

MOLECULAR CHARACTERIZATION OF SEVERE AND MILD CASES OF INFLUENZA A (H1N1) 2009 STRAIN FROM ARGENTINA

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Abstract While worldwide pandemic influenza A(H1N1) pdm case fatality rate (CFR) was 0.4%, Argentina's was 4.5%. A total of 34 strains from mild and severe cases were analyzed. A full genome sequencing was carried out on 26 of these, and a partial sequencing on the remaining eight. We observed no evidence that the high CFR can be attributed to direct virus changes. No evidence of re-assortment, mutations associated with resistance to antiviral drugs, or genetic drift that might contribute to virulence was observed. Although the mutation D225G associated with severity in the latest reports from the Ukraine and Norway is not observed among the Argentine strains, an amino acid change in the area (S206T) surrounding the HA receptor binding domain was observed, the same previously established worldwide.

Key words: H1N1 pdm, influenza, Argentina

Resumen *Caracterización molecular de cepas de influenza A (H1N1) 2009 de casos leves y graves de la Argentina.* Mientras que la tasa de letalidad (CFR) para (H1N1)pdm en todo el mundo era del 0.4%, en la Argentina la mortalidad observada fue de 4.5%. La secuenciación del genoma completo de 26 cepas de virus argentinos de influenza A (H1N1)pdm de casos leves y graves y de 8 cepas secuenciadas parcialmente no mostró evidencia de que la elevada tasa de letalidad se pueda atribuir directamente a cambios en el virus. No se encontraron hallazgos de recombinación, de mutaciones asociadas con la resistencia a los medicamentos antivirales ni de variaciones genéticas que puedan contribuir a la virulencia observada. Si bien la mutación D225G asociada con la gravedad, comunicada en informes procedentes de Ucrania y Noruega, no se ha encontrado en las cepas argentinas estudiadas, se ha observado un cambio aminoacídico en la región (S206T) en torno al dominio del sitio de unión al receptor en la HA, el mismo hallado en cepas distribuidas alrededor del mundo.

Palabras clave: H1N1 pdm, influenza, Argentina

On June 11, 2009, in acknowledgement of sustained global human to human transmission, the WHO declared that the novel influenza A virus (H1N1pdm) was pandemic. Although H1N1pdm has not been reported as more virulent than seasonal H1N1 influenza, its high transmissibility in an immunologically naïve population implies potential for substantial morbidity and mortality and mandates close surveillance for evolution toward increased virulence.

Based on worldwide surveillance data and mathematical modeling, the case fatality rate (CFR) of H1N1pdm was estimated at 0.4%^{1,2}. In contrast, although the first case in Argentina was detected in May 17, 3056 cases with 137 deaths were reported by July 16, representing a computed CFR of 4.5%².

The pathogenesis of infectious diseases is the result of the interaction between the pathogen, the host and the environment (e.g. secondary infections, health care system). Although it is likely that severity will vary from country to country depending on health care resources and the public health measures adopted to mitigate impact, a second possible cause for a significantly higher mortality rate may be changes in the virus itself. Influenza viruses may change in virulence and host range as a function of

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reassortment with strains that contain virulence factors³, or through genetic drift. To investigate these possibilities as explanations for the increased CFR in Argentina we sequenced the genomes of H1N1pdm virus obtained from cases of mild and severe disease.

Materials and Methods

Samples collected included: 50 nasal swabs containing H1N1pdm and 50 virus isolates. Severe cases were defined as having a fatal outcome ($n = 21$) or requiring ventilation support ($n = 15$), (Table 1). Samples here analyzed had been routinely collected for use in approved H1N1 tests at INEI-AN-LIS Malbrán. Samples were identified before being re-analyzed at Columbia University. Data were analyzed anonymously.

RNA was isolated using *TriReagent* (Molecular Research Center, Cincinnati, OH, USA). DNA was removed from RNA preparations by treatment with DNase I (DNA-free, Ambion, Austin, TX, USA). Reverse transcription (RT) reactions were performed using the Superscript II Reverse Transcriptase (Invitrogen, Carlsbad, CA, USA).

Two protocols were used to amplify template for 454 pyrosequencing (Roche Life Sciences, CT, USA). In one protocol, first strand synthesis was initiated with a random octamer linked to an specific artificial primer sequence known as the sequence independent amplification (SIA) primer⁴; a modified protocol was also used wherein first strand synthesis was initiated with the SIA primer doped with a primer mixture containing the same specific sequence linked to influenza virus A (FLUAV), influenza virus B (FLUBV) and influenza virus C (FLUCV) sequences representing the conserved termini of influenza virus genome segments⁵. Random libraries prepared with this method were pyrosequenced. The raw sequencing reads were trimmed and analyzed using GS Reference Mapper (Roche Life Sciences, CT, USA) against the reference sequence A/H1N1/4/California/2009 (accession numbers FJ966079– FJ966086).

Alternatively, random primed cDNA were amplified with primers spanning the whole genome of H1N1pdm for conventional Sanger sequencing. Primers were designed using SCPrimer (<http://scprimer.cpmc.columbia.edu>)⁶ to amplify 800 bp regions of the H1N1pdm genome. A 500 bp staggered overlap strategy was employed to improve fidelity of the sequences. Terminal sequences were added by PCR using a universal influenza A primer, targeting the conserved viral termini (5' ATA TCG TCT CGT ATT AGT AGA AAC AAG G),⁷ combined with specific primers positioned near the ends of the genome segments. Real-time PCR SYBR Green assays (Applied Biosystems, Foster City, CA) were conducted to determine the viral load in each sample (primers available upon request). The libraries yielded between 2475 and 42 188 non-host sequence reads. Mapping indicated coverage ranging from 21% to 99% distributed along the influenza virus 8 segments. Virus sequence representation was directly proportional to viral load. A titer of at least 20 000 genome copies/reaction was necessary to achieve >90% coverage.

For the clustering analysis we performed the Principal Component Analysis (PCA) followed by consensus K-means clustering. A consensus clustering out of the 1000 individual K-means trials was built. Parsimony tree was built of the 8 concatenated segments using the *dnapars* program from the PHYLIP package⁸. Clusters previously found with the K-means algorithm correspond to particular clades in the tree.

Results

Twenty-six complete genomes and eight partial genomes containing complete HA and NA gene were obtained (Table 1). They were analyzed for re-assortment, presence of mutations that confer antiviral resistance and phylogenetic clustering. All 26 genomes obtained contained the S31N mutation in the M2 protein, which confers cross-resistance to the adamantane class of anti-influenza drugs. The most commonly detected mutation in oseltamivir-resistant viruses (H274Y) was not detected in any of the genomes. None of the Argentinean genomes contain the mutation D225G in within the influenza HA receptor binding site (Fig. 1). No re-assortment with regular seasonal influenza strains (either H1N1 or H3N2) was detected.

By analysis of the eigenvectors of PCA, we selected 17 single nucleotide polymorphisms (SNPs) that reliably classified 5 clusters, wherein the difference between any pair of clusters was at least 2 SNPs (Table 2). Each of the clusters is named after the earliest isolate that belongs to it. Our analysis identifies five different clusters of H1N1pdm sequences. Cluster 1 is composed of sequences first identified in Mexico in early April and sequences obtained shortly thereafter in the US, Canada, China and Europe. Cluster 2 contains sequences detected in California, US in early April and their relatives, isolated in the US, Mexico, Canada, Dominican Republic, Japan, China and Europe. Cluster 3 is defined by sequences obtained in Japan in the second half of May. Cluster 4 is composed of isolates from New England, US and Canada obtained in May and June. Cluster 5 is the largest cluster, dates back to the earliest isolates from Mexico (April 19, 2009) and has wide geographical spread, including Mexico, the United States, Argentina, Canada, China and several European countries. Specimens from Argentina belong to cluster 5. It is distinguished by 6 mutations: G2163A(PB2) G658A(HA) G1248A(NP) G492A(MA) G600A(MA) A367G(NS) (Fig.2). The Argentinean isolates (except for #8989) contain a characteristic polymorphic marker T1623C in the PA segment. This substitution does not change the amino acid.

Discussion

PCA analyses revealed that all Argentinean isolates are related suggesting a recent common origin, but given that the newer splits (especially those in the Cluster 5) cannot be resolved completely, we cannot determine whether there was a single H1N1pdm introduction in Argentina or several events involving highly related viruses. Interestingly, one of the phylogenetically informative sites that define a large sub-clade in cluster 5 is the transversion A-T in position 658, that results in a Thr-Ser change in position 206 (H3 numbering). This change occurs in the defined

TABLE 1.– Data for the specimens presented in this study

Sample	Date	Age	Sex	Location, Province	Symptoms	Sample	Accession
7649	15-Jun	63 yrs.	F	Salta City, Salta	mild	isolate	CY047776-83
7785	15-Jun	2 mth.	M	Unknown, Buenos Aires	severe	isolate	CY047784-91
7937	17-Jun	Unknown	Unknown	L. Zamora, Buenos Aires	mild	isolate	CY047792-99
7953	17-Jun	46 yrs.	F	Neuquén, Neuquén	mild	isolate	CY047800-07
7967	17-Jun	35 yrs.	F	Quilmes, Buenos Aires	mild	isolate	CY047808-15
7980	17-Jun	25 yrs.	F	City of Buenos Aires	mild	isolate	CY047816-23
8019	18-Jun	10 yrs.	F	City of Buenos Aires	mild	isolate	CY047824-31
8551	19-Jun	26 yrs.	M	Ezpeleta, Buenos Aires	severe	isolate	CY047832-39
8574	21-Jun	18 mth.	F	C. Suárez, Buenos Aires	severe	isolate	CY047840-47
8673	19-Jun	66 yrs.	F	Bariloche, Rio Negro	mild	isolate	CY047848-55
8989	21-Jun	16 yrs.	F	Olavarria, Buenos Aires	mild	isolate	CY047856-63
8994	22-Jun	30 yrs.	M	Olavarria, Buenos Aires	mild	isolate	CY047864-71
9004	22-Jun	12 yrs.	F	San Pedro, Buenos Aires	mild	isolate	CY047872-79
9180	23-Jun	8 yrs.	M	General Pico, La Pampa	mild	isolate	CY047880-87
9333	20-Jun	58 yrs.	F	Rio Gallegos, Santa Cruz	mild	isolate	CY047888-95
9384	23-Jun	20 yrs.	M	Neuquén, Neuquén	mild	isolate	CY047896-03
9452	22-Jun	9 mth.	M	City of Buenos Aires	mild	isolate	CY047904-11
9576	23-Jun	9 mth.	F	Unknown, Buenos Aires	severe	isolate	CY047912-07
9579	23-Jun	5 yrs.	M	Unknown, Buenos Aires	severe	isolate	CY047920-27
9583	24-Jun	5 yrs.	Unknown	Unknown, Buenos Aires	severe	isolate	CY047928-35
9588	24-Jun	1 mth.	F	Unknown, Buenos Aires	severe	isolate	CY047936-43
9597	24-Jun	1 yrs.	F	Unknown, Buenos Aires	severe	isolate	CY047944-51
9705	24-Jun	9 yrs.	F	Puerto Madryn, Chubut	mild	isolate	CY047952-59
9711	24-Jun	36 yrs.	Unknown	Ushuaia, Tierra del Fuego	mild	isolate	CY047960-67
9721	24-Jun	52 yrs.	F	Ushuaia, Tierra del Fuego	mild	isolate	CY047968-75
12295	27-Jun	Unknown	Unknown	Jujuy, Jujuy	mild	NS*	CY047752-53
12508	27-Jun	Unknown	Unknown	Neuquén, Neuquén	mild	NS*	CY047754-55
13777	30-Jun	50 yrs.	M	Cipolletti, Rio Negro	mild	NS*	CY047756-63
14044	1-Jul	Unknown	Unknown	Tucumán, Tucumán	mild	NS*	CY047764-65
14192	3-Jul	19 mth.	Unknown	Unknown, Buenos Aires	severe	NS*	CY047766-67
14332	3-Jul	1 mth.	Unknown	City of Buenos Aires	severe	NS*	CY047768-69
14591	3-Jul	39 yrs.	M	S. Fernando, Buenos Aires	severe	NS*	CY047770-71
15765	6-Jul	Unknown	M	Cipolletti, Rio Negro	mild	NS*	CY047772-73
18426	9-Jul	48 yrs.	M	City of Buenos Aires	severe	NS*	CY047774-75

*NS: nasal swab

receptor-binding site of FLUAV H1N1. Since the change is observed in both clinical samples and virus isolates, it cannot be attributed to selection in cell culture. It has been modeled extensively and the sites of interaction between the sialic based carbohydrates and the hemagglutinin are well characterized⁹⁻¹³. Moreover, this same region had been described as one of the more significant antigenic epitopes of H1-subtype influenza A virus and related with vaccine efficacy¹⁴. Three secondary structure elements, the 190 helix (residues 190-198); the 130 loop (residues 135 to 138), and the 220 loop (residues 221 to 228) form the sides of the receptor-binding subdomain. Changes in this area (D225G, D190E, K222L^{9, 11}) or others outside

of this area that result in a change in the conformation of the domain (e.g. Thr189¹³), result in changes in the affinity of the receptor-binding site and are believed to be responsible for human, swine or avian hosts adaptation. In addition, the substitution K222L results in complete abolition of binding⁹. Of note, the mutation D225G in this same area has also been observed in fatal cases from the Ukraine and Norway^{15, 16}. Given the limited availability of data on the H1N1pdm strains, we have not been able to demonstrate if this change is positively selected.

In summary, sequencing results of 26 complete genomes of H1N1pdm virus obtained from cases of mild and severe disease indicate continuous virus evolution by

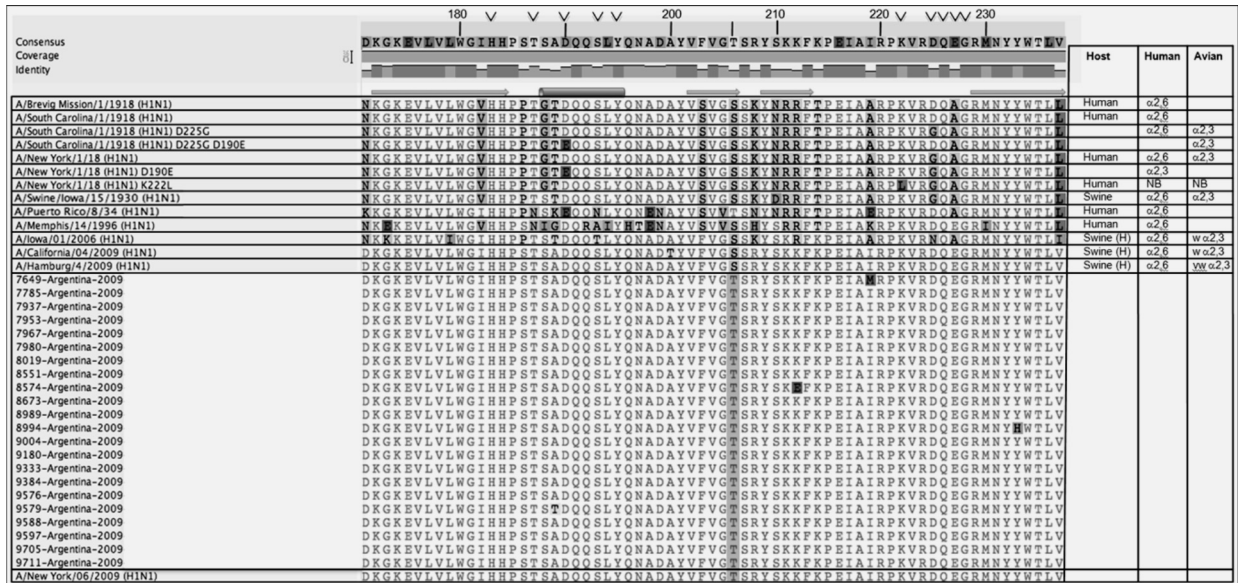


Fig. 1.— Amino acid alignment (in one-letter code) within the influenza HA receptor binding site (position 171- 237; H3 numbering). Positions that had been implicated in receptor specificity are indicated by Δ. Positions 183, 190, 222, 225, 226, 227⁹⁻¹¹, 13 and 189, 193 and 195⁹ are marked. Positions with amino acid sequence changes are highlighted. The box over position 206 highlights the amino acid change in Cluster 5 HA segments. Original host and type of receptor binding specificity are also indicated; Swine (H) are human isolates of influenza virus of swine origin.

TABLE 2.— Single Nucleotide Polymorphisms (SNPs) used to classify 5 clusters

Locus	Consensus	Cluster 1	Cluster 2	Cluster 3	Cluster 4	Cluster 5
888(PB2)	C	*	*	*	*	T
1711(PB2)	T		*	C	*	*
1945(PB2)	G	*	*	A	*	*
603(PB1)	G	*	*	*	*	A
636(PB1)	G	*	*	A	*	*
2000(PB1)	T	*	*	C	*	*
2076(PB1)	C	*	*	*	*	T
936(PA)	G	*	*	A	*	*
1741(PA)	A	C	*	*	*	*
4(HA)	A	*	*	*	*	G
1408(HA)	T	C	C	C	*	*
298(NP)	A	G	G	*	*	*
1118(NP)	C	T	*	*	*	*
1143(NP)	A	G	G	*	*	*
316(NA)	A	G	G	*	*	*
742(NA)	G	A	A	A	*	*
659(NS)	G	*	*	A	*	*

*: no change compared with the consensus

drift, but show no evidence that the Argentinean high case fatality rate can be attributed to changes in viral virulence or resistance to antivirals.

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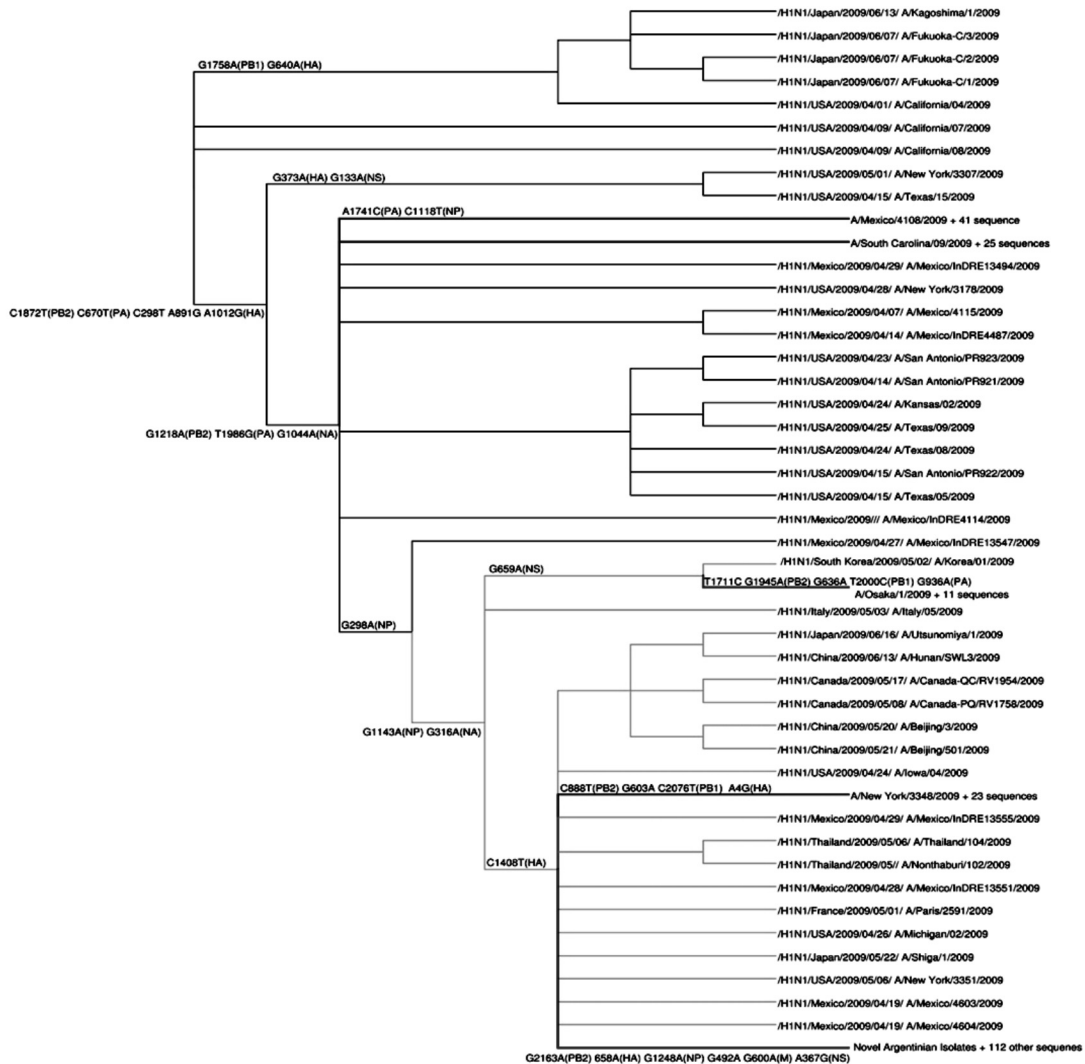


Fig. 2.– Phylogenetic analysis of the H1N1pdm sequences using the Maximum Parsimony method. For building the tree we use the PHYLIP package⁸, and for visualizing it we use the Dendroscope program¹⁷.

Conflicts of interest: The funding organizations had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. S. Hutchison and M. Egholm are 454 Life Sciences employees.

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