

UNIVERSITY OF WESTMINSTER



**WestminsterResearch**

<http://www.wmin.ac.uk/westminsterresearch>

**Impaired phagocytic function of polymorpho-nuclear neutrophils in B chronic lymphocytic leukemia.**

**K. Gabunia<sup>1</sup>**  
**N. Gachechiladze<sup>1</sup>**  
**L. Burjanadze<sup>1</sup>**  
**T. Roschupkina<sup>1</sup>**  
**D. Baloyan<sup>1</sup>**  
**Lela Kardava<sup>1</sup>**  
**D. Ghirdaladze<sup>2</sup>**  
**G. Iosava<sup>2</sup>**  
**Andrew P. Jewell<sup>3</sup>**  
**Peter M. Lydyard<sup>4</sup>**  
**Nino Porakishvili<sup>1</sup>**

<sup>1</sup> Department of Immunology, Tbilisi State University, Georgia

<sup>2</sup> Institute of Haematology & Blood Transfusion, Tbilisi, Georgia

<sup>3</sup> School of Life Sciences, Kingston University, UK

<sup>4</sup> Department of Immunology, Royal Free & University College Medical School, London, UK

This is an electronic version of an article published in *Haematologica*, 85 (Supplement 7). pp. 28-29, July 2000. *Haematologica* is available online at:

<http://www.haematologica.org/free/venezia.pdf>

---

The WestminsterResearch online digital archive at the University of Westminster aims to make the research output of the University available to a wider audience. Copyright and Moral Rights remain with the authors and/or copyright owners. Users are permitted to download and/or print one copy for non-commercial private study or research. Further distribution and any use of material from within this archive for profit-making enterprises or for commercial gain is strictly forbidden.

---

Whilst further distribution of specific materials from within this archive is forbidden, you may freely distribute the URL of WestminsterResearch. (<http://www.wmin.ac.uk/westminsterresearch>).

In case of abuse or copyright appearing without permission e-mail [watts@wmin.ac.uk](mailto:watts@wmin.ac.uk).

# IMPAIRED PHAGOCYTOTIC FUNCTION OF POLYMORPHONUCLEAR NEUTROPHILS IN B CHRONIC LYMPHOCYTIC LEUKEMIA

Gabunia K,\* Gachechiladze N,\* Burjanadze L,\* Roschupkina T,\* Baloyan D,\* Kardava L,\* Ghirdaladze D,# Iosava G,# Jewell AP,° Lydyard PM,† Porakishvili N\*

\*Department of Immunology, Tbilisi State University; #Institute of Haematology and Blood Transfusiology, Tbilisi, Georgia;

†Department of Immunology, Royal Free and University College London Medical School; °School of Life Sciences, Kingston University, Surrey, UK

**Introduction.** The functional activity of polymorphonuclear neutrophils (PMNs) in patients with B-cell chronic lymphocyte leukemia (B-CLL) is still controversial due to the heterogeneity of the disease and individual variability, although normal absolute numbers of PMNs have been described. We have recently shown<sup>1</sup> a significant ( $p < 0.001$ ) decrease in the ability to release reactive oxygen intermediates in the PMA-induced nitroblue tetrazolium test (NBT) by PMNs of B-CLL patients ( $37.9 \pm 19.0\%$ , range 12-83%) compared with normal controls ( $81.5 \pm 12.7\%$ , range 59-94%). Both, the number of NBT<sup>+</sup> PMNs and the intensity of the formation of formazan crystals were decreased.

**Methods.** The detection of immature PMNs was measured by the activity of leukocyte alkaline phosphatase (LAP) using a Sigma LAP kit in 10 untreated Rai staged patients (age range 48-77) and 10 age-matched controls. PMNs were analyzed morphologically to determine banding. A possible overall decrease in enzymes, related to oxygen-dependent bactericidal function was evaluated by the expression of myeloperoxidase (MPO) in PMNs using a cytochemical assay (Sigma). The ability of PMNs to attach and engulf opsonized target cells was also tested. For this purpose, *Staphylococcus aureus* were opsonized with specific rabbit polyclonal IgG antibodies (Molecular Probes) and added to PMNs isolated on double Ficoll gradients. Cells were incubated for 30 minutes at 37°C, washed and stained with Giemsa. The numbers of PMNs with internalized and/or attached opsonized bacteria were evaluated per 200 PMNs.

**Results:** The intracellular expression of LAP by blood PMNs in B-CLL patients ( $67.7 \pm 13.2\%$ , range 52-81%) did not differ from that in healthy controls ( $71.7 \pm 15.2\%$ , range 50-85%), excluding immaturity of the PMNs as a factor. Morphologic analysis confirmed that there were very few band-type immature PMNs in the blood of B-CLL patients. The expression of MPO by B-CLL PMNs ( $94.3 \pm 1.5\%$ ) was no different from that of normal controls ( $92.7 \pm 1.9\%$ ), suggesting that the oxygen-dependent peroxidase system was not impaired. Although attachment of the opsonized particles was similar to that in controls, there was, nevertheless, a significant decrease in internalization by B-CLL PMNs (Table 1).

**Table 1. Internalization of opsonized *S.aureus*.**

Source of cells	PMN with or without <i>S. aureus</i> , %			
	Attached only	Attached and internalized	Internalized only	None
B-CLL patients	36.0±11.1	5.6±5.9	14.3±10.9	42.3±25.3
Controls	37.5±4.0	17.4±8.6	40.2±5.9	4.9±0.8
P	> 0.05	> 0.05	< 0.001	< 0.01

\*Mann-Whitney test (the data represent mean ± standard deviation).

*Conclusions.* Our data are consistent with a reduced phagocytic function of PMNs of B-CLL patients which might contribute to the increased susceptibility to infection in this disease.

## References

1. Gabunia Kh, Roschupkina T, Kardava L, et al. Decreased function of polymorphonuclear cells in B-cell chronic lymphocytic leukaemia is related to high white blood cell count. *Blood* 1999; 94:10 (Suppl 1) 40b.