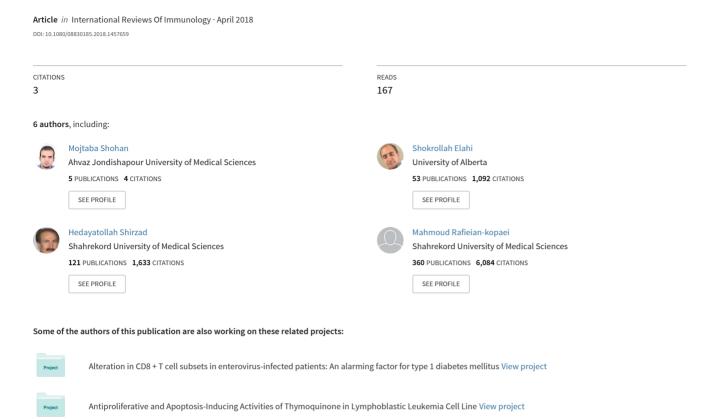
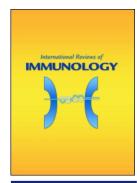
### Th9 Cells: Probable players in ulcerative colitis pathogenesis





### **International Reviews of Immunology**



ISSN: 0883-0185 (Print) 1563-5244 (Online) Journal homepage: http://www.tandfonline.com/loi/iiri20

# Th9 Cells: Probable players in ulcerative colitis pathogenesis

Mojtaba Shohan, Shokrollah Elahi, Hedayatollah Shirzad, Mahmoud Rafieian-Kopaei, Nader Bagheri & Emad Soltani

To cite this article: Mojtaba Shohan, Shokrollah Elahi, Hedayatollah Shirzad, Mahmoud Rafieian-Kopaei, Nader Bagheri & Emad Soltani (2018): Th9 Cells: Probable players in ulcerative colitis pathogenesis, International Reviews of Immunology, DOI: 10.1080/08830185.2018.1457659

To link to this article: <a href="https://doi.org/10.1080/08830185.2018.1457659">https://doi.org/10.1080/08830185.2018.1457659</a>

	Published online: 19 Apr 2018.
	Submit your article to this journal 🗗
Q <sup>L</sup>	View related articles 🗹
CrossMark	View Crossmark data 🗗





### Th9 Cells: Probable players in ulcerative colitis pathogenesis

Mojtaba Shohan<sup>a</sup>, Shokrollah Elahi<sup>b</sup>, Hedayatollah Shirzad<sup>c</sup>, Mahmoud Rafieian-Kopaei <sup>od</sup>, Nader Bagheri<sup>a</sup>, and Emad Soltani<sup>a</sup>

<sup>a</sup>Department of Microbiology and Immunology, Faculty of Medicine, Shahrekord University of Medical Sciences, Shahrekord, Iran; <sup>b</sup>Department of Dentistry, Department of Medical Microbiology and Immunology, Faculty of Medicine and Dentistry, University of Alberta, Edmonton, Alberta, Canada; <sup>c</sup>Cellular and Molecular Research Center, Basic Health Sciences Institute, Shahrekord University of Medical Sciences, Shahrekord, Iran; <sup>d</sup>Medical Plants Research Center, Basic Health Sciences Institute, Shahrekord University of Medical Sciences, Shahrekord, Iran

#### **ABSTRACT**

T lymphocytes represent an important part of adaptive immune system undertaking different functions to regulate immune responses. CD4+ T cells are the most important activator cells in inflammatory conditions. Depending on the type of induced cells and inflamed sites, expression and activity of different subtypes of helper T cells are changed. Recent studies have confirmed the existence of a new subset of helper T lymphocytes called Th9. Naive T cells can differentiate into Th9 subtypes if they are exposed simultaneously by interleukin (IL) 4 and transforming growth factor  $\beta$ and also secondary activation of a complicated network of transcription factors such as interferon regulatory factor 4 (IRF4) and Smads which are essential for adequate induction of this phenotype. Th9 cells specifically produce interleukin 9 and their probable roles in promoting intestinal inflammation are being investigated in human subjects and experimental models of ulcerative colitis (UC). Recently, infiltration of Th9 cells, overexpression of IL-9, and certain genes associated with Th9 differentiation have been demonstrated in inflammatory microenvironment of UC. Intestinal oversecretion of IL-9 protein is likely to break down epithelial barriers and compromise tolerance to certain commensal microorganisms which leads to inflammation. Th9 pathogenicity has not yet been adequately explored in UC and they are far from being considered as inflammatory cells in this milieu, therefore precise understanding the role of these newly identified cells in particular their potential role in gut pathogenesis may enable us to develop novel therapeutic approaches for inflammatory bowel disease. So, this article tries to discuss the latest knowledge on the above-mentioned field.

#### **ARTICLE HISTORY**

Received 12 September 2017 Accepted 22 March 2018

#### KEYWORDS

Intestinal inflammation; IL-9; Th9; ulcerative colitis

#### **Abbreviations**

CMC Chronic mucocutaneous candidiasis DSS Dextran sodium sulfate Experimental autoimmune encephalomyelitis EAE FOXP3 Forkhead box P3 **GATA** Family of transcription factors bind to "GATA" sequences IBDs Inflammatory bowel diseases **IELs** Intraepithelial lymphocytes ILInterleukin ILCs Innate lymphoid cells IRF4 Interferon regulatory factor 4 Janus kinase IAK LPS Lipopolysaccharide NF-κB Nuclear factor kappa B NKT Natural killer cell NOS2 Nitric oxide synthase 2

	member 4
PU1	An ETS family transcription factor
ROS	Reactive oxygen species
RSV	Respiratory syncytial virus
STAT	Signal transducer and activator of transcription
TCR	T cell receptor
TGF- $\beta$	Transforming growth factor beta
TLR	Toll-like receptor
TNBS	Trinitrobenzenesulfonic acid
Tregs	Regulatory T cells
TSLP	Thymic stromal lymphopoietin
UC	Ulcerative colitis

Tumor necrosis factor receptor superfamily

#### 1. Introduction

OX40

Effective immunologic responses are strongly dependent on the accurate induction of T cells. By expressing CD40 ligand, they provide required signals for clonal

expansion of B cells, immunoglobulin switching, and antibody production. They also regulate innate immunity through affecting macrophages and neutrophils, create memory T cell response, and manage inflammatory responses to pathogens.<sup>2</sup> However, uncontrolled T cell response can cause pathological alterations which may leads to allergy and autoimmunity.<sup>3</sup> A naive T cell in the presence of interleukin (IL)-4 and transforming growth factor (TGF)- $\beta$  secretes IL-9, which drives cellular responses toward the differentiation of a novel characterized type of CD4+ cells named Th9.4 The existence of Th9 cells and their roles as novel players in gut immunity have been confirmed in a number of studies. For example, compromised tissue integrity and sustained inflammatory responses during flare of ulcerative colitis (UC) in colon are related to IL-9 production by Th9 cells.<sup>5,6</sup> Tight junction proteins as principal components of intercellular adhesion mechanisms are important molecules, which maintain intestinal tissue integrity and prevent luminal antigens from being exposed to inflammatory cells present in lamina propria.<sup>7</sup> Recent studies have also shown some alterations in expression of these proteins by Th9-derived IL-9 that result in further gut inflammation. This article tries to discuss the harmful roles of Th9 cells in the gut.

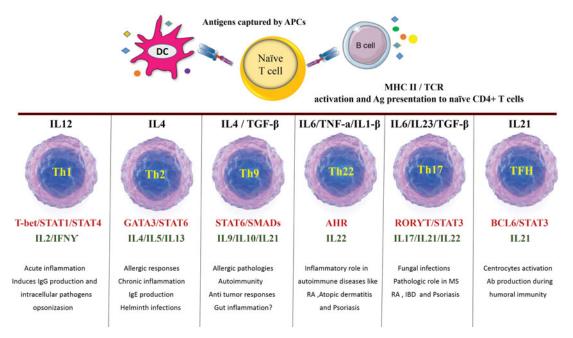
## 2. Various CD4+ cells are distinctively differentiated

Multiple immune responses of the host subsequent to pathogen's crossing the innate immunity barriers are dependent on the appropriate function of T lymphocytes. According to their identifying markers and activities, T cells are divided into three major classes as follows: CD8+ T lymphocytes that are cytotoxic or killer cells and are known to be involved in response to tumor, metastatic cells, and viral infections.8 Regulatory T cells (Tregs) that play a significant role in suppressing inflammatory responses, inducing immunological tolerance, and regulating immune responses to prevent autoimmunity through secretion of certain immunomodulatory cytokines such as IL-10, TGF- $\beta$ , and IL-35 or via cell-cell contact.9,10 CD4+ helper T cells that are major players in inflammatory milieu contributing to antibody production, regulating innate immunity, and inducing immunologic memory. Predominant types of these cell lineages including Th1, Th17, Th22, Th9, Th2, and TFH, are developed through T cell receptor (TCR) signaling and being affected by various cytokines and transcription factors (Figure 1).<sup>11–13</sup>

# 3. Th9 cells: Signaling pathways and cytokine profile

# 3.1. Characteristics and secretion sources of interleukin 9

IL-9 is a multifunctional cytokine and the most important source of IL-9 is Th9 lymphocytes. Therefore, they are most probably involved in the responses associated with increased IL-9 level during the process of inflammatory damages caused by immune system hyperactivity.<sup>22-24</sup> Although IL-9 is structurally different from lymphocytes growth factors, it was first identified and cloned as a growth factor of murine mast cells and T cells (T cell growth factor III) in 1980.<sup>23,25</sup> IL-9 is a cytokine with a common gamma-chain receptor similar to IL-4, -2, and -15, and upon binding to its receptor on target cells induces a signal through activation of janus kinase 1 (JAK1) and JAK3, creating a dimer between signal transducer and activator of transcription 3 (STAT3), STAT5, and STAT1.<sup>26-28</sup> The alpha subunit of IL-9 receptor contains WSXWS and BOX motifs and therefore is considered as a member of hematopoietin family.<sup>22</sup> The encoding gene of the protein of 144 amino acids is localized to chromosome 5 in human and chromosome 13 in mouse.<sup>29,30</sup> Th9 cells secrete abundant amounts of IL-9, but no specific and definite transcription factor has yet been defined for their complete differentiation.<sup>31</sup> First, because of increased release of IL-9 from Leishmaniainfected murine Th2 cells, it was thought that IL-9 was generated only by Th2 cells therefore, its role has been investigated mainly in Th2-related pathologies. 32 It seems that in different immunological conditions, various factors impact induction of IL-9.4 Via the simultaneous effects of IL-4 and TGF- $\beta$  on naive T cells, their downstream signaling proteins such as GATA3, interferon regulatory factor 4 (IRF4), STAT6, and PU1 are being activated and induce IL-9 promoter transcription.<sup>33</sup> PU1deficient mice exhibit potent inhibitory responses against the IL-9-dependent allergic airway inflammation. Such mice have a normal Th2 response despite having low amounts of IL-9. This indicates that PU1 play a critical role in Th9 development. 17 Evidence remains inconsistent on factors that physically interact to activate IL-9 gene. In some allergic responses such as lung inflammation and certain anaphylactic conditions, Th2 and Th9 cells are deeply proliferated.<sup>34–36</sup> Th17 cells that are induced in IL-6 and TGF- $\beta$  containing environment, can generate IL-9 under certain conditions, however, IL-23 which promotes induction of Th17 cells was reported to decrease IL-9 generation. This indicates that IL-9 expression by Th17 completely depends on types of signals in inflammatory environment and does not occur constantly.<sup>37,38</sup>



**Figure 1.** Differentiation pathways of CD4+ T cells. Antigen presenting cells such as dendritic cells and B lymphocytes uptake antigens from the environment and present to naïve T cells. Depending on the type of antigens, cytokines released in the environment and activation of transcription factors, various types of CD4+ cells are induced and interact with different cell types and drive inflammatory responses with respect to related proteins and chemokine receptors. <sup>13–21</sup>

In certain autoimmunities, such as experimental autoimmune encephalomyelitis (EAE) and type I diabetes, IL-9 is produced by Th17 phenotype.<sup>39</sup> During EAE, IL-9 plays two different functions. It suppresses EAE-induced inflammation by influencing Tregs function but intensifies inflammatory mediators through affecting mast cells and Th17 cells in an autocrine manner. 37,38,40-42 In type I diabetes, an autoimmune disease with Th17 dominancy, the role of IL-9 has not yet been explained.<sup>43</sup> Current evidence on secretion of IL-9 by Tregs are inconsistent. Although coexpression of forkhead box P3 (FOXP3) and IL-9 has been observed in murine T cells in vivo, it has not yet been determined whether FOXP3-expressing T cells can generate IL-9 or not.44 IL-9 is an activator of human mast cell progenitors. As crucial regulators of innate immunity, mast cells expresses pattern recognition receptors like Toll-like receptor (TLRs) to recognize pathogen-associated molecular patterns and also are capable of producing IL-9 protein in response to stem cell factor, IL-10 and IL-3.45,46 By expressing IL-9 receptors, they can be activated by IL-9 in an autocrine manner and also can pathologically release this protein during allergic diseases such as asthma and allergic rhinitis. 47,48 Mucosal mast cells are dominant secretors of IL-9 in experimental models of intestinal food allergy.<sup>45</sup> Although IL-9 is first known as a growth factor of mast cells that was named P40, they can generate IL-9 in response to lipopolysaccharide (LPS) and IL-1 through nuclear factor kappa B (NF- $\kappa$ B) activation.<sup>49</sup> In addition, there are some

evidences indicating that histamine and IL-1 $\beta$  induce IL-9 secretion from murine bone marrow-derived mast cells.<sup>50,51</sup> Interestingly, remarkable infiltration of both chymase+ and trypase+ mast cells were observed by immunohistochemical staining of chronic UC lesions and it is stated that mast cells are the major source of several inflammatory mediators which can increase intestinal permeability and inflammation during gutrelated pathologies. 52,53 There are reports that murine naïve natural killer T cells can also secrete IL-9 in the presence of IL-2. However, due to certain similarities in signaling pathways between IL-9 and IL-4, -5, and IL13, these proteins are secreted from these cells following IL-9 induction.<sup>54</sup> In natural killer cell (NKT)-CD1d deficient mouse, allergic airway inflammation leads to decreased production of IL-9 and decreased infiltration of mast cells in the lung, which confirms NKT's ability to produce IL-9 in vivo.<sup>55</sup> As a growth factor, IL-9 can cause NKTs transformation into nasal NKT cells lymphoma. In tissue sections of patients with nasal NKT cells lymphoma, the trafficking of IL-9 generating NKTs was reported.<sup>56</sup> Innate lymphoid cells are recently defined subtypes, which contribute to both innate and adaptive immune responses. They have diverse distribution in lymphoid and mucosal tissues and are involved in tissue remodeling and homeostasis. Type 2 innate lymphoid cells (ILCs) which are more characterized by Th2-like responses are mainly induced by effects of IL-25, IL-33, thymic stromal lymphopoietin (TSLP), IL-4, IL-7, and IL-9.57 ILC2s are also important regulators of epithelial responses in acute lung allergy which their effective induction in such condition is mainly dependent on IRF4 and IL-9.58 During Th2-mediated intestinal inflammation, IL-33 is produced by damaged epithelial cells which affects ILC2s to release more IL-9 and finally more eosinophil recruitment, mucus secretion, and mastocytosis in response to IL-5 and IL-13, respectively.<sup>59</sup> We have given a snapshot of different sources of IL-9. Obviously, this protein plays significant roles in different diseases via its widely various functions. Now, we touch on cytokines, genes, and signaling pathways involved in Th9 development.

#### 3.2. Differentiation pathways of Th9 cells

IL-4 and TGF- $\beta$  activate two distinct pathways in T cells. The former causes transcription of GATA3 and STAT6 genes that leads to cell response toward Th2, and the latter activates FOXP3 and induces Tregs differentiation.<sup>60,61</sup> The simultaneous effects of these two cytokines on T cells lead to generation of IL-9 and induction of Th9 phenotype.<sup>62</sup> It has not yet been definitely determined which transcription factors physically interact and bind to IL-9 promoter. Different studies have reported inconsistent findings on this issue that will be distinctively quoted. Multiple proteins are considered as IL-4 and TGF- $\beta$  downstream signals that contribute to amplification of IL-9 promoter as a complicate network $^{63-65}$  (Figure 2).

#### 3.2.1. Positive regulators

As shown in Figure 2, all of these proteins contribute somehow to the induction of Th9 cells. Here, they will be discussed individually. IRF4 is the first factor of interest whose key role has been demonstrated in increasing

transcription of IL-9 gene in T lymphocytes. Phosphorylation of STAT6 occurs following IL-4's binding to its receptor, and then IRF4 is activated and binds to IL-9 gene. 66,67 However, appropriate induction of other cells such as Th17 and Th2 depends on the presence of this factor.<sup>68,69</sup> IRF4-deficient mouse cannot induce IL-9-secreting T cells, and also silencing of IRF4 mRNA by relevant SiRNA prevents development of Th9 phenotype and activation of IL-9 promoter.<sup>70</sup> GATA3 is the other factor that is activated by STAT6 and causes destruction of T-Bet and FOXP3. It has not yet been determined whether GATA3 binds to the IL-9 gene directly or not.<sup>30</sup> After TGF- $\beta$  binding to its receptor, a factor called PU1 activates IL-9 promoter through binding to IRF4.71 PU1 is an ETS family transcription factor whose increased expression has been reported in certain Th9-dependent pathologies such as allergic inflammation and inflammatory bowel disease (IBDs).<sup>17,72</sup> Although the signaling pathways of Th9 and Th2 cells are partly similar, increased expression of PU1 causes downregulation in Th2 response. This indicates that differentiation of these two subsets may be completely separated under different conditions.<sup>73</sup> Effects of PU1 on inducing Th9 cells is due to histone modifications caused by this factor, and T cells that do not have such factor are not able to generate IL-9 in IL-4 and TGF-β-containing culture media. As with IRF4, silencing of PU1-related mRNA by SiRNAs leads to decreased response of Th9 and also inability to generate IL-9.17 Other factors which become activated following TGF- $\beta$ 's effect on T cells are SMADs molecules that potentially play a role in activating IRF4. A study with an experimental model of asthma demonstrated that SMAD2 and SMAD3 bound to IRF4 directly and exerted exponential effects on IL-9 production

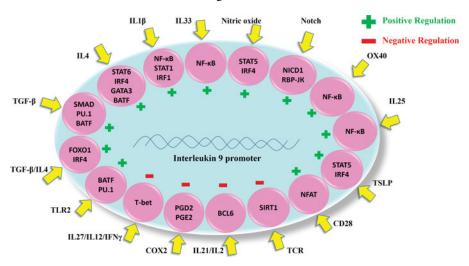


Figure 2. A complex set of cytokines and transcription factors involved in Th9 phenotype induction and IL-9 production by murine and human lymphocytes. Interactions among these factors in different diseases to bind to the gene and amplify IL-9 mRNA varies, which represents the plasticity of T helper cells and dependency of their differentiation on the types of inflammatory responses involved in the environment.

by T cells. This study, inconsistent with the previous studies, demonstrated that instead of PU1, SMAD2 and SMAD3 directly bound to IRF4 via MH1 and MH2 domains and therefore activated IL-9 gene.<sup>74</sup> Differentiation of Th9 cells seems to be varies under different conditions depending on the involved transcription factors and the way they get activated. Besides that, TSLP, nitric oxide, and IL-1 $\beta$  play important roles in activating Th9 cells, such that increased expression of IL-9 has been reported in response to such stimuli in mouse and human T lymphocytes. 75,76 In addition to the effects of IL-4 and TGF- $\beta$  on T cells, TCR signaling causes increase in certain factors such as intracellular domains NFAT, AP1, Notch1, and NF- $\kappa$ B that are likely to contribute to binding to the IL-9 gene promoter.<sup>77</sup> Differentiation of Th9 cells and IL-9 secretion can even be dependent on type I interferons and IL-21.<sup>78</sup> It has been shown that basic leucine zipper ATF-like transcription factor (BATF), as an AP1 family transcription factor, can directly affect Th9 induction, such that the IL-9-dependent secretion of BATF is greater in atopic children's T cells compared to healthy children's T cells. In addition, BATF-deficient T cells cannot produce IL-9 through allergic responses in the mouse model.<sup>79</sup> In addition, engagement of TLR2 signaling in CD4+ T cells under Th9 differentiation increased the expression of BATF and PU1, which are crucial for IL-9 transcription.80 Recently a study indicated that induction of IL-9 protein could be positively dependent on transcription factor FOXO1 in cooperation with IRF4. FOXO1 is a member of forkhead family transcription factors which has been shown to promote both Th9 and Th17 allergic responses through IL-9 induction.<sup>81</sup> It has also been shown that nitric oxide can positively impact on Th9 polarization and expression of IRF4/STAT5 by increasing IL-4R and TGF- $\beta$ R on human and mouse CD4+ T cells. Accordingly, in an ovalbumin (OVA)-induced airway inflammation, nitric oxide synthase 2 (NOS2)-negative mice exhibited less Th9 response in comparison with wild-type model.<sup>82</sup>

#### 3.2.2. Negative regulators

Th9 cells have been shown to considerably promote antitumor activity in a pulmonary melanoma model through a CCR6-dependent recruitment of specific CD8+ CTLs in tumor tissue; however, Th9-related tumor suppression can be promoted by a deficiency in histone deacetylase SIRT1, which cooperates with mTOR-TAK1-HIF1α complex in glycolysis pathway. 83,84 SiRNA-related or conventional inhibition of SIRT1 in human and mouse CD4+ T cells promoted both pathologic allergic responses and antitumor activity by Th9 cells.83 The transcriptional B cell repressor lymphoma 6 (BCL6), which is mainly required for follicular helper T cell development, can

impair Th9 induction and IL-9 transactivation. Ectopic expression of IFNγ, IL-2, and IL-21 in EAE results in BCL6-dependent inhibition of IL-9 which can exacerbate the disease symptoms. 65,85 Also, it has been reported that IFN $\gamma$  can increase the expression of IL-27 by mast cells through the activation of STAT1.86 As indicated before, more pronounced Th9 response is significantly relate to allergic lung inflammation. In an experimental model of allergic lung failure, it was observed that cyclooxygenase enzyme 2 was the main inducer of prostaglandins D2 and E2 which were the pivotal downregulators of Th9derived IL-9 through the inhibition of IL-17 receptor B. In addition, Cox2-negative CD4+ T cells can considerably express IL-9 protein in a culture media containing IL-4 and TGF- $\beta$ , which confirms the negative effects of Cox2 on Th9 activation.<sup>87</sup>

#### 4. Th9 cells: Infection and allergic responses

IL-9 and Th9 cells play significant roles in mucosal immunogenicity throughout asthma-induced allergic responses, airway inflammation, and allergic rhinitis.88 Genetically, chromosome 5, IL-9 and its receptor, and the secretion rate of serum IgE are directly associated.<sup>89,90</sup> The pathogenicity of IL-9 has been frequently demonstrated in the mouse model of allergy such that injecting mice with this protein has led to increased serum IgE levels, eosinophils infiltration, and airway hyperresponsiveness (AHR).<sup>91,92</sup> In mouse aspergillosis, IL-9 causes such responses in transgenic state.<sup>93</sup> In addition, in experimental models of asthma, several factors such as TSLP, tumor necrosis factor receptor superfamily member 4 (OX40), TRAF6 ligase, BATF kinase, NIK, and NF- $\kappa$ B are the stimulators of IL-9 promoter, and therefore play critical roles in activating Th9 cell-dependent inflammatory pathways in these models.<sup>75,94–97</sup> Goblet cell metaplasia is also one of the IL-9 effects throughout allergic inflammation which is associated with IL-13 and the activation of hematopoietic cells.<sup>35</sup> Anti-IL-9 antibody therapy in a mouse model of allergy alleviated the symptoms, although allergic reactions and the infiltration of mast cells may occur even in the experimental model of asthma in transgenic mice in the absence of the gene. 98,99 In asthmatic patients, the secretion levels of IL-9 and its receptor increase significantly. 100 Besides that, IL-9 can cause increase in mucus secretion, infiltration of eosinophils, mastocytes, and neutrophils, and disruption of airway tissue collagenous structure. 101 High amounts of IL-9 have also been demonstrated in allergic rhinitis, which is directly correlated with the infiltration rate of nasal eosinophilia. 102 Recently, traces of Th9 cells and IL-9 have been reported in certain infections. Regarding parasitic infections, evidence on IL-9 effects

are inconsistent. Increase in IgG generation and bowel hypermotility as well as effect on muscles to excrete the parasite, the activation of tissue mastocytosis, and ultimately increased activity of eosinophils to fight nematodes such as Trichinella and Trichuris muris are all dependent on IL-9 and its protective effects. On the other hand, blocking IL-9 by antibody in these models led to opposite outcomes that confirm its effect on intestinal clearance. 103,104 In contrast, in mouse experimental models of Giardia lamblia and Nippostrongylus brasiliensis infections, no effect was reported on defense mechanisms against the parasites in the absence of IL-9, and they were easily cleared of infection. 105 Besides that, in mouse model of schistosomiasis, it is noteworthy that granuloma derived from Schistosomia eggs was formed even in the absence of IL-9.106 In patients with alveolar echinococcosis, who are predisposed to acute liver damages, the secretion rate of IL-9 and the expression of IRF4 and PU1 were dramatically increased in peripheral blood mononuclear cells (PBMCs) and inflamed liver tissues, which represents enhanced activity of Th9 cells in this condition. 107 Certain effects of IL-9 have also been reported in viral infections. Respiratory syncytial virus (RSV) is one of the most common causes of viral infections in children that is closely associated with asthma incidence and also causes bronchitis or chronic inflammation of the bronchi. 108 In experimental model, increased IL-9 in RSV infection and rapid clearance of the virus in case of blocking this protein were reported. 109 Throughout infection with gramnegative bacteria such as Pseudomonas, the symptoms are relieved after injecting IL-9 to mice infected with this bacterium, which is due to decrease in TNFα and IFN $\gamma$ , and increase in IL-10 secretion. <sup>110</sup> In patients with chronic mucocutaneous candidiasis (CMC), the levels of IL-9 were decreased compared to healthy controls, which indicates that Th1 and Th17 responses are dominant in this milieu. 111 Table 1 demonstrates the effects of IL-9 and Th9 cells during certain infections. 107,110,112-119

#### 5. Th9 and intestinal inflammation

#### 5.1. Intestinal tolerance: A unique feature

The digestive immune system is the evolutionary oldest and the most sophisticated organ of the body, which designed pivotal tolergenic mechanisms and a specific cellular map to maintain the mucosal layers sterile and separated from the luminal antigens that is estimated to be 100 million microbial cells.<sup>120,121</sup> Effective mucosal homeostasis is mostly dependent on early encountering to various microorganisms of normal flora and also proper nutrition. For instance, early exposure to some clostridium subtypes is necessary for accumulation of

CD4+CD25+ Tregs in the colon which have protective effects on IBDs. Accordingly, reduced clostridium serotypes in IBD patients were observed. 122-124 Intraepithelial lymphocytes (IELs) are other important cells participated in colon homeostasis which are involved in mucosal defense and wound repair and their differentiation is associated mainly with aryl hydrocarbon receptors. It has been reported that a cruciferos vegetables-depleted nutrition negatively affects Ahr and IELs polarization leads to invasion of pathogens and intensified inflammatory responses.<sup>125</sup> Also deficiency in vitamins A and D, tryptophan, and milk-derived taurocholic acid all are related to Tregs dysfunction and impaired antimicrobial peptides which may result in epithelial damage and intestinal inflammation. 126,127 Goblet cells are the main source of glycosylated mucins which construct two distinct mucosal layers on epithelium surface and restrict the direct adhesion of microbial pathogens to the epithelium. Severe functional defects of goblet cells specially MUC2 gene have been reported in UC patients. 128-130 Paneth cells are other protective cells that play an important role in maintaining hemostasis in small intestine through the secretion of antimicrobial peptides. Genetical defects in autophagy-related genes NOD2, ATGL16, and XBP1, which are severely associated with Crohns disease incidence, can impair the release of antimicrobial proteins from paneth cells. 120,131 As a secondary source of IL-9, NKT cells recruit to inflammation sites in response to CXCL16 chemokine and produce IL-13, which promotes UC pathogenesis. It has been reported that intestinal colonization of a neonatal mouse with microbial normal flora inhibits infiltration of iNKT cells and protects from oxazolone-induced colitis. 132,133

#### 5.2. Th9 and experimental ulcerative colitis

Recent studies on mouse models of IBD and patients with IBD especially UC, demonstrated that the production of IL-9 and Th9-related transcription factors are increased.<sup>6,72,134,135</sup> Here we discuss how these cells potentially exhibit pathological activity through the inflammatory responses in the large intestine. Crohn's disease and UC are two important types of IBD that cause inflammation of the gastrointestinal tract lining tissues due to genetical predisposition and effects of environmental factors. Although these diseases share certain characteristics, they are completely different in terms of pathophysiology, the types of cell response, and genes involved in the incidence of the disease. 136 Crohn's disease causes deep inflammation alongside sporadic (but not continuous) involving of submucosa throughout the whole gastrointestinal tract from the mouth to the anus, while UC causes superficial inflammation restricted



**Table 1.** Pathologic and protective activity of Th9 cells in several microbial infections.

Type of infection	Secretion sources of IL-9	Related effects	Ref.
Parasitic—Schistosoma japonicum	Th9 cells-CDS+ cells-NKT cells-yo T cells	Promote liver failure and hepatic damages	115
Parasitic— <i>Trichinella spiralis</i>	Th9 cells-Th2 cells	Promote bowel mastocytosis and intestinal permeability leading to parasite expulsion	112
Parasitic— <i>Trichuris muris</i>	Th9 cells-Th2 cells	Increase muscle contraction and goblet cells hyperplasia and promote worm expulsion	112
Parasitic—H. polygyrus	CD4+ T cells (CTLA4 dependent)	Single infection: low amount, decreased mastocytosis in mice coinfection with <i>T. spiralis</i> : high amount, increased mast cells hyperplasia and worm expulsion in mice	112,114
Parasitic—Strongyloides stercolaris	Th9 cells	Increase Ag-specific responses and protection in human patients	112,114
Parasitic—Giardia lambiia, Nippostrongylus brasiliensis	Th9 cells	Complete parasite clearance even in absence of IL-9	112,114
Parasitic—Echinococcus mutilocularis	Th9 cells	Increase parasite persistence in host in human patient	107,112,114
Bacterial—Pseudomonas aeruginosa	T cells	Protective effects through suppression of TNF-a and induction of IL-10	110
Bacterial— <i>Chlamydia</i> muridarum	CD4+ T cells	No significant effect on Ab response and protection against lung infection in mice	116
Viral—Coxsackievirus B3	CD4+ and CD8+ T cells	Protective effects—inhibited virus replication	118
Viral—RSV	CD4+ T cells	Increase mucus production and lung failure	108,117
Fungal—Candida albicans	Th2, Th9 cells	General defect in IL-9 induction—protective effects in CMC	119

in the large intestine. 137 In Crohn's disease, Th1- and Th17-related cytokines are involved, while in UC, Th2 responses are dominant. 138 Due to the similarity and dependence of Th2 and Th9 responses on each other, increased IL-9 and Th9 cells represents a component of UC pathogenesis. In other words, these cells do not receive their required signals to completely differentiate in Crohn's disease inflammatory milieu. First, we touch on evidence about the presence of Th9 cells in the experimental models of IBD. In mouse model of oxazolone-induced colitis where Th2 responses are dominant as with human colitis, the gene expression of the IL-9 in the inflamed intestinal tissue and CD4+ cells in the spleen are increased compared to wild-type model. In this model, immunohistochemical analysis confirmed the presence and increased frequency of mucosal CD4+IL-9+ cells. Although the production of small amounts of IL-9 by innate lymphoid cells, CD117+ mast cells, and CD11c+ cells has been reported, Th9 lymphocytes are the most important source of this protein in oxazoloneinduced colitis. 105 After rectal injection of oxazolone and inducing intestinal inflammation in mice, Gerlach et al. induced IL-9 deficiency in mice via two methods as follow: using anti-IL-9 antibody and inducing genetic defect in producing this cytokine. In both models of IL-9 deficiency, histopathological investigations demonstrated that inflammation was much lower compared to the wild-type model according to weight loss, clinical scores calculated by high-resolution miniendoscopy, and reactive oxygen species (ROS) bioluminescence evaluation. These evidences are indicative of the effective role of IL-9 in regulating intestinal inflammatory responses in this model. Inducing deficiency in PU1, a critical factor of Th9

induction in this model, caused decrease in the expression of IL-9 and Th2 cytokines and significantly decreased the disease symptoms. Increased expression of the IL-9 receptor on the Epcam+ intestinal epithelial cells, also confirms IL-9 activity against the tolergenic property of this tissue.<sup>6</sup> In both patients with UC and experimental model of this disease, pathogenic changes were induced by IL-9 in the intestinal epithelial barriers and tight junction proteins. In oxazolone-induced colitis where Th2 response is dominant, expression of intestinal tissue-permeable proteins such as claudins 1 and 2 increases, but in contrast, claudin 3 and occludin that increase the inhibitory activity of lining barriers significantly, were not changed. Bacterial translocation and fluorescein isothiocyanate-dextran effect on decreased permeability of the intestinal tissue lining membrane, demonstrated that the permeability of these membranes to many antigens decreased if the IL-9 was eliminated.<sup>6</sup> Through using organoid cell culture, creating three-dimensional crypts of the intestine, and exerting IL-9's effect on them, a study reported that IL-9 caused disruption in tissue remodeling mechanisms and cell proliferation.<sup>6</sup> Gerlach et al. conducted surgery to induce injury in mouse intestinal tissue to investigate the slowdown of tissue repair process and IL-9-dependent wound healing. Two days after surgery, recombinant IL-9's effect showed that the duration of wound healing increased considerably, which confirmed the potential role of IL-9 in wound healing process.<sup>6</sup> In an experimental model, namely trinitrobenzenesulfonic acid (TNBS)induced colitis, in which Th1 response is dominant as with human Crohn's disease, in contrast to the former model, IL-9 had no effect on the expression of claudin 2 but affected the expression of occludin and claudin 1. This

indicates that in addition to IL-9, other factors regulate the expression of these molecules in different inflammatory conditions. In TNBS-induced colitis in wild-type and IL-9 deficient mice, the rates of inflammation and weight loss in IL-9 knockout mice were much lower than those in wild-type mice. Besides that, goblet cell dysfunction, wound induction, and mononuclear cells accumulation were much less pronounced in IL-9 knockout model. The immunofluorescence study of lamina propria cells revealed increased frequency of PU1+CD4+ cells in the wild-type model, representing that the expression of this factor depends on the frequency of the Th9 cells and the amount of IL-9 in this model.<sup>5</sup> Real-time PCR, western blotting, and study of the tight junction proteins expression in TNBS-induced colitis showed that unlike claudin 1, the expression of claudins 4 and 7, Jam-A, and occludin increased in the IL9-deficient model compared to the wild-type model. Inconsistency in findings on claudins expression in the models of oxazolone and TNBSinduced colitis is due to different dominant cell responses of these two models, such that it can be argued that IL-9 and Th9 cells exhibit regulatory activity in the TNBSinduced colitis model due to similarity of this condition to Crohn's disease and Th1/Th17 dominancy.<sup>5</sup> Immunohistochemical analyses of another experimental model of intestinal colitis that was induced by the effect of dextran sodium sulphate (DSS), showed an increase in Th9 cells expressing PU1 and CD3 markers. In addition, anti-IL-9 antibody treatment for 2 weeks caused relief of the disease symptoms and decreased the inflammatory mediators due to decline in activated lymphocytes in the lamina propria of mouse intestine. 139 Taken together, it can be argued that although Th9 cells need different transcription factors to differentiate in various inflammatory conditions, these cells increase in CD4+ cells-related inflammation, dysfunction of the intestinal lining barriers, and tolergenic dysfunction of epithelial cells by changing the expression of tight junction proteins, through secretion of IL-9 in the experimental models of IBDs especially UC (Figure 3).

#### 5.3. Th9 clinical manifestations in UC

Imbalance between the effector cells and immunomodulatory cells is one of the most important reasons for the incidence and intensification of inflammatory responses particularly in gastrointestinal diseases such as gastritis or peptic ulcers. 140-143 Meanwhile, increased in T cells, such as Th17, and associated upregulation of the expression of inflammatory mediators such as IL-6, IL-23, and IL-33 is one of the important causes of different clinical presentations of Helicobacter pylori-related infections in the stomach such as duodenal ulcers and gastritis. 144-148 The role of helper T

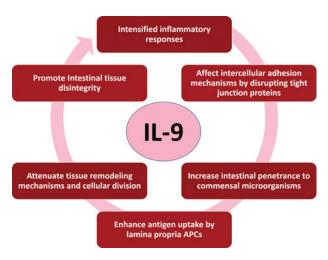


Figure 3. Pathological effects of IL-9 on expression of tight junction proteins and tissue repair mechanisms leads to dysfunction of epithelial barriers and declined tolergenic nature of intestinal tissue.

cells in producing inflammatory mediators has also been demonstrated during IBD. For example, Th1 and Th17 cells in Crohn's disease, and Th2 cells-related cytokines such as IL-4 and IL-13 in UC are increased in expression and function. 149,150 Following the elimination of proteins that maintain tight intercellular junctions, integrity of epithelial cells will be compromised resulting in easy pass of luminal antigens and subsequently intensification of IBD-induced inflammatory responses. Different proteins have been identified in regards to this issue such as IL-17 and IL-22.151-153 Here we discuss mainly on the role of IL-9 and possibly Th9 cells in expediting inflammation and the pathogenicity of IBDs. The role of these cells have been studied in different diseases such as allergies, certain autoimmune conditions such as lupus, and different infections as we already discussed. 100,154 Studies on patients with UC demonstrated an important role for Th9 cells in UC pathogenesis. The study of geneexpression levels of IL-9 alongside other inflammatory cytokines such as IL-6 and IL-17A which are associated with Th17 subtype, confirmed increased expression of these genes in inflamed biopsies taken from the intestines of UC patients. Interestingly, the expression level of IL-9 mRNA was directly correlated with inflammation score.<sup>72</sup> Additionally, two-color immunofluorescence staining on tissue samples demonstrated an increase in the frequency of mucosal CD3+IL9+ cells in such patients. IRF4 and PU1 have positive effects on IL-9 expression in T cells in the presence of TGF- $\beta$  and IL-4 and have been known as two main factors for the differentiation of Th9 cells.66 The gene-expression levels of these molecules increase in patients with intestinal colitis and are directly correlated with intensification of inflammation and disease symptoms.<sup>72</sup> After culturing peripheral blood lymphocytes



under necessary conditions to induce Th9 cells, 6% of these cells secrete IL-9, and the coexpression of CD3, IRF4, and IL-9 was reported in 25% of these cells. In addition, the expression of the  $\alpha 4/\beta 7$  integrin on the IL-9-generating lymphocytes increased in peripheral blood of patients with UC, which indicates their homing ability in the epithelial tissue of the large intestine. Furthermore, flow cytometric analysis has shown increased expression of IL-9 receptor on the polymorphonuclear leukocytes of the peripheral blood in patients with UC compared to controls. Finally, this study has demonstrated that the resistance of polymorphonuclear cells (PMNs) to apoptosis with IL-9 is increased in a dose-dependent manner. As a result, this protein can serve as a significant inducer of antiapoptotic signals of peripheral blood mononuclear cells in such patients.<sup>72</sup> Immunohistochemical staining of  $\alpha E\beta 7$  and  $\alpha 4\beta 7$  integrins on T lymphocytes revealed a remarkable mucosal accumulation of such cells in IBD patients. Therapeutically, targeting T cells homing in the gut is a growing field of IBD treatment now. As reported, intestinal recruitment of Th9 cells in IBD patients can be inhibited using antibody blockade of  $\alpha E\beta 7$  integrin. Etrolizumab monoclonal antibody to  $\alpha E\beta 7$  integrin effectively blocked Th9 adhesion on mucosal E-cadherin and MadCam1.155

#### 6. Conclusion

Th9 cells are a novel described subtype of helper T cells. It seems that Th9 cells promote allergic damages in patients and experimental models. In addition, the traces of Th9 cells have been found in certain microbial infections, which require further investigation. We have already provided several evidences on the role of Th9 cells in IBD-induced pathologies. As we discussed increased frequency of CD4+IL-9+ cells, so called Th9 cells are reported in animal models and patients with UC. The changes in gene expression of IL-9 in biopsy specimens of patients with IBD, the effects of such cytokines on the expression of proteins that maintain tight intercellular junctions and their potential role on immune tolerance in the gut merits further investigations. Although the cited studies offered some answers to many questions on the role of Th9 cells and IL-9 in the pathogenesis of UC, many questions remained unanswered. Identification of specific transcription factors involved in the differentiation of these cells in inflammatory conditions such as UC and in depth understanding of how expression of this cytokine at gene and protein levels can be manipulated are essential for potential therapeutic strategies.

#### **Competing interests**

None declared.

#### **Acknowledgement**

The authors are grateful to the staffs of Students Research Committee, Shahrekord University of Medical Sciences, Shahrekord, Iran.

#### **Funding**

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

#### **ORCID**

Mahmoud Rafieian-Kopaei http://orcid.org/0000-0003-3190-7863

#### References

- 1. Zhu J, Paul WE. CD4 T cells: fates, functions, and faults. Blood. 2008;112(5):1557-1569. doi:10.1182/blood-2008-05-078154. PMID:18725574.
- 2. Luckheeram RV, Zhou R, Verma AD, Xia B. CD4+T cells: differentiation and functions. Clin Dev Immunol. 2012;2012:925135.
- 3. Hirahara K, Nakayama T. CD4+ T-cell subsets in inflammatory diseases: beyond the Th1/Th2 paradigm. Int Immunol. 2016;28(4):163–171. doi:10.1093/intimm/dxw006. PMID:26874355.
- 4. Jabeen R, Kaplan MH. The symphony of the ninth: the development and function of Th9 cells. Curr Opin Immunol. 2012;24(3):303-307. doi:10.1016/j.coi.2012.02.001. PMID:22365614.
- 5. Gerlach K, McKenzie AN, Neurath MF, Weigmann B. IL-9 regulates intestinal barrier function in experimental T cell-mediated colitis. Tissue barriers. 2015;3(1-2):e983777. doi:10.4161/21688370.2014.983777. PMID:25838986.
- 6. Gerlach K, Hwang Y, Nikolaev A, et al. TH9 cells that express the transcription factor PU. 1 drive T cellmediated colitis via IL-9 receptor signaling in intestinal epithelial cells. Nat Immunol. 2014;15(7):676-686. doi:10.1038/ni.2920. PMID:24908389.
- 7. Turner JR. Intestinal mucosal barrier function in health and disease. Nat Rev Immunol. 2009;9(11):799. doi:10.1038/nri2653. PMID:19855405.
- 8. Zhang N, Bevan MJ. CD8+ T cells: foot soldiers of the immune system. Immunity. 2011;35(2):161-168. doi:10.1016/j.immuni.2011.07.010. PMID:21867926.
- 9. Elahi S, Horton H. Association of HLA-alleles with the immune regulation of chronic viral infections. Int J Biochem Cell Biol. 2012;44(8):1361-1365. doi:10.1016/j.biocel.2012.05.003. PMID:22595281.
- 10. Sakaguchi S, Miyara M, Costantino CM, Hafler DA. FOXP3+ regulatory T cells in the human immune system. Nat Rev Immunol. 2010;10(7):490. doi:10.1038/nri2785. PMID:20559327.
- 11. Nakayamada S, Takahashi H, Kanno Y, O'Shea JJ. Helper T cell diversity and plasticity. Curr Opin Immunol. 2012;24(3):297-302. doi:10.1016/j.coi.2012.01.014. PMID:22341735.



- 12. Basu R, O'Quinn DB, Silberger DJ, et al. Th22 cells are an important source of IL-22 for host protection against enteropathogenic bacteria. *Immunity.* 2012;37(6):1061–1075. doi:10.1016/j.immuni.2012.08.024. PMID:23200827.
- 13. Jia L, Wu C. The biology and functions of Th22 cells. *Adv Exp Med Biol.* 2014;841:209–230.
- 14. Palmer MT, Weaver CT. Autoimmunity: increasing suspects in the CD4+ T cell lineup. *Nat Immunol.* 2010;11(1):36–40. doi:10.1038/ni.1802. PMID:20016508.
- 15. Jäger A, Kuchroo VK. Effector and regulatory T-cell subsets in autoimmunity and tissue inflammation. *Scand J Immunol.* 2010;72(3):173–184. doi:10.1111/j.1365-3083.2010.02432.x. PMID:20696013.
- Lu Y, Hong S, Li H, et al. Th9 cells promote antitumor immune responses in vivo. *J Clin Invest*. 2012;122(11):4160. doi:10.1172/JCI65459. PMID: 23064366.
- 17. Chang H-C, Sehra S, Goswami R, et al. The transcription factor PU. 1 is required for the development of IL-9-producing T cells and allergic inflammation. *Nat Immunol.* 2010;11(6):527–534. doi:10.1038/ni.1867. PMID:20431622.
- 18. Pan H-F, Leng R-X, Li X-P, Zheng SG, Ye D-Q. Targeting T-helper 9 cells and interleukin-9 in autoimmune diseases. *Cytokine Growth Factor Rev.* 2013;24(6):515–522. doi:10.1016/j.cytogfr.2013.09.001. PMID:25215394.
- 19. Maddur MS, Miossec P, Kaveri SV, Bayry J. Th17 cells: biology, pathogenesis of autoimmune and inflammatory diseases, and therapeutic strategies. *Am J Pathol.* 2012;181(1):8–18. doi:10.1016/j.ajpath.2012.03.044. PMID:22640807.
- 20. Crotty S. T follicular helper cell differentiation, function, and roles in disease. *Immunity*. 2014;41(4):529–542. doi:10.1016/j.immuni.2014.10.004. PMID:25367570.
- 21. Rojas-Zuleta WG, Vásquez G. Th9 lymphocytes: a recent history from IL-9 to its potential role in rheumatic diseases. *Autoimmunity Rev.* 2016;15(7):649–655. doi:10.1016/j.autrev.2016.02.020.
- Goswami R, Kaplan MH. A brief history of IL-9. *J Immunol*. 2011;186(6):3283–3288. doi:10.4049/jimmunol.1003049. PMID:21368237.
- 23. Zhao P, Xiao X, Ghobrial RM, Li XC. IL-9 and Th9 cells: progress and challenges. *Int Immunol.* 2013;25(10):547–551. doi:10.1093/intimm/dxt039. PMID:24027199.
- 24. Li J, Chen S, Xiao X, et al. IL-9 and Th9 cells in health and diseases—from tolerance to immunopathology. *Cytokine Growth Factor Rev.* 2017;37:47–55. doi:10.1016/j.cytogfr.2017.07.004.
- 25. Wilhelm C, Turner J-E, Van Snick J, Stockinger B. The many lives of IL-9: a question of survival [quest]. *Nat Immunol.* 2012;13(7):637–641. doi:10.1038/ni.2303. PMID:22713829.
- 26. Bauer JH, Liu KD, You Y, et al. Heteromerization of the  $\gamma$ c chain with the interleukin-9 receptor  $\alpha$  subunit leads to STAT activation and prevention of apoptosis. *J Biol Chem.* 1998;273(15):9255–9260. doi:10.1074/jbc.273.15.9255. PMID:9535918.
- 27. Demoulin J-B, Uyttenhove C, Van Roost E, et al. A single tyrosine of the interleukin-9 (IL-9) receptor is required for STAT activation, antiapoptotic activity, and growth

- regulation by IL-9. *Mol Cell Biol.* 1996;16(9):4710–4716. doi:10.1128/MCB.16.9.4710. PMID:8756628.
- 28. Demoulin J-B, Van Roost E, Stevens M, et al. Distinct roles for STAT1, STAT3, and STAT5 in differentiation gene induction and apoptosis inhibition by interleukin-9. *J Biol Chem.* 1999;274(36):25855–25861. doi:10.1074/jbc.274.36.25855. PMID:10464327.
- Mock BA, Krall M, Kozak CA, et al. IL9 maps to mouse chromosome 13 and human chromosome 5. *Immuno-genetics*. 1990;31(4):265–270. doi:10.1007/BF00204898. PMID:1970335.
- Stassen M, Schmitt E, Bopp T. From interleukin-9 to T helper 9 cells. *Ann N Y Acad Sci.* 2012;1247(1):56–68. doi:10.1111/j.1749-6632.2011.06351.x. PMID:22235761.
- 31. Veldhoen M, Uyttenhove C, Van Snick J, et al. Transforming growth factor-β 'reprograms' the differentiation of T helper 2 cells and promotes an interleukin 9-producing subset. *Nat Immunol.* 2008;9(12):1341–1346. doi:10.1038/ni.1659. PMID:18931678.
- Gessner A, Blum H, Röllinghoff M. Differential regulation of IL-9-expression after infection with Leishmania major in susceptible and resistant mice. *Immunobiology*. 1993;189(5):419–435. doi:10.1016/S0171-2985(11)80414-6. PMID:8125519.
- 33. Kaplan MH. Th9 cells: differentiation and disease. *Immunol Rev.* 2013;252(1):104–115. doi:10.1111/imr.12028. PMID:23405898.
- 34. Sitkauskiene B, Rådinger M, Bossios A, et al. Airway allergen exposure stimulates bone marrow eosinophilia partly via IL-9. *Respir Res.* 2005;6(1):33. doi:10.1186/1465-9921-6-33. PMID:15823208.
- 35. Steenwinckel V, Louahed J, Orabona C, et al. IL-13 mediates vivo IL-9 in activities cells epithelial but not on hematopoilung etic cells. Immunol. 2007;178(5):3244-3251. doi:10.4049/jimmunol.178.5.3244. PMID:17312173.
- Osterfeld H, Ahrens R, Strait R, et al. Differential roles for the IL-9/IL-9 receptor α-chain pathway in systemic and oral antigen-induced anaphylaxis. *J Allergy Clin Immunol*. 2010;125(2):469–476. e2. doi:10.1016/j.jaci.2009.09.054. PMID:20159257.
- 37. Nowak EC, Weaver CT, Turner H, et al. IL-9 as a mediator of Th17-driven inflammatory disease. *J Exp Med*. 2009;206(8):1653–1660. doi:10.1084/jem.20090246. PMID:19596803.
- 38. Elyaman W, Bradshaw EM, Uyttenhove C, et al. IL-9 induces differentiation of TH17 cells and enhances function of FoxP3+ natural regulatory T cells. *Proc Natl Acad Sci USA*. 2009;106(31):12885–12890. doi:10.1073/pnas.0812530106. PMID:19433802.
- 39. Noelle RJ, Nowak EC. Cellular sources and immune functions of interleukin-9. *Nat Rev Immunol*. 2010;10(10):683–687. doi:10.1038/nri2848. PMID:20847745.
- 40. Li H, Nourbakhsh B, Ciric B, et al. Neutralization of IL-9 ameliorates experimental autoimmune encephalomyelitis by decreasing the effector T cell population. *J Immunol.* 2010;185(7):4095–4100. doi:10.4049/jimmunol.1000986. PMID:20805418.
- 41. Jäger A, Dardalhon V, Sobel RA, et al. Th1, Th17, and Th9 effector cells induce experimental autoimmune encephalomyelitis with different pathological



- phenotypes. J Immunol. 2009;183(11):7169-7177. doi:10.4049/jimmunol.0901906. PMID:19890056.
- 42. Peron JPS, Yang K, Chen M-L, et al. Oral tolerance reduces Th17 cells as well as the overall inflammation in the central nervous system of EAE mice. J Neuroimmunol. 2010;227(1):10-17. doi:10.1016/j.jneuroim.2010.06.002. PMID:20580440.
- 43. Beriou G, Bradshaw EM, Lozano E, et al. TGF- $\beta$  induces IL-9 production from human Th17 cells. J Immunol. 2010;185(1):46-54. doi:10.4049/jimmunol.1000356. PMID:20498357.
- 44. Lu L-F, Lind EF, Gondek DC, et al. Mast cells are essential intermediaries in regulatory T-cell tolerance. Nature. 2006;442(7106):997-1002. doi:10.1038/nature05010. PMID:16921386.
- 45. Mukai K, Tsai M, Saito H, Galli SJ. Mast cells as sources of cytokines, chemokines, and growth factors. Immunol 2018;282(1):121-150. doi:10.1111/imr.12634. PMID:29431212.
- 46. John ALS, Abraham SN. Innate immunity and its regulation by mast cells. J Immunol. 2013;190(9):4458-4463. doi:10.4049/jimmunol.1203420. PMID:23606723.
- 47. Chen C-Y, Lee J-B, Liu B, et al. Induction of interleukin-9-producing mucosal mast cells promotes susceptibility to IgE-mediated experimental food allergy. Immunity. 2015;43(4):788-802. doi:10.1016/j.immuni.2015.08.020. PMID:26410628.
- 48. Sehra S, Yao W, Nguyen ET, et al. TH9 cells are required for tissue mast cell accumulation during allergic inflammation. J Allergy Clin Immunol. 2015;136(2):433-440. doi:10.1016/j.jaci.2015.01.021. PMID:25746972.
- 49. Stassen M, Müller C, Arnold M, et al. IL-9 and IL-13 production by activated mast cells is strongly enhanced in the presence of lipopolysaccharide: NF-κB is decisively involved in the expression of IL-9. J Immunol. 2001;166(7):4391-4398. doi:10.4049/jimmunol.166.7.4391. PMID:11254693.
- 50. Stassen M, Arnold M, Hültner L, et al. Murine bone marrow-derived mast cells as potent producers of IL-9: costimulatory function of IL-10 and kit ligand in the presence of IL-1. J Immunol. 2000;164(11):5549-5555. doi:10.4049/jimmunol.164.11.5549. PMID:10820228.
- 51. Wiener Z, Falus A, Toth S. IL-9 increases the expression of several cytokines in activated mast cells, while the IL-9-induced IL-9 production is inhibited in mast cells of histamine-free transgenic mice. Cytokine. doi:10.1016/j.cyto.2004.01.006. 2004;26(3):122-130. PMID:15135806.
- 52. Farhadi A, Fields JZ, Keshavarzian A. Mucosal mast cells are pivotal elements in inflammatory bowel disease that connect the dots: stress, intestinal hyperpermeability and inflammation. World J Gastroenterol. 2007;13(22):3027. doi:10.3748/wjg.v13.i22.3027. PMID:17589915.
- 53. Stoyanova II, Gulubova MV. Mast cells and inflammatory mediators in chronic ulcerative colitis. Acta Histochem. 2002;104(2):185-192. doi:10.1078/0065-1281-00641. PMID:12086339.
- 54. Lauwerys BR, Garot N, Renauld J-C, Houssiau FA. Cytokine production and killer activity of NK/T-NK cells derived with IL-2, IL-15, or the combination of IL-12 and IL-18. J Immunol. 2000;165(4):1847-1853. doi:10.4049/jimmunol.165.4.1847. PMID:10925263.

- 55. Jones TG, Hallgren J, Humbles A, et al. Antigeninduced increases in pulmonary mast cell progenitor numbers depend on IL-9 and CD1d-restricted NKT cells. J Immunol. 2009;183(8):5251-5260. doi:10.4049/jimmunol.0901471. PMID:19783672.
- 56. Nagato T, Kobayashi H, Kishibe K, et al. Expression of interleukin-9 in nasal natural killer/T-cell lymphoma cell lines and patients. Clin Cancer Res. 2005;11(23):8250-8257. doi:10.1158/1078-0432.CCR-05-1426. PMID:16322282.
- 57. Klose CS, Artis D. Innate lymphoid cells as regulators of immunity, inflammation and tissue homeostasis. Nat Immunol. 2016;17(7):765. doi:10.1038/ni.3489. PMID:27328006.
- 58. Mohapatra A, Van Dyken SJ, Schneider C, Nussbaum JC, Liang H-E, Locksley RM. Group 2 innate lymphoid cells utilize the IRF4-IL-9 module to coordinate epithelial cell maintenance of lung homeostasis. Mucosal Immunol. 2016;9(1):275. doi:10.1038/mi.2015.59. PMID:26129648.
- 59. Wilhelm C, Turner J-E, Van Snick J, Stockinger B. The many lives of IL-9: a question of survival? Nat Immunol. 2012;13(7):637. doi:10.1038/ni.2303. PMID:22713829.
- 60. Kaplan MH, Schindler U, Smiley ST, Grusby MJ. Stat6 is required for mediating responses to IL-4 and for the development of Th2 cells. Immunity. 1996;4(3):313-319. doi:10.1016/S1074-7613(00)80439-2. PMID:8624821.
- 61. Lu L, Ma J, Wang X, et al. Synergistic effect of TGF- $\beta$  superfamily members on the induction of Foxp3+ Treg. Eur J Immunol. 2010;40(1):142-152. doi:10.1002/eji.200939618. PMID:19943263.
- 62. Cosmi L, Maggi L, Santarlasci V, et al. T helper cells plasticity in inflammation. Cytometry Part A. 2014;85(1):36-42. doi:10.1002/cyto.a.22348.
- 63. Kaplan MH. The transcription factor network in Th9 cells. Seminars Immunopathol. 2017;39(1):11-20.
- 64. Koh B, Hufford MM, Pham D, et al. The ETS family transcription factors Etv5 and PU. 1 function in parallel to promote Th9 cell development. J Immunol. 2016;197(6):2465-2472. doi:10.4049/jimmunol.1502383. PMID:27496971.
- 65. Bassil R, Orent W, Olah M, et al. BCL6 controls Th9 cell development by repressing Il9 transcription. J Immunol. 2014;193(1):198-207. doi:10.4049/jimmunol.1303184. PMID:24879792.
- 66. Perumal NB, Kaplan MH. Regulating Il9 transcription in T helper cells. Trends Immunol. 2011;32(4):146-150. doi:10.1016/j.it.2011.01.006. PMID:21371941.
- 67. Staudt V, Bothur E, Klein M, et al. Interferon-regulatory factor 4 is essential for the developmental program of T helper 9 cells. Immunity. 2010;33(2):192-202. doi:10.1016/j.immuni.2010.07.014. PMID:20674401.
- 68. Huber M, Brüstle A, Reinhard K, et al. IRF4 is essential for IL-21-mediated induction, amplification, and stabilization of the Th17 phenotype. Proc Natl Acad Sci USA. 2008;105(52):20846-20851. doi:10.1073/pnas.0809077106. PMID:19088203.
- 69. Zheng Y, Chaudhry A, Kas A, et al. Regulatory T-cell suppressor program co-opts transcription factor IRF4 to control TH2 responses. Nature. 2009;458(7236):351doi:10.1038/nature07674. PMID: 356. 19182775.



- Brüstle A, Heink S, Huber M, et al. The development of inflammatory TH-17 cells requires interferon-regulatory factor 4. *Nat Immunol.* 2007;8(9):958–966. doi:10.1038/ni1500. PMID:17676043.
- 71. Schmitt E, Klein M, Bopp T. Th9 cells, new players in adaptive immunity. *Trends Immunol.* 2014;35(2):61–68. doi:10.1016/j.it.2013.10.004. PMID:24215739.
- 72. Nalleweg N, Chiriac MT, Podstawa E, et al. IL-9 and its receptor are predominantly involved in the pathogenesis of UC. *Gut.* 2015;64(5):743–755.
- 73. Chang H-C, Han L, Jabeen R, Carotta S, Nutt SL, Kaplan MH. PU. 1 regulates TCR expression by modulating GATA-3 activity. *J Immunol.* 2009;183(8):4887–4894. doi:10.4049/jimmunol.0900363. PMID:19801513.
- 74. Tamiya T, Ichiyama K, Kotani H, et al. Smad2/3 and IRF4 play a cooperative role in IL-9-producing T cell induction. *J Immunol.* 2013;191(5):2360–2371. doi:10.4049/jimmunol.1301276. PMID:23913959.
- 75. Yao W, Zhang Y, Jabeen R, et al. Interleukin-9 is required for allergic airway inflammation mediated by the cytokine TSLP. *Immunity.* 2013;38(2):360–372. doi:10.1016/j.immuni.2013.01.007. PMID:23376058.
- 76. Niedbala W, Besnard A-G, Nascimento DC, et al. Nitric oxide enhances Th9 cell differentiation and airway inflammation. *Nat Commun.* 2014;5:4575. doi:10.1038/ncomms5575. PMID:25099390.
- 77. Elyaman W, Bassil R, Bradshaw EM, et al. Notch receptors and Smad3 signaling cooperate in the induction of interleukin-9-producing T cells. *Immunity*. 2012;36(4):623–634. doi:10.1016/j.immuni.2012.01.020. PMID:22503540.
- 78. Wong MT, Jessica JY, Alonso MN, et al. Regulation of human Th9 differentiation by type I interferons and IL-21. *Immunol Cell Biol.* 2010;88(6):624–631. doi:10.1038/icb.2010.53. PMID:20421880.
- Jabeen R, Goswami R, Awe O, et al. Th9 cell development requires a BATF-regulated transcriptional network. J Clin Invest. 2013;123(11):4641. doi:10.1172/JCI69489. PMID:24216482.
- 80. Karim AF, Reba SM, Li Q, et al. Toll like receptor 2 engagement on CD4+ T cells promotes TH9 differentiation and function. *Eur J Immunol.* 2017;47(9):1513–1524. doi:10.1002/eji.201646846. PMID:28665005.
- 81. Malik S, Sadhu S, Elesela S, et al. Transcription factor Foxo1 is essential for IL-9 induction in T helper cells. *Nat Commun.* 2017;8(1):815. doi:10.1038/s41467-017-00674-6. PMID:28993609.
- 82. Niedbala W, Besnard A-G, Nascimento DC, et al. Nitric oxide enhances Th9 cell differentiation and airway inflammation. *Nat Commun.* 2014;5:4575. doi:10.1038/ncomms5575. PMID:25099390.
- 83. Wang Y, Bi Y, Chen X, et al. Histone deacetylase SIRT1 negatively regulates the differentiation of interleukin-9-producing CD4+ T cells. *Immunity.* 2016;44(6):1337–1349. doi:10.1016/j.immuni.2016.05.009. PMID:27317260.
- 84. Lu Y, Hong S, Li H, et al. Th9 cells promote antitumor immune responses in vivo. *J Clin Invest.* 2012;122(11):4160–4171. doi:10.1172/JCI65459. PMID:23064366.
- 85. Liao W, Spolski R, Li P, et al. Opposing actions of IL-2 and IL-21 on Th9 differentiation correlate

- with their differential regulation of BCL6 expression. *Proc Natl Acad Sci U S A.* 2014;111(9):3508–3513. doi:10.1073/pnas.1301138111. PMID:24550509.
- 86. Murugaiyan G, Beynon V, Da Cunha AP, et al. IFN-γ limits Th9-mediated autoimmune inflammation through dendritic cell modulation of IL-27. *J Immunol.* 2012;189(11):5277–5283. doi:10.4049/jimmunol.1200808. PMID:23125412.
- 87. Li H, Edin ML, Bradbury JA, et al. Cyclooxygenase-2 inhibits T helper cell type 9 differentiation during allergic lung inflammation via down-regulation of IL-17RB. *Am J Respir Crit Care Med.* 2013;187(8):812–822. doi:10.1164/rccm.201211-2073OC. PMID:23449692.
- 88. Neurath MF, Finotto S. IL-9 signaling as key driver of chronic inflammation in mucosal immunity. *Cytokine Growth Factor Rev.* 2016;29:93–99. doi:10.1016/j.cytogfr.2016.02.002. PMID:26976761.
- 89. Doull I, Lawrence S, Watson M, et al. Allelic association of gene markers on chromosomes 5q and 11q with atopy and bronchial hyperresponsiveness. *Am J Respir Crit Care Med.* 1996;153(4):1280–1284. doi:10.1164/ajrccm.153.4.8616554. PMID:8616554.
- 90. Kauppi P, Laitinen T, Ollikainen V, et al. The IL9R region contribution in asthma is supported by genetic association in an isolated population. *Eur J Hum Genet*. 2000;8(10):788. doi:10.1038/sj.ejhg.5200541. PMID:11039580.
- 91. Reader JR, Hyde DM, Schelegle ES, et al. Interleukin-9 induces mucous cell metaplasia independent of inflammation. *Am J Respir Cell Mol Biol.* 2003;28(6):664–672. doi:10.1165/rcmb.2002-0207OC. PMID:12760964.
- 92. Levitt RC, McLane MP, MacDonald D, et al. IL-9 pathway in asthma: new therapeutic targets for allergic inflammatory disorders. *J Allergy Clin Immunol*. 1999;103(5):S485–S491. PMID:10329852.
- 93. McLane MP, Haczku A, Van De Rijn M, et al. Interleukin-9 promotes allergen-induced eosinophilic inflammation and airway hyperresponsiveness in transgenic mice. *Am J Respir Cell Mol Biol.* 1998;19(5):713–720. PMID:9806735.
- 94. Sehra S, Yao W, Nguyen ET, et al. TH9 cells are required for tissue mast cell accumulation during allergic inflammation. *J Allergy Clin Immunol.* 2015;136(2):433–440. PMID:25746972.
- 95. Visekruna A, Ritter J, Scholz T, et al. Tc9 cells, a new subset of CD8+ T cells, support Th2-mediated airway inflammation. *Eur J Immunol.* 2013;43(3):606–618. PMID:23254356.
- 96. Xiao X, Balasubramanian S, Liu W, et al. OX40 signaling favors the induction of TH9 cells and airway inflammation. *Nat Immunol.* 2012;13(10):981–990. PMID:22842344.
- 97. Übel C, Sopel N, Graser A, et al. The activating protein 1 transcription factor basic leucine zipper transcription factor, ATF-like (BATF), regulates lymphocyte-and mast cell-driven immune responses in the setting of allergic asthma. *J Allergy Clin Immunol.* 2014;133(1):198–206. e9.
- 98. Cheng G, Arima M, Honda K, et al. Anti-interleukin-9 antibody treatment inhibits airway inflammation and hyperreactivity in mouse asthma model. *Am J Respir Crit Care Med.* 2002;166(3):409–416. PMID:12153980.
- 99. Steenwinckel V, Louahed J, Lemaire MM, et al. IL-9 promotes IL-13-dependent paneth cell hyperplasia



- and up-regulation of innate immunity mediators in intestinal mucosa. J Immunol. 2009;182(8):4737-4743. PMID:19342650.
- 100. Shimbara A, Christodoulopoulos P, Soussi-Gounni A, et al. IL-9 and its receptor in allergic and nonallergic lung disease: increased expression in asthma. J Allergy Clin Immunol. 2000;105(1):108-115. PMID:10629460.
- 101. Erpenbeck VJ, Hohlfeld JM, Volkmann B, et al. Segmental allergen challenge in patients with atopic asthma leads to increased IL-9 expression in bronchoalveolar lavage fluid lymphocytes. J Allergy Clin Immunol. 2003;111(6):1319-1327. PMID:12789235.
- 102. Nouri-Aria KT, Pilette C, Jacobson MR, et al. IL-9 and c-Kit+ mast cells in allergic rhinitis during seasonal allergen exposure: effect of immunotherapy. J Allergy Clin Immunol. 2005;116(1):73-79. PMID:15990777.
- 103. Blankenhaus B, Reitz M, Brenz Y, et al. Foxp3+ regulatory T cells delay expulsion of intestinal nematodes by suppression of IL-9-driven mast cell activation in BALB/c but not in C57BL/6 mice. PLoS Pathog. 2014;10(2):e1003913. PMID:24516385.
- 104. Grencis RK. Immunity to helminths: resistance, regulation, and susceptibility to gastrointestinal nematodes. Annu Rev Immunol. 2015;33:201-225. PMID:25533702.
- 105. Neurath MF, Kaplan MH. Th9 cells in immunity and immunopathological diseases. Semin Immunopathol. 2017;39(1):1-4.
- 106. Nowak EC, Noelle RJ. Interleukin-9 as a T helper type 17 cytokine. Immunology. 2010;131(2):169-173. PMID:20673237.
- 107. Tuxun T, Apaer S, Ma H-Z, et al. The potential role of Th9 cell related cytokine and transcription factors in patients with hepatic alveolar echinococcosis. J Immunol Res. 2015;2015:895416. PMID:26509179.
- 108. Carroll KN, Gebretsadik T, Escobar GJ, et al. Respiratory syncytial virus immunoprophylaxis in high-risk infants and development of childhood asthma. J Allergy Clin Immunol. 2017;139(1):66-71. e3. PMID:27212083.
- 109. Dodd JS, Lum E, Goulding J, et al. IL-9 regulates pathology during primary and memory responses to respiratory syncytial virus infection. J Immunol. 2009;183(11):7006-7013. PMID:19915054.
- 110. Grohmann U, Van Snick J, Campanile F, et al. IL-9 protects mice from Gram-negative bacterial shock: suppression of TNF- $\alpha$ , IL-12, and IFN- $\gamma$ , and induction of IL-10. *J Immunol.* 2000;164(8):4197–4203. PMID:10754315.
- 111. Becker K, Rösler B, Wang X, et al. Th2 and Th9 responses in patients with chronic mucocutaneous candidiasis and hyper-IgE syndrome. Clin Exp Allergy. 2016;46(12):1564-1574. PMID:27474157.
- 112. Licona-Limón P, Henao-Mejia J, Temann AU, et al. Th9 cells drive host immunity against gastrointestinal worm infection. Immunity. 2013;39(4):744-757. PMID:24138883.
- 113. Pang N, Zhang F, Ma X, et al. Th9/IL-9 profile in human echinococcosis: their involvement in immune response during infection by Echinococcus granulosus. Mediators Inflamm. 2014;2014:781649.
- 114. Licona-Limón P, Arias-Rojas A, Olguín-Martínez E. IL-9 and Th9 in parasite immunity. Semin Immunopathol. 2017;39(1):29-38.

- 115. Li L, Xie H, Wang M, et al. Characteristics of IL-9 induced by Schistosoma japonicum infection in C57BL/6 mouse liver. Sci Rep. 2017;7(1):2343.
- 116. Peng Y, Gao X, Yang J, et al. Chlamydial lung infection induces transient IL-9 production which is redundant for host defense against primary infection. PloS One. 2015;10(2):e0115195. PMID:25646821.
- 117. Christiaansen AF, Knudson CJ, Weiss KA, Varga SM. The CD4 T cell response to respiratory syncytial virus infection. Immunol Res. 2014;59(1-3):109-117. PMID:24838148.
- 118. Yu M, Long Q, Li H-H, et al. IL-9 inhibits viral replication in coxsackievirus B3-induced myocarditis. Front Immunol. 2016;7:409.
- 119. Becker K, Rösler B, Wang X, et al. Th2 and Th9 responses in patients with chronic mucocutaneous candidiasis and hyper-IgE syndrome. Clin Exp Allergy. 2016;46(12):1564-1574. PMID:27474157.
- 120. Cader MZ, Kaser A. Recent advances in inflammatory bowel disease: mucosal immune cells in intestinal inflammation. Gut. 2013;62(11):1653-1664. PMID:24104886.
- 121. Ley RE, Peterson DA, Gordon JI. Ecological and evolutionary forces shaping microbial diversity in the human intestine. Cell. 2006;124(4):837-848. PMID:16497592.
- 122. Atarashi K, Tanoue T, Shima T, et al. Induction of colonic regulatory T cells by indigenous Clostridium species. Science. 2011;331(6015):337-341. PMID:21205640.
- 123. Petersen ER, Claesson MH, Schmidt EGW, et al. Consumption of probiotics increases the effect of regulatory T cells in transfer colitis. Inflamm Bowel Dis. 2011;18(1):131-142. PMID:21495121.
- 124. Di Giacinto C, Marinaro M, Sanchez M, et al. Probiotics ameliorate recurrent Th1-mediated murine colitis by inducing IL-10 and IL-10-dependent TGF-βbearing regulatory cells. J Immunol. 2005;174(6):3237-3246. PMID:15749854.
- 125. Li Y, Innocentin S, Withers DR, et al. Exogenous stimuli maintain intraepithelial lymphocytes via aryl hydrocarbon receptor activation. Cell. 2011;147(3):629-640. PMID:21999944.
- 126. Devkota S, Wang Y, Musch MW, et al. Dietary-fatinduced taurocholic acid promotes pathobiont expansion and colitis in IL10-/- mice. *Nature*. 2012;487(7405):104. PMID:22722865.
- 127. Veldhoen M, Brucklacher-Waldert V. Dietary influences on intestinal immunity. Nat Rev Immunol. 2012;12(10):696. PMID:23007570.
- 128. Johansson ME, Larsson JMH, Hansson GC. The two mucus layers of colon are organized by the MUC2 mucin, whereas the outer layer is a legislator of hostmicrobial interactions. Proc Natl Acad Sci U S A. 2011;108(Supplement 1):4659-4665. PMID:20615996.
- 129. Jass J, Walsh M. Altered mucin expression in the gastrointestinal tract: a review. J Cell Mol Med. 2001;5(3):327-351. PMID:12067494.
- 130. Van der Sluis M, De Koning BA, De Bruijn AC, et al. Muc2-deficient mice spontaneously develop colitis, indicating that MUC2 is critical for colonic protection. Gastroenterol. 2006;131(1):117-129.
- 131. Gutierrez O, Pipaon C, Inohara N, et al. Induction of Nod2 in myelomonocytic and intestinal epithelial



- cells via nuclear factor- $\kappa$ B activation. *J Biol Chem.* 2002;277(44):41701–41705. PMID:12194982.
- Olszak T, An D, Zeissig S, et al. Microbial exposure during early life has persistent effects on natural killer T cell function. *Science*. 2012;336(6080):489–493. PMID:22442383.
- Lawson V. Turned on by danger: activation of CD1drestricted invariant natural killer T cells. *Immunology*. 2012;137(1):20–27. PMID:22734667.
- 134. Atreya R, Neurath MF. IBD pathogenesis in 2014: molecular pathways controlling barrier function in IBD. *Nat Rev Gastroenterol Hepatol.* 2015;12(2):67–68. PMID:25446731.
- 135. Kim H, Chung D. IL-9-producing invariant NKT cells protect against DSS-induced colitis in an IL-4-dependent manner. *Mucosal Immunol.* 2013;6(2):347–357. PMID:22892939.
- 136. Baumgart DC, Carding SR. Inflammatory bowel disease: cause and immunobiology. *Lancet*. 2007;369(9573):1627–1640. PMID:17499605.
- 137. Silverberg MS, Satsangi J, Ahmad T, et al. Toward an integrated clinical, molecular and serological classification of inflammatory bowel disease: report of a Working Party of the 2005 Montreal World Congress of Gastroenterology. *Can J Gastroenterol Hepatol.* 2005;19(Suppl A):5A–36A.
- 138. Singh UP, Singh NP, Murphy EA, et al. Chemokine and cytokine levels in inflammatory bowel disease patients. *Cytokine*. 2016;77:44–49. PMID:26520877.
- 139. Yuan A, Yang H, Qi H, et al. IL-9 antibody injection suppresses the inflammation in colitis mice. *Biochem Biophys Res Commun.* 2015;468(4):921–926. PMID:26603936.
- 140. Rahimian G, Sanei MH, Shirzad H, et al. Virulence factors of Helicobacter pylori vacA increase markedly gastric mucosal TGF-*β*1 mRNA expression in gastritis patients. *Microb Pathog.* 2014;67:1–7. PMID:24462401.
- 141. Bagheri N, Shirzad H, Elahi S, et al. Downregulated regulatory T cell function is associated with increased peptic ulcer in Helicobacter pylori-infection. *Microb Pathog.* 2017;110:165–175. PMID:28666843.
- 142. Salimzadeh L, Bagheri N, Zamanzad B, et al. Frequency of virulence factors in Helicobacter pylori-infected patients with gastritis. *Microb Pathog.* 2015;80:67–72. PMID:25656240.
- 143. Bagheri N, Azadegan-Dehkordi F, Rahimian G, Rafieian-Kopaei M, Shirzad H. Role of regulatory Tcells in different clinical expressions of helicobacter pylori infection. Arch Med Res. 2016;47(4):245–254. PMID:27664483.
- 144. Shirzad H, Bagheri N, Azadegan-Dehkordi F, et al. New insight to IL-23/IL-17 axis in Iranian infected adult

- patients with gastritis: effects of genes polymorphisms on expression of cytokines. *Acta Gastroenterol Belg.* 2015;78. PMID:26151690.
- 145. Bagheri N, Azadegan-Dehkordi F, Rahimian G, et al. Altered Th17 cytokine expression in Helicobacter pylori patients with TLR4 (D299G) polymorphism. *Immunol Invest.* 2016;45(2):161–171. PMID:26853914.
- 146. Bagheri N, Azadegan-Dehkordi F, Shirzad H, Rafieian-Kopaei M, Rahimian G, Razavi A. The biological functions of IL-17 in different clinical expressions of Helicobacter pylori-infection. *Microb Pathog.* 2015;81:33–38. PMID:25773771.
- 147. Bagheri N, Razavi A, Pourgheysari B, et al. Up-regulated Th17 cell function is associated with increased peptic ulcer disease in Helicobacter pylori-infection. *Infection, Genet Evol.* 2018.
- 148. Azadegan-Dehkordi F, Bagheri N, Shirzad M, et al. Correlation between mucosal IL-6 mRNA expression level and virulence factors of Helicobacter pylori in Iranian adult patients with chronic gastritis. *Jundishapur J Microbiol.* 2015;8(8).
- 149. Kobayashi T, Okamoto S, Hisamatsu T, et al. IL23 differentially regulates the Th1/Th17 balance in ulcerative colitis and Crohn's disease. *Gut.* 2008;57(12):1682–1689. PMID:18653729.
- 150. Brand S. Crohn's disease: Th1, Th17 or both? The change of a paradigm: new immunological and genetic insights implicate Th17 cells in the pathogenesis of Crohn's disease. *Gut.* 2009;58(8):1152–1167. PMID:19592695.
- 151. Brand S, Beigel F, Olszak T, et al. IL-22 is increased in active Crohn's disease and promotes proinflammatory gene expression and intestinal epithelial cell migration. *Am J Physiol Gastrointest Liver Physiol*. 2006;290(4):G827–G838. PMID:16537974.
- 152. Li H, Rostami A. IL-9: basic biology, signaling pathways in CD4+ T cells and implications for autoimmunity. *J Neuroimmune Pharmacol.* 2010;5(2):198–209. PMID:20020328.
- 153. Sarra M, Pallone F, MacDonald TT, Monteleone G. IL-23/IL-17 axis in IBD. *Inflamm Bowel Dis.* 2010;16(10):1808–1013. PMID:20222127.
- 154. Ouyang H, Shi Y, Liu Z, et al. Increased interleukin9 and CD4+ IL-9+ T cells in patients with systemic lupus erythematosus. *Mol Med Rep.* 2013;7(3):1031–1037. PMID:23291628.
- 155. Zundler S, Schillinger D, Fischer A, et al. Blockade of  $\alpha E \beta 7$  integrin suppresses accumulation of CD8+ and Th9 lymphocytes from patients with IBD in the inflamed gut in vivo. *Gut.* 2016;gutjnl-2016-312439.