

# A role for the extended amygdala in the fear-enhancing effects of acute selective serotonin reuptake inhibitor treatment

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Selective serotonin reuptake inhibitors (SSRIs) are reported to exacerbate symptoms of anxiety when treatment is initiated. These clinical findings have been extended to animal models wherein SSRIs also potentiate anxiety and fear learning, which depend on the amygdala. Yet, little is known about the role of specific amygdalar circuits in these acute effects of SSRIs. Here, we first confirmed that a single injection of fluoxetine 1 h before auditory fear conditioning potentiated fear memory in rats. To probe the neural substrates underlying this enhancement, we analyzed the expression patterns of the immediate early gene, Arc (activity-regulated cytoskeleton-associated protein). Consistent with previous reports, fear conditioning induced Arc protein expression in the lateral and basal amygdala. However, this was not enhanced further by pre-treatment with fluoxetine. Instead, fluoxetine significantly enhanced expression of Arc in the central amygdala (CeA) and the bed nucleus of the stria terminalis (BNST). Next, we tested whether direct targeted infusions of fluoxetine into the CeA, or BNST, leads to the same fear-potentiating effect. Strikingly, direct infusion of fluoxetine into the BNST, but not the CeA, was sufficient to enhance fear memory. Moreover, this behavioral effect was also accompanied by robust Arc expression in the CeA, similar to the systemic injection. Our results identify a novel role for the BNST in the acute fear-enhancing effects of SSRIs. These findings highlight the need to look beyond the traditional focus on input nuclei of the amygdala and add to accumulating evidence implicating these microcircuits in gating fear and anxiety.

*Translational Psychiatry* (2013) 3, e209; doi:10.1038/tp.2012.137; published online 15 January 2013

## Introduction

Antidepressant drugs, such as selective serotonin reuptake inhibitors (SSRIs), are commonly administered for treating a range of anxiety disorders.<sup>1–4</sup> Although long-term treatment with SSRIs yields therapeutic benefits, they exacerbate symptoms of anxiety when treatment is initiated, potentially leading to an increased risk of suicidal ideation.<sup>5–10</sup> Although the acute anxiogenic effects of SSRIs have been demonstrated in numerous rodent models of anxiety-like behavior,<sup>11–15</sup> the cellular substrates of these acute effects are largely unknown.

Acute SSRI treatment, before auditory fear conditioning, leads to stronger fear memory,<sup>16</sup> which is consistent with its known anxiogenic effects. Auditory fear conditioning offers a significant advantage as a robust model of associative emotional learning, as the neural circuitry and molecular mechanisms have been elucidated in detail in the amygdala.<sup>17–20</sup> The lateral nucleus of the amygdala (LA) serves as the major input interface receiving sensory information from the thalamus and cortex.<sup>21,22</sup> Information processed within the LA and basal nucleus of the amygdala (BA) is then relayed to output structures. The central nucleus of the amygdala (CeA) and the bed nucleus of the stria terminalis (BNST), which together comprise the extended amygdala, serve as the

principal output structures of the amygdaloid complex that project to the brainstem and hypothalamus and mediate the expression of fear responses.<sup>23,24</sup> While the CeA mediates expression of conditioned fear responses to brief stimuli, the BNST mediates generalized and sustained fear responses to more diffuse stimulus contingencies.<sup>25,26</sup>

Current understanding of the cellular and circuit mechanisms of fear conditioning gives rise to several predictions about how acute treatment with SSRIs may enhance fear memory. Plasticity in the LA is crucial for the formation of conditioned fear memory.<sup>20</sup> Activity-regulated cytoskeleton-associated protein (Arc), a plasticity-related immediate early gene (IEG), is induced by auditory fear conditioning, in the LA and BA, and is essential for fear memory expression.<sup>27</sup> Thus, SSRIs may potentiate fear memory by enhancing Arc expression in the LA or BA. Alternatively, SSRIs may affect the output nuclei, which have also been implicated in fear memory formation.<sup>19,28,29</sup> In this study, we explore these possibilities by combining auditory fear conditioning, systemic treatment and targeted infusions of the SSRI fluoxetine into the amygdala and analysis of Arc expression patterns to identify components of the amygdalar fear circuitry that may underlie the acute behavioral effects of SSRIs.

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**Keywords:** anxiety; bed nucleus of the stria terminalis; central amygdala; fear conditioning; fluoxetine; immediate early gene

Received 23 August 2012; revised 27 October 2012; accepted 10 November 2012

## Materials and methods

**Subjects.** Male Sprague–Dawley rats (250–300 g) were housed in groups of two or three with *ad libitum* access to food and water. All animal experiments were conducted at the National Centre for Biological Sciences (NCBS) with the exception of experiments involving local drug infusions, which were carried out at the animal facility at Barnard College. Animals were maintained in a temperature-controlled room, with a 14/10-h day/night cycle at NCBS and 12/12-h day/night cycle at the Barnard College animal facility. All procedures were approved by the Institutional Animal Ethics Committee for NCBS or Columbia University's Animal Care and Use Committee.

**Drug administration.** Fluoxetine hydrochloride was obtained from Hikal Ltd (Bangalore, India) and Sigma-Aldrich (St Louis, MO, USA).

**Systemic injections.** For systemic drug administration, fluoxetine was dissolved in 0.9% sterile saline and injected intraperitoneally (i.p.) at a dose of  $10 \text{ mg kg}^{-1}$ .<sup>28–30</sup>

**Local drug infusions.** Cannulae were surgically implanted to locally infuse fluoxetine or saline into the CeA or BNST. Rats were anesthetized and maintained in deep anesthesia with a mixture of isoflurane and oxygen and placed in a stereotaxic apparatus. Body temperature was maintained with a heated gel pad. Betadine was applied to the scalp and a local anesthetic (bupivacaine, subcutaneous) was injected under the scalp. The skull was exposed, and small holes were drilled. Stainless steel guide cannulae (22 gauge; Plastics One, Roanoke, VA, USA) fitted with infusion cannulae (28 gauge) that extended (2 mm) beyond the base of the guide were positioned bilaterally above the CeA (–2.5 mm anteroposterior (AP),  $\pm$ 4.3 mm mediolateral (ML), –8.0 mm dorsoventral (DV)) or BNST (–0.6 mm AP,  $\pm$ 1.5 ML, –6.5 mm DV).<sup>31</sup> The guide cannulae were secured to the skull using surgical screws and acrylic dental cement. Infusion cannulae were replaced with dummy cannulae that did not extend beyond the guide cannulae to prevent clogging. After surgery, rats received an analgesic (carprofen,  $5 \text{ mg kg}^{-1}$ , i.p.) and 5 ml of lactated ringer (subcutaneous) and recovered for 1 week before drug infusion and behavioral testing.

Fluoxetine was infused into the CeA or BNST at a dose of 6 or  $9 \mu\text{g}$  per side (in  $0.5 \mu\text{l}$  saline). This choice of dose was based on the behavioral effects reported by earlier studies that have infused fluoxetine directly into the brain at these doses.<sup>32,33</sup> The drug solution was infused at a rate of  $2 \mu\text{l min}^{-1}$  through infusion cannulae attached to a  $1\text{-}\mu\text{l}$  Hamilton syringe via polyethylene tubing. The cannulae were left in place for an additional 2 min after the infusion to allow for diffusion of the solution away from the cannula tip, after which the dummy cannulae were replaced and the rat was returned to its home cage.

To verify the placement of the injection cannula tips after behavioral testing, rats were anesthetized with Euthasol (containing pentobarbital;  $100 \text{ mg kg}^{-1}$ , i.p.) and transcardially perfused with 0.9% saline followed by ice-cold

4% paraformaldehyde in phosphate-buffered saline (PBS). Brains were removed and stored in 4% paraformaldehyde for 24 h and then cryoprotected in 20% followed by 30% sucrose (in  $0.1 \text{ M}$  phosphate buffer, pH 7.4). Subsequently, brains were blocked and cut in  $60 \mu\text{m}$  sections through the CeA or BNST using a cryostat. After standard histological Nissl or hematoxylin staining, sections were examined on a light microscope to localize the cannula tip.

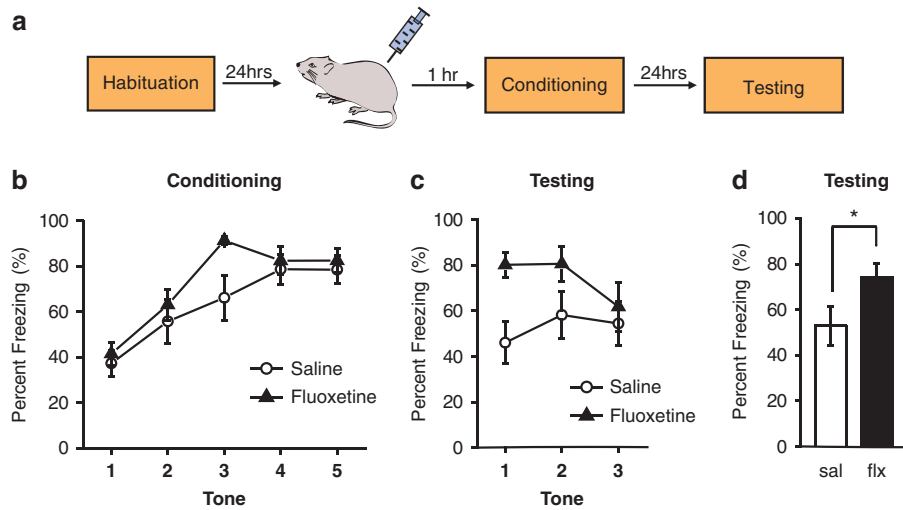
## Behavioral procedures

**Apparatus and stimuli.** Rats were fear conditioned in chambers made of aluminium and Plexiglass walls with stainless steel rod flooring attached to a shock generator (Coulbourn Instruments, Allentown, PA, USA). The chamber was illuminated by an overhead yellow light, all of which was located within a sound attenuating box (Coulbourn Instruments). The testing context was made distinct by changing the walls, floors, lighting and introducing a peppermint odor to the chamber. A video camera was used to record behavioral conditioning and testing sessions for subsequent offline analysis. Stimulus presentation was controlled by a computer with Graphic State 2 software (Coulbourn Instruments). The conditioning and testing chambers were thoroughly cleaned with alcohol between sessions.

**Fear conditioning.** For experiments involving systemic drug administration, auditory fear conditioning consisted of a 3-day procedure. On day 1, all rats were first habituated to the conditioning chamber followed by the testing chamber for 10 min. On day 2, rats were injected with fluoxetine or saline, and 60 min later they were fear conditioned in the conditioning chamber. Fear conditioning involved five pairings of a conditioned stimulus (CS; 5 kHz tone, 75 db, 20 s) that co-terminated with an unconditioned stimulus (0.5 mA footshock, 0.5 s). The inter-trial interval varied randomly between 60 and 120 s. On day 3, rats were placed in the testing chamber where they received three CS presentations with an inter-trial interval that varied randomly between 60 and 120 s. Behavior during the conditioning and testing sessions was videotaped for subsequent manual off-line analysis of freezing behavior. Freezing, defined as cessation of all movement except for respiration, was used as a measure of conditioned fear. Freezing is reported as a percentage of total tone presentation time.

For the local drug infusion experiments, rats were allowed to recover for 1 week after the surgery. On day 1, rats were habituated to the conditioning chamber for 20 min. On day 2, rats were infused with fluoxetine or saline 15 min before they were fear conditioned with two pairings of the CS (5 kHz tone, 75 db, 30 s) and unconditioned stimulus (0.5 mA footshock, 0.5 s). On day 3, they were tested in the testing chamber with 10 CS presentations.

**Immunohistochemistry.** Two hours after the conditioning session, rats were killed and their brains were analyzed for behaviorally induced Arc protein expression. The 2-h time point after fear conditioning was chosen to examine Arc expression based on an earlier study that reported an induction of Arc protein expression in the LA and BA 2 h after fear conditioning.<sup>27</sup> Rats were deeply anesthetized with



**Figure 1** Acute systemic administration of fluoxetine enhances conditioned fear memory. (a) Schematic representation of the experimental protocol. Rats were habituated to the conditioning and testing context and injected with saline or fluoxetine ( $10 \text{ mg kg}^{-1}$ , i.p.) on the following day. One hour after injection, they were conditioned with five conditioned stimulus–unconditioned stimulus (CS–US) pairings and tested for conditioned fear to three CS presentations on the following day. (b) Percentage of time spent freezing in response to tone presentations on the conditioning day. (c) Percentage of time spent freezing in response to tone presentations on the testing day. (d) Percentage of time spent freezing to all three tones on the testing day. Saline (sal):  $N = 13$ , fluoxetine (fix):  $N = 14$ . Error bars indicate mean  $\pm$  s.e.m. and  $*P < 0.05$ .

halothane followed by ketamine (0.05 ml per 100 g, i.p.) and xylazine (0.2 ml per 100 g, i.p.) and transcardially perfused with 0.1 M PBS, followed by ice-cold 4% paraformaldehyde–PBS. Brains were removed and postfixed in 4% paraformaldehyde–PBS for 24 h and then cryoprotected in 20% followed by 30% sucrose (in 0.1 M phosphate buffer, pH 7.4). A cryostat was used to cut 40- $\mu\text{m}$  thick coronal brain sections containing the amygdala. Every sixth section was then processed for Arc immunoreactivity. After blocking in PBS containing 1% bovine serum albumin (BSA Fraction V, Sigma-Aldrich)–0.1% Triton X-100, slices were incubated overnight at room temperature in anti-Arc primary antibody (mouse monoclonal, 1:1000 dilution; Santa Cruz Biotechnology, Santa Cruz, CA, USA) in PBS–1% BSA–0.1% Triton X-100. Sections were then incubated with a biotinylated secondary antibody (anti-mouse IgG, 1:200 dilution, Vectastain Elite ABC kit, Vector Laboratories, Burlingame, CA, USA) for 1 h at room temperature. Arc-expressing cells were then visualized using VectaStain Elite ABC kit (Vector Laboratories) followed by DAB staining (DAB peroxidase substrate, Vector Laboratories). Sections were then mounted onto Fisherbrand electrostatic slides. Sections from comparable anterior–posterior levels were selected for scoring ( $-2.04$  to  $-2.92$  mm posterior to Bregma for the CeA,  $0.36$  to  $-0.96$  mm posterior to Bregma for the BNST and  $-2.92$  and  $-3.96$  mm posterior to Bregma for the dentate gyrus (DG)). Cell counts were taken from at least three sections (six hemispheres) per rat. Cells were counted manually using the NeuroLucida image analysis system (MicroBrightField, Wiliston, VT, USA) attached to an Olympus BX61 microscope (Olympus, Tokyo, Japan;  $\times 40$ , 0.75 numerical aperture).

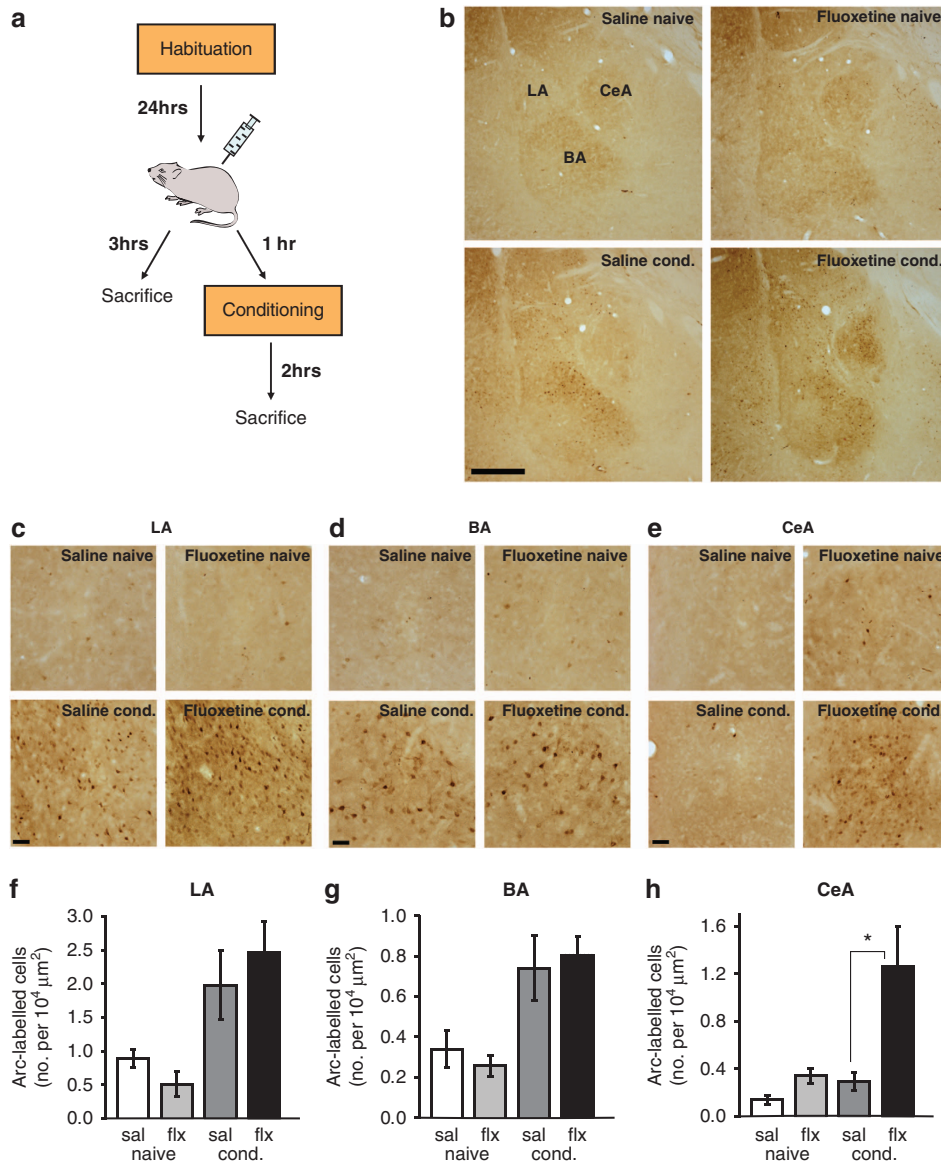
**Statistics.** Statistical analysis was performed using OriginPro-8 (OriginLab, Northampton, MA, USA) or SPSS-9 (SPSS Inc., Chicago, IL, USA). Fear conditioning data

were analyzed using a repeated measures analysis of variance (ANOVA) for tone-wise freezing scores (within subject factor—tone presentation, between subject factor—drug treatment). Student’s *t*-test was used to compare freezing scores averaged across the first three tones. Arc protein expression was analyzed separately in the LA, BA, CeA, BNST and DG using a two-way ANOVA (between subject factors—fear conditioning and drug treatment) or Student’s *t*-test for independent samples, where applicable. Significance was accepted for  $P < 0.05$ .

## Results

### Acute systemic administration of fluoxetine enhances conditioned fear memory.

We first tested the effects of the SSRI fluoxetine on Pavlovian auditory fear conditioning, an amygdala-dependent form of fear learning in which an animal rapidly learns to associate a previously innocuous tone with an aversive footshock. Re-exposure to the tone alone elicits a conditioned response, such as cessation of locomotor activity or ‘freezing’, which we used as a behavioral measure for the learned association. Rats were administered a single injection of fluoxetine ( $10 \text{ mg kg}^{-1}$ , i.p.) or saline 1 h before fear conditioning (Figure 1a). We found that fluoxetine did not significantly change freezing during the conditioning session when all tones were included in the analysis (Figure 1b; repeated measures ANOVA:  $F_{(1,25)} = 2.73$ ,  $P = 0.11$ ). However, we did find that fluoxetine-treated rats exhibited higher freezing than saline-treated rats to the third tone ( $t_{(12.96)} = 2.46$ ,  $P < 0.05$ ), consistent with previous studies showing an enhancement in learning following acute SSRI administration.<sup>16</sup> Thus, there is a possibility that in our experiment, the fluoxetine group learned fear faster but then approached maximal levels of freezing faster. Interestingly, presentation of the tone alone the next day during the testing session elicited significantly higher freezing in fluoxetine-

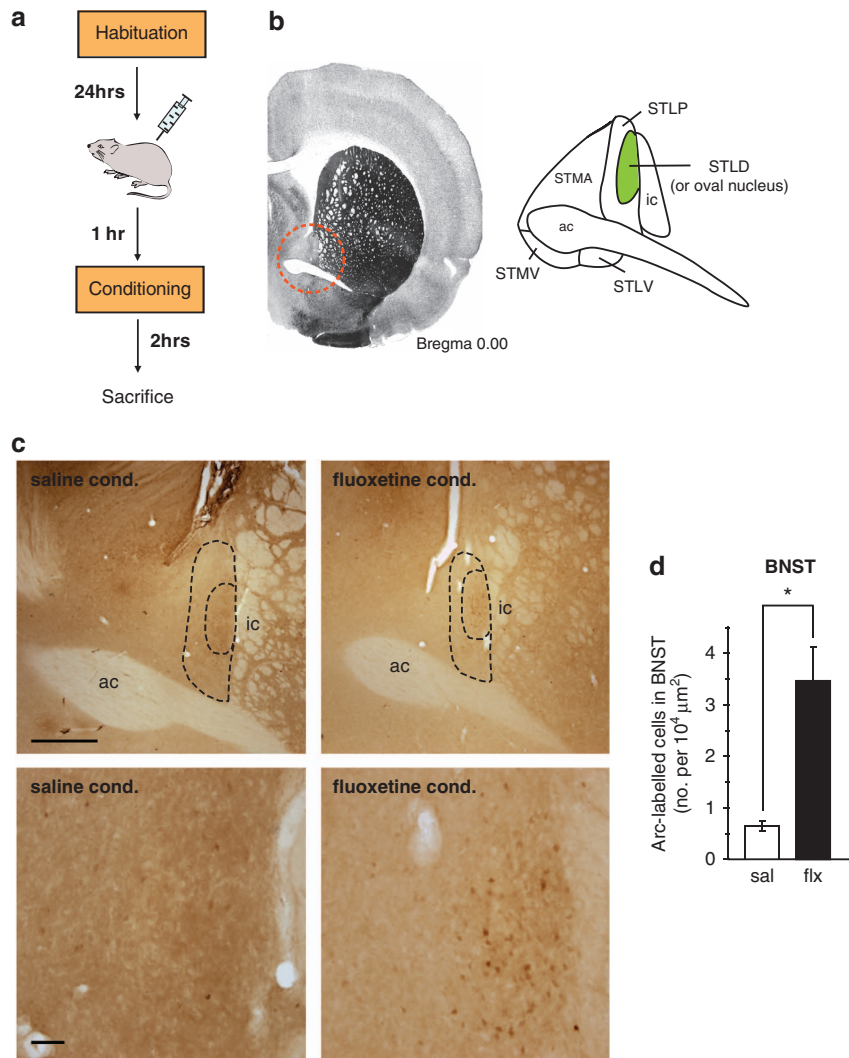


**Figure 2** Acute systemic fluoxetine treatment before fear conditioning leads to an increase in Arc (activity-regulated cytoskeleton-associated protein) expression in the central nucleus (CeA), but not lateral (LA) or basal nucleus (BA) of the amygdala. **(a)** Schematic representation of the experimental protocol. Rats were habituated to the conditioning context and injected with saline or fluoxetine ( $10 \text{ mg kg}^{-1}$ , i.p.) on the following day. Rats were either conditioned an hour after the injection and killed 2 h later (saline- and fluoxetine-conditioned groups) or were directly killed 3 h after the injection (saline and fluoxetine naive groups). The brain tissue was then processed for immunohistochemistry. **(b)** Low magnification images of brain sections from the four treatment groups to show Arc protein expression in the amygdala; bar =  $500 \mu\text{m}$ . **(c–e)** Representative images of Arc protein expression in the LA, BA and CeA, respectively; bar =  $40 \mu\text{m}$ . **(f–h)** Quantification of Arc-expressing cells across the four treatment groups in the LA, BA and CeA, respectively. Saline naive (sal naive):  $N = 4$ , fluoxetine naive (flx naive):  $N = 4$ , saline conditioned (sal cond.):  $N = 6$ , fluoxetine conditioned (flx cond.):  $N = 6$ . Error bars indicate mean  $\pm$  s.e.m. and  $*P < 0.05$ .

treated rats than saline-treated controls (Figure 1c; repeated measures ANOVA:  $F_{(1,25)} = 4.33$ ,  $P < 0.05$  and Figure 1d;  $t_{(25)} = 2.1$ ,  $P < 0.05$ ). The baseline levels of freezing measured during the 30-s period immediately before tone onset did not differ between the groups ( $t_{(25)} = 0.08$ ,  $P = 0.94$ ), indicating that the enhancement in fear elicited by fluoxetine was specific to the learned association. These results are consistent with a previous study showing that a single injection of the SSRI citalopram before fear conditioning also enhances the acquisition of auditory fear memory.<sup>16</sup>

**Acute systemic fluoxetine administration enhances Arc protein expression in the CeA but not LA or BA.** We next investigated which specific components of the neural circuitry underlying fear learning might serve as substrates for the fear-enhancing effect of fluoxetine. An effective strategy for such an analysis comes from numerous studies in which the IEG Arc has been used to identify cells that are activated and undergo plasticity in response to behaviorally relevant stimuli.<sup>34–38</sup> Using this approach, a recent study has shown that auditory fear conditioning induces Arc expression in the

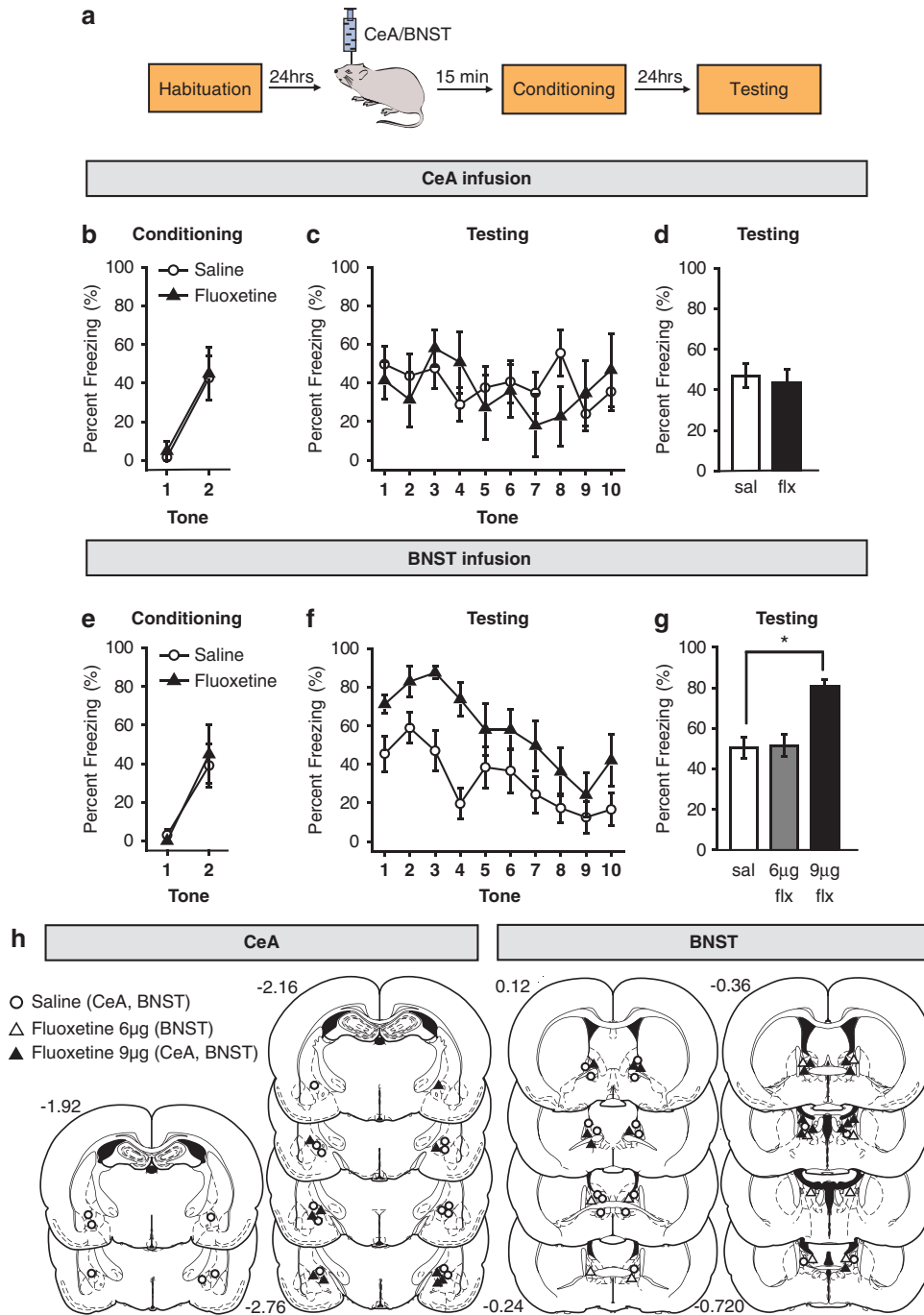




**Figure 3** Acute systemic fluoxetine treatment induces Arc (activity-regulated cytoskeleton-associated protein) expression in the bed nucleus of the stria terminalis (BNST). (a) Schematic representation of the experimental protocol. Rats were habituated to the conditioning context and injected with saline or fluoxetine ( $10 \text{ mg kg}^{-1}$ , i.p.) on the following day. Rats were conditioned an hour after the injection and killed 2 h later. The brain tissue was then processed for immunohistochemistry. (b) Left, sample coronal brain section containing the BNST, marked in a red dotted circle (Paxinos and Watson<sup>31</sup>); right, schematic of the principal BNST sub-regions (STLD, BNST lateral-dorsal or oval nucleus; STLP, BNST lateral-posterior; STL, BNST lateral-ventral; STMV, BNST medial-ventral; STMA, BNST medial-anterior; ac, anterior commissure, ic, internal capsule). (c) Representative images of Arc protein expression in the BNST. Top panel: low magnification images of the BNST from the saline (left)- and fluoxetine (right)-injected rats; bar =  $500 \mu\text{m}$ ; bottom panel: higher magnification images of the BNST from the top panel; bar =  $50 \mu\text{m}$ . (d) Quantification of Arc-expressing cells in the BNST. Saline conditioned (sal):  $N = 3$ , fluoxetine conditioned (flx):  $N = 3$ . Error bars indicate mean  $\pm$  s.e.m. and  $*P < 0.05$ .

LA and BA and that this upregulation of Arc is essential for the expression of fear memory.<sup>27</sup> We therefore hypothesized that acute fluoxetine administration may enhance fear memory by promoting plasticity in the LA and BA, thereby leading to an increase in Arc protein expression in these regions. To test this hypothesis, we administered a single injection of fluoxetine ( $10 \text{ mg kg}^{-1}$ , i.p.) to rats 1 h before fear conditioning. Two hours after conditioning, the rats were killed and behaviorally induced Arc protein expression was quantified in the LA, BA and CeA (Figure 2a). To dissociate the effects of fluoxetine on baseline amygdala activity from those specifically triggered by fear conditioning, we injected a separate group of naive rats with the same dose of fluoxetine, returned them to their home cage and killed them

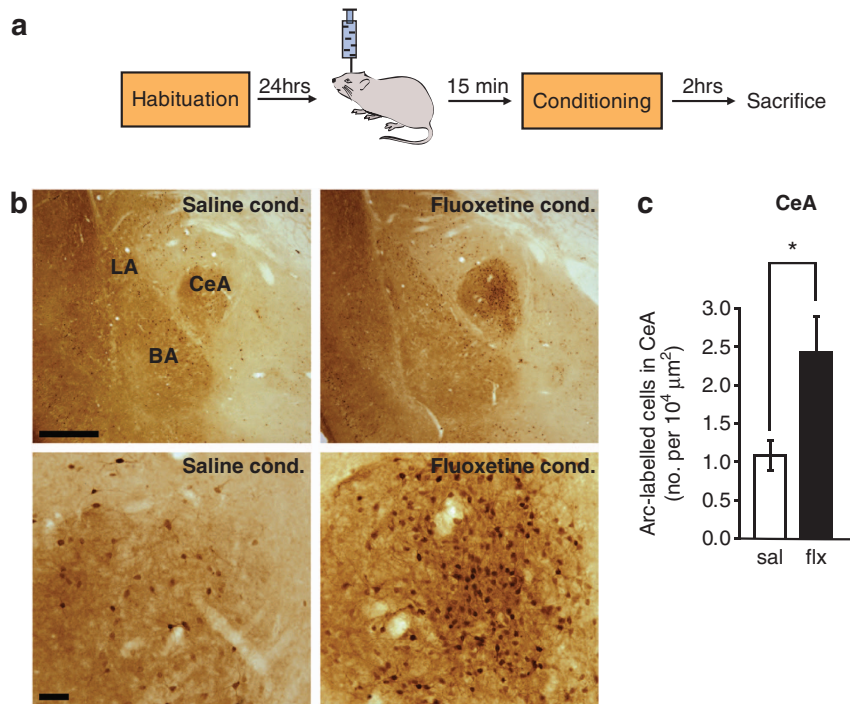
3 h later (Figure 2a). Statistical analysis using a two-way ANOVA showed a significant effect of fear conditioning in the LA and BA (LA:  $F_{(1,16)} = 11.81$ ,  $P < 0.01$ ; BA:  $F_{(1,16)} = 15.11$ ,  $P < 0.01$ ), indicating that there was more Arc protein expression in both the regions of fear conditioned animals than naive controls (Figures 2b–d, f and g). This is in agreement with earlier work showing that Arc expression is induced by auditory fear conditioning in these sub-regions of the amygdala.<sup>27</sup> Strikingly, Arc expression in the LA and BA was not affected by pre-treatment with fluoxetine (LA:  $F_{(1,16)} = 0.10$ ,  $P = 0.76$ ; BA:  $F_{(1,16)} = 0.002$ ,  $P = 0.96$ ; Figures 2b–d, f and g), and there was no interaction between fear conditioning and fluoxetine treatment (LA:  $F_{(1,16)} = 0.95$ ,  $P = 0.34$ ; BA:  $F_{(1,16)} = 0.37$ ,  $P = 0.55$ ). By contrast, a two-way



**Figure 4** Direct local infusion of fluoxetine into the bed nucleus of the stria terminalis (BNST), but not to the central nucleus of the amygdala (CeA), before fear conditioning enhances conditioned fear memory. (a) Schematic representation of the experimental protocol. Rats were habituated to the conditioning context and infused with saline or fluoxetine (9 µg per side unless mentioned otherwise) in the CeA or BNST on the following day. Fifteen minutes after the drug infusion, they were conditioned with two conditioned stimulus–unconditioned stimulus (CS-US) pairings. Rats were then tested for conditioned fear in response to 10 CS presentations on the next day. (b, e) Percentage of time spent freezing in response to tone presentations on the conditioning day for the intra-CeA and intra-BNST infusion animals, respectively. (c, f) Percentage of time spent freezing in response to tone presentations on the testing day for the intra-CeA and intra-BNST infusion animals, respectively. (d, g) Percentage of time spent freezing in response to the first three tones on the testing day for the intra-CeA and intra-BNST infusion animals, respectively. (h) Cannula tip placements from rats infused used with saline, fluoxetine-6 µg per side and fluoxetine-9 µg per side in the CeA (first and second column; Bregma -1.92 to -2.76 mm) and BNST (third and fourth column; Bregma +0.12 to -0.72 mm) before fear conditioning. CeA saline infusion (sal): *N* = 11, CeA fluoxetine infusion - 9 µg per side (flx): *N* = 5, BNST saline infusion (sal): *N* = 9, BNST fluoxetine infusion - 6 µg per side (6 µg flx): *N* = 6, BNST fluoxetine infusion - 9 µg per side (9 µg flx): *N* = 8. Error bars indicate mean ± s.e.m. and \**P* < 0.05.

ANOVA revealed a significant main effect of fluoxetine treatment on Arc protein expression in the CeA ( $F_{(1,16)}=9.26$ ,  $P<0.01$ ; Figures 2b, e and h). Pairwise

comparisons reveal that while systemic fluoxetine treatment had no significant effect in naive animals, rats treated with fluoxetine before fear conditioning showed significantly



**Figure 5** Direct local infusion of fluoxetine into the bed nucleus of the stria terminalis (BNST) before fear conditioning induces Arc (activity-regulated cytoskeleton-associated protein) expression in the central nucleus of the amygdala (CeA). (a) Schematic representation of the experimental protocol. Rats were habituated to the conditioning context and infused with saline or fluoxetine (9 μg per side) in the CeA or BNST on the following day. Fifteen minutes after the drug infusion, they were conditioned with two conditioned stimulus–unconditioned stimulus (CS-US) pairings. Two hours after conditioning, they were killed, and the brain tissue was processed for immunohistochemistry. (b) Representative images of Arc protein expression in the CeA. Top panel: low magnification images of the amygdala from the saline-infused (left) and fluoxetine-infused (right) rats; bar = 500 μm; bottom panel: high magnification images of the CeA from the top panel; bar = 50 μm. (c) Quantification of Arc-expressing cells in the CeA. Saline infusion (sal):  $N = 10$ , fluoxetine infusion (flx) – 9 μg per side:  $N = 9$ . Error bars indicate mean  $\pm$  s.e.m. and  $*P < 0.05$ . BA, basal nucleus of the amygdala; LA, lateral nucleus of the amygdala.

enhanced Arc protein expression in the CeA compared with saline-treated rats (Tukey's HSD (honestly significant difference),  $P < 0.05$ ; Figure 2h). Fear conditioning also had a significant effect on Arc expression in the CeA ( $F_{(1,16)} = 5.89$ ,  $P < 0.05$ ; Figure 2h). However, the interaction between fear conditioning and fluoxetine treatment was not significant ( $F_{(1,16)} = 2.98$ ,  $P = 0.10$ ). These results suggest that the facilitation of fear memory by acute systemic treatment with fluoxetine is accompanied by activation of the CeA and raise the possibility that this activation may underlie the behavioral effect.

**Acute systemic fluoxetine administration induces Arc protein expression in the BNST.** The CeA and the BNST serve as the principal output structures of the extended amygdaloid complex that project to the brainstem and hypothalamus and mediate the expression of fear responses.<sup>23,24,39</sup> In addition to being highly interconnected, the BNST and CeA have been shown to have important roles in modulating fear and anxiety.<sup>40–42</sup> Given this anatomical and functional relationship and our above finding that acute systemic fluoxetine enhanced Arc expression in the CeA, we examined the pattern of Arc protein in the BNST 2 h after rats pre-treated with acute systemic fluoxetine were fear conditioned (Figure 3a). Consistent with our hypothesis, we found that a single injection of fluoxetine also triggered Arc

expression in the dorsolateral region of the BNST (Figures 3c and d;  $t_{(2,1)} = 4.28$ ,  $P < 0.05$ ).

Finally, we examined whether Arc expression is enhanced by fluoxetine in other brain regions involved in fear learning. Previous work has shown that an acute SSRI injection decreases contextual fear conditioning,<sup>43,44</sup> a form of fear learning that is hippocampal-dependent.<sup>45,46</sup> Strikingly, in contrast to the CeA and the BNST, we found that fluoxetine treatment had the opposite effect on the DG of the hippocampus, a brain region that has been previously implicated in antidepressant action.<sup>47</sup> Acute fluoxetine treatment significantly decreased Arc protein expression in the DG in both the naive and fear-conditioned animals (Supplementary Figure S1;  $F_{(1,16)} = 19.41$ ,  $P < 0.001$ ), while fear conditioning itself had no effect on Arc expression (Supplementary Figure S1;  $F_{(1,16)} = 0.07$ ,  $P = 0.79$ ).

**Direct local infusion of fluoxetine into the CeA does not enhance conditioned fear memory.** Our results demonstrate that a systemic injection of fluoxetine enhances Arc protein expression in two output nuclei of the extended amygdala: the BNST and the CeA. To determine whether fluoxetine is acting directly on neurons in these two structures to enhance fear conditioning, we tested the effects of local infusions of fluoxetine targeted at either the CeA or the BNST.

We first tested the possibility that activation of CeA by fluoxetine is sufficient to modify fear behavior. If this is true, then local infusion of fluoxetine into the CeA before conditioning should also enhance auditory fear memory, as was observed with systemic treatment. We bilaterally infused fluoxetine (9 µg per side) directly into the CeA (−2.5 mm AP, +/−4.3 mm ML, −8.0 mm DV) 15 min before the conditioning session and measured CS-elicited freezing behavior 24 h later (Figure 4a). Intra-CeA fluoxetine infusions did not affect freezing during the conditioning session (Figure 4b; repeated measures ANOVA:  $F_{(1,14)} = 0.59$ ,  $P = 0.46$ ). Surprisingly, we found that intra-CeA fluoxetine infusions before conditioning also did not affect freezing measured during the testing session (Figure 4c; repeated measures ANOVA:  $F_{(1,14)} = 0.10$ ,  $P = 0.76$  and Figure 4d;  $t_{(14)} = 0.2528$ ,  $P = 0.80$ ). This suggests that targeted infusion of fluoxetine into the CeA alone is not sufficient to produce the same behavioral effects as acute systemic treatment.

**Direct local infusion of fluoxetine into the bed nucleus of the stria terminalis (BNST), is sufficient to enhance conditioned fear memory.** Given that Arc expression was also enhanced in the BNST following systemic fluoxetine administration coupled with fear conditioning, we next examined whether local infusions of fluoxetine into the BNST would enhance fear conditioning. We bilaterally infused rats with fluoxetine (6 or 9 µg per side) in the BNST 15 min before fear conditioning and measured the freezing response to the CS 24 h later (Figure 4a). These infusions targeted the dorsolateral and dorsomedial nuclei of the BNST (−0.6 mm AP, +/−1.5 mm ML, −6.5 mm DV), and infusion cannulae tracks in all the experimental animals were limited to 0.12 to −0.72 mm Bregma (Figure 4h). These coordinates were chosen based on earlier studies<sup>48</sup> that characterized the electrophysiological properties of BNST neurons in the area dorsal to the anterior commissure, between 0.0 and −0.6 mm Bregma. Intra-BNST fluoxetine infusions did not affect freezing during the conditioning session (Figure 4e; repeated measures ANOVA:  $F_{(1,15)} = 0.01$ ,  $P = 0.91$ ). Interestingly, we found that the intra-BNST fluoxetine enhanced fear memory recall during the testing session 24 h later. Moreover, this fear-enhancing effect was dose-dependent (Figure 4g; one-way ANOVA:  $F_{(2,20)} = 7.80$ ,  $P < 0.01$ ). Although the lower dose of 6 µg per side did not affect memory recall (Figure 4g; Tukey's HSD:  $P = 0.99$ ), a higher dose of 9 µg per side was sufficient to elicit a significant enhancement in fear memory during the testing session (Figure 4f; repeated measures ANOVA:  $F_{(1,15)} = 8.52$ ,  $P < 0.05$  and Figure 4g; Tukey's HSD:  $P < 0.01$ ). Thus, a single infusion of fluoxetine directly into the BNST produced the same behavioral effect on fear memory recall as a single systemic injection of fluoxetine.

**Direct local infusion of fluoxetine into the BNST before fear conditioning enhances Arc protein expression in the CeA.** Together, our results suggest that the BNST may be a primary site of action for fluoxetine and may mediate the behavioral enhancement of fear memory seen after acute systemic fluoxetine treatment. However, as shown earlier, systemic fluoxetine treatment also led to enhanced Arc

expression in the CeA. Does direct infusion of fluoxetine into the BNST cause the same effect in the CeA? To address this question, behaviorally induced Arc protein expression was evaluated in the CeA of rats infused with fluoxetine (9 µg per side) in the BNST 15 min before fear conditioning and killed 2 h later (Figure 5a). We found that intra-BNST fluoxetine infusion did enhance Arc protein expression in the CeA (Figures 5b and c;  $t_{(10.7)} = 2.6$ ,  $P < 0.5$ ). Thus, similar to systemic acute administration of fluoxetine, the facilitation of fear memory recall caused by intra-BNST fluoxetine infusions is also accompanied by the induction of Arc protein expression in the CeA.

## Discussion

Consistent with earlier reports,<sup>16</sup> we found that a single injection of fluoxetine immediately before conditioning enhances the recall of fear memory a day later. Surprisingly, fluoxetine pretreatment was accompanied by increased Arc protein expression in the CeA and BNST but not in the LA or BA. This is consistent with recent reports identifying a critical role for the CeA in the acquisition of auditory fear memory.<sup>19,49</sup> Although enhanced Arc expression in the CeA suggested a potential role for this structure in enhancing fear memory after acute systemic treatment with SSRIs, we found that local infusion of fluoxetine directly into the CeA did not affect fear memory. Instead, local infusion of fluoxetine into the BNST, a closely related component of the extended amygdala, was sufficient to enhance both fear memory and Arc protein expression in the CeA. Together these results suggest that systemically administered fluoxetine may act within the BNST to drive both Arc expression in the CeA and enhance fear learning.

## The acute effects of SSRI treatment on fear and anxiety.

The acute effects of SSRIs have been demonstrated in numerous models of anxiety-like behavior.<sup>11–15</sup> The amygdala is known to be an important component of anxiety circuits in the brain<sup>50,51</sup> and aberrant amygdala activity is associated with affective symptoms of numerous psychiatric disorders.<sup>52–55</sup> Furthermore, there are reports that acute administration of SSRIs increase amygdala activity<sup>56,57</sup> and fear-potentiated startle responses in healthy humans.<sup>58</sup> Although this work implicates the amygdala in the acute anxiogenic effects of SSRIs, the key unresolved issue was to identify specific components of the amygdala that mediate these effects.

## Identifying amygdalar circuits involved in the acute effects of SSRIs.

To probe the neural substrates underlying the acute effects of fluoxetine, we analyzed the impact of fluoxetine on the expression of Arc, an IEG that is induced by behaviorally relevant stimuli and has a critical role in consequent synaptic plasticity.<sup>59,60</sup> Arc mRNA undergoes rapid transport to dendrites and is enriched at local sites of synaptic activity, a process requiring NMDA (*N*-methyl-D-aspartate)-receptor activation.<sup>61</sup> Fear conditioning, which also requires activation of NMDA receptors,<sup>62,63</sup> is known to induce the expression of Arc protein in both the LA and BA. Furthermore, this upregulation of Arc is required for the subsequent expression of fear memory.<sup>27</sup> Although previous studies report that acute SSRI treatment modulates IEG



expression in naive as well as in stressed animals in several brain areas, including the basolateral amygdala (BLA), medial amygdala, CeA and BNST,<sup>15,64–66</sup> nothing is known about how SSRIs affect the induction of Arc expression in sub-regions of the amygdala following cued fear conditioning. We hypothesized that acute SSRI treatment may potentiate fear memory by affecting activity of the LA and BA. Surprisingly, we found that fluoxetine pre-treatment had no effect on Arc expression in the LA or BA, despite fear conditioning itself enhancing Arc expression in these areas in the absence of fluoxetine. However, Arc expression in the CeA was significantly enhanced with fluoxetine treatment, suggesting a role for CeA activation in fluoxetine-induced potentiation of fear memory.

It is interesting to note that in our study the effect of acute fluoxetine is evident only in the CeA of fear conditioned, but not naive, animals. It is possible that fluoxetine, which increases extracellular levels of serotonin by blocking the serotonin transporter, does not change Arc levels under baseline conditions, because levels of serotonin are low. During fear conditioning, serotonin levels increase<sup>67–69</sup> and are further elevated by blockade of the serotonin transporter, both of which appear to be required for Arc expression in the CeA to increase.

**Role of the extended amygdala.** The CeA is the primary output nucleus of the amygdaloid complex that mediates the expression of conditioned fear responses via its projections to downstream brain areas.<sup>23</sup> Although much attention has focused on the role of the BLA in the acquisition of auditory fear conditioning, several recent studies show that the CeA may function in parallel with the BLA to make distinct contributions to fear and anxiety-like behavior. Indeed, inactivation of the CeA impairs fear learning,<sup>70,71</sup> and rats with BLA lesions can learn conditioned fear responses in an associative, CeA-dependent manner.<sup>72,73</sup> Importantly, activity-dependent plasticity within the CeA is essential for the acquisition of fear conditioning.<sup>25,65,66</sup> Moreover, diverse agents ranging from alcohol to neuropeptides like oxytocin are known to produce anxiolytic effects by modulating inhibitory microcircuits in the CeA.<sup>74,75</sup>

Although our data suggest that elevation of Arc expression in CeA may be a neural correlate of the facilitation of fear memory by systemic fluoxetine, direct infusion of fluoxetine into the CeA failed to enhance fear memory. This indicates that fluoxetine does not mediate its behavioral effects on fear conditioning by acting directly on the CeA and instead raises the possibility that fluoxetine may act on other brain areas that, in turn, project to the CeA to modulate fear memory. Indeed, systemic fluoxetine treatment similarly increased Arc protein expression in the BNST. We found that direct infusion of fluoxetine into the BNST before conditioning was sufficient to enhance both fear memory, as well as Arc expression in the CeA. In other words, the direct action of fluoxetine on the BNST has the same functional consequences as systemic treatment.

To evaluate whether the upregulation of Arc expression is specific to the CeA and BNST, we also analyzed Arc expression in the DG of the hippocampus. We found that fluoxetine treatment decreased Arc expression in the DG

regardless of whether or not animals were fear conditioned, providing further support for a specific role of the extended amygdala in the acute effects of fluoxetine. This result is consistent with earlier reports of a decrease in hippocampus-dependent contextual fear memory following acute SSRI treatment.<sup>43,44</sup> These results are also interesting in light of studies that implicate the hippocampus in antidepressant effects<sup>47</sup> and reports of contrasting effects of stress on the hippocampus and amygdala<sup>76</sup> and warrant further investigation in future studies.

The involvement of both the BNST and CeA is intriguing because of substantial evidence that the two regions mediate distinct fear responses, despite their extensive similarities. The CeA is an integral part of fear circuits that mediate stimulus-specific conditioned fear responses to discrete sensory cues and is essential for the acquisition and expression of auditory fear conditioning.<sup>17,26</sup> On the other hand, the BNST is not necessary for acquisition or expression of fear to a discrete conditioned cue.<sup>23,77–79</sup> Instead, the BNST mediates non-specific, ‘anxiety-like’ sustained fear states in response to more diffuse environmental contingencies, such as light-induced startle and corticotropin-releasing factor-induced anxiety.<sup>26</sup> However, many recent studies suggest that the two fear systems do interact and are not completely independent. For instance, although the BNST is not required for conditioned fear responses to brief sensory cues, it is required for conditioned responses to long duration cues paired with a shock.<sup>80,81</sup> The BNST is also required for contextual fear conditioning.<sup>77,82,83</sup> Further, animals with BNST lesions display less fear generalization.<sup>82</sup> Together, these studies suggest that although the BNST is not involved directly in the formation of brief stimulus-specific memory, other aspects of conditioned fear, such as the effect of context and stimulus specificity, may be modulated by the BNST.

Our results suggest that although the BNST is not normally required for the acquisition and expression of conditioned fear memory, acute systemic fluoxetine treatment recruits the BNST into the fear circuitry, leading to an increase in fear memory, possibly mediated via the CeA. Although our experiments suggest that pre-treatment with fluoxetine engages the BNST during fear learning, our data does not address whether the BNST is also active during memory recall. Future studies aimed at measuring IEG expression in the BNST during fear expression may help in addressing this possibility. Further, studies involving transient inactivation of the BNST during the conditioning or testing sessions in different groups of rats pre-treated with fluoxetine will help verify a role for the BNST in the acquisition and expression of conditioned fear.

The results presented here raise questions regarding how SSRIs modulate BNST function. The increased availability of serotonin (5-hydroxytryptamine (5-HT)) due to SSRIs may act on numerous candidate 5-HT receptors in the BNST.<sup>41,84</sup> Interestingly, activation of the 5-HT<sub>2A/2C/7</sub> receptor sub-types *in vitro* mediates an excitatory response to 5-HT that may be anxiogenic.<sup>41</sup> Consistent with this, pre-treatment of rats with a 5-HT<sub>2C</sub> antagonist blocked the acute fear-enhancing effect of the SSRI citalopram.<sup>28</sup> Conversely, systemic activation of 5-HT<sub>2C</sub> receptors in rats is anxiogenic<sup>85</sup> and increases the expression of c-fos in the anterolateral BNST.<sup>86</sup> The activation

of BNST by acute SSRI treatment may therefore be mediated by the activation of specific 5-HT receptors on BNST neurons, a possibility that awaits further analysis.

In conclusion, our findings demonstrate that the direct action of fluoxetine in BNST successfully replicates the consequences of acute systemic fluoxetine treatment in terms of a molecular marker of activity and plasticity (induction of Arc expression), its enhanced expression in a specific circuit within the amygdala (CeA) and its behavioral impact (enhanced fear memory recall). The BNST appears to have a novel modulatory role in generating stimulus-specific conditioned fear responses, which are also known to involve the CeA.<sup>87</sup> In this regard, our results reflect the importance of expanding our focus from the BLA to output structures of the amygdaloid complex, such as the CeA and BNST. Future studies will be needed to address what the precise role of the CeA is in the acute effects of SSRIs on fear memory and whether this is any different from its normal role during fear acquisition and expression in the absence of SSRIs. This will require an understanding of what specific cellular populations and connections between the CeA and BNST mediate such modulatory influences. Such an approach is likely to provide a more comprehensive experimental framework for studying these short-term undesirable effects as well as the therapeutic effects of long-term SSRI treatment.

### Conflict of interest

The authors declare no conflict of interest.

**Acknowledgements.** This research was supported by the National Centre for Biological Sciences (Grant no. 1143 to SC), the SfN-Neuroscience Scholars Program to NSB and a grant to Barnard College from the Undergraduate Science Education Program of the Howard Hughes Medical Institute. We thank Prof. Andreas Luthi for his helpful advice.

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