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Role of Bile Acids in Colorectal Carcinogenesis

F.M. Nagengast, M.J.A.L. Grubben and I.P. van Munster

Dietary factors are considered important environmental risk determinants for colorectal cancer development. Epidemiological studies have shown that a high fat (or meat) intake is associated positively and a high starch, fibre (non-starch polysaccharide), vegetable and fruit intake negatively with colorectal cancer incidence. One mechanism by which these effects are possibly exerted is through the metabolism of secondary bile acids. Secondary bile acids are formed after enzymatic deconjugation and dehydroxylation of primary bile acids in the large bowel by anaerobic bacteria. It has been shown that these compounds can have tumour-promoting capacities in animal experiments. In epidemiological studies, colonic cancer risk is related to the faecal bile acid concentration. In serum and bile of patients with colonic adenomas, more deoxycholic acid was detected than in healthy controls. Secondary bile acids are toxic to several cell systems at physiological concentrations. The exact mechanism by which these amphiphilic molecules exert their action is not well understood. It might act through membrane damage, intracellular mitochondrial action or genotoxic effects. So far the evidence that bile acids are involved in colonic carcinogenesis is largely circumstantial. It is, however, well accepted that environmental factors, such as dietary habits influence genetic susceptibility. Bile acids could play a promoting role in this process.

Key words: bile acids and salts, cytotoxicity, cell proliferation, colonic carcinogenesis, dietary factors Eur J Cancer, Vol. 31A, Nos 7/8, pp. 1067–1070, 1995

INTRODUCTION

The role of dietary factors in colonic carcinogenesis THE INCIDENCE of colorectal cancer is high in Western countries and is related to dietary habits. Currently it is assumed that dietary factors modulate a genetic susceptibility. In epidemiological observations, the consumption of animal fat is positively related to the incidence of colon cancer [1, 2]. The intake of fibre is possibly negatively related to this incidence; however, many inconsistencies exist [3]. Migrant studies revealed that inhabitants moving from low-incidence to high-incidence areas acquired the colonic cancer risk of the region they moved to [4]. Within a given high-risk population, groups with different life styles have different colon cancer risks. Seventh Day Adventists in the U.S.A. have a lower incidence than the general population, probably because they consume a diet low in fat and high in fibre [5]. of these bile acids is even more increased. However, other studies in the U.S.A., U.K. and New Zealand have failed to demonstrate a correlation between high fat intake and colorectal cancer incidence [8]. Case-control studies have shown conflicting results in this respect [9].

Fat

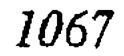
The hypothesis postulates that a high fat diet enhances the formation and degradation of bile acids and neutral sterols exerting a promoting effect in colonic carcinogenesis. Indeed, it has been found that dietary fat increases the output and faecal concentration of bile acids [6]. Epidemiological evidence has shown that populations with a high incidence of colorectal cancer and consuming a high fat and animal protein diet, excrete about twice the amount of secondary bile acids [7]. The concentration

Bile acid metabolism

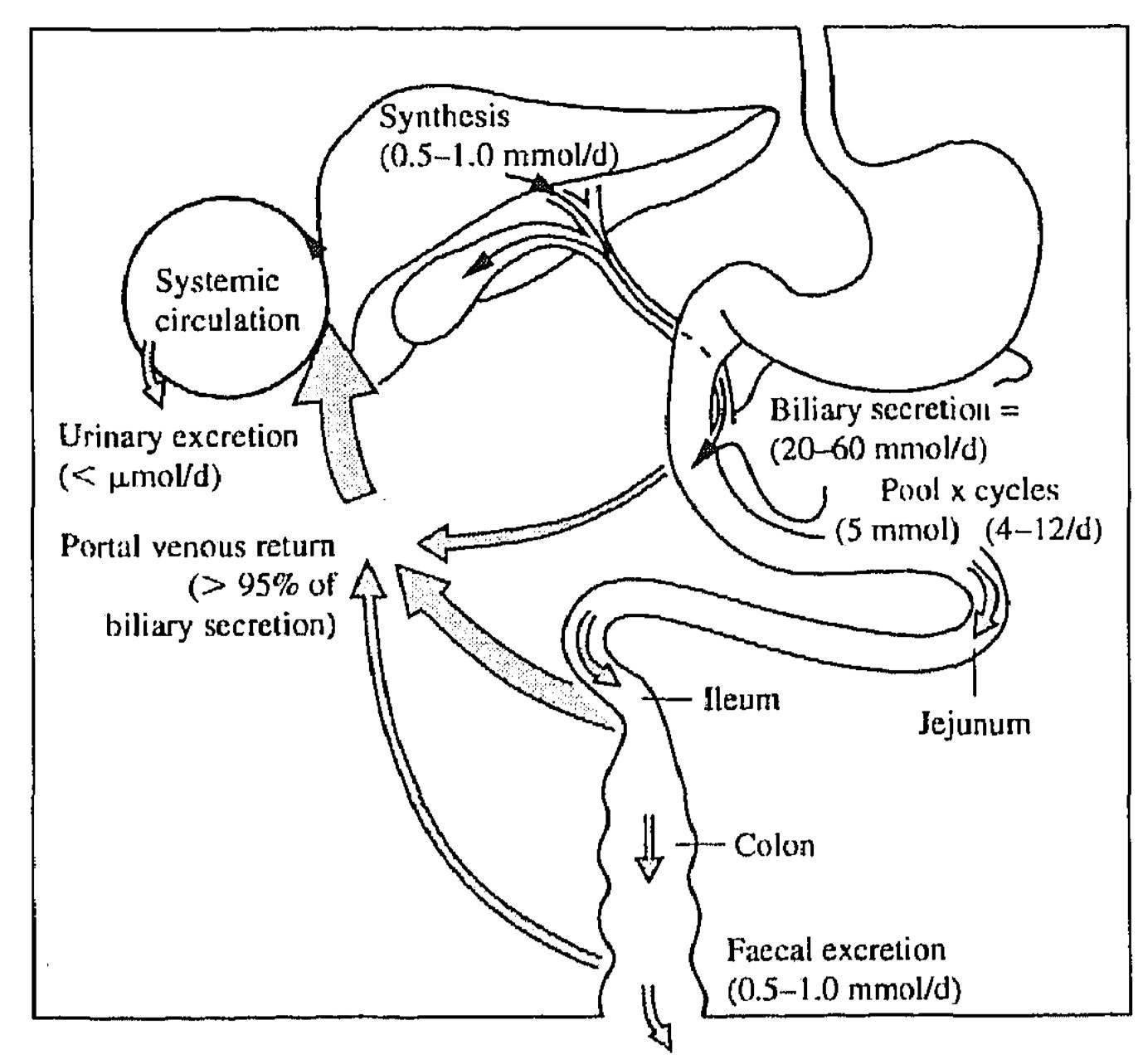
Bile acids are the major end products of cholesterol metabolism and are synthesised in the liver. The primary bile acids cholic acid (CA) and chenodeoxycholic acid (CDCA) are derived via several intermediate steps from cholesterol and secreted in bile as glycine or taurine conjugates.

They serve as cholesterol solubilising agents by the formation of micelles, and play an important role in the digestion and absorption of lipids in the small intestine. More than 95% of the bile acids passing through the ileum are reabsorbed and return to the liver through the portal vein. An efficient conservation in the so-called enterohepatic circulation is thus achieved. The proportion of bile acids not absorbed in the terminal ileum is 2-5% per cycle, and amounts to an average loss of 20% of the bile acid pool with 6-12 enterohepatic circulations per day. Bile acids that escape absorption in the ileum, are metabolised in the large bowel by the anaerobic bacterial flora. First, deconjugation takes place and the amino acid molecule on the carboxyl group is removed. Secondly, the primary bile acids CA and CDCA are dehydroxylated and converted into the secondary bile acids deoxycholic acid (DCA) and lithocholic acid (LCA), respectively. Further bacterial degradation in the large bowel and alterations in the liver produce the tertiary bile acids. DCA is partly absorbed in the colon and enters the enterohepatic circulation, where it is conjugated in the liver and secreted in

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Animal experiments

Secondary bile acids can act as tumour promoters in animal experiments, which are ideally performed in rodents. Because spontaneous colon cancer rarely occurs in rodents, initiating carcinogens such as azoxymethane have to be used [13]. Studies have been performed both by dietary manipulation (fat and fibre) and by direct application of bile acids to the colonic mucosa. Feeding high fat diets resulted in a higher tumour yield and an increased faecal bile acid concentration. Fibre addition has an opposite effect, although results were conflicting in this respect [13]. After diversion of bile ducts or small bowel resection more tumours can occur [22]. Direct installation of bile acids in the large bowel can be tumour promoting [23]. In one study, infusion of DCA led to damage of the mucosa thereby provoking an increased cell renewal. This was accomplished by increased cell proliferation which might be the key mechanism in the effect of bile acids in colonic carcinogenesis [24]. From this and other experiments, the concept has emerged that the concentration of soluble bile acids rather than the total faecal bile acid concentration determines possible cytotoxic effects of these molecules. The former is reflected by the concentration in the aqueous phase of the stool.

Figure 1. The enterohepatic circulation of bile acids in man.

bile; LCA is almost insoluble and very little is reabsorbed. Both secondary bile acids are excreted in the stool and make up to 95% of the total amount of excreted bile acids. In the stool, the major part of the bile acids are bound to dietary and bacterial residues. In the circulating bile acid pool, CA and CDCA each comprise about 30-40%, DCA about 20-30% and LCA less than 5% of the total amount [10]. Figure 1 shows the enterohepatic circulation of bile acids.

EXPERIMENTAL EVIDENCE

The hypothesis that the relationship between diet and colorectal cancer is established through bile acid metabolism has led to

Genotoxicity and mutagenicity

Secondary bile acids can have co-mutagenic effects as has been shown in the Ames test [25]. It has also been demonstrated that LCA can transform hamster embryo cells in culture. LCA can break DNA strands in cultured L1210 cells and enhance the activity of repair mechanisms after DNA strand breakage caused by 2-aminoanthracene [26, 27].

Both LCA and DCA stimulated the incorporation of tritiated thymidine in mouse liver and biliary tract epithelium, suggestive of enhanced cell proliferation [28]. Among components contributing to faecal mutagenicity are reactive glyceryl ethers, known as fecapentaenes. Their biosynthesis might be stimulated by bile salts [29]. In general conjugated bile salts have less or no genotoxic effects and unconjugated dihydroxy and monohydroxy bile salts are more genotoxic. However, it must be kept in mind that the most abundant monohydroxy bile acid in the human colonic lumen, LCA, is very poorly soluble in water.

many studies on cell systems, to experimental studies in animals (mostly rodents) and metabolic and interventional studies in humans.

Epidemiology

Colon cancer incidence is positively related to dietary fat intake and negatively to fibre and probably even more to total

Part of the discrepancy can be attributed to confounding potential of bile acids, there is also evidence of a direct stimufactors, such as age and dietary consumption, which were latory effect of several bile acids on proliferation. Bile salts not controlled for in these investigations. Faecal bile acid (DCA) can release prostaglandin E_2 (PGE₂) from colonic tissues. concentration proves to be age-dependent and inversely related The proliferative activity of colonic epithelial cells is among to dietary fibre intake [14]. In some studies bile acid kinetics or other things suppressed by PGE₂. Bile salts can enhance the biliary and serum bile acids were measured. The absorption of release of arachidonate from colonocytes and subsequently the DCA from the large bowel is also age-dependent [15, 16], higher synthesis of PGE_2 . This could be another explanation of the link in adenoma patients than in age-matched controls, and coincides between cell proliferation and bile acids [35]. A third effect with a more anaerobic environment [17, 18]. One study showed a higher biliary CDCA fraction in adenoma and carcinoma known as protein kinases. Protein kinase C appears to play a patients [19]. Recently it was demonstrated that adenoma paticritical role in tumour promotion and in the action of growth ents have a higher serum DCA concentration than healthy factors [36]. Bile acids might have a direct stimulatory effect on subclasses of these enzymes [37]. controls [20, 21].

Cytotoxicity of bile acids

Damaging effects of various bile acids on the colonic mucosa starch intake [11]. A high fat consumption leads to a higher bile have been described at the concentrations present in the aqueous acid excretion. Several epidemiological studies have shown that phase of stool [24]. Bile acids can disrupt the integrity of the cell the concentration of faecal bile acids is positively related to membrane of colonic mucosal cells [24, 30–32]. The increased cell loss will stimulate a compensatory cell renewal by increased colonic cancer incidence [12]. Case-control experiments, however, have shown conflicting results: in some a higher faecal bile mucosal proliferation. Thus, dietary manipulation resulting in a acid concentration was found in patients with adenomas or rise in colonic bile acid concentration can cause increased cancer, others found no difference between cases and controls mucosal proliferation [33, 34]. In addition to the attractive hypothesis that hyperproliferation is induced by the cytotoxic [13]. could be on a family of enzymes within the cell membrane

Bile Acids in Colorectal Carcinogenesis

Recently, a putative mechanism of hepatocyte necrosis was published. Toxic bile salts (in the liver GCDC) impair mitochondrial function, leading to an inhibition of oxidative phosphorylation and enhanced formation of toxic oxygen species by the mitochondrial respiratory chain. This results in oxidative stress and ATP depletion causing an increase in Ca²⁺ concentration with stimulation of hydrolases. This could lead to hydrolysis of lipid membranes and structural proteins causing cell death by necrosis [38].

We do not know whether this mechanism could be operative in colonocytes, since the type of cell and amount of cytotoxic bile salts differs considerably from the liver, but it remains an attractive hypothesis for cytotoxicity. Indirect evidence that this mechanism might also be applicable to cells other than hepatocytes, comes from experiments in which cytotoxicity of bile salts is determined in colon cancer cell lines (HT-29, CACO-2). In these cell lines cytotoxicity can be measured by the MTT assay, in which the mitochondrial function of the cell is tested. In viable cells MTT (3-(4,5-dimethylthizol-2-yl)-2,5-diphenyl tetrazolium bromide) is converted by the mitochondria to formazan, a blue dye, which can be detected in a fluorometer. With this assay, we and others have shown that the unconjugated dihydroxy bile acids DCA and CDCA are cytotoxic in a range that can be found in faecal water [32, 39]. Conjugated dihydroxy bile acids and cholic acid are not cytotoxic in this assay. So, as stated previously, the bile acid-induced increase in mucosal proliferation may be the key step in the association between bile acids and colon carcinogenesis. It has been demonstrated that a hyperproliferative colonic mucosa is more susceptible to carcinogens than a quiescent mucosa [40-42]. When proliferation is increased, the fraction of cells in Sphase (target cells) is relatively high, possibly resulting in an increased potency of intraluminal mutagenic substances. Large bowel neoplasms are associated with changes in proliferative characteristics, and in patients with colonic adenomas and cancer, an overall increased colonic mucosal proliferation has been demonstrated [43]. Furthermore, the proliferative compartment expands from the basal part of the crypts to the luminal surface. Similar changes in proliferative acitivity can be seen in patients with familial adenomatous polyposis, who are at high risk of developing colonic cancer. So, within the adenoma-carcinoma sequence, hyperproliferation might be a relatively early event leading towards an increased susceptibility to colonic cancer. Bile acids possibly play an important intermediate role in this process. It has to be kept in mind, however, that many other factors contribute to this cascade of events. Most intriguing, of course, are the successive genetic events that occur in the adenoma-carcinoma sequence [44]. In Figure 2, a hypothesis of colonic carcinogenesis is shown with special reference to the role of bile acids.

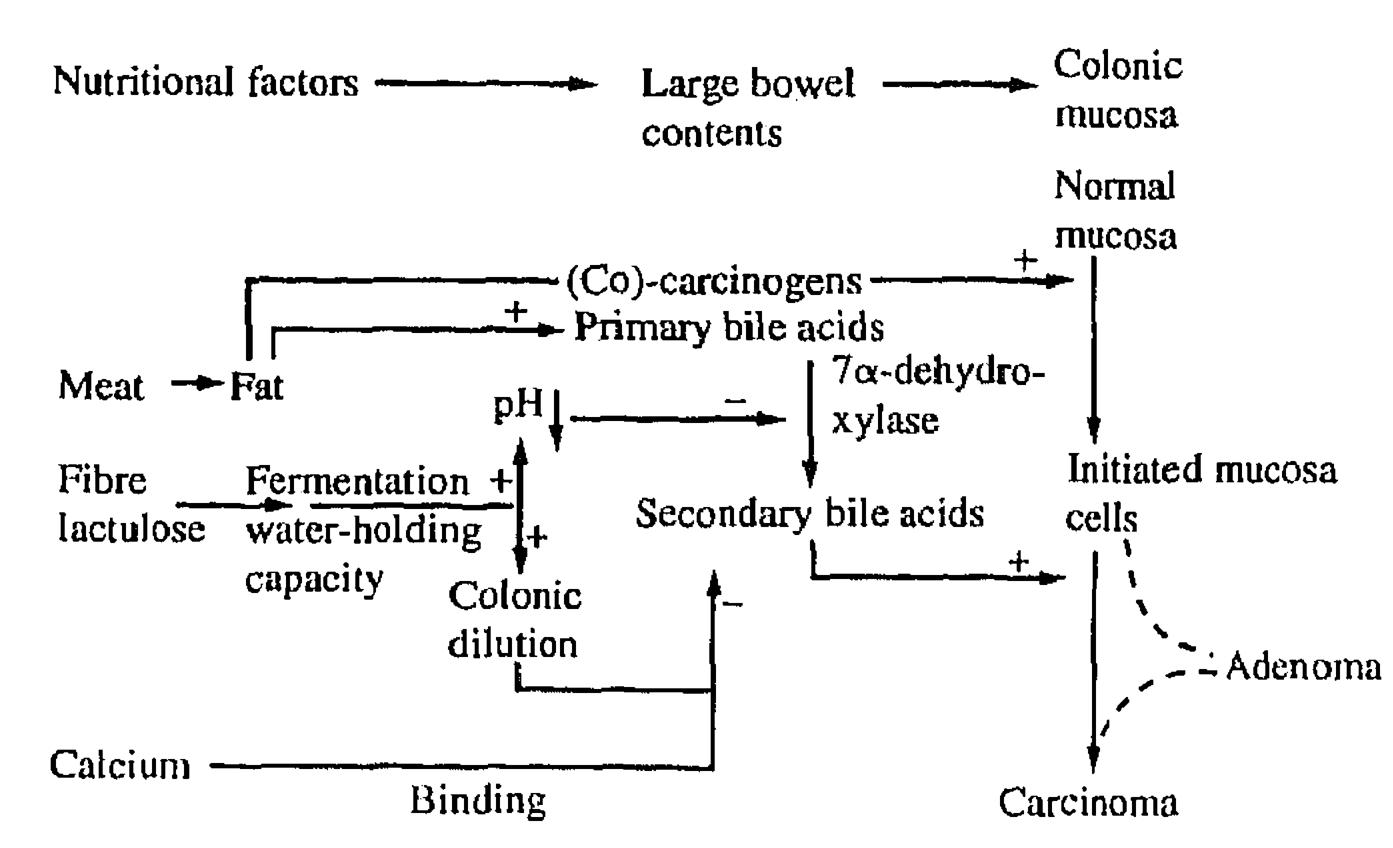


Figure 2. Hypothesis for the role of bile acids in colonic carcinogenesis.

is largely circumstantial and is derived from epidemiological and experimental studies in both animals and humans. Bile acids are probably cytotoxic to colonocytes and lead to a compensatory cell proliferation. The mechanism of cytotoxicity is not well understood, but can be attributed to membrane as well as intracellular effects. The increase in colonic cell proliferation is probably one of the key steps in the risk of development of colorectal cancer. Further studies are necessary to elucidate the mechanisms which are involved in the interaction of luminal events with the cascade of genetic changes that occur within the colonic mucosa during colonic carcinogenesis.

- 1. Enstrom JE. Colorectal cancer and consumption of beef and fat. Br *J Cancer* 1975, 32, 321–329.
- 2. Wynder EL, Shigematsu T. Environmental factors of cancer of the colon and rectum. Cancer 1967, 20, 1520–1561. 3. Trock B, Lanza E, Greenwald P. Dietary fiber, vegetables, and colon cancer: critical review and meta-analyses of the epidemiological evidence. *J Natl Cancer Inst* 1990, 8, 650–661. 4. Haenszel W, Kurihara M. Studies of Japanese migrants-I. Mortality from cancer and other diseases among Japanese in the United States. J Natl Cancer Inst 1968, 35, 291–297. 5. Phillips RL. Role of lifestyle and dietary habits in risk of cancer among Seventh-Day Adventists. Cancer Res 1975, 35, 3515–3522. 6. Cummings JH, Wiggins HS, Jenkins DJ, et al. Influence of diets high and low in animal fat on bowel habit, gastrointestinal transit time, fecal microflora, bile acid, and fat excretion. J clin Invest 1978, **61**, 953–963. 7. Reddy BS, Ekelund G, Bohe M. Metabolic epidemiology of colon cancer: dietary pattern and fecal sterol concentrations of three populations. *Nutr Cancer* 1983, 5, 34-40. 8. McMichael AJ, Potter JD, Hetzel BS. Time trends in colorectal cancer mortality in relation to food and alcohol consumption: United States, United Kingdom, Australia and New Zealand. Int J Epidemiol 1979, 8, 295–303. 9. Jain M, Cook GM, Davis FG, Grace MG, Howe GR, Miller AB. A case-control study of diet and colorectal cancer. Int J Cancer 1980, 26, 757-768. 10. Carey MC. The enterohepatic circulation. In Arias I, Popper H,

CONCLUSIONS

Bile acids (salts) are amphiphilic molecules synthesised in the liver from cholesterol. They play an important role in the solubility of cholesterol in bile and in the digestive process in

the small bowel through formation of micelles. An effective enterohepatic circulation keeps most of these bile acids within the body. During every cycle approximately 5% of the primary bile acids are lost into the large bowel. Here extensive degradation by the anaerobic flora occurs. The main events are deconjugation and dehydroxylation leading to the formation of unconjugated secondary bile acids. These last compounds have been incriminated in colonic carcinogenesis. Thus far, evidence EUC 31:7/8-0 Schachter D, Shafritz DA, eds. The Liver: Biology and Pathobiology. New York, Raven Press, 1982, 429–465.

- Cummings JH, Bingham SA, Heaton KW, Eastwood MA. Fecal weight; colon cancer risk, and dietary intake of nonstarch polysaccharides (dietary fiber). Gastroenterology 1992, 103, 1783-1789.
 Reddy BS, Weisburger JH, Wynder EL. Effects of high risk and low risk diets for colon carcinogenesis on fecal microflora and steroids in man. J Nutr 1975, 105, 878-884.
- 13. Nagengast FM. Bile acids and colonic carcinogenesis. Scand J Gastroenterol 1988, 23, 76-81.

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- 14. Nagengast FM, van der Werf SDJ, Lamers HLM, Hectors MPC, Buys WCAM, van Tongeren JHM. The influence of age, intestinal transit time and dietary composition on fecal bile acid profiles in healthy subjects. *Dig Dis Sci* 1988, 33, 673-678.
- 15. van der Werf SDJ, Huybregts AWM, Lamers HLM, van Berge Henegouwen GP, van Tongeren JHM. Age dependent differences in human bile acid metabolism and 7α-dehydroxylation. Eur J clin Invest 1981, 11, 425-431.
- 16. Salemans JMJI, Nagengast FM, Tangerman A, et al. Effect of aging on postprandial conjugated and unconjugated serum bile acid levels in healthy subjects. Eur J clin Invest 1993, 23, 192–198.
- 17. van der Werf SDJ, Nagengast FM, van Berge Henegouwen GP, Huybregts AWM, van Tongeren JHM. Colonic absorption of secondary bile acids in patients with adenomatous polyps and in matched controls. *Lancet* 1982, 1, 759-762.
- 18. van der Werf SDJ, Nagengast FM, van Berge, Henegouwen GP, Huybregts AWM, Tongeren JHM. Intracolonic environment and the presence of colonic adenomas in man. Gut 1983, 24, 876–880.
- 19. Moorehead RJ, Campbell GR, Donaldson DP, McKelvey STD. Relationship between duodenal bile acids and colorectal neoplasia. Gut 1987, 28, 1454–1459. 20. Bayerdorffer E, Mannes GA, Richter WO, et al. Increased serum deoxycholic acid levels in men with colorectal adenomas. Gastroen*terology* 1993, 104, 145–151. 21. Bayerdorffer E, Mannes GA, Ochsenkuhn T, Dirschedl P, Wiebecke B, Paumgartner G. Unconjugated secondary bile acids in the serum of patients with colorectal adenomas. Gut 1995, 36, 268-273. 22. Chomchai C, Bhadrachari N, Nigro ND. The effect of bile on the induction of experimental intestinal tumors in rats. Dis Colon *Rectum* 1974, 17, 310–312. 23. Narisawa T, Magadia NE, Weisburger JH, Wynder EL. Promoting effects of bile acids on colon carcinogenesis after intrarectal installation of MNNG in rats. *J Natl Cancer Inst* 1974, 53, 1093–1097. 24. Rafter JJ, Eng VW, Furrer R, Medline A, Bruce WR. Effects of calcium and pH on the mucosal damage produced by deoxycholic acid in the rat colon. Gut 1986, 27, 1320–1329. 25. Wilpart M, Roberfroid M. Effect of secondary bile acids on the mutagenicity of MNNG, 2AAF, and 2-nitrofluorene towards Salmonella typhimurium strains. Carcinogenesis 1986, 7, 703-706. 26. Kulkarni MS, Heidipreim PM, Yielding KM. Production by lithocholic acid of DNA strand breaks in L1210 cells. Cancer Res 1980, 40, 2666–2669.

- 29. Van Tassell RL, Macdonald DK, Wilkins TD. Stimulation of mutagen production in human feces by bile and bile acids. *Mutat Res* 1982, 103, 233-239.
- 30. Valhouny GV, Satchithanandram S, Lightfood F. Morphological disruption of colonic mucosa by free or cholestyramine-bound bile acids. Dig Dis Sci 1984, 29, 439-442.
- 31. Lapre JA, van der Meer R. Diet-induced increase of colonic bile acids stimulates lytic activity of fecal water and proliferation of colonic cells. *Carcinogenesis* 1992, 13, 41-44.
- 32. van Munster IP, Tangerman A, de Haan AFJ, Nagengast FM. A new method for the determination of bile acids and aqueous phase of stool: the effect of calcium. *Eur J clin Invest* 1993, 23, 773-777.
- 33. Deschner EE, Cohen BI, Raicht RF. Acute and chronic effect of dietary cholic acid on colonic epithelial cell proliferation. *Digestion* 1981, 21, 290-296.
- 34. Deschner EE, Raicht RF. Influence of bile on kinetic behaviours of colonic epithelial cells of the rat. *Digestion* 1979, **19**, 322-327.
- 35. DeRubertis FR, Craven PA, Saito R. Bile salt stimulation of colonic epithelial proliferation. Evidence for involvement of lipoxygenase

 Kanagalingam K, Strause E. Influence of lithocholic acid on 2aminoanthracene-induced alterations in DNA synthesis in rat tissues in vivo. Cancer Biochem Biophys 1980, 4, 159-166.
Bagheri SA, Bolt MS, Boyer JD, Palmer RH. Stimulation of thymidine incorporation in mouse liver and biliary tract epithelium by lithocholate and deoxycholate. Gastroenterology 1978, 74, 188-192.

- products. J clin Invest 1984, 74, 1614-1624.
- 36. Nishizuka Y. The role of protein kinase C in cell surface signal transduction and tumor promotion. *Nature* 1984, 308, 693-698.
- 37. Craven PA, Pfanstiel J, DeRubertis FR. Role of activation of protein kinase C in the stimulation of colonic epithelial proliferation and reactive oxygen formation by bile acids. J clin Invest 1987, 79, 532-541.
- 38. Rosser BG, Gores GJ. Liver cell necrosis: mechanisms and clinical implications. *Gastroenterology* 1995, **108**, 252–272.
- 39. Latta RK, Fiander H, Ross NW, Simpson C, Schneider H. Toxicity of bile acids to colon cancer cell lines. Cancer Lett 1993, 70, 167–173.
- 40. Barthold SW, Beck D. Modification of early dimethylhydrazine carcinogenesis by colonic mucosal hyperplasia. Cancer Res 1980, 40, 4451-4455.
- 41. Deschner EE, Long FC, Hakissan M, Herrman SL. Differential susceptibility of AKR, C57BL/6J, and CF1 mice to 1,2-dimethylhy-drazine-induced colonic tumor formation predicted by proliferative characteristics of colonic epithelial cells. J Natl Cancer Inst 1983, 70, 279-282.
- 42. Deschner EE, Long FC, Hakissian M, Cupo SH. Differential susceptibility of inbred mouse strains forecast by acute colonic proliferative response to methylazoxymethanol. J Natl Cancer Inst 1984, 72, 195–198.
- 43. Terpstra OT, van-Blankenstein M, Dees J, Eilers GA. Abnormal pattern of cell proliferation in the entire colonic mucosa of patients with colon adenoma or cancer. *Gastroentrology* 1987, 92, 704–708.
- 44. Fearon ER, Vogelstein B. A genetic model for colorectal tumorigenesis. Cell 1990, 61, 759-767.

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