Experimental evaluation of travoprost-induced changes in biomechanical behavior of ex-vivo rabbit corneas

Authors

XiaoBo Zheng ^{1,2}, Yuan Wang ¹, YiPing Zhao ¹, Si Cao ¹, Rong Zhu ¹, Wei Huang ¹, Ayong Yu ¹, JinHai Huang ¹, QinMei Wang ^{1,2}, JunJie Wang ^{1,2*}, FangJun Bao ^{1,2*}, Ahmed Elsheikh ^{3,4}

Affiliations

- ¹ Eye Hospital, WenZhou Medical University, Wenzhou, 325027, China
- ² The institution of ocular biomechanics, Wenzhou Medical University, Wenzhou, Zhejiang Province 325027, China
- ³ School of Engineering, University of Liverpool, Liverpool L69 3GH, UK
- ⁴ National Institute for Health Research (NIHR) Biomedical Research Centre at Moorfields Eye Hospital NHS Foundation Trust and UCL Institute of Ophthalmology, UK

Financial Support

This study was supported by the National Natural Science Foundation of China (81600712, 31771020), the Natural Science Foundation of Zhejiang Province (LY16H120005, LY18A020008), the Science and Technology Plan Project of Wenzhou Science and Technology Bureau (Y20170198), the Projects of medical and Health technology development program in ZheJiang Province (2016ZHB012, 2018RC057).

Abbreviated title

Experimental evaluation of travoprost-induced biomechanical change of rabbit cornea

Acknowledgements

The authors thank Shen LJ from WenZhou Medical University for technical assistance

with the study.

Contributors

Design of the study: Zheng XB, Wang JJ, Bao FJ, Elsheikh A

Acquisition of data, analysis and interpretation of data: Zheng XB, Wang Y, Zhao

YP, Cao S, Zhu R, Huang W, Yu AY, Huang JH, Wang QM, Wang JJ, Bao FJ, Elsheikh

Α

Revising the article critically for important intellectual content: Zheng XB, Wang

JJ, Bao FJ, Elsheikh A

Final approval of the version to be published: Zheng XB, Wang Y, Zhao YP, Cao S,

Zhu R, Huang W, Yu AY, Huang JH, Wang QM, Wang JJ, Bao FJ, Elsheikh A

PRECIS:

Prostaglandin F2α analogue travoprost causes a stiffness reduction effect on corneal

biomechanical properties under low applied stresses

Co-Corresponding author

Dr. JunJie Wang

No. 270 XueYuan West Road, WenZhou City, ZheJiang Prov, 325027, China

e-mail: w.wangjunjie@outlook.com

Tel: 86-577-88068862

Corresponding author

Dr. FangJun Bao

No. 270 XueYuan West Road, WenZhou City, ZheJiang Prov, 325027, China

e-mail: bfjmd@126.com

Tel: 86-577-88067937

Fax: 86-577-88824115

Abstract

Purpose: To assess the effects of prostaglandin F2α analogues travoprost on the

biomechanical behavior of ex-vivo rabbit cornea.

Methods: 18 Japanese white rabbits were included in the study. The left eye (treated

group, Tr) of each rabbit was preserved for 10 days in storage medium Eusol-C solution

with 1:10 travoprost diluent, while the contralateral eye (control group, Co) was

preserved in a similar but travoprost-free medium. Strips of corneal tissue were

dissected and tested under cyclic load conditions with up to 0.1 N uniaxial tension force.

The resulting load-elongation data were used to derive the stress-strain behavior and

the tangent modulus (Et) of the tissue. Differences in Et between the treated (Et-_{Tr}) and

control group (Et-Co) were assessed statistically to determine the biomechanical effects

of travoprost on the cornea.

Results: Central corneal thickness remained similar in the two groups before (p = 0.073)

and after storage (p= 0.303), although it became significantly thicker in both groups

after storage (P<0.01). Compared with the control group, the travoprost treated corneas

exhibited lower Et values but the differences reduced and became insignificant with

rises in stress to which the tissue was subjected (1 - Et- $_{Tr}$ /Et- $_{Co}$ = -11.7±41.8%, p< 0.05

at 10 kPa stress, -9.2±36.1%, p> 0.05 at 20 kPa, -7.3±35.4%, p>0.05 at 30 kPa).

Conclusions: Significant reductions in corneal stiffness, that are associated with the

use of travoprost, were observed experimentally under low applied stresses. This

stiffness-reduction effect should be considered in clinical management, especially in

primary open angle glaucoma treatment.

Keywords: prostaglandin; cornea; tangent modulus, tensile test

Introduction

Glaucoma, the second leading cause of blindness worldwide ¹, is a form of optic neuropathy associated with progressive degeneration of retinal ganglion cells and irreversible vision loss ². Raised intraocular pressure (IOP) remains the most important risk factor, and reduction of IOP can slow the progression of optic neuropathy ³ and is reportedly the most effective management method for glaucoma. Pharmacologic therapy is the initial treatment for glaucoma, and the most commonly prescribed classes of topical hypotensive agents are prostaglandin analogs (PGAs), especially for primary open angle glaucoma (POAG) ⁴. Prostaglandin F2α analogues (PGF 2α) upregulates the activity of matrix metalloproteinase (MMP) and downregulates the tissue inhibitor of metalloproteinase (TIMP) ⁵⁻⁷, which results in remodeling of the extracellular matrix, increasing the space between the bundles of smooth muscle cells, allowing better outflow and leading to lowering of IOP.

However, in addition to the effect of PGAs-induced IOP reduction, PGF 2α has been shown to decrease the collagen fibril density and corneal thickness ^{8,9}. Fibroblasts and extracellular matrix are the main structural components of the cornea, responsible to a large extent for determining corneal biomechanical properties. Collagen degradation caused by long-term topical prostaglandin therapy could influence corneal biomechanical behavior and induce reductions in corneal hysteresis (CH), corneal resistance factor (CRF) – both measured by the Ocular Response Analyzer, ORA) ¹⁰ – and the deformation amplitude (DA) provided by the Corvis ST (CVS) ¹¹. Changes in corneal biomechanical properties after long-term topical prostaglandin therapy possibly introduces inaccuracies in IOP measurement and in other applications that require knowledge of corneal biomechanics such as planning of surgical procedures, assessment of stiffness deterioration associated with keratoconus and optimization of corneal cross-linking treatment ¹².

Corneal biomechanical metrics provided by the ORA and CVS have been widely used

to assess the general biomechanical response of the cornea. Nevertheless, these metrics may be influenced by corneal shape and the intraocular pressure (IOP), and their links to standard mechanical properties, such as the tangent modulus of tissue (Et), have not been established $^{12-14}$. This study aims to address this shortfall through an experimental investigation of whether the usage of PGF 2α (in particular travoprost) influences the biomechanics of corneal tissue.

Materials and methods

2.1. Experimental animals

Eighteen Japanese white rabbits (2-3 kg) from the Animal Breeding Unit at Wenzhou Medical University were included in this study. All animals were treated in agreement with the ARVO Statement for Use of Animals in Ophthalmic and Vision Research, and every effort was made to minimize suffering. This study was approved by the Animal Care and Ethics Committee of the University's Eye Hospital.

2.2. Experimental design

After being euthanized by intravenous injection of high concentrations of pentobarbital sodium (Merok, Germany), bilateral eyes of each rabbit were immediately enucleated. The entire cornea, with the adjacent 3 mm wide scleral strip, was extracted from each ocular globe while all other ocular components were removed. The left eyes of the 18 rabbits, which form the treated group (Tr), were placed in storage medium of Eusol-C solution (Alchimia S.r.l, Ponte S. Nicolo', Italy) with 0.0004% travoprost (Travatan; Alcon Laboratories, Inc., Fort Worth, TX) diluent (1:10 dilution of stock solution). The corresponding 18 right eyes constituted the control group (Co), and were placed in the same medium but without travoprost. Based on other studies ^{15, 16}, the 1/10 dilution was selected to take into account the relatively short duration of storage adopted in the study compared to the long-term exposure in clinical usage. All corneas were incubated in standard incubator conditions (37°C, 5% CO₂) for 10 days as described in a previous study ¹⁷.

2.3. Biomechanical Tensile Testing

A 2-mm-wide corneoscleral strip centered on the cornea was excised from each specimen using two parallel surgical blades along the inferior-superior direction. The strips were connected to a pair of mechanical clamps, leaving a distance of 10mm in between. An electronic caliper (Exploit 033004, Exploit Tools Group, Yiwu, China) was used to measure the thickness (t) and width (w) of the strip in 5 equally-spaced locations along this length. Mechanical tests were conducted using a material testing machine (EZ-Test, Shimadzu, Kyoto, Japan) equipped with a 50 N capacity load cell at a room temperature of 22°C, Figure 1. The initial distance between the clamps was measured by a vernier caliper and recorded as Lo. The specimens were conditioned by four cycles of loading and unloading with 1mm/min elongation rate and 0.10N max load, and the behavior recorded in the fourth cycle was considered representative of specimens' stable behavior ¹⁸. The order of testing paired specimens, obtained from the same animal, was randomized and recorded. Strips were covered with gauze soaked with Phosphate buffered saline (PBS, Maixin, China) to keep them moist during the test procedure.

The load-displacement (F- Δ L) data obtained from the fourth cycle were used to calculate the stress under each load, F, as $\sigma = \frac{F}{w \cdot t}$, where t was the average corneal thickness and w the average specimen width. The related strain was obtained as $\varepsilon = \Delta$ L / Lo. The stress-strain results were fitting to an exponential function $\sigma = A \cdot (e^{B \cdot \varepsilon} - 1)$, where A and B were constants, and the tangent modulus (E_t) was calculated as $E_t = \frac{d\sigma}{d\varepsilon} = A \cdot B \cdot e^{B \cdot \varepsilon} = B \cdot (\sigma + A)$.

2.6 Statistical analysis

All analyses were performed using the PASW Statistics 20.0 (SPSS Inc., Chicago, USA). Comparisons of biomechanical and geometrical parameters in the two specimen groups were performed using the paired T-test. P values less than 0.05 were considered indicative of statistical significance.

Results

3.1. Corneal thickness

After 10 days of incubation in culture medium with 0.0004% travoprost diluent, the central corneal thickness (CCT) of the treated group increased from $369.4\pm22.5~\mu m$ to $658.9\pm184.4~\mu m$ (p<0.01), and in control group from $363.8\pm19.3~\mu m$ to $602.4\pm208.1~\mu m$ (p<0.01). There were no significant differences in corneal thickness between the two groups before storage (p=0.073), and the difference in corneal thickness between the groups remained statistically non-significant (p=0.303) after incubation.

3.2 Biomechanical behavior

There was a clear difference in the load-displacement behavior observed for the two specimen groups as shown in Figure 2. With material parameters A and B determined (Table 1), the stress-strain (σ - ϵ) relationships (Figure 3), and hence the tangent modulus (Et = $d\sigma/d\epsilon$) at any stress level can be obtained. For statistical evaluation purposes, the Et values were compared at 10, 20 and 30 kPa stresses, the first two of which were within the tissue's nonlinear stage, while the third was within the later linear part. At 10 kPa stress, Et was significantly lower in the treated group (Et- $_{Tr}$), compared to the control groups (Et- $_{Co}$), but this difference reduced in value and became insignificant under 20 and 30 kPa (Table 2).

Discussion

Topical medication is commonly used in the primary management of glaucoma. Among the several anti-glaucoma eye drops developed, PGF 2α are considered highly effective first-line agents because of their significant success in lowering IOP levels ¹⁹, that is in spite of reported side effects including eyelid skin darkening ²⁰, iris pigmentation ²¹, conjunctival hyperemia ²² and ocular irritation ²³. While these biological side effects have been considered previously ^{15, 24}, little attention has been given to the effect of PGF 2α on corneal biomechanics. PGF 2α have previously been found to accelerate

collagen degradation ²⁵, decrease fibronectin protein content ²⁶, stimulate collagen gel contraction ²⁷ and change collagen distribution in corneal stroma ²⁶. Since collagen fibrils are the main load carrying components of the cornea , these effects may lead to material stiffness reduction. This study, which attempted to address this point, showed that travoprost eye drops significantly reduced the mechanical stiffness (as measured by the tangent modulus, Et) of the ex-vivo rabbit cornea.

Earlier studies that relied on the ORA and CVS to provide indications of the biomechanical effects of PGAs produced inconsistent results. While some studies reported increases in corneal hysteresis parameter (CH – a measure of corneal viscoelasticity) with PGA treatment ²⁸⁻³³, others reported decreases ¹⁰. Also, there was no agreement on the effect of PGAs on the corneal resistance factor (CRF – a measure of corneal stiffness) with reported increases ³¹, decreases ^{10,33}, and no significant change ^{28,32}. However, after adjusting for IOP, CCT and other factors, which may influence corneal behavior, a significant reduction in the CVS's deformation amplitude (DA – a measure of corneal stiffness) was detected after PGA therapy ¹¹. Nevertheless, since the biomechanical metrics provided by the ORA and CVS cannot be linked directly to the traditional measures of tissue stiffness (primarily Et), and could be influenced by factors such as IOP and CCT ¹²⁻¹⁴, the present study relied instead of the classic tensile test in quantifying the effect of PGAs on corneal biomechanics.

The uniaxial tension test is a simple and well-accepted experimental technique for characterizing the mechanical behavior of tissue ³⁴. In spite of the limitations caused by the initially curved form of specimens and the termination of fibrils along the specimen sides ³⁵, the test method remains viable for comparative studies, such as the present research, where the focus is on the variation in tissue behavior due to different treatment regimes. All specimens exhibited clear nonlinear behavior, as indicated in a previous study ³⁶, with an initial low stiffness increasing gradually until a stage of constant stiffness was reached at stresses slightly below 30 kPa. In order to ensure the test results were repeatable, three loading-unloading cycles were carried out before using the

results of the forth cycle as representative of specimens' stable behavior 18 . The results showed significant decreases in Et in the treated group, by -11.7±41.8% (p< 0.05) at a stress of 10 kPa compared to the control group. However, these differences decreased and became insignificant with higher stress levels (-9.2±36.1%, p> 0.05 at 20kPa and -7.3±35.4%, p> 0.05 at 30 kPa).

The changes in tissue stiffness reported in this study may well influence the accuracy of IOP measurement – needed for glaucoma management ³⁷. Most tonometry techniques, contact or non-contact, depend on applying a mechanical force and correlating corneal resistance to deformation under this force to the value of IOP. While simple and easy to implement, this measurement concept makes the estimation of IOP dependent on corneal biomechanical properties ³⁸. With the application of PGAs leading to reductions in corneal stiffness, and hence underestimations of true IOP, the result may be an overestimation of the effect of PGAs in lowering IOP, which can have significant implications for glaucoma management.

There was a significant thickness increase observed during the storage period – due to tissue swelling – which correlated with the anaerobic state and increased lacate concentration 39 caused by the storage medium. The increase in corneal thickness, which affected both treated and control groups, may have masked the thickness reduction effect caused by PGAs usage as reported in earlier studies $^{8, 9}$, possibly as PGF 2α can induce excessive production of MMPs and inhibit the production of TIMP, both of which lead to an accelerated matrix degradation and decrease in CCT.

The present study relied on rabbit eyes due to their similarity to human eyes in biomechanical behavior ^{40, 41}, and the difficulty in obtaining human donor eyes in sufficient numbers for research. The tests were also done ex vivo, and while concerted efforts were made to preserve the tissue and test it within 2 hours post-mortem, there may have some degradation, which can affect the results obtained.

To the best of our knowledge, this is the first study to investigate the effect of PGAs hypotensive medications on corneal biomechanical property changes measured in standard biomechanical experiments. Corneal material stiffness reduced significantly with the use of PGF 2α (travoprost diluent, 0.0004%), causing concern over the accuracy of IOP measurement in patients undergoing chronic PGA therapy. This finding warrants caution when evaluating IOP measurements and the results of patient follow-up. Further investigation is required to quantify the effect of the stiffness reduction reported herein on the IOP measurements with commonly used tonometers, and hence the management of glaucoma.

References:

- 1. Quigley HA, Broman AT. The number of people with glaucoma worldwide in 2010 and 2020. Br J Ophthalmol 2006;90:262-7.
- 2. Weinreb RN, Aung T, Medeiros FA. The pathophysiology and treatment of glaucoma: a review. JAMA 2014;311:1901-11.
- 3. INVESTIGATORS. TA. The Advanced Glaucoma Intervention Study (AGIS): 7. The relationship between control of intraocular pressure and visual field deterioration. The AGIS Investigators. Am J Ophthalmol 2000;130:429-40.
- 4. Camras CB. Comparison of latanoprost and timolol in patients with ocular hypertension and glaucoma: a six-month masked, multicenter trial in the United States. The United States Latanoprost Study Group. Ophthalmology 1996;103:138-47.
- 5. Lindsey JD, Kashiwagi K, Boyle D, Kashiwagi F, Firestein GS, Weinreb RN. Prostaglandins increase proMMP-1 and proMMP-3 secretion by human ciliary smooth muscle cells. Curr Eye Res 1996;15:869-75.
- 6. Schachtschabel U, Lindsey JD, Weinreb RN. The mechanism of action of prostaglandins on uveoscleral outflow. Curr Opin Ophthalmol 2000;11:112-5.
- 7. Weinreb RN, Lindsey JD, Marchenko G, Marchenko N, Angert M, Strongin A. Prostaglandin FP agonists alter metalloproteinase gene expression in sclera. Invest Ophthalmol Vis Sci 2004;45:4368-77.
- 8. Schlote T, Tzamalis A, Kynigopoulos M. Central corneal thickness during treatment with travoprost 0.004% in glaucoma patients. J Ocul Pharmacol Ther 2009;25:459-62.
- 9. Zhong Y, Shen X, Yu J, Tan H, Cheng Y. The comparison of the effects of latanoprost, travoprost, and bimatoprost on central corneal thickness. Cornea 2011;30:861-4.
- 10. Meda R, Wang Q, Paoloni D, Harasymowycz P, Brunette I. The impact of chronic use of prostaglandin analogues on the biomechanical properties of the cornea in patients with primary openangle glaucoma. Br J Ophthalmol 2017;101:120-125.
- 11. Wu N, Chen Y, Yu X, Li M, Wen W, Sun X. Changes in Corneal Biomechanical Properties after Long-Term Topical Prostaglandin Therapy. PLoS One 2016;11:e0155527.
- 12. Bao F, Deng M, Wang Q, et al. Evaluation of the relationship of corneal biomechanical metrics with physical intraocular pressure and central corneal thickness in ex vivo rabbit eye globes. Exp Eye Res

2015;137:11-7.

- 13. Kotecha A, Elsheikh A, Roberts CR, Zhu H, Garway-Heath DF. Corneal thickness- and age-related biomechanical properties of the cornea measured with the ocular response analyzer. Invest Ophthalmol Vis Sci 2006;47:5337-47.
- 14. Chang PY, Chang SW, Wang JY. Assessment of corneal biomechanical properties and intraocular pressure with the Ocular Response Analyzer in childhood myopia. Br J Ophthalmol 2010;94:877-81.
- 15. Guenoun JM, Baudouin C, Rat P, Pauly A, Warnet JM, Brignole-Baudouin F. In vitro comparison of cytoprotective and antioxidative effects of latanoprost, travoprost, and bimatoprost on conjunctiva-derived epithelial cells. Invest Ophthalmol Vis Sci 2005;46:4594-9.
- 16. Guenoun JM, Baudouin C, Rat P, Pauly A, Warnet JM, Brignole-Baudouin F. In vitro study of inflammatory potential and toxicity profile of latanoprost, travoprost, and bimatoprost in conjunctiva-derived epithelial cells. Invest Ophthalmol Vis Sci 2005;46:2444-50.
- 17. Ni S, Yu J, Bao F, Li J, Elsheikh A, Wang Q. Effect of glucose on the stress-strain behavior of ex-vivo rabbit cornea. Exp Eye Res 2011;92:353-60.
- 18. Elsheikh A, Alhasso D. Mechanical anisotropy of porcine cornea and correlation with stromal microstructure. Exp Eye Res 2009;88:1084-91.
- 19. Al-Jazzaf AM, DeSantis L, Netland PA. Travoprost: a potent ocular hypotensive agent. Drugs Today (Barc) 2003;39:61-74.
- 20. Yang HK, Park KH, Kim TW, Kim DM. Deepening of eyelid superior sulcus during topical travoprost treatment. Jpn J Ophthalmol 2009;53:176-9.
- 21. Huang P, Zhong Z, Wu L, Liu W. Increased iridial pigmentation in Chinese eyes after use of travoprost 0.004%. J Glaucoma 2009;18:153-6.
- 22. Stewart WC, Kolker AE, Stewart JA, Leech J, Jackson AL. Conjunctival hyperemia in healthy subjects after short-term dosing with latanoprost, bimatoprost, and travoprost. Am J Ophthalmol 2003;135:314-20.
- 23. Day DG, Sharpe ED, Atkinson MJ, Stewart JA, Stewart WC. The clinical validity of the treatment satisfaction survey for intraocular pressure in ocular hypertensive and glaucoma patients. Eye (Lond) 2006;20:583-90.
- 24. Kahook MY, Ammar DA. In vitro toxicity of topical ocular prostaglandin analogs and preservatives on corneal epithelial cells. J Ocul Pharmacol Ther 2010;26:259-63.
- 25. Maruyama Y, Mori K, Ikeda Y, Ueno M, Kinoshita S. Effects of long-term topical prostaglandin therapy on central corneal thickness. J Ocul Pharmacol Ther 2014;30:440-4.
- 26. Wu KY, Wang HZ, Hong SJ. Effect of latanoprost on cultured porcine corneal stromal cells. Curr Eye Res 2005;30:871-9.
- 27. Liu Y, Yanai R, Lu Y, Hirano S, Sagara T, Nishida T. Effects of antiglaucoma drugs on collagen gel contraction mediated by human corneal fibroblasts. J Glaucoma 2006;15:255-9.
- 28. Tsikripis P, Papaconstantinou D, Koutsandrea C, Apostolopoulos M, Georgalas I. The effect of prostaglandin analogs on the biomechanical properties and central thickness of the cornea of patients with open-angle glaucoma: a 3-year study on 108 eyes. Drug design, development and therapy 2013;7:1149-56.
- 29. Agarwal DR, Ehrlich JR, Shimmyo M, Radcliffe NM. The relationship between corneal hysteresis and the magnitude of intraocular pressure reduction with topical prostaglandin therapy. Br J Ophthalmol 2012;96:254-7.
- 30. McCullough SJ, Little JA, Breslin KM, Saunders KJ. Comparison of refractive error measures by the

IRX3 aberrometer and autorefraction. Optom Vis Sci 2014;91:1183-90.

- 31. Pinero DP, Teus MA. Clinical outcomes of small-incision lenticule extraction and femtosecond laser-assisted wavefront-guided laser in situ keratomileusis. J Cataract Refract Surg 2016;42:1078-93.
- 32. Lazcano-Gomez G, Ancona-Lezama D, Gil-Carrasco F, Jimenez-Roman J. Effects of topical travoprost 0.004% on intraocular pressure and corneal biomechanical properties in an animal model. Digital journal of ophthalmology: DJO / sponsored by Massachusetts Eye and Ear Infirmary 2016;22:1-5.
- 33. Detry-Morel M, Jamart J, Pourjavan S. Evaluation of corneal biomechanical properties with the Reichert Ocular Response Analyzer. Eur J Ophthalmol 2011;21:138-48.
- 34. Williams M, Lewis W, Franco W. Loss of Tryptophan Fluorescence Correlates With Mechanical Stiffness Following Photo-Crosslinking Treatment of Rabbit Cornea. Invest Ophthalmol Vis Sci 2017;58:1110-1115.
- 35. Elsheikh A, Anderson K. Comparative study of corneal strip extensometry and inflation tests. J R Soc Interface 2005;2:177-85.
- 36. Liu X, Wang L, Ji J, et al. A mechanical model of the cornea considering the crimping morphology of collagen fibrils. Invest Ophthalmol Vis Sci 2014;55:2739-46.
- 37. Clayson K, Pan X, Pavlatos E, et al. Corneoscleral stiffening increases IOP spike magnitudes during rapid microvolumetric change in the eye. Exp Eye Res 2017;165:29-34.
- 38. Liu J, Roberts CJ. Influence of corneal biomechanical properties on intraocular pressure measurement: quantitative analysis. J Cataract Refract Surg 2005;31:146-55.
- 39. Redbrake C, Salla S, Frantz A, Reim M. [Energy metabolism of the human cornea in various culture systems]. Klin Monbl Augenheilkd 1997;210:213-8.
- 40. Sjoquist B, Basu S, Byding P, Bergh K, Stjernschantz J. The pharmacokinetics of a new antiglaucoma drug, latanoprost, in the rabbit. Drug Metab Dispos 1998;26:745-54.
- 41. Pellinen P, Huhtala A, Tolonen A, Lokkila J, Maenpaa J, Uusitalo H. The cytotoxic effects of preserved and preservative-free prostaglandin analogs on human corneal and conjunctival epithelium in vitro and the distribution of benzalkonium chloride homologs in ocular surface tissues in vivo. Curr Eye Res 2012;37:145-54.

- 1 Figure Captions:
- 2 Figure 1 Corneal specimen and experimental platform
- 3 Figure 2 Mean load-displacement behavior in treated and control groups
- 4 Figure 3 Mean stress-strain behavior of corneas in each specimen group error bars
- 5 represent standard deviation of strain values

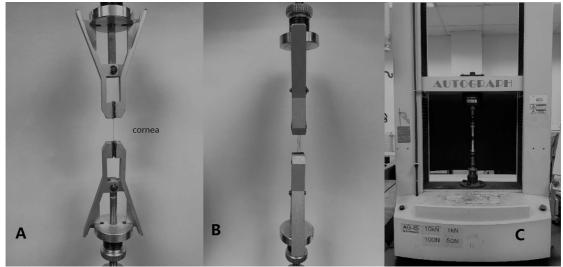


Figure1 Corneal specimen and experimental platform, A. front view of testing strip specimens after assembly, B. side view of testing strip specimens after assembly, C. material testing machine

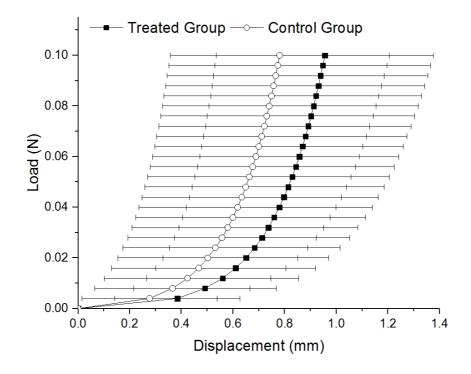


Figure 2 Mean load-displacement behavior in treated and control groups

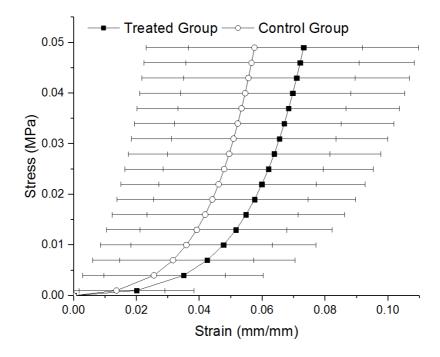


Figure 3 Mean stress-strain behavior of corneas in the treated and control groups –

error bars represent the standard deviation of strain values

Table Captions:

Table 1 Mean and standard deviation of constitutive parameters A and B in two specimen groups

Table 1 Mean and standard deviation of constitutive parameters A and B in the two specimen groups

Group	A	В	RMS, mm
Treated group	0.002 ± 0.003	66.481 ± 23.706	0.0017 ± 0.0014
Control group	0.004 ± 0.006	71.495 ± 26.946	0.0012 ± 0.0008

Table 2 Average and standard deviation values of tangent modulus (MPa) in treated and control groups at three stress levels

Stress (kPa)	Tangent Mod	Tangent Modulus, Et (MPa)		Et /Et 0/
	Tr	Co	– р	Et- _{Tr} /Et- _{Co} , %
10	0.78 ± 0.27	1.00 ± 0.45	0.025	88.3 ± 41.8
20	1.45 ± 0.49	1.71 ± 0.64	0.059	90.8 ± 36.1
30	2.11 ± 0.72	2.43 ± 0.88	0.119	92.7 ± 35.4

Tr = treated group, Co = control group; Et-Tr/Et-Co = ratio between tangent modulus in treated group (Et-Tr) and control group (Et-Co)