

Contents lists available at ScienceDirect

Autoimmunity Reviews

journal homepage: www.elsevier.com/locate/autrev

1 Review

Q1 The role of microRNAs in the pathogenesis of autoimmune diseases

Q2 Ji-Qing Chen ^a, Gábor Papp ^a, Péter Szodoray ^b, Margit Zeher ^{a,*}

4 ^a Division of Clinical Immunology, Faculty of Medicine, University of Debrecen, Móricz Zs. str. 22, H-4032 Debrecen, Hungary

5 ^b Centre for Immune Regulation, Department of Immunology, University of Oslo, Oslo University Hospital, Rikshospitalet, Oslo, Norway

6

7 A R T I C L E I N F O

8 *Article history:*
9 Received 6 July 2016
10 Accepted 10 July 2016
11 Available online xxx

12 *Keywords:*

26 MicroRNAs (miRs)
27 Systemic lupus erythematosus (SLE)
28 Primary Sjögren's syndrome (SS)
29 Rheumatoid arthritis (RA)
30 Systemic sclerosis (SSc)
31 Multiple sclerosis (MS)
32 Psoriasis

33 *34*

36

37 *38* Contents

40	1. Introduction	0
41	2. The biology of miRNAs	0
42	3. miRNAs in immune system	0
43	3.1. Innate immunity	0
44	3.2. Adaptive immunity	0
45	4. MiRNAs in autoimmune diseases	0
46	4.1. Systemic lupus erythematosus	0
47	4.2. Primary Sjögren's syndrome	0
48	4.3. Rheumatoid arthritis	0
49	4.4. Systemic sclerosis	0
50	4.5. Multiple sclerosis	0
51	4.6. Psoriasis	0
52	5. Conclusions and future perspectives	0
53	Take-home messages	0
54	Conflict of interest	0
55	Acknowledgments	0
56	References	0

57

58 1. Introduction

59 MicroRNAs (miRNAs) constitute a recently discovered family of
60 small RNAs, ranging from 18 to 25 nucleotides in length. They are

single-stranded, endogenous non-coding RNAs playing critical roles in
61 regulating gene expression [1,2]. 62

63 miRNAs regulate approximately 90% of protein-coding genes, and
64 play a central role in various biological processes including immune
65 cell lineage commitment, differentiation, proliferation, apoptosis and
66 maintenance of immune homeostasis. It is not surprising that
67 alterations in the expression of miRNAs potentially contribute to the

* Corresponding author.
E-mail address: zeher@iibel.dote.hu (M. Zeher).

development of certain pathological conditions and clinical disorders. Nowadays, the pathogenetical role of miRNAs is most intensively studied in malignant diseases as well as autoimmune conditions. Changes in miRNA expression profiles have been identified in different autoimmune diseases such as systemic lupus erythematosus (SLE), rheumatoid arthritis (RA) and Sjögren's syndrome (SS) [3–5]. In this review, we summarize the characteristics of miRNAs and their functional role in the immune system and autoimmune diseases including SLE, primary SS, RA, systemic sclerosis (SSc), multiple sclerosis (MS) and psoriasis.

2. The biology of miRNAs

The majority of miRNA genes derived from the intergenic regions or in oriented antisense to form independent transcription units. Most of the others reside in the intron region of protein-coding genes [6]. Human miRNAs are not always genomically isolated; sometimes several miRNAs are assembled as clusters for further transcription and expression [7].

The miRNA biogenesis and maturation occur first in the nucleus and then in the cytoplasm with the help of several proteins and enzymes (Fig. 1). The first step in the miRNA biogenesis is the generation of primary miRNA transcripts (pri-miRNAs) from DNA molecules in the nucleus of the cell. Most miRNA genes are transcribed by RNA polymerase II to produce a few hundred to thousand nucleotide-long pri-miRNA [6]. The pri-miRNAs are both capped and polyadenylated with a typical hairpin structure [8]. These pri-miRNAs are recognized by an enzyme-protein complex and further cleaved into 70–100 nucleotide-long precursor miRNA (pre-miRNA). This complex is composed of Drosha and DiGeorge syndrome critical region gene 8 (DGCR8) and denoted as microprocessor complex [9]. Drosha is one of the two members of the RNase III family while DGCR8 is the double-stranded RNA-binding protein which is deleted in DiGeorge syndrome [10]. The pre-miRNA then exported to cytoplasm through exportin 5, which is a member of the karyopherin family of nucleocytoplasmic proteins. The exportin 5 recognizes a two-nucleotide overhang left by Drosha at the 3' end of the pre-miRNA hairpin, requiring the GTP-bound form of the Ran GTPase for providing energy [11].

The noncanonical miRNA biogenesis pathway bypasses the microprocessor complex cleavage processing for another sort of pre-miRNAs, known as mirtrons, which directly spliced out of introns by spliceosome. The branched pre-mirtrons then undergo lariat-mediated debranching to mimic the structural features of pre-miRNAs [12,13]. Interestingly, mirtrons can not only be found in *Caenorhabditis elegans* and *Drosophila*, but also reported in mammals [14].

The pre-miRNAs have further processing to yield mature miRNA in the cytoplasm. The second member of the RNase III family named Dicer interacts with both 5' and 3' ends of the pre-miRNA and cleaves the hairpin loop, processing to a 19–25 nucleotides miRNA/miRNA* duplex [15,16]. The miRNA* was regarded as passenger strand since it is less-stable, while the miRNA as guide strand. The miRNA/miRNA* duplex releases the helix structure after loaded into the argonaute (Ago) proteins. The guide strand remains the interaction with Ago to generate the RNA-induced silencing complex (RISC), which facilitate miRNAs binds to their targets [17]. The passenger strand as complementary strand of the guide strand is degraded as a RISC complex substrate. However recent study demonstrates that several miRNA* are stably expressed and may play an important role, as well [18].

The mature miRNA interacts with the 3'-UTR of specific messenger RNA (mRNA) to regulate gene expression. Target mRNA is recognized by the 2–7 nucleotides of the 'seed' region of the miRNA [19]. The complementary degree of the base pairing between the miRNA seed region and mRNA defines the mechanism of gene regulation [20]. When the complementary base pairing is perfect or near-perfect, Ago protein of the RISC complex induces the endonucleotic cleavage of the target mRNA resulting in deadenylation and degradation of mRNA fragments.

When the base pairing is incomplete, the formation of double-stranded RNA, resulting from the binding of miRNA, leads to translational repression [2,21,22]. Repressed mRNAs aggregate in cytoplasmic foci called P-bodies, which are known sites of mRNA destabilization [23,24].

3. miRNAs in immune system

The miRNAs play critical roles not only in the development of immune system but also the regulation of both innate and adaptive immunity [5,25]. MiRNAs function as translational repressors during stem cell fate and differentiation [26]. MiR-181, miR-223 and miR-142s are strongly expressed in hematopoietic cells and shown regulatory roles during hematopoietic lineage differentiation [27,28].

3.1. Innate immunity

The innate immune system is the first line of host defense and important in mechanisms against invading microorganisms; moreover, it forms the basis of the development of adaptive immunity. Host cells express diverse pattern recognition receptors (PRRs), including toll-like receptors (TLRs), C-type lectin-like receptors (CLRs), retinoic acid-inducible gene (RIG)-I-like-receptors (RLRs) and nucleotide-binding oligomerization domain (NOD)-like receptors (NLRs). These can recognize a wide range of pathogen-associated molecular patterns (PAMPs). These mechanisms trigger the intracellular signaling pathways, which results in releasing of proinflammatory cytokines, chemokines, and interferons (IFNs), as well as lead to the expression of co-stimulatory molecules [29]. TLRs are the most characterized PRRs, which are capable of potentially activating different cell types, which could be highly expressed on most immune cells [30]. Their downstream signaling pathways lead to the production of a wide range of immune-stimulatory cytokines and chemokines. Aberrant activation of TLRs may result in unrestricted inflammatory responses therefore the family of TLRs may play a pivotal role in the development of autoimmune diseases [31]. Among all ten TLR subtypes, TLR1, TLR2, TLR4, TLR5, TLR6 and TLR10 are generally regarded as extracellular receptors, while the family of TLR3, TLR7, TLR8 and TLR9 are intracellular receptors located in endosomal compartments and responsible for the recognition of nucleic acids derived from viruses, bacteria and the host [32–35]. TLR4 can recognize lipopolysaccharides (LPSs), which is the typical endotoxin for gram-negative bacteria. The LPS-mediated inflammatory responses consequently induce overexpression of miR-146a/b, miR-132 and miR-155. Upregulation of miR-146 leads to translational repression of its target genes interleukin-1 receptor-associated kinase (IRAK) 1 and tumor necrosis factors receptor associated factor (TRAF) 6 [36]. miR-146 was recognized as a negative regulator of RLRs in the *in vitro* model of mouse macrophages through targeting IRAK1, IRAK2 and TRAF6 [37]. Exposure to LPS stimulates tumor necrosis factors (TNF)- α secretion. Overexpression of miR-155 and lower expression of miR-125b may relate with elevated level of TNF- α . It was indicated that miR-155 targets transcript coding gene for several proteins enhancing TNF- α translation, including Fas-associated death domain protein (FADD), I κ B kinase epsilon (IKKepsilon) and TNFR superfamily-interacting serine-threonine kinase 1 (Ripk1), while miR-125b targets the 3'-UTR of TNF- α transcripts [38]. In miR-147 knockout mice, increased inflammatory cytokine expression found in macrophages upon TLR stimulation such as ligands to TLR2, TLR3 and TLR4. Thus miR-147 was regarded as a negative regulator in TLR-activated inflammatory responses [39]. The miR-1303 production is also regulated by the NF- κ B pathway. A recent study revealed negative regulation of mycobacteria-induced Atg2B protein production related with autophagy process [40].

The miR-146a and miR-155 influence IFN-type I synthesis in plasmacytoid dendritic cells mediated by TLR-7 and TLR-9, while in T and B cells, group of miRNAs including miR-21, miR-126, miR-146a, miR-155, miR-1246 and others might correlate with epigenetic

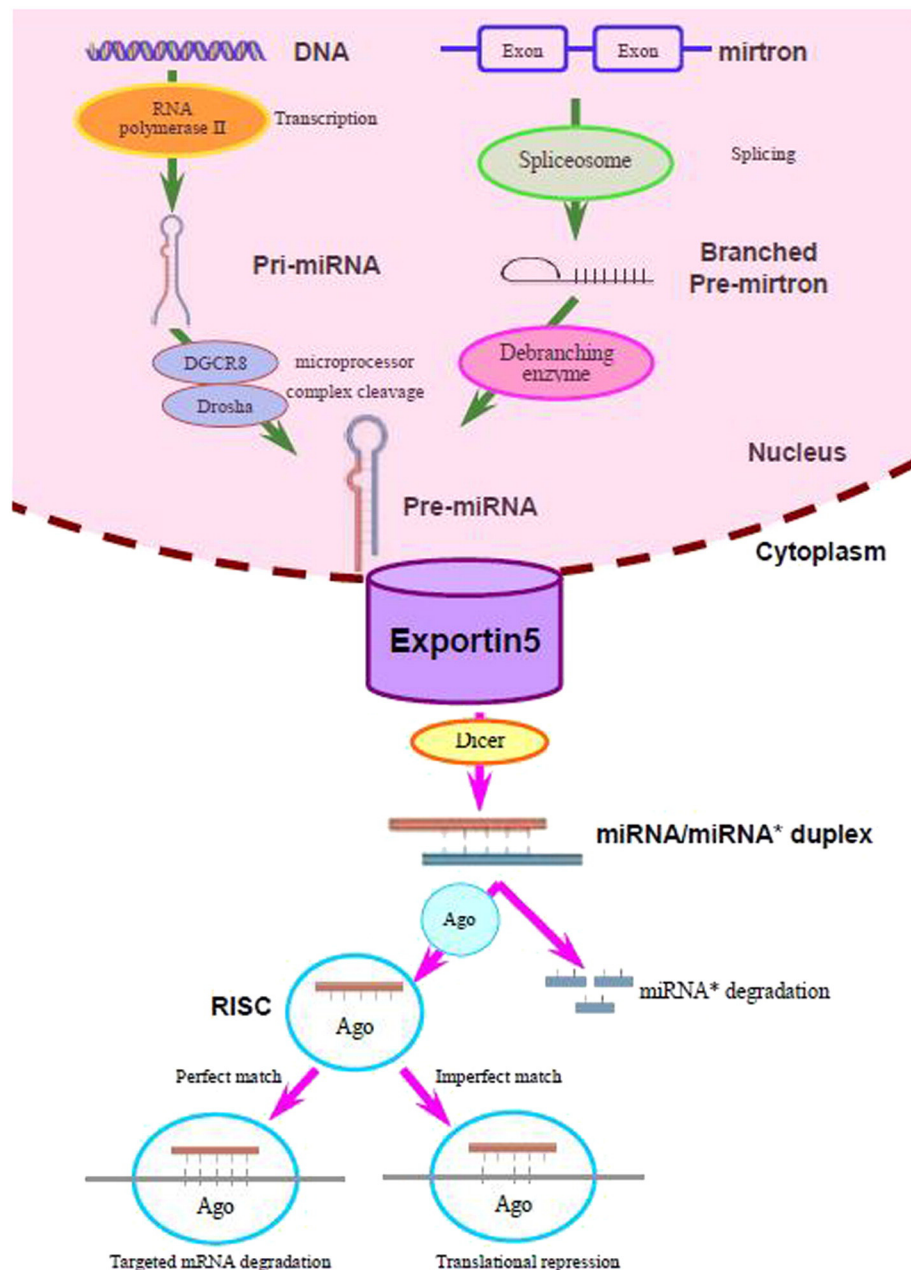


Fig. 1. microRNA biogenesis and mechanisms of action. Most miRNA are transcribed from genomic DNA by RNA polymerase II to generate typical hairpin structured primary miRNA transcripts (pri-miRNAs). These pri-miRNAs are recognized by the microprocessor complex (Drosha and DGCR8) and further cleaved into precursor miRNA (pre-miRNA). The noncanonical miRNA biogenesis pathway starts from mirtrons, which directly spliced out of introns by spliceosome. The branched pre-mirtrons then undergo lariat-mediated debranching to mimic the structural features of pre-miRNAs. The pre-miRNA then exported to cytoplasm through exportin 5. The pre-miRNA is further cleaved by Dicer into miRNA/miRNA* duplex. The guide strand loaded into the argonaute (Ago) proteins to generate the RNA-induced silencing complex (RISC), while the passenger strand (miRNA*) would eventually degrade. The perfect complementary base pairing match between miRNA and target messenger RNA (mRNA) induces target mRNA degradation, while the imperfect match results in the repression of the mRNA translation.

193 modifications, support abnormal cytosine release, differentiation of cell
194 subsets, B cell hyperactivity and autoantibody production [41].

195 3.2. Adaptive immunity

196 The adaptive immune system involved both T and B lymphocytes as
197 major cellular components. One of the RNase III family enzymes, Dicer
198 as mentioned previously is important in the biogenesis of miRNA. In
199 the early stage of T cell development, depletion of Dicer leads to reduction
200 of T cell numbers both in the thymus and peripheral lymphoid organs
201 [42]. Dicer-deficient T helper (Th) cells show aberrant cytokine
202 secretion, such as increased expression of IFN- γ in the absence of exogenous
203 cytokines and blocking antibodies [43]. In early B cell progenitors,

204 depletion of Dicer results in blocking at the pro- to pre-B cell transition
205 since miR-17 mostly target the genes that upregulated in Dicer-
206 deficient pro-B cells [44].

207 Interleukin (IL)-17 produced by Th17 cells are closely related to
208 miR-326 and miR-155. It is shown that overexpression of miR-326 results
209 in increased number of Th17 cells through targeting Ets-1 in multiple
210 sclerosis patients and severe experimental autoimmune encephalomyelitis (EAE)
211 mice [45]. MiR-155 on the other hand is essential for dendritic cell
212 production of cytokines which induce Th17 cell formation. MiR-155
213 knock-out mice are recognized resistant to EAE [46]. MiR-155 is
214 down-regulated in human monocyte-derived dendritic cells in response
215 to LPS-induced inflammatory processes [47]. MiR-155 expression is
216 necessary for maintaining regulatory T (Treg) cell

Table 1
Differential expression of miRNAs in autoimmune diseases.

Disease	Sample	miRNA expression	
		Up-regulated	Down-regulated
SLE	PBMCs	miR-516a-3p [54]	miR-126 [53]
		miR-525-5p [55]	miR-17-5p [55]
		miR-629	miR-112
		miR-21 [55]	miR-141
		miR-61	miR-184
		miR-78	miR-196a
		miR-142-3p	miR-383
		miR-189	miR-409-3p
		miR-198	miR-146a [57]
		miR-298	miR-155 [59]
		miR-299-3p	
		miR-342	
		miR-410 [60]	
			miR-26a [61]
APS	Urinary exosomes	miR-146a-3p [66]	
		miR-146a-5p	
		miR-155	
		miR-210	
		miR-146a-3p [66]	
		miR-146a-5p	
		miR-155	
		miR-210	
		hsa-miR-768-3p [71]	hsa-miR-574 [71]
		miR-16 [69]	
		miR-200b-3p	
		miR-223	miR200b-5p [69]
		miR-483-5p	
		miR-146a/b [72]	
		miR-155 [74]	miR-155 [76]
miR-181a [77]			
miR-34b-3p [78]			
miR-300			
miR-609			
miR-877-3p			
miR-3162-3p			
miR-4701-5p			
miR-146a [80]			
miR-155 [81]			
miR-301a-3p [82]	miR-16 [82]		
miR-223 [82]	miR-146a [82]		
	miR-155 [82]		
	miR-188-5p [85]		
miR-16 [83]			
miR-132 [83]			
miR-146a [83]			
miR-223 [83]			
miR-146a [84]			
miR-155 [84]			
miR-223 [84]			
miR-146a [87]	miR-363 [87]		
	miR-498 [87]		
miR-223 [88]	miR-99a [88]		
	miR-100 [88]		
	miR-125b [88]		
	miR-199-3p [88]		
	miR-199-5p [88]		
	miR-152 [88]		
	miR-214 [88]		
miR-21 [99-105]	miR-15b [91-98]		
miR-92a [99-105]	miR-16 [91-98]		
miR-133 [99-105]	miR-27a/b [91-98]		
miR-142-3p [99-105]	miR-132 [91-98]		
miR-200a/b [99-105]	miR-150 [91-98]		
miR-590 [99-105]	miR-335 [91-98]		
	miR-29a [106,107]		
	miR-135b [108]		
	miR-193b [109]		
miR-21 [111,112]	miR-214 [114]		
miR-146a/b [111,112]	miR-140-5p [115]		
miR-155 [111,112]	miR-572 [116]		
miR-326 [113]			
miR-27a [114]			
miR-150 [117]			

Table 1 (continued)

Disease	Sample	miRNA expression	
		Up-regulated	Down-regulated
Psoriasis	PBMCs	miR-142-3p [119]	miR-99a [119]
		miR-146a [119]	miR-125b [119]
		miR-155 [119]	miR-181a [119]
		miR-224 [119]	
		miR-378 [119]	
	Serum	miR-146a [120]	
	Th17	miR-223 [119]	miR-193b [119]
	Hair shaft	miR-424 [121]	
	Lesional skin	miR-26b-5p [122]	

Abbreviations
 SLE: systemic lupus erythematosus.
 APS: antiphospholipid syndrome.
 SS: Sjögren's syndrome.
 RA: rheumatoid arthritis.
 SSc: systemic sclerosis.
 MS: multiple sclerosis.
 PBMCs: peripheral blood mononuclear cells.
 MSGs: minor salivary glands.
 SGECs: salivary gland epithelial cells.
 FFPE: formalin-fixed paraffin-embedded synovial tissue.
 CSF: cerebral spinal fluid.
 Th: T helper cell.

proliferative activity under Foxp3 regulation in controlling the IL-2 signaling pathway by targeting the suppressor of cytokine signaling (SOCS) 1 [48]. Like miR-155, miR-146a is not only relevant to the innate immune system but also critical in the adaptive immune system. Overexpression of miR-146a was found in Treg cells as a response to activation of signal transducer and activator transcription (STAT) 1. The negative regulator of STAT1 phosphorylation downstream of the IFN-γ receptor is SOCS1, which additionally associated with Th1-mediated autoimmunity [49]. In activated B cells, miR-181b results in the down-regulation of activation-induced cytidine deaminase (AID) mRNA and protein levels. By restricting AID activity, miR-181b may prevent B cell malignant transformation [50].

The overexpression of miR-148a results in impaired B cell tolerance, which accelerates the development of autoimmune diseases. Moreover, miR-148a inhibits the expression of the autoimmune suppressor Gadd45α, the tumor suppressor phosphatase and tensin homolog (PTEN) and the pro-apoptotic protein Bim and protects immature B cells from apoptosis induced by engagement of B cell antigen receptor [51].

4. MiRNAs in autoimmune diseases

Alterations in miRNA regulation seem to be highly related to the development of immune dysfunctions and autoimmunity. Recently several studies have focused on the role of miRNAs in autoimmune diseases and different expression profiles have been identified as biomarkers of certain autoimmune conditions, such as SLE, RA and SS. Table 1 summarized the differential expression of miRNAs in autoimmune diseases.

4.1. Systemic lupus erythematosus

SLE is one of the most prevalent systemic autoimmune disorders. SLE has a large spectrum of clinical presentations since the disease can affect multiple organs, including skin, joints, kidneys, lungs, nervous system, and serous membranes. The diversity of its clinical features is matched by the complexity of pathogenic factors including genetic, hormonal, and environmental factors [52].

A recent study demonstrated that blood plasma level of miRNA-126 was significantly lower in SLE patients compared to that in normal controls. In addition, both plasma levels of IFN-α and interferon-inducible gene ISG56 mRNA in peripheral blood mononuclear cells (PBMCs) showed higher levels in SLE patients compared to controls. Based on

these observations, miRNA-126 may inhibit the production of IFN- α and decrease in its expression level is possibly involved in the pathogenesis of SLE [53].

Zhu et al. reported high expression levels of miRNA-516a-3p, miRNA-629 and miRNA-525-5p in the PBMCs of paediatric SLE (pSLE) patients compared to healthy children. In addition, the increased expression levels of these three miRNAs were positively correlated with the SLEDAI scores and CRP levels. The target genes of these three miRNAs, namely Yinyang1 (YY1), Kruppel-like factor 13 (KLF13) and interferon regulatory factor 5 (IRF5), were found to be important in the pathogenesis of pSLE [54]. Dai et al. indicated 16 miRNAs with altered expression pattern in PBMCs, based on a microarray analysis involving 23 SLE patients from Han population, as following: seven miRNAs are decreased expression in SLE: miR-17-5p, miR-112, miR-141, miR-184, miR-196a, miR-383, and miR-409-3p; the other nine miRNAs are overexpressed in SLE: miR-21, miR-61, miR-78, miR-142-3p, miR-189, miR-198, miR-298, miR-299-3p, and miR-342 [55]. Two years later, they analyzed miRNAs in kidney biopsy samples of class II lupus nephritis (LN) patients, compared to renal tumor patients' kidney resection samples. They reported 66 miRNAs differentially regulated in lupus nephritis patients. Among them, 36 are up-regulated and the rest 30 are down-regulated [56].

The downregulation of miR-146a also contributes to the development of SLE. It was revealed that miR-146a is a negative regulator of type I IFN pathway by targeting IFN regulatory factor 5, STAT 1, IRAK1 and TRAF6 [57]. Two recent studies focused on miR-155; the miR-155 expression level correlated negatively with the expression of CD1d in B cells of SLE mice. Additionally, it was found that lower expression level of CD1d on B cells was decreased by targeting Ets-1 through activation of TLR9. Moreover, in juvenile SLE patients, miR-155 is downregulated in PBMCs compared to that of healthy controls. It was reported that miR-155 expression level was negatively correlated with Systemic Lupus Erythematosus Disease Activity Index (SLEDAI) score [58,59].

It is observed that up-regulation of miR-410 significantly reduced the expression levels of fibrosis factors such as transforming growth factor- β 1 (TGF- β 1) by inhibiting secretion of IL-6 in the pathogenesis of LN [60]. The epigenetic modulator EZH2 might shift implicating effector in lupus naïve CD4 + T cells and opposes inhibitory TGF- β signaling. The expression level of miR-26a, which is sensitive to glucose availability and targets EZH2, correlated negatively with SLEDAI [61].

A current study indicated that miR-148a-3p expression level was significantly higher in blood serum and glomerular cells in SLE with active LN. Up-regulation of miR-148a-3p accelerated glomerular cell proliferation and proliferating cell nuclear antigen (PCNA) expression, consequently reducing the PTEN expression level [62]. The significant overexpression of miR-130b-3p was demonstrated in serum of SLE patients with early stage LN, compared with that measured in healthy controls. Serum miR-130b-3p did not affect SLE disease activity (SLEDAI, ds-DNA, and complements levels) but correlated with renal damage since the expression of serum miR-130b-3p correlated positively with 24-h proteinuria and chronicity index (histological chronicity index and glomerular sclerosis) [63]. On the other hand, miR-29c expression in urinary exosomes showed a strong negative correlation with the chronicity but not with renal function (eGFR and creatinine levels). Urinary exosomes are micro-vesicles released by the epithelial cell facing the urinary space and proposed a novel and ideal source of markers for evaluating stage of LN [64]. Furthermore, expressions of several miRNAs were elevated in the urinary exosome fraction compared to the cell-free and exosome-depleted supernatant fraction, especially with LN. Among the exosomal miRNAs, miR-146a was the most overexpressed in SLE patients with active LN compared to the control group or to the SLE patients in the absence of LN [65].

Circulating antiphospholipid antibodies (aPLs) increase the risk of pregnancy complications, which leads to an autoimmune disorder named antiphospholipid syndrome (APS). APS patients with adverse pregnancy outcomes showed significantly higher levels of circulating

exosomal-associated miR-146a-3p compared to healthy pregnant controls. The specific aPL significantly induced trophoblasts to express higher level of miR-146a-5p, miR-146a-3p, miR-155 and miR-210. Except miR-155, the other miRNAs were inhibited by the TLR4 antagonist. The suppression of miR-146a-3p significantly reduced aPL-induced trophoblast IL-8 secretion regulated by the TLR8 [66].

4.2. Primary Sjögren's syndrome

Primary SS is a slowly progressive systemic autoimmune inflammatory disease that primarily affects middle-aged women (female to male ratio: 9:1), although it may be found in all ages including childhood. The target organs are primarily exocrine glands, such as salivary and lacrimal glands. Therefore, patients show typically symptoms of dry mouth and dry eyes [67]. Besides the pathognomonic glandular symptoms (GS), other systemic symptoms, denoted as extraglandular manifestations (EGMs) (e.g. polyarthritis, myositis, vasculitis, polyneuropathy etc.) can also develop during the disease course in approximately one third of the patients [68].

The increase of Ro/SSA and La/SSB autoantigens is a common feature in SS patients. The miRNAs which are suspected to target Ro/SSA and La/SSB mRNAs in primary SS are as follows: let-7b, miR-16, miR-181a, miR-200b-3p, miR-200b-5p, miR-223 and miR483-5p. The overexpression of miR-16 in minor salivary glands (MSGs), miR-200b-3p in salivary gland epithelial cells (SGECs) and miR-223 together with miR-483-5p in PBMCs of 29 SS patients compared to 24 sicca-complaining controls has been shown previously. Significant lower-expression of miR200b-5p levels was reported in SS patients with mucosa-associated lymphoid tissue (MALT) lymphoma compared to primary SS patients [69]. Another study demonstrated the positive correlation between the expression levels of La/SSB and the Dicer enzyme in connection with cancer prognosis. La/SSB promotes global microRNA expression and identifies stem-loop [70]. Alevisos et al. generated microRNA microarray profiles from the minor salivary glands of patients with SS who had low-grade or high-grade inflammation and impaired or normal saliva production, and compared the results with that observed in healthy control subjects. They found hsa-miR-768-3p overexpression, while hsa-miR-574 was underexpressed in patients' biopsies; additionally, their inverse correlations to focus scores were also demonstrated [71]. Previously, our workgroup not only confirmed the over-expression of miR-146a/b in PBMCs of SS but also demonstrated the unanticipated over-expression of its functionally targeted gene, TRAF 6. Furthermore, we also reported decreased gene expression of IRAK 1 [72]. The over-expression of TRAF6 is surprising since miR-146a could inhibit the expression of TRAF6 [73]. Recently, enhanced expression of miRNA-155 was reported in untreated Sjögren's syndrome [74]. Of note, SS patients treated with immunosuppressants also showed the over expression of miR-155. On the contrary, in Asian population the relative expression of miR-155 was lower in PBMCs of SS patients not receiving any immunosuppressive treatment than the controls, which may emphasize the importance of the diverse genetic background of different ethnicities [75,76]. A recent study demonstrated the over-expression of miR-181a in the PBMCs of pSS patients, which was associated with the up-regulation of several virus-derived miRNAs, suggesting that viral infection of PBMC plays a role in the disease [77].

Up-regulated expression of miR-34b-3p, miR-4701-5p, miR-609, miR-300, miR-3162-3p, and miR-877-3p in SS monocytes compared to controls may relate with opposing of TGF- β signaling pathway and TLR/NF- κ B pathways induced pro-inflammatory IL-12 secretion [78].

4.3. Rheumatoid arthritis

RA is a frequent autoimmune disorder with prevalence rates approximately 1% of the adults worldwide. The disease primarily affects the synovial joints, and the chronic inflammatory process consequently causes the destruction of the articular tissue [79].

Associations between the alterations in miRNA expressions and the pathomechanisms of the disease have been shown previously. Elevated expression of miR-146a and miR-155 was determined both in whole blood samples and PBMCs of RA in Canadian cohort in comparison with healthy individuals [80]. The expression of the transcription factors (ROR γ t and STAT3) of Th17 cells was significantly increased in the PBMCs of RA patients while miR-301a-3p was also found overexpressed. Levels of miR-301a-3p showed positive correlation with the frequency of Th17 cells in RA patients [81]. MiR-146a, miR-155 and miR-16 were found to have lower expression levels in the serum of early stage of RA patients who were prior to and after 3 and 12 months of antirheumatic drugs therapy compared to established RA. Based on a recent observation, miR-223 may be a potential marker of disease activity since decreased serum level of miR-223 was found after therapy in early RA [82].

MiR-16, miR-132, miR-146a, and miR-223 were found to be overexpressed in synovial fluid and blood plasma of patients compared to healthy controls. No correlation was identified between plasma and synovial fluid miRNAs although concentrations of miRNAs in synovial fluid were significantly lower compared to that of plasma levels [83]. A very recent study showed altered expression levels of certain miRNAs in formalin-fixed paraffin-embedded synovial tissue (FFPE) samples of patients with RA compared to osteoarthritis (OA) patients. It was reported that miR-146a, miR-155, and miR-223 were upregulated significantly in FFPE samples of established RA patients [84].

It was also shown that miR-188-5p is downregulated in synovial tissue samples of RA patients as well as in RA synovial fibroblasts (RASf). Moreover, it was revealed, that miR-188-5p is directly and indirectly regulating the expression of genes confirmed by gene expression profiling in RASf, including hyaluronan binding protein KIAA1199 as well as collagens COL1A1 and COL12A1, which may correlate with extracellular matrix formation and destruction in RA [85].

MiR-573 might be a negative regulator in RA since miR-573 could suppress the activation of mitogen-activated protein kinase (MAPK) which is regarded as one of the potential targets for RA treatment [86].

Regarding CD4+ T cells of RA patients, miRNA expression analysis indicated significant upregulation of miR-146a expression, while miR-363 and miR-498 were downregulated [87].

The miRNA expression in macrophages from patients with active RA and OA was recently determined. Seven miRNAs, namely miR-99a, miR-100, miR-125b, miR-199-3p, miR-199-5p, miR-152 and miR-214 were downregulated and only miR-223 was upregulated in macrophages in RA, compared to the results from OA samples. It was also implied that high miR-223 levels functionally impair the AHR (aryl hydrocarbon receptor)/ARNT (AHR nuclear translocator) pathway in myeloid cells by reducing ARNT protein levels. The AHR activation may be linked to the pathogenesis of RA, since AHR agonists inhibit pro-inflammatory cytokine expression in macrophages [88].

A recent study investigated single nucleotide polymorphisms (SNP) rs22928323 of miR-149 in 200 RA patients and 120 healthy controls. Rs22928323 showed correlation with RA development but was not associated with further clinical characteristics [89].

4.4. Systemic sclerosis

SSc is characterized by accelerated fibrosis and tissue damages in the skin and visceral organs such as heart, lungs and kidneys. SSc can be classified into two sub-groups based on the extent of skin thickening: limited SSc and diffuse SSc. Patients with the limited form are at lower risk of having visceral involvement, while the diffuse form involves several systems of internal organs [90].

Different study groups reported how miRNAs regulate fibrogenesis. The miR-15b, miR-16, miR-27a, miR-27b, miR-132, miR-150, and miR-335 seem to play an important role in the induction of myofibroblast proliferation and resistance to apoptosis [91–98]. On the contrary,

miR-21, miR-92a, miR-133, miR-142-3p, miR-200a/b, and miR-590 have been shown to suppress fibrotic processes [99–105].

Regarding other miRNAs, miR-29a was considered as the most direct regulator of extracellular matrix (ECM) synthesis. It targets the gene TAB1 and may lead to apoptosis of the dermal fibroblasts resulting to lower TIMP-1 production and promote collagen degradation by increasing MMP-1 production, suggesting that miR-29a may be a potential therapeutic target for SSc [106]. The restoration of miR-29a decreased TNF- α production in dermal fibroblasts of SSc patients. Moreover, Bcl-2 expression was upregulated in SSc fibroblasts and the ratio of Bax:Bcl-2 in fibroblasts was significantly lower compared to normal controls. However, miR-29a disrupted the expression profiling of Bcl-2 family proteins (Bax, Bcl-2 and Bcl-XL), which proved that miR-29a is an anti-fibrotic factor induce apoptosis and an attenuator cause ECM production in SSc fibroblasts [107].

Additionally, miR-135b expression is significantly lower both in serum and isolated CD14+ monocytes from patients compared to controls. T cell-derived IL-13 increased collagen expression in dermal fibroblasts which was dependent on STAT6 and miR-135b. Besides, miR-135b is repressed by methylation and could be mediated by the repressive protein methyl cap binding protein 2 (MeCP2), which is significantly enhanced in SSc dermal fibroblasts compared to controls [108].

Iwamoto et al. reported the downregulation of miR-193b in SSc fibroblasts and skin sections. Knockdown of miR-193b induced the expression of mRNA and urokinase-type plasminogen activator (uPA) enzyme, which was strongly expressed in vascular smooth muscle cells in SSc skin section and contributed to the proliferative vasculopathy with intimal hyperplasia characteristic for SSc [109].

4.5. Multiple sclerosis

Multiple sclerosis is an autoimmune neurological disease which affects the brain and the spinal cord thus leading to the main triad symptoms of inflammation, demyelination and gliosis. The damage of the protective covering of the myelin sheath surrounding the nerve cells result in single or multiple symptoms including motoric, speech, swallowing, and visual disabilities and other neuronal problems [110].

MiR expression profile analysis indicated significant overexpression of miR-21, miR-146a, miR-146b and miR-155 in PBMCs of relapsing remitting MS patients compared to controls [111,112]. MiR-326 promotes differentiation by targeting Ets-1, furthermore, its overexpression leads to Th17 cell proliferation and disease aggravation in experimental autoimmune encephalomyelitis [113]. Upregulation of miR-27a was observed in relapsing phase of MS compared to remitting phase and healthy controls; on the contrary, miR-214 was underexpressed in relapsing phase of MS, which implied that miR-27a may inhibit Th17 cell differentiation, while miR-214 may promote Th17 cell differentiation [114].

The expression of miR-140-5p was found to be significantly decreased in the PBMCs of MS patients compared to those in controls, and miR-140-5p level was inversely correlated with disease severity. Transfection of synthetic miR-140-5p in PBMCs inhibited activation of STAT1 and consequently suppressed the encephalitogenic Th1 differentiation, which suggests that miR-140-5p may be a novel marker involved in the pathogenesis of MS [115].

Another group recently reported significantly lower expression of miR-572 in overall MS patients, compared to healthy controls. MiR-572 was found to be significantly upregulated in secondary progressive and relapsing remitting MS, while it was downregulated in primary progressive MS. Consequently, with the different potential, this miRNA could be regarded as a non-invasive biomarker for remyelination [116].

The expression level of miR-150 was elevated in cerebral spinal fluid (CSF) from patients with clinically isolated syndrome (CIS) who convert to MS later, compared to those CIS who did not convert during follow-up (median period of 52 months). The miR-150 may be regarded as a marker of CNS inflammation, since higher levels of miR-150 correlate

with higher levels of CSF biomarkers, involving C-X-C motif chemokine 13 (CXCL13), matrix metalloproteinase 9 (MMP-9) and osteopontin. Additionally, the level of miR-150 in CSF decreased after treating with natalizumab for one year and remains unchanged with fingolimod, while level of miR-150 in plasma increased after the treatment with natalizumab and decreased after fingolimod therapy [117].

4.6. Psoriasis

Psoriasis is a chronic and frequently relapsing inflammatory skin disease characterized by pathologic features such as accelerated epidermopoiesis, marked hyperkeratosis with parakeratosis, vascular dilatation, and inflammatory cell infiltration. The most common form of the disorder is the chronic plaque psoriasis with rounded erythematous, dry, scaling patches. The lesions have a predilection site as nails, scalp, genitalia, extensor surfaces, and the lumbosacral region [118].

Recently, a study group discovered 24 dysregulated miRNAs in the epidermis of psoriatic skin and 37 dysregulated miRNAs in the dermal inflammatory infiltrates of patients. Among those, miR-99a, miR-125b and miR-181a were significantly lower expressed in PBMCs while miR-142-3p, miR-146a, miR-155, miR-224 and miR-378 were upregulated. Moreover, miR-193b was downregulated and miR-223 was upregulated in Th17 cells, while miR-125b was downregulated in T regulatory cells [119]. MiR-146a level was up-regulated in blood samples from patients of psoriasis in comparison with healthy controls, but no significant positive relation was revealed with PASI scores in patients. However after 12 weeks of treatment with Narrow-Band Ultraviolet B phototherapy or treatment with methotrexate, expression of miR-146a decreased dramatically, which suggests that miR-146a may be useful in evaluating and screening the effect of treatment of psoriasis objectively [120].

Even though miR-424 levels were not correlated with disease activity markers, such as PASI (psoriasis area and severity index), hair shaft; miR-424 levels were significantly upregulated in psoriasis patients compared with normal controls and those with atopic dermatitis [121].

A recent study reported increased level of miR-26b-5p in subcutaneous adipose tissue under lesional psoriasis skin compared to nonlesional psoriatic skin. miR-26b-5p down-regulates neutral cholesterol ester hydrolase 1 enzyme, which is essential for cholesterol efflux, in monocytes/macrophages, adipocytes, vascular endothelial cells and fibroblasts [122].

Additionally, the G allele of SNP rs2910164 in miR-146a regarded as a risk factor, which would impair its suppression on the proliferation of keratinocytes through the decreased inhibition of the target gene [123].

5. Conclusions and future perspectives

The discovery of miRNAs and the recognition of their critical role in modulating gene expression changed the way we think about genetic control. The intensive research over the last decade shed light on multiple pathways and modes how miRNAs regulate cell development and differentiation. The central role of miRNAs in modulating immune system responses was also recognized, although, there are numerous questions about miRNAs, yet to be answered.

In the last years much attention was drawn to the function of miRNAs in autoimmunity. Changes in the expression levels of certain miRNAs in the circulation or in different cells and tissues are characteristics for various autoimmune conditions and presumably contribute to disease development. Consequently, some of these molecules may be regarded as novel and attractive biomarkers specific for different autoimmune disorders. However, functional experimental studies are required to verify and establish the causal association between the aberrantly expressed miRNAs and the development of disease. Additionally, the mechanisms underlying the aberrant expression of miRNAs, as well as the influence of other factors that regulate miRNAs, also remained to be investigated.

Genome-wide surveys identified many single nucleotide polymorphisms (SNPs) in the predicted miRNA target sites, as well as in miRNAs themselves. In some instances, SNPs have been shown to alter miRNA function, thus possibly contributing to disease development. The better understanding of the immune regulatory mechanisms of miRNAs by pathway-based exploratory analyses and the mapping and characterization of miRNA SNPs may help not only to elucidate the pathogenesis of autoimmune conditions but also can lead to the development of complex therapeutic approaches in patients with immunological disorders.

Take-home messages

- Alterations of miRNAs expression are involved in the development of autoimmune conditions.
- Certain miRNAs could be regarded as novel and specific biomarkers for different autoimmune diseases.
- Exploration of miRNA target genes will define their role in autoimmunity and reveal novel targets and therapeutic approaches.

Conflict of interest

No disclosure to report.

Acknowledgments

This work was supported by the Chinese Scholarship Council, the grants of the Hungarian National Scientific Research Fund (OTKA) and TÁMOP-4.2.2.A-11/1/KONV-2012-0023 project, which is co-financed by the European Union and European Social Fund.

References

- [1] Chua JH, Armugam A, Jeyaseelan K. MicroRNAs: biogenesis, function and applications. *Curr Opin Mol Ther* 2009;11:189–199.
- [2] Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004;116:281–297.
- [3] Wang H, Peng W, Ouyang X, Li W, Dai Y. Circulating microRNAs as candidate biomarkers in patients with systemic lupus erythematosus. *Transl Res* 2012;160:198–206. <http://dx.doi.org/10.1016/j.trsl.2012.04.002>.
- [4] Kapsogeorgou EK, Gourzi VC, Manoussakis MN, Moutsopoulos HM, Tzioufas AG. Cellular microRNAs (miRNAs) and Sjögren's syndrome: candidate regulators of autoimmune response and autoantigen expression. *J Autoimmun* 2011;37:129–135. <http://dx.doi.org/10.1016/j.jaut.2011.05.003>.
- [5] Furer V, Greenberg JD, Attur M, Abramson SB, Pillinger MH. The role of microRNA in rheumatoid arthritis and other autoimmune diseases. *Clin Immunol* 2010;136:1–15. <http://dx.doi.org/10.1016/j.clim.2010.02.005>.
- [6] Lee Y, Kim M, Han J, Yeom K-H, Lee S, Baek SH, et al. MicroRNA genes are transcribed by RNA polymerase II. *EMBO J* 2004;23:4051–4060. <http://dx.doi.org/10.1038/sj.emboj.7600385>.
- [7] Cullen BR. Transcription and processing of human microRNA precursors. *Mol Cell* 2004;16:861–865. <http://dx.doi.org/10.1016/j.molcel.2004.12.002>.
- [8] Cai X, Hagedorn CH, Cullen BR. Human microRNAs are processed from capped, polyadenylated transcripts that can also function as mRNAs. *RNA* 2004;10:1957–1966. <http://dx.doi.org/10.1261/rna.7135204>.
- [9] Gregory RI, Yan K-P, Amuthan G, Chendrimada T, Doratotaj B, Cooch N, et al. The microprocessor complex mediates the genesis of microRNAs. *Nature* 2004;432:235–240. <http://dx.doi.org/10.1038/nature03120>.
- [10] Lee Y, Ahn C, Han J, Choi H, Kim J, Yim J, et al. The nuclear RNase III Drosha initiates microRNA processing. *Nature* 2003;425:415–419. <http://dx.doi.org/10.1038/nature01957>.
- [11] Yi R, Qin Y, Macara IG, Cullen BR. Exportin-5 mediates the nuclear export of pre-microRNAs and short hairpin RNAs. *Genes Dev* 2003;17:3011–3016. <http://dx.doi.org/10.1101/gad.1158803>.
- [12] Ruby JG, Jan CH, Bartel DP. Intronic microRNA precursors that bypass Drosha processing. *Nature* 2007;448:83–86. <http://dx.doi.org/10.1038/nature05983>.
- [13] Okamura K, Hagen JW, Duan H, Tyler DM, Lai EC. The mirtron pathway generates microRNA-class regulatory RNAs in *Drosophila*. *Cell* 2007;130:89–100. <http://dx.doi.org/10.1016/j.cell.2007.06.028>.
- [14] Berezhikov E, Chung W-J, Willis J, Cuppen E, Lai EC. Mammalian mirtron genes. *Mol Cell* 2007;28:328–336. <http://dx.doi.org/10.1016/j.molcel.2007.09.028>.
- [15] Park J-E, Heo I, Tian Y, Simanshu DK, Chang H, Jee D, et al. Dicer recognizes the 5' end of RNA for efficient and accurate processing. *Nature* 2011;475:201–205. <http://dx.doi.org/10.1038/nature10198>.
- [16] Lund E, Dahlberg JE. Substrate selectivity of exportin 5 and Dicer in the biogenesis of microRNAs. *Cold Spring Harb Symp Quant Biol* 2006;71:59–66. <http://dx.doi.org/10.1101/sqb.2006.71.050>.
- [17] Yoda M, Cifuentes D, Izumi N, Sakaguchi Y, Suzuki T, Giraldez AJ, et al. Poly(A)-specific ribonuclease mediates 3'-end trimming of Argonaute2-cleaved precursor microRNAs. *Cell Rep* 2013;5:715–726. <http://dx.doi.org/10.1016/j.celrep.2013.09.029>.

- [18] Fabian MR, Sonenberg N, Filipowicz W. Regulation of mRNA translation and stability by microRNAs. *Annu Rev Biochem* 2010;79:351–379. <http://dx.doi.org/10.1146/annurev-biochem-060308-103103>.
- [19] Lewis BP, Burge CB, Bartel DP. Conserved seed pairing, often flanked by adenosines, indicates that thousands of human genes are microRNA targets. *Cell* 2005;120:15–20. <http://dx.doi.org/10.1016/j.cell.2004.12.035>.
- [20] Carthew RW, Sontheimer EJ. Origins and mechanisms of miRNAs and siRNAs. *Cell* 2009;136:642–655. <http://dx.doi.org/10.1016/j.cell.2009.01.035>.
- [21] Valencia-Sanchez MA, Liu J, Hannon GJ, Parker R. Control of translation and mRNA degradation by miRNAs and siRNAs. *Genes Dev* 2006;20:515–524. <http://dx.doi.org/10.1101/gad.1399806>.
- [22] Meister G, Landthaler M, Patkaniowska A, Dorsett Y, Teng G, Tuschl T. Human Argonaute2 mediates RNA cleavage targeted by miRNAs and siRNAs. *Mol Cell* 2004;15:185–197. <http://dx.doi.org/10.1016/j.molcel.2004.07.007>.
- [23] Pillai RS, Bhattacharyya SN, Filipowicz W. Repression of protein synthesis by miRNAs: how many mechanisms? *Trends Cell Biol* 2007;17:118–126. <http://dx.doi.org/10.1016/j.tcb.2006.12.007>.
- [24] Bruno I, Wilkinson MF. P-bodies react to stress and nonsense. *Cell* 2006;125:1036–1038. <http://dx.doi.org/10.1016/j.cell.2006.06.003>.
- [25] Dai R, Ahmed SA. MicroRNA, a new paradigm for understanding immunoregulation, inflammation, and autoimmune diseases. *Transl Res* 2011;157:163–179. <http://dx.doi.org/10.1016/j.trsl.2011.01.007>.
- [26] Gangaraju VK, Lin H. MicroRNAs: key regulators of stem cells. *Nat Rev Mol Cell Biol* 2009;10:116–125. <http://dx.doi.org/10.1038/nrm2621>.
- [27] Chen C-Z, Li L, Lodish HF, Bartel DP. MicroRNAs modulate hematopoietic lineage differentiation. *Science* 2004;303:83–86. <http://dx.doi.org/10.1126/science.1091903>.
- [28] Kąghuwanshi S, Karnati HK, Sarvothaman S, Gutti U, Saladi RG, Tummala PR, et al. microRNAs: key players in hematopoiesis. *Adv Exp Med Biol* 2015;887:171–211. http://dx.doi.org/10.1007/978-3-319-22380-3_10.
- [29] Akira S, Uematsu S, Takeuchi O. Pathogen recognition and innate immunity. *Cell* 2006;124:783–801. <http://dx.doi.org/10.1016/j.cell.2006.02.015>.
- [30] Santegoets KCM, van Bon L, van den Berg WB, Wenink MH, Radstake TRDJ. Toll-like receptors in rheumatic diseases: are we paying a high price for our defense against bugs? *FEBS Lett* 2011;585:3660–3666. <http://dx.doi.org/10.1016/j.febslet.2011.04.028>.
- [31] Mogensen TH, Paludan SR. Reading the viral signature by Toll-like receptors and other pattern recognition receptors. *J Mol Med (Berl)* 2005;83:180–192. <http://dx.doi.org/10.1007/s00109-004-0620-6>.
- [32] Kawai T, Akira S. Toll-like receptors and their crosstalk with other innate receptors in infection and immunity. *Immunity* 2011;34:637–650. <http://dx.doi.org/10.1016/j.immuni.2011.05.006>.
- [33] Roach JC, Glusman G, Rowen L, Kaur A, Purcell MK, Smith KD, et al. The evolution of vertebrate Toll-like receptors. *Proc Natl Acad Sci U S A* 2005;102:9577–9582. <http://dx.doi.org/10.1073/pnas.0502272102>.
- [34] Takeda K, Akira S. Toll-like receptors in innate immunity. *Int Immunol* 2005;17:1–14. <http://dx.doi.org/10.1093/intimm/dxh186>.
- [35] Chen J-Q, Szodoray P, Zeher M. Toll-like receptor pathways in autoimmune diseases. *Clin Rev Allergy Immunol* 2015. <http://dx.doi.org/10.1007/s12016-015-8473-z>.
- [36] Taganov KD, Boldin MP, Chang K-J, Baltimore D. NF-kappaB-dependent induction of microRNA miR-146, an inhibitor targeted to signaling proteins of innate immune responses. *Proc Natl Acad Sci U S A* 2006;103:12481–12486. <http://dx.doi.org/10.1073/pnas.0605298103>.
- [37] Hou J, Wang P, Lin L, Liu X, Ma F, An H, et al. MicroRNA-146a feedback inhibits RIG-I-dependent Type I IFN production in macrophages by targeting TRAF6, IRAK1, and IRAK2. *J Immunol* 2009;183:2150–2158. <http://dx.doi.org/10.4049/jimmunol.0900707>.
- [38] Tili E, Michaille J-J, Cimino A, Costinean S, Dumitru CD, Adair B, et al. Modulation of miR-155 and miR-125b levels following lipopolysaccharide/TNF-alpha stimulation and their possible roles in regulating the response to endotoxin shock. *J Immunol* 2007;179:5082–5089.
- [39] Liu G, Friggeri A, Yang Y, Park Y-J, Tsuruta Y, Abraham E. miR-147, a microRNA that is induced upon Toll-like receptor stimulation, regulates murine macrophage inflammatory responses. *Proc Natl Acad Sci U S A* 2009;106:15819–15824. <http://dx.doi.org/10.1073/pnas.0901216106>.
- [40] Au KY, Pong JCH, Ling WL, Li JCB. MiR-1303 regulates mycobacteria induced autophagy by targeting Atg2B. *PLoS One* 2016;11, e0146770. <http://dx.doi.org/10.1371/journal.pone.0146770>.
- [41] Húsáková M. MicroRNAs in the key events of systemic lupus erythematosus pathogenesis. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub* 2016. <http://dx.doi.org/10.5507/bp.2016.004>.
- [42] Cobb BS, Nesterova TB, Thompson E, Hertweck A, O'Connor E, Godwin J, et al. T cell lineage choice and differentiation in the absence of the RNase III enzyme Dicer. *J Exp Med* 2005;201:1367–1373. <http://dx.doi.org/10.1084/jem.20050572>.
- [43] Muljo SA, Ansel KM, Kanellopoulou C, Livingston DM, Rao A, Rajewsky K. Aberrant T cell differentiation in the absence of Dicer. *J Exp Med* 2005;202:261–269. <http://dx.doi.org/10.1084/jem.20050678>.
- [44] Koralov SB, Muljo SA, Galler GR, Krek A, Chakraborty T, Kanellopoulou C, et al. Dicer ablation affects antibody diversity and cell survival in the B lymphocyte lineage. *Cell* 2008;132:860–874. <http://dx.doi.org/10.1016/j.cell.2008.02.020>.
- [45] Du C, Liu C, Kang J, Zhao G, Ye Z, Huang S, et al. MicroRNA miR-326 regulates TH-17 differentiation and is associated with the pathogenesis of multiple sclerosis. *Nat Immunol* 2009;10:1252–1259. <http://dx.doi.org/10.1038/ni.1798>.
- [46] O'Connell RM, Kahn D, Gibson WSJ, Round JL, Scholz RL, Chaudhuri AA, et al. MicroRNA-155 promotes autoimmune inflammation by enhancing inflammatory T cell development. *Immunity* 2010;33:607–619. <http://dx.doi.org/10.1016/j.immuni.2010.09.009>.
- [47] Ceppi M, Pereira PM, Dunand-Sauthier I, Barras E, Reith W, Santos MA, et al. MicroRNA-155 modulates the interleukin-1 signaling pathway in activated human monocyte-derived dendritic cells. *Proc Natl Acad Sci U S A* 2009;106:2735–2740. <http://dx.doi.org/10.1073/pnas.0811073106>.
- [48] Lu L-F, Thai T-H, Calado DP, Chaudhry A, Kubo M, Tanaka K, et al. Foxp3-dependent microRNA155 confers competitive fitness to regulatory T cells by targeting SOCS1 protein. *Immunity* 2009;30:80–91. <http://dx.doi.org/10.1016/j.immuni.2008.11.010>.
- [49] Lu L-F, Boldin MP, Chaudhry A, Lin L-L, Taganov KD, Hanada T, et al. Function of miR-146a in controlling Treg cell-mediated regulation of Th1 responses. *Cell* 2010;142:914–929. <http://dx.doi.org/10.1016/j.cell.2010.08.012>.
- [50] de Yébenes VG, Belver L, Pisano DG, González S, Villasante A, Croce C, et al. miR-181b negatively regulates activation-induced cytidine deaminase in B cells. *J Exp Med* 2008;205:2199–2206. <http://dx.doi.org/10.1084/jem.20080579>.
- [51] Gonzalez-Martín A, Adams BD, Lai M, Shepherd J, Salvador-Bernaldez M, Salvador JM, et al. The microRNA miR-148a functions as a critical regulator of B cell tolerance and autoimmunity. *Nat Immunol* 2016;17:433–440. <http://dx.doi.org/10.1038/ni.3385>.
- [52] Rothfield N. Clinical aspects and treatment of systemic lupus erythematosus. *Curr Opin Rheumatol* 1989;1:327–331.
- [53] Liu Y-J, Fan W-J, Bai J-Z. microRNA-126 expression and its mechanism of action in patients with systemic lupus erythematosus. *Eur Rev Med Pharmacol Sci* 2015;19:3838–3842.
- [54] Zhu J, Huang X, Su G, Wang L, Wu F, Zhang T, et al. High expression levels of microRNA-629, microRNA-525-5p and microRNA-516a-3p in paediatric systemic lupus erythematosus. *Clin Rheumatol* 2014;33:807–815. <http://dx.doi.org/10.1007/s10067-014-2583-5>.
- [55] Dai Y, Huang Y-S, Tang M, Lv T-Y, Hu C-X, Tan Y-H, et al. Microarray analysis of microRNA expression in peripheral blood cells of systemic lupus erythematosus patients. *Lupus* 2007;16:939–946. <http://dx.doi.org/10.1177/0961203307084158>.
- [56] Dai Y, Sui W, Lan H, Yan Q, Huang H, Huang Y. Comprehensive analysis of microRNA expression patterns in renal biopsies of lupus nephritis patients. *Rheumatol Int* 2009;29:749–754. <http://dx.doi.org/10.1007/s00296-008-0758-6>.
- [57] Tang Y, Luo X, Cui H, Ni X, Yuan M, Guo Y, et al. MicroRNA-146 A contributes to abnormal activation of the type I interferon pathway in human lupus by targeting the key signaling proteins. *Arthritis Rheum* 2009;60:1065–1075. <http://dx.doi.org/10.1002/art.24436>.
- [58] Liu F, Fan H, Ren D, Dong G, Hu E, Ji J, et al. TLR9-induced miR-155 and Ets-1 decrease expression of CD1d on B cells in SLE. *Eur J Immunol* 2015;45:1934–1945. <http://dx.doi.org/10.1002/eji.201445286>.
- [59] Lashine YA, Salah S, Aboulelein HR, Abdelaziz AI. Correcting the expression of miRNA-155 represses PP2Ac and enhances the release of IL-2 in PBMCs of juvenile SLE patients. *Lupus* 2015;24:240–247. <http://dx.doi.org/10.1177/0961203314552117>.
- [60] Liu D, Zhang N, Zhang J, Zhao H, Wang X. miR-410 suppresses the expression of interleukin-6 as well as renal fibrosis in the pathogenesis of lupus nephritis. *Clin Exp Pharmacol Physiol* 2016;43:616–625. <http://dx.doi.org/10.1111/1440-1681.12576>.
- [61] Coit P, Dozmorov MG, Merrill JT, McCune WJ, Maksimowicz-McKinnon K, Wren JD, et al. Epigenetic reprogramming in naïve CD4+ T cells favoring T cell activation and non-Th1 effector T cell immune response as an early event in lupus flares. *Arthritis Rheumatol* 2016. <http://dx.doi.org/10.1002/art.39720>.
- [62] Qingjuan L, Xiaojuan F, Wei Z, Chao W, Pengpeng K, Hongbo L, et al. miR-148a-3p overexpression contributes to glomerular cell proliferation by targeting PTEN in lupus nephritis. *Am J Physiol Cell Physiol* 2016;310:C470–C478. <http://dx.doi.org/10.1152/ajpcell.00129.2015>.
- [63] Wang W, Mou S, Wang L, Zhang M, Shao X, Fang W, et al. Up-regulation of serum miR-130b-3p level is associated with renal damage in early lupus nephritis. *Sci Rep* 2015;5:12644. <http://dx.doi.org/10.1038/srep12644>.
- [64] Solé C, Cortés-Hernández J, Felip ML, Vidal M, Ordi-Ros J. miR-29c in urinary exosomes as predictor of early renal fibrosis in lupus nephritis. *Nephrol Dial Transplant* 2015;30:1488–1496. <http://dx.doi.org/10.1093/ndt/gfv128>.
- [65] Perez-Hernandez J, Forner MJ, Pinto C, Chaves JF, Cortes R, Redon J. Increased urinary exosomal microRNAs in patients with systemic lupus erythematosus. *PLoS One* 2015;10, e0138618. <http://dx.doi.org/10.1371/journal.pone.0138618>.
- [66] Gysler SM, Mulla MJ, Guerra M, Brosens JJ, Salmon JE, Chamley LW, et al. Antiphospholipid antibody-induced miR-146a-3p drives trophoblast interleukin-8 secretion through activation of Toll-like receptor 8. *Mol Hum Reprod* 2016. <http://dx.doi.org/10.1093/molehr/gaw027>.
- [67] Zeher M, Szodoray P. Sjögren's syndrome and associated disorders. vol. Sjögren's Syndrome. Kerala, India: Transworld Research Network; 2009.
- [68] Szanto A, Szodoray P, Kiss E, Kapitany A, Szegedi G, Zeher M. Clinical, serologic, and genetic profiles of patients with associated Sjögren's syndrome and systemic lupus erythematosus. *Hum Immunol* 2006;67:924–930. <http://dx.doi.org/10.1016/j.humimm.2006.06.006>.
- [69] Goutz VC, Kapsogeorou EK, Kyriakidis NC, Tzioufas AG. Study of microRNAs (miRNAs) that are predicted to target the autoantigens Ro/SSA and La/SSB in primary Sjögren's Syndrome. *Clin Exp Immunol* 2015;182:14–22. <http://dx.doi.org/10.1111/cei.12664>.
- [70] Liang C, Xiong K, Szulwach KE, Zhang Y, Wang Z, Peng J, et al. Sjögren syndrome antigen B (SSB)/La promotes global microRNA expression by binding microRNA precursors through stem-loop recognition. *J Biol Chem* 2013;288:723–736. <http://dx.doi.org/10.1074/jbc.M112.401323>.
- [71] Alevizos I, Alexander S, Turner RJ, Illei GG. MicroRNA expression profiles as biomarkers of minor salivary gland inflammation and dysfunction in Sjögren's syndrome. *Arthritis Rheum* 2011;63:535–544. <http://dx.doi.org/10.1002/art.30131>.
- [72] Zilahi E, Tarr T, Papp G, Griger Z, Sipka S, Zeher M. Increased microRNA-146a/b, TRAF6 gene and decreased IRAK1 gene expressions in the peripheral mononuclear cells of patients with Sjögren's syndrome. *Immunol Lett* 2012;141:165–168. <http://dx.doi.org/10.1016/j.imlet.2011.09.006>.
- [73] Bhaumik D, Scott GK, Schokrpur S, Patil CK, Campisi J, Benz CC. Expression of microRNA-146 suppresses NF-kappaB activity with reduction of metastatic potential in breast cancer cells. *Oncogene* 2008;27:5643–5647. <http://dx.doi.org/10.1038/onc.2008.171>.
- [74] Chen J-Q, Zilahi E, Papp G, Sipka S, Zeher M. Simultaneously increased expression of microRNA-155 and suppressor of cytokine signaling 1 (SOCS1) gene in the peripheral blood mononuclear cells of patients with primary Sjögren's syndrome. *Int J Rheum Dis* 2015. <http://dx.doi.org/10.1111/1756-185X.12804>.
- [75] Pauley KM, Stewart CM, Gauna AE, Dupre LC, Kulkarni R, Chan AL, et al. Altered miR-146a expression in Sjögren's syndrome and its functional role in innate immunity. *Eur J Immunol* 2011;41:2029–2039. <http://dx.doi.org/10.1002/eji.201040757>.
- [76] Shi H, Zheng L, Zhang P, Yu C. miR-146a and miR-155 expression in PBMCs from patients with Sjögren's syndrome. *J Oral Pathol Med* 2014;43:792–797. <http://dx.doi.org/10.1111/jop.12187>.
- [77] Peng L, Ma W, Yi F, Yang Y-J, Lin W, Chen H, et al. MicroRNA profiling in Chinese patients with primary Sjögren syndrome reveals elevated miRNA-181a in peripheral blood

- mononuclear cells. *J Rheumatol* 2014;41:2208–2213. <http://dx.doi.org/10.3899/jrheum.131154>.
- [78] Williams AEG, Choi K, Chan AL, Lee YJ, Reeves WH, Bubbs MR, et al. Sjögren's syndrome-associated microRNAs in CD14(+) monocytes unveils targeted TGF β signaling. *Arthritis Res Ther* 2016;18:95. <http://dx.doi.org/10.1186/s13075-016-0987-0>.
- [79] Gibofsky A. Overview of epidemiology, pathophysiology, and diagnosis of rheumatoid arthritis. *Am J Manag Care* 2012;18:S295–S302.
- [80] Mookherjee N, El-Gabalawy HS. High degree of correlation between whole blood and PBMC expression levels of miR-155 and miR-146a in healthy controls and rheumatoid arthritis patients. *J Immunol Methods* 2013;400–401:106–110. <http://dx.doi.org/10.1016/j.jim.2013.10.001>.
- [81] Tang X, Yin K, Zhu H, Tian J, Shen D, Yi L, et al. Correlation between the expression of microRNA-301a-3p and the proportion of Th17 cells in patients with rheumatoid arthritis. *Inflammation* 2016. <http://dx.doi.org/10.1007/s10753-016-0304-8>.
- [82] Filková M, Aradi B, Senolt L, Ospelt C, Vettori S, Mann H, et al. Association of circulating miR-223 and miR-16 with disease activity in patients with early rheumatoid arthritis. *Ann Rheum Dis* 2014;73:1898–1904. <http://dx.doi.org/10.1136/annrheumdis-2012-202815>.
- [83] Murata K, Yoshitomi H, Tanida S, Ishikawa M, Nishitani K, Ito H, et al. Plasma and synovial fluid microRNAs as potential biomarkers of rheumatoid arthritis and osteoarthritis. *Arthritis Res Ther* 2010;12:R86. <http://dx.doi.org/10.1186/ar3013>.
- [84] Kriegsmann M, Randau TM, Gravius S, Lisenko K, Altmann C, Arens N, et al. Expression of miR-146a, miR-155, and miR-223 in formalin-fixed paraffin-embedded synovial tissues of patients with rheumatoid arthritis and osteoarthritis. *Virchows Arch* 2016. <http://dx.doi.org/10.1007/s00428-016-1939-4>.
- [85] Ruedel A, Dietrich P, Schubert T, Hofmeister S, Hellerbrand C, Bosserhoff A-K. Expression and function of microRNA-188-5p in activated rheumatoid arthritis synovial fibroblasts. *Int J Clin Exp Pathol* 2015;8:4953–4962.
- [86] Wang L, Song G, Zheng Y, Wang D, Dong H, Pan J, et al. miR-573 is a negative regulator in the pathogenesis of rheumatoid arthritis. *Cell Mol Immunol* 2015. <http://dx.doi.org/10.1038/cmi.2015.63>.
- [87] Li J, Wan Y, Guo Q, Zou L, Zhang J, Fang Y, et al. Altered microRNA expression profile with miR-146a upregulation in CD4+ T cells from patients with rheumatoid arthritis. *Arthritis Res Ther* 2010;12:R81. <http://dx.doi.org/10.1186/ar3006>.
- [88] Ogando J, Nardáguila M, Díaz-Alderete A, Usategui A, Miranda-Ramos V, Martínez-Herrera DJ, et al. Notch-regulated miR-223 targets the aryl hydrocarbon receptor pathway and increases cytokine production in macrophages from rheumatoid arthritis patients. *Sci Rep* 2016;6:20223. <http://dx.doi.org/10.1038/srep20223>.
- [89] Xiao M, Ma Y, Chen X, Kuang B. Single nucleotide polymorphism of miR-149 and susceptibility of rheumatoid arthritis. *Zhong Nan Da Xue Xue Bao Yi Xue Ban* 2015;40:495–498. <http://dx.doi.org/10.11817/j.jissn.1672-7347.2015.05.006>.
- [90] Casale R, Buonocore M, Matucci-Cerinic M. Systemic sclerosis (scleroderma): an integrated challenge in rehabilitation. *Arch Phys Med Rehabil* 1997;78:767–773.
- [91] Chen C, Wu C-Q, Zhang Z-Q, Yao D-K, Zhu L. Loss of expression of miR-335 is implicated in hepatic stellate cell migration and activation. *Exp Cell Res* 2011;317:1714–1725. <http://dx.doi.org/10.1016/j.yexcr.2011.05.001>.
- [92] Venugopal SK, Jiang J, Kim T-H, Li Y, Wang S-S, Torok NJ, et al. Liver fibrosis causes downregulation of miRNA-150 and miRNA-194 in hepatic stellate cells, and their overexpression causes decreased stellate cell activation. *Am J Physiol Gastrointest Liver Physiol* 2010;298:G101–G106. <http://dx.doi.org/10.1152/ajpgi.00220.2009>.
- [93] Guo C-J, Pan Q, Li D-G, Sun H, Liu B-W. miR-15b and miR-16 are implicated in activation of the rat hepatic stellate cell: an essential role for apoptosis. *J Hepatol* 2009;50:766–778. <http://dx.doi.org/10.1016/j.jhep.2008.11.025>.
- [94] Freischmidt A, Müller K, Ludolph AC, Weishaupt JH. Systemic dysregulation of TDP-43 binding microRNAs in amyotrophic lateral sclerosis. *Acta Neuropathol Commun* 2013;1:42. <http://dx.doi.org/10.1186/2051-5960-1-42>.
- [95] Ji J, Zhang J, Huang G, Qian J, Wang X, Mei S. Over-expressed microRNA-27a and 27b influence fat accumulation and cell proliferation during rat hepatic stellate cell activation. *FEBS Lett* 2009;583:759–766. <http://dx.doi.org/10.1016/j.febslet.2009.01.034>.
- [96] Honda N, Jinnin M, Kira-Etoh T, Makino K, Kajihara I, Makino T, et al. miR-150 downregulation contributes to the constitutive type I collagen overexpression in scleroderma dermal fibroblasts via the induction of integrin β 3. *Am J Pathol* 2013;182:206–216. <http://dx.doi.org/10.1016/j.ajpath.2012.09.023>.
- [97] Makino K, Jinnin M, Aoi J, Hirano A, Kajihara I, Makino T, et al. Discoidin domain receptor 2-microRNA 196a-mediated negative feedback against excess type I collagen expression is impaired in scleroderma dermal fibroblasts. *J Invest Dermatol* 2013;133:110–119. <http://dx.doi.org/10.1038/jid.2012.252>.
- [98] Honda N, Jinnin M, Kajihara I, Makino T, Makino K, Masuguchi S, et al. TGF- β -mediated downregulation of microRNA-196a contributes to the constitutive upregulated type I collagen expression in scleroderma dermal fibroblasts. *J Immunol* 2012;188:3323–3331. <http://dx.doi.org/10.4049/jimmunol.1100876>.
- [99] Makino K, Jinnin M, Kajihara I, Honda N, Sakai K, Masuguchi S, et al. Circulating miR-142-3p levels in patients with systemic sclerosis. *Clin Exp Dermatol* 2012;37:34–39. <http://dx.doi.org/10.1111/j.1365-2230.2011.04158.x>.
- [100] Sing T, Jinnin M, Yamane K, Honda N, Makino K, Kajihara I, et al. microRNA-92a expression in the sera and dermal fibroblasts increases in patients with scleroderma. *Rheumatology (Oxford)* 2012;51:1550–1556. <http://dx.doi.org/10.1093/rheumatology/kes120>.
- [101] Zhu H, Li Y, Qu S, Luo H, Zhou Y, Wang Y, et al. MicroRNA expression abnormalities in limited cutaneous scleroderma and diffuse cutaneous scleroderma. *J Clin Immunol* 2012;32:906–913. <http://dx.doi.org/10.1007/s10875-011-9647-y>.
- [102] Shan H, Zhang Y, Lu Y, Zhang Y, Pan Z, Cai B, et al. Downregulation of miR-133 and miR-590 contributes to nicotine-induced atrial remodeling in canines. *Cardiovasc Res* 2009;83:465–472. <http://dx.doi.org/10.1093/cvr/cvp130>.
- [103] Wang B, Koh P, Winbanks C, Coughlan MT, McClelland A, Watson A, et al. miR-200a prevents renal fibrogenesis through repression of TGF- β 2 expression. *Diabetes* 2011;60:912–913. <http://dx.doi.org/10.2337/db10-0892>.
- [104] Liu G, Friggeri A, Yang Y, Milosevic J, Ding Q, Thannickal VJ, et al. miR-21 mediates fibrogenic activation of pulmonary fibroblasts and lung fibrosis. *J Exp Med* 2010;207:1589–1597. <http://dx.doi.org/10.1084/jem.20100035>.
- [105] Zhong X, Chung ACK, Chen H-Y, Meng X-M, Lan HY. Smad3-mediated upregulation of miR-21 promotes renal fibrosis. *J Am Soc Nephrol* 2011;22:1668–1681. <http://dx.doi.org/10.1681/ASN.2010111168>.
- [106] Ciechomska M, O'Reilly S, Suwara M, Bogunia-Kubik K, van Laar JM. MiR-29a reduces TIMP-1 production by dermal fibroblasts via targeting TGF- β activated kinase 1 binding protein 1, implications for systemic sclerosis. *PLoS One* 2014;9:e115596. <http://dx.doi.org/10.1371/journal.pone.0115596>.
- [107] Jafarnejad-Farsangi S, Farazmand A, Mahmoudi M, Gharibdoost F, Karimzadeh E, Noorbakhsh F, et al. MicroRNA-29a induces apoptosis via increasing the Bax:Bcl-2 ratio in dermal fibroblasts of patients with systemic sclerosis. *Autoimmunity* 2015;48:926–932. <http://dx.doi.org/10.3109/08916934.2015.1030616>.
- [108] O'Reilly S, Ciechomska M, Fullard N, Przyborski S, van Laar JM. IL-13 mediates collagen deposition via STAT6 and microRNA-135b: a role for epigenetics. *Sci Rep* 2016;6:25066. <http://dx.doi.org/10.1038/srep25066>.
- [109] Iwamoto N, Vettori S, Maurer B, Brock M, Pachera E, Jüngel A, et al. Downregulation of miR-193b in systemic sclerosis regulates the proliferative vasculopathy by urokinase-type plasminogen activator expression. *Ann Rheum Dis* 2016;75:303–310. <http://dx.doi.org/10.1136/annrheumdis-2014-205326>.
- [110] Polman CH, Reingold SC, Edan G, Filippi M, Hartung H-P, Kappos L, et al. Diagnostic criteria for multiple sclerosis: 2005 revisions to the “McDonald Criteria”. *Ann Neurol* 2005;58:840–846. <http://dx.doi.org/10.1002/ana.20703>.
- [111] Fenoglio C, Cantoni C, De Riz M, Ridolfi E, Cortini F, Serpente M, et al. Expression and genetic analysis of miRNAs involved in CD4+ cell activation in patients with multiple sclerosis. *Neurosci Lett* 2011;504:9–12. <http://dx.doi.org/10.1016/j.neulet.2011.08.021>.
- [112] Devier DJ, Lovera JF, Lukiw WJ. Increase in NF- κ B-sensitive miRNA-146a and miRNA-155 in multiple sclerosis (MS) and pro-inflammatory neurodegeneration. *Front Mol Neurosci* 2015;8:5. <http://dx.doi.org/10.3389/fnmol.2015.00005>.
- [113] Du C, Liu C, Kang J, Zhao G, Ye Z, Huang S, et al. MicroRNA miR-326 regulates TH-17 differentiation and is associated with the pathogenesis of multiple sclerosis. *Nat Immunol* 2009;10:1252–1259. <http://dx.doi.org/10.1038/ni.1798>.
- [114] Ahmadian-Elmi M, Bidmehski Pour A, Naghavian R, Ghaedi K, Tanhaei S, Izadi T, et al. miR-27a and miR-214 exert opposite regulatory roles in Th17 differentiation via mediating different signaling pathways in peripheral blood CD4(+) T lymphocytes of patients with relapsing-remitting multiple sclerosis. *Immunogenetics* 2016;68:43–54. <http://dx.doi.org/10.1007/s00251-015-0881-y>.
- [115] Guan H, Singh UP, Rao R, Mrelashvili D, Sen S, Hao H, et al. Inverse correlation of expression of microRNA-140-5p with progression of multiple sclerosis and differentiation of encephalitogenic T helper type 1 cells. *Immunology* 2016;147:488–498. <http://dx.doi.org/10.1111/imm.12583>.
- [116] Mancuso R, Hervis A, Agostini S, Rovaris M, Caputo D, Clerici M. MicroRNA-572 expression in multiple sclerosis patients with different patterns of clinical progression. *J Transl Med* 2015;13:148. <http://dx.doi.org/10.1186/s12967-015-0504-2>.
- [117] Bergman P, Picket E, Khademi M, James T, Brundin L, Olsson T, et al. Circulating miR-150 in CSF is a novel candidate biomarker for multiple sclerosis. *Neuro Neuroimmunol Neuroinflammation* 2016;3:e219. <http://dx.doi.org/10.1212/NXI.0000000000000219>.
- [118] Christophers E. Psoriasis—epidemiology and clinical spectrum. *Clin Exp Dermatol* 2001;26:314–320.
- [119] Lövendorf MB, Mitsui H, Zibert JR, Røpke MA, Hafner M, Dyring-Andersen B, et al. Laser capture microdissection followed by next-generation sequencing identifies disease-related microRNAs in psoriatic skin that reflect systemic microRNA changes in psoriasis. *Exp Dermatol* 2015;24:187–193. <http://dx.doi.org/10.1111/exd.12604>.
- [120] Ele-Refaei AM, El-Esawy FM. Effect of narrow-band ultraviolet B phototherapy and methotrexate on microRNA (146a) levels in blood of psoriatic patients. *Dermatol Res Pract* 2015;2015:145769. <http://dx.doi.org/10.1155/2015/145769>.
- [121] Tsuru Y, Jinnin M, Ichihara A, Fujisawa A, Moriya C, Sakai K, et al. miR-424 levels in hair shaft are increased in psoriatic patients. *J Dermatol* 2014;41:382–385. <http://dx.doi.org/10.1111/1346-8138.12460>.
- [122] Cheung L, Fisher RM, Kuzmina N, Li D, Li X, Werngren O, et al. Psoriasis skin inflammation-induced microRNA-26b targets NCEH1 in underlying subcutaneous adipose tissue. *J Invest Dermatol* 2016;136:640–648. <http://dx.doi.org/10.1016/j.jid.2015.12.008>.
- [123] Zhang W, Yi X, Guo S, Shi Q, Wei C, Li X, et al. A single-nucleotide polymorphism of miR-146a and psoriasis: an association and functional study. *J Cell Mol Med* 2014;18:978–980. <http://dx.doi.org/10.1111/jcmm.12359>.