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The Alzheimer's Disease Neuroimaging Initiative 3: continued innovation for clinical trial improvement

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Abstract

INTRODUCTION—The overall goal of the Alzheimer's Disease Neuroimaging Initiative (ADNI) is to validate biomarkers for Alzheimer's disease (AD) clinical trials. ADNI-3, beginning August 1, 2016, is a five year renewal of the current ADNI-2 study.

METHODS—ADNI-3 will follow current and additional subjects with normal cognition, mild cognitive impairment (MCI) and AD using innovative technologies such as tau imaging, magnetic resonance imaging (MRI) sequences for connectivity analyses, and a highly-automated immunoassay platform and mass spectroscopy approach for CSF biomarker analysis. A Systems Biology/Pathway approach will be used to identify genetic factors for subject selection/enrichment. Amyloid positron emission tomography (PET) scanning will be standardized using by the Centiloid method. The Brain Health Registry will help recruit subjects and monitor subject cognition.

RESULTS—Multi-modal analyses will provide insight into AD pathophysiology and disease progression.

DISCUSSION—ADNI-3 will aim to inform AD treatment trials and facilitate development of AD disease-modifying treatments.

Keywords

Alzheimer's disease; tau imaging; amyloid phenotyping; Centiloid method; Brain Health Registry; functional connectivity; clinical trial biomarkers

1. Introduction

The number of Americans with Alzheimer's disease (AD) is projected to increase from 5.2 million in 2016 to 13.8 million in 2050 [1]. On the current trajectory, the total cost for AD and other dementias during this time is predicted to rise from \$236 billion in 2015 to more than \$1 trillion in 2050 [1]. The Alzheimer's Association estimates that a treatment that

delays the onset of AD by five years would save an estimated \$935 billion in just the first 10 years. Indeed, the ~30% reductions in AD incidence in people >60 with a high school education reported from the Framingham study [2] are estimated to have resulted in >\$300 M in health care cost savings. The Alzheimer's Disease Neuroimaging Initiative (ADNI) was launched in 2004 with overarching aims of validating biomarkers for, and informing the design of, therapeutic trials in AD [3]. Funded by a unique public—private partnership, ADNI has now been running for 12 years, studying subjects with AD dementia (AD) and amnestic mild cognitive impairment (MCI), and cognitively normal (CN) elders. Beginning with an initial five-year study termed ADNI-1 [4], followed by a two-year extension which enrolled early MCI subjects, termed ADNI-GO, and then by a further five-year competitive renewal termed ADNI-2 [5]. Over this time, ADNI has made a profound impact on nearly all aspects of AD pathobiology and patient oriented research [6]. A five-year competitive renewal of ADNI-2, termed ADNI-3, will begin on August 1, 2016. ADNI has been the subject of several journal Special Issues [7–9], has generated over 1000 publications, and has been the subject of many iterative reviews of its progress and major milestones [3, 10, 11].

ADNI has played and continues to play a central role in improving treatment trials. The development of AD therapeutics has stalled in efforts to move beyond modestly effective symptomatic drugs, which are likely to have an impact at the dementia stage, to diseasemodifiers, requiring treatment at earlier pre-dementia or even presymptomatic stages of disease. There are many reasons for this failure, including issues of target selection, offtarget toxicity, subject selection, and insufficient pharmacokinetic and pharmacodynamics data to support trial design. In the past, AD and MCI were diagnosed clinically. Unfortunately, the clinical diagnosis lacked specificity (some patients diagnosed with MCI or dementia due to AD did not have AD pathology) and sensitivity (it is not possibly to identify cognitively normal subjects who have amyloid pathology using clinical measurements). One of the major accomplishments of ADNI has been to validate amyloid phenotyping, which detects the presence of β -amyloid (A β) pathology in living subjects with amyloid positron emission tomography (PET) [12-14] and cerebrospinal fluid (CSF) measures of Aβ [15–17]. In the past, subjects with clinical AD but without AD pathology have likely been enrolled in trials due to lack of amyloid phenotyping [18–20]. This has now changed, in part due to the contributions of ADNI. ADNI investigators have advanced the design of pre-dementia trials in the statistical [21-24], methodological [25-34], cognitive [35] and clinical [31, 32, 36, 37] literature, and with regulators [25] in the US and abroad facilitating the design of major completed and ongoing trials (avagacestat, gantanerumab, aducanumab, solanezumab, Anti Amyloid Treatment in Asymptomatic Alzheimer's Disease (A4) study, and A5 study). These advances have included the move from time-to-endpoint designs to continuous outcome measures as primaries [21, 25], the use of biomarker-based subject selection [22], single primary outcomes in prodromal trials [25], and cognitive endpoints in pre-dementia clinical trials [38–41].

Current clinical trials use measures of memory, cognition, and/or function as outcomes. These are imperfect measures as they are subject to high test-retest variability and are influenced by factors other than changes due to AD. A 'biological marker' that represents progression of AD pathology, correlates with symptomatology (especially memory and cognitive decline), and is not affected by non-AD pathology could be used as a surrogate

outcome measure, overcoming the problems associated with the current clinical measures. Despite some early hopes, brain Aβ burden, measured with amyloid PET or in CSF, does not correlate well with disease severity and has not yet been proven an effective surrogate outcome [42, 43]. Several counterintuitive results in which increased brain atrophy was reported in response to anti-Aβ therapy [44–49] have also ruled out volumetric magnetic resonance imaging (MRI) measures as satisfactory surrogate markers for therapeutic Aβ reduction. However, MR measures may still be a highly useful outcome measure for neuroprotective interventions. What therefore might be the ideal surrogate outcome measure? Pathological studies have indicated that AD symptomatology is more closely associated with tau tangles than AB deposits [43, 50, 51], and that brain tau correlates with cognition [43, 50, 51], suggesting a cause-effect relationship between tau tangles, synaptic dysfunction/synapse loss/neurodegeneration, and cognitive function. Recently, [18F]-T807 (also known as AV1451) and other tau PET ligands have been developed to detect tau in humans [52], raising the possibility that tau PET could ultimately be used as a surrogate marker for AD clinical trials. However, synaptic loss correlates most closely with cognitive impairment in AD so it is likely that CSF synaptic protein biomarkers such as neurogranin could play a significant role as a surrogate marker in concert with biomarkers of AD pathology [53].

ADNI-3 aims to directly improve clinical trials in four major ways. Firstly, it will study the use of tau PET for subject selection, as a baseline covariate and as a potential surrogate outcome measure. Secondly, it will investigate other signals such as change in CSF biomarkers, and functional imaging techniques ([18 F]-fluorodeoxyglucose-PET (FDG-PET), arterial spin labelling perfusion (ASL) MRI, and task-free functional MRI (TF-fMRI) that may detect treatment effects in phase 2 trials. [54]. Thirdly, it will directly address the lack of reliability of A β and tau phenotyping through the standardization of different amyloid PET tracers in the 'Centiloid project', and the development of new immunoassay platforms and mass spectroscopy techniques to improve the reliability of CSF analysis of A β and tau as well as assess the utility of CSF synaptic protein biomarkers such as neurogranin [55]. Lastly, in conjunction with the Brain Health Registry (BHR; www.brainhealthregistry.org [56]), it will implement web-based methods for recruitment and characterization of subjects to overcome the problems of slow recruitment, and high trial costs due to high "screen fails".

A secondary goal of ADNI-3 is to deepen our understanding of the progression and pathophysiology of AD. Continuation of the longitudinal phenotyping of AD from preclinical to prodromal to dementia stages will provide further insight into the disease process. Tau PET studies will investigate the relationship between A β and tau across the spectrum of cognition. The provision of biofluid samples to outside 'omics' projects such as lipidomics and metabolomics [57, 58] will facilitate a Systems biology/Pathway analysis approach to characterizing AD. Improvement of methods for measuring functional and structural connectivity using MRI 'Connectome-like' sequences will provide insight into the role of connectivity in this disease. Finally, ADNI-3 will continue to serve three Department of Defense ADNI grants investigating the relationship of traumatic brain injury and post-traumatic stress disorder on development of AD in Vietnam Veterans [59].

Given that ADNI has had a myriad of impacts on diverse aspects of AD [6] and in fields beyond the mandate of the Initiative, and that these impacts continue to multiply with the growing pool of shared data, is expected that ADNI-3 will make even greater inroads into enabling successful clinical trials of AD therapies and that its multimodal, longitudinal approach will provide important insights into disease progression.

2. Methods

2.1 Study design

ADNI-3 will be a continuation of ADNI-2, and is projected to retain 697 subjects from ADNI-2 (295 CNCN274 amnestic MCI, and 128 AD). To provide sufficient power and compensate for loss of drop outs, ADNI-3 will enroll an additional 133 CN (with and without the subjective memory concerns), 151 amnestic MCI (both early and late MCI) and 87 AD subjects (371 total new subjects) which will result in a cohort with 40% CN, 40% MCI and 20% AD subjects (Table 1). The BHR, an innovative internet-based registry for recruitment, assessment, and longitudinal monitoring for neuroscience studies, will be used for the identification and screening of eligible ADNI-3 participants, and for subsequent cognitive longitudinal monitoring using online neuropsychological tests. Enrollment is expected to be completed in 2017. There will be 3–4 years of follow-up for all newly enrolled subjects. MCI and AD subjects will be seen every year for a clinical visit and MRI. Most CN subjects will be seen on alternating years for clinical visits and phone checks although some CN subjects selected to receive a tau PET will have annual clinical visits. All MCI and CN subjects will receive a tau PET scan at baseline and in Year 5. Amyloid positive subjects may be randomly selected to receive two additional tau PET scans on the basis that A\beta level correlates with the level of brain tau tangles. AD subjects will receive a tau PET scan every year. All subjects receive amyloid PET and lumbar punctures every other year. MCI and AD subjects will receive a FDG-PET scan at baseline. CN and MCI subjects will be followed through the entire project while AD subjects will be followed for 24 months.

2.2 Clinical Core

The Clinical Core/Coordinating Center, led by Paul Aisen and Ronald Petersen, will continue to be responsible for managing the day-to-day clinical operations of ADNI, including the retention and follow-up of ADNI-2 subjects, and the enrolment of new subjects. Most clinical assessments from ADNI-2 will be continued in ADNI-3 to preserve the value of the longitudinal dataset. The proprietary Boston Naming Test will be replaced with the license-free Multilingual Naming Test [60]. The Core will also incorporate a performance-based functional assessment, the Financial Capacity Instrument.-Short Form [61].

The BHR will be used to assist recruitment of new enrollees and for at home longitudinal monitoring. Like the BHR, ADNI-3 will use theweb-based computerized cognitive assessment, CogState, which may be more sensitive early in the disease course, is simple to perform, culture free, and exhibits minimal learning effects [62, 63]. BHR currently has over

40,000 participants, 37% of whom are over age 55 and report memory concerns, and 29% of whom endorse a first degree relative with dementia.

2.3 PET Core

The PET Core, led by William Jagust, will establish harmonized protocols for the collection of tau PET and amyloid PET data, quality control of all acquired data, and data analysis. Florbetaben (Neuraceq) [64] will be incorporated as a second amyloid tracer in addition to florbetapir. Those subjects from ADNI-2 will continue to be evaluated with [18F]florbetapir whereas new subjects enrolled in ADNI-3 will be evaluated with longitudinal [18F]florbetaben. Tau PET data acquired using the tau ligand [18F]T807 will be examined using region-of-interest based approaches that recapitulate Braak staging in order to define tracer uptake by topography. The Banner Alzheimer Institute will examine whole-brain voxelwise approaches to tau PET data and will calculate a cerebral tau index to define extent and magnitude of brain tau deposition. The PET Core will continue FDG-PET imaging at the baseline examination on all subjects. Multimodal analysis including all PET and MRI data will form the basis for testing a proposed set of hypotheses that examine the ability of different PET biomarkers to predict outcomes at different stages of the AD pathophysiological process, how the biomarkers relate to one another, and how changes in biomarkers are related to clinical change.

- **2.3.1. Centiloid scale for the comparison of amyloid PET tracers**—To allow direct comparison of amyloid tracers ([18F]florbetapir, [18F]florbetaben, [¹¹C]Pittsburgh Compound (PiB)), regional brain standardized uptake value (SUV) outcomes will be converted into Centiloids, a process that reports amyloid tracer retention on a 0–100 scale using [¹¹C]PIB as a reference. We hypothesize that different amyloid imaging agents will have similar effect sizes for prediction of decline and detection of longitudinal change when placed on the Centiloid scale. Combining different amyloid imaging agents on this scale will increase statistical power.
- **2.3.2. Tau imaging**—A major feature of ADNI-3 is the incorporation of multisite, longitudinal tau PET imaging, which, in conjunction with clinical/cognitive assessments amyloid PET, MRI, CSF analysis, genetics, and widespread data sharing is expected to make a substantial contribution to our understanding of the role of tau in AD pathophysiology. Previous exploratory work using both region of interest (ROI) and voxel-based approaches have suggested a correlation between tau and cognition. In the ROI approach, patients were classified according to Braak staging determined by T1 MRI scan. Higher PIB retention and smaller hippocampal volumes were significantly associated with higher Braak stages even excluding those subjects with manifest clinical AD. There was a strong correlation (β = -3.191, p = .008) between performance on a standard laboratory episodic memory factor score and retention of [18 F]T807 in Braak stage 1/2 ROIs, and between global cognition and [18 F]T807 retention in Braak stage 3/4 and 5/6 ROIs (Figure 1) [65]. Voxelwise analysis using a cerebral tau index (CTI) differentiated between patient groups and young controls. In these data, CTI also correlated with the Mini Mental State Exam (MMSE) (r=.558 p=.016 in old controls, MCI, AD).

2.4 MRI Core

The MRI Core, led by Clifford Jack, will continue to improve clinical trials by developing and standardizing protocols, operationalizing definitions of clinical subgroups to accommodate biological heterogeneity, and optimizing inclusion/stratification and outcome metrics. ADNI-2 acquisition protocols will be expanded in ADNI-3 to include seven sequences (structural MRI, FLAIR and T2*GRE, diffusion MRI (dMRI), TF-fMRI, ASL perfusion MRI, and a high resolution coronal T2 fast spin echo) in all subjects. In addition, the dMRI and TF-fMRI protocols will be implemented using both standard and advanced protocols. The advanced dMRI and TF-fMRI acquisitions will be implemented on systems that are capable and will resemble those performed in the Human Connectome Project (HCP). ADNI-3 will be the largest multi-site, multi-vendor study to leverage several advanced MRI methods. HCP-like dMRI will offer more precise region-based fractional anisotropy and mean diffusivity measures as well as higher-fidelity characterization of white matter tract geometry. HCP-like TF-fMRI acquisitions will offer many advantages over standard TF-fMRI, including greater temporal and spatial resolution, less noisy connectivity measures, and time-varying connectivity. ASL perfusion MRI in ADNI-3 will be acquired using the 3D Pseudo-Continuous ASL protocol recommended by the International Society for Magnetic Resonance in Medicine perfusion work group, on systems where this is possiblee. High-resolution medial temporal lobe (MTL) subregion imaging offers quantification of changes in hippocampal subfields and parahippocampal gyrus subregions, which are the location of the earliest stages of tau pathology [66-69]. All ADNI-3 scans will be acquired at 3T.

2.5 Biomarker Core

The Biomarker Core, led by Leslie Shaw and John Trojanowski, will continue the ADNI Biofluid Biobank and distribution of samples to investigators, provide highly standardized $A\beta_{1-42}$, t-tau and p-tau₁₈₁ measurements on CSF samples, collaborate in the development of new tests for blood biomarkers (eg, ApoE4 protein in plasma; t-tau in plasma; exosomal fraction) and CSF biomarkers that detect co-pathologies such as Lewy Bodies and TDP43 proteinopathy in addition to other informative CSF proteins such as neurogranin.

Previously, the Core found close agreement in the measurement of CSF $A\beta1$ –42 with a codeveloped reference method that uses the same sample preparation steps but different HPLC and mass spectrometry instrumentation (Figure 2). An effort involving four laboratories in the Alzheimer's Association (AA) Global Biomarker Standardization Consortium (GBSC) showed very good concordance across 12 CSF pool samples (R²=0.98; average intralaboratory %CV of 4.7%; inter-laboratory %CV of 12.2%) that improved further when adjusted using a common calibrator[70]. Validation efforts for the fully-automated accuracy-and precision-based Roche Elecsys immunoassay platform for the measurement of $A\beta_{1-42}$, t-tau and p-tau₁₈₁ in all ADNI CSF samples are ongoing. One study over multiple centers has been completed and suggests that this method shows the best inter-laboratory performance reported for any method to date: the total measurement error across four participating laboratories and three different reagent lots and across five days of runs ranged from 2.2 to 5.1% over the five different CSF pools used for this study [71]. This system will be fully implemented in ADNI-3. In addition, a newly validated mass spectrometry assay for

 $A\beta_{1-42}$, $A\beta_{1-40}$ and $A\beta_{1-38}$ using a 2D-UPLC/MS-MS platform [55, 72], calibrated with a surrogate calibrator matrix prepared from artificial CSF plus 4 mg/mL bovine serum albumin, will be implemented in all ADNI-3 CSF samples. A mass spectrometry-based measurement of t-tau using a high sensitivity MRM mass spectrometry system will be validated and implemented for t-tau measurement in CSF of all ADNI-2 and ADNI-3 study subjects.

Using these improved methods of measurement, we predict that the CSF biomarkers alone and in combination for subject selection will reduce sample sizes thus improving efficiency for treatment trials, and that pathologic concentrations of $A\beta_{1-42}$ will predict decline in measures of memory, cognition and function. Furthermore, we hypothesize that rates of change of CSF $A\beta_{1-42}$ will be predictive of decline in memory, cognition, and function and ADNI-3 subjects.

2.6 Genetics Core

The Genetics Core, led by Andrew Saykin, will continue to provide genomic biosample banking and genotyping, identify and validate genetic markers to enhance clinical trial design and drug discovery, and provide an organizational framework to foster collaboration on genomic studies within ADNI-3. In addition to protocols employed in ADNI-2, peripheral blood mononuclear cells will be banked for use in development of induced pluripotent stem cells, functional drug development-related assays and other purposes. Systems biology modeling approaches yielding polygenic risk scores and gene pathway- and network-based metrics will be used to predict disease progression and outcomes. Associations between variation in the *MAPT* gene, encoding tau protein, and other pathways, and tau PET will be investigated and the influence of genetic variation on proteomics and metabolomics biomarker assays will be assessed.

We anticipate that ADNI data will demonstrate that the efficiency of clinical trials can be improved by enrichment with genetic markers beyond *APOE*, thereby reducing sample size, time required to complete trials, and lowering costs. Beginning with AD candidate genes nominated by large genome wide association studies (GWAS) [73] sequencing studies (e.g., *TREM2* [56–59], *PLD3* [60]), prior studies of AD endophenotypes in ADNI [74, 75] and other studies, followed by GWA (e.g., [76, 77]), we expect to identify variants that improve prediction of disease trajectory (i.e., onset, course, and outcome). We also predict that variants associated with biomarkers may yield clues to biological mechanisms and serve as potential targets for enrichment or therapeutic development. Examples of candidate trial enrichment markers that will be further studied in ADNI-3 include *BCHE* [76] and *IL1RAP* [78]. Additional data generated by ADNI-3 will enhance power by increasing participants with complete longitudinal data (and the range of phenotypes).

We envision that the innovative Systems Biology modeling approaches yielding polygenic risk scores and gene pathway- and network-based metrics will prove more powerful than single variants in predicting disease progression and outcomes, and that variation in the *MAPT* gene and other pathways will be associated with tau PET. Finally, we predict that controlling for the influence of genetic variation on proteomics (from studies of ADNI plasma and CSF samples) and metabolomics (through collaboration with the AD

Metabolomics Consortium), biomarker assays will improve the performance of *-omics* biomarkers in predicting disease progression and outcomes.

2.7 Neuropathology Core

The Neuropathology Core, led by John Morris and Nigel Cairns, will continue to foster and facilitate a voluntary brain autopsy for each ADNI participant at each site, to maintain a repository of frozen and fixed brain tissue from ADNI participants, and to validate the clinical, CSF biomarker and neuroimaging data of participants collected during the period of the grant. Additionally in ADNI-3, the contribution of common comorbidities (Lewy bodies, TDP-43 proteinopathy, vascular disease, hippocampal sclerosis, and tau astrogliopathy) to the variance in clinical, CSF biomarker, and neuroimaging data will be investigated, and the relationships between neuropathology and genomic data in multimodal studies of ADNI participants will be characterized. In participants who have undergone tau PET imaging, the spatial organization of tau burden in the postmortem brain will be correlated with tau PET data in collaboration with the PET Core.

Since the inception of the ADNI Neuropathology Core, the overall autopsy rate (number of autopsies/number of deaths) is 62%, with tissue received from 49 of the 52 patients who have come to autopsy at one of the ADNI sites. Currently, 42 of 57 ADNI-2 sites are fully operational to obtain autopsy consent and brain donation and we anticipate that during ADNI-3, a number of additional sites will become operational. Thus far, nearly half (18/41) of autopsied patients diagnosed with documented plaque and tangle AD pathology also had some form of comorbidity, most commonly Lewy bodies (synucleinopathy) and TDP-43 proteinopathy in the medial temporal lobe as well as argyrophilic grain disease and hippocampal sclerosis. We hypothesize that these comorbidities contribute to the variance in clinical, CSF biomarker, and neuroimaging data. By obtaining at least 100 total brains by the end of the next grant cycle, we calculate we will have 80% power to detect a correlation coefficient accounting for 8% of variation ($r = \pm 0.28$). We expect to gain insight into the relationship between these comorbidities and genetics using comprehensive integrative genomics and bioinformatics analyses with ADNI genome sequencing data in collaboration with the Genetics Core. We hypothesize that tau PET will be a better predictor of cognitive decline than other imaging and CSF biomarkers.

2.8 Biostatistics Core

The Biostatistics Core, led by Laurel Beckett, will carry out analyses of ADNI-3 data, separately, and in combination with data from previous phases with the aim of validating the potential of key clinical, functional, MRI, PET, CSF, and genetic biomarkers. Baseline biomarker distribution and performance, and longitudinal biomarker change will both be characterized as predictors of cognitive, functional, and biological change, of progression from CN to MCI and MCI to AD, and for use as inclusion/exclusion criteria, screening and stratification. New biostatistical methodologies will be developed to support ADNI-3 goals including those that account for missing and/or skipped data. A model for disease progression [79] will be extended to include tau PET data and to capture heterogeneity among sub-populations in the order of marker progression. Generalized Mallows models

will be extended to estimate the most likely sequence of progression events and its variation within and between sub-populations.

By combining results from PET tracers for amyloid, tau, and metabolism (FDG), and other measures, we predict that metabolism and tau, but not amyloid, will be correlated with cognition, and that metabolism will be negatively correlated with tau, but not correlated with amyloid. Moreover, we hypothesize that longitudinal changes in tau will be most strongly related to cognitive decline in all subject groups, whereas amyloid and metabolism will be very weakly related, and moderately related, respectively, to longitudinal cognitive decline. We hypothesize that future cognitive decline will be predicted by baseline PET, with amyloid and tau having stronger predictive power in controls than metabolism, and that all three PET imaging agents will be predictive in MCI, with tau the most predictive. Finally, we predict that in all groups except AD, individuals with more brain amyloid will have more tau in the neocortex and that longitudinally, those with amyloid will show increases in neocortical tau over time.

2.8.1. Assessment of the contribution of non-AD pathology—We predict that hypoperfusion, altered diffusion, and atrophy on structural MRI will predict concurrent tau PET ligand uptake, and that the severity of cerebrovascular disease and cerebral microbleeds will modify the ability of MRI and other modalities to predict future cognitive decline. In combination with PET, biofluids, and clinical measures, we will operationalize the definitions of subgroups within the ADNI population. Formal definitions of groups like SNAP (suspected non-Alzheimer's pathophysiology) [80, 81] and cerebrovascular phenotypes are needed to accommodate the biological heterogeneity within clinical trials populations. We anticipate that the development of new/optimized analysis methods and the creation of "AD-signature" summary numeric measures for each MR modality, and the optimization of inclusion/stratification, outcome metrics, and trial design will lower sample sizes and increase power of clinical trials.

2.9. Informatics Core

The Informatics Core, led by Arthur Toga, will continue to provide an information infrastructure to support the operational and research aims of each of the ADNI Cores and to provide data access and information resources for the wider research community. All new data acquired and produced as part of ADNI-3 will be stored in the Informatics Core ADNI repository at the Laboratory of Neuroimaging (LONI) at the University of Southern California. Data across all ADNI phases and data sources will be harmonized to enable coherent search functionality for interested investigators, and visualization on an interactive data platform. ADNI-3 data along with previous ADNI data will be provided in an 'analysis-ready' form for searching and downloads. They will also be provided for data aggregation efforts such as the Global Alzheimer's Association Interactive Network.

3. Conclusions

ADNI-3 is poised to make substantial contributions to the improvement of clinical trials for AD therapies through the use of a variety of innovative approaches that build on the basis of knowledge accumulated in the study over the last 12 years. Recent results with tau PET

suggest that increased misfolded or aggregated tau levels and brain (which occurs in concert with reduced levels of soluble brain tau [82] correlate with cognition, correlate with CSF tau levels, correlate with the presence of amyloidosis and show longitudinal progression. Thus tau PET is a promising biomarker to track change and treatment effects. Tau PET may also demonstrate the features required to serve as a surrogate marker in AD treatment trials, greatly reducing the number of subjects and length of trials. The development of the Centiloid approach for comparing amyloid tracers will facilitate use of multiple amyloid PET tracers for diagnosis and clinical trial enrollment. Use of the new automated Roche immunoassay platform by the Biomarker Core may reduce the previous variance problems with CSF measurements of $A\beta_{1-42}$ and tau/p-tau₁₈₁. The Brain Health Registry will facilitate recruitment of characterized participants for ADNI-3, and provide at home assessments. ADNI-3 will be augmented by the addition of MRI imaging techniques that target structural and functional connectivity in the brain and allow sub-regional examination of the medial temporal lobe. A Systems Biology approach promises to uncover novel genetic contributions to AD that may be used for subject selection or enrichment. Continuation of the longitudinal phenotyping across all stages of the disease process, and with the inclusion of tau imaging and connectivity analyses will provide further insights into AD pathophysiology. Therefore, the innovative approach of ADNI-3 will facilitate the validation of biomarkers for AD trials, enabling development of effective preventive or diseasemodifying treatments for AD. Ultimately, we hope to build models that enable the implementation of precision medicine approaches to stratifying patients for clinical trials and therapy including combination therapy [83, 84].

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References

- Association As. 2016 Alzheimer's disease facts and figures. Alzheimer's & Dementia. 2016; 12:459–509.
- Satizabal CL, Beiser AS, Chouraki V, Chene G, Dufouil C, Seshadri S. Incidence of Dementia over Three Decades in the Framingham Heart Study. N Engl J Med. 2016; 374:523–32. [PubMed: 26863354]

3. Weiner MW, Veitch DP, Aisen PS, Beckett LA, Cairns NJ, Cedarbaum J, et al. 2014 Update of the Alzheimer's Disease Neuroimaging Initiative: A review of papers published since its inception. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2015; 11:e1–e120.

- Mueller SG, Weiner MW, Thal LJ, Petersen RC, Jack C, Jagust W, et al. The Alzheimer's disease neuroimaging initiative. Neuroimaging Clin N Am. 2005; 15:869–77. xi–xii. [PubMed: 16443497]
- 5. Weiner MW, Aisen PS, Jack CR Jr, Jagust WJ, Trojanowski JQ, Shaw L, et al. The Alzheimer's disease neuroimaging initiative: progress report and future plans. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2010; 6:202–11 e7.
- Weiner MW, Veitch DP, Aisen PS, Beckett LA, Cairns NJ, Cedarbaum J, et al. Impact of the Alzheimer's Disease Neuroimaging Initiative, 2004 to 2014. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2015; 11:865–84.
- 7. Frisoni GB, Weiner MW. Alzheimer's Disease Neuroimaging Initiative special issue. Neurobiology of aging. 2010; 31:1259–62. [PubMed: 20570400]
- 8. Thies WH. Alzheimer's Disease Neuroimaging Initiative: A decade of progress in Alzheimer's disease. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2015; 11:727–9.
- Khachaturian ZS. Perspective on the Alzheimer's disease neuroimaging initiative: progress report and future plans. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2010; 6:199–201.
- 10. Weiner MW, Veitch DP, Aisen PS, Beckett LA, Cairns NJ, Green RC, et al. The Alzheimer's Disease Neuroimaging Initiative: a review of papers published since its inception. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2012; 8:S1–68.
- 11. Weiner MW, Veitch DP, Aisen PS, Beckett LA, Cairns NJ, Green RC, et al. The Alzheimer's Disease Neuroimaging Initiative: a review of papers published since its inception. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2013; 9:e111–94.
- 12. Landau SM, Breault C, Joshi AD, Pontecorvo M, Mathis CA, Jagust WJ, et al. Amyloid-beta imaging with Pittsburgh compound B and florbetapir: comparing radiotracers and quantification methods. Journal of nuclear medicine: official publication, Society of Nuclear Medicine. 2013; 54:70–7.
- 13. Landau SM, Fero A, Baker SL, Koeppe R, Mintun M, Chen K, et al. Measurement of longitudinal beta-amyloid change with 18F-florbetapir PET and standardized uptake value ratios. Journal of nuclear medicine: official publication, Society of Nuclear Medicine. 2015; 56:567–74.
- 14. Landau SM, Thomas BA, Thurfjell L, Schmidt M, Margolin R, Mintun M, et al. Amyloid PET imaging in Alzheimer's disease: a comparison of three radiotracers. European journal of nuclear medicine and molecular imaging. 2014; 41:1398–407. [PubMed: 24647577]
- 15. Shaw LM, Vanderstichele H, Knapik-Czajka M, Clark CM, Aisen PS, Petersen RC, et al. Cerebrospinal fluid biomarker signature in Alzheimer's disease neuroimaging initiative subjects. Annals of neurology. 2009; 65:403–13. [PubMed: 19296504]
- Shaw LM, Vanderstichele H, Knapik-Czajka M, Figurski M, Coart E, Blennow K, et al. Qualification of the analytical and clinical performance of CSF biomarker analyses in ADNI. Acta neuropathologica. 2011; 121:597–609. [PubMed: 21311900]
- 17. Kang JH, Korecka M, Toledo JB, Trojanowski JQ, Shaw LM. Clinical utility and analytical challenges in measurement of cerebrospinal fluid amyloid-beta(1–42) and tau proteins as Alzheimer disease biomarkers. Clin Chem. 2013; 59:903–16. [PubMed: 23519967]
- 18. Salloway S, Sperling R, Gregg K, Yu P, Joshi AD, Lu M, et al. Incidence and Clinical Progression of Placedbo-Treated Amyloid-Negative Subjects with Mild-Moderate Alzheimer's Disease: Results from the Phase 3 PET Sub-studies of bapineuzumab and solanezumab. AAIC. 2013; 9(4 Suppl):P888–9.
- 19. Vellas B, Carrillo MC, Sampaio C, Brashear HR, Siemers E, Hampel H, et al. Designing drug trials for Alzheimer's disease: what we have learned from the release of the phase III antibody trials: a report from the EU/US/CTAD Task Force. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2013; 9:438–44.
- 20. Siemers ER, Sundell KL, Carlson C, Case M, Sethuraman G, Liu-Seifert H, et al. Phase 3 solanezumab trials: Secondary outcomes in mild Alzheimer's disease patients. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2016; 12:110–20.

 Donohue MC, Gamst AC, Thomas RG, Xu R, Beckett L, Petersen RC, et al. The relative efficiency of time-to-threshold and rate of change in longitudinal data. Contemporary clinical trials. 2011; 32:685–93. [PubMed: 21554992]

- 22. Donohue MC, Gamst AC, Aisen PS. Requiring an amyloid-beta1–42 biomarker for prodromal Alzheimer's disease or mild cognitive impairment does not lead to more efficient clinical trials. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2011; 7:245–6. author reply 7–9.
- 23. Donohue MC, Aisen PS. Mixed model of repeated measures versus slope models in Alzheimer's disease clinical trials. The journal of nutrition, health & aging. 2012; 16:360–4.
- 24. Donohue MC, Jacqmin-Gadda H, Le Goff M, Thomas RG, Raman R, Gamst AC, et al. Estimating long-term multivariate progression from short-term data. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2014; 10:S400–10.
- Aisen PS, Andrieu S, Sampaio C, Carrillo M, Khachaturian ZS, Dubois B, et al. Report of the task force on designing clinical trials in early (predementia) AD. Neurology. 2011; 76:280–6.
 [PubMed: 21178097]
- 26. Vellas B, Hampel H, Rouge-Bugat ME, Grundman M, Andrieu S, Abu-Shakra S, et al. Alzheimer's disease therapeutic trials: EU/US Task Force report on recruitment, retention, and methodology. The journal of nutrition, health & aging. 2012; 16:339–45.
- 27. Bernick C, Cummings J, Raman R, Sun X, Aisen P. Age and rate of cognitive decline in Alzheimer disease: implications for clinical trials. Archives of neurology. 2012; 69:901–5. [PubMed: 22431834]
- 28. Grill JD, Raman R, Ernstrom K, Aisen P, Karlawish J. Effect of study partner on the conduct of Alzheimer disease clinical trials. Neurology. 2013; 80:282–8. [PubMed: 23255824]
- 29. Henley DB, Dowsett SA, Chen YF, Liu-Seifert H, Grill JD, Doody RS, et al. Alzheimer's disease progression by geographical region in a clinical trial setting. Alzheimer's research & therapy. 2015; 7:43.
- 30. Grill JD, Raman R, Ernstrom K, Aisen P, Dowsett SA, Chen YF, et al. Comparing recruitment, retention, and safety reporting among geographic regions in multinational Alzheimer's disease clinical trials. Alzheimer's research & therapy. 2015; 7:39.
- 31. Sperling RA, Jack CR Jr, Aisen PS. Testing the right target and right drug at the right stage. Sci Transl Med. 2011; 3:111cm33.
- 32. Vellas B, Aisen PS, Sampaio C, Carrillo M, Scheltens P, Scherrer B, et al. Prevention trials in Alzheimer's disease: an EU-US task force report. Progress in neurobiology. 2011; 95:594–600. [PubMed: 21925234]
- Aisen PS, Vellas B, Hampel H. Moving towards early clinical trials for amyloid-targeted therapy in Alzheimer's disease. Nature reviews Drug discovery. 2013; 12:324.
- 34. Andrieu S, Coley N, Lovestone S, Aisen PS, Vellas B. Prevention of sporadic Alzheimer's disease: lessons learned from clinical trials and future directions. The Lancet Neurology. 2015; 14:926–44. [PubMed: 26213339]
- 35. Salmon DP, Ferris SH, Thomas RG, Sano M, Cummings JL, Sperling RA, et al. Age and apolipoprotein E genotype influence rate of cognitive decline in nondemented elderly. Neuropsychology. 2013; 27:391–401. [PubMed: 23876113]
- 36. Sperling RA, Aisen PS, Beckett LA, Bennett DA, Craft S, Fagan AM, et al. Toward defining the preclinical stages of Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2011; 7:280–92.
- 37. Sperling RA, Rentz DM, Johnson KA, Karlawish J, Donohue M, Salmon DP, et al. The A4 study: stopping AD before symptoms begin? Sci Transl Med. 2014; 6:228fs13.
- 38. Sano M, Raman R, Emond J, Thomas RG, Petersen R, Schneider LS, et al. Adding Delayed Recall to the Alzheimer Disease Assessment Scale is Useful in Studies of Mild Cognitive Impairment But Not Alzheimer Disease. Alzheimer Dis Assoc Disord. 2011; 25:122–7. [PubMed: 20921876]
- Donohue MC, Sperling RA, Salmon DP, Rentz DM, Raman R, Thomas RG, et al. The preclinical Alzheimer cognitive composite: measuring amyloid-related decline. JAMA neurology. 2014; 71:961–70. [PubMed: 24886908]

40. Liu-Seifert H, Siemers E, Sundell K, Price K, Han B, Selzler K, et al. Cognitive and functional decline and their relationship in patients with mild Alzheimer's dementia. Journal of Alzheimer's disease: JAD. 2015; 43:949–55. [PubMed: 25125457]

- 41. Amariglio RE, Donohue MC, Marshall GA, Rentz DM, Salmon DP, Ferris SH, et al. Tracking early decline in cognitive function in older individuals at risk for Alzheimer disease dementia: the Alzheimer's Disease Cooperative Study Cognitive Function Instrument. JAMA neurology. 2015; 72:446–54. [PubMed: 25706191]
- 42. Hedden T, Oh H, Younger AP, Patel TA. Meta-analysis of amyloid-cognition relations in cognitively normal older adults. Neurology. 2013; 80:1341–8. [PubMed: 23547267]
- 43. Nelson PT, Alafuzoff I, Bigio EH, Bouras C, Braak H, Cairns NJ, et al. Correlation of Alzheimer disease neuropathologic changes with cognitive status: a review of the literature. J Neuropathol Exp Neurol. 2012; 71:362–81. [PubMed: 22487856]
- 44. Fox NC, Black RS, Gilman S, Rossor MN, Griffith SG, Jenkins L, et al. Effects of Abeta immunization (AN1792) on MRI measures of cerebral volume in Alzheimer disease. Neurology. 2005; 64:1563–72. [PubMed: 15883317]
- 45. Salloway S, Sperling R, Keren R, Porsteinsson AP, van Dyck CH, Tariot PN, et al. A phase 2 randomized trial of ELND005, scyllo-inositol, in mild to moderate Alzheimer disease. Neurology. 2011; 77:1253–62. [PubMed: 21917766]
- 46. Salloway S, Sperling R, Gilman S, Fox NC, Blennow K, Raskind M, et al. A phase 2 multiple ascending dose trial of bapineuzumab in mild to moderate Alzheimer disease. Neurology. 2009; 73:2061–70. [PubMed: 19923550]
- 47. Salloway S, Sperling R, Fox NC, Blennow K, Klunk W, Raskind M, et al. Two phase 3 trials of bapineuzumab in mild-to-moderate Alzheimer's disease. N Engl J Med. 2014; 370:322–33. [PubMed: 24450891]
- 48. Novak G, Einstein S, Tudor IC, Gregg K, Collins P, Wyman B, et al. The rate of clinical progression and brain atrophy is greater with increasing severity of Alzheimer's disease: Results from the volumetric MRI substudies of two phase III trials with bapineuzumab. Alzheimer's & Dementia: The Journal of the Alzheimer's Association. 2013; 9
- 49. Saumier D, Aisen PS, Gauthier S, Vellas B, Ferris SH, Duong A, et al. Lessons learned in the use of volumetric MRI in therapeutic trials in Alzheimer's disease: the ALZHEMED (Tramiprosate) experience. J Nutr Health Aging. 2009; 13:370–2. [PubMed: 19300884]
- 50. Braak H, Braak E. Staging of Alzheimer's disease-related neurofibrillary changes. Neurobiology of aging. 1995; 16:271–8. discussion 8–84. [PubMed: 7566337]
- 51. Ossenkoppele R, Schonhaut DR, Baker SL, O'Neil JP, Janabi M, Ghosh PM, et al. Tau, amyloid, and hypometabolism in a patient with posterior cortical atrophy. Annals of neurology. 2015; 77:338–42. [PubMed: 25448043]
- 52. Villemagne VL, Fodero-Tavoletti MT, Masters CL, Rowe CC. Tau imaging: early progress and future directions. The Lancet Neurology. 2015; 14:114–24. [PubMed: 25496902]
- 53. Portelius E, Zetterberg H, Skillback T, Tornqvist U, Andreasson U, Trojanowski JQ, et al. Cerebrospinal fluid neurogranin: relation to cognition and neurodegeneration in Alzheimer's disease. Brain: a journal of neurology. 2015
- 54. Sperling RA, Jack CR Jr, Black SE, Frosch MP, Greenberg SM, Hyman BT, et al. Amyloid-related imaging abnormalities in amyloid-modifying therapeutic trials: recommendations from the Alzheimer's Association Research Roundtable Workgroup. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2011; 7:367–85.
- 55. Kang JH, Korecka M, Figurski MJ, Toledo JB, Blennow K, Zetterberg H, et al. The Alzheimer's Disease Neuroimaging Initiative 2 Biomarker Core: A review of progress and plans. Alzheimers Dement. 2015; 11:772–91. [PubMed: 26194312]
- 56. Mackin RSIP, Truran D, Nosheny R, Finkey S, Mohlenhoff B, Flenniken D, Crothers R, Bickford D, Maruff P, Harel B, Weiner MW. Evaluating the potential of online neuropsychological assessments to accelerate neuroscience trials: The Brain Health Registry. American Journal of Psychiatry. 2016 Submitted.
- 57. Kaddurah-Daouk R, Zhu H, Sharma S, Bogdanov M, Rozen SG, Matson W, et al. Alterations in metabolic pathways and networks in Alzheimer's disease. Translational psychiatry. 2013; 9:18.

58. Motsinger-Reif AA, Zhu H, Kling MA, Matson W, Sharma S, Fiehn O, et al. Comparing metabolomic and pathologic biomarkers alone and in combination for discriminating Alzheimer's disease from normal cognitive aging. Acta Neuropathol Commun. 2013; 1:2051–5960.

- 59. Weiner MW, Veitch DP, Hayes J, Neylan T, Grafman J, Aisen PS, et al. Effects of traumatic brain injury and posttraumatic stress disorder on Alzheimer's disease in veterans, using the Alzheimer's Disease Neuroimaging Initiative. Alzheimers Dement. 2014; 10:S226–35. [PubMed: 24924673]
- 60. Ivanova I, Salmon DP, Gollan TH. The multilingual naming test in Alzheimer's disease: clues to the origin of naming impairments. Journal of the International Neuropsychological Society: JINS. 2013; 19:272–83. [PubMed: 23298442]
- 61. Gerstenecker A, Eakin A, Triebel K, Martin R, Swenson-Dravis D, Petersen RC, et al. Age and education corrected older adult normative data for a short form version of the Financial Capacity Instrument. Psychological assessment. 2016; 28:737–49. [PubMed: 26168311]
- 62. Maruff P, Lim YY, Darby D, Ellis KA, Pietrzak RH, Snyder PJ, et al. Clinical utility of the cogstate brief battery in identifying cognitive impairment in mild cognitive impairment and Alzheimer's disease. BMC Psychol. 2013; 1:30. [PubMed: 25566378]
- 63. Darby DG, Pietrzak RH, Fredrickson J, Woodward M, Moore L, Fredrickson A, et al. Intraindividual cognitive decline using a brief computerized cognitive screening test. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2012; 8:95–104.
- 64. Sabri O, Sabbagh MN, Seibyl J, Barthel H, Akatsu H, Ouchi Y, et al. Florbetaben PET imaging to detect amyloid beta plaques in Alzheimer disease: Phase 3 study. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2015
- 65. Scholl M, Lockhart SN, Schonhaut DR, O'Neil JP, Janabi M, Ossenkoppele R, et al. PET Imaging of Tau Deposition in the Aging Human Brain. Neuron. 2016; 89:971–82. [PubMed: 26938442]
- 66. de Flores R, La Joie R, Landeau B, Perrotin A, Mezenge F, de La Sayette V, et al. Effects of age and Alzheimer's disease on hippocampal subfields: comparison between manual and FreeSurfer volumetry. Human brain mapping. 2015; 36:463–74. [PubMed: 25231681]
- 67. Mueller S, Yushkevich P, Wang L, Van Leemput K, Mezher A, Iglesias JE, et al. Collaboration for a systematic comparison of different techniques to measure subfield volumes: Announcement and first results. Alzheimer's & Dementia: The Journal of the Alzheimer's Association. 2013; 9:P51.
- 68. Pluta J, Yushkevich P, Das S, Wolk D. In vivo analysis of hippocampal subfield atrophy in mild cognitive impairment via semi-automatic segmentation of T2-weighted MRI. Journal of Alzheimer's disease: JAD. 2012; 31:85–99. [PubMed: 22504319]
- 69. Yushkevich PA, Pluta JB, Wang H, Xie L, Ding SL, Gertje EC, et al. Automated volumetry and regional thickness analysis of hippocampal subfields and medial temporal cortical structures in mild cognitive impairment. Human brain mapping. 2015; 36:258–87. [PubMed: 25181316]
- 70. Pannee J, Gobom J, Shaw LM, Korecka M, Chambers EE, Lame M, et al. Round robin test on quantification of amyloid-beta 1–42 in cerebrospinal fluid by mass spectrometry. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2016; 12:55–9.
- 71. Bittner T, Zetterberg H, Teunissen C, Ostlund RE, Militello M, Andreasson U, et al. Technical performance of a novel, fully automated electrochemiluminescence immunoassay for the quantitation of β -amyloid (1–42) in human cerebrospinal fluid. Alz Dementia. 2015 in press.
- 72. Korecka M, Waligorska T, Figurski M, Toledo JB, Arnold SE, Grossman M, et al. Qualification of a surrogate matrix-based absolute quantification method for amyloid-beta(4)(2) in human cerebrospinal fluid using 2D UPLC-tandem mass spectrometry. Journal of Alzheimer's disease: JAD. 2014; 41:441–51. [PubMed: 24625802]
- 73. Lambert JC, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C, et al. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. Nature genetics. 2013; 45:1452–8. [PubMed: 24162737]
- 74. Saykin AJ, Shen L, Yao X, Kim S, Nho K, Risacher SL, et al. Genetic studies of quantitative MCI and AD phenotypes in ADNI: Progress, opportunities, and plans. Alzheimer's & dementia: the journal of the Alzheimer's Association. 2015; 11:792–814.
- 75. Shen L, Thompson PM, Potkin SG, Bertram L, Farrer LA, Foroud TM, et al. Genetic analysis of quantitative phenotypes in AD and MCI: imaging, cognition and biomarkers. Brain imaging and behavior. 2014; 8:183–207. [PubMed: 24092460]

 Mukherjee S, Kim S, Ramanan VK, Gibbons LE, Nho K, Glymour MM, et al. Gene-based GWAS and biological pathway analysis of the resilience of executive functioning. Brain imaging and behavior. 2014; 8:110–8. [PubMed: 24072271]

- 77. Ramanan VK, Risacher SL, Nho K, Kim S, Shen L, McDonald BC, et al. GWAS of longitudinal amyloid accumulation on 18F-florbetapir PET in Alzheimer's disease implicates microglial activation gene IL1RAP. Brain: a journal of neurology. 2015; 138:3076–88. [PubMed: 26268530]
- Jahanshad N, Rajagopalan P, Hua X, Hibar DP, Nir TM, Toga AW, et al. Genome-wide scan of healthy human connectome discovers SPON1 gene variant influencing dementia severity. Proceedings of the National Academy of Sciences of the United States of America. 2013; 110:4768–73. [PubMed: 23471985]
- 79. Donohue MC, Jacqmin-Gadda H, Le Goff M, Thomas RG, Raman R, Gamst AC, et al. Estimating long-term multivariate progression from short-term data. Alzheimers Dement. 2014; 10:S400–10. [PubMed: 24656849]
- 80. Jack CR Jr, Knopman DS, Chetelat G, Dickson D, Fagan AM, Frisoni GB, et al. Suspected non-Alzheimer disease pathophysiology concept and controversy. Nat Rev Neurol. 2016; 12:117–24. [PubMed: 26782335]
- 81. Jack CR Jr, Knopman DS, Weigand SD, Wiste HJ, Vemuri P, Lowe V, et al. An operational approach to National Institute on Aging-Alzheimer's Association criteria for preclinical Alzheimer disease. Annals of neurology. 2012; 71:765–75. [PubMed: 22488240]
- 82. Bramblett GT, Trojanowski JQ, Lee VM. Regions with abundant neurofibrillary pathology in human brain exhibit a selective reduction in levels of binding-competent tau and accumulation of abnormal tau-isoforms (A68 proteins). Lab Invest. 1992; 66:212–22. [PubMed: 1735956]
- 83. Montine TJ, Montine KS. Precision medicine: Clarity for the clinical and biological complexity of Alzheimer's and Parkinson's diseases. J Exp Med. 2015; 212:601–5. [PubMed: 25941321]
- 84. Perry D, Sperling R, Katz R, Berry D, Dilts D, Hanna D, et al. Building a roadmap for developing combination therapies for Alzheimer's disease. Expert Rev Neurother. 2015; 15:327–33. [PubMed: 25708309]
- 85. Leinenbach A, Pannee J, Dulffer T, Huber A, Bittner T, Andreasson U, et al. Mass spectrometry-based candidate reference measurement procedure for quantification of amyloid-beta in cerebrospinal fluid. Clin Chem. 2014; 60:987–94. [PubMed: 24842955]
- 86. Alsop DC, Detre JA, Golay X, Gunther M, Hendrikse J, Hernandez-Garcia L, et al. Recommended implementation of arterial spin-labeled perfusion MRI for clinical applications: A consensus of the ISMRM perfusion study group and the European consortium for ASL in dementia. Magn Reson Med. 2015; 73:102–16. [PubMed: 24715426]

Research in context

- 1. Systematic review: The authors reviewed literature pertaining to the achievements of ADNI and the development of future methodologies for the ADNI-3 study using traditional sources such as PubMed. Extensive literature supports the widespread impact of the ADNI study in multiple areas and particularly in the improvement of clinical trials such as the ongoing A4 and A5 studies
- 2. Interpretation: Results from analysis of ADNI-1 and ADNI-2 data combined with latest advances in technology have informed the structure and approach of the upcoming ADNI-3 study.
- 3. Future directions: innovative technologies such as longitudinal tau imaging, MRI connectivity analyses, and a mass spectroscopy approach to biomarker analysis, in addition to the standardization of amyloid PET scanning using the Centiloid method, and continued multimodal analysis will be employed to further understand AD pathophysiology and disease progression. The Brain Health Registry will help recruit subjects and monitor their cognition.

2.5		Cale Da		A D
0.7	Braak 0	Braak I/II	Braak III/IV	Braak V/VI
n (YA / OA / AD)	5/2/0	0/25/0	0/6/2	0/0/13
ApoE ε4 (0 / 1 / 2 allele)	2 / 0 / 0 (5 N/A)	18/7/0	4/3/0 (1 N/A)	6/5/2
% APOE ε4 +	28	28	38	54
MMSE	28.9 (1.5)	28.8 (1.3)	27.4 (3.2)	18.7 (5.1)
PiB DVR index	1.01 (.14) (5 N/A)	1.13 (.2)	1.35 (.36) (1 N/A)	1.80 (.17) (2 N/A
% PiB + (≥1.06)	0 (5 N/A)	44	71 (1 N/A)	100 (2 N/A)
Hippocampal volume (%ICV)	.54 (.05)	.44 (.05)	.42 (.09)	.44 (.05)

Figure 1. In-vivo Braak staging

Braak staging of healthy young adults (YA, n=5), healthy older adults (OA, n=15) and AD patients (n=15) based on AV-1451 Braak region of interest uptake, with participant characterization by in-vivo assigned Braak stage. ICV, intracranial volume. Values for Mini Mental State Exam (MMSE), Pittsburgh compound B distribution volume ratio (PIB DVR) index, and hippocampal volume are mean (SD). Reproduced with permission from [65].

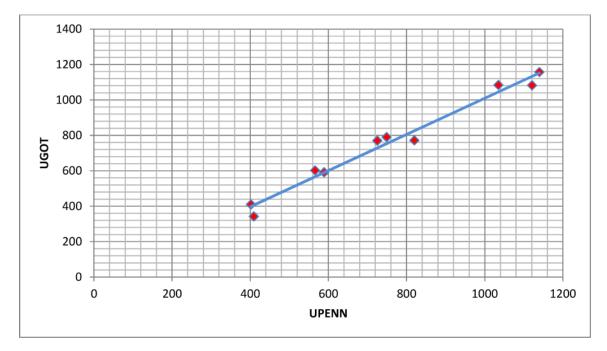


Figure 2. Comparison of $A\beta_{1_42}$ results (pg/mL) for 10 patient CSF samples UGOT, University of Gothenburg; UPENN, University of Pennsylvania. Each laboratory used their multiple reaction monitoring/tandem mass spectrometry method [72, 85], each approved by the Joint Committee for Traceability in Laboratory Medicine as reference methods.

Table 1

Enrollment Plan

	Rollover (enrolled YR01)	New (enrolled YR01)	New (enrolled YR02)	Total
Normal cognition (CN)	295	65	68	428
Amnestic MCI	274	65	86	425
AD	128	17	70	215
Total	697	147	224	1,068

Table 2

MRI sequences in ADNI-3

Modality	System	Measures	Notes
3D T1 volume	MPRAGE – Siemens/Phiips IRFSPGR – GE	Structural changes	Approximate 1mm ³ resolution
3D-FLAIR	All	Cerebral microbleeds	As in ADNI-2
Diffusion MRI	Advanced if available or basic	WM tract geometry	Similar to Human Connectome Project
Task-free fMRI	Advanced if available or basic	Functional connectivity	Similar to Human Connectome Project
Arterial Spin Labeling	Capable systems	Whole brain cerebral blood flow	Follows recommendations of ISMRM Perfusion Study Group and the European Consortium for ASL in Dementia [86].
Coronal high resolution T2	All	MTL subregion analysis	