MEL levels are low to further reduce cardiotoxicity from DOX treatment. ³⁵⁹ Mitochondria are a common target for both MEL and many anticancer agents. ¹⁰⁷ The heart is particularly susceptible to oxidative damage induced by anticancer agents because it is abundant in mitochondria, ¹⁰⁷ which are both sources and targets for ROS. ³⁶⁰ Furthermore, the heart has an elevated rate of oxygen consumption and limited antioxidant defense systems when compared with other tissues. Thus abnormalities in mitochondrial functions may be the primary causative factors in the pathogenesis of anticancer drug-induced cardiotoxicity, ^{107,361}

In view of the abovementioned information, the general clinical approach to attenuate anticancer drug-induced cardiotoxicity is to utilize natural antioxidants such as MEL. The dark hormone holds amphiphilic properties. The term "amphiphilic properties" refers to a chemical compound possessing both hydrophilic and lipophilic properties. MEL has shed light on this therapeutic avenue with being dually oncostatic and cardioprotective. 107 Another appealing and unique property of MEL, which other antioxidants do not possess, is that its metabolites also exhibit antioxidant activity by scavenging ROS and reactive nitrogen species (RNS). 107 In addition, MEL concentrations in subcellular compartments (cell membrane, cytosol, nucleus, and mitochondria) fluctuate independently of the circadian rhythm during a 24-hr period. 107 From a therapeutic perspective, clinicians should bear in mind that the subcellular levels of MEL are controlled by regulatory mechanisms; therefore, MEL has low toxicity when administered at high doses.³⁶² MEL is the first known natural antioxidant molecule that may have both curative and palliative actions in the treatment of human neoplasms. 363 To date, studies regarding the direct effect of MEL on the mitochondrial respiratory chain and complexes in relation to anticancer agent-induced cardiotoxicity, including MTT, seem to be lacking. 107,364 This crosstalk between MEL and the mitochondria is quite important as it can enable to overcome these pharmacological hurdles for greater clinical impact. Because almost all clinically used antitumor drugs exhibit toxic side effects that affect heart function, 365 the discovery of natural phytochemicals as chronopharmaceuticals for clinical use becomes more and more important. Aside from MEL, other natural phytochemicals such as coenzyme Q10, L-carnitine, resveratrol, curcumin, and ginkgo biloba also counteract the cardiotoxic side effects of cancer chemotherapy. 365 Thus, co-administration of these natural cardioprotective agents seems promising to improve the clinical outcomes of cardiotoxicity.

In light of emerging chronotherapeutic approaches, ³⁶⁶ various chronoprogrammable drug delivery systems have been developed. ^{329,339,367–381} These approaches include chronomodulating infusion pumps, controlled-release microchip strategies, floating pulsatile systems, nanotechnology, press-coating approaches, micro-electrochemical systems, osmotic pressure, microchips, liposomes, thermosensitive hydrogels, micro- and nanocarriers, and microparticle-based systems. The development of programmable time pumps has enabled the safe and highly effective delivery of combination chronotherapy protocols. ³³⁷ Unfortunately, no current drug delivery system can satisfy all the requirements of chronotherapeutics. ³³⁰ From the applied and clinical perspectives, one of the most important issues is

to consider the ease of manufacturing and cost-effectiveness during the selection of the appropriate chronopharmaceutical technology.

5 | RESEARCH GAPS, CHALLENGES, PERSPECTIVES, AND OPPORTUNITIES TO REDUCE MTT-INDUCED CARDIOTOXICITY

5.1 | The gaps in cross-disciplinary research

The lack of a mechanism-based curative treatment of cardiotoxicity for many compounds is a treatment concern in the clinic, and only general cardioprotective care or discontinuing cancer therapy (premature discontinuation of chemotherapy) is currently applied to the patients. Each molecular drug or therapeutic modality may have a unique biological mechanism of action because each drug shows a unique toxicity profile regardless of whether the drugs are in the same class and exhibit structural similarity, 382 This feature is also supported by the observation that transcriptome-wide response profiles in cardiomyocytes using a cohort of KIs showed a limited overlap.³⁸³ Therefore, general cardioprotective strategies (ACE inhibitors, beta-blockers, digitalis, etc.) may not work effectively. Disease-specific mechanisms must be addressed in preclinical research (basic and translational) for a mechanistic understanding of the cardiotoxicity induced by novel anticancer therapies. Drug-induced cardiac dysfunction and cardiomyopathies reflect serious, clinically adverse effects of drug actions, and they can be used in experimental models to study the pathogenic mechanisms of these cardiac disorders, offering the advantage of precise control of the onset time, and can often be studied in a longitudinal fashion.³⁸⁴ The gaps currently observed between clinical observations and pharmacological innovation might be reduced by a better application of the concept of reverse translational research with a bidirectional research paradigm - from practice (bedside) to research (benchside) - and a back flow of information. This research paradigm aims to transfer clinical insights or observations into hypotheses that can be investigated in the basic research laboratory and that can subsequently inform clinical practice. With such a bidirectional research paradigm, clinical cardiotoxicity becomes an important source of observations and ideas to feed into fundamental research for further mechanistic insights. In turn, these new mechanistic insights allow proposing novel concepts regarding these drugs and accelerating clinical advances in knowledge to further bridge knowledge gaps. Such a mechanistic approach should yield disease-onset, mechanism-based biomarkers that identify the initial disease stages. These markers are valuable tools for developing strategies to prevent the progression of diseases that can be translated into clinic for diagnosis and to measure disease progress and/or the effects of treatment. In line with the expectations, using various modeling systems (in vitro, in vivo, computational biology or their combination) to explore the underlying mechanisms that drive the cardiotoxicity is a high-priority research area in preclinical research.

The development of new PD, prognostic, and surrogate biomarkers not only enhances the efficiency of cancer treatment (e.g., targeting neoantigens) but also mitigates the risk of cardiotoxicity. Thus, this development is urgently needed in this era of precision oncology. According to the hierarchical tree of various sources of biomarkers in nature (genes and genetic variants, RNA, microRNA, proteins, and metabolites), some caveats should be kept in mind: (1) each category of markers is suited to a specific purpose, (2) caution should be taken regarding cross-species translatability, and (3) biomarker values fluctuate in response to the pathophysiology of the organism over time. Taken together, genomic and transcriptomic markers can provide information about a person's risk of developing a specific disorder or how a patient may respond to treatment. Proteins and metabolites are more dynamic in a disease course and thus carry more diagnostic information than DNA or RNA. It is important to note that DNA, RNA, and proteins are more species-dependent; thus, these biomarkers should be best identified and characterized using human materials. In contrast, small molecules or metabolites are less species-dependent, 385,386 and miRNAs are also well conserved among species, ranging from worms to humans; 324,387 therefore, many of these markers from animal studies can be translated into clinical practice. 324,388 Metabolic biomarkers are subtly sensitive indicators of health status that provide early profiles on drug efficacy, toxicity, and mechanism. 45,389 miRNAs are sensitive and specific toxicity reporters as



well.^{224,390} Considering the sensitivity and interspecies translatability, identifying metabolic biomarkers, especially metabolic imaging biomarkers and/or circulating/tissue miRNA markers, should be prioritized in preclinical studies using model organisms, allowing a direct comparison of animal models with human studies. Altered ionic homeostasis is considered the foremost change in the early phase of responses of myocardium to environmental toxicants or drugs.⁴⁵ Preclinical research on the molecular basis of cardiac ion channels in relation to regulation and drug sensitivity of the cardiac ionic currents should be performed.

5.2 | Modeling systems for clinical cardiotoxicity

5.2.1 Overview of living modeling systems

The next step is to select a proper modeling system for predicting clinical cardiotoxicity as precisely as possible. A proper model should encompass adequate sensitivity, predictability (similarity to human myocardial biology), measurable functional parameters of cardiac dynamics/performance, real-time data acquisition over time, and high throughput, all of which should be considered collectively. For example, in vitro assays that use primary animal cardiomyocytes, human-induced pluripotent stem cell (hiPSC)-derived cardiomyocytes, hiPSC-derived endothelial cells, and hiPSC-derived cardiac fibroblasts are primarily designed as electrophysiological models to assess the interaction of drug candidates with the main ion channels that are involved in maintaining the cardiac action potential (electrophysiology), ^{391,392} contractility, ³⁹² kinase phosphorylation profiling ³⁹² and more. Complementary to these in vitro assays, mathematical or computational approaches can model the propagation of the action potential and ion concentration dynamics of the heart.³⁹³ Recently, the xCELLigence RTCA Cardio System was developed to monitor cardiac contractility and arrhythmogenic properties in vitro based on impedance measurement. This technology can be used for a high-throughput screening of functional cardiotoxicity. ³⁹¹ However, many in vitro models may lack or incompletely recapitulate important aspects of human biology in vivo, 8,394 allowing certain toxicities to escape detection. Cardiac performance is the result of an entire functioning organ, including electrophysiological function (rhythms or heartbeats), pump function (cardiac output, stroke volume, stroke work, ejection fraction) and mechanical muscle function (force velocity curve, maximum velocity of shortening (Vmax), LVEF, ventricular contractility (dP/dt)). Although ex vivo experiments using isolated hearts from live animals (e.g., the Langendorff heart assay) can be used to assess cardiac performance, these experiments present technical challenges because they are labor intensive and not amenable to a high-throughput analysis. However, addressing in vivo cardiac effects of MTT necessitates a suitable animal model. Thus, it is important to select the most appropriate animal model for mechanistic studies and/or as a predictive preclinical model.

5.2.2 Rodent model

In biomedical sciences, *Mus musculus* is the most commonly used species for animal models due to its genetic, physiological, and anatomical similarities to the human system in general. However, the expenses associated with the use of this species limit its application in large-scale molecular and/or therapeutic screening or modeling. To date, using a mouse model solely for drug screening (i.e., a single factor experimental study) is very rare, while multi-factorial experiments have often been performed, for example, simultaneous evaluation of antitumor efficacy and cardiotoxicity of cancer drugs, ^{395–406} and/or studies of cardioprotective strategies after using cancer drugs. ^{407–412} More importantly, many drugs, including oncologic pharmaceuticals, can often lead to cardiotoxic electrophysiological effects (e.g., QT prolongation, atrioventricular conduction blocks, and ventricular arrhythmias including torsades do pointes); thus, adverse electrophysiological effects represent important phenotypes of drug-induced cardiotoxicity. ^{75,391,413} In this regard, the intrinsic species-specific differences of cardiac electrophysiology require attention during model selection. The cardiac electrophysiological properties in mice are significantly different from their human counterpart. ⁴¹⁴ Furthermore, mice are nocturnal animals; therefore, this CTS of this species is mismatched with the human counterpart from the perspective of chronobiology and chronopharmacology. These practical concerns may lead one to consider mice as a second choice model organism for studying drug-induced cardiotoxicity.

5.2.3 | Zebrafish model

Zebrafish have filled a niche in the phylogenetic gap between invertebrates and mammals as one of the most successful vertebrate models for studying human physiology and diseases. 415 Approximately 70% of human genes have at least one obvious zebrafish orthologue and mutations in homologous genes lead to similar phenotypes. 416 The genome of zebrafish is well conserved, and its physiology shows good resemblance to mammals in general. Additionally, most zebrafish organs perform the same functions as their human counterparts. 417 Regarding the cardiovascular system, zebrafish stand out for their highly conserved integrative physiology of the cardiovascular system and a pharmacological response similar to that of human beings. 417,418 The key morphological, functional, mechanobiological, electrophysiological, metabolic, and molecular profiles, as well as many cardiac events, overlap with those in humans, 418-422 including the kinome profiles, where most kinase inhibitors interact with the kinases. 423,424 Moreover, the cardiac electrophysiological properties of the hearts of zebrafish larvae and adults resemble those of humans in many aspects. 413,425-428 It is of particular importance to evaluate the pharmacological response of cardiac function in an animal model relevant to the human heart for an appropriate assessment of the safety (drug-induced cardiotoxicity) and efficacy of drugs that target cardiovascular diseases (cardiopharmacology). Many human cardiovascular drugs have identical effects on zebrafish physiology, and numerous human cardiovascular disorders have been recapitulated in zebrafish genetic models. 417,429 Importantly, many drugs that cause QT prolongation in humans consistently cause bradycardia and AV block in zebrafish, 413 suggesting that zebrafish are a rational, predictive model for cardiac safety liability of chemicals/drugs and also a valued, mechanistic model of human cardiovascular diseases. The overall physiology, genetics, and cardiovascular pharmacology similarities strongly support the use of zebrafish as the most reasonable approach for modeling human cardiovascular diseases or events in vivo, $^{420,428,430-433}$ monitoring drug-induced in $vivo\ cardiotoxicity, ^{413,417,418,434-436}\ and\ in\ vivo\ cardiovascular\ drug\ discovery. ^{429}\ Furthermore, zebrafish\ are\ a\ diurnal$ vertebrate; 437 thus, the zebrafish CTS is expected to be compatible with the human counterpart for studies of chronobiology and chronopharmacology. 437-454 Recently, zebrafish have been increasingly recognized as a model for circadian rhythm disorder. 455 Lastly, the general advantages of the zebrafish model, 456 e.g., convenience of drug delivery, rapid development, ease of genetic manipulation, low cost of maintenance relative to other model animals, high fecundity, transparency, small size, and the availability of various transgenic lines, make this animal model an almost ideal complement to rodent-based biomedical research. This organism has been considered the "new mice" to replace the so-called "higher vertebrates" (e.g., mice, rats) according to the 3R (replacement, reduction, and refinement) perspective for animals used for experimentation. 456-458 Research in exploring the potential use of zebrafish as a mechanistic model organism to study cancer treatment-induced cardiomyopathy and myocardial dysfunction or to exploit the predictability of clinical cardiotoxicity, particularly for MTT, should be highly promoted. To date, extensive studies of the mechanisms involved in relation to MTT have not yet been undertaken, and published data in the preclinical setting using zebrafish remain highly limited.⁴³⁴ A research program is being started in our laboratory using zebrafish as a predictive and mechanistic model to address clinical cardiotoxicity induced by MTT to identify potential mechanistic biomarkers.

Regardless of the aforementioned merits, several important caveats in modeling kinase inhibitors on a zebrafish model should be considered: (1) Confounding effects: transgenic fluorescent animals are genetically modified, and the confounding effects of an inbred strain background may harbor compensatory mechanisms that can alter biological events. 459 Thus, any data obtained from transgenic lines must be further confirmed by wild-type fish lines. (2) Cardiac regeneration: the zebrafish heart maintains the ability to regenerate throughout adulthood with scar-free regeneration of a damaged heart. In contrast, the adult mammalian heart exhibits a limited signaling powerhouse and cell reservoir for regeneration. 460,461 The limited cardiac regenerative capacity in humans is witnessed by scar formation following myocardial infarction. (3) The human kinome shares many catalytic domains of zebrafish homologs, 423,462 which is important for tyrosine kinases. 462 However, small changes in the sequence of amino acids in a protein may reduce or enhance certain inhibitor interactions, consequently leading to under- or over-estimation of toxicity. 423 To extend zebrafish utility as a functional model organism for toxicity and efficacy in studying kinase inhibitors, more knowledge of these relationships may be required. (4) Phenylthiourea (PTU) is a well-known inhibitor of tyrosinase

is widely used in zebrafish research to suppress pigmentation in the developing embryos/larvae of small animals. ⁴⁶⁴ In modeling the toxicity of kinase inhibitors on zebrafish, PTU is not recommended to avoid potential synergistic effects from PTU and a test drug. (5) Pseudokinases are generally less conserved between zebrafish and humans. ⁴²³ Thus, it is unsuitable to model pseudokinases in a zebrafish model. (6) The drug exposure method for zebrafish is mainly bathing, and caution should be taken with insoluble drugs and drug metabolism. ^{465,466} Therapeutic antibodies are large therapeutic molecules that differ from small molecule agents in absorption, distribution, and elimination properties between the two classes of drugs in a systematic manner. ⁴⁶⁷ (1) Absorption: the absorption of small and large molecules differs with respect to their common extravascular routes of delivery (oral for small molecule agents versus injection for antibodies). (2) Elimination: small molecule agents are commonly distributed into the tissues but are eliminated primarily by liver metabolism. Meanwhile, catabolism and target-mediated drug disposition are unique features of antibody distribution and elimination. ⁴⁶⁸ The above knowledge must be considered when testing humanized antibodies in animals.

5.3 | Challenges, perspectives, and opportunities

5.3.1 Kinase inhibitors versus "precision oncology" on efficacy and cardiotoxicity/toxicity

Introduction of the "MTT for cancer" concept in the past 15 years has led to optimistic expectations that we can precisely target the molecular underpinnings or molecular drivers of a patient's disease, and this concept is hailed as a revolutionizing approach in the treatment of cancer.⁴⁶⁹ Most impressively, the concept has served as the foundation for personalized medicine or precision oncology/medicine to guide the selection of treatments in the clinic.⁴⁷⁰ Precision oncology aims to deliver the right therapy to the right patient at the right time through promising identification of genetic alterations in human cancer and either the signaling pathways or specific biological processes that are essential for the development and progression of tumors, followed by therapy targeted to a patient's unique genetic or other relevant characteristics. 471-479 Many kinase inhibitors that target oncogenic mutations in protein kinases have been developed under such theoretical expectations and perspectives. 115,480 Consequently, most kinase inhibitors that are currently developed and clinically used largely focus on anticancer effects. In real-world practice, an individually tailored cancer treatment is determined or guided by specific molecular biomarkers based on a genetic understanding of their diseases (biomarker-informed treatment).^{2,481} These biomarkers are somatic genomic alterations (mutated, amplified, or overexpressed), often called "driver mutations," and are considered responsible for the initiation and progression of cancer. Thus, these "driver mutations" are optimal biomarkers for selecting patients with targeted therapies. 481 Because mutations and dysregulation of protein kinases play causal roles in cancers, kinase inhibitors have become the largest class of new anticancer drugs, serving as the cornerstone of the development of molecularly targeted therapeutics to date, 2,109,472 although the effectiveness/potency and potential cardiotoxicity/toxicity of those very recently approved therapeutics remains unclear.² In light of the promise of precision oncology and based on the concept of MTT over the past 15 years, many concerns and challenges have been raised in the real world, and the medical, pharmacological, and research communities must face those challenges. First, clinically, intrinsic (primary), and acquired (secondary) resistance to targeted agents has emerged as a primary challenge. 471,482,483 This resistance is typically reflected by the limited effectiveness of targeted therapies on long-term clinical benefits. Almost all patients acquire resistance in less than two years from therapy initiation, and a small subset of patients (10-20%) simply fail to respond at all, demonstrating intrinsic (primary) resistance. 116,471,482,483 This resistance is because of alternative oncogenic pathways taking over and/or other mechanisms (e.g., bypass pathways), leading to the occurrence of drugresistant variants. 101,471,482,484-489 Cancer cells may become drug resistant much more rapidly than would be predicted from the rates of conventional mutation 116 by differential utilization of genomic and epigenetic strategies at various levels through point mutations, deletions, translocations, amplifications, altered microRNA levels, and epigenetic anomalies (epimutations). 134,490,491 Therefore, irreversible inhibitors may also need to be combined with other targeted agents and/or chemotherapy to address drug resistance. 101 In addition, secondary mutations in the ATPbinding site, such as threonine to methionine at position 790, serve as a mechanism of resistance to ATP-competitive kinase inhibitors.⁴⁹² The complex and adaptive nature of most tumors frustrates the optimistic predictions of MTT for cancer.⁴⁹³ Second, most of these PKIs prolong survival in cancer patients only weeks or months longer than standard cytotoxic therapies.¹⁰⁹ In other words, generally targeted therapies by inhibiting protein kinases or other molecular events are usually not curative, even when combined with other chemotherapy or radiotherapy.^{101,494} Third, the number of patients who gained clinical benefits from genomic profiling or the analysis of their tumor genomes followed using MTTs against those "driver" mutations is currently quite small,^{470,495} and responsive patients may be designated "exceptional responders" for largely unexplored reasons.^{494,496–500} To this end, inhibiting tumor growth through targeting defects in cancer genetics does not seem working meaningfully, while life-threatening complications such as cardiotoxicity are unavoidable, uncontrollable, and unpredictable. Fourth, the majority of cardiotoxicities induced by MTTs are unexpectedly associated with the second- and third-generation kinase inhibitors;⁵⁰¹ a new face on an old problem. Fifth, the paradigm of precision oncology remains a theoretical promise or hypothesis rather than a real-time reality in many areas of clinical oncology.^{479,495,502}

5.3.2 Kinase inhibitors versus cancer driver or actionable mutations on efficacy and cardiotoxicity/toxicity

Kinase inhibitors that are designed to target cancer "driver" mutations, which are identified by cancer genetic profiling, are being challenged for their clinical endpoint—i.e., the clinical beneficial effects from targeted agents are not generalizable to a large patient population. 101,109,471,482,483,494 First, biologically, the "driver"-centric paradigm on tumorigenesis is being challenged not only by the clinical reality but also by a novel paradigm. 503-506 The novel paradigm envisions that those traditionally considered "passenger" mutations (i.e., those mutations with no roles in carcinogenesis) affect the course of cancer progression. 504-506 Further, cancer evolution and progression are a balance of "driver" and "passenger" mutations, rather than being solely determined by "driver" mutations. 505,506 In line with the novel paradigm, the role of a targeted therapeutic approach is to interfere with an existing balance of "driver" and "passenger" mutations of cancer and then to develop a new interaction network to build a new balance of "drivers" and "passengers". Such an iterative cycle can occur through an entire treatment course with different kinase inhibitors or combined with other categories of anticancer drugs in real time. As observed in the clinic, most tumors can escape from the inhibition of any single kinase, as manifested as a partial and/or non-durable response with these therapeutics, and multiple target inhibitors are necessary to sort the effects. 101,153,483,507,508 The balance between "drivers" and "passengers" reflects the clinical reality, and this novel paradigm may encourage the clinical and pharmaceutical communities to rethink and refine the current concept of MTT under the umbrella of precision oncology, with respect to the pharmaceutical innovation against the effects on the clinical endpoint. Second, the emerging concept of "actionable mutations", defined as those mutations that are potentially responsive to a targeted therapy (actionability), 509,510 remains a huge challenge with respect to the identification of such targetable mutations. 494,509 From the perspective of onco-pharmacology, the most significant success has been achieved in oncology by targeting mutationally activated "oncogenic" driver kinases, including Bcr-Abl, EGFR, c-Kit, PDGFR, ALK, and b-RAF (Tables 1 and 2). 105,469 To date, only a small subset of the human kinome has been studied, ¹⁰⁴ and many potential candidate kinases for cancer therapies remain unexplored. 469,511 Consequently, the cardiac safety liabilities of those potential future kinase inhibitors in the clinical endpoint are unknown. Third, with more than 518 protein kinases in the human kinome, 108 including the cardiac kinome, 512 it remains largely ambiguous in relation to the target specificity or selectivity of individual kinases and their biological roles in the cells or tissue-specific contexts, and most of the identified kinases remain largely uncharacterized or unexplored for their functions and tissue expressions, 513 including the majority of kinases expressed in the heart. 514 Many molecular mechanisms relevant to cardiac disease are also relevant to tumor biology, suggesting that cancer and cardiovascular disease have a shared biological mechanism.⁵¹⁵ Indeed, numerous overlapping signaling pathways that drive tumorigenesis are also essential for normal cardiac function, 3.7,512,516 Unfortunately, these molecular signaling pathways are often targeted in molecularly targeted cancer therapies. 8,517 Fourth, promiscuous targeting is a unique feature of ATP-competitive kinase inhibitors, and the potential is greater than almost any other type of drugs regardless of multi-targeted or highly specific targeted kinase inhibitors. 50,153,518 Multi-targeted agents are frequently more efficacious than selective agents, ¹¹⁶ and partial inhibition of a small number of targets can be more efficient than the complete inhibition of a single target. ⁵¹⁹ However, a major concern using combination approaches is the possibility of more uncontrollable and unpredictable toxicities on multiple organs/tissues than that observed with single target approaches. Therefore, target selectivity/specificity represents the biggest challenge for drug design and the development of kinase inhibitors^{116,483} because most inhibitors interact with the highly conserved ATP-binding sites of kinases. Additionally, developing pharmacological agents that target only one of the more than 500 kinases present in humans remains a formidable challenge. ¹⁶⁴

5.3.3 | Effects of kinase inhibitors on the immune system and concerns for autoimmune cardiovascular diseases

Kinases play pivotal roles in tumor immunity and tumor immune evasion; thus, they could serve as relevant therapeutic intervention points far beyond various antibody-based therapies. 520 However, it is important that a kinase inhibitor does not co-target components of the immune system that are essential for mounting an immune response. 101 Unfortunately, some drugs may bind directly and reversibly to immune receptors, e.g., major histocompatibility complex or TCR, thereby stimulating the cells in a manner similar to a pharmacological activation of other receptors. 521 This concept has been termed "pharmacological interaction with immune receptors" or the "p-i concept". 521-523 In line with the importance of kinases in the regulation of an immune response as an intrinsic defense mechanism, 524 many oncogenic $signaling\ pathways\ in\ tumor\ cells, such\ as\ MAPK\ or\ PI3K/AKT/mTOR, are\ also\ critical\ in\ the\ regulation\ of\ immune\ cells,$ and those oncogenic signaling pathways are often targeted by anticancer kinase inhibitors.⁵²⁵ Thus, many targeted agents might have "off-target" effects (either beneficial or detrimental) on immune cells beyond their intended effects on the respective signaling pathways in cancer cells. 526,527 Immunomodulatory effects of kinase inhibitors can act indi $vidually \ on \ DCs, effector \ T cells, and immunosuppressive cells \ or their combination. \\ ^{528} \ Indeed, such effects \ are induced \ on \ DCs, effector \ T cells, and immunosuppressive cells or their combination. \\ ^{528} \ Indeed, such effects \ are induced \ on \ DCs, effector \ T cells, and immunosuppressive cells or their combination. \\ ^{528} \ Indeed, such effects \ are induced \ on \ DCs, effector \ T cells, and immunosuppressive cells or their combination. \\ ^{528} \ Indeed, such effects \ are induced \ on \ DCs, effector \ T cells, and immunosuppressive cells or their combination. \\ ^{528} \ Indeed, such effects \ are induced \ on \ DCs, effector \ T cells, and immunosuppressive cells or their combination. \\ ^{528} \ Indeed, such effects \ are induced \ on \ DCs, effector \ T cells, and immunosuppressive cells or their combination. \\ ^{528} \ Indeed, such effects \ are induced \ on \ DCs, effector \ T cells, and immunosuppressive cells or their combination. \\ ^{528} \ Indeed, such effects \ on \ DCs, effector \ T cells, and immunosuppressive cells or their combination. \\ ^{528} \ Indeed, such effects \ on \ DCs, effector \ T cells, and immunosuppressive cells or their combination. \\ ^{528} \ Indeed, such effects \ On \ DCs, effector \ T cells, and immunosuppressive cells or their combination. \\ ^{528} \ Indeed, such effects \ On \ DCs, effector \ T cells, and immunosuppressive cells or their cells of \ On \ DCs, effector \ T cells, and immunosuppressive cells or \ On \ DCs, effector \ T cells, and immunosuppressive cells or \ On \ DCs, effector \ T cells, and \ On \ DCs, effector \ T cells, and \ On \ DCs, effector \ T cells \ On \ DCs, effector \ T cells \ On \ DCs, effector \$ by some FDA-approved kinase inhibitors, e.g., temsirolimus, sorafenib, sunitinib, imatinib, and dasatinib. 101,525,529 The secondary pharmacological interaction of kinase inhibitors with the immune system have yet to be studied in detail in cancer patients, 525 and the exact in vivo mechanisms remain to be further clarified in human studies. This crosstalk may open new possibilities for using either a stimulatory or inhibitory function of the immune system for defined targeting.⁵²¹ The consequences of these kinase inhibitors on the functionality of immune effector cells alter immune cell infiltration (immune subset conditioning), increase the frequency and function of effector immune elements, and decrease the number and function of immune suppressor cells.^{528,529} Although the regulation of immune cell signaling with kinase inhibitors has produced robust evidence, whether autoimmune cardiovascular diseases can be induced by the pathway when patients are treated with kinase inhibitors remains unknown. Further studies are warranted to determine the causal direction of this relationship.

5.3.4 | Targeting non-coding cancer drivers by kinase inhibitors and concerns of cardiac safety liabilities

Cancer may arise from the accumulation of multiple driver mutations.⁵³⁰ However, cancers harbor a large number of molecular alterations, and targeting one or some of the many molecular alterations to achieve a clinically significant, sustained effect might not be realistic for most tumors.⁴⁷⁹ In addition, the overwhelming majority of oncogenic variants, both somatic and germline, occur in non-coding portions of the genome⁵³¹ as opposed to protein-coding regions, which have been incorporated into the paradigm of precision oncology in the clinic. The concept of MTT developed to target protein-coding cancer drivers is being challenged by the emerging paradigm of non-coding cancer drivers.^{531–533} The non-protein-coding cancer genome remains widely unexplored,⁵³² but this field will have a profound impact on many disciplines beyond cancer research. For example, such genetic events noticed in cancer are also observed in autoimmune disorders, in which approximately 90% of the causal variants are non-coding, and most map to immune cell enhancer regions.⁵³⁴ Given the central role in cellular signaling of kinases in the heart and many other organs aside from cancers, accumulating pharmacological and pathological evidence has revealed that kinases are also

promising drug targets for numerous non-oncological indications. ^{101,535-540} The successful approval of tofacitinib for the treatment of arthritis is a typical example, ⁵³⁶ although the exact mode of action of tofacitinib in the setting of autoimmune disease has yet to be unraveled. ⁵³⁷ The importance of cardiac safety liabilities with kinase inhibitors goes beyond cancer treatment. Insights gained from the clinical endpoints of oncology show that an optimization of the therapeutic equation (efficacy vs. toxicity) appears to be very hard to achieve in many cases with kinase inhibitors. ¹¹⁶ It is highly important to use robust systemic approaches, ^{154,541,542} including kinase panels ⁵⁴³ and chemical proteomics, ⁵⁴⁴ for understanding chemical interactions with biological systems, characterizing drug-induced molecular changes in affected cells and tissues of interest, and permitting kinome-wide analysis of candidate molecules. ¹¹⁶ These strategies may allow researchers to start with these theoretical approaches to initially identify virtual cardiotoxicity by focusing on pathways that are common to all or many cardiotoxicities. ^{154,545} Then, a multitude of data will help reach a better characterization of "specific" tumor or disease signaling pathways that are unique to cancer cells or diseases and do not significantly affect normal cells such as heart muscle.

6 | CARDIOTOXICITY INDUCED BY ICIS AND T-CELL THERAPY

Immune therapies represent an advance in the fight against cancer. In a broad sense, this field encompasses a number of treatment approaches that utilize distinct components of the immune system in the fight against cancer. ^{44,546} The main forms of immunotherapy strategies that are used or in active clinical development today include ICIs, therapeutic monoclonal antibodies, cancer treatment vaccines, immune system modulators, and immune cell therapy (including different forms of adoptive cell transfer (ACT), such as tumor-infiltrating lymphocytes (TILs) and CART). Currently at the forefront of immunotherapy are ICIs and CART. Here, we focus on ICIs and CART cell therapy and their potential to induce cardiotoxicity. To date, clinical cardiotoxicity induced by ICIs and CART seems to be less frequently reported compared with MTT and other categories of anticancer drugs. The clinical data remain limited because the approved ICIs are still limited, and CART has not yet been approved. Thus, serious side effects such as cardiotoxicity are still being documented. Nevertheless, clinical cardiotoxicity induced by ICIs^{12–28,546–550} and CART^{30–44} have been reported (Table 3).

In the physiological situation, immune checkpoint proteins limit the strength and duration of immune responses and normally act as a type of "off switch" that helps keep the T cells from attacking normal cells in the body,⁵⁵¹ which in turn reduces autoimmunity and promotes self-tolerance.⁵⁵²⁻⁵⁵⁴ Recently approved ICIs that target receptors that are involved in the immune escape of cancer cells, such as cytotoxic T lymphocyte-associated antigen-4 (CTLA-4), programmed cell death protein-1 (PD-1), and programmed cell death protein ligand-1 (PD-L1), are increasingly being used for therapeutic benefit in a number of cancers. 546 Thus far, the development of ICIs has focused on these major targets. 14 These therapeutic monoclonal antibodies interact with specific co-stimulatory or co-inhibitory molecules that are expressed on the surface of activated T cells and strengthen the immune response against cancer cells and minimize tumor evasion from host immunity.555,556 ICIs work by blocking inhibitory pathways of Tcell activation, leading to an immune response directed against tumors; thus, this activation represents a nonspecific immunologic activation that can lead to immune-related adverse events (IRAEs).⁵⁵⁷ To date, FDA-approved ICIs ^{26,27,547,548,550,558} include Ipilimumab (CTLA-4), Pembrolizumab (PD-1), Nivolumab (PD-1), Atezolizumab (PD-L1), Avelumab (PD-L1), and Durvalumab (PD-L1). Based on the working principles, ICI-induced toxicity typically involves autoimmune disorders, ^{28,54,555,557,559} including autoimmune myocarditis. ^{12–28,93,550} The importance of ICIs in the heart has been addressed in preclinical data, and the data give some mechanistic insights into the clinical observations of autoimmune myocarditis. 560,561 In addition, although ICIs are an attractive concept in the therapeutic development of cancer treatment, many cancer patients do not respond to treatments with ICIs, partly because of the lack of tumor-infiltrating effector T cells^{562–565} or primary or acquired resistance due to various mechanisms.⁵⁶⁵ The overall response rates against melanoma, bladder cancer, Hodgkin's lymphoma and non-small cell lung cancer using ICIs is approximately 30%, and complete response rates (eradication of a patient's tumors) are as low as 5%. 566 Recently, macrophage activation syndrome (MAS), a very severe, new complication of ICIs with



a 50% mortality rate, has been reported.⁵⁶⁷ More serious concerns have clearly been raised about the clinical use of ICIs.

Immune cell therapy (T-cell based therapy) consists of several forms, TILs, cytotoxic T lymphocytes (CTLs), transgenic T-cell receptor (tgTCR) T cells and CART. 196,568,569 CART is considered the best-in-class example that the genetic engineering of T cells can lead to deep and durable responses in CD19⁺ B cell malignancies.⁷³ The receptors allow the modified T cells to attach to specific proteins on the surface of cancer cells, and the modified T cells become activated and attack the cancer cells once bound.⁵⁷⁰ CD19 CART has demonstrated remarkable success in treating hematologic cancers, prominently including acute and chronic B cell leukemias.⁵⁷¹ Currently, a growing number of clinical trials have focused on solid tumors and targeted surface proteins including carcinoembryonic antigen, diganglioside GD2, mesothelin, interleukin 13 receptor α , human epidermal growth factor receptor 2, fibroblast activation protein, L1 cell adhesion molecule, 568,571 and melanoma-associated antigen 3.32 However, the clinical results in solid tumors have been much less encouraging, 71,570 Historically, three generations of CAR constructs have been developed, and the third-generation CARs are the most recent 196,572 and contain more than one additional co-stimulatory molecule. Recently, fourth-generation CAR T cells redirected for universal cytokine killing (TRUCK) have been described and are designed to express and release transgenic products that accumulate in the targeted tissue. 572-575 However, despite tremendous efforts to date CART targeting, CD19 remains the beststudied example^{205,570,576}; CD19 is a cell surface molecule on B cells and B cell malignancies. The FDA granted a 'breakthrough therapy' designation to anti-CD19 CART on 1 July 2014. 196,205 To date, the challenge for targeted CART continues to be the identification of suitable epitopes to ensure on-target specificity. 51,53,55,577 The identification of suitable epitopes to ensure on-target specificity. fication of truly tumor-specific antigens has become the greatest general challenge in cancer immunotherapy⁵⁷² because true tumor-specific antigens are rare, and many tumor-associated antigens are shared by both tumor and normal tissues, resulting in off-tumor, on-target toxicity.53,578 Cardiotoxicity caused by CART has been reported (Table 3),³⁰⁻⁴⁴ but the causes are diverse, and the mechanisms are not fully understood.^{30,31} In general, the unpredictable specificities of targeted tumor antigens and cytokine release syndrome are recognized as two major factors that are accountable for "on-target, off-tumor" toxicity in multiple organs, including cardiotoxicity.^{51,53,83,196,579–582} Conceptually, neoantigens are the antigens encoded by tumor-specific mutated genes (known as driver mutations) and specifically expressed in the tumor; thus, neoantigens are theoretically ideal targets for cancer immunotherapy because they are less likely to induce normal tissue toxicity (non-exclusive antigens).^{564,583} However, the identification of neoantigens depends on targeting "driver" mutations first, and many questions remain unanswered regarding how to precisely define and distinguish between "driver" mutations or clinically actionable mutations (responsive to targeted therapies) from the much larger set of passenger alterations embedded in tumor DNA. 494 The emerging concept of non-coding drivers adds additional challenges to precisely define or identify "neoantigens." In addition, the use of host lymphodepletion-chemotherapy with immunosuppressive agents (e.g., cyclophosphamide) is required to augment the antitumor effects of CART.⁵⁸⁴ Unfortunately, these concomitant therapies can lead to clinical cardiotoxicity.585-588

The recent experiences with severe, life-threatening episodes of cardiotoxicity associated with ICIs and CART gives rise to some important concerns that are biologically and clinically relevant for future preclinical studies, oncology trials, and clinical practice to limit the uncontrolled activation of immune responses. Immunotherapies have significant potential, yet there is room for further improvement. The immune system is species-specific; thus, immunotherapy-induced toxicity/cardiotoxicity may not be readily predictable in animal models because of the differences in both gene expression and amino acid sequences of peptides derived from homologous proteins. The insights derived from animal models are limited by significant functional differences of the cardiac and immune systems between animals and humans. For instance, idiosyncratic drug reactions (drug- or individual human leukocyte antigen restriction-specific) are unpredictable, occur only in certain susceptible patients and have a complex dose-dependent relationship. Furthermore, the immune system is a complex network of organs, different cell types and molecules that interact among themselves and with other organs, cells, and local and systemic factors. Thus, it is very difficult to predict the behavior of the system from any of its components studied in isolation. Page 12 Finally, antigen presentation is not a static event but

spatiotemporally dynamic,⁵⁶⁴ which may lead to drug-mediated immunotoxicity due to unstable immunophenotypes (immunophenotypic drift).^{31,55,590,591} Multi-organ toxicity resulting from ICIs and CART may reflect the complex scenarios of the immune system in vivo. These problems may be minimized by combining in vitro data with mechanistic mathematical models, which describe intracellular metabolism, fluid-flow, substrate, hormone, and nutrient distribution and provide the opportunity to mimic the in vivo scenario.^{394,577}

7 | CONCLUSION AND OUTLOOK

Anticancer drug-induced toxicities occur because agents are not selective, or the targets are not unique to cancer cells. The research gaps resulting from the problematic specificity/selectivity of drugs leave a bottleneck on effective clinical management (curative care) because of uncontrollable toxicities on multiple organs. Improving the specificity/selectivity of drug target selection (druggable molecular targets and tumor-specific antigens) is considered the single most important factor, ⁵⁹² and this strategy will substantially minimize the risk of potential cardiovascular safety liabilities and other organ toxicity. The pharmaceutical industry may have to move toward more selective agents to meet clinical challenges. In line with the mission "oncology drugs still a pipeline priority" (innovation drives the race), as suggested by the US FDA,⁵⁹³ numerous approved anticancer drugs and those under investigation have been included in pharmaceutical pipeline databases. 594,595 Given the explosive rate of new anticancer drug development, there is an urgent need for a synergistic improvement of preclinical studies, clinical trials, pharmacovigilance and post-marketing surveillance as a whole, including an awareness of biosimilars in oncology. Cardiovascular toxicity has become a very challenging issue during cancer therapy, while consensual guidelines remain lacking for its effective management. 596 The development of novel therapeutic modalities such as chronotherapy and chronoprevention using natural products of antioxidants should be highly encouraged for both researchers and clinicians to simultaneously address clinical cardiotoxicity and drug efficacy. Furthermore, working with multidisciplinary teams should be considered compulsory to decrease morbidity and mortality from both cardiotoxicity and cancer itself, ⁵⁹⁷ and multidisciplinary collaboration is helpful to address the interdisciplinary differences and dilemmas in the meantime. Lastly, kinase inhibitors used in patients with cancer have given a new face to cardiovascular medicine, providing unprecedented insights into the functional roles played by numerous kinases in the cardiovascular system.² This insight will aid in the fundamental understanding of kinase inhibitors and their direct clinical applications in oncology, cardiology, and the cardio-oncology interface.

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REFERENCES

- Moslehi JJ, Deininger M. Tyrosine kinase inhibitor-associated cardiovascular toxicity in chronic myeloid leukemia. J Clin Oncol. 2015;33(35):4210-4218.
- Bellinger AM, Arteaga CL, Force T, Humphreys BD, Demetri GD, Druker BJ, Moslehi JJ. Cardio-Oncology: how new targeted cancer therapies and precision medicine can inform cardiovascular discovery. *Circulation*. 2015;132(23):2248– 2258.
- 3. Cheng H, Force T. Molecular mechanisms of cardiovascular toxicity of targeted cancer therapeutics. *Circ Res.* 2010;106(1):21–34.
- 4. Force T, Krause DS, Van Etten RA. Molecular mechanisms of cardiotoxicity of tyrosine kinase inhibition. *Nat Rev Cancer*. 2007;7(5):332–344.
- 5. Svoboda M, Poprach A, Dobes S, Kiss I, Vyzula R. Cardiac toxicity of targeted therapies used in the treatment for solid tumours: a review. *Cardiovasc Toxicol*. 2012;12(3):191–207.

- 6. Curigliano G, Cardinale D, Dent S, Criscitiello C, Aseyev O, Lenihan D, Cipolla CM. Cardiotoxicity of anticancer treatments: epidemiology, detection, and management. *CA Cancer J Clin*. 2016;66(4):309–325.
- 7. Force T, Kolaja KL. Cardiotoxicity of kinase inhibitors: the prediction and translation of preclinical models to clinical outcomes. *Nat Rev Drug Discov.* 2011;10(2):111–126.
- 8. Mellor HR, Bell AR, Valentin JP, Roberts RR. Cardiotoxicity associated with targeting kinase pathways in cancer. *Toxicol Sci.* 2011;120(1):14–32.
- Li W, Cornell RF, Lenihan D, Slosky D, Jagasia M, Piazza G, Moslehi J. Cardiovascular complications of novel multiple myeloma treatments. Circulation. 2016;133(9):908–912.
- 10. Stellitano A, Fedele R, Barilla S, Iaria A, Rao CM, Martino M. Chemotherapy and cardiotoxicity in hematologic malignancies: a Review. Curr Cancer Drug Targets. 2017;17(4):311–324.
- 11. Orphanos GS, Ioannidis GN, Ardavanis AG. Cardiotoxicity induced by tyrosine kinase inhibitors. *Acta Oncol.* 2009;48(7):964–970.
- 12. Heinzerling L, Ott PA, Hodi FS, Husain AN, Tajmir-Riahi A, Tawbi H, Pauschinger M, Gajewski TF, Lipson EJ, Luke JJ. Cardiotoxicity associated with CTLA4 and PD1 blocking immunotherapy. *J Immunother Cancer*. 2016;4:50.
- 13. Laubli H, Balmelli C, Bossard M, Pfister O, Glatz K, Zippelius A. Acute heart failure due to autoimmune myocarditis under pembrolizumab treatment for metastatic melanoma. *J Immunother Cancer*. 2015;3:11.
- Champiat S, Lambotte O, Barreau E, Belkhir R, Berdelou A, Carbonnel F, Cauquil C, Chanson P, Collins M, Durrbach A, Ederhy S, Feuillet S, Francois H, Lazarovici J, Le Pavec J, De Martin E, Mateus C, Michot JM, Samuel D, Soria JC, Robert C, Eggermont A, Marabelle A. Management of immune checkpoint blockade dysimmune toxicities: a collaborative position paper. Ann Oncol. 2016;27(4):559–574.
- Eggermont AM, Chiarion-Sileni V, Grob JJ, Dummer R, Wolchok JD, Schmidt H, Hamid O, Robert C, Ascierto PA, Richards JM, Lebbe C, Ferraresi V, Smylie M, Weber JS, Maio M, Konto C, Hoos A, de Pril V, Gurunath RK, de Schaetzen G, Suciu S, Testori A. Adjuvant ipilimumab versus placebo after complete resection of highrisk stage III melanoma (EORTC 18071): a randomised, double-blind, phase 3 trial. *Lancet Oncol.* 2015;16(5):522– 530.
- Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, Drake CG, Camacho LH, Kauh J, Odunsi K, Pitot HC, Hamid O, Bhatia S, Martins R, Eaton K, Chen S, Salay TM, Alaparthy S, Grosso JF, Korman AJ, Parker SM, Agrawal S, Goldberg SM, Pardoll DM, Gupta A, Wigginton JM. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. N Engl J Med. 2012;366(26):2455–2465.
- 17. Geisler BP, Raad RA, Esaian D, Sharon E, Schwartz DR. Apical ballooning and cardiomyopathy in a melanoma patient treated with ipilimumab: a case of takotsubo-like syndrome. *J Immunother Cancer*. 2015;3:4.
- 18. Yun S, Vincelette ND, Mansour I, Hariri D, Motamed S. Late onset ipilimumab-induced pericarditis and pericardial effusion: a rare but life threatening complication. *Case Rep Oncol Med*. 2015;2015:794842.
- 19. Gibson R, Delaune J, Szady A, Markham M. Suspected autoimmune myocarditis and cardiac conduction abnormalities with nivolumab therapy for non-small cell lung cancer. *BMJ Case Rep.* 2016;2016:pii: bcr2016216228.
- 20. Moslehi JJ. Cardiovascular toxic effects of targeted cancer therapies. N Engl J Med. 2016;375(15):1457-1467.
- 21. Heinzerling L, Goldinger SM. A review of serious adverse effects under treatment with checkpoint inhibitors. *Curr Opin Oncol.* 2017;29(2):136–144.
- 22. Cheng F, Loscalzo J. Autoimmune cardiotoxicity of cancer immunotherapy. Trends Immunol. 2017;38(2):77–78.
- 23. Semper H, Muehlberg F, Schulz-Menger J, Allewelt M, Grohe C. Drug-induced myocarditis after nivolumab treatment in a patient with PDL1- negative squamous cell carcinoma of the lung. *Lung Cancer*. 2016;99:117–119.
- Kushnir I, Wolf I. Nivolumab-induced pericardial tamponade: a case report and discussion. Cardiology. 2017;136(1):49–51.
- 25. Dasanu CA, Jen T, Skulski R. Late-onset pericardial tamponade, bilateral pleural effusions and recurrent immune monoarthritis induced by ipilimumab use for metastatic melanoma. *J Oncol Pharm Pract.* 2017;23(3):231–234.
- 26. Atezolizumab. http://www.accessdata.fda.gov/drugsatfda_docs/label/2016/761041lbl.pdf.
- 27. Alexander W. The checkpoint immunotherapy revolution: what started as a trickle has become a flood, despite some daunting adverse effects; new drugs, indications, and combinations continue to emerge. *PT*. 2016;41(3):185–191.
- 28. Teply BA, Lipson EJ. Identification and management of toxicities from immune checkpoint-blocking drugs. *Oncology* (Williston Park). 2014;28 Suppl 3:30–38.
- 29. Wang DY, Okoye GD, Neilan TG, Johnson DB, Moslehi JJ. Cardiovascular toxicities associated with cancer immunotherapies. *Curr Cardiol Rep.* 2017;19(3):21.

- Brudno JN, Kochenderfer JN. Toxicities of chimeric antigen receptor T cells: recognition and management. Blood. 2016;127(26):3321–3330.
- Bonifant CL, Jackson HJ, Brentjens RJ, Curran KJ. Toxicity and management in CAR T-cell therapy. Mol Ther Oncolytics. 2016;3:16011.
- 32. Linette GP, Stadtmauer EA, Maus MV, Rapoport AP, Levine BL, Emery L, Litzky L, Bagg A, Carreno BM, Cimino PJ, Binder-Scholl GK, Smethurst DP, Gerry AB, Pumphrey NJ, Bennett AD, Brewer JE, Dukes J, Harper J, Tayton-Martin HK, Jakobsen BK, Hassan NJ, Kalos M, June CH. Cardiovascular toxicity and titin cross-reactivity of affinity-enhanced T cells in myeloma and melanoma. *Blood*. 2013;122(6):863–871.
- 33. Morgan RA, Yang JC, Kitano M, Dudley ME, Laurencot CM, Rosenberg SA. Case report of a serious adverse event following the administration of T cells transduced with a chimeric antigen receptor recognizing ERBB2. *Mol Ther*. 2010;18(4):843–851.
- 34. Lee DW, Kochenderfer JN, Stetler-Stevenson M, Cui YK, Delbrook C, Feldman SA, Fry TJ, Orentas R, Sabatino M, Shah NN, Steinberg SM, Stroncek D, Tschernia N, Yuan C, Zhang H, Zhang L, Rosenberg SA, Wayne AS, Mackall CL. T cells expressing CD19 chimeric antigen receptors for acute lymphoblastic leukaemia in children and young adults: a phase 1 dose-escalation trial. *Lancet*. 2015;385(9967):517–528.
- 35. Kalos M, Levine BL, Porter DL, Katz S, Grupp SA, Bagg A, June CH. T cells with chimeric antigen receptors have potent antitumor effects and can establish memory in patients with advanced leukemia. *Sci Transl Med*. 2011;3(95):95ra73.
- 36. van den Berg JH, Gomez-Eerland R, van de Wiel B, Hulshoff L, van den Broek D, Bins A, Tan HL, Harper JV, Hassan NJ, Jakobsen BK, Jorritsma A, Blank CU, Schumacher TN, Haanen JB. Case report of a fatal serious adverse event upon administration of T cells transduced with a MART-1-specific T-cell receptor. *Mol Ther*. 2015;23(9):1541–1550.
- 37. Maus MV, Haas AR, Beatty GL, Albelda SM, Levine BL, Liu X, Zhao Y, Kalos M, June CH. T cells expressing chimeric antigen receptors can cause anaphylaxis in humans. *Cancer Immunol Res.* 2013;1(1):26–31.
- Maude SL, Frey N, Shaw PA, Aplenc R, Barrett DM, Bunin NJ, Chew A, Gonzalez VE, Zheng Z, Lacey SF, Mahnke YD, Melenhorst JJ, Rheingold SR, Shen A, Teachey DT, Levine BL, June CH, Porter DL, Grupp SA. Chimeric antigen receptor T cells for sustained remissions in leukemia. N Engl J Med. 2014;371(16):1507–1517.
- 39. Davila ML, Riviere I, Wang X, Bartido S, Park J, Curran K, Chung SS, Stefanski J, Borquez-Ojeda O, Olszewska M, Qu J, Wasielewska T, He Q, Fink M, Shinglot H, Youssif M, Satter M, Wang Y, Hosey J, Quintanilla H, Halton E, Bernal Y, Bouhassira DC, Arcila ME, Gonen M, Roboz GJ, Maslak P, Douer D, Frattini MG, Giralt S, Sadelain M, Brentjens R. Efficacy and toxicity management of 19–28z CAR T cell therapy in B cell acute lymphoblastic leukemia. Sci Transl Med. 2014;6(224):224ra225.
- 40. Kochenderfer JN, Dudley ME, Carpenter RO, Kassim SH, Rose JJ, Telford WG, Hakim FT, Halverson DC, Fowler DH, Hardy NM, Mato AR, Hickstein DD, Gea-Banacloche JC, Pavletic SZ, Sportes C, Maric I, Feldman SA, Hansen BG, Wilder JS, Blacklock-Schuver B, Jena B, Bishop MR, Gress RE, Rosenberg SA. Donor-derived CD19-targeted T cells cause regression of malignancy persisting after allogeneic hematopoietic stem cell transplantation. *Blood*. 2013;122(25):4129-4139.
- 41. Brudno JN, Somerville RP, Shi V, Rose JJ, Halverson DC, Fowler DH, Gea-Banacloche JC, Pavletic SZ, Hickstein DD, Lu TL, Feldman SA, Iwamoto AT, Kurlander R, Maric I, Goy A, Hansen BG, Wilder JS, Blacklock-Schuver B, Hakim FT, Rosenberg SA, Gress RE, Kochenderfer JN. Allogeneic T cells that express an anti-CD19 chimeric antigen receptor induce remissions of B-cell malignancies that progress after allogeneic hematopoietic stem-cell transplantation without causing graft-versus-host disease. *J Clin Oncol.* 2016;34(10):1112–1121.
- 42. Kochenderfer JN, Dudley ME, Kassim SH, Somerville RP, Carpenter RO, Stetler-Stevenson M, Yang JC, Phan GQ, Hughes MS, Sherry RM, Raffeld M, Feldman S, Lu L, Li YF, Ngo LT, Goy A, Feldman T, Spaner DE, Wang ML, Chen CC, Kranick SM, Nath A, Nathan DA, Morton KE, Toomey MA, Rosenberg SA. Chemotherapy-refractory diffuse large B-cell lymphoma and indolent B-cell malignancies can be effectively treated with autologous T cells expressing an anti-CD19 chimeric antigen receptor. *J Clin Oncol.* 2015;33(6):540–549.
- 43. Kochenderfer JN, Dudley ME, Feldman SA, Wilson WH, Spaner DE, Maric I, Stetler-Stevenson M, Phan GQ, Hughes MS, Sherry RM, Yang JC, Kammula US, Devillier L, Carpenter R, Nathan DA, Morgan RA, Laurencot C, Rosenberg SA. B-cell depletion and remissions of malignancy along with cytokine-associated toxicity in a clinical trial of anti-CD19 chimeric-antigen-receptor-transduced T cells. Blood. 2012;119(12):2709–2720.
- 44. Nelson MH, Paulos CM. Novel immunotherapies for hematologic malignancies. Immunol Rev. 2015;263(1):90-105.
- 45. Kang YJ, Molecular and cellular mechanisms of cardiotoxicity. Environ Health Perspect. 2001;109 Suppl 1:27-34.
- 46. Coviello JS, Cardiac side effects of targeted therapies. Semin Oncol Nurs. 2014;30(3):175-182.
- 47. Hedhli N, Russell KS. Cardiotoxicity of molecularly targeted agents. Curr Cardiol Rev. 2011;7(4):221-233.

- 48. Raschi E, De Ponti F. Cardiovascular toxicity of anticancer-targeted therapy: emerging issues in the era of cardio-oncology. *Intern Emerg Med*. 2012;7(2):113–131.
- 49. Ederhy S, Izzedine H, Massard C, Dufaitre G, Spano JP, Milano G, Meuleman C, Besse B, Boccara F, Kahyat D, Cohen A, Soria JC. Cardiac side effects of molecular targeted therapies: towards a better dialogue between oncologists and cardiologists. *Crit Rev Oncol Hematol.* 2011; 80(3):369–379.
- 50. Hasinoff BB, Patel D. The lack of target specificity of small molecule anticancer kinase inhibitors is correlated with their ability to damage myocytes in vitro. *Toxicol Appl Pharmacol.* 2010;249(2):132–139.
- 51. van Loenen MM, de Boer R, Amir AL, Hagedoorn RS, Volbeda GL, Willemze R, van Rood JJ, Falkenburg JH, Heemskerk MH. Mixed T cell receptor dimers harbor potentially harmful neoreactivity. *Proc Natl Acad Sci U S A*. 2010;107(24):10972–10977.
- 52. Liu S, Kurzrock R. Understanding toxicities of targeted agents: implications for anti-tumor activity and management. Semin Oncol. 2015;42(6):863–875.
- 53. Gross G, Eshhar Z. Therapeutic potential of T cell chimeric antigen receptors (CARs) in cancer treatment: counteracting off-tumor toxicities for safe CAR T cell therapy. *Annu Rev Pharmacol Toxicol*. 2016;56:59–83.
- 54. Villadolid J, Amin A. Immune checkpoint inhibitors in clinical practice: update on management of immune-related toxicities. *Transl Lung Cancer Res.* 2015;4(5):560–575.
- 55. Jethwa H, Adami AA, Maher J. Use of gene-modified regulatory T-cells to control autoimmune and alloimmune pathology: is now the right time? Clin Immunol. 2014;150(1):51–63.
- 56. Kizhedath A, Wilkinson S, Glassey J. Applicability of predictive toxicology methods for monoclonal antibody therapeutics: status quo and scope. *Arch Toxicol*. 2017;91(4):1595–1612.
- 57. Dy GK, Adjei AA. Understanding, recognizing, and managing toxicities of targeted anticancer therapies. *CA Cancer J Clin.* 2013;63(4):249–279.
- 58. Seruga B, Sterling L, Wang L, Tannock IF. Reporting of serious adverse drug reactions of targeted anticancer agents in pivotal phase III clinical trials. *J Clin Oncol.* 2011;29(2):174–185.
- 59. Rosland GV, Engelsen AS. Novel points of attack for targeted cancer therapy. *Basic Clin Pharmacol Toxicol*. 2015;116(1):9–18.
- 60. Siegel R, DeSantis C, Virgo K, Stein K, Mariotto A, Smith T, Cooper D, Gansler T, Lerro C, Fedewa S, Lin C, Leach C, Cannady RS, Cho H, Scoppa S, Hachey M, Kirch R, Jemal A, Ward E. Cancer treatment and survivorship statistics, 2012. *CA Cancer J Clin*. 2012;62(4):220–241.
- 61. Shah DR, Shah RR, Morganroth J. Tyrosine kinase inhibitors: their on-target toxicities as potential indicators of efficacy. Drug Saf. 2013;36(6):413–426.
- 62. Chen YM, Immune checkpoint inhibitors for nonsmall cell lung cancer treatment. J Chin Med Assoc. 2017;80(1):7-14.
- 63. de Mello RA, Veloso AF, Esrom Catarina P, Nadine S, Antoniou G. Potential role of immunotherapy in advanced non-small-cell lung cancer. *Onco Targets Ther*. 2017;10:21–30.
- 64. Donin NM, Lenis AT, Holden S, Drakaki A, Pantuck A, Belldegrun A, Chamie K. Immunotherapy for the treatment of urothelial carcinoma. *J Urol.* 2017;197(1):14–22.
- 65. Du L, Herbst RS, Morgensztern D. Immunotherapy in lung cancer. Hematol Oncol Clin North Am. 2017;31(1):131-141.
- 66. Hude I, Sasse S, Engert A, Brockelmann PJ. The emerging role of immune checkpoint inhibition in malignant lymphoma. *Haematologica*. 2017;102(1):30–42.
- 67. Mohamed H, Eltobgy M, Abdel-Rahman O. Immune checkpoints aberrations and malignant mesothelioma: assessment of prognostic value and evaluation of therapeutic potentials. *Anticancer Agents Med Chem.* 2017. [Epub ahead of print] https://www.ncbi.nlm.nih.gov/pubmed/28042777.
- 68. Remon J, Besse B. Immune checkpoint inhibitors in first-line therapy of advanced non-small cell lung cancer. *Curr Opin Oncol.* 2017;29(2):97–104.
- 69. Yu LY, Tang J, Zhang CM, Zeng WJ, Yan H, Li MP, Chen XP. New immunotherapy strategies in breast cancer. *Int J Environ Res Public Health*. 2017;14(1):pii: E68. https://doi.org/10.3390/ijerph14010068.
- 70. Zhou TC, Sankin Al, Porcelli SA, Perlin DS, Schoenberg MP, Zang X. A review of the PD-1/PD-L1 checkpoint in bladder cancer: from mediator of immune escape to target for treatment. *Urol Oncol.* 2017;35(1):14–20.
- 71. Guo Y, Wang Y, Han W. Chimeric antigen receptor-modified T cells for solid tumors: challenges and prospects. *J Immunol Res.* 2016;2016:3850839.
- 72. Kenderian SS, Porter DL, Gill S. Chimeric antigen receptor T cells and hematopoietic cell transplantation: how not to put the CART before the horse. *Biol Blood Marrow Transplant*. 2017;23(2):235–246.

- 73. Ruella M, June CH. Chimeric antigen receptor T cells for B cell neoplasms: choose the right CAR for you. *Curr Hematol Malig Rep.* 2016;11(5):368–384.
- 74. Smith C, Khanna R. Adoptive cellular immunotherapy for virus-associated cancers: a new paradigm in personalized medicine. *Immunol Cell Biol.* 2017;95(4):364–371.
- 75. Viganego F, Singh R, Fradley MG. Arrhythmias and other electrophysiology issues in cancer patients receiving chemotherapy or radiation. *Curr Cardiol Rep.* 2016;18(6):52.
- Manrique CR, Park M, Tiwari N, Plana JC, Garcia MJ. Diagnostic strategies for early recognition of cancer therapeuticsrelated cardiac dysfunction. Clin Med Insights Cardiol. 2017;11:1179546817697983.
- 77. Tajiri K, Aonuma K, Sekine I. Cardio-oncology: a multidisciplinary approach for detection, prevention and management of cardiac dysfunction in cancer patients. *Jpn J Clin Oncol*. 2017;47(8):678–682.
- 78. Hamo CE, Bloom MW, Cardinale D, Ky B, Nohria A, Baer L, Skopicki H, Lenihan DJ, Gheorghiade M, Lyon AR, Butler J. Cancer therapy-related cardiac dysfunction and heart failure: part 2: prevention, treatment, guidelines, and future directions. *Circ Heart Fail*. 2016;9(2):e002843.
- 79. Thavendiranathan P, Nolan MT. An emerging epidemic: cancer and heart failure. Clin Sci (Lond). 2017;131(2):113-121.
- 80. Cautela J, Lalevee N, Ammar C, Ederhy S, Peyrol M, Debourdeau P, Serin D, Le Dolley Y, Michel N, Orabona M, Barraud J, Laine M, Bonello L, Paganelli F, Barlesi F, Thuny F. Management and research in cancer treatment-related cardiovascular toxicity: challenges and perspectives. *Int J Cardiol*. 2016;224:366–375.
- 81. Mailankody S, Prasad V. Implications of proposed medicare reforms to counteract high cancer drug prices. *JAMA*. 2016;316(3):271–272.
- 82. Mailankody S, Prasad V. Five years of cancer drug approvals: innovation, efficacy, and costs. *JAMA Oncol.* 2015;1(4):539–540.
- 83. Restifo NP, Dudley ME, Rosenberg SA. Adoptive immunotherapy for cancer: harnessing the T cell response. *Nat Rev Immunol*. 2012;12(4):269–281.
- 84. Maude S, Barrett DM. Current status of chimeric antigen receptor therapy for haematological malignancies. *Br J Haematol*. 2016;172(1):11–22.
- 85. Zhang W, CAR T-cell therapy: opportunities and challenges. Immunotherapy. 2016;8(3):245-247.
- 86. Kalaitsidou M, Kueberuwa G, Schutt A, Gilham DE. CAR T-cell therapy: toxicity and the relevance of preclinical models. Immunotherapy. 2015;7(5):487–497.
- 87. Camacho LH. Current status of biosimilars in oncology. Drugs. 2017;77(9):985-997.
- 88. Jacobs I, Ewesuedo R, Lula S, Zacharchuk C. Biosimilars for the treatment of cancer: a systematic review of published evidence. *BioDrugs*. 2017;31(1):1–36.
- 89. Declerck P, Danesi R, Petersel D, Jacobs I. The language of biosimilars: clarification, definitions, and regulatory aspects. *Drugs*. 2017;77(6):671–677.
- Madonna R, Cadeddu C, Deidda M, Mele D, Monte I, Novo G, Pagliaro P, Pepe A, Spallarossa P, Tocchetti CG, Zito C, Mercuro G. Improving the preclinical models for the study of chemotherapy-induced cardiotoxicity: a position paper of the Italian working group on drug cardiotoxicity and cardioprotection. *Heart Fail Rev.* 2015;20(5):621–631.
- 91. Turner JR, Integrated cardiovascular safety: multifaceted considerations in drug development and therapeutic use. Expert Opin Drug Saf. 2017;16(4):481–492.
- 92. Larsen CM, Mulvagh SL. Cardio-oncology: what you need to know now for clinical practice and echocardiography. *Echo Res Pract*. 2017;4(1):R33–R41.
- 93. Cheng F, Loscalzo J. Autoimmune cardiotoxicity of cancer immunotherapy. Trends Immunol. 2017;38(2):77-78.
- 94. Lenihan DJ, Hartlage G, DeCara J, Blaes A, Finet JE, Lyon AR, Cornell RF, Moslehi J, Oliveira GH, Murtagh G, Fisch M, Zeevi G, lakobishvili Z, Witteles R, Patel A, Harrison E, Fradley M, Curigliano G, Lenneman CG, Magalhaes A, Krone R, Porter C, Parasher S, Dent S, Douglas P, Carver J. Cardio-Oncology Training: a proposal from the international cardioncology society and Canadian Cardiac Oncology Network for a new multidisciplinary specialty. J Card Fail. 2016;22(6):465-471.
- 95. Gujral DM, Lloyd G, Bhattacharyya S. Provision and clinical utility of cardio-oncology services for detection of cardiac toxicity in cancer patients. *J Am Coll Cardiol*. 2016;67(12):1499–1500.
- Givertz MM, Cancer and the heart: training subspecialists, optimizing care, and moving the field forward. J Card Fail. 2016;22(6):472-474.

- 97. Yang H, Qin C, Li YH, Tao L, Zhou J, Yu CY, Xu F, Chen Z, Zhu F, Chen YZ. Therapeutic target database update 2016: enriched resource for bench to clinical drug target and targeted pathway information. *Nucleic Acids Res.* 2016;44(D1):D1069-D1074.
- 98. Rask-Andersen M, Zhang J, Fabbro D, Schioth HB. Advances in kinase targeting: current clinical use and clinical trials. Trends Pharmacol Sci. 2014;35(11):604–620.
- 99. Xu J, Wang P, Yang H, Zhou J, Li Y, Li X, Xue W, Yu C, Tian Y, Zhu F. Comparison of FDA approved kinase targets to clinical trial ones: insights from their system profiles and drug-target interaction networks. *Biomed Res Int*. 2016;2016:2509385.
- 100. Qin C, Zhang C, Zhu F, Xu F, Chen SY, Zhang P, Li YH, Yang SY, Wei YQ, Tao L, Chen YZ. Therapeutic target database update 2014: a resource for targeted therapeutics. *Nucleic Acids Res.* 2014;42(Database issue):D1118-D1123.
- Levitzki A, Tyrosine kinase inhibitors: views of selectivity, sensitivity, and clinical performance. Annu Rev Pharmacol Toxicol. 2013;53:161–185.
- 102. Roskoski R, Jr Classification of small molecule protein kinase inhibitors based upon the structures of their drug-enzyme complexes. *Pharmacol Res.* 2016;103:26–48.
- 103. Carmi C, Mor M, Petronini PG, Alfieri RR. Clinical perspectives for irreversible tyrosine kinase inhibitors in cancer. *Biochem Pharmacol.* 2012;84(11):1388–1399.
- 104. Wu P, Nielsen TE, Clausen MH. FDA-approved small-molecule kinase inhibitors. *Trends Pharmacol Sci.* 2015;36(7):422–439.
- 105. Liu Q, Sabnis Y, Zhao Z, Zhang T, Buhrlage SJ, Jones LH, Gray NS. Developing irreversible inhibitors of the protein kinase cysteinome. *Chem Biol.* 2013;20(2):146–159.
- 106. Krause DS, Van Etten RA. Tyrosine kinases as targets for cancer therapy. N Engl J Med. 2005;353(2):172-187.
- 107. Govender J, Loos B, Marais E, Engelbrecht AM. Mitochondrial catastrophe during doxorubicin-induced cardiotoxicity: a review of the protective role of melatonin. *J Pineal Res.* 2014;57(4):367–380.
- 108. Manning G, Whyte DB, Martinez R, Hunter T, Sudarsanam S. The protein kinase complement of the human genome. *Science*. 2002;298(5600):1912–1934.
- 109. Roskoski R, Jr A historical overview of protein kinases and their targeted small molecule inhibitors. *Pharmacol Res.* 2015;100:1–23.
- 110. Zhang H, Photiou A, Grothey A, Stebbing J, Giamas G. The role of pseudokinases in cancer. *Cell Signal*. 2012;24(6):1173–1184
- 111. Pazin MJ, Williams LT. Triggering signaling cascades by receptor tyrosine kinases. *Trends Biochem Sci.* 1992;17(10):374–379
- 112. Bolen JB, Nonreceptor tyrosine protein kinases. Oncogene. 1993;8(8):2025-2031.
- 113. Lipovka Y, Konhilas JP. AMP-activated protein kinase signalling in cancer and cardiac hypertrophy. *Cardiovasc Pharm Open Access*. 2015;4(3):154.
- 114. Zhang J, Yang PL, Gray NS. Targeting cancer with small molecule kinase inhibitors. Nat Rev Cancer. 2009;9(1):28–39.
- 115. Wong KK, Engelman JA, Cantley LC. Targeting the PI3K signaling pathway in cancer. Curr Opin Genet Dev. 2010;20(1):87-
- 116. Fischer PM. Approved and experimental small-molecule oncology kinase inhibitor drugs: a mid-2016 overview. *Med Res Rev.* 2017;37(2):314–367.
- 117. Sutherland JJ, Gao C, Cahya S, Vieth M. What general conclusions can we draw from kinase profiling data sets? *Biochim Biophys Acta*. 2013;1834(7):1425–1433.
- 118. Frezza M, Schmitt S, Dou QP. Targeting the ubiquitin-proteasome pathway: an emerging concept in cancer therapy. *Curr Top Med Chem.* 2011;11(23):2888–2905.
- 119. Shaikh AY, Shih JA. Chemotherapy-induced cardiotoxicity. Curr Heart Fail Rep. 2012;9(2):117-127.
- 120. Roy R, Evens AM, Patton D, Gallot L, Larson A, Rademaker A, Cilley J, Spies S, Variakojis D, Gordon LI, Winter JN. Bortezomib may be safely combined with Y-90-ibritumomab tiuxetan in patients with relapsed/refractory follicular non-Hodgkin lymphoma: a phase I trial of combined induction therapy and bortezomib consolidation. *Leuk Lymphoma*. 2013;54(3):497–502.
- 121. Enrico O, Gabriele B, Nadia C, Sara G, Daniele V, Giulia C, Antonio S, Mario P. Unexpected cardiotoxicity in haematological bortezomib treated patients. *Br J Haematol*. 2007;138(3):396–397.
- 122. Subedi A, Sharma LR, Shah BK. Bortezomib-induced acute congestive heart failure: a case report and review of literature. *Ann Hematol.* 2014;93(10):1797–1799.

- 123. Xiao Y, Yin J, Wei J, Shang Z. Incidence and risk of cardiotoxicity associated with bortezomib in the treatment of cancer: a systematic review and meta-analysis. *PLoS One.* 2014;9(1):e87671.
- 124. Maurea N, Spallarossa P, Cadeddu C, Madonna R, Mele D, Monte I, Novo G, Pagliaro P, Pepe A, Tocchetti CG, Zito C, Mercuro G. A recommended practical approach to the management of target therapy and angiogenesis inhibitors cardiotoxicity: an opinion paper of the working group on drug cardiotoxicity and cardioprotection, Italian Society of Cardiology. *J Cardiovasc Med (Hagerstown)*. 2016;17 Suppl 1:e93–e104.
- 125. Bortezomib. https://www.uptodate.com/contents/bortezomib-drug-information?source=search_result&search=Borte zomib&selectedTitle=1~100.
- 126. Grandin EW, Ky B, Cornell RF, Carver J, Lenihan DJ. Patterns of cardiac toxicity associated with irreversible proteasome inhibition in the treatment of multiple myeloma. *J Card Fail*. 2015;21(2):138–144.
- 127. Danhof S, Schreder M, Rasche L, Strifler S, Einsele H, Knop S. 'Real-life' experience of preapproval carfilzomib-based therapy in myeloma analysis of cardiac toxicity and predisposing factors. *Eur J Haematol.* 2016;97(1):25–32.
- 128. Siegel D, Martin T, Nooka A, Harvey RD, Vij R, Niesvizky R, Badros AZ, Jagannath S, McCulloch L, Rajangam K, Lonial S. Integrated safety profile of single-agent carfilzomib: experience from 526 patients enrolled in 4 phase II clinical studies. *Haematologica*. 2013;98(11):1753–1761.
- 129. Harvey RD, Incidence and management of adverse events in patients with relapsed and/or refractory multiple myeloma receiving single-agent carfilzomib. *Clin Pharmacol*. 2014;6:87–96.
- 130. Carfilzomib. https://www.uptodate.com/contents/carfilzomib-drug-information?source=search_result&search=Carfilzomib&selectedTitle=1~21.
- 131. Laubach JP, Moslehi JJ, Francis SA, San Miguel JF, Sonneveld P, Orlowski RZ, Moreau P, Rosinol L, Faber EA, Jr., Voorhees P, Mateos MV, Marquez L, Feng H, Desai A, van de Velde H, Elliott J, Shi H, Dow E, Jobanputra N, Esseltine DL, Niculescu L, Anderson KC, Lonial S, Richardson PG. A retrospective analysis of 3954 patients in phase 2/3 trials of bortezomib for the treatment of multiple myeloma: towards providing a benchmark for the cardiac safety profile of proteasome inhibition in multiple myeloma. *Br J Haematol.* 2017;178(4):547–560.
- 132. Lakshmaiah KC, Jacob LA, Aparna S, Lokanatha D, Saldanha SC. Epigenetic therapy of cancer with histone deacetylase inhibitors. *J Cancer Res Ther.* 2014;10(3):469–478.
- 133. Song SH, Han SW, Bang YJ. Epigenetic-based therapies in cancer: progress to date. Drugs. 2011;71(18):2391–2403.
- 134. Liu M, Zhou J, Chen Z, Cheng AS. Understanding the epigenetic regulation of tumours and their microenvironments: opportunities and problems for epigenetic therapy. *J Pathol.* 2017;241(1):10–24.
- 135. Hronek J, Reed M. Nursing roles in cardiac safety: romidepsin in Patients with T-cell lymphoma. *Oncol Nurs Forum*. 2016;43(2):227–234.
- 136. Shah MH, Binkley P, Chan K, Xiao J, Arbogast D, Collamore M, Farra Y, Young D, Grever M. Cardiotoxicity of histone deacetylase inhibitor depsipeptide in patients with metastatic neuroendocrine tumors. *Clin Cancer Res.* 2006;12(13):3997–4003.
- 137. Federico M, Bagella L. Histone deacetylase inhibitors in the treatment of hematological malignancies and solid tumors. *J Biomed Biotechnol.* 2011;2011;475641.
- 138. Piekarz RL, Frye AR, Wright JJ, Steinberg SM, Liewehr DJ, Rosing DR, Sachdev V, Fojo T, Bates SE. Cardiac studies in patients treated with depsipeptide, FK228, in a phase II trial for T-cell lymphoma. *Clin Cancer Res.* 2006;12(12):3762–3773.
- 139. Romidepsin. https://www.uptodate.com/contents/romidepsin-drug-information?source=search_result&search=Romi depsin&selectedTitle=1~10.
- 140. Laubach JP, San-Miguel JF, Hungria V, Hou J, Moreau P, Lonial S, Lee JH, Einsele H, Alsina M, Richardson PG. Deacetylase inhibitors: an advance in myeloma therapy? *Expert Rev Hematol*. 2017;10(3):229–237.
- 141. Yeo W, Chung HC, Chan SL, Wang LZ, Lim R, Picus J, Boyer M, Mo FK, Koh J, Rha SY, Hui EP, Jeung HC, Roh JK, Yu SC, To KF, Tao Q, Ma BB, Chan AW, Tong JH, Erlichman C, Chan AT, Goh BC. Epigenetic therapy using belinostat for patients with unresectable hepatocellular carcinoma: a multicenter phase I/II study with biomarker and pharmacokinetic analysis of tumors from patients in the Mayo Phase II Consortium and the Cancer Therapeutics Research Group. *J Clin Oncol.* 2012;30(27):3361–3367.
- 142. Belinostat. https://www.uptodate.com/contents/belinostat-drug-information?source=search_result&search=Belinostat&selectedTitle=1~7.
- 143. Redic KA, Hough SM, Price EM. Clinical developments in the treatment of relapsed or relapsed and refractory multiple myeloma: impact of panobinostat, the first-in-class histone deacetylase inhibitor. *Onco Targets Ther.* 2016;9:2783–2793.

- 144. Panobinostat. https://www.uptodate.com/contents/panobinostat-drug-information?source=search_result&search= Panobinostat&selectedTitle=1~12.
- 145. Eglen R, Reisine T. Drug discovery and the human kinome: recent trends. Pharmacol Ther. 2011;130(2):144-156.
- 146. Singh J, Petter RC, Baillie TA, Whitty A. The resurgence of covalent drugs. Nat Rev Drug Discov. 2011;10(4):307-317.
- 147. Landi L, Cappuzzo F. Irreversible EGFR-TKIs: dreaming perfection. Transl Lung Cancer Res. 2013;2(1):40-49.
- 148. Baillie TA, Future of toxicology-metabolic activation and drug design: challenges and opportunities in chemical toxicology. *Chem Res Toxicol*. 2006;19(7):889–893.
- 149. Liebler DC, Protein damage by reactive electrophiles: targets and consequences. Chem Res Toxicol. 2008;21(1):117-128.
- 150. Fasano M, Della Corte CM, Califano R, Capuano A, Troiani T, Martinelli E, Ciardiello F, Morgillo F. Type III or allosteric kinase inhibitors for the treatment of non-small cell lung cancer. Expert Opin Investig Drugs. 2014;23(6):809–821.
- 151. Dar AC, Shokat KM. The evolution of protein kinase inhibitors from antagonists to agonists of cellular signaling. *Annu Rev Biochem.* 2011;80:769–795.
- 152. Lamba V, Ghosh I. New directions in targeting protein kinases: focusing upon true allosteric and bivalent inhibitors. *Curr Pharm Des.* 2012;18(20):2936–2945.
- 153. Knight ZA, Lin H, Shokat KM. Targeting the cancer kinome through polypharmacology. *Nat Rev Cancer*. 2010;10(2):130–137.
- 154. Cross MJ, Berridge BR, Clements PJ, Cove-Smith L, Force TL, Hoffmann P, Holbrook M, Lyon AR, Mellor HR, Norris AA, Pirmohamed M, Tugwood JD, Sidaway JE, Park BK. Physiological, pharmacological and toxicological considerations of drug-induced structural cardiac injury. *Br J Pharmacol*. 2015;172(4):957–974.
- 155. Doenst T, Nguyen TD, Abel ED. Cardiac metabolism in heart failure: implications beyond ATP production. *Circ Res.* 2013;113(6):709–724.
- 156. Weisner J, Gontla R, van der Westhuizen L, Oeck S, Ketzer J, Janning P, Richters A, Muhlenberg T, Fang Z, Taher A, Jendrossek V, Pelly SC, Bauer S, van Otterlo WA, Rauh D. Covalent-allosteric kinase inhibitors. *Angew Chem Int Ed Engl.* 2015;54(35):10313–10316.
- 157. Trametinib. https://www.uptodate.com/contents/trametinib-drug-information?source=search_result&search=Tramet inib&selectedTitle=1~26.
- 158. Flaherty KT, Robert C, Hersey P, Nathan P, Garbe C, Milhem M, Demidov LV, Hassel JC, Rutkowski P, Mohr P, Dummer R, Trefzer U, Larkin JM, Utikal J, Dreno B, Nyakas M, Middleton MR, Becker JC, Casey M, Sherman LJ, Wu FS, Ouellet D, Martin AM, Patel K, Schadendorf D, Group MS. Improved survival with MEK inhibition in BRAF-mutated melanoma. N Engl J Med. 2012;367(2):107–114.
- 159. Kim KB, Kefford R, Pavlick AC, Infante JR, Ribas A, Sosman JA, Fecher LA, Millward M, McArthur GA, Hwu P, Gonzalez R, Ott PA, Long GV, Gardner OS, Ouellet D, Xu Y, DeMarini DJ, Le NT, Patel K, Lewis KD. Phase II study of the MEK1/MEK2 inhibitor Trametinib in patients with metastatic BRAF-mutant cutaneous melanoma previously treated with or without a BRAF inhibitor. *J Clin Oncol.* 2013;31(4):482–489.
- 160. Ibrutinib. https://www.uptodate.com/contents/ibrutinib-drug-information?source=search_result&search=lbrutinib&selectedTitle=1~36.
- Lampson BL, Yu L, Glynn RJ, Barrientos JC, Jacobsen ED, Banerji V, Jones JA, Walewska R, Savage KJ, Michaud GF, Moslehi JJ, Brown JR. Ventricular arrhythmias and sudden death in patients taking ibrutinib. *Blood*. 2017;129(18):2581–2584.
- 162. Sendur MA, Aksoy S, Altundag K. Cardiotoxicity of novel HER2-targeted therapies. *Curr Med Res Opin.* 2013;29(8):1015–1024.
- 163. Afatinib. https://www.uptodate.com/contents/afatinib-drug-information?source=search_result&search=Afatinib&selectedTitle=1~15.
- 164. Gower CM, Chang ME, Maly DJ. Bivalent inhibitors of protein kinases. Crit Rev Biochem Mol Biol. 2014;49(2):102-115.
- 165. Bailey FP, Byrne DP, McSkimming D, Kannan N, Eyers PA. Going for broke: targeting the human cancer pseudokinome. *Biochem J.* 2015;465(2):195–211.
- 166. Hammaren HM, Virtanen AT, Silvennoinen O. Nucleotide-binding mechanisms in pseudokinases. *Biosci Rep.* 2015;36(1):e00282.
- 167. Reiterer V, Eyers PA, Farhan H. Day of the dead: pseudokinases and pseudophosphatases in physiology and disease. Trends Cell Biol. 2014;24(9):489–505.
- 168. Mondal D, Mathur A, Chandra PK. Tripping on TRIB3 at the junction of health, metabolic dysfunction and cancer. *Biochimie*. 2016;124:34–52.

- 169. Eyers PA, Keeshan K, Kannan N. Tribbles in the 21st century: The evolving roles of tribbles pseudokinases in biology and disease. *Trends Cell Biol.* 2017;27(4):284–298.
- 170. Vainchenker W, Constantinescu SN. JAK/STAT signaling in hematological malignancies. *Oncogene*. 2013;32(21):2601–2613
- 171. Baselga J, Swain SM. Novel anticancer targets: revisiting ERBB2 and discovering ERBB3. *Nat Rev Cancer*. 2009;9(7):463–475.
- 172. Salazar M, Lorente M, Orea-Soufi A, Davila D, Erazo T, Lizcano J, Carracedo A, Kiss-Toth E, Velasco G. Oncosuppressive functions of tribbles pseudokinase 3. *Biochem Soc Trans*. 2015;43(5):1122–1126.
- 173. Xie T, Lim SM, Westover KD, Dodge ME, Ercan D, Ficarro SB, Udayakumar D, Gurbani D, Tae HS, Riddle SM, Sim T, Marto JA, Janne PA, Crews CM, Gray NS. Pharmacological targeting of the pseudokinase Her3. *Nat Chem Biol.* 2014;10(12):1006–1012.
- 174. Cowan-Jacob SW, Jahnke W, Knapp S. Novel approaches for targeting kinases: allosteric inhibition, allosteric activation and pseudokinases. *Future Med Chem.* 2014;6(5):541–561.
- 175. Karvonen H, Niininen W, Murumagi A, Ungureanu D. Targeting ROR1 identifies new treatment strategies in hematological cancers. *Biochem Soc Trans*. 2017;45(2):457–464.
- 176. Rebagay G, Yan S, Liu C, Cheung NK. ROR1 and ROR2 in human malignancies: potentials for targeted therapy. *Front Oncol.* 2012;2:34.
- 177. Borcherding N, Kusner D, Liu GH, Zhang W. ROR1, an embryonic protein with an emerging role in cancer biology. *Protein Cell*. 2014;5(7):496–502.
- 178. Aghebati-Maleki L, Shabani M, Baradaran B, Motallebnezhad M, Majidi J, Yousefi M. Receptor tyrosine kinase-like orphan receptor 1 (ROR-1): an emerging target for diagnosis and therapy of chronic lymphocytic leukemia. *Biomed Pharmacother*. 2017;88:814–822.
- 179. Shabani M, Naseri J, Shokri F. Receptor tyrosine kinase-like orphan receptor 1: a novel target for cancer immunotherapy. *Expert Opin Ther Targets*. 2015;19(7):941–955.
- 180. Asem MS, Buechler S, Wates RB, Miller DL, Stack MS. Wnt5a signaling in cancer. Cancers (Basel). 2016;8(9):79.
- 181. Dave H, Anver MR, Butcher DO, Brown P, Khan J, Wayne AS, Baskar S, Rader C. Restricted cell surface expression of receptor tyrosine kinase ROR1 in pediatric B-lineage acute lymphoblastic leukemia suggests targetability with therapeutic monoclonal antibodies. *PLoS One.* 2012;7(12):e52655.
- 182. Hojjat-Farsangi M, Moshfegh A, Daneshmanesh AH, Khan AS, Mikaelsson E, Osterborg A, Mellstedt H. The receptor tyrosine kinase ROR1-an oncofetal antigen for targeted cancer therapy. *Semin Cancer Biol.* 2014;29:21–31.
- 183. Richey EA, Lyons EA, Nebeker JR, Shankaran V, McKoy JM, Luu TH, Nonzee N, Trifilio S, Sartor O, Benson AB, 3rd, Carson KR, Edwards BJ, Gilchrist-Scott D, Kuzel TM, Raisch DW, Tallman MS, West DP, Hirschfeld S, Grillo-Lopez AJ, Bennett CL. Accelerated approval of cancer drugs: improved access to therapeutic breakthroughs or early release of unsafe and ineffective drugs? *J Clin Oncol.* 2009;27(26):4398–4405.
- 184. Mertens AC, Yasui Y, Neglia JP, Potter JD, Nesbit ME, Jr., Ruccione K, Smithson WA, Robison LL. Late mortality experience in five-year survivors of childhood and adolescent cancer: the Childhood Cancer Survivor Study. *J Clin Oncol.* 2001;19(13):3163–3172.
- 185. Yeh ET, Bickford CL. Cardiovascular complications of cancer therapy: incidence, pathogenesis, diagnosis, and management. *J Am Coll Cardiol*. 2009;53(24):2231–2247.
- 186. Meacham LR, Chow EJ, Ness KK, Kamdar KY, Chen Y, Yasui Y, Oeffinger KC, Sklar CA, Robison LL, Mertens AC. Cardiovascular risk factors in adult survivors of pediatric cancer–a report from the childhood cancer survivor study. *Cancer Epidemiol Biomarkers Prev.* 2010;19(1):170–181.
- 187. Rose-Felker K, Border WL, Hong BJ, Chow EJ. Cardio-oncology related to heart failure: pediatric considerations for cardiac dysfunction. *Heart Fail Clin*. 2017;13(2):311–325.
- 188. Armstrong GT, Joshi VM, Ness KK, Marwick TH, Zhang N, Srivastava D, Griffin BP, Grimm RA, Thomas J, Phelan D, Collier P, Krull KR, Mulrooney DA, Green DM, Hudson MM, Robison LL, Plana JC. Comprehensive echocardiographic detection of treatment-related cardiac dysfunction in adult survivors of childhood cancer: results from the St. Jude Lifetime Cohort Study. *J Am Coll Cardiol.* 2015;65(23):2511–2522.
- 189. Armstrong GT, Plana JC, Zhang N, Srivastava D, Green DM, Ness KK, Daniel Donovan F, Metzger ML, Arevalo A, Durand JB, Joshi V, Hudson MM, Robison LL, Flamm SD. Screening adult survivors of childhood cancer for cardiomyopathy: comparison of echocardiography and cardiac magnetic resonance imaging. *J Clin Oncol.* 2012;30(23):2876–2884.
- 190. Chen MH, Colan SD, Diller L. Cardiovascular disease: cause of morbidity and mortality in adult survivors of childhood cancers. *Circ Res.* 2011;108(5):619–628.

- 191. Lipshultz SE, Cochran TR, Franco VI, Miller TL. Treatment-related cardiotoxicity in survivors of childhood cancer. *Nat Rev Clin Oncol.* 2013;10(12):697–710.
- 192. Lipshultz SE, Franco VI, Miller TL, Colan SD, Sallan SE. Cardiovascular disease in adult survivors of childhood cancer. Annu Rev Med. 2015;66:161–176.
- 193. Sarosiek KA, Fraser C, Muthalagu N, Bhola PD, Chang W, McBrayer SK, Cantlon A, Fisch S, Golomb-Mello G, Ryan JA, Deng J, Jian B, Corbett C, Goldenberg M, Madsen JR, Liao R, Walsh D, Sedivy J, Murphy DJ, Carrasco DR, Robinson S, Moslehi J, Letai A. Developmental regulation of mitochondrial apoptosis by c-Myc governs age- and tissue-specific sensitivity to cancer therapeutics. *Cancer Cell*. 2017;31(1):142–156.
- 194. Toro-Salazar OH, Gillan E, O'Loughlin MT, Burke GS, Ferranti J, Stainsby J, Liang B, Mazur W, Raman SV, Hor KN. Occult cardiotoxicity in childhood cancer survivors exposed to anthracycline therapy. *Circ Cardiovasc Imaging*. 2013;6(6):873–880.
- 195. No authors listed. PD-1 inhibitor becomes "breakthrough therapy". Cancer Discov. 2013;3(7):OF14.
- 196. Ataca P, Arslan O. Chimeric antigen receptor T cell therapy in hematology. Turk J Haematol. 2015;32(4):285-294.
- 197. Batlevi CL, Matsuki E, Brentjens RJ, Younes A. Novel immunotherapies in lymphoid malignancies. *Nat Rev Clin Oncol.* 2016;13(1):25–40.
- 198. Gill S, June CH. Going viral: chimeric antigen receptor T-cell therapy for hematological malignancies. *Immunol Rev.* 2015;263(1):68–89.
- 199. Helleday T. PARP inhibitor receives FDA breakthrough therapy designation in castration resistant prostate cancer: beyond germline BRCA mutations. *Ann Oncol.* 2016;27(5):755–757.
- 200. Kim ES. Olmutinib: First global approval. Drugs. 2016;76(11):1153-1157.
- 201. Lim JS, Sundar R, Chenard-Poirier M, Lopez J, Yap TA. Emerging biomarkers for PD-1 pathway cancer therapy. *Biomark Med*. 2017;11(1):53–67.
- 202. Mahoney KM, Atkins MB. Prognostic and predictive markers for the new immunotherapies. *Oncology (Williston Park)*. 2014;3(28 Suppl):39–48.
- 203. Mahoney KM, Freeman GJ, McDermott DF. The next immune-checkpoint inhibitors: PD-1/PD-L1 blockade in melanoma. Clin Ther. 2015;37(4):764–782.
- 204. Mangini NS, Wesolowski R, Ramaswamy B, Lustberg MB, Berger MJ. Palbociclib: A novel cyclin-dependent kinase inhibitor for hormone receptor-positive advanced breast cancer. *Ann Pharmacother*. 2015;49(11):1252–1260.
- 205. Miller BC, Maus MV. CD19-targeted CAR T cells: a new tool in the fight against B cell malignancies. *Oncol Res Treat*. 2015;38(12):683–690.
- 206. Nichols M. New directions for drug-resistant breast cancer: the CDK4/6 inhibitors. *Future Med Chem.* 2015;7(12):1473–1481.
- 207. Santarpia M, Altavilla G, Rosell R. Alectinib: a selective, next-generation ALK inhibitor for treatment of ALK-rearranged non-small-cell lung cancer. Expert Rev Respir Med. 2015;9(3):255–268.
- 208. Skoulidis F, Papadimitrakopoulou VA. Personalized medicine tackles clinical resistance: alectinib in ALK-positive non-small cell lung cancer progressing on first-generation ALK inhibitor. *Clin Cancer Res.* 2016;22(21):5177–5182.
- 209. Kesselheim AS, Woloshin S, Eddings W, Franklin JM, Ross KM, Schwartz LM. Physicians' knowledge about FDA approval standards and perceptions of the "breakthrough therapy" designation. JAMA. 2016;315(14):1516–1518.
- 210. Krishnamurti T, Woloshin S, Schwartz LM, Fischhoff B. A randomized trial testing US Food and Drug Administration "Breakthrough" language. *JAMA Intern Med.* 2015;175(11):1856–1858.
- 211. Casartelli A, Lanzoni A, Comelli R, Crivellente F, Defazio R, Dorigatti R, Fasdelli N, Faustinelli I, Pagliarusco S, Tontodonati M, Cristofori P. A novel and integrated approach for the identification and characterization of drug-induced cardiac toxicity in the dog. *Toxicol Pathol.* 2011;39(2):361–371.
- 212. Lage A, Crombet T. Control of advanced cancer: the road to chronicity. Int J Environ Res Public Health. 2011;8(3):683-697.
- 213. Wickramasinghe CD, Nguyen KL, Watson KE, Vorobiof G, Yang EH. Concepts in cardio-oncology: definitions, mechanisms, diagnosis and treatment strategies of cancer therapy-induced cardiotoxicity. Future Oncol. 2016;12(6):855–870.
- 214. Pantziarka P, Bouche G, Meheus L, Sukhatme V, Sukhatme VP, Vikas P. The Repurposing Drugs in Oncology (ReDO) Project. *Ecancermedicalscience*. 2014;8:442.
- 215. Pantziarka P, Cairns L. Recycling existing drugs for cancer therapy: delivering low cost cancer care. *Ecancermedicalscience*. 2014;8:ed40.
- 216. Florescu M, Cinteza M, Vinereanu D. Chemotherapy-induced Cardiotoxicity. Maedica (Buchar). 2013;8(1):59-67.

- Dolci A, Dominici R, Cardinale D, Sandri MT, Panteghini M. Biochemical markers for prediction of chemotherapy-induced cardiotoxicity: systematic review of the literature and recommendations for use. Am J Clin Pathol. 2008;130(5):688–695.
- 218. Broder H, Gottlieb RA, Lepor NE. Chemotherapy and cardiotoxicity. Rev Cardiovasc Med. 2008;9(2):75-83.
- 219. Plana JC, Galderisi M, Barac A, Ewer MS, Ky B, Scherrer-Crosbie M, Ganame J, Sebag IA, Agler DA, Badano LP, Banchs J, Cardinale D, Carver J, Cerqueira M, DeCara JM, Edvardsen T, Flamm SD, Force T, Griffin BP, Jerusalem G, Liu JE, Magalhaes A, Marwick T, Sanchez LY, Sicari R, Villarraga HR, Lancellotti P. Expert consensus for multimodality imaging evaluation of adult patients during and after cancer therapy: a report from the American Society of Echocardiography and the European Association of Cardiovascular Imaging. Eur Heart J Cardiovasc Imaging. 2014;15(10):1063–1093.
- 220. Ewer MS, Ewer SM. Cardiotoxicity of anticancer treatments: what the cardiologist needs to know. *Nat Rev Cardiol*. 2010;7(10):564–575.
- 221. Billingham ME, Mason JW, Bristow MR, Daniels JR. Anthracycline cardiomyopathy monitored by morphologic changes. *Cancer Treat Rep.* 1978;62(6):865–872.
- 222. Bristow MR, Thompson PD, Martin RP, Mason JW, Billingham ME, Harrison DC. Early anthracycline cardiotoxicity. Am J Med. 1978;65(5):823–832.
- 223. Jiji RS, Kramer CM, Salerno M. Non-invasive imaging and monitoring cardiotoxicity of cancer therapeutic drugs. *J Nucl Cardiol*. 2012;19(2):377–388.
- 224. Sandhu H, Maddock H. Molecular basis of cancer-therapy-induced cardiotoxicity: introducing microRNA biomarkers for early assessment of subclinical myocardial injury. *Clin Sci (Lond)*. 2014;126(6):377–400.
- 225. Zajicek G. Cancer as a systemic disease. Med Hypotheses. 1978;4(3):193-207.
- 226. Murphy KT. The pathogenesis and treatment of cardiac atrophy in cancer cachexia. *Am J Physiol Heart Circ Physiol.* 2016;310(4):H466-H477.
- 227. McLean AS, Huang SJ, Salter M. Bench-to-bedside review: the value of cardiac biomarkers in the intensive care patient. *Crit Care*. 2008;12(3):215.
- 228. Witteles RM. Biomarkers as predictors of cardiac toxicity from targeted cancer therapies. *J Card Fail*. 2016;22(6):459–464.
- 229. Weber M, Hamm C. Role of B-type natriuretic peptide (BNP) and NT-proBNP in clinical routine. *Heart*. 2006;92(6):843–849.
- 230. Shah KS, Yang EH, Maisel AS, Fonarow GC. The role of biomarkers in detection of cardio-toxicity. *Curr Oncol Rep.* 2017;19(6):42.
- 231. Michos ED, Wilson LM, Yeh HC, Berger Z, Suarez-Cuervo C, Stacy SR, Bass EB. Prognostic value of cardiac troponin in patients with chronic kidney disease without suspected acute coronary syndrome: a systematic review and metaanalysis. Ann Intern Med. 2014;161(7):491–501.
- 232. Pavo N, Raderer M, Hulsmann M, Neuhold S, Adlbrecht C, Strunk G, Goliasch G, Gisslinger H, Steger GG, Hejna M, Kostler W, Zochbauer-Muller S, Marosi C, Kornek G, Auerbach L, Schneider S, Parschalk B, Scheithauer W, Pirker R, Drach J, Zielinski C, Pacher R. Cardiovascular biomarkers in patients with cancer and their association with all-cause mortality. *Heart*. 2015;101(23):1874–1880.
- 233. Lyon AR. Disparate worlds drawing closer together: cardiovascular biomarkers predict cancer outcomes in treatment-naive patients. *Heart*. 2015;101(23):1853–1854.
- 234. Wang X, Raulji P, Mohapatra SS, Patel R, Hellermann G, Kong X, Vera PL, Meyer-Siegler KL, Coppola D, Mohapatra S. Natriuretic peptide receptor a as a novel target for prostate cancer. *Mol Cancer*. 2011;10:56.
- 235. Wieshammer S, Dreyhaupt J, Muller D, Momm F, Jakob A. Limitations of N-terminal Pro-B-type natriuretic peptide in the diagnosis of heart disease among cancer patients who present with cardiac or pulmonary symptoms. *Oncology*. 2016;90(3):143–150.
- 236. Thavendiranathan P, Poulin F, Lim KD, Plana JC, Woo A, Marwick TH. Use of myocardial strain imaging by echocardiography for the early detection of cardiotoxicity in patients during and after cancer chemotherapy: a systematic review. J Am Coll Cardiol. 2014;63(25 Pt A):2751–2768.
- 237. Bottinor WJ, Migliore CK, Lenneman CA, Stoddard MF. Echocardiographic assessment of cardiotoxic effects of cancer therapy. *Curr Cardiol Rep.* 2016;18(10):99.
- 238. Chen-Scarabelli C, McRee C, Leesar MA, Hage FG, Scarabelli TM. Comprehensive review on cardio-oncology: role of multimodality imaging. *J Nucl Cardiol*. 2016;24(3):906–935.
- 239. Bezerra HG, Costa RA, Reiber JH, Rybicki FJ, Schoenhagen P, Stillman AA, De Sutter J, Van de Veire NR. Cardiovascular imaging 2013 in the International Journal of Cardiovascular Imaging. *Int J Cardiovasc Imaging*. 2014;30(4):683–695.

- 240. Collier P, Phelan D, Klein A. A test in context: myocardial strain measured by speckle-tracking echocardiography. *J Am Coll Cardiol*. 2017;69(8):1043–1056.
- 241. Nguyen KL, Hu P, Ennis DB, Shao J, Pham KA, Chen JJ. Cardiac MRI: a translational imaging tool for characterizing anthracycline-induced myocardial remodeling. *Curr Oncol Rep.* 2016;18(8):48.
- 242. Jeong D, Patel A, Francois CJ, Gage KL, Fradley MG. Cardiac magnetic resonance imaging in oncology. *Cancer Control.* 2017;24(2):147–160.
- 243. Pitekova B, Ravi S, Shah SV, Mladosievicova B, Heitner S, Ferencik M. The role of imaging with cardiac computed tomography in cardio-oncology patients. *Curr Cardiol Rep.* 2016;18(9):87.
- 244. Drazner MH. The progression of hypertensive heart disease. Circulation. 2011;123(3):327-334.
- 245. Lessick J, Sideman S, Azhari H, Shapiro E, Weiss JL, Beyar R. Evaluation of regional load in acute ischemia by three-dimensional curvatures analysis of the left ventricle. *Ann Biomed Eng.* 1993;21(2):147–161.
- 246. Beyar R, Weiss JL, Shapiro EP, Graves WL, Rogers WJ, Weisfeldt ML. Small apex-to-base heterogeneity in radius-to-thickness ratio by three-dimensional magnetic resonance imaging. *Am J Physiol.* 1993;264(1 Pt 2):H133–140.
- 247. Cooley D, A. What is concentric remodeling of the left ventricle with adequate systolic function? Ask a Texas Heart Institute Doctor. 2014. http://www.texasheart.org/HIC/HeartDoctor/answer_3342.cfm
- 248. Pugliese NR, Fabiani I, La Carrubba S, Conte L, Antonini-Canterin F, Colonna P, Caso P, Benedetto F, Santini V, Carerj S, Romano MF, Citro R, Di Bello V. Italian Society of Cardiovascular E. Classification and prognostic evaluation of left ventricular remodeling in patients with asymptomatic heart failure. Am J Cardiol. 2017;119(1):71–77.
- 249. Konstam MA, Kramer DG, Patel AR, Maron MS, Udelson JE. Left ventricular remodeling in heart failure: current concepts in clinical significance and assessment. *JACC Cardiovasc Imaging*. 2011;4(1):98–108.
- 250. Udelson JE, Konstam MA. Ventricular remodeling fundamental to the progression (and regression) of heart failure. *J Am Coll Cardiol*. 2011;57(13):1477–1479.
- 251. Seo Y, Ishizu T, Aonuma K. Current status of 3-dimensional speckle tracking echocardiography: a review from our experiences. *J Cardiovasc Ultrasound*. 2014;22(2):49–57.
- 252. Avelar E, Strickland CR, Rosito G. Role of imaging in cardio-oncology. Curr Treat Options Cardiovasc Med. 2017;19(6):46.
- 253. Luis SA, Yamada A, Khandheria BK, Speranza V, Benjamin A, Ischenko M, Platts DG, Hamilton-Craig CR, Haseler L, Burstow D, Chan J. Use of three-dimensional speckle-tracking echocardiography for quantitative assessment of global left ventricular function: a comparative study to three-dimensional echocardiography. *J Am Soc Echocardiogr.* 2014;27(3):285–291.
- 254. Lin ACW, Strugnell W, Riley R, Schmitt B, Zenge M, Schmidt M, Morris NR, Hamilton-Craig C. Higher resolution cine imaging with compressed sensing for accelerated clinical left ventricular evaluation. *J Magn Reson Imaging*. 2017;45(6):1693–1699.
- 255. Madore B, Hoge WS, Chao TC, Zientara GP, Chu R. Retrospectively gated cardiac cine imaging with temporal and spatial acceleration. *Magn Reson Imaging*. 2011;29(4):457–469.
- 256. Malayeri AA, Johnson WC, Macedo R, Bathon J, Lima JA, Bluemke DA. Cardiac cine MRI: quantification of the relationship between fast gradient echo and steady-state free precession for determination of myocardial mass and volumes. *J Magn Reson Imaging*. 2008;28(1):60–66.
- 257. Nakano S, Takahashi M, Kimura F, Senoo T, Saeki T, Ueda S, Tanno J, Senbonmatsu T, Kasai T, Nishimura S. Cardiac magnetic resonance imaging-based myocardial strain study for evaluation of cardiotoxicity in breast cancer patients treated with trastuzumab: a pilot study to evaluate the feasibility of the method. *Cardiol J.* 2016;23(3):270–280.
- 258. Bengel FM. Clinical cardiovascular molecular imaging. J Nucl Med. 2009;50(6):837-840.
- 259. Wollenweber T, Bengel FM. Cardiac molecular imaging. Semin Nucl Med. 2014;44(5):386-397.
- 260. Dobrucki LW, Sinusas AJ. PET and SPECT in cardiovascular molecular imaging. Nat Rev Cardiol. 2010;7(1):38-47.
- 261. Sinusas AJ. Molecular imaging in nuclear cardiology: translating research concepts into clinical applications. *Q J Nucl Med Mol Imaging*. 2010;54(2):230–240.
- 262. Saraste A, Nekolla SG, Schwaiger M. Cardiovascular molecular imaging: an overview. *Cardiovasc Res.* 2009;83(4):643–652
- 263. Sosnovik DE, Nahrendorf M, Weissleder R. Targeted imaging of myocardial damage. *Nat Clin Pract Cardiovasc Med.* 2008;2(5 Suppl):S63–S70.
- 264. Kasama S, Toyama T, Kurabayashi M. Usefulness of cardiac sympathetic nerve imaging using (123)lodine-Metaiodobenzylguanidine Scintigraphy for Predicting sudden cardiac death in patients with heart failure. *Int Heart J.* 2016;57(2):140–144.

- 265. Bengel FM, Thackeray JT. Altered cardiac innervation predisposes to ventricular arrhythmia: targeted positron emission tomography identifies risk in ischemic cardiomyopathy. *J Am Coll Cardiol*. 2014;63(2):150–152.
- 266. Thackeray JT, Bengel FM. Assessment of cardiac autonomic neuronal function using PET imaging. *J Nucl Cardiol*. 2013;20(1):150–165.
- 267. Wollenweber T, Bengel FM. Molecular imaging to predict ventricular arrhythmia in heart failure. *J Nucl Cardiol*. 2014;21(6):1096–1109.
- Guimaraes SL, Brandao SC, Andrade LR, Maia RJ, Markman Filho B. Cardiac sympathetic hyperactivity after chemotherapy: early sign of cardiotoxicity. Arq Bras Cardiol. 2015;105(3):228–234.
- 269. Stokkel MP, de Wit-van der Veen LJ, Boekhout A. I-123-MIBG myocardial imaging in trastuzumab-based cardiotoxicity: the first experience. *Nucl Med Commun.* 2013;34(1):19–24.
- 270. Swiger KJ, Singh J, Lenihan DJ. Cardiomyopathic toxicity from chemotherapy: is there an opportunity for preemptive intervention? Curr Treat Options Cardiovasc Med. 2017;19(3):20.
- 271. Kang J, Chang JH, Wilson BC, Veilleux I, Bai Y, DaCosta R, Kim K, Ha S, Lee JG, Kim JS, Lee SG, Kim SM, Lee HJ, Ahn YB, Han S, Yoo Y, Song TK. A prototype hand-held tri-modal instrument for in vivo ultrasound, photoacoustic, and fluorescence imaging. *Rev Sci Instrum.* 2015;86(3):034901.
- 272. Wang LV, Yao J. A practical guide to photoacoustic tomography in the life sciences. *Nat Methods*. 2016;13(8):627–638.
- 273. Valluru KS, Willmann JK. Clinical photoacoustic imaging of cancer. Ultrasonography. 2016;35(4):267–280.
- 274. Wang S, Lin J, Wang T, Chen X, Huang P. Recent Advances in photoacoustic imaging for deep-tissue biomedical applications. *Theranostics*. 2016;6(13):2394–2413.
- 275. Yao J, Xia J, Wang LV. Multiscale functional and molecular photoacoustic tomography. *Ultrason Imaging*. 2016;38(1):44–62
- 276. Zhang YS, Wang LV, Xia Y. Seeing through the surface: non-invasive characterization of biomaterial-tissue interactions using photoacoustic microscopy. *Ann Biomed Eng.* 2016;44(3):649–666.
- 277. Zhou Y, Yao J, Wang LV. Tutorial on photoacoustic tomography. J Biomed Opt. 2016;21(6):61007.
- 278. Sivasubramanian K, Pramanik M. High frame rate photoacoustic imaging at 7000 frames per second using clinical ultrasound system. *Biomed Opt Express*. 2016;7(2):312–323.
- 279. Dong B, Sun C, Zhang HF. Optical detection of ultrasound in photoacoustic imaging. *IEEE Trans Biomed Eng.* 2017;64(1):4–15.
- 280. Miao Q, Pu K. Emerging Designs of activatable photoacoustic probes for molecular imaging. *Bioconjug Chem.* 2016;27(12):2808–2823.
- 281. Liu C, Gong X, Lin R, Liu F, Chen J, Wang Z, Song L, Chu J. Advances in imaging techniques and genetically encoded probes for photoacoustic imaging. *Theranostics*. 2016;6(13):2414–2430.
- 282. Lemaster JE, Jokerst JV. What is new in nanoparticle-based photoacoustic imaging? Wiley Interdiscip Rev Nanomed Nanobiotechnol. 2017;9(1):https://doi.org/10.1002/wnan.1404
- 283. Hu S. Emerging concepts in functional and molecular photoacoustic imaging. Curr Opin Chem Biol. 2016;33:25-31.
- 284. Jeon M, Kim J, Kim C. Multiplane spectroscopic whole-body photoacoustic imaging of small animals in vivo. *Med Biol Eng Comput.* 2016;54(2–3):283–294.
- 285. Wang LV, Hu S. Photoacoustic tomography: in vivo imaging from organelles to organs. *Science*. 2012;335(6075):1458–1462.
- 286. Wang LV. Multiscale photoacoustic microscopy and computed tomography. Nat Photonics. 2009;3(9):503–509.
- 287. Wang D, Wu Y, Xia J. Review on photoacoustic imaging of the brain using nanoprobes. *Neurophotonics*. 2016;3(1):010901.
- 288. Li R, Wang P, Lan L, Lloyd FP, Jr., Goergen CJ, Chen S, Cheng JX. Assessing breast tumor margin by multispectral photoacoustic tomography. *Biomed Opt Express*. 2015;6(4):1273–1281.
- 289. Xia W, Piras D, Singh MK, van Hespen JC, van Leeuwen TG, Steenbergen W, Manohar S. Design and evaluation of a laboratory prototype system for 3D photoacoustic full breast tomography. *Biomed Opt Express*. 2013;4(11):2555–2569.
- 290. Pramanik M, Ku G, Li C, Wang LV. Design and evaluation of a novel breast cancer detection system combining both thermoacoustic (TA) and photoacoustic (PA) tomography. *Med Phys.* 2008;35(6):2218–2223.

- 291. Maeda A, Bu J, Chen J, Zheng G, DaCosta RS. Dual in vivo photoacoustic and fluorescence imaging of HER2 expression in breast tumors for diagnosis, margin assessment, and surgical guidance. *Mol Imaging*. 2015;14(1):7290201400043. https://doi.org/10.2310/7290.2014.00043.
- 292. Zhang M, Kim HS, Jin T, Yi A, Moon WK. Ultrasound-guided photoacoustic imaging for the selective detection of EGFR-expressing breast cancer and lymph node metastases. *Biomed Opt Express*. 2016;7(5):1920–1931.
- 293. Chen BZ, Yang JG, Wu D, Zeng DW, Yi Y, Yang N, Jiang HB. Photoacoustic imaging of cerebral hypoperfusion during acupuncture. *Biomed Opt Express*. 2015;6(9):3225–3234.
- 294. Nasiriavanaki M, Xia J, Wan H, Bauer AQ, Culver JP, Wang LV. High-resolution photoacoustic tomography of restingstate functional connectivity in the mouse brain. *Proc Natl Acad Sci U S A*. 2014;111(1):21–26.
- 295. Dahal S, Cullum BM. Characterization of multiphoton photoacoustic spectroscopy for subsurface brain tissue diagnosis and imaging. *J Biomed Opt.* 2016;21(4):47001.
- 296. van Es P, Biswas SK, Bernelot Moens HJ, Steenbergen W, Manohar S. Initial results of finger imaging using photoacoustic computed tomography. *J Biomed Opt.* 2014;19(6):060501.
- 297. Erpelding TN, Kim C, Pramanik M, Jankovic L, Maslov K, Guo Z, Margenthaler JA, Pashley MD, Wang LV. Sentinel lymph nodes in the rat: noninvasive photoacoustic and US imaging with a clinical US system. *Radiology*. 2010;256(1):102–110.
- 298. Taruttis A, Herzog E, Razansky D, Ntziachristos V. Real-time imaging of cardiovascular dynamics and circulating gold nanorods with multispectral optoacoustic tomography. *Opt Express*. 2010;18(19):19592–19602.
- 299. Kang D, Huang Q, Li Y. Noninvasive photoacoustic measurement of the composite indicator dilution curve for cardiac output estimation. *Biomed Opt Express*. 2015;6(2):536–543.
- 300. Laufer J, Norris F, Cleary J, Zhang E, Treeby B, Cox B, Johnson P, Scambler P, Lythgoe M, Beard P. In vivo photoacoustic imaging of mouse embryos. *J Biomed Opt.* 2012;17(6):061220.
- 301. Song KH, Wang LV. Noninvasive photoacoustic imaging of the thoracic cavity and the kidney in small and large animals. *Med Phys.* 2008;35(10):4524–4529.
- 302. Dean-Ben XL, Ford SJ, Razansky D. High-frame rate four dimensional optoacoustic tomography enables visualization of cardiovascular dynamics and mouse heart perfusion. *Sci Rep.* 2015;5:10133.
- 303. Horiguchi A, Tsujita K, Irisawa K, Kasamatsu T, Hirota K, Kawaguchi M, Shinchi M, Ito K, Asano T, Shinmoto H, Tsuda H, Ishihara M. A pilot study of photoacoustic imaging system for improved real-time visualization of neurovascular bundle during radical prostatectomy. *Prostate*. 2016;76(3):307–315.
- 304. Tang S, Chen J, Samant P, Stratton K, Xiang L. Transurethral photoacoustic endoscopy for prostate cancer: a simulation study. *IEEE Trans Med Imaging*. 2016;35(7):1780–1787.
- 305. Schwarz M, Buehler A, Aguirre J, Ntziachristos V. Three-dimensional multispectral optoacoustic mesoscopy reveals melanin and blood oxygenation in human skin in vivo. *J Biophotonics*. 2016;9(1–2):55–60.
- Valluru KS, Wilson KE, Willmann JK. Photoacoustic imaging in oncology: translational preclinical and early clinical experience. Radiology. 2016;280(2):332–349.
- 307. Diot G, Dima A, Ntziachristos V. Multispectral opto-acoustic tomography of exercised muscle oxygenation. *Opt Lett.* 2015;40(7):1496–1499.
- 308. Liu Y, Nie L, Chen X. Photoacoustic molecular imaging: from multiscale biomedical applications towards early-stage theranostics. *Trends Biotechnol.* 2016;34(5):420–433.
- 309. V KS, Hong XJ, V MM, M B, Tin A. Progress in anterior chamber angle imaging for glaucoma risk prediction a review on clinical equipment, practice and research. *Med Eng Phys.* 2016;38(12):1383–1391.
- 310. Bourantas CV, Jaffer FA, Gijsen FJ, van Soest G, Madden SP, Courtney BK, Fard AM, Tenekecioglu E, Zeng Y, van der Steen AFW, Emelianov S, Muller J, Stone PH, Marcu L, Tearney GJ, Serruys PW. Hybrid intravascular imaging: recent advances, technical considerations, and current applications in the study of plaque pathophysiology. *Eur Heart J*. 2017;38(6):400–412.
- 311. McNally LR, Mezera M, Morgan DE, Frederick PJ, Yang ES, Eltoum IE, Grizzle WE. Current and emerging clinical applications of Multispectral Optoacoustic Tomography (MSOT) in oncology. Clin Cancer Res. 2016;22(14):3432–3439.
- 312. Wang LV, Gao L. Photoacoustic microscopy and computed tomography: from bench to bedside. *Annu Rev Biomed Eng.* 2014;16:155–185.
- 313. La Riviere P. WE-H-206-00: advances in preclinical imaging. Med Phys. 2016;43(6):3847-3848.
- 314. Yao J, Wang LV. Breakthrough in Photonics 2013: photoacoustic tomography in biomedicine. *IEEE Photonics J*. 2014;6(2):pii: 0701006.

- 315. Wang L. WE-H-206-01: Photoacoustic tomography: multiscale imaging from organelles to patients by ultrasonically beating the optical diffusion limit. *Med Phys.* 2016;43(6):3847–3848.
- 316. Holotta M, Grossauer H, Kremser C, Torbica P, Volkl J, Degenhart G, Esterhammer R, Nuster R, Paltauf G, Jaschke W. Photoacoustic tomography of ex vivo mouse hearts with myocardial infarction. *J Biomed Opt.* 2011;16(3):036007.
- 317. Zhang C, Cheng YJ, Chen J, Wickline S, Wang LV. Label-free photoacoustic microscopy of myocardial sheet architecture. *J Biomed Opt.* 2012;17(6):060506.
- 318. Wang L, Maslov K, Xing W, Garcia-Uribe A, Wang LV. Video-rate functional photoacoustic microscopy at depths. *J Biomed Opt.* 2012;17(10):106007.
- 319. Zemp RJ, Song L, Bitton R, Shung KK, Wang LV. Realtime photoacoustic microscopy of murine cardiovascular dynamics. *Opt Express.* 2008;16(22):18551–18556.
- 320. Taruttis A, Claussen J, Razansky D, Ntziachristos V. Motion clustering for deblurring multispectral optoacoustic tomography images of the mouse heart. *J Biomed Opt.* 2012;17(1):016009.
- 321. Robinson WH, Lindstrom TM, Cheung RK, Sokolove J. Mechanistic biomarkers for clinical decision making in rheumatic diseases. *Nat Rev Rheumatol.* 2013;9(5):267–276.
- 322. Ritchlin CT. An integrative approach to biomarker development in psoriatic arthritis. *J Rheumatol Suppl.* 2015;93:43–47.
- 323. Antoine DJ, Dear JW, Lewis PS, Platt V, Coyle J, Masson M, Thanacoody RH, Gray AJ, Webb DJ, Moggs JG, Bateman DN, Goldring CE, Park BK. Mechanistic biomarkers provide early and sensitive detection of acetaminophen-induced acute liver injury at first presentation to hospital. *Hepatology*. 2013;58(2):777–787.
- 324. Wang W, Shi Q, Mattes WB, Mendrick DL, Yang X. Translating extracellular microRNA into clinical biomarkers for druginduced toxicity: from high-throughput profiling to validation. *Biomark Med.* 2015;9(11):1177–1188.
- 325. Kosaka N, Iguchi H, Yoshioka Y, Takeshita F, Matsuki Y, Ochiya T. Secretory mechanisms and intercellular transfer of microRNAs in living cells. *J Biol Chem*. 2010;285(23):17442–17452.
- 326. Iguchi H, Kosaka N, Yoshioka Y, Ochiya T. Secretory microRNAs as a versatile communication tool. *Commun Integr Biol.* 2010;3(5):478–481.
- 327. Smolensky MH, Peppas NA. Chronobiology, drug delivery, and chronotherapeutics. Adv Drug Deliv Rev. 2007;59(9–10):828–851.
- 328. Levi F, Okyar A. Circadian clocks and drug delivery systems: impact and opportunities in chronotherapeutics. *Expert Opin Drug Deliv*. 2011;8(12):1535–1541.
- 329. Lin SY, Kawashima Y. Current status and approaches to developing press-coated chronodelivery drug systems. *J Control Release*. 2012;157(3):331–353.
- 330. Mirza Aami M, Shakeel F, Iqbal Z. An overview of the regulatory and developmental strategies of chronotherapeutics. Therapeutic Innovation & Regulatory Science. 2016;50(4):450–454.
- 331. Levi F. The circadian timing system, a coordinator of life processes. Implications for the rhythmic delivery of cancer therapeutics. Conf Proc IEEE Eng Med Biol Soc. 2006;Suppl:6736–6739.
- 332. Fu L, Kettner NM. The circadian clock in cancer development and therapy. *Prog Mol Biol Transl Sci.* 2013;119:221–282.
- 333. Sukumaran S, Almon RR, DuBois DC, Jusko WJ. Circadian rhythms in gene expression: relationship to physiology, disease, drug disposition and drug action. *Adv Drug Deliv Rev.* 2010;62(9–10):904–917.
- 334. Bordyugov G, Westermark PO, Korencic A, Bernard S, Herzel H. Mathematical modeling in chronobiology. *Handb Exp Pharmacol.* 2013;217:335–357.
- 335. Okamura H. Suprachiasmatic nucleus clock time in the mammalian circadian system. *Cold Spring Harb Symp Quant Biol.* 2007;72:551–556.
- 336. Kaur G, Phillips C, Wong K, Saini B. Timing is important in medication administration: a timely review of chronotherapy research. *Int J Clin Pharm.* 2013;35(3):344–358.
- 337. Innominato PF, Roche VP, Palesh OG, Ulusakarya A, Spiegel D, Levi FA. The circadian timing system in clinical oncology. *Ann Med.* 2014;46(4):191–207.
- 338. Sewlall S, Pillay V, Danckwerts MP, Choonara YE, Ndesendo VM, du Toit LC. A timely review of state-of-the-art chronopharmaceuticals synchronized with biological rhythms. *Curr Drug Deliv*. 2010;7(5):370–388.
- 339. Youan BB. Chronopharmaceutical drug delivery systems: hurdles, hype or hope. *Adv Drug Deliv Rev.* 2010;62(9–10):898–903.

- 340. Ballesta A, Innominato PF, Dallmann R, Rand DA, Levi FA. Systems chronotherapeutics. *Pharmacol Rev.* 2017;69(2):161–199.
- 341. Traynor K, Newton DW, Hrushesky WJ, Reiter RJ. A pharmacist's primer on chronotherapeutics. *Am Pharm.* 1992;NS32(3):77–83.
- 342. Mandal AS, Biswas N, Karim KM, Guha A, Chatterjee S, Behera M, Kuotsu K. Drug delivery system based on chronobiology–a review. *J Control Release*. 2010;147(3):314–325.
- 343. Tran TH, Lee BJ. On-off pulsed oral drug-delivery systems: a possible tool for drug delivery in chronotherapy. *Ther Deliv*. 2011;2(9):1199–1214.
- 344. Sewlall S, Pillay V, Danckwerts MP, Choonara YE, Ndesendo VM, du Toit LC. A timely review of state-of-the-art chronopharmaceuticals synchronized with biological rhythms. *Curr Drug Deliv*. 2010;7(5):370–388.
- 345. Singh R, Sharma PK, Malviya R. Circadian cycle and chronotherapeutics: recent trend for the treatment of various biological disorders. *Recent Pat Drug Deliv Formul*. 2012;6(1):80–91.
- 346. Librodo P, Buckley M, Luk M, Bisso A. Chronotherapeutic drug delivery. J Infus Nurs. 2015;6(38 Suppl):S18-S23.
- 347. Levi F, Karaboue A, Gorden L, Innominato PF, Saffroy R, Giacchetti S, Hauteville D, Guettier C, Adam R, Bouchahda M. Cetuximab and circadian chronomodulated chemotherapy as salvage treatment for metastatic colorectal cancer (mCRC): safety, efficacy and improved secondary surgical resectability. *Cancer Chemother Pharmacol*. 2011;67(2):339–348
- 348. Ohdo S. Chronopharmaceutics: pharmaceutics focused on biological rhythm. Biol Pharm Bull. 2010;33(2):159-167.
- 349. Altinok A, Levi F, Goldbeter A. A cell cycle automaton model for probing circadian patterns of anticancer drug delivery. *Adv Drug Deliv Rev.* 2007;59(9-10):1036–1053.
- 350. Altinok A, Levi F, Goldbeter A. Identifying mechanisms of chronotolerance and chronoefficacy for the anticancer drugs 5-fluorouracil and oxaliplatin by computational modeling. *Eur J Pharm Sci.* 2009;36(1):20–38.
- 351. Innominato PF, Giacchetti S, Moreau T, Smaaland R, Focan C, Bjarnason GA, Garufi C, Iacobelli S, Tampellini M, Tumolo S, Carvalho C, Karaboue A, Levi F, Group AIC. Prediction of survival by neutropenia according to delivery schedule of oxaliplatin-5-Fluorouracil-leucovorin for metastatic colorectal cancer in a randomized international trial (EORTC 05963). Chronobiol Int. 2011;28(7):586–600.
- 352. Grimaldi B. Therapeutic implications of cancer-specific clocks. Ann Pharmacol Pharm. 2017;2(4):1-2.
- 353. De Mei C, Ercolani L, Parodi C, Veronesi M, Lo Vecchio C, Bottegoni G, Torrente E, Scarpelli R, Marotta R, Ruffili R, Mattioli M, Reggiani A, Wade M, Grimaldi B. Dual inhibition of REV-ERBbeta and autophagy as a novel pharmacological approach to induce cytotoxicity in cancer cells. *Oncogene*. 2015;34(20):2597–2608.
- 354. Sajan J, Cinu TA, Chacko AJ, Litty J, Jaseeda T. Chronotherapeutics and chronotherapeutic drug delivery systems. *Trop J Pharm Res.* 2009;8(5):467–475.
- 355. Levi F, Okyar A. Circadian clocks and drug delivery systems: impact and opportunities in chronotherapeutics. *Opin Drug Deliv*. 2011;8(12):1535–1541.
- 356. Ohdo S, Koyanagi S, Matsunaga N, Hamdan A. Molecular basis of chronopharmaceutics. *J Pharm Sci.* 2011;100(9):3560–3576.
- 357. Altun A, Ugur-Altun B. Melatonin: therapeutic and clinical utilization. Int J Clin Pract. 2007;61(5):835-845.
- 358. Zmrzljak UP, Rozman D. Circadian regulation of the hepatic endobiotic and xenobitoic detoxification pathways: the time matters. Chem Res Toxicol. 2012;25(4):811–824.
- 359. Arif IS, Hooper CL, Greco F, Williams AC, Boateng SY. Increasing doxorubicin activity against breast cancer cells using PPARgamma-ligands and by exploiting circadian rhythms. *Br J Pharmacol*. 2013;169(5):1178–1188.
- 360. Doroshaw JH. Anthracyclin antibiotic-stimulated superoxide, hydrogen peroxide, and hydroxyl radical production by NADH dehydrogenase. *Cancer Res.* 1983;43(10):4543–4551.
- 361. Kaiserova H, Simunek T, Sterba M, den Hartog GJ, Schroterova L, Popelova O, Gersl V, Kvasnickova E, Bast A. New iron chelators in anthracycline-induced cardiotoxicity. *Cardiovasc Toxicol*. 2007;7(2):145–150.
- 362. Venegas C, García JA, Escames G, Ortiz F, Lopez A, Doerrier C, Garcia-Corzo L, Lopez LC, Reiter RJ, Acuna-Castroviejo D. Extrapineal melatonin: analysis of its subcellular distribution and daily fluctuations. *J Pineal Res.* 2012;52(2):217–227.
- 363. Lissoni P, Barni S, Mandala M, Ardizzoia A, Paolorossi F, Vaghi M, Longarini R, Malugani F, Tancini G. Decreased toxicity and increased efficacy of cancer chemotherapy using the pineal hormone melatonin in metastatic solid tumour patients with poor clinical status. *Eur J Cancer.* 1999;35(12):1688–1692.
- 364. Lucius K, Trukova K. Integrative therapies and cardiovascular disease in the breast cancer population: a review, part 1. *Integr Med (Encinitas)*. 2015;14(4):22–29.

- 365. Piasek A, Bartoszek A, Namiesnik J. [Phytochemicals that counteract the cardiotoxic side effects of cancer chemotheraSpy] Substancje pochodzenia roslinnego przeciwdzialające kardiotoksyczności towarzyszacej chemioterapii nowotworow. Postepy Hig Med Dosw (Online). 2009;63:142–158.
- 366. Gandhi BR, Mundada AS, Gandhi PP. Chronopharmaceutics: as a clinically relevant drug delivery system. *Drug Deliv.* 2011;18(1):1–18.
- 367. Hradetzky D. Transdermal drug delivery devices for chronotherapy. Med Device Technol. 2008;19(3):45-47.
- 368. Lin SY. Chronotherapeutic approach to design a thermoresponsive membrane for transdermal drug delivery. *Curr Drug Deliv.* 2004;1(3):249–263.
- 369. Sutradhar KB, Sumi CD. Implantable microchip: the futuristic controlled drug delivery system. *Drug Deliv.* 2016;23(1):1–11.
- 370. Hamidi M, Azadi A, Rafiei P, Ashrafi H. A pharmacokinetic overview of nanotechnology-based drug delivery systems: an ADME-oriented approach. *Crit Rev Ther Drug Carrier Syst.* 2013;30(5):435–467.
- 371. Balmayor ER, Azevedo HS, Reis RL. Controlled delivery systems: from pharmaceuticals to cells and genes. *Pharm Res.* 2011;28(6):1241–1258.
- 372. Sharma A, Madhunapantula SV, Robertson GP. Toxicological considerations when creating nanoparticle-based drugs and drug delivery systems. Expert Opin Drug Metab Toxicol. 2012;8(1):47–69.
- 373. Patra CN, Swain S, Sruti J, Patro AP, Panigrahi KC, Beg S, Rao ME. Osmotic drug delivery systems: basics and design approaches. *Recent Pat Drug Deliv Formul*. 2013;7(2):150–161.
- 374. Sunil SA, Srikanth MV, Rao NS, Uhumwangho MU, Latha K, Murthy KV. Chronotherapeutic drug delivery systems: an approach to circadian rhythms diseases. *Curr Drug Deliv*. 2011;8(6):622–633.
- 375. Aragao-Santiago L, Bohr A, Delaval M, Dalla-Bona AC, Gessler T, Seeger W, Beck-Broichsitter M. Innovative formulations for controlled drug delivery to the lungs and the technical and toxicological challenges to overcome. *Curr Pharm Des.* 2016;22(9):1147–1160.
- 376. Vhora I, Patil S, Bhatt P, Gandhi R, Baradia D, Misra A. Receptor-targeted drug delivery: current perspective and challenges. *Ther Deliv.* 2014;5(9):1007–1024.
- 377. Robitzki AA, Kurz R. Biosensing and drug delivery at the microscale: novel devices for controlled and responsive drug delivery. *Handb Exp Pharmacol*. 2010;197:87–112.
- 378. Bale S, Khurana A, Reddy AS, Singh M, Godugu C. Overview on Therapeutic Applications of Microparticulate Drug Delivery Systems. Crit Rev Ther Drug Carrier Syst. 2016;33(4):309–361.
- 379. Cooper DL, Conder CM, Harirforoosh S. Nanoparticles in drug delivery: mechanism of action, formulation and clinical application towards reduction in drug-associated nephrotoxicity. *Expert Opin Drug Deliv.* 2014;11(10):1661–1680.
- 380. Gong C, Qi T, Wei X, Qu Y, Wu Q, Luo F, Qian Z. Thermosensitive polymeric hydrogels as drug delivery systems. *Curr Med Chem.* 2013;20(1):79–94.
- 381. Patil SS, Shahiwala A. Patented pulsatile drug delivery technologies for chronotherapy. *Expert Opin Ther Pat.* 2014;24(8):845–856.
- 382. Doherty KR, Wappel RL, Talbert DR, Trusk PB, Moran DM, Kramer JW, Brown AM, Shell SA, Bacus S. Multi-parameter in vitro toxicity testing of crizotinib, sunitinib, erlotinib, and nilotinib in human cardiomyocytes. *Toxicol Appl Pharmacol*. 2013;272(1):245–255.
- 383. van Hasselt J, Hansen J, Xiong Y, Shim Y, Pickard A, Jayaraman G, Hu B, Mahajan M, Gallo J, Sobie E, Birtwistle M, Azeloglu E, Iyengar R. Clinically-weighted transcriptomic signatures for protein kinase inhibitor associated cariotoxicity. *BioRxiv*. 2016. http://www.biorxiv.org/content/early/2016/09/19/075754.
- 384. Gwathmey JK, Davidoff AJ. Pathophysiology of cardiomyopathies: Part II. Drug-induced and other interventions. *Curr Opin Cardiol*. 1994;9(3):369–378.
- 385. Barr J, Vazquez-Chantada M, Alonso C, Perez-Cormenzana M, Mayo R, Galan A, Caballeria J, Martin-Duce A, Tran A, Wagner C, Luka Z, Lu SC, Castro A, Le Marchand-Brustel Y, Martinez-Chantar ML, Veyrie N, Clement K, Tordjman J, Gual P, Mato JM. Liquid chromatography-mass spectrometry-based parallel metabolic profiling of human and mouse model serum reveals putative biomarkers associated with the progression of nonalcoholic fatty liver disease. *J Proteome Res.* 2010;9(9):4501–4512.
- 386. Palomo L, Casal E, Royo F, Cabrera D, van-Liempd S, Falcon-Perez JM. Considerations for applying metabolomics to the analysis of extracellular vesicles. *Front Immunol.* 2014;5:651.
- 387. Friedman RC, Farh KK, Burge CB, Bartel DP. Most mammalian mRNAs are conserved targets of microRNAs. *Genome Res.* 2009;19(1):92–105.

- 388. Heijne WH, Kienhuis AS, van Ommen B, Stierum RH, Groten JP. Systems toxicology: applications of toxicogenomics, transcriptomics, proteomics and metabolomics in toxicology. *Expert Rev Proteomics*. 2005;2(5):767–780.
- 389. Schnackenberg LK, Beger RD. Metabolomic biomarkers: their role in the critical path. *Drug Discov Today Technol.* 2007;4(1):13–16.
- 390. Chaudhari U, Nemade H, Gaspar JA, Hescheler J, Hengstler JG, Sachinidis A. MicroRNAs as early toxicity signatures of doxorubicin in human-induced pluripotent stem cell-derived cardiomyocytes. *Arch Toxicol.* 2016;90(12):3087–3098.
- 391. Xi B, Wang T, Li N, Ouyang W, Zhang W, Wu J, Xu X, Wang X, Abassi YA. Functional cardiotoxicity profiling and screening using the xCELLigence RTCA cardio system. *J Lab Autom*. 2011;16(6):415–421.
- 392. Sharma A, Burridge PW, McKeithan WL, Serrano R, Shukla P, Sayed N, Churko JM, Kitani T, Wu H, Holmstrom A, Matsa E, Zhang Y, Kumar A, Fan AC, Del Alamo JC, Wu SM, Moslehi JJ, Mercola M, Wu JC. High-throughput screening of tyrosine kinase inhibitor cardiotoxicity with human induced pluripotent stem cells. *Sci Transl Med.* 2017;9(377):eaaf2584. https://doi.org/10.1126/scitranslmed.aaf2584
- 393. Penaranda A, Cantalapiedra IR, Bragard J, Echebarria B. Cardiac dynamics: a simplified model for action potential propagation. *Theor Biol Med Model*. 2012;9:50. https://doi.org/10.1186/1742-4682-9-50.
- 394. Williams DP, Shipley R, Ellis MJ, Webb S, Ward J, Gardner I, Creton S. Novel in vitro and mathematical models for the prediction of chemical toxicity. *Toxicol Res (Camb)*. 2013;2(1):40–59.
- 395. Pawar S, Mahajan K, Vavia P. In vivo anticancer efficacy and toxicity studies of a novel polymer conjugate N-acetyl glucosamine (NAG)-PEG-doxorubicin for targeted cancer therapy. AAPS PharmSciTech. 2017. https://doi.org/10.1208/s12249-017-0787-0 [Epub ahead of print].
- 396. Alderton PM, Gross J, Green MD. Comparative study of doxorubicin, mitoxantrone, and epirubicin in combination with ICRF-187 (ADR-529) in a chronic cardiotoxicity animal model. *Cancer Res.* 1992;52(1):194–201.
- 397. Ali M, Kamjoo M, Thomas HD, Kyle S, Pavlovska I, Babur M, Telfer BA, Curtin NJ, Williams KJ. The clinically active PARP inhibitor AG014699 ameliorates cardiotoxicity but does not enhance the efficacy of doxorubicin, despite improving tumor perfusion and radiation response in mice. *Mol Cancer Ther.* 2011;10(12):2320–2329.
- 398. Bellamy WT, Peng YM, Odeleye A, Ellsworth L, Xu MJ, Grogan TM, Weinstein RS. Cardiotoxicity in the SCID mouse following administration of doxorubicin and cyclosporin A. *Anticancer Drugs*. 1995;6(6):736–743.
- 399. Cao J, Cui S, Li S, Du C, Tian J, Wan S, Qian Z, Gu Y, Chen WR, Wang G. Targeted cancer therapy with a 2-deoxyglucose-based adriamycin complex. *Cancer Res.* 2013;73(4):1362–1373.
- 400. D'Avino C, Paciello R, Riccio G, Coppola C, Laccetti P, Maurea N, Raines RT, De Lorenzo C. Effects of a second-generation human anti-ErbB2 ImmunoRNase on trastuzumab-resistant tumors and cardiac cells. *Protein Eng Des Sel.* 2014;27(3):83–88.
- 401. Fedele C, Riccio G, Malara AE, D'Alessio G, De Lorenzo C. Mechanisms of cardiotoxicity associated with ErbB2 inhibitors. Breast Cancer Res Treat. 2012;134(2):595–602.
- 402. Mukherjee A, Dutta S, Sanyal U. Evaluation of antitumor efficacy and toxicity of novel 6-nitro-2-(3-chloropropyl)-1H-benz[de]isoquinoline-1,3-dione in vivo in mouse. *J Cancer Res Ther*. 2013;9(3):442-446.
- 403. Nasr M, Nafee N, Saad H, Kazem A. Improved antitumor activity and reduced cardiotoxicity of epirubicin using hepatocyte-targeted nanoparticles combined with tocotrienols against hepatocellular carcinoma in mice. *Eur J Pharm Biopharm*. 2014;88(1):216–225.
- 404. Ryu JS, Lee HS, Hong YS, Lee JJ, Sohn UD, Kim TY. In vivo antitumor efficacy and cardiotoxicity of novel anthracycline ID6105 (11-hydroxy-aclacinomycin X, Hyrubicin). *Cancer Chemother Pharmacol.* 2006;57(6):811–818.
- 405. Santucci L, Mencarelli A, Renga B, Ceccobelli D, Pasut G, Veronese FM, Distrutti E, Fiorucci S. Cardiac safety and antitumoral activity of a new nitric oxide derivative of pegylated epirubicin in mice. *Anticancer Drugs.* 2007;18(9):1081–1091.
- 406. Zhou ZY, Wan LL, Yang QJ, Han YL, Li D, Lu J, Guo C. Nilotinib reverses ABCB1/P-glycoprotein-mediated multidrug resistance but increases cardiotoxicity of doxorubicin in a MDR xenograft model. *Toxicol Lett.* 2016;259:124–132.
- 407. Fulbright JM, Egas-Bejar DE, Huh WW, Chandra J. Analysis of redox and apoptotic effects of anthracyclines to delineate a cardioprotective strategy. *Cancer Chemother Pharmacol.* 2015;76(6):1297–1307.
- 408. Goyal V, Bews H, Cheung D, Premecz S, Mandal S, Shaikh B, Best R, Bhindi R, Chaudhary R, Ravandi A, Thliveris J, Singal PK, Niraula S, Jassal DS. The cardioprotective role of N-Acetyl cysteine amide in the prevention of doxorubicin and trastuzumab-mediated cardiac dysfunction. *Can J Cardiol*. 2016;32(12):1513–1519.
- 409. Liu Y, Nguyen P, Baris TZ, Poirier MC. Molecular analysis of mitochondrial compromise in rodent cardiomyocytes exposed long term to nucleoside reverse transcriptase inhibitors (NRTIs). Cardiovasc Toxicol. 2012;12(2):123– 134.

- 410. Sacco G, Bigioni M, Evangelista S, Goso C, Manzini S, Maggi CA. Cardioprotective effects of zofenopril, a new angiotensin-converting enzyme inhibitor, on doxorubicin-induced cardiotoxicity in the rat. *Eur J Pharmacol.* 2001;414(1):71–78.
- 411. Zhu H, Luo P, Fu Y, Wang J, Dai J, Shao J, Yang X, Chang L, Weng Q, Yang B, He Q. Dihydromyricetin prevents cardiotoxicity and enhances anticancer activity induced by adriamycin. *Oncotarget*. 2015;6(5):3254–3267.
- 412. Zhu W, Zhang W, Shou W, Field LJ. P53 inhibition exacerbates late-stage anthracycline cardiotoxicity. *Cardiovasc Res.* 2014;103(1):81–89.
- 413. Milan DJ, Peterson TA, Ruskin JN, Peterson RT, MacRae CA. Drugs that induce repolarization abnormalities cause bradycardia in zebrafish. *Circulation*. 2003;107(10):1355–1358.
- 414. Boukens BJ, Rivaud MR, Rentschler S, Coronel R. Misinterpretation of the mouse ECG: 'musing the waves of Mus musculus'. *J Physiol.* 2014;592(21):4613–4626.
- 415. Shin JT, Fishman MC. From Zebrafish to human: modular medical models. *Annu Rev Genomics Hum Genet*. 2002;3:311–340.
- 416. Howe K, Clark MD, Torroja CF, Torrance J, Berthelot C, Muffato M, Collins JE, Humphray S, McLaren K, Matthews L, McLaren S, Sealy I, Caccamo M, Churcher C, Scott C, Barrett JC, Koch R, Rauch GJ, White S, Chow W, Kilian B, Quintais LT, Guerra-Assuncao JA, Zhou Y, Gu Y, Yen J, Vogel JH, Eyre T, Redmond S, Banerjee R, Chi J, Fu B, Langley E, Maguire SF, Laird GK, Lloyd D, Kenyon E, Donaldson S, Sehra H, Almeida-King J, Loveland J, Trevanion S, Jones M, Quail M, Willey D, Hunt A, Burton J, Sims S, McLay K, Plumb B, Davis J, Clee C, Oliver K, Clark R, Riddle C, Elliot D, Threadgold G, Harden G, Ware D, Begum S, Mortimore B, Kerry G, Heath P, Phillimore B, Tracey A, Corby N, Dunn M, Johnson C, Wood J, Clark S, Pelan S, Griffiths G, Smith M, Glithero R, Howden P, Barker N, Lloyd C, Stevens C, Harley J, Holt K, Panagiotidis G, Lovell J, Beasley H, Henderson C, Gordon D, Auger K, Wright D, Collins J, Raisen C, Dyer L, Leung K, Robertson L, Ambridge K, Leongamornlert D, McGuire S, Gilderthorp R, Griffiths C, Manthravadi D, Nichol S, Barker G, Whitehead S, Kay M, Brown J, Murnane C, Gray E, Humphries M, Sycamore N, Barker D, Saunders D, Wallis J, Babbage A, Hammond S, Mashreghi-Mohammadi M, Barr L, Martin S, Wray P, Ellington A, Matthews N, Ellwood M, Woodmansey R, Clark G, Cooper J, Tromans A, Grafham D, Skuce C, Pandian R, Andrews R, Harrison E, Kimberley A, Garnett J, Fosker N, Hall R, Garner P, Kelly D, Bird C, Palmer S, Gehring I, Berger A, Dooley CM, Ersan-Urun Z, Eser C, Geiger H, Geisler M, Karotki L, Kirn A, Konantz J, Konantz M, Oberlander M, Rudolph-Geiger S, Teucke M, Lanz C, Raddatz G, Osoegawa K, Zhu B, Rapp A, Widaa S, Langford C, Yang F, Schuster SC, Carter NP, Harrow J, Ning Z, Herrero J, Searle SM, Enright A, Geisler R, Plasterk RH, Lee C, Westerfield M, de Jong PJ, Zon LI, Postlethwait JH, Nusslein-Volhard C, Hubbard TJ, Roest Crollius H, Rogers J, Stemple DL. The zebrafish reference genome sequence and its relationship to the human genome. Nature. 2013;496(7446):498-503.
- 417. MacRae CA, Peterson RT. Zebrafish as tools for drug discovery. Nat Rev Drug Discov. 2015;14(10):721-731.
- 418. Lin KY, Chang WT, Lai YC, Liau I. Toward functional screening of cardioactive and cardiotoxic drugs with zebrafish in vivo using pseudodynamic three-dimensional imaging. *Anal Chem.* 2014;86(4):2213–2220.
- 419. Stainier DY, Fishman MC. The zebrafish as a model system to study cardiovascular development. *Trends Cardiovasc Med.* 1994;4(5):207–212.
- 420. Leong IU, Skinner JR, Shelling AN, Love DR. Zebrafish as a model for long QT syndrome: the evidence and the means of manipulating zebrafish gene expression. *Acta Physiol (Oxf)*. 2010;199(3):257–276.
- 421. Shin JT, Pomerantsev EV, Mably JD, MacRae CA. High-resolution cardiovascular function confirms functional orthology of myocardial contractility pathways in zebrafish. *Physiol Genomics*. 2010;42(2):300–309.
- 422. Vornanen M, Hassinen M. Zebrafish heart as a model for human cardiac electrophysiology. *Channels (Austin)*. 2016;10(2):101–110.
- 423. Wlodarchak N, Tariq R, Striker R. Comparative analysis of the human and zebrafish kinomes: focus on the development of kinase inhibitors. *Trends Cell Mol Biol.* 2015;10:49–75.
- 424. Rakshambikai R, Srinivasan N, Gadkari RA. Repertoire of protein kinases encoded in the genome of zebrafish shows remarkably large population of PIM kinases. *J Bioinform Comput Biol.* 2014;12(1):1350014.
- 425. Verkerk AO, Remme CA. Zebrafish: a novel research tool for cardiac (patho) electrophysiology and ion channel disorders. *Front Physiol.* 2012;3:255.
- 426. Burns CG, Milan DJ, Grande EJ, Rottbauer W, MacRae CA, Fishman MC. High-throughput assay for small molecules that modulate zebrafish embryonic heart rate. *Nat Chem Biol.* 2005;1(5):263–264.
- 427. Chi NC, Shaw RM, Jungblut B, Huisken J, Ferrer T, Arnaout R, Scott I, Beis D, Xiao T, Baier H, Jan LY, Tristani-Firouzi M, Stainier DY. Genetic and physiologic dissection of the vertebrate cardiac conduction system. *PLoS Biol.* 2008;6(5):e109.

- 428. Lin E, Craig C, Lamothe M, Sarunic MV, Beg MF, Tibbits GF. Construction and use of a zebrafish heart voltage and calcium optical mapping system, with integrated electrocardiogram and programmable electrical stimulation. *Am J Physiol Regul Integr Comp Physiol.* 2015;308(9):R755–R768.
- 429. Asnani A, Peterson RT. The zebrafish as a tool to identify novel therapies for human cardiovascular disease. *Dis Model Mech.* 2014;7(7):763–767.
- 430. Zheng PP, Severijnen LA, van der Weiden M, Willemsen R, Kros JM. A crucial role of caldesmon in vascular development in vivo. *Cardiovasc Res.* 2009;81(2):362–369.
- 431. Zheng PP, Severijnen LA, Willemsen R, Kros JM. Images in cardiovascular medicine. Functional cardiac phenotypes in zebrafish caldesmon morphants: a digital motion analysis. *Circulation*. 2009;120(17):e145–e146.
- 432. Zheng PP, Romme E, van der Spek PJ, Dirven CM, Willemsen R, Kros JM. Glut 1/SLC2A1 is crucial for the development of the blood-brain barrier in vivo. *Ann Neurol.* 2010;68(6):835–844.
- 433. Tsutsui H, Higashijima S, Miyawaki A, Okamura Y. Visualizing voltage dynamics in zebrafish heart. *J Physiol.* 2010;588(Pt 12):2017–2021.
- 434. Cheng H, Kari G, Dicker AP, Rodeck U, Koch WJ, Force T. A novel preclinical strategy for identifying cardiotoxic kinase inhibitors and mechanisms of cardiotoxicity. *Circ Res.* 2011;109(12):1401–1409.
- 435. McGrath P, Li CQ. Zebrafish: a predictive model for assessing drug-induced toxicity. *Drug Discov Today.* 2008;13(9-10):394-401.
- 436. Zhu JJ, Xu YQ, He JH, Yu HP, Huang CJ, Gao JM, Dong QX, Xuan YX, Li CQ. Human cardiotoxic drugs delivered by soaking and microinjection induce cardiovascular toxicity in zebrafish. *J Appl Toxicol*. 2014;34(2):139–148.
- 437. Akle V, Stankiewicz AJ, Kharchenko V, Yu L, Kharchenko PV, Zhdanova IV. Circadian kinetics of cell cycle progression in adult neurogenic niches of a diurnal vertebrate. *J Neurosci.* 2017;37(7):1900–1909.
- 438. Ben-Moshe Z, Foulkes NS, Gothilf Y. Functional development of the circadian clock in the zebrafish pineal gland. *Biomed Res Int.* 2014;2014:235781.
- 439. Cahill GM, Hurd MW, Batchelor MM. Circadian rhythmicity in the locomotor activity of larval zebrafish. *Neuroreport*. 1998;9(15):3445–3449.
- 440. Dang Y, Wang J, Giesy JP, Liu C. Responses of the zebrafish hypothalamic-pituitary-gonadal-liver axis PCR array to prochloraz are dependent on timing of sampling. *Aquat Toxicol*. 2016;175:154–159.
- 441. Dekens MP, Santoriello C, Vallone D, Grassi G, Whitmore D, Foulkes NS. Light regulates the cell cycle in zebrafish. *Curr Biol.* 2003;13(23):2051–2057.
- 442. Dickmeis T, Lahiri K, Nica G, Vallone D, Santoriello C, Neumann CJ, Hammerschmidt M, Foulkes NS. Glucocorticoids play a key role in circadian cell cycle rhythms. *PLoS Biol.* 2007;5(4):e78.
- 443. Hirayama J, Cardone L, Doi M, Sassone-Corsi P. Common pathways in circadian and cell cycle clocks: light-dependent activation of Fos/AP-1 in zebrafish controls CRY-1a and WEE-1. *Proc Natl Acad Sci U S A*. 2005;102(29):10194–10199.
- 444. Idda ML, Bertolucci C, Vallone D, Gothilf Y, Sanchez-Vazquez FJ, Foulkes NS. Circadian clocks: lessons from fish. *Prog Brain Res.* 2012;199:41–57.
- 445. Idda ML, Kage E, Lopez-Olmeda JF, Mracek P, Foulkes NS, Vallone D. Circadian timing of injury-induced cell proliferation in zebrafish. *PLoS One.* 2012;7(3):e34203.
- 446. Moore HA, Whitmore D. Circadian rhythmicity and light sensitivity of the zebrafish brain. PLoS One. 2014;9(1):e86176.
- 447. Noche RR, Lu PN, Goldstein-Kral L, Glasgow E, Liang JO. Circadian rhythms in the pineal organ persist in zebrafish larvae that lack ventral brain. *BMC Neurosci.* 2011;12:7.
- 448. Smadja Storz S, Tovin A, Mracek P, Alon S, Foulkes NS, Gothilf Y. Casein kinase 1delta activity: a key element in the zebrafish circadian timing system. *PLoS One*. 2013;8(1):e54189.
- 449. Tovin A, Alon S, Ben-Moshe Z, Mracek P, Vatine G, Foulkes NS, Jacob-Hirsch J, Rechavi G, Toyama R, Coon SL, Klein DC, Eisenberg E, Gothilf Y. Systematic identification of rhythmic genes reveals camk1gb as a new element in the circadian clockwork. *PLoS Genet*. 2012;8(12):e1003116.
- 450. Uchida Y, Hirayama J, Nishina H. A common origin: signaling similarities in the regulation of the circadian clock and DNA damage responses. *Biol Pharm Bull.* 2010;33(4):535–544.
- 451. Vallone D, Gondi SB, Whitmore D, Foulkes NS. E-box function in a period gene repressed by light. *Proc Natl Acad Sci U S A*. 2004;101(12):4106–4111.
- 452. Vansteensel MJ, Michel S, Meijer JH. Organization of cell and tissue circadian pacemakers: a comparison among species. Brain Res Rev. 2008;58(1):18–47.

- 453. Vatine G, Vallone D, Gothilf Y, Foulkes NS. It's time to swim! Zebrafish and the circadian clock. FEBS Lett. 2011;585(10):1485-1494.
- 454. Whitmore D, Cermakian N, Crosio C, Foulkes NS, Pando MP, Travnickova Z, Sassone-Corsi P. A clockwork organ. *Biol Chem.* 2000;381(9-10):793–800.
- 455. Xiao B, Cui LQ, Ding C, Wang H. Effects of lithium and 2,4-dichlorophenol on zebrafish: circadian rhythm disorder and molecular effects. *Zebrafish*. 2017;14(3):209–215.
- 456. Guerra-Varela J, Cabezas-Sainz P, Yebra-Pimentel E, Gutierrez-Lovera C, Cedron VP, Otero Obarrio MA, Sciara AA, Rodriguez N, Araujo J, Millan A, Sanchez L. "A zebra in the water": inspiring science in Spain. *Zebrafish*. 2016;13(4):241–247.
- 457. Doke SK, Dhawale SC. Alternatives to animal testing: a review. Saudi Pharm J. 2015;23(3):223-229.
- 458. Strahle U, Scholz S, Geisler R, Greiner P, Hollert H, Rastegar S, Schumacher A, Selderslaghs I, Weiss C, Witters H, Braunbeck T. Zebrafish embryos as an alternative to animal experiments—a commentary on the definition of the onset of protected life stages in animal welfare regulations. *Reprod Toxicol.* 2012;33(2):128–132.
- 459. Kacew S. Confounding factors in toxicity testing. *Toxicology*. 2001;160(1-3):87–96.
- 460. Kennedy-Lydon T, Rosenthal N. Cardiac regeneration: epicardial mediated repair. *Proc Biol Sci.* 2015; 282(1821):20152147.
- 461. Masters M, Riley PR. The epicardium signals the way towards heart regeneration. Stem Cell Res. 2014;13(3 Pt B):683–692.
- 462. Challa AK, Chatti K. Conservation and early expression of zebrafish tyrosine kinases support the utility of zebrafish as a model for tyrosine kinase biology. *Zebrafish*. 2013;10(3):264–274.
- 463. Hall AM, Orlow SJ. Degradation of tyrosinase induced by phenylthiourea occurs following Golgi maturation. *Pigment Cell Res.* 2005;18(2):122–129.
- 464. Wang WD, Wang Y, Wen HJ, Buhler DR, Hu CH. Phenylthiourea as a weak activator of aryl hydrocarbon receptor inhibiting 2,3,7,8-tetrachlorodibenzo-p-dioxin-induced CYP1A1 transcription in zebrafish embryo. *Biochem Pharmacol.* 2004;68(1):63–71.
- 465. Sarmah S, Marrs JA. Zebrafish as a vertebrate model system to evaluate effects of environmental toxicants on cardiac development and function. *Int J Mol Sci.* 2016;17(12):2123.
- 466. Sabeh MK, Kekhia H, Macrae CA. Optical mapping in the developing zebrafish heart. *Pediatr Cardiol.* 2012;33(6):916–922.
- 467. Ferl GZ, Theil FP, Wong H. Physibodies: a minireview on physiologically based pharmacokineyic models of small molecules and therapeutic antibodies: a minreview on fundamental concepts and applications. *Biopharm Drug Dispos*. 2016;37(2):75–92.
- 468. Gibiansky L, Gibiansky E. Target-mediated drug disposition model and its approximations for antibody-drug conjugates. *J Pharmacokinet Pharmacodyn.* 2014;41(1):35–47.
- 469. Haber DA, Gray NS, Baselga J. The evolving war on cancer. Cell. 2011;145(1):19-24.
- 470. MacConaill LE, Lindeman NI, Rollins BJ. Brave-ish New World-what's needed to make precision oncology a practical reality. JAMA Oncol. 2015;1(7):879–880.
- 471. Ellis LM, Hicklin DJ. Resistance to targeted therapies: refining anticancer therapy in the era of molecular oncology. *Clin Cancer Res.* 2009;15(24):7471–7478.
- 472. Gharwan H, Groninger H. Kinase inhibitors and monoclonal antibodies in oncology: clinical implications. *Nat Rev Clin Oncol.* 2016;13(4):209–227.
- 473. Kurian AW, Friese CR. Precision medicine in breast cancer care: an early glimpse of impact. *JAMA Oncol.* 2015;1(8):1109–1110.
- 474. Rubin R. A precision medicine approach to clinical trials. JAMA. 2016;316(19):1953-1955.
- 475. Blumenthal GM, Mansfield E, Pazdur R. Next-generation sequencing in oncology in the era of precision medicine. *JAMA Oncol.* 2016;2(1):13–14.
- 476. Harris MH, DuBois SG, Glade Bender JL, Kim A, Crompton BD, Parker E, Dumont IP, Hong AL, Guo D, Church A, Stegmaier K, Roberts CW, Shusterman S, London WB, MacConaill LE, Lindeman NI, Diller L, Rodriguez-Galindo C, Janeway KA. Multicenter feasibility study of tumor molecular profiling to inform therapeutic decisions in advanced pediatric solid tumors: the individualized cancer therapy (iCat) study. *JAMA Oncol.* 2016;2(5):608–615.
- 477. Horlings HM, Shah SP, Huntsman DG. Using somatic mutations to guide treatment decisions: context matters. *JAMA Oncol.* 2015;1(3):275–276.

- 478. Pasche B, Absher D. Whole-genome sequencing: a step closer to personalized medicine. *JAMA*. 2011;305(15):1596–1597
- 479. Prasad V, Fojo T, Brada M. Precision oncology: origins, optimism, and potential. *Lancet Oncol.* 2016;17(2):e81–e86.
- 480. Gibbs JB. Mechanism-based target identification and drug discovery in cancer research. *Science*. 2000;287(5460):1969–1973
- 481. Abrams J, Conley B, Mooney M, Zwiebel J, Chen A, Welch JJ, Takebe N, Malik S, McShane L, Korn E, Williams M, Staudt L, Doroshow J. National Cancer Institute's precision medicine initiatives for the new national clinical trials network. *Am Soc Clin Oncol Educ Book*. 2014:71–76.
- 482. Asic K. Dominant mechanisms of primary resistance differ from dominant mechanisms of secondary resistance to targeted therapies. *Crit Rev Oncol Hematol.* 2016;97:178–196.
- 483. Shoshan MC, Linder S. Target specificity and off-target effects as determinants of cancer drug efficacy. *Expert Opin Drug Metab Toxicol*. 2008;4(3):273–280.
- 484. Holohan C, Van Schaeybroeck S, Longley DB, Johnston PG. Cancer drug resistance: an evolving paradigm. *Nat Rev Cancer*. 2013;13(10):714–726.
- 485. Hammerman PS, Janne PA, Johnson BE. Resistance to epidermal growth factor receptor tyrosine kinase inhibitors in non-small cell lung cancer. *Clin Cancer Res.* 2009;15(24):7502–7509.
- 486. Milojkovic D, Apperley J. Mechanisms of resistance to imatinib and second-generation tyrosine inhibitors in chronic myeloid leukemia. *Clin Cancer Res.* 2009;15(24):7519–7527.
- 487. Janne PA, Gray N, Settleman J. Factors underlying sensitivity of cancers to small-molecule kinase inhibitors. *Nat Rev Drug Discov.* 2009;8(9):709–723.
- 488. Bracci L, Schiavoni G, Sistigu A, Belardelli F. Immune-based mechanisms of cytotoxic chemotherapy: implications for the design of novel and rationale-based combined treatments against cancer. *Cell Death Differ*. 2014;21(1):15–25.
- 489. Ray Chaudhuri A, Callen E, Ding X, Gogola E, Duarte AA, Lee JE, Wong N, Lafarga V, Calvo JA, Panzarino NJ, John S, Day A, Crespo AV, Shen B, Starnes LM, de Ruiter JR, Daniel JA, Konstantinopoulos PA, Cortez D, Cantor SB, Fernandez-Capetillo O, Ge K, Jonkers J, Rottenberg S, Sharan SK, Nussenzweig A. Replication fork stability confers chemoresistance in BRCA-deficient cells. *Nature*. 2016;535(7612):382–387.
- 490. Fojo T. Multiple paths to a drug resistance phenotype: mutations, translocations, deletions and amplification of coding genes or promoter regions, epigenetic changes and microRNAs. *Drug Resist Updat*. 2007;10(1–2):59–67.
- 491. Duesberg P, Stindl R, Hehlmann R. Origin of multidrug resistance in cells with and without multidrug resistance genes: chromosome reassortments catalyzed by aneuploidy. *Proc Natl Acad Sci U S A*. 2001;98(20):11283–11288.
- 492. Pao W, Miller VA, Politi KA, Riely GJ, Somwar R, Zakowski MF, Kris MG, Varmus H. Acquired resistance of lung adenocarcinomas to gefitinib or erlotinib is associated with a second mutation in the EGFR kinase domain. *PLoS Med.* 2005;2(3):e73.
- 493. Joyner MJ, Paneth N, Ioannidis JP. What happens when underperforming big ideas in research become entrenched? JAMA. 2016;316(13):1355–1356.
- 494. Chau NG, Lorch JH. Exceptional Responders inspire change: lessons for drug development from the bedside to the bench and back. *Oncologist*. 2015;20(7):699–701.
- 495. Prasad V. Perspective: the precision-oncology illusion. *Nature*. 2016;537(7619):S63.
- 496. Subbiah IM, Subbiah V. Exceptional responders: in search of the science behind the miracle cancer cures. *Future Oncol.* 2015;11(1):1–4.
- 497. Takebe N, McShane L, Conley B. Biomarkers: exceptional responders-discovering predictive biomarkers. *Nat Rev Clin Oncol.* 2015;12(3):132–134.
- 498. Mullard A. Learning from exceptional drug responders. *Nat Rev Drug Discov.* 2014;13(6):401–402.
- 499. Poh A. In search of exceptional responders. Cancer Discov. 2015;5(1):8.
- 500. Sheridan C. Cancer centers zero in on exceptional responders. Nat Biotechnol. 2014;32(8):703-704.
- 501. Damrongwatanasuk R, Fradley MG. Cardiovascular complications of targeted therapies for chronic myeloid leukemia. *Curr Treat Options Cardiovasc Med.* 2017;19(4):24.
- 502. Berger MF, Van Allen EM. Delivering on the promise of precision cancer medicine. Genome Med. 2016;8(1):110.
- 503. Lawrence MS, Stojanov P, Polak P, Kryukov GV, Cibulskis K, Sivachenko A, Carter SL, Stewart C, Mermel CH, Roberts SA, Kiezun A, Hammerman PS, McKenna A, Drier Y, Zou L, Ramos AH, Pugh TJ, Stransky N, Helman E, Kim J, Sougnez C, Ambrogio L, Nickerson E, Shefler E, Cortes ML, Auclair D, Saksena G, Voet D, Noble M, DiCara D, Lin P, Lichtenstein L,

- Heiman DI, Fennell T, Imielinski M, Hernandez B, Hodis E, Baca S, Dulak AM, Lohr J, Landau DA, Wu CJ, Melendez-Zajgla J, Hidalgo-Miranda A, Koren A, McCarroll SA, Mora J, Lee RS, Crompton B, Onofrio R, Parkin M, Winckler W, Ardlie K, Gabriel SB, Roberts CW, Biegel JA, Stegmaier K, Bass AJ, Garraway LA, Meyerson M, Golub TR, Gordenin DA, Sunyaev S, Lander ES, Getz G. Mutational heterogeneity in cancer and the search for new cancer-associated genes. *Nature*. 2013;499(7457):214–218.
- 504. Zhao J, Cheng F, Wang Y, Arteaga CL, Zhao Z. Systematic prioritization of druggable mutations in approximately 5000 genomes across 16 cancer types using a structural genomics-based approach. *Mol Cell Proteomics*. 2016;15(2):642–656.
- 505. McFarland CD, Korolev KS, Kryukov GV, Sunyaev SR, Mirny LA. Impact of deleterious passenger mutations on cancer progression. *Proc Natl Acad Sci U S A.* 2013;110(8):2910–2915.
- 506. McFarland CD, Mirny LA, Korolev KS. Tug-of-war between driver and passenger mutations in cancer and other adaptive processes. *Proc Natl Acad Sci U S A*. 2014;111(42):15138–15143.
- 507. Carragher NO, Unciti-Broceta A, Cameron DA. Advancing cancer drug discovery towards more agile development of targeted combination therapies. *Future Med Chem.* 2012;4(1):87–105.
- 508. Groenendijk FH, Bernards R. Drug resistance to targeted therapies: deja vu all over again. *Mol Oncol.* 2014;8(6):1067–1083.
- 509. Carr TH, McEwen R, Dougherty B, Johnson JH, Dry JR, Lai Z, Ghazoui Z, Laing NM, Hodgson DR, Cruzalegui F, Hollingsworth SJ, Barrett JC. Defining actionable mutations for oncology therapeutic development. *Nat Rev Cancer*. 2016;16(5):319–329.
- 510. Dancey JE, Bedard PL, Onetto N, Hudson TJ. The genetic basis for cancer treatment decisions. *Cell.* 2012;148(3):409–420.
- 511. Rask-Andersen M, Masuram S, Schioth HB. The druggable genome: evaluation of drug targets in clinical trials suggests major shifts in molecular class and indication. *Annu Rev Pharmacol Toxicol*. 2014;54:9–26.
- 512. Fuller SJ, Osborne SA, Leonard SJ, Hardyman MA, Vaniotis G, Allen BG, Sugden PH, Clerk A. Cardiac protein kinases: the cardiomyocyte kinome and differential kinase expression in human failing hearts. *Cardiovasc Res.* 2015;108(1):87–98.
- 513. Fedorov O, Muller S, Knapp S. The (un)targeted cancer kinome. Nat Chem Biol. 2010;6(3):166-169.
- 514. Lal H, Kolaja KL, Force T. Cancer genetics and the cardiotoxicity of the therapeutics. *J Am Coll Cardiol*. 2013;61(3):267–274.
- 515. Blaes A, Prizment A, Koene RJ, Konety S. Cardio-oncology related to heart failure: common risk factors between cancer and cardiovascular disease. *Heart Fail Clin*. 2017;13(2):367–380.
- 516. Chen MH, Kerkela R, Force T. Mechanisms of cardiac dysfunction associated with tyrosine kinase inhibitor cancer therapeutics. *Circulation*. 2008;118(1):84–95.
- 517. Force T, Kuida K, Namchuk M, Parang K, Kyriakis JM. Inhibitors of protein kinase signaling pathways: emerging therapies for cardiovascular disease. *Circulation*. 2004;109(10):1196–1205.
- Davis MI, Hunt JP, Herrgard S, Ciceri P, Wodicka LM, Pallares G, Hocker M, Treiber DK, Zarrinkar PP. Comprehensive analysis of kinase inhibitor selectivity. Nat Biotechnol. 2011;29(11):1046–1051.
- 519. Csermely P, Agoston V, Pongor S. The efficiency of multi-target drugs: the network approach might help drug design. *Trends Pharmacol Sci.* 2005;26(4):178–182.
- 520. Gross S, Rahal R, Stransky N, Lengauer C, Hoeflich KP. Targeting cancer with kinase inhibitors. *J Clin Invest*. 2015;125(5):1780–1789.
- 521. Pichler WJ, Beeler A, Keller M, Lerch M, Posadas S, Schmid D, Spanou Z, Zawodniak A, Gerber B. Pharmacological interaction of drugs with immune receptors: the p-i concept. *Allergol Int.* 2006;55(1):17–25.
- 522. Pichler WJ. Pharmacological interaction of drugs with antigen-specific immune receptors: the p-i concept. *Curr Opin Allergy Clin Immunol*. 2002;2(4):301–305.
- 523. Pichler WJ. The p-i concept: pharmacological interaction of drugs with immune receptors. World Allergy Organ J. 2008;1(6):96–102.
- 524. Handley ME, Rasaiyaah J, Chain BM, Katz DR. Mixed lineage kinases (MLKs): a role in dendritic cells, inflammation and immunity? Int J Exp Pathol. 2007;88(2):111–126.
- 525. Ott PA, Adams S. Small-molecule protein kinase inhibitors and their effects on the immune system: implications for cancer treatment. *Immunotherapy*. 2011;3(2):213–227.
- 526. Mohty M, Blaise D, Olive D, Gaugler B. Imatinib: the narrow line between immune tolerance and activation. *Trends Mol Med*. 2005;11(9):397–402.

- 527. Seggewiss R, Price DA, Purbhoo MA. Immunomodulatory effects of imatinib and second-generation tyrosine kinase inhibitors on T cells and dendritic cells: an update. *Cytotherapy*. 2008;10(6):633–641.
- 528. Seliger B, Massa C, Rini B, Ko J, Finke J. Antitumour and immune-adjuvant activities of protein-tyrosine kinase inhibitors. *Trends Mol Med.* 2010;16(4):184–192.
- 529. Kwilas AR, Donahue RN, Tsang KY, Hodge JW. Immune consequences of tyrosine kinase inhibitors that synergize with cancer immunotherapy. *Cancer Cell. Microenviron.* 2015;2(1):pii: e677.
- 530. Vogelstein B, Papadopoulos N, Velculescu VE, Zhou S, Diaz LA, Jr., Kinzler KW. Cancer genome landscapes. *Science*. 2013;339(6127):1546–1558.
- 531. Khurana E, Fu Y, Chakravarty D, Demichelis F, Rubin MA, Gerstein M. Role of non-coding sequence variants in cancer. *Nat Rev Genet.* 2016;17(2):93–108.
- 532. Weinhold N, Jacobsen A, Schultz N, Sander C, Lee W. Genome-wide analysis of noncoding regulatory mutations in cancer. *Nat Genet.* 2014;46(11):1160–1165.
- 533. Mularoni L, Sabarinathan R, Deu-Pons J, Gonzalez-Perez A, Lopez-Bigas N. OncodriveFML: a general framework to identify coding and non-coding regions with cancer driver mutations. *Genome Biol.* 2016;17(1):128.
- 534. Farh KK, Marson A, Zhu J, Kleinewietfeld M, Housley WJ, Beik S, Shoresh N, Whitton H, Ryan RJ, Shishkin AA, Hatan M, Carrasco-Alfonso MJ, Mayer D, Luckey CJ, Patsopoulos NA, De Jager PL, Kuchroo VK, Epstein CB, Daly MJ, Hafler DA, Bernstein BE. Genetic and epigenetic fine mapping of causal autoimmune disease variants. *Nature*. 2015;518(7539):337–343.
- 535. Ben-Bassat H, Klein BY. Inhibitors of tyrosine kinases in the treatment of psoriasis. *Curr Pharm Des.* 2000;6(9):933–942
- 536. Kontzias A, Laurence A, Gadina M, O'Shea JJ. Kinase inhibitors in the treatment of immune-mediated disease. F1000 Med Rep. 2012;4:5.
- 537. Patterson H, Nibbs R, McInnes I, Siebert S. Protein kinase inhibitors in the treatment of inflammatory and autoimmune diseases. *Clin Exp Immunol.* 2014;176(1):1–10.
- 538. Belmonte SL, Blaxall BC. G protein coupled receptor kinases as therapeutic targets in cardiovascular disease. *Circ Res.* 2011;109(3):309–319.
- 539. Satoh K, Fukumoto Y, Shimokawa H. Rho-kinase: important new therapeutic target in cardiovascular diseases. *Am J Physiol Heart Circ Physiol*. 2011;301(2):H287–H296.
- 540. Dhalla NS, Muller AL. Protein kinases as drug development targets for heart disease therapy. *Pharmaceuticals (Basel)*. 2010;3(7):2111–2145.
- 541. Mardinoglu A, Nielsen J. Systems medicine and metabolic modelling. J Intern Med. 2012;271(2):142–154.
- 542. Kell DB. The virtual human: towards a global systems biology of multiscale, distributed biochemical network models. *IUBMB Life*. 2007;59(11):689–695.
- 543. Goldstein DM, Gray NS, Zarrinkar PP. High-throughput kinase profiling as a platform for drug discovery. *Nat Rev Drug Discov*. 2008;7(5):391–397.
- 544. Patricelli MP, Szardenings AK, Liyanage M, Nomanbhoy TK, Wu M, Weissig H, Aban A, Chun D, Tanner S, Kozarich JW. Functional interrogation of the kinome using nucleotide acyl phosphates. *Biochemistry*. 2007;46(2):350–358.
- 545. Scheiber J, Chen B, Milik M, Sukuru SC, Bender A, Mikhailov D, Whitebread S, Hamon J, Azzaoui K, Urban L, Glick M, Davies JW, Jenkins JL. Gaining insight into off-target mediated effects of drug candidates with a comprehensive systems chemical biology analysis. *J Chem Inf Model*. 2009;49(2):308–317.
- 546. Jain V, Bahia J, Mohebtash M, Barac A. Cardiovascular complications associated with novel cancer immunotherapies. Curr Treat Options Cardiovasc Med. 2017;19(5):36.
- 547. Avelumab. https://www.uptodate.com/contents/avelumab-drug-information?source=search_result&search=Bavencio &selectedTitle=1~12.2017.
- 548. Avelumab. https://www.bavencio.com/en_US/for-patients-and-caregivers.html. 2017.
- 549. Durvalumab. https://www.uptodate.com/contents/durvalumab-drug-information?source=search_result&search= lmfinzi&selectedTitle=1~11.2017.
- 550. Varricchi G, Marone G, Mercurio V, Galdiero MR, Bonaduce D, Tocchetti CG. Immune checkpoint inhibitors and cardiac toxicity: an emerging issue. *Curr Med Chem.* 2017. https://doi.org/10.2174/0929867324666170407125017 [Epub ahead of print].
- 551. Michot JM, Bigenwald C, Champiat S, Collins M, Carbonnel F, Postel-Vinay S, Berdelou A, Varga A, Bahleda R, Hollebecque A, Massard C, Fuerea A, Ribrag V, Gazzah A, Armand JP, Amellal N, Angevin E, Noel N, Boutros C, Mateus C,

- Robert C, Soria JC, Marabelle A, Lambotte O. Immune-related adverse events with immune checkpoint blockade: a comprehensive review. *Eur J Cancer.* 2016;54:139–148.
- 552. Li Y, Li F, Jiang F, Lv X, Zhang R, Lu A, Zhang G. A mini-review for cancer immunotherapy: molecular understanding of PD-1/PD-L1 pathway & translational blockade of immune checkpoints. *Int J Mol Sci.* 2016;17(7).
- 553. Francisco LM, Sage PT, Sharpe AH. The PD-1 pathway in tolerance and autoimmunity. *Immunol Rev.* 2010;236:219–242.
- 554. Fife BT, Pauken KE. The role of the PD-1 pathway in autoimmunity and peripheral tolerance. *Ann N Y Acad Sci.* 2011;1217:45–59.
- 555. Gelao L, Criscitiello C, Esposito A, Goldhirsch A, Curigliano G. Immune checkpoint blockade in cancer treatment: a double-edged sword cross-targeting the host as an "innocent bystander". *Toxins (Basel)*. 2014;6(3):914–933.
- 556. Romano E, Romero P. The therapeutic promise of disrupting the PD-1/PD-L1 immune checkpoint in cancer: unleashing the CD8 T cell mediated anti-tumor activity results in significant, unprecedented clinical efficacy in various solid tumors. *J Immunother Cancer.* 2015;3:15.
- 557. Cappelli LC, Shah AA, Bingham CO, 3rd. Immune-related adverse effects of cancer immunotherapy- implications for rheumatology. *Rheum Dis Clin North Am.* 2017;43(1):65–78.
- 558. Atezolizumab. http://www.fda.gov/Drugs/InformationOnDrugs/ApprovedDrugs/ucm525780.htm.
- 559. Kong YC, Flynn JC. Opportunistic autoimmune disorders potentiated by immune-checkpoint inhibitors anti-CTLA-4 and anti-PD-1. Front Immunol. 2014;5:206.
- 560. Nishimura H, Okazaki T, Tanaka Y, Nakatani K, Hara M, Matsumori A, Sasayama S, Mizoguchi A, Hiai H, Minato N, Honjo T. Autoimmune dilated cardiomyopathy in PD-1 receptor-deficient mice. *Science*. 2001;291(5502):319–322.
- 561. Okazaki T, Tanaka Y, Nishio R, Mitsuiye T, Mizoguchi A, Wang J, Ishida M, Hiai H, Matsumori A, Minato N, Honjo T. Autoantibodies against cardiac troponin I are responsible for dilated cardiomyopathy in PD-1-deficient mice. *Nat Med.* 2003;9(12):1477–1483.
- 562. Kyi C, Postow MA. Checkpoint blocking antibodies in cancer immunotherapy. FEBS Lett. 2014;588(2):368–376.
- 563. Kleponis J, Skelton R, Zheng L. Fueling the engine and releasing the break: combinational therapy of cancer vaccines and immune checkpoint inhibitors. *Cancer Biol Med.* 2015;12(3):201–208.
- 564. Gad AZ, El-Naggar S, Ahmed N. Realism and pragmatism in developing an effective chimeric antigen receptor T-cell product for solid cancers. *Cytotherapy*. 2016;18(11):1382–1392.
- 565. O'Donnell JS, Long GV, Scolyer RA, Teng MW, Smyth MJ. Resistance to PD1/PDL1 checkpoint inhibition. *Cancer Treat Rev.* 2017;52:71–81.
- 566. News. 'Immunoswitch' particles may be key to more effective cancer immunotherapy. 2017. https://www.drugtargetreview.com/news/23754/immunoswitch-particles-may-key-effective-cancer-immunotherapy/?utm_medium =email&utm_campaign=DTR%20-%20Newsletter%2023%202017&utm_content=DTR%20-%20Newsletter%2023%202017+CID_baf709250c33534ce98ca5b551a0fceb&utm_source=Email%20marketing&utm_term=Immunoswitch%20particles%20may%20be%20key%20to%20more%20effective%20cancer%20immunotherapy.
- 567. Malissen N, Lacotte J, Du-Thanh A, Gaudy-Marqueste C, Guillot B, Grob JJ. Macrophage activation syndrome: a new complication of checkpoint inhibitors. *Eur J Cancer*. 2017;77:88–89.
- 568. Fousek K, Ahmed N. The evolution of T-cell therapies for solid malignancies. *Clin Cancer Res.* 2015;21(15):3384–3392.
- 569. Wu R, Forget MA, Chacon J, Bernatchez C, Haymaker C, Chen JQ, Hwu P, Radvanyi LG. Adoptive T-cell therapy using autologous tumor-infiltrating lymphocytes for metastatic melanoma: current status and future outlook. *Cancer J*. 2012;18(2):160–175.
- 570. Newick K, O'Brien S, Moon E, Albelda SM. CAR T cell therapy for solid tumors. Annu Rev Med. 2017;68:139-152.
- 571. Gill S, Maus MV, Porter DL. Chimeric antigen receptor T cell therapy: 25 years in the making. *Blood Rev.* 2016;30(3):157–167.
- 572. Figueroa JA, Reidy A, Mirandola L, Trotter K, Suvorava N, Figueroa A, Konala V, Aulakh A, Littlefield L, Grizzi F, Rahman RL, Jenkins MR, Musgrove B, Radhi S, D'Cunha N, D'Cunha LN, Hermonat PL, Cobos E, Chiriva-Internati M. Chimeric antigen receptor engineering: a right step in the evolution of adoptive cellular immunotherapy. *Int Rev Immunol.* 2015;34(2):154–187.
- 573. Dai H, Wang Y, Lu X, Han W. Chimeric antigen receptors modified T-cells for cancer therapy. *J Natl Cancer Inst.* 2016;108(7):pii: djv439.
- 574. Chmielewski M, Abken H. TRUCKs: the fourth generation of CARs. Expert Opin Biol Ther. 2015;15(8):1145-1154.

- 575. Han EQ, Li XL, Wang CR, Li TF, Han SY. Chimeric antigen receptor-engineered T cells for cancer immunotherapy: progress and challenges. *J Hematol Oncol.* 2013;6:47.
- 576. Maude SL, Teachey DT, Porter DL, Grupp SA. CD19-targeted chimeric antigen receptor T-cell therapy for acute lymphoblastic leukemia. *Blood*. 2015;125(26):4017–4023.
- 577. Cameron BJ, Gerry AB, Dukes J, Harper JV, Kannan V, Bianchi FC, Grand F, Brewer JE, Gupta M, Plesa G, Bossi G, Vuidepot A, Powlesland AS, Legg A, Adams KJ, Bennett AD, Pumphrey NJ, Williams DD, Binder-Scholl G, Kulikovskaya I, Levine BL, Riley JL, Varela-Rohena A, Stadtmauer EA, Rapoport AP, Linette GP, June CH, Hassan NJ, Kalos M, Jakobsen BK. Identification of a Titin-derived HLA-A1-presented peptide as a cross-reactive target for engineered MAGE A3-directed T cells. Sci Transl Med. 2013;5(197):197ra103.
- 578. King J, Waxman J, Stauss H. Advances in tumour immunotherapy. QJM. 2008;101(9):675-683.
- 579. Maude SL, Barrett D, Teachey DT, Grupp SA. Managing cytokine release syndrome associated with novel T cell-engaging therapies. *Cancer J.* 2014;20(2):119–122.
- 580. Maus MV, Levine BL. Chimeric antigen receptor T-cell therapy for the community oncologist. *Oncologist*. 2016;21(5):608-617.
- 581. Zhang E, Xu H. A new insight in chimeric antigen receptor-engineered T cells for cancer immunotherapy. *J Hematol Oncol.* 2017;10(1):1.
- 582. Shank BR, Do B, Sevin A, Chen SE, Neelapu SS, Horowitz SB. Chimeric Antigen receptor T cells in hematologic malignancies. *Pharmacotherapy*. 2017;37(3):334–345.
- 583. Lu YC, Robbins PF. Cancer immunotherapy targeting neoantigens. Semin Immunol. 2016;28(1):22-27.
- 584. Saida Y, Watanabe S, Tanaka T, Baba J, Sato K, Shoji S, Igarashi N, Kondo R, Okajima M, Koshio J, Ichikawa K, Nozaki K, Ishikawa D, Koya T, Miura S, Tanaka J, Kagamu H, Yoshizawa H, Nakata K, Narita I. Critical roles of chemoresistant effector and regulatory T cells in antitumor immunity after lymphodepleting chemotherapy. *J Immunol.* 2015;195(2):726–735.
- 585. de Jonge ME, Huitema AD, Rodenhuis S, Beijnen JH. Clinical pharmacokinetics of cyclophosphamide. *Clin Pharmacokinet*. 2005;44(11):1135–1164.
- 586. Dhesi S, Chu MP, Blevins G, Paterson I, Larratt L, Oudit GY, Kim DH. Cyclophosphamide-induced cardiomyopathy: a case report, review, and recommendations for management. *J Investig Med High Impact Case Rep.* 2013;1(1):2324709613480346.
- 587. Mehta J, Singhal S, Or R. Cyclophosphamide-induced cardiomyopathy during bone marrow transplantation for severe aplastic anemia. *J Assoc Physicians India*. 1994;42(2):159–160.
- 588. Taniguchi I. Clinical significance of cyclophosphamide-induced cardiotoxicity. Intern Med. 2005;44(2):89-90.
- 589. DiMaggio PA, Jr., Subramani A, Judson RS, Floudas CA. A novel framework for predicting in vivo toxicities from in vitro data using optimal methods for dense and sparse matrix reordering and logistic regression. *Toxicol Sci.* 2010;118(1):251–265.
- 590. Ngo NT, Brodie C, Giles C, Horncastle D, Klammer M, Lampert IA, Rahemtulla A, Naresh KN. The significance of tumour cell immunophenotype in myeloma and its impact on clinical outcome. *J Clin Pathol.* 2009;62(11):1009–1015.
- 591. Crow J, Youens K, Michalowski S, Perrine G, Emhart C, Johnson F, Gerling A, Kurtzberg J, Goodman BK, Sebastian S, Rehder CW, Datto MB. Donor cell leukemia in umbilical cord blood transplant patients: a case study and literature review highlighting the importance of molecular engraftment analysis. *J Mol Diagn.* 2010;12(4):530–537.
- 592. Csermely P, Korcsmaros T, Kiss HJ, London G, Nussinov R. Structure and dynamics of molecular networks: a novel paradigm of drug discovery: a comprehensive review. *Pharmacol Ther.* 2013;138(3):333–408.
- 593. Buffery D. The 2015 oncology drug pipeline: innovation drives the race to cure cancer. *Am Health Drug Benefits*. 2015;8(4):216–222.
- 594. Listing CD. https://www.cancer.gov/about-cancer/treatment/drugs. NCI Database.
- 595. Wang H, Yin Y, Wang P, Xiong C, Huang L, Li S, Li X, Fu L. Current situation and future usage of anticancer drug databases. *Apoptosis*. 2016;21(7):778–794.
- 596. Jovenaux L, Cautela J, Resseguier N, Pibarot M, Taouqi M, Orabona M, Pinto J, Peyrol M, Barraud J, Laine M, Bonello L, Paganelli F, Barlesi F, Thuny F. Practices in management of cancer treatment-related cardiovascular toxicity: a cardio-oncology survey. *Int J Cardiol.* 2017;241:387–392.
- 597. Lopez-Fernandez T, Martin Garcia A, Santaballa Beltran A, Montero Luis A, Garcia Sanz R, Mazon Ramos P, Velasco Del Castillo S, Lopez de Sa Areses E, Barreiro-Perez M, Hinojar Baydes R, Perez de Isla L, Valbuena Lopez SC, Dalmau Gonzalez-Gallarza R, Calvo-Iglesias F, Gonzalez Ferrer JJ, Castro Fernandez A, Gonzalez-Caballero E, Mitroi

C, Arenas M, Virizuela Echaburu JA, Marco Vera P, Iniguez Romo A, Zamorano JL, Plana Gomez JC, Lopez Sendon Henchel JL. Cardio-onco-hematology in clinical practice. Position paper and recommendations. *Rev Esp Cardiol (Engl Ed)*. 2017;70(6):474–486.

SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

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