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Circadian Disruptions in the *Myshkin* Mouse Model of Mania Are Independent of Deficits in Suprachiasmatic Molecular Clock Function

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ABSTRACT

BACKGROUND: Alterations in environmental light and intrinsic circadian function have strong associations with mood disorders. The neural origins underpinning these changes remain unclear, although genetic deficits in the molecular clock regularly render mice with altered mood-associated phenotypes.

METHODS: A detailed circadian and light-associated behavioral characterization of the Na⁺/K⁺-ATPase $\alpha 3$ *Myshkin* (*Myk/+*) mouse model of mania was performed. Na⁺/K⁺-ATPase $\alpha 3$ does not reside within the core circadian molecular clockwork, but *Myk/+* mice exhibit concomitant disruption in circadian rhythms and mood. The neural basis of this phenotype was investigated through molecular and electrophysiological dissection of the master circadian pacemaker, the suprachiasmatic nuclei (SCN). Light input and glutamatergic signaling to the SCN were concomitantly assessed through behavioral assays and calcium imaging.

RESULTS: In vivo assays revealed several circadian abnormalities including lengthened period and instability of behavioral rhythms, and elevated metabolic rate. Grossly aberrant responses to light included accentuated resetting, accelerated re-entrainment, and an absence of locomotor suppression. Bioluminescent recording of circadian clock protein (PERIOD2) output from ex vivo SCN revealed no deficits in *Myk/+* molecular clock function. Optic nerve crush rescued the circadian period of *Myk/+* behavior, highlighting that afferent inputs are critical upstream mediators. Electrophysiological and calcium imaging SCN recordings demonstrated changes in the response to glutamatergic stimulation as well as the electrical output indicative of altered retinal input processing.

CONCLUSIONS: The *Myshkin* model demonstrates profound circadian and light-responsive behavioral alterations independent of molecular clock disruption. Afferent light signaling drives behavioral changes and raises new mechanistic implications for circadian disruption in affective disorders.

Keywords: Bipolar, Circadian, Light, Mania, Mood, Suprachiasmatic

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Bipolar disorder (BPD) is a debilitating mental health condition that affects approximately 0.7% to 0.9% of the population of Western societies (1). BPD is characterized by episodes of depression, euthymia, and mania, but its etiology and neural substrates remain poorly understood (2). Disruption of sleep and circadian rhythms is prevalent in many mental health diseases, including BPD, and as such, underlying circadian systems are implicated within BPD pathophysiology (3). Indeed, treatment of circadian rhythm abnormalities can alleviate symptoms of affective disorders (4,5), while circadian clock gene polymorphisms represent risk factors across neuropsychiatric conditions (6). Therefore, studying the circadian system may provide insight into the mechanisms and root causes of BPD.

Coordinated circadian rhythms in mammals, including humans, originate from the master circadian clock located in the hypothalamic suprachiasmatic nuclei (SCN) (7). SCN

neurons contain an intracellular gene–protein transcription–translation feedback loop (TTFL) that is the molecular basis of circadian timekeeping, and the *Period1/2* (*Per1/2*) genes and their protein products PERIOD1/2 (PER1/2) are key components of this intracellular molecular clock (8). The TTFL drives the SCN neural network to exhibit electrically excited states during the day and relatively quiescent states at night (9). Such variation is key for individual SCN neurons to coordinate their internal clocks, as well as for the SCN to signal and exert temporal control on behavior and physiology (10). Consistent alignment of these central circadian rhythms to the external environmental light/dark (LD) cycle is important for health and well-being (11). Light information signaled directly from intrinsically photosensitive retinal ganglion cells to the SCN is critical in this process (12). This non-image-forming light input pathway, the retinohypothalamic tract, uses the excitatory neurotransmitter glutamate to activate SCN neurons, resetting

the phase of the TTFL and ultimately the timing of behavioral and brain states such as sleeping and waking (13,14). Further, both SCN-dependent and SCN-independent actions of light exert a powerful influence on mood pathology (15).

Intriguingly, in mice, targeted disruption of core TTFL components alters circadian rhythms and consistently elevates the expression of aberrant behaviors resembling those of human affective disorders (16,17). However, because circadian clock genes are also expressed in mood-regulating brain centers (18,19), it is challenging to ascribe behavioral deficits directly to specific brain loci (20). Further, in human BPD, it is unclear whether sleep and circadian disruption arise as etiological drivers or as a consequence of wider pathophysiology. To gain insight into these problems, we used the *Myshkin* (*Myk/+*) mouse, which possesses a heterozygous inactivating mutation in the neuron-specific Na^+/K^+ -ATPase (NKA) $\alpha 3$ subunit, encoded by *Atp1a3*, and models the manic phase of BPD with face, construct, and predictive validity (20,21). Importantly, this mouse has no known TTFL deficit. We report that *Myk/+* animals exhibit behavioral circadian rhythm disruption as well as unusually heightened behavioral responses to light and enhanced activation of SCN neurons in vitro to a neurochemical mimic of light input. Intriguingly, we also found that the *Myk/+* SCN TTFL rhythms are intact, while the electrophysiological output of the *Myk/+* SCN neural network was damped. Critically, we show that period-lengthening effects on behavior of the *Myshkin* mutation are ameliorated through removal of the light input pathway. Circadian abnormalities in the *Myk/+* mice arise through alterations in light signaling and processing by the SCN. This model provides new insights into the etiological mechanisms of circadian disruption in animal models of affective disorders that are independent of core circadian clock gene perturbation.

METHODS AND MATERIALS

Animal Housing and Breeding

Adult congenic *Myk/+* and wild-type (+/+) animals (2–6 months of age) used in this study were bred from pairs (male *Myk/+* × female +/+) of animals that had been backcrossed on to the C57BL/6Ncr strain for 20 generations (20). Pilot investigations revealed no obvious sex differences in behavioral measures in either genotype, so the data from male and female mice were combined (see also Supplemental Figure S1). To generate mice in which the dynamic activities of the molecular clock can be monitored in tissues ex vivo, *Myk/+* mice were crossed with *mPer2^{LUC}* mice bearing a knock-in PER2-luciferase (LUC) construct (referred to here as PER2::LUC mice) (22). Congenic +/+ × PER2::LUC (+/+PER2) and *Myk/+* × PER2::LUC animals were generated through crosses of heterozygous male *Myk/+* and female PER2::LUC animals. All behavioral and in vitro studies of mice on the PER2::LUC background were performed on filial 1 generation animals. See Supplemental Methods for further details.

For assessment of daily rhythms in locomotor activity (with or without a running wheel), ingestive behavior (feeding and drinking), and metabolic activity, animals were housed singly as previously described (23,24). Most studies were conducted under 12-hour LD conditions; however, in some instances

animals were assessed under a day-length (16-hour light/8-hour dark cycle) condition or in constant dark (DD) or constant light (LL). These in vivo investigations were conducted using previously established protocols (23,25); see the Supplement for study-specific details.

For in vitro assessments of electrophysiological activity, calcium transients, and bioluminescence rhythms in PER2::LUC, SCN-containing brain slices were made from adult mice using previously published protocols (26). Whole-cell current-clamp recordings and assessment of calcium transients were performed as previously described (9,26). Rhythms in whole SCN slice PER2::LUC expression were assessed using luminometry, while single cells in SCN slices were visualized and imaged using a Hamamatsu Image EM9100-13 electron-multiplying-CCD (Hamamatsu, Welwyn Garden City, UK) (27,28).

Data Analysis and Statistics

Unless stated otherwise, genotype comparisons were made using two-tailed Student *t* test or by two-way analysis of variance with Sidak post hoc comparisons. For within-genotype comparisons, a one-way analysis of variance was applied with Sidak corrections unless otherwise stated. Nonparametric equivalents and corrections for unequal variances were utilized where appropriate and are detailed in figure captions. The threshold for statistical significance was set at $p < .05$. See the Supplement for further details.

RESULTS

Circadian mechanisms as well as visual and non-image-forming light pathways influence normal and pathophysiological states including metabolism and mood behaviors (15). As such, we sought to determine if and how murine daily and circadian rhythms are influenced by the *Myshkin* mutation.

When singly housed under standard 12-hour LD conditions, the mania phenotype of *Myk/+* mice was characterized by an inability to restrict the normal nocturnally elevated wheel-running activity to the dark phase. Instead, *Myk/+* animals sustained vigorous activity into the first 3 to 4 hours of the lights-on phase (Figure 1A). Consequently, in comparison with +/+ animals, the duration of the daily active (alpha) phase was significantly lengthened by ~2.1 hours (Figure 1B), with the percentage of wheel running occurring during the day elevated by ~6.9% (Figure 1C) in *Myk/+* mice. Some *Myk/+* animals also showed unstable LD rhythms, with activity onsets variably preceding the initiation of the dark phase (Figure 1A). The effects did not rely on the provision of a running wheel, because monitoring of general locomotor activity via infrared detector indicated near-identical disruption in *Myk/+* animals (Supplemental Figure S4A–D). Similar effects of the *Myshkin* mutation on wheel-running activity were observed in a separate cohort of female mice (Supplemental Figure S1A–E).

In humans, extending daily exposure to light (>14 hours per 24 hours) associated with summer months can exacerbate the symptoms of mania (29,30). When transferred into longer day length (16 hours light/8 hours dark), *Myk/+* mice exhibited longer alpha, weaker rhythms, and ~40% higher wheel running in the light phase than +/+ mice (Supplemental Figure S2A–D). This reveals that the *Myshkin* mutation compromises behavioral consolidation and that increasing the

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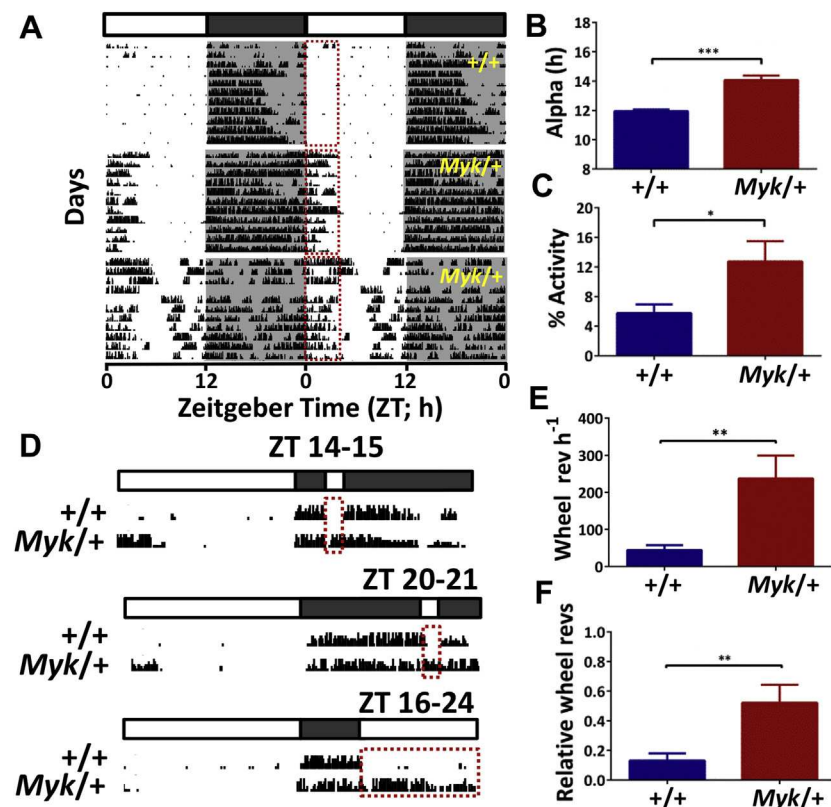


Figure 1. The *Myshkin* mutation alters the suppression of wheel-running behavior by light. **(A)** Example double-plotted actograms from wild-type (+/+) ($n = 37$) and *Myk/+* ($n = 38$) mice maintained under 12-hour light/dark conditions. Red boxes indicate typical region when “tails” in *Myk/+* light-phase activity occur. Gray-shaded areas of actograms delineate lights off (dark). Zeitgeber time 0 (ZT0) = lights on; ZT12 = lights off. **(B)** Alpha duration under these 12-hour light/dark conditions is elongated in *Myk/+* animals (+/+ : 11.95 ± 0.12 hours, *Myk/+* : 14.07 ± 0.32 hours; $p < .0001$). **(C)** Percentage of total daily activity in the lights-on phase is increased by the *Myshkin* mutation (+/+ : $5.8 \pm 1.2\%$, *Myk/+* : $12.7 \pm 2.7\%$; $p = .043$). **(D)** Example single-plotted actograms showing the presence (+/+ mice) and absence (*Myk/+* animals) of negative masking responses to 1-hour or 8-hour light pulses given during the lights-off phase. **(E)** Wheel revolutions per hour exhibited during these 1-hour or 8-hour light pulses are increased by the *Myshkin* mutation (+/+ : 45 ± 13 revolutions/hour $^{-1}$ (rev h $^{-1}$), *Myk/+* : 239 ± 62 rev h $^{-1}$; $p = .005$). **(F)** Locomotor activity during light pulse (running wheel revolutions) normalized to each animal’s daily mean is higher in *Myk/+* mice (+/+ : 0.13 ± 0.05 relative wheel revolutions, *Myk/+* : 0.52 ± 0.12 relative wheel revolutions; $p = .002$). Data are plotted as mean \pm SEM. * $p < .05$; ** $p < .01$; *** $p < .001$.

duration of the lights-on phase overtly disrupts rhythmic control of behavior.

Circadian disruption and mental illness can alter body weight regulation (31–33). Indeed, bipolar patients with mania can exhibit elevated basal metabolic rate (34), so we subsequently profiled metabolic activity in *Myk/+* and +/+ mice. Using indirect calorimetry and monitoring of ingestion activity for 6.5 days under a 12-hour LD cycle, *Myk/+* animals were found to exhibit elevated basal metabolic rate, heat production, and drinking activity (Figure 2A–C, E). The duration of their elevated metabolic activity was sustained into the lights-on phase (Figure 2A, F); ingestive behavior was increased during the lights-on phase but was reduced over 24 hours (Figure 2D, G).

In nocturnal rodents, light exposure typically suppresses locomotor behaviors (negative masking), but when exposed to 1-hour or 8-hour pulses of light during the night, *Myk/+* mice maintained activity or increased wheel-running behavior during the pulses, while +/+ animals reduced locomotor activity (Figure 1D–F). Further, when released into an illuminated ($\sim 1.5 \mu\text{W}/\text{cm}^2$) open-field test arena for 15 minutes during the early night (Zeitgeber time 15–18 [ZT15–18]), hyperlocomotor activity (as assessed by distance traversed) was marked in *Myk/+* but not in +/+ mice (Supplemental Figure S3). Therefore, unlike other nocturnal rodents such as the Syrian hamster (35) as well as mice with TTFL mutations (36), *Myk/+* mice do not exhibit pronounced negative masking, an effect overtly manifested in wheel running during the day.

Because *Myk/+* mice exhibited disrupted rhythms under LD conditions, their intrinsic circadian rhythms in wheel-running

behavior were initially assessed over 14 days in the absence of light (DD). Consistent with a previous report (20), behavioral rhythms of *Myk/+* mice differed significantly from those of +/+ animals. *Myk/+* mice displayed a lengthened period (Figure 3A–D; ~ 24.2 hours vs. ~ 23.5 hours) and an unusually elongated active phase (Figure 3E: alpha; ~ 19.0 hours vs. ~ 13.1 hours), while the amplitude, or strength (percent variance as measured by chi-square periodogram), of their behavioral rhythms was also markedly reduced (Figure 3F). In a separate all-female cohort, *Myk/+* animals also showed similar changes in circadian rhythms of wheel running in DD (Supplemental Figure S1A, B, F–H). These effects of the *Myshkin* mutation were not dependent on the provision of a running wheel (37,38), as similar changes in general locomotor activity rhythms were exhibited by animals monitored with a passive infrared system without a functioning running wheel (Supplemental Figure S4A, B, E, F).

When assessed for an additional 14 to 21 days in DD, all +/+ animals sustained rhythmic wheel-running activity, whereas that of some *Myk/+* mice ($n = 3$ of 38; 8%) weakened and they became circadianly arrhythmic (Figure 3C). Additionally, some *Myk/+* animals (but no +/+ mice) exhibited gradual unusual changes in free-running period ($n = 8$ of 38; 21%) either spontaneously or following transfer to a clean cage (Supplemental Figure S5A–C). Because rhythm amplitude and free-running stability are metrics for the output strength of the circadian system, this indicates that *Myk/+* animals possess diminished and unstable central circadian regulation of behavior and physiology.

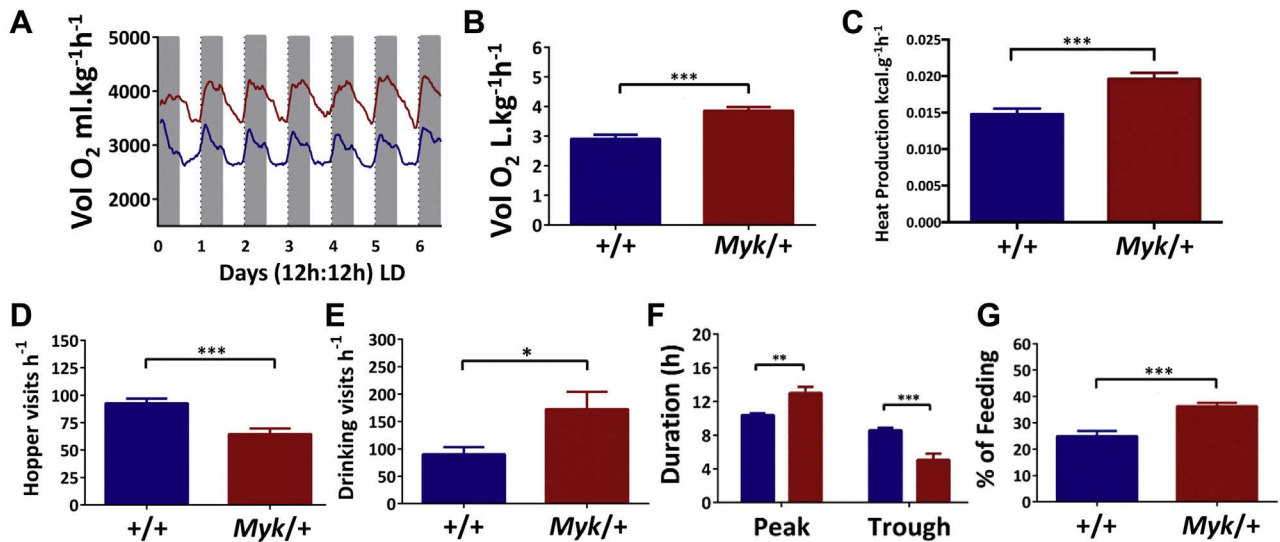


Figure 2. Altered metabolic rhythms in *Myk/+* mice. **(A)** Smoothed traces of wild-type (+/+) mice (blue; $n = 10$) and *Myk/+* mice (red; $n = 12$) maximal oxygen consumption (Vol O_2) over 6.5 days. Gray-shaded columns delineate lights off (dark). **(B)** Mean Vol O_2 consumption is elevated in *Myk/+* animals (+/+ : 2.91 ± 0.14 L/kg/hour, *Myk/+* : 3.86 ± 0.13 L/kg/hour; $p < .0001$). **(C)** Mean hourly heat production is elevated in the *Myk/+* mice (+/+ : 0.0148 ± 0.0007 kcal/g/hour, *Myk/+* : 0.196 ± 0.0008 kcal/g/hour; $p = .0003$). **(D)** Altered ingestive behaviors in *Myk/+* animals. Mean hourly food hopper visits were reduced by the *Myshkin* mutation (+/+ : 92.6 ± 4.3 visits, *Myk/+* : 64.6 ± 5.1 visits; $p = .0005$). **(E)** Mean hourly visits to drinking spout were increased by the *Myshkin* mutation (+/+ : 90.0 ± 13.2 visits/hour, *Myk/+* : 172.6 ± 31.7 visits/hour; $p = .03$). **(F)** Altered duration of daily peak (+/+ : 10.37 ± 0.23 hours, *Myk/+* : 13 ± 0.75 hours; $p = .003$) and nadir (+/+ : 8.59 ± 0.30 hours, *Myk/+* : 5.05 ± 0.75 hours; $p = .0004$) in Vol O_2 activity. **(G)** The *Myshkin* mutation increases percentage of daily feeding occurring during the lights-on phase (+/+ : $25.0 \pm 1.9\%$, *Myk/+* : $36.3 \pm 1.3\%$; $p < .0001$). Data in panels **(C–G)** are plotted as mean \pm SEM. * $p < .05$, ** $p < .01$, *** $p < .001$. LD, light/dark.

For rodents in DD, light exposure during the night shifts subsequent onsets of activity; early night exposure (circadian time 14–18 [CT14–18]) delays circadian rhythms, while light given later in the night (CT20–24) advances rhythm onsets (39), so we next sought to determine if the *Myshkin* mutation affected photic resetting of the circadian system. Mice were released from 12-hour LD conditions into DD and, after 14 days, were exposed to a 1-hour light pulse either late (CT20–21; Figure 4A, B) or early (CT14–15; Figure 4C, D)

in their active subjective night (an Aschoff type I protocol). The subsequent phase of the onset of their wheel-running rhythms was then measured (23). When exposed to light pulses at CT14, *Myk/+* mice showed much larger phase delays than +/+ mice (~ -2.32 hours vs. ~ -1.6 hours, $p = .008$; Figure 4C–E). Light pulse treatment at CT20 evoked phase delays in *Myk/+* activity that were unusual, as they were of the opposite direction to typical advances elicited at this time in +/+ mice (~ -1.3 hours vs. $+0.5$ hours, $p \leq .02$;

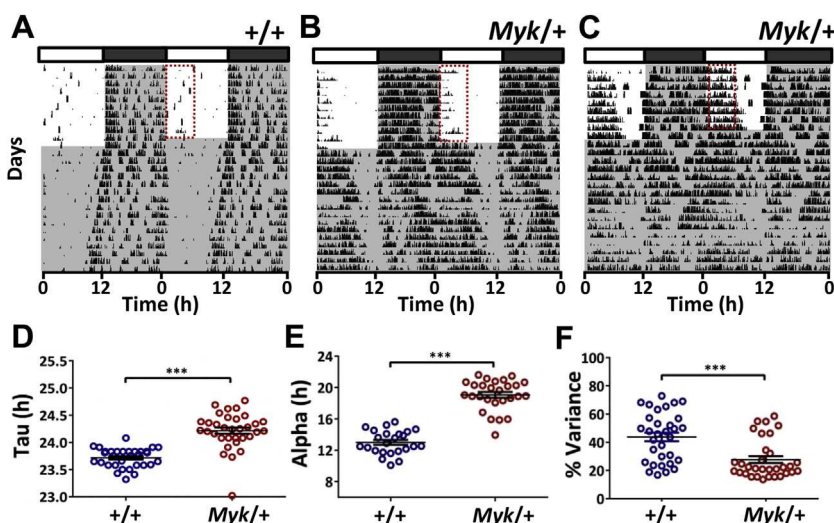
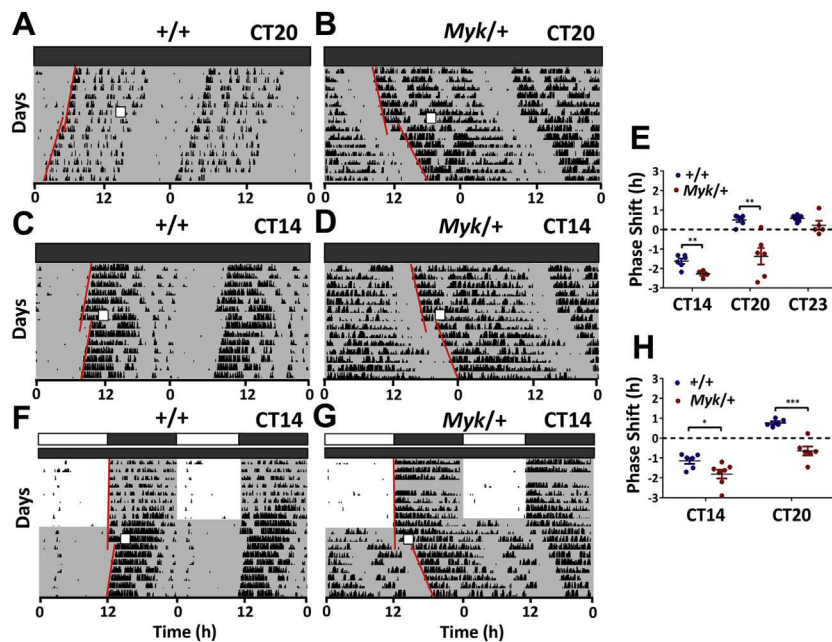


Figure 3. Altered free-running rhythms in *Myk/+* mice. **(A–C)** Example double-plotted actograms of wild-type (+/+) and *Myk/+* animals released into constant dark (indicated by gray shading) following entrainment to 12-hour light/dark cycle. Most *Myk/+* mice maintained rhythms in constant dark **(B)**, but exceptionally, some **(C)** became arrhythmic. Red rectangles delineate timing of typical elevated wheel running in the *Myk/+* mice. **(D)** The *Myshkin* mutation lengthened free-running period (+/+ : 23.72 ± 0.03 hours, *Myk/+* : 24.22 ± 0.02 hours; $p < .0001$) and **(E)** alpha duration (+/+ : 13.14 ± 0.26 hours, *Myk/+* : 18.99 ± 0.37 hours; $p < .0001$). **(F)** The amplitude of daily wheel-running rhythms is reduced by the *Myshkin* mutation (as measured by chi-square periodogram; +/+ : $43.8 \pm 3.1\%$ variance, *Myk/+* : $27.7 \pm 2.4\%$ variance; $p = .001$). Data in panels **(D–F)** are graphed as scatter plots with mean \pm SEM. *** $p < .001$.

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responses to 1-hour light pulses presented under Aschoff type II protocol (CT14, $+/+$: -1.1 ± 0.1 hours, $Myk/+$: -1.8 ± 0.2 hours, $p = .03$; CT20, $+/+$: 0.76 ± 0.08 hours, $Myk/+$: -0.48 ± 0.18 hours, $p = .0001$). Panels (E) and (H) are graphed as scatter plots with mean \pm SEM. * $p < .05$, ** $p < .01$, *** $p < .001$.

Figure 4A, B, E). No genotype differences were found with a light pulse given near the subjective night-day transition (CT23; Figure 4E). This experiment was repeated with 1-hour light pulses given within 48 hours following release from 12-hour LD into DD (an Aschoff type II protocol) with similarly altered resetting patterns observed (Figure 4F–H). This indicates that the altered phase shifts to light observed in $Myk/+$ mice do not emerge as a consequence of long-term adaptation to DD. Thus, the *Myshkin* mutation both accentuates and alters the temporal pattern of the resetting effects of light on murine rhythms in behavior.

We next assessed if the *Myshkin* mutation influenced how mice respond to simulated jetlag. Jetlag and other external disruptors to normal activity rhythms are associated with the presentation of episodes in BPD, and therefore sensitivity to external perturbation represents an important measure (40,41). In response to the 8-hour advance (Figure 5A–D) or delay (Supplemental Figure S5D, E) of the LD cycle, $Myk/+$ mice rapidly altered their daily pattern of wheel running within 2 to 4 days, whereas $+/+$ animals took 6 to 8 days to resynchronize. Comparable rapid resynchronization to an 8-hour advance in the LD cycle was also observed in a separate cohort of female $Myk/+$ mice (Supplemental Figure S1A, B, I, J). To directly test if the *Myshkin* mutation enhances the photic resetting capabilities of the neural circadian system, animals were next subjected to a transient 7-hour advance of the LD cycle for 2 days then released into free-running DD conditions (Figure 5E, F). Again, $Myk/+$ mice demonstrated unusually large advances in the new phase of their activity onsets, and on the first day in DD these were of much greater magnitude (~ 8.3 hours vs. ~ 2.2 hours) than were those shown by $+/+$ animals (Figure 5G, H). This finding indicates that mechanisms that

normally brake the circadian system to prevent extraordinarily large resetting are dysfunctional in $Myk/+$ animals (42,43).

In rodents, exposure to LL suppresses wheel-running behavior and lengthens the period of circadian rhythms (24,44), so we next assessed how $Myk/+$ mice adapt to LL. Consistent with previous research, all 8 $+/+$ animals showed a suppression of wheel running in LL (reduced by $\sim 90\%$ from LD) and exhibited free-running rhythms with a period of ~ 25 hours (Supplemental Figure S1A, B, K). $Myk/+$ animals also showed longer period rhythms in LL (~ 25 hours), but most (5 of 7) exhibited markedly elevated wheel running in LL ($\sim 403\%$ increase from LD). Therefore, while the period-lengthening effects of LL are observed in $Myk/+$ animals, some individuals sustain increased rather than decreased locomotor activity.

The NKA $\alpha 3$ pump is localized to several central nervous system structures, including retinal ganglion cells whose axons project along the optic nerve (45,46), so we next investigated if retinal input to the SCN contributed to the altered circadian rhythms of $Myk/+$ mice. To do so, mice free running in DD either underwent a sham surgical procedure or had their optic nerves crushed (see the Supplement for procedural details). Optic nerve crush in $Myk/+$ mice markedly shortened their circadian period by ~ -0.7 hours to ~ 23.6 hours, while $+/+$ animals showed no obvious change in circadian period (Figure 6A–C). Similarly, $Myk/+$ mice undergoing sham surgical procedure showed no change in free-running period (Figure 6C). This indicates that aberrant intrinsic activity of the light input pathway to the SCN contributes to the period-lengthening effect of the *Myshkin* mutation.

Glutamate is the main neurochemical of the retinal input pathway to the SCN, and stimulation of the alpha-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA)-type

Figure 4. Enhanced circadian resetting responses to light in $Myk/+$ mice. (A, B) Example double-plotted actograms showing resetting effects of a 1-hour light pulse (Aschoff type I protocol) given under constant dark conditions to wild-type ($+/+$) and $Myk/+$ animals (white boxes indicate light pulse) during the late subjective night (circadian time [CT] 20). Note the opposing direction (delay) of the phase shift in $Myk/+$ activity onset compared with the advance in activity onset of the $+/+$ animal. (C, D) Example double-plotted actograms illustrating the shifting effects of a 1-hour light pulse (Aschoff type I protocol) given early in the subjective night (CT14) on the free-running rhythms of $+/+$ and $Myk/+$ animals. $Myk/+$ animals exhibited larger phase delays at CT14. (E) Individual responses to 1-hour light pulses given under Aschoff type I protocol (CT14, $+/+$: -1.6 ± 0.1 hours, $Myk/+$: -2.3 ± 0.1 hours, $p = .008$; CT20, $+/+$: 0.5 ± 0.1 hours, $Myk/+$: -1.3 ± 0.4 hours, $p = .002$; CT23, $+/+$: 0.6 ± 0.1 hours, $Myk/+$: 0.2 ± 0.2 hours, $p = .14$). (F, G) Double-plotted actograms showing the phase resetting responses of $+/+$ and $Myk/+$ mice to 1-hour light pulse given early in the subjective night (CT14) following transfer from light/dark to constant dark conditions (Aschoff type II protocol). Animals were exposed to a 1-hour light pulse within 48 hours following release into constant dark. (H) Individual

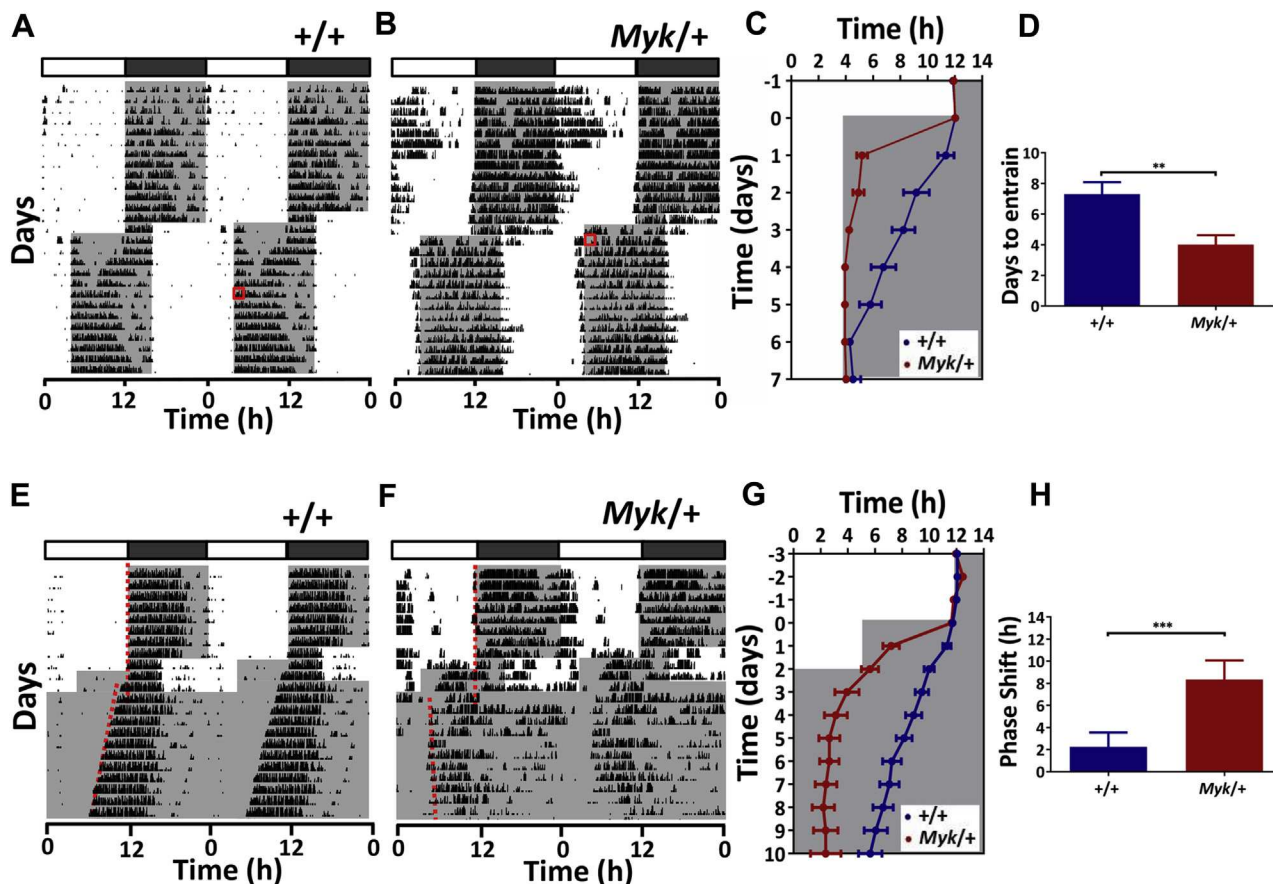


Figure 5. Rapid adaptation to simulated jetlag in *Myk/+* mice. (A, B) Double-plotted actograms of mice exposed to simulated jetlag protocol; here the light/dark (LD) cycle was advanced by 8 hours. Red boxes indicate the time point at which the animals were designated as having re-entrained to the new LD cycle. Gray-shaded areas in actograms and in panels (C) and (G) delineate lights off (dark). (C) *Myk/+* mice readjust their daily onset in wheel-running activity to the new LD phase more rapidly than do wild-type (+/+) mice. Note the large readjustment in the onset of daily wheel running observed in the *Myk/+* mice on day 1 following the advance in the LD cycle. (D) *Myk/+* mice take fewer days to re-entrain their wheel-running activity to the new LD cycle than do +/+ animals (+/+ : 6.5 ± 0.7 days vs. *Myk/+* : 3.6 ± 0.6 days; $p = .006$). (E, F) Double-plotted example actograms of +/+ and *Myk/+* animals responding to transient jetlag. To validate resetting of the circadian system to simulated jetlag seen in panels (A, B), mice were exposed to a 7-hour advance of the 12-hour LD cycle for 48 hours and then released into constant dark. *Myk/+* animals show a large magnitude advance in the timing of their wheel-running activity. (G) The larger readjustment of the onset in daily wheel running of *Myk/+* mice is sustained in constant dark, indicating that the circadian system of these animals has reset to a much larger extent than +/+ mice. (H) Mean phase shift on first 24 hours in constant dark is significantly larger in *Myk/+* mice (+/+ : 2.2 ± 0.5 hours, *Myk/+* : 8.3 ± 0.8 hours; $p < .0001$). Data in panels (C, D, G, H) plotted as mean ± SEM. ** $p < .01$, *** $p < .001$.

glutamate receptors excites SCN neurons, elevates intracellular Ca^{2+} , and can shift the phase of behavioral rhythms (47,48). To determine if acute responses of SCN neurons to glutamate were altered by the *Myshkin* mutation, the actions of AMPA (5–20 μ M) on intracellular Ca^{2+} were assessed in +/+ and *Myk/+* hypothalamic SCN brain slices. Neurons were loaded with the fluorescent calcium indicator dye Fura-2-acetoxymethyl ester to enable recording of somatic intracellular Ca^{2+} changes across populations of single SCN cells (Supplemental Figure S6A).

During the day (ZT4–10), AMPA treatments evoked changes in intracellular Ca^{2+} of similar magnitude and duration in both genotypes (Supplemental Figure S6B, C). However, with applications made during the night (ZT14–18), AMPA elicited increases in intracellular Ca^{2+} that were significantly larger in *Myk/+* neurons compared with +/+ SCN neurons

(Figure 6D, E). At this time, response magnitude was dose dependent (5 and 10 μ M), with baseline recovery from the 10- μ M application taking significantly longer in the *Myk/+* SCN neurons (Figure 6F). Such changes in AMPA responses indicate that, in addition to altering retinohypothalamic tract activity, the *Myshkin* mutation enhances the processing of a neurochemical mimic of this light input pathway to the SCN.

To gain further insight into the etiology of circadian disturbances arising from the *Myshkin* mutation, we next investigated the anatomy and TFL molecular pacemaking of the *Myk/+* SCN. The SCN exhibit characteristic, spatially distinct neuropeptide topography. Immunohistochemical staining for major SCN neuropeptides, vasoactive intestinal polypeptide and arginine vasopressin, showed no overt genotype differences in the pattern of expression (Supplemental Figure S7). Molecular clock activity was then tracked through expression

Altered Circadian Rhythms and Responses to Light in a Manic Mouse

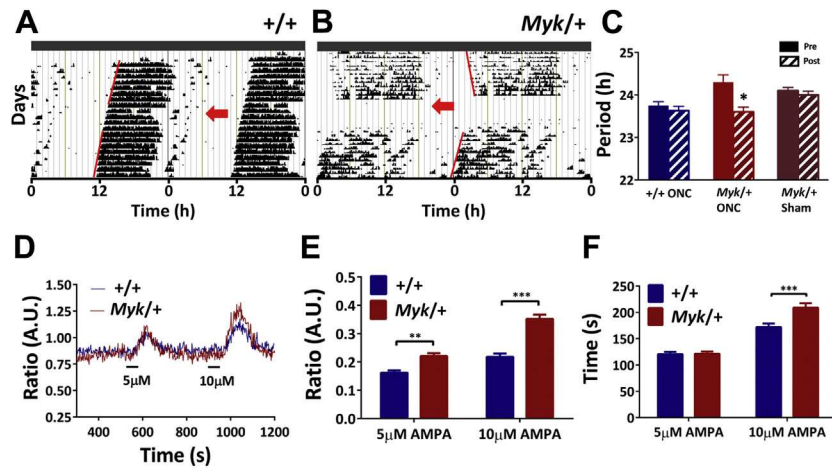


Figure 6. Altered retinal signals and excitatory processing within the suprachiasmatic nuclei (SCN) underlie *Myk/+* circadian behavioral phenotypes. **(A, B)** Double-plotted wheel-running actograms illustrating the effects of optic nerve crush (ONC) in wild-type (+/+) and *Myk/+* animals maintained in constant dark conditions. Red arrows delineate the day of ONC. Red lines are fitted through activity onsets pre- and postsurgical intervention. The free-running period of *Myk/+* mice was restored by ONC but not sham procedure. **(C)** Mean period of cohorts undergoing ONC or sham procedure (+/+ ONC $n = 6$: ~ -0.1 hours, $p = .13$; *Myk/+* ONC $n = 6$: -0.75 hours, $p = .025$; *Myk/+* sham $n = 4$: ~ -0.1 hours, $p = .12$). **(D)** Example responses of single SCN neurons loaded with Fura-2 to alpha-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA) (5 and 10 μ M) tested during subjective night (+/+ : $n = 88$, *Myk/+* : $n = 148$). **(E)** When tested during the subjective night (Zeitgeber time 14–18 [ZT14–18]), *Myk/+* SCN neurons exhibited larger AMPA-evoked cellular increases in intracellular Ca^{2+} than did +/+ SCN neurons (two-way analysis of variance: interaction $p < .0001$; +/+ 5 μ M: 0.16 ± 0.01 arbitrary units (A.U.), *Myk/+* 5 μ M: 0.22 ± 0.01 A.U., Sidak $p = .0029$; +/+ 10 μ M: 0.22 ± 0.01 A.U., *Myk/+* 10 μ M: 0.35 ± 0.01 , $p < .0001$). **(F)** During subjective night (ZT14–18), the washout duration following AMPA treatment (10 μ M) was longer in the *Myk/+* neurons compared with +/+ SCN neurons (+/+ 5 μ M: 121 ± 4 seconds, *Myk/+* 5 μ M: 121 ± 4 seconds, $p = .99$; +/+ 10 μ M: 172 ± 4 seconds, *Myk/+* 10 μ M: 209 ± 8 seconds, $p = .0007$). Data in panels **(C, E, F)** are plotted as mean \pm SEM. * $p < .05$, ** $p < .01$, *** $p < .001$.

of the circadian clock protein PER2 via PER2::LUC expression in SCN explants (Supplemental Figure S8). The characteristics of circadian rhythms in wheel running seen in *Myk/+* mice (increased circadian period and alpha) were maintained in *Myk/+*PER2::LUC mice (Supplemental Figure S8A–E). Surprisingly, in SCN explants cultured ex vivo, no overt genotype differences were found in period or amplitude of whole-tissue PER2::LUC expression (Figure 7A–C). Similarly, in SCN explants imaged at single-cell resolution, neither the period nor the damping rate of single-cell rhythms nor the intercellular synchrony within SCN slices differed between the genotypes (Figure 7D–F). The persistence of +/+PER2-like rhythms ex vivo indicates that, in isolation, the *Myk/+* SCN is a stable pacemaker. Together with the findings from the optic nerve crush experiment, this reinforces the idea that, in vivo, afferent signals such as those coming from the eye act to diminish the *Myk/+* SCN’s control of physiology and behavior.

Neurons of the SCN control behavior and physiology in part by varying their spontaneous firing rate (SFR), with higher frequency discharge during the day than at night (49). Because neurophysiological studies suggest that the NKA pump influences SCN neurons (50), and because NKA $\alpha 3$ subunits affect membrane excitability (51,52), we next made whole-cell electrophysiological recordings and assessed the electrical activity of +/+ and *Myk/+* SCN neurons maintained in brain slices. SCN neurons exhibit distinct electrophysiological states (9,53) and these were evident in both +/+ and *Myk/+* SCN recordings, but no genotype differences were detected in most passive properties of these neurons (Supplemental Table S1). However, comparison of the SFR of *Myk/+* and +/+ SCN neurons based on the time of recording indicated clear genotype-related differences. Unexpectedly, the mean SFR of *Myk/+* SCN neurons did not differ from day to night recordings, whereas +/+ SCN neurons had significantly higher SFR during the day as compared with the night (Figure 7G).

Day/night variation in SFR is a key characteristic of the SCN network both in vitro and in vivo, and because the TTFL appears to be intact in the *Myk/+* SCN, this damping in *Myk/+* SCN neuronal activity most likely arises from exposure to altered photic afferent signals.

DISCUSSION

Here we have demonstrated that, in addition to increases in the period and active phase duration of circadian rhythms in behavior, the *Myk/+* mouse exhibits instability in behavioral rhythms and unusually heightened circadian resetting/re-entrainment responses to light. Other mouse models expressing mania-like states, including *Clock*^{Δ19}, *Reverba*^{α-/-}, and *DAT*^{-/-} mice, also show heightened phase-shifting responses to light (54–56). Interestingly, in patients with mania, locomotor rhythms may also weaken with increasing severity of manic symptoms (57), and sensitivity to both white and blue light is heightened during manic episodes (58–60). Similar to bipolar patients (61,62), *Myk/+* mice exhibit both altered sleep patterns (20) and circadian rhythm disturbance as well as elevated metabolic activity (34). Notably, the nocturnally elevated metabolic rate and locomotor activity of *Myk/+* mice were not suppressed by light, indicating the absence of negative masking. Consistent with this, *Myk/+* mice did not behaviorally adjust to a long day length, and in LL they increased wheel running, whereas +/+ mice markedly reduced it. Indeed, even in the absence of light, alterations in retinal afferents were sufficient to drive circadian behavioral disruption, because removal of afferent photic input to the SCN in vivo restored the circadian period of behavioral rhythms. Further, when cultured in vitro and assessed in isolation from retinal inputs, *Myk/+* single-cell PER2 rhythms and whole-SCN explants behaved as stable pacemakers, comparable to +/+ SCN tissue. This indicates that the SCN molecular clock is largely intact in this mouse model of mania, a finding

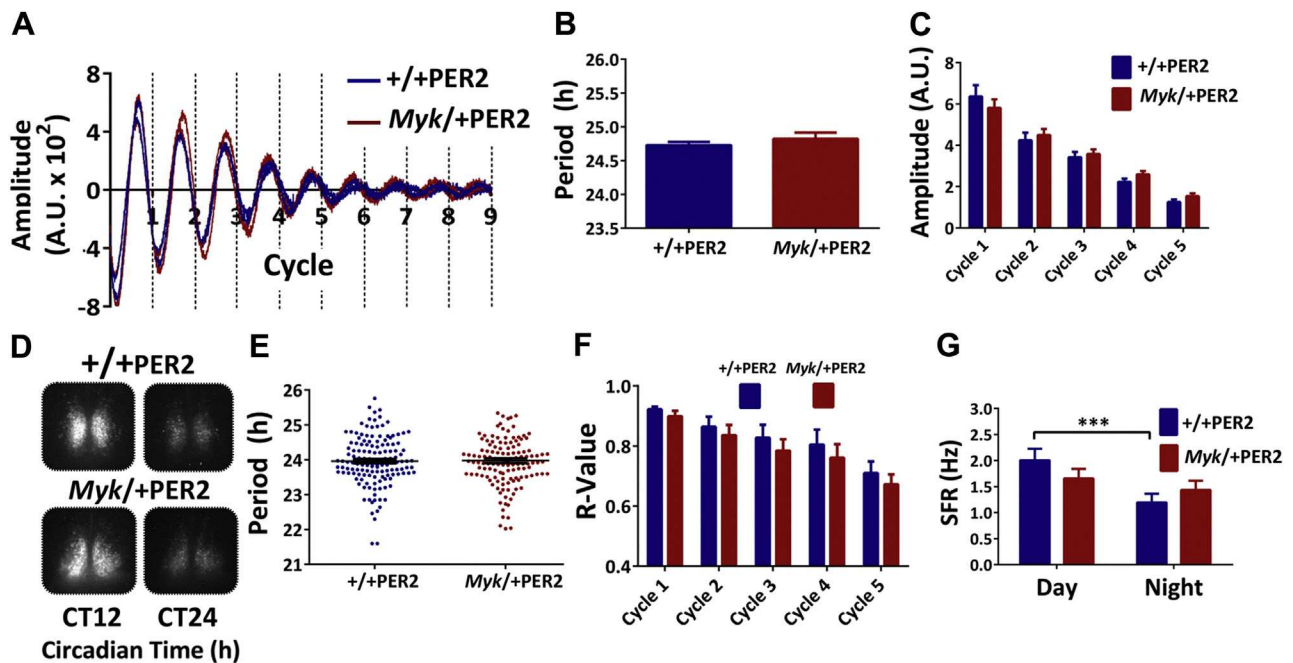


Figure 7. The *Myshkin* mutation does not affect bioluminescent rhythms of PER2::LUC in whole suprachiasmatic nuclei (SCN) brain slices or single cells, but rather damps electrophysiological activity. **(A)** Example luminometric recordings of rhythms of PER2::LUC output of whole SCN tissue explants from +/+PER2::LUC (+/+PER2) and *Myk*/+PER2::LUC (*Myk*/+PER2). **(B)** No genotype differences were detected in the period of PER2::LUC oscillations (+/+PER2: 24.73 ± 0.05 hours, *Myk*/+PER2: 24.82 ± 0.09 hours; $p = .39$). **(C)** Peak amplitude of PER2::LUC rhythms across the first 5 days (24-hour cycles) in culture did not differ between the genotypes (two-way analysis of variance: genotype $p = .78$; interaction $p = .13$). **(D)** Example of single-cell imaging from +/+PER2 ($n = 4$) and *Myk*/+PER2 ($n = 4$) explants at two time points over initial 24 hours ex vivo. **(E)** The period of single-cell rhythms did not differ between the genotypes (+/+PER2: $n = 140$, 23.96 ± 0.06 hours; *Myk*/+PER2: $n = 132$, 23.96 ± 0.07 hours; $p = .99$). **(F)** The synchrony (R) between single cells in the SCN slices did not differ between the genotypes at any of the 5 days (5 × 24-hour cycles) ex vivo (two-way analysis of variance: genotype $p = .70$). **(G)** Mean spontaneous firing rate (SFR) from whole-cell current-clamp recordings made over a 24-hour cycle. Day/night variation in SFR is seen across +/+ SCN neurons but not *Myk*/+ SCN neurons (two-way analysis of variance: time $p = .0079$; +/+ day: 2.0 ± 0.2 Hz, +/+ night: 1.2 ± 0.2 Hz, $p = .0009$; *Myk*/+ day: 1.6 ± 0.2 Hz, *Myk*/+ night: 1.4 ± 0.2 Hz; $p = .31$). Data in panel **(E)** graphed as scatter plot with mean ± SEM. Data in panels **(B, C, F, G)** plotted as mean ± SEM. *** $p < .001$. A.U., arbitrary units; CT, circadian time.

concordant with the observation that the molecular clock is also intact in fibroblasts from patients with BPD (63). However, *Myk*/+ SCN slices lacked day/night variation in neuronal firing rate. Importantly, the low amplitude, long period behavioral rhythms, and damped SCN electrical activity of *Myk*/+ mice resemble similar measures made from +/+ mice exposed to long day lengths or LL (44,64–66). These findings reveal an important role for light inputs to the SCN in this mouse model of mania.

NKA $\alpha 3$ is localized to central and peripheral neurons, including retinal ganglion cells (46,67), and glutamate is the main neurotransmitter of image- and non-image-forming pathways from the eye (12,49). Both SCN and extra-SCN sites are implicated in masking effects of light (68,69) and the absence of negative masking in *Myk*/+ mice presumably arises as a consequence of altered glutamatergic signaling at these sites. Glutamatergic synapses are linked with mood disorders (70), and it is suggested that measures of glutamate in the brain vary in BPD, becoming elevated during mania and reduced in episodes of depression (71). Further, mood stabilizers used to treat BPD, such as lithium and valproate, act to restore glutamate levels (72,73) and can reduce sensitivity to light and directly alter SCN function (74,75). Interestingly, colocalization and functional coupling between NKA $\alpha 3$ and

glutamate transporters has been demonstrated in rat brain, with $\alpha 3$ having similar neuronal localization to excitatory amino acid transporter 2/glutamate transporter 1, the most abundant subtype of glutamate transporter in the central nervous system (76). Previously, Kirshenbaum *et al.* (20) found that the duration of glutamate-evoked $[Ca^{2+}]_i$ transients was prolonged in cultured cortical neurons from *Myk*/+ mice (20), raising the possibility that this mutation alters glutamatergic signaling in other brain pathways. Because metabolic rate is elevated in *Myk*/+ mice, it is plausible that the *Myshkin* mutation influences energy balance centers in the mediobasal hypothalamus (77).

In high-firing hippocampal and cerebellar neurons, loss or reduction of NKA $\alpha 3$ is associated with neuronal hyperexcitability (20,51,78,79). *Atp1a3* is expressed in the SCN (80), but *Myk*/+ SCN neurons that have a reduction in functional NKA $\alpha 3$ show damped daytime firing rate. To discharge action potentials, high-firing cells can require considerable adenosine triphosphate, and a likely consequence of a reduction in NKA pump activity is ionic imbalance and chronic depolarization (81). Because individual SCN neurons are comparatively low firing (typically <5 Hz), this suggests that their adenosine triphosphate requirements are low, such that a reduction in NKA $\alpha 3$ activity does not drive the cell into a chronic

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depolarized state. Indeed, because some classes of intrinsically photosensitive retinal ganglion cells can spontaneously fire at high frequencies (70–90 Hz) (82), it is probable that they are more readily hyperexcited through the reduction in NKA $\alpha 3$ activity, as is evidenced by the associated changes in retinal input to the SCN in *Myk/+* animals.

NKA α subunits ($\alpha 1$ – $\alpha 3$) are associated with BPD, although subunit-specific roles in pathology remain to be defined (20,83,84). A heterozygous missense mutation (V129M) in NKA $\alpha 3$ was identified in a 9-year-old boy with DSM-5 schizophrenia and a clinical presentation including mood swings (85). Multiple *Atp1a3*-specific mutant mouse models exhibit variable presentations of neurological deficits (86,87), but one key commonality is that all show heightened psychomotor states. Missense mutations in NKA $\alpha 3$ have been identified in other neurological conditions including rapid-onset dystonia parkinsonism, alternating hemiplegia of childhood (AHC), and CAPOS (cerebellar ataxia, areflexia, pes cavus, optic atrophy, and sensorineural hearing loss) syndrome (88,89). Patients with rapid-onset dystonia parkinsonism show greater incidence of mood and psychotic symptoms than control family members (90). AHC patients are easily aroused and prone to behavioral and psychiatric symptoms, such as impulsivity, lack of attention control, and episodes of hyperactivity (H. Rosewich, M.D., personal communication, July 6, 2016). Clinical presentation of AHC is sensitive to changes in lighting and sleep induction. Indeed, acute presentation of AHC symptoms such as hemiplegic attacks can be triggered by stimulation of the optic system (H. Rosewich, personal communication) and attenuated through occlusion of the eyes (91). In addition to circadian-associated behaviors, therefore, our findings support a wider role of light and retinal signaling in the pathology of *ATP1A3*-related disorders. As the activity of NKA $\alpha 3$ was recently found to be impaired by its aberrant association with misfolded and aggregated amyloid- β in Alzheimer's disease (92) and α -synuclein in Parkinson's disease (93), NKA $\alpha 3$ dysfunction may also contribute to the circadian sleep disturbances in these common age-related neurodegenerative conditions (94–96).

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JWST, NK, HRS, TA-M, and AT LH conducted the behavioral and metabolic assessments of the mice. JWST made the electrophysiological and intracellular calcium recordings. JWST, NK, and VB performed the bioluminescence recordings. GSK and MRR conducted the optic nerve crush experiment. MDCB advised on the design of the electrophysiology and calcium imaging experiments. SJC supplied the *Myshkin* mouse line, advised on experimental design and edited the manuscript. JWST, SJC, and HDP wrote the manuscript.

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ARTICLE INFORMATION

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REFERENCES

1. Wittchen HU, Jacobi F, Rehm J, Gustavsson A, Svensson M, Jonsson B, et al. (2011): The size and burden of mental disorders and other disorders of the brain in Europe 2010. *Eur Neuropharmacol* 21:655–679.
2. Phillips ML, Kupfer DJ (2013): Bipolar disorder diagnosis: challenges and future directions. *Lancet* 381:1663–1671.
3. Wulff K, Gatti S, Wettstein JG, Foster RG (2010): Sleep and circadian rhythm disruption in psychiatric and neurodegenerative disease. *Nat Rev Neurosci* 11:589–599.
4. Wu JC, Kelsoe JR, Schachar C, Bunney BG, DeModena A, Golshan S, et al. (2009): Rapid and sustained antidepressant response with sleep deprivation and chronotherapy in bipolar disorder. *Biol Psychiatry* 66:298–301.
5. Dallaspesza S, Suzuki M, Benedetti F (2015): Chronobiological therapy for mood disorders. *Curr Psychiatry Rep* 17:95.
6. Etain B, Milhiet V, Bellivier F, Leboyer M (2011): Genetics of circadian rhythms and mood spectrum disorders. *Eur Neuropsychopharmacol* 21(suppl 4):S676–S682.
7. Welsh DK, Takahashi JS, Kay SA (2010): Suprachiasmatic nucleus: cell autonomy and network properties. *Annu Rev Physiol* 72:551–577.
8. Partch CL, Green CB, Takahashi JS (2014): Molecular architecture of the mammalian circadian clock. *Trends Cell Biol* 24:90–99.
9. Belle MD, Diekmann CO, Forger DB, Piggins HD (2009): Daily electrical silencing in the mammalian circadian clock. *Science* 326:281–284.
10. Colwell CS (2011): Linking neural activity and molecular oscillations in the SCN. *Nat Rev Neurosci* 12:553–569.
11. Roenneberg T, Allebrandt KV, Meroz M, Vetter C (2012): Social jetlag and obesity. *Curr Biol* 22:939–943.
12. Lucas RJ, Peirson SN, Berson DM, Brown TM, Cooper HM, Czeisler CA, et al. (2014): Measuring and using light in the melanopsin age. *Trends Neurosci* 37:1–9.
13. Castel M, Belenky M, Cohen S, Ottersen OP, Storm-Mathisen J (1993): Glutamate-like immunoreactivity in retinal terminals of the mouse suprachiasmatic nucleus. *Eur J Neurosci* 5:368–381.
14. Golombek DA, Rosenstein RE (2010): Physiology of circadian entrainment. *Physiol Rev* 90:1063–1102.
15. LeGates TA, Fernandez DC, Hattar S (2014): Light as a central modulator of circadian rhythms, sleep and affect. *Nat Rev Neurosci* 15:443–454.
16. Landgraf D, McCarthy MJ, Welsh DK (2014): Circadian clock and stress interactions in the molecular biology of psychiatric disorders. *Curr Psychiatry Rep* 16:483.
17. Roybal K, Theobald D, Graham A, DiNieri JA, Russo SJ, Krishnan V, et al. (2007): Mania-like behavior induced by disruption of CLOCK. *Proc Natl Acad Sci U S A* 104:6406–6411.

18. Sidor MM, Spencer SM, Dziras K, Parekh PK, Tye KM, Warden MR, *et al.* (2015): Daytime spikes in dopaminergic activity drive rapid mood-cycling in mice. *Mol Psychiatry* 20:1406–1419.
19. Chung S, Lee EJ, Yun S, Choe HK, Park SB, Son HJ, *et al.* (2014): Impact of circadian nuclear receptor REV-ERB α on midbrain dopamine production and mood regulation. *Cell* 157:858–868.
20. Kirshenbaum GS, Clapcote SJ, Duffy S, Burgess CR, Petersen J, Jarowek KJ, *et al.* (2011): Mania-like behavior induced by genetic dysfunction of the neuron-specific Na⁺, K⁺-ATPase α 3 sodium pump. *Proc Natl Acad Sci U S A* 108:18144–18149.
21. Logan RW, McClung CA (2016): Animal models of bipolar mania: The past, present and future. *Neuroscience* 321:163–188.
22. Yoo SH, Yamazaki S, Lowrey PL, Shimomura K, Ko CH, Buhr ED, *et al.* (2004): PERIOD2::LUCIFERASE real-time reporting of circadian dynamics reveals persistent circadian oscillations in mouse peripheral tissues. *Proc Natl Acad Sci U S A* 101:5339–5346.
23. Bechtold DA, Brown TM, Luckman SM, Piggins HD (2008): Metabolic rhythm abnormalities in mice lacking VIP-VPAC2 signaling. *Am J Physiol Regul Integr Comp Physiol* 294:R344–R351.
24. Marston OJ, Williams RH, Canal MM, Samuels RE, Upton N, Piggins HD (2008): Circadian and dark-pulse activation of orexin/hypocretin neurons. *Mol Brain* 1:19.
25. Hughes AT, Piggins HD (2008): Behavioral responses of *Vipr2*^{-/-} mice to light. *J Biol Rhythms* 23:211–219.
26. Belle MD, Hughes AT, Bechtold DA, Cunningham P, Pierucci M, Burdakov D, *et al.* (2014): Acute suppressive and long-term phase modulation actions of orexin on the mammalian circadian clock. *J Neurosci* 34:3607–3621.
27. Guilding C, Hughes AT, Brown TM, Namvar S, Piggins HD (2009): A riot of rhythms: neuronal and glial circadian oscillators in the mediobasal hypothalamus. *Mol Brain* 2:28.
28. Guilding C, Scott F, Bechtold DA, Brown TM, Wegner S, Piggins HD (2013): Suppressed cellular oscillations in after-hours mutant mice are associated with enhanced circadian phase-resetting. *J Physiol* 591:1063–1080.
29. Geoffroy PA, Bellivier F, Scott J, Etain B (2014): Seasonality and bipolar disorder: a systematic review, from admission rates to seasonality of symptoms. *J Affect Disord* 168:210–223.
30. Wang B, Chen D (2013): Evidence for seasonal mania: a review. *J Psychiatr Pract* 19:301–308.
31. Asher G, Schibler U (2011): Crosstalk between components of circadian and metabolic cycles in mammals. *Cell Metab* 13:125–137.
32. Sestan-Pesa M, Horvath TL (2016): Metabolism and mental illness. *Trends Mol Med* 22:174–183.
33. Turek FW, Joshu C, Kohsaka A, Lin E, Ivanova G, McDearmon E, *et al.* (2005): Obesity and metabolic syndrome in circadian Clock mutant mice. *Science* 308:1043–1045.
34. Caliyurt O, Altıay G (2009): Resting energy expenditure in manic episode. *Bipolar Disord* 11:102–106.
35. Li X, Gilbert J, Davis FC (2005): Disruption of masking by hypothalamic lesions in Syrian hamsters. *J Comp Physiol A Neuroethol Sens Neural Behav Physiol* 191:23–30.
36. Pendergast JS, Yamazaki S (2011): Masking responses to light in period mutant mice. *Chronobiol Int* 28:657–663.
37. Goh J, Ladiges W (2015): Voluntary wheel running in mice. *Curr Protoc Mouse Biol* 5:283–290.
38. Kirshenbaum GS, Burgess CR, Dery N, Fahnestock M, Peever JH, Roder JC (2014): Attenuation of mania-like behavior in Na(+), K(+)-ATPase α 3 mutant mice by prospective therapies for bipolar disorder: melatonin and exercise. *Neuroscience* 260:195–204.
39. Jud C, Schmutz I, Hampp G, Oster H, Albrecht U (2005): A guideline for analyzing circadian wheel-running behavior in rodents under different lighting conditions. *Biol Proced Online* 7:101–116.
40. Inder ML, Crowe MT, Porter R (2016): Effect of transmeridian travel and jetlag on mood disorders: Evidence and implications. *Aust N Z J Psychiatry* 50:220–227.
41. Frank E, Swartz HA, Kupfer DJ (2000): Interpersonal and social rhythm therapy: Managing the chaos of bipolar disorder. *Biol Psychiatry* 48:593–604.
42. Jagannath A, Butler R, Godinho SI, Couch Y, Brown LA, Vasudevan SR, *et al.* (2013): The CRTCL1-SIK1 pathway regulates entrainment of the circadian clock. *Cell* 154:1100–1111.
43. Yamaguchi Y, Suzuki T, Mizoro Y, Kori H, Okada K, Chen Y, *et al.* (2013): Mice genetically deficient in vasopressin V1a and V1b receptors are resistant to jet lag. *Science* 342:85–90.
44. Hughes AT, Croft CL, Samuels RE, Myung J, Takumi T, Piggins HD (2015): Constant light enhances synchrony among circadian clock cells and promotes behavioral rhythms in VPAC2-signaling deficient mice. *Sci Rep* 5:14044.
45. Specht SC, Sweadner KJ (1984): Two different Na, K-ATPases in the optic nerve: Cells of origin and axonal transport. *Proc Natl Acad Sci U S A* 81:1234–1238.
46. Wetzel RK, Arystarkhova E, Sweadner KJ (1999): Cellular and sub-cellular specification of Na, K-ATPase α and β isoforms in the postnatal development of mouse retina. *J Neurosci* 19:9878–9889.
47. Michel S, Itri J, Colwell CS (2002): Excitatory mechanisms in the suprachiasmatic nucleus: The role of AMPA/KA glutamate receptors. *J Neurophysiol* 88:817–828.
48. Mizoro Y, Yamaguchi Y, Kitazawa R, Yamada H, Matsuo M, Fustin JM, *et al.* (2010): Activation of AMPA receptors in the suprachiasmatic nucleus phase-shifts the mouse circadian clock in vivo and in vitro. *PLoS One* 5:e10951.
49. Brown TM, Piggins HD (2007): Electrophysiology of the suprachiasmatic circadian clock. *Prog Neurobiol* 82:229–255.
50. Wang YC, Huang RC (2006): Effects of sodium pump activity on spontaneous firing in neurons of the rat suprachiasmatic nucleus. *J Neurophysiol* 96:109–118.
51. Dobretsov M, Stimers JR (2005): Neuronal function and α 3 isoform of the Na/K-ATPase. *Front Biosci* 10:2373–2396.
52. Azarias G, Kruusmagi M, Connor S, Akkuratov EE, Liu XL, Lyons D, *et al.* (2013): A specific and essential role for Na, K-ATPase α 3 in neurons co-expressing α 1 and α 3. *J Biol Chem* 288:2734–2743.
53. Pennartz CM, De Jeu MT, Geurtsen AM, Sluiter AA, Hermes ML (1998): Electrophysiological and morphological heterogeneity of neurons in slices of rat suprachiasmatic nucleus. *J Physiol* 506:775–793.
54. Landgraf D, Long JE, Proulx CD, Barandas R, Malinow R, Welsh DK (2016): Genetic disruption of circadian rhythms in the suprachiasmatic nucleus causes helplessness, behavioral despair, and anxiety-like behavior in mice. *Biol Psychiatry* 80:827–835.
55. Vitaterna MH, Ko CH, Chang AM, Buhr ED, Fruechte EM, Schook A, *et al.* (2006): The mouse Clock mutation reduces circadian pacemaker amplitude and enhances efficacy of resetting stimuli and phase-response curve amplitude. *Proc Natl Acad Sci U S A* 103:9327–9332.
56. Preitner N, Damiola F, Lopez-Molina L, Zakany J, Duboule D, Albrecht U, *et al.* (2002): The orphan nuclear receptor REV-ERB α controls circadian transcription within the positive limb of the mammalian circadian oscillator. *Cell* 110:251–260.
57. Jones SH, Hare DJ, Evershed K (2005): Actigraphic assessment of circadian activity and sleep patterns in bipolar disorder. *Bipolar Disord* 7:176–186.
58. Dallaspezia S, Benedetti F (2015): Chronobiology of bipolar disorder: Therapeutic implication. *Curr Psychiatry Rep* 17:606.
59. Henriksen TE, Skrede S, Fasmer OB, Hamre B, Gronli J, Lund A (2014): Blocking blue light during mania - markedly increased regularity of sleep and rapid improvement of symptoms: A case report. *Bipolar Disord* 16:894–898.
60. von Knorring L (1978): Visual averaged evoked responses in patients with bipolar affective disorders. *Neuropsychobiology* 4:314–320.
61. Soreca I (2014): Circadian rhythms and sleep in bipolar disorder: Implications for pathophysiology and treatment. *Curr Opin Psychiatry* 27:467–471.
62. Bradley AJ, Webb-Mitchell R, Hazu A, Slater N, Middleton B, Gallagher P, *et al.* (2017): Sleep and circadian rhythm disturbance in bipolar disorder. *Psychol Med* 1–12.
63. Yang S, Van Dongen HP, Wang K, Berrettini W, Bucan M (2009): Assessment of circadian function in fibroblasts of patients with bipolar disorder. *Mol Psychiatry* 14:143–155.

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64. VanderLeest HT, Houben T, Michel S, Deboer T, Albus H, Vansteensel MJ, *et al.* (2007): Seasonal encoding by the circadian pacemaker of the SCN. *Curr Biol* 17:468–473.
65. Ohta H, Yamazaki S, McMahon DG (2005): Constant light desynchronizes mammalian clock neurons. *Nat Neurosci* 8:267–269.
66. Brown TM, Piggins HD (2009): Spatiotemporal heterogeneity in the electrical activity of suprachiasmatic nuclei neurons and their response to photoperiod. *J Biol Rhythms* 24:44–54.
67. Bottger P, Tracz Z, Heuck A, Nissen P, Romero-Ramos M, Lykke-Hartmann K (2011): Distribution of Na/K-ATPase alpha 3 isoform, a sodium-potassium P-type pump associated with rapid-onset of dystonia parkinsonism (RDP) in the adult mouse brain. *J Comp Neurol* 519:376–404.
68. Thompson S, Foster RG, Stone EM, Sheffield VC, Mrosovsky N (2008): Classical and melanopsin photoreception in irradiance detection: Negative masking of locomotor activity by light. *Eur J Neurosci* 27:1973–1979.
69. Doyle SE, Yoshikawa T, Hillson H, Menaker M (2008): Retinal pathways influence temporal niche. *Proc Natl Acad Sci U S A* 105:13133–13138.
70. de Sousa RT, Loch AA, Carvalho AF, Brunoni AR, Haddad MR, Henter ID, *et al.* (2017): Genetic studies on the tripartite glutamate synapse in the pathophysiology and therapeutics of mood disorders. *Neuropsychopharmacology* 42:787–800.
71. Dickerson F, Stallings C, Vaughan C, Origoni A, Khushalani S, Yolken R (2012): Antibodies to the glutamate receptor in mania. *Bipolar Disord* 14:547–553.
72. Friedman SD, Dager SR, Parow A, Hirashima F, Demopoulos C, Stoll AL, *et al.* (2004): Lithium and valproic acid treatment effects on brain chemistry in bipolar disorder. *Biol Psychiatry* 56:340–348.
73. Strawn JR, Patel NC, Chu WJ, Lee JH, Adler CM, Kim MJ, *et al.* (2012): Glutamatergic effects of divalproex in adolescents with mania: A proton magnetic resonance spectroscopy study. *J Am Acad Child Adolesc Psychiatry* 51:642–651.
74. Hallam KT, Olver JS, Norman TR (2005): Effect of sodium valproate on nocturnal melatonin sensitivity to light in healthy volunteers. *Neuropsychopharmacology* 30:1400–1404.
75. Seggie J, Carney PA, Parker J, Grof E, Grof P (1989): Effect of chronic lithium on sensitivity to light in male and female bipolar patients. *Prog Neuropsychopharmacol Biol Psychiatry* 13:543–549.
76. Rose EM, Koo JC, Antflick JE, Ahmed SM, Angers S, Hampson DR (2009): Glutamate transporter coupling to Na, K-ATPase. *J Neurosci* 29:8143–8155.
77. Sohn JW (2015): Network of hypothalamic neurons that control appetite. *BMB Rep* 48:229–233.
78. Vaillend C, Mason SE, Cuttle MF, Alger BE (2002): Mechanisms of neuronal hyperexcitability caused by partial inhibition of Na⁺/K⁺-ATPases in the rat CA1 hippocampal region. *J Neurophysiol* 88:2963–2978.
79. Fremont R, Calderon DP, Maleki S, Khodakhah K (2014): Abnormal high-frequency burst firing of cerebellar neurons in rapid-onset dystonia-parkinsonism. *J Neurosci* 34:11723–11732.
80. Lein ES, Hawrylycz MJ, Ao N, Ayres M, Bensinger A, Bernard A, *et al.* (2007): Genome-wide atlas of gene expression in the adult mouse brain. *Nature* 445:168–176.
81. Le Masson G, Przedborski S, Abbott LF (2014): A computational model of motor neuron degeneration. *Neuron* 83:975–988.
82. Zhao X, Stafford BK, Godin AL, King WM, Wong KY (2014): Photo-response diversity among the five types of intrinsically photosensitive retinal ganglion cells. *J Physiol* 592:1619–1636.
83. el-Mallakh RS, Wyatt RJ (1995): The Na, K-ATPase hypothesis for bipolar illness. *Biol Psychiatry* 37:235–244.
84. Goldstein I, Lerer E, Laiba E, Mallet J, Muijaheed M, Laurent C, *et al.* (2009): Association between sodium- and potassium-activated adenosine triphosphatase alpha isoforms and bipolar disorders. *Biol Psychiatry* 65:985–991.
85. Smedemark-Margulies N, Brownstein CA, Vargas S, Tembulkar SK, Towne MC, Shi J, *et al.* (2016): A novel de novo mutation in ATP1A3 and childhood-onset schizophrenia. *Cold Spring Harb Mol Case Stud* 2:a001008.
86. Hunanyan AS, Fainberg NA, Linabarger M, Arehart E, Leonard AS, Adil SM, *et al.* (2015): Knock-in mouse model of alternating hemiplegia of childhood: Behavioral and electrophysiologic characterization. *Epilepsia* 56:82–93.
87. Holm TH, Isaksen TJ, Glerup S, Heuck A, Bottger P, Fuchtbauer EM, *et al.* (2016): Cognitive deficits caused by a disease-mutation in the alpha3 Na⁺/K⁺-ATPase isoform. *Sci Rep* 6:31972.
88. Heinzen EL, Arzimanoglou A, Brashear A, Clapcote SJ, Gurrieri F, Goldstein DB, *et al.* (2014): Distinct neurological disorders with ATP1A3 mutations. *Lancet Neurol* 13:503–514.
89. Maas RP, Schieving JH, Schouten M, Kamsteeg EJ, van de Warrenburg BP (2016): The genetic homogeneity of CAPOS syndrome: Four new patients with the c.2452G>A (p.Glu818Lys) mutation in the ATP1A3 gene. *Pediatr Neurol* 59:71–75.e71.
90. Brashear A, Cook JF, Hill DF, Amponsah A, Snively BM, Light L, *et al.* (2012): Psychiatric disorders in rapid-onset dystonia-parkinsonism. *Neurology* 79:1168–1173.
91. Rosewich H, Baethmann M, Ohlenbusch A, Gartner J, Brockmann K (2014): A novel ATP1A3 mutation with unique clinical presentation. *J Neurol Sci* 341:133–135.
92. Ohnishi T, Yanazawa M, Sasahara T, Kitamura Y, Hiroaki H, Fukazawa Y, *et al.* (2015): Na, K-ATPase alpha3 is a death target of Alzheimer patient amyloid-beta assembly. *Proc Natl Acad Sci U S A* 112:E4465–E4474.
93. Shrivastava AN, Redeker V, Fritz N, Pieri L, Almeida LG, Spolidoro M, *et al.* (2015): alpha-synuclein assemblies sequester neuronal alpha3-Na⁺/K⁺-ATPase and impair Na⁺ gradient. *EMBO J* 34:2408–2423.
94. Mattis J, Sehgal A (2016): Circadian rhythms, sleep, and disorders of aging. *Trends Endocrinol Metab* 27:192–203.
95. Musiek ES (2015): Circadian clock disruption in neurodegenerative diseases: Cause and effect? *Front Pharmacol* 6:29.
96. Videnovic A, Golombek D (2013): Circadian and sleep disorders in Parkinson's disease. *Exp Neurol* 243:45–56.