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Abstract

Myocardial infarction leads to compensatory ventricular remodeling. Disturbances in myocardial contractility depend on the active transport of Ca²⁺ and Na⁺, which are regulated by Na⁺-K⁺ ATPase. Inappropriate regulation of Na⁺-K⁺ ATPase activity leads to excessive loss of K⁺ and gain of Na⁺ by the cell. We determined the participation of Na⁺-K⁺ ATPase in ventricular performance early and late after myocardial infarction. Wistar rats (8-10 per group) underwent left coronary artery ligation (infarcted, Inf) or sham-operation (Sham). Ventricular performance was measured at 3 and 30 days after surgery using the Langendorff technique. Left ventricular systolic pressure was obtained under different ventricular diastolic pressures and increased extracellular Ca²⁺ concentrations (Ca²⁺_e) and after low and high ouabain concentrations. The baseline coronary perfusion pressure increased 3 days after myocardial infarction and normalized by 30 days (Sham 3 = 88 ± 6; Inf 3 = 130 ± 9; Inf 30 = 92 ± 7 mmHg; P < 0.05). The inotropic response to Ca²⁺_e and ouabain was reduced at 3 and 30 days after myocardial infarction (Ca²⁺ = 1.25 mM; Sham 3 = 70 ± 3; Inf 3 = 45 ± 2; Inf 30 = 29 ± 3 mmHg; P < 0.05), while the Frank-Starling mechanism was preserved. At 3 and 30 days after myocardial infarction, ventricular Na⁺-K⁺ ATPase activity and contractility were reduced. This Na⁺-K⁺ ATPase hypoactivity may modify the Na⁺, K⁺ and Ca²⁺ transport across the sarcolemma resulting in ventricular dysfunction.

Key words: Heart failure; Ouabain; Na*-K* ATPase; Myocardial infarction; Calcium handling

Introduction

Myocardial infarction induces a progressive geometric, structural, and functional remodeling of both ventricles (1-4). Several adaptive changes resulting from the myocardium infarction, including myocyte hypertrophy (5), increased deposition of extracellular matrix components (6), and chamber enlargement (7,8), seem to depend either on hemodynamic disturbances determined by the loss of contractile tissue (9) or on the neurohumoral activation triggered by myocardial ischemia (10-12).

Studies in intact and isolated preparations have suggested that contractile abnormalities may depend on changes in transient calcium concentrations (13) and on the decreased Ca²⁺ sensitivity in the contractile machinery of the ventricular myocardium (4,14). The influences of the coronary vasculature and its perfusion on the diastolic properties of the heart and on cardiac contraction have also been well established. Coronary perfusion affects the diastolic ventricular properties through changes in the lengthwise stiffness of the coronary vasculature. Under low oxygen conditions, an increase in perfusion enhances cardiac muscle contractility via the increased oxygen supply (15). However, the best described disturbances in myocardial contractility are intimately dependent on the active transport of ions, namely Ca2+ and Na+, which are regulated by SERCA and Na⁺-K⁺ ATPase. Na⁺-K⁺-ATPase is a heteromeric protein consisting of α and β subunits. While the α subunit contains the amino acids involved in catalytic function, ion transport and cardiac glycoside binding, the function of the β subunit is not yet fully understood, although it is essential for the normal activity of the enzyme and is involved in the transport of the functional Na⁺-K⁺ ATPase in the plasma membrane (16). Ouabain binds to

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Na⁺-K⁺ ATPase at a site formed in the extracellular part of the catalytic α subunit by the H1-H2, H3-H4, and H5-H6 loops. Reversible interaction of cardiotonic steroids with this site may induce a conformational change in the Na⁺/K⁺ ATPase protein. In the active Na⁺-K⁺ ATPase, cardiotonic steroids are fixed leading to the enzyme's inactivation. In cardiac myocytes, ouabain induces an intracellular increase of Na⁺ concentration, resulting from inhibition of the Na⁺-K⁺ ATPase, subsequently increasing the intracellular Ca²⁺ concentration via Na⁺/Ca²⁺ exchanger inhibition, causing a positive inotropic effect (17). Thus, inappropriate regulation of Na⁺-K⁺ ATPase activity, which leads to excessive loss of K⁺ from the cell and gain of Na⁺ into the cell, could result in disturbances in Ca²⁺ signaling and contractility.

The Na⁺-K⁺ ATPase has a binding site for endogenous cardiac glycosides (18,19). It has been proposed that an increase in endogenous ouabain levels could be involved in the pathogenesis of congestive heart failure (20). Furthermore, although inhibition of Na⁺-K⁺ ATPase by cardiac glycosides is well documented and increases in intracellular Na⁺ and Ca²⁺ have been observed in the presence of cardiotonic steroids, a direct connection between a reduction in Na⁺-K⁺ ATPase activity and contractility in the heart failure following myocardial infarction (MI) has not been demonstrated.

Since levels of endogenous digitalis-like factor increase in the plasma of humans (18,19) after MI and during heart failure, we designed this study to assess the exogenous effect of ouabain on heart contractility performance early and late after myocardial infarction in rats. Since rodents exhibit an ouabain-insensitive α_1 -isoenzyme (21), we perfused a higher concentration of ouabain in the isolated heart (50 nM) than usually observed in human plasma during heart failure (18). Our hypothesis was that a decrease in Na⁺-K⁺ ATPase activity could contribute to the reduced myocardial contractility occurring early and late after MI.

Material and Methods

Animals and coronary artery occlusion

Adult male Wistar rats aged 8-12 weeks at the beginning of the study were used. They were housed under controlled temperature and light conditions with free access to standard rat chow and water. Care and use of the laboratory animals were in accordance with NIH guidelines and the local Ethics Committee at EMESCAM (Escola Superior de Ciências da Santa Casa de Misericórdia de Vitória; Protocol #03/2007).

Animals were anesthetized with ether and a thoracotomy was performed at the level of the fourth left intercostal space to exteriorize the heart. The anterior descending branches of the left coronary artery were occluded between the tip of the left atrium appendage and the right ventricle outflow tract with 6-0 mononylon thread (3). After coronary ligature the chest was rapidly closed and spontaneous respiratory movements were resumed. Most of the animals developed an anterolateral transmural infarction involving 20-40% of the left ventricular surface. Sham-operated rats, used as control, were similarly treated except for the coronary artery ligation. The mortality rate was approximately 30%, with most cases occurring within 30 min of coronary ligation. After recovery from anesthesia, the animals were kept in collective cages in animal care and control facility of the department.

Hemodynamic and left ventricular function

Surgical instrumentation was performed using aseptic surgical procedures. Rats were anesthetized with chloral hydrate (0.3 g/kg, *ip*) and the carotid artery was cannulated with a polyethylene catheter (PE 50, Clay-Adams, USA), and filled with heparin in saline (50 U/mL). The cannulas were advanced into the left ventricular chamber and connected to pressure transducers (TSD 104A) and a Biopac System (MP 100; USA) to measure ventricular pressure. After stabilization the pressure waves were recorded for 1 to 3 min. The following parameters were analyzed: left ventricular systolic and end-diastolic pressures (LVSP and LVEDP, respectively), their first derivative (positive and negative, dP/dt_{max} and dP/dt_{min}, respectively) and heart rate.

Isolated heart perfusion

Groups of infarcted and Sham-operated rats were assessed at 3 and 30 days after surgery. The rats were treated with heparin (50 IU, ip) and anesthetized with pentobarbital sodium (40 mg/kg body weight, ip). Ten minutes later, the heart was excised, mounted in an isolated organ chamber and perfused according to the Langendorff technique at a constant flow (10 mL/min) with Krebs-Henseleit bicarbonate-buffered solution containing: 120 mM NaCl, 5.4 mM KCl, 1.2 mM CaCl₂, 2.5 mM MgSO₄, 1.2 mM Na₂SO₄, 2.0 mM NaH₂PO₄, 24 mM NaHCO₃, and 11 mM glucose. This solution was filtered through a 0.8-µm nitrocellulose filter (Millipore; Sigma, USA) and aerated with 5% CO₂ and 95% O₂, pH 7.4, at 35°C. The left atrium was opened and a soft distensible balloon mounted at the tip of a rigid plastic tube was inserted into the left ventricular cavity through the atrioventricular valve. The balloon was connected via a Y piece to a pressure transducer (Gould P23XL, USA) and to a syringe so that the diastolic pressure of the left ventricle cavity could be adjusted to predetermined values by injecting water into the balloon. The developed pressure was registered with a chart recorder (FUNBEC, RG 300, Brazil).

The hearts were driven with isolated suprathreshold rectangular pulses (5-ms duration) at a constant rate (3.3 Hz) through a pair of Ag/AgCl electrodes attached to the upper region of the right ventricle. All measurements began 30 min to 1 h after mounting to allow the beating preparation to adapt to the *in vitro* conditions. The coronary perfusion pressure (CPP) was continuously recorded by connecting a

pressure transducer (Gould P23XL) to the inflow of the aortic pressure tube. Since coronary flow was maintained constant throughout the experiment (10 mL/min), changes of CPP depended on changes of coronary resistance. Ventricular function curves were obtained by measuring the LVSP developed while diastolic pressure was increased from 0 to 30 mmHg in steps of 5 mmHg. This curve was obtained under control conditions and repeated 30 min after the addition of 50 nM ouabain (Sigma) to the perfusion solution. In order to determine the inotropic response to Ca²⁺, the diastolic pressure was measured at increasing Ca²⁺ concentrations (0.62, 1.25, and 2.5 mM) before and after the addition of 50 nM ouabain (30 min) to the perfusion solution.

At the end of experimental protocol, the inotropic response to increasing concentrations of ouabain (1, 10, and 100 μ M) was tested in an environment with an extracellular Ca²⁺ concentration of 1.25 mM. The concentration of ouabain used here was significantly higher than the plasma concentrations of ouabain or the dose-delivered pharmacological treatment. It was only used as a tool to understand the functional participation of Na⁺-K⁺ ATPase during the initial and late phase after myocardial infarction.

During the protocols performed in this study, experiments without ouabain were used as the control condition to be compared with ouabain perfusion. The only exception was in Figure 3. In this case, the CPP and LVSP responses were analyzed in the presence of increasing ouabain concentrations (1, 10, and 100 μ M). In these experiments, we considered the low dose of ouabain as a baseline condition (control, using 50 nM ouabain). The justification for this was our interest in determining the effects of the increased ouabain concentration compared to the lowest concentration (50 nM ouabain) that is closest to the endogenous plasma ouabain concentration.

Determination of infarct size

The presence of infarction was initially determined by gross observation of the heart. At the end of each experiment the ventricles were separated, blotted and weighed and their wet weights were normalized to the respective body weight. The infarct size 3 days after coronary ligation was evaluated by staining four coronal sections of the left ventricle in 1% tetrazolium chloride for 5 min (22). After staining, the infarcted tissue was dissected from the surviving muscle under microscopic visualization. Both fragments were blotted and weighed and the scar size was reported as the percentage of the left ventricle not stained during the staining procedure. In the groups sacrificed 30 days after surgery, the fibrous scar was separated from the remaining left ventricle muscle. The outlines of both fragments were drawn on graph paper and the areas were measured using the cross-point method (3). The interventricular septum was considered to be part of the left ventricle. Only hearts with a fibrotic scar covering more than 30% and less than 40%

of the left ventricular endocardial surface were included in this study.

Na⁺-K⁺ ATPase activity

The enzymatic material was extracted from the left ventricle by the method of Velema and Zaagsma (23) with the following modifications: the left ventricle was homogenized in a solution containing 20 mM Tris-HCl and 1 mM EDTA, pH 7.5. The homogenized tissue was centrifuged at 10,000 g for 20 min and the precipitate was discarded. The same volume of solution was added to the supernatant and centrifuged at 10,500 g again for 1 h. The precipitate was resuspended in 20 mM Tris-HCl and 1 mM EDTA, pH 7.5, in a final volume of 400 µL. Na+-K+ ATPase activity was assayed by measuring Pi liberation from 3 mM ATP in the presence of 125 mM NaCl, 3 mM MgCl₂, 20 mM KCl and 50 mM Tris-HCI, pH 7.5. The enzyme was preincubated for 5 min at 37°C and the reaction was initiated with the addition of ATP. Incubation times and protein concentration were chosen in order to ensure that the measurements were made in the linear part of the reaction. The reaction was stopped by the addition of 200 µL 10% trichloroacetic acid. Controls containing added enzyme preparation after the addition of trichloroacetic acid were used to correct for nonenzymatic hydrolysis of the substrate. All samples were run in triplicate. Specific activity is reported as nmol Pi released · min⁻¹·mg protein⁻¹ unless otherwise stated. The specific activity of the enzyme was determined in the presence and absence of 50 nM ouabain. Protein concentration was measured by the method of Bradford (24) using bovine serum albumin as standard.

Statistical analysis

Data are reported as means \pm SEM. Continuous variables were compared by one-way and two-way ANOVA followed by the Tukey test to determine differences between groups. Relationships between variables were evaluated by univariate linear regression analysis and comparisons between the two groups were made using the Student *t*-test. A P value of < 0.05 was considered to be statistically significant.

Results

Hemodynamic and heart parameters are shown in Table 1. At 30 days after MI, rats presented a marked increase of LVEDP and a concomitant reduction in indices of left ventricular contractility and relaxation (dP/dt). The infarct area at 3 and 30 days after left coronary occlusion covered about 1/3 of the left ventricular endocardial surface, representing 36 ± 4 and $32.1 \pm 2\%$, respectively. Our previous experience (3,4) has indicated that ligation of the left main coronary artery with this range of scar area resulted in left ventricular dysfunction characterized by a reduction in the maximum positive and negative dP/dt, decreased LVSP and

	3 days		30 days	
	Sham (N = 8)	Infarcted (N = 8)	Sham (N = 10)	Infarcted (N = 10)
Body weight (g)	320 ± 16	315 ± 12	345 ± 13	325 ± 15
LV/BW (mg/g)	2.1 ± 0.09	2.30 ± 0.1	2.16 ± 0.12	2.26 ± 0.12
RV/BW (mg/g)	0.56 ± 0.02	0.62 ± 0.04	0.55 ± 0.03	$0.67 \pm 0.05^{*}$
Infarct size (%)		36 ± 4		32.1 ± 2
Aortic SP (mmHg)	115 ± 3	112 ± 3.4	102 ± 2.1	98 ± 4.1
Aortic DP (mmHg)	82 ± 6.1	89 ± 4.6	79 ± 3	70 ± 3
LVSP (mmHg)	125 ± 8	118 ± 5	101.5 ± 7	100.2 ± 8
LVEDP (mmHg)	2.5 ± 0.7	$10.5 \pm 0.3^*$	2.9 ± 0.7	18.9 ± 1.2*
Heart rate (bpm)	303 ± 17	352 ± 13*	319 ± 16	340 ± 19
LV +dP/dt (mmHg/s)	7628 ± 492	6846 ± 221	6928 ± 355	$4694 \pm 207^{*+}$
LV -dP/dt (mmHg/s)	7296 ± 198	7081 ± 327	6833 ± 710	$4029 \pm 480^{*+}$

Table 1. Characteristics of the animal groups.

Data are reported as means \pm SEM for N rats in each group. LV/BW, RV/BW = left and right wet ventricular weight-to-body weight ratio, respectively; SP = systolic pressure; DP = diastolic pressure; LVSP = left ventricular systolic pressure; LVEDP = left ventricular end diastolic pressure; +dP/dt and -dP/dt = first derivative of pressure positive and negative, respectively. *P < 0.05 compared to appropriate Sham group and *P < 0.01 compared to infarcted group at 3 days (unpaired Student *t*-test).

increased LVEDP. Subsequent hypertrophic growth was observed in both chambers. In the present study, coronary artery occlusion did not affect normal body growth because body weight was not different among the groups. The left ventricle weight-to-body weight ratio did not change in either infarct groups; however, right ventricle hypertrophy was observed at 30 days after coronary occlusion.

Coronary perfusion pressure and ventricular performance

Figure 1 shows that the hearts of the 3-day infarcted (Inf) group perfused with normal Krebs solution (1.25 mM Ca²⁺) presented increased CPP when compared to the respective Sham group. One interesting feature is that CPP tended to decrease towards control values in the 30-day group. This behavior was maintained at lower (0.62 mM) and higher (2.5 mM) extracellular Ca²⁺. In the Sham groups, CPP was constant throughout the observation period.

CPP was measured after extracellular Ca²⁺ was increased from 0.62 to 2.5 mM for both groups at 3 and 30 days after left coronary occlusion (Figure 2, top). The baseline CPP was higher in the Inf group at 3 days but returned to normal at 30 days (Sham 3 = 88 ± 6; Inf 3 = 130 ± 9; Inf 30 = 92 ± 7 mmHg; P < 0.05). Coronary perfusion pressure variation produced by an increase in extracellular Ca²⁺ from 0.62 to 2.5 mM before and after 50 nM ouabain perfusion was similar between groups at 3 days (Sham before ouabain: 15.4 ± 4.3 vs after ouabain: 43.6 ± 8.5 mmHg; Inf before ouabain: 32.6 ± 6 vs after ouabain: 38.4 ± 10 mmHg) and at 30 days (Sham before ouabain: 12.5 ± 8 vs after ouabain: 33.8 ± 9 mmHg; Inf before ouabain:

24.6 \pm 3 *vs* after ouabain: 40.6 \pm 4 mmHg) as determined by two-way ANOVA.

Figure 2 shows that extracellular calcium enhanced the LVSP at 3 and 30 days after infarction or Sham operation. However, the infarcted hearts developed a smaller LVSP compared to the Sham group. Second implication of these results is that the inotropic response to the increase in extracellular Ca²⁺ was depressed in both infarcted groups. The reduced ventricular performance was observed by a smaller increment in the relative LVSP when extracellular Ca²⁺ was increased from 0.62 to 2.5 mM. The percent changes in LVSP increased by 300% in the Sham group at 3 days compared to 138% in the Inf group. A similar reduction of the inotropic response to Ca²⁺ was also observed at 30 days.

Ouabain responsiveness

Ouabain (50 nM) perfused for 30 min did not produce an inotropic effect in the Inf group but altered the ventricular performance of the Sham groups at 3 and 30 days (Figure 2, bottom). In the Sham groups, 3 and 30 days after MI, the increment of extracellular Ca²⁺ concentration increased LVSP. However, when 50 nM ouabain was used, a positive inotropic effect was obtained only at 0.62 mM Ca²⁺. At 1.25 mM, the perfusion of Ca²⁺ plus ouabain produced an increase in LVSP similar to that observed in the ouabainfree condition. At 2.5 mM Ca²⁺ plus ouabain, the increase of the LVSP was less than that observed in the absence of ouabain.

Figure 3 (top) presents the changes in CPP at increasing concentrations of ouabain. In both groups only 100 μ M ouabain increased CPP. Although at 3 days CPP was larger in the Inf than in the Sham group, the CPP increase occurred in parallel in both groups. Even at 30 days, when the baseline CPP was similar, ouabain produced the same CPP response in both groups. Figure 3 (bottom) shows the changes in LVSP as ouabain concentration was increased in the perfusion solution from 50 nM to 100 μ M. In both groups the inotropic effect of ouabain was observed only at 100 μ M. The baseline LVSP in the Inf group was lower compared to the Sham group and remained lower when the ouabain concentration was increased. The absolute change of LVSP in response to 100 μ M ouabain was smaller in the Inf groups (Sham 3 days = 25.7 ± 3.5 vs Inf 3 days = 15.3 ± 2.5 mmHg; Sham 30 days = 26.2 ± 5 vs Inf 30 days = 11.3 ± 2.4 mmHg; P < 0.05).

Frank-Starling mechanism of the heart

The Frank-Starling mechanism of the heart was used to determine the intrinsic inotropic response of the cardiac muscle after MI (Figure 4). To evaluate the effects of the increase of LVEDP, the experiments were performed using



Figure 1. Changes in coronary perfusion pressure in Sham and infarcted hearts 3 and 30 days after left coronary occlusion during perfusion with 1.25 mM extracellular Ca²⁺. Diastolic pressure was maintained constant at 5 mmHg. Data are reported as means \pm SEM for 8-10 rats in each group. *P < 0.05 compared to 3-day Sham group (two-way ANOVA and Tukey test).

0.62 mM Ca²⁺. This Ca²⁺ concentration was used because, at least at 3 days, both the Sham and Inf groups presented similar LVSP values at the beginning of the ventricular function curve protocol (Figure 2). Thus, any difference in LVSP achieved after increases of diastolic pressure could be interpreted as being dependent on the ventricular stretch. At 3 days after surgery, under low Ca²⁺ concentration, the increase of diastolic pressure from 0 to 30 mmHg produced a similar inotropic effect in both groups. Ouabain (50 nM) perfused for 30 min produced a further parallel upward



Figure 2. Changes in coronary perfusion pressure (CPP) and left ventricular systolic pressure (LVSP) in response to extracellular Ca²⁺ in Sham-operated rat hearts before (filled circles) and after 50 nM ouabain (open circles), and infarcted rat hearts before (filled squares) and after 50 nM ouabain (open squares) at 3 and 30 days after surgery. Ouabain perfusion increased LVSP at 0.62 mM and decreased LVSP at 2.5 mM Ca²⁺ in the Sham but not in the infarcted hearts. Diastolic pressure was maintained constant at 5 mmHg. Data are reported as means ± SEM for 8-10 rats in each group. *P < 0.05 for Sham group compared to infarcted group (two-way ANOVA plus Tukey test).



Figure 3. Coronary perfusion pressure (CPP) and left ventricle isovolumic systolic pressure (LVSP) response to increased ouabain concentration (1, 10 and 100 μ M) at 3 and 30 days after surgery. C = control using 50 nM ouabain. Diastolic pressure was maintained constant at 5 mmHg at 0.62 mM extracellular Ca²⁺. In the infarcted group, baseline CPP was higher than in the Sham group at 3 days, but equal at 30 days. Data are reported as means ± SEM for 8-10 rats in each group. *P < 0.01 for infarcted *vs* Sham; *P < 0.05 compared to its own control (two-way ANOVA plus Tukey test).



Figure 4. *Top*, Left ventricle systolic pressure curves (LVSP) obtained at different diastolic pressures (DP). Sham-operated before (filled circles) and after 50 nM ouabain (open circles), and infarcted hearts before (filled squares) and after 50 nM ouabain (open squares) at 3 and 30 days after surgery. *Bottom*, LVSP (%) = left ventricular curves normalized to DP of 5 mmHg. Hearts were perfused using 0.62 mM extracellular Ca²⁺. Data are reported as means ± SEM for 8-10 rats in each group. *P < 0.05 compared to the curve without ouabain perfusion (two-way ANOVA and Tukey test).

displacement of the ventricular function curve only in the Sham group. However, at 30 days LVSP was reduced in the Inf group for all diastolic pressures in the presence and absence of 50 nM ouabain. The relative increase of LVSP (Figure 4, bottom) was calculated considering the values obtained at 5 mmHg as 100%. The normalized ventricular function curves were similar in both groups at 3 and 30 days after surgery.

CPP remained unchanged during the diastolic pressure increase for both groups, even during ouabain perfusion (50 nM).

Na⁺-K⁺ ATPase activity

Maximal Na⁺-K⁺ ATPase activity, estimated as nmol Pi released min⁻¹·mg protein⁻¹ in the presence and absence of 50 nM ouabain was reduced after MI (Figure 5). At 3 days, the activity was 69% lower in Inf animals compared to Sham animals, whereas at 30 days the activities were 60% lower in Inf animals compared to Sham animals.

Discussion

In the present study, we analyzed the effects of ouabain on coronary resistance and myocardial contractility at 3 and 30 days after MI in rats. Coronary resistance increased 3 days after MI at all calcium concentrations and normalized by 30 days. Ouabain did not affect these responses. MI reduced the force developed, the calcium responsiveness and the inotropic effects of ouabain, while the Frank-Starling mechanism of the heart was preserved. It is well-known that ouabain increases LVSP by inhibiting Na*-K* ATPase, which leads to an increase of intracellular Na⁺ and, indirectly, in intracellular Ca2+. Different Na+-K+ATPase subunit isoforms are present in the heart. Na⁺-K⁺ ATPase- α_1 is predominant, although there are variable amounts of Na⁺-K⁺ATPase- α_2 in the adult ventricular myocytes of most species (25). Despa and Bers (25) demonstrated that the functional density of Na⁺-K⁺ ATPase- α_2 is 4.5 times higher in the T-tubules than in the sarcolemma, whereas Na⁺-K⁺ ATPase- α_1 is almost uniformly distributed between the T-tubules and sarcolemma. The intracellular Ca2+ increase is believed to be the mechanism responsible for the enhancement of mvofibrillar force development.

In this study, we indirectly measured the coronary resistance after MI to extracellular Ca²⁺, evaluated before and after a low ouabain concentration. Our study demonstrated that the baseline CPP was higher at 3 days but normalized at 30 days after left coronary occlusion (Figure 1). We raised two possibilities to account for this. First, a high CPP in the acute phase could be associated with a reduction in the coronary resistance distribution. If we assume that the coronary circulation is a parallel circuit, then the total resistance is equal to the individual resistance in each resistive element divided by the element number. Thus, the occlusion of one of these elements, for the same flow, would increase the

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Figure 5. Left ventricle Na⁺-K⁺ ATPase activity in a control nonoperated group of rats (CT) and in 3 and 30 days Sham and infarcted (Inf) rats. Data are reported as means \pm SEM for 6 rats in each group. *P < 0.05 *vs* control (one-way ANOVA plus Tukey test).

total resistance. In order to test this hypothesis, we occluded the coronary circulation in vitro in a similar way to that used to produce MI. CPP increased by approximately 15 mmHg (results not shown), while the average CPP increment at 3 days after MI was approximately 25 mmHg. Thus, the increase in total coronary resistance could contribute, at least in part, to the high CPP in the acute phase after MI. However, after 30 days, the CPP was virtually normal and at 90 days it was normal (results not shown). This normalization could be due the angiogenesis process installed at this time (26). An increase in capillary density and in the capillary/myocyte ratio in the surviving myocardium might explain the CPP normalization. The second possibility arises from a change in coronary resistance. We subsequently tested the influence of extracellular Ca2+ and ouabain on coronary resistance. In our studies, the elevation of extracellular Ca2+ from 0.62 mM to 2.5 mM increased CPP in all groups, although the increase was higher in the Inf group when compared with the Sham group. In contrast, a decreased contractile response to the same extracellular Ca2+ concentration was observed in the left ventricular at 3 and 30 days after MI. A low concentration of ouabain did not affect coronary resistance at 3 days and did not seem to affect coronary resistance under the conditions studied here. Another point to consider is that coronary pressureflow relationships are strongly determined by autoregulation. Furthermore, flow reserve depends on autoregulation, and the supply-to-demand ratio assumes that flow is related to muscle metabolism and is negligible in systole. Also, CPP depends on vasomotor tone and cardiac contractility and thus on the mechanical properties of the vasculature and the cardiac muscle (15).

In the present study, Na⁺-K⁺ATPase activity was reduced

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at 3 and 30 days after MI compared to Sham-operated animals. We also observed a large increase in Na⁺-K⁺ ATPase activity in the Sham-operated animals compared to control. Our interpretation of this phenomenon is based on the fact that the control group was used in our study as a non-operated group of rats, although the best control for an infarction is the Sham group. The reason why a Sham group is widely used as a control for MI is the fact that these animals undergo the entire surgical procedure without having the coronary artery occluded. After 3 days the animals may present some inflammatory response due to the surgical procedure, which would be absent in the control group. Thus, Sham animals are a better control for MI than rats not subjected to a surgical procedure (control). The inflammatory response may induce a slight increase in cardiac Na⁺-K⁺ ATPase activity since during the inflammatory response proinflammatory molecules may activate enzymes. An increase in vascular mRNA for inducible nitric oxide synthase was demonstrated in the acute phase of MI (27). In fact, results from our laboratory have demonstrated that the inducible nitric oxide synthase isoform was not detected in the tail artery of control rats not subjected to surgery (27), but was increased in Sham animals 3 days after surgery (28). Another study demonstrated that prostaglandins, key inflammatory mediators, induce vascular smooth muscle relaxation by stimulating Na+-K+ ATPase activity (29). Thus, the increase of cardiac Na⁺-K⁺ ATPase activity early in the Sham group could be associated with the inflammatory response following the surgery.

The inhibition of Na⁺-K⁺ ATPase by glycosides initiates an increase in the force dependent on the decrease in Ca²⁺ extrusion by the Na⁺-Ca²⁺ exchanger, which in turn increases the cellular content of Ca²⁺ (30). In this study, ouabain induced an increase in the contraction force only in the presence of low extracellular calcium (0.62 mM). In the presence of high concentrations of extracellular calcium, ouabain induced a reduction in force in the Sham group but not in the Inf group. It is possible that in the Sham group the ouabain-induced negative inotropic effect in the presence of high intracellular calcium could be due to the condition of calcium overload as previously demonstrated in rat heart in the presence of high calcium concentrations (31,32). However, the ouabain-induced negative inotropic effect on high intracellular calcium was not seen in the Inf group, a fact probably due to a reduced Na⁺-K⁺ ATPase activity in these animals. It has been well established that length-dependent positive inotropic effects on the myocardium are dependent on the myofibrillar responsiveness as well as on the internal Ca2+ handling after each excitationcontraction cycle. The positive inotropic response induced by increased left ventricle diastolic pressure did not change after ouabain perfusion in the Inf group at 3 and 30 days.

It is possible that myocytes from the Inf group were not able to properly use the available intracellular Ca²⁺, which could be due to a reduction of Ca2+ responsiveness and impairment of Na+-K+ ATPase activity. This hypothesis is corroborated by our previous results showing a reduced left ventricular Ca2+-responsiveness after MI. This has been explained by a change in intracellular Ca²⁺ handling (31) or may be due to a reduction in myofilament intracellular Ca²⁺ responsiveness (14,33). This can be explained by the reduction of myocardial ouabain binding sites and Na⁺-K⁺ATPase activity observed after experimental MI (34) after heart failure in dogs (35) and in rats (36). Messenger RNA levels of phospholemman, a member of the FXYD family of small single span membrane proteins with putative ion-transport regulatory properties, were increased in postinfarction (MI) rat myocytes (37). Indeed, it has been proposed that, in addition to reduced expression of Na+-K+-ATPase (25,30,34-36), overexpression of phospholemman may also account for the depressed Na⁺-K⁺ATPase activity observed in post-infarction animals (37).

These results were different in failing human hearts, where increased inotropic response and toxic effects of ouabain have been observed (38). Furthermore, increased plasma levels of endogenous ouabain have been described as a consequence of MI (18) and have been interpreted as a homeostatic response to the diminished cardiac output of patients with heart failure. It is probable that in these situations the physiological role of ouabain is global rather then limited to the myocardium. This may explain why a tissue distal from the heart is also affected. This is the case for skeletal muscle from rats subjected to MI in which ouabain sites were reduced even though their affinity was not different (36,39). Furthermore, our results demonstrate that the inotropic response to ouabain was Ca²⁺-dependent in the Sham group. The positive inotropic response in the Sham group was only seen in the presence of low extracellular Ca2+ concentrations. This is considered to be a pharmacological action of digitalis. However, when extracellular Ca2+ was increased to 2.5 mM, a reduction in the positive inotropic effect was detected (Figure 2). This situation suggests a condition known as calcium overload, in which the contraction force is reduced by a mechanism involving reduced ATP production in myocytes (40). In our study some infarcted hearts (data not shown) did not survive perfusion with a high extracellular Ca2+ concentration associated with 50 nM ouabain, probably due to the development of calcium overload.

In conclusion, the reduced ventricular Na⁺-K⁺ ATPase activity may contribute to the decrease in left ventricular performance during inotropic interventions early and late after myocardial infarction in rats.

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