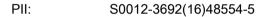
Accepted Manuscript

Human epididymis protein 4 (HE4): a novel serum inflammatory biomarker in cystic fibrosis

Béla Nagy, Jr., MD, PhD, Béla Nagy, MD, PhD, Libor Fila, MD, Luka A. Clarke, PhD, Ferenc Gönczy, MD, Olga Bede, MD, PhD, Dóra Nagy, MD, Rita Újhelyi, MD, PhD, Ágnes Szabó, MD, Andrea Anghelyi, MD, Miklós Major, MD, Zsolt Bene, MD, Zsolt Fejes, MSc, Péter Antal-Szalmás, MD, PhD, Harjit Pal Bhattoa, MD, PhD, György Balla, MD, PhD, János Kappelmayer, MD, PhD, Margarida D. Amaral, PhD, Milan Macek, Jr., MD, PhD, István Balogh, PhD



DOI: 10.1016/j.chest.2016.04.006

Reference: CHEST 434

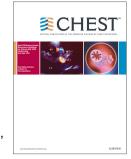
To appear in: CHEST

Received Date: 31 March 2016

Accepted Date: 4 April 2016

Please cite this article as: Nagy B Jr, Nagy B, Fila L, Clarke LA, Gönczy F, Bede O, Nagy D, Újhelyi R, Szabó Á, Anghelyi A, Major M, Bene Z, Fejes Z, Antal-Szalmás P, Bhattoa HP, Balla G, Kappelmayer J, Amaral MD, Macek M Jr, Balogh I, Human epididymis protein 4 (HE4): a novel serum inflammatory biomarker in cystic fibrosis, *CHEST* (2016), doi: 10.1016/j.chest.2016.04.006.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



Human epididymis protein 4 (HE4): a novel serum inflammatory biomarker in cystic fibrosis

Béla Nagy Jr MD, PhD^{1,*}; Béla Nagy MD, PhD³; Libor Fila MD⁴; Luka A. Clarke PhD⁵, Ferenc Gönczy MD⁶; Olga Bede MD, PhD⁷; Dóra Nagy MD⁷, Rita Újhelyi MD, PhD⁸; Ágnes Szabó MD⁷; Andrea Anghelyi MD⁹; Miklós Major MD¹⁰; Zsolt Bene MD³; Zsolt Fejes MSc¹; Péter Antal-Szalmás MD, PhD¹; Harjit Pal Bhattoa MD, PhD¹; György Balla MD, PhD³; János Kappelmayer MD, PhD¹; Margarida D. Amaral PhD⁵, Milan Macek Jr MD, PhD¹¹; István Balogh PhD^{1,2}

¹Department of Laboratory Medicine, ²Division of Clinical Genetics, ³Institute of Pediatrics, Faculty of Medicine, University of Debrecen, Debrecen, Hungary, ⁴Department of Pulmonology, Charles University, 2nd Faculty of Medicine, Motol University Hospital, Prague, Czech Republic, ⁵University of Lisboa, Faculty of Sciences, BiolSI-Biosystems & Integrative Sciences Institute, Lisboa, Portugal, ⁶Kenézy Gyula County Hospital, Debrecen, Hungary, ⁷Department of Pediatrics, Szent-Györgyi Albert Medical University, Szeged, Hungary, ⁸Heim Pál Children's Hospital, Budapest, Hungary, ⁹Petz Aladár County Hospital, Győr, Hungary, ¹⁰Markusovszky Lajos County Hospital, Szombathely, Hungary, ¹¹Department of Biology and Medical Genetics, Motol University Hospital, 2nd Faculty of Medicine, Charles University, Prague, Czech Republic

Key words: cystic fibrosis, HE4, sweat test, CFTR mutations, CRP, inflammation, FEV1

Running title: Elevated serum levels of HE4 in cystic fibrosis

Conflict of interest: MDA has received compensation for scientific seminars from Vertex Pharmaceuticals, PTC Pharmaceuticals and Gilead; research grants from Gilead Genese and Vertex Pharmaceuticals. All other authors declare no conflict of interest.

*Corresponding authors:

Béla Nagy Jr MD, PhD, Department of Laboratory Medicine, University of Debrecen,

Nagyerdei krt. 98., H-4032 Debrecen, Hungary Email: nagyb80@gmail.com

Abstract

Background: Increased expression of the human epididymis protein 4 (HE4) was previously described in lung biopsy samples from cystic fibrosis (CF) patients, but it has remained unknown whether serum HE4 concentrations are elevated in CF.

Methods: Seventy-seven children with CF from six Hungarian CF centers and 57 adult CF patients from a Czech center were enrolled. In addition, 94 individuals with non-CF lung diseases, and 117 normal controls without pulmonary disorders were analyzed. Serum HE4 was measured by an immunoassay and its expression was further investigated via the quantification of HE4 mRNA using RT-qPCR in CF versus non-CF respiratory epithelium biopsies. The expression of the potential regulator miR-140-5p was analyzed using an UPL-based RT-qPCR assay. HE4 was measured in the supernatants from unpolarized and polarized cystic fibrosis bronchial epithelial (CFBE) cells expressing wt- or F508del-CFTR.

Results: Serum HE4 levels were significantly elevated in children with CF (99.5 [73.1-128.9] pmol/L) compared to controls (36.3 [31.1-43.4] pmol/L; P<0.0001). This observation was replicated in CF adults (115.7 [77.8-148.7] pmol/L; P<0.0001). In contrast, abnormal but lower HE4 concentrations were found in cases of severe bronchitis, asthma, pneumonia or bronchiectasis. In CF patients, the concentrations of HE4 were positively correlated with overall disease severity and C-reactive protein concentrations, while a significant inverse relationship was found between HE4 and the spirometric FEV₁ value. Relative HE4 mRNA levels were significantly upregulated (P=0.011) with a decreased miR-140-5p expression (P=0.020) in the CF versus non-CF airway biopsies. There were 2-fold higher HE4 concentrations in the supernatant of polarized F508del-CFTR CFBE cells compared to wt cells.

Conclusions: HE4 serum levels positively correlate with the overall severity of CF and the degree of pulmonary dysfunction. HE4 may thus be utilized as novel inflammatory biomarker and possibly also as measure of treatment efficacy in CF lung disease.

Introduction

Cystic fibrosis (CF) is an autosomal recessive disorders and is linked to a disease-causing variation in the CF transmembrane conductance regulator (*CFTR*) gene ¹. CF is a multisystem disorder in which abnormal ionic transport mediated by CFTR may lead to alteration of properties in the airway surface fluid and mucus ². The diagnosis of CF is based on typical clinical features associated with elevated sweat chloride concentrations, and the detection of 2 CF-causing *CFTR* mutations in *trans* ^{3,4}. More than 2000 genetic alterations have already been identified in the *CFTR* gene with the major p.Phe508del mutation accounting for approximately 70% of all CF alleles in Hungary and the Czech Republic ⁵⁻⁷. The inflammatory response in response to chronic pulmonary infections is related to a number of overexpressed cytokines and metalloproteinases that are considered to be major pathogenetic components in CF ⁸⁻¹².

Although several diagnostic guidelines have been published for CF within the last two decades ^{3,4,13}, few studies have employed potential serum biomarkers. These biomarkers may provide the desired quantifiable "endophenotype" which could be utilized to monitor disease severity. The clinical course of CF could then be indirectly assessed using e.g., increased serum concentrations of the C-reactive protein (CRP), interleukin (IL)-1β, and myeloperoxidase ¹⁴ that is linked to neutrophilia ¹⁵, while the measurement of plasma soluble CD14 ¹⁶, as well as CRP and plasma IL-8 ¹⁷ concentrations might be predictive of pulmonary exacerbations. Cytokine concentrations were also thoroughly analyzed in the bronchoalveolar lavage fluid (BALF) ^{9,18} and in sputum samples from CF patients ¹⁹. Moreover, tumor markers, such as cancer antigens (CA) 125 and CA 19-9, were connected with *Pseudomonas aeruginosa* lung colonization and impaired lung function ²⁰, while CA 19-9 may discriminate cases with borderline sweat chloride

concentrations²¹. However, assessment of various biomarkers has been primarily considered within the scientific domain, rather than as part of routine CF laboratory diagnostics.

The human epididymis protein 4 (HE4) is encoded by the WFDC2 gene ²² and is known to be a consistent tumor marker in epithelial ovarian cancers ^{23,24}, lung malignancies ^{25,26} and endometrial carcinomas ²⁷. This protein is a member of the whey acidic protein (WAP) family that is homologous to other serine proteinase inhibitors, comprising e.g., elafin and secretory leukocyte protease inhibitor (SLPI) ^{28,29}. However, in these mediators different control mechanisms, compared to HE4, have been suggested ¹⁴. Elafin and SLPI proteins have a major role in host defense of the respiratory tract via inhibition of neutrophil elastase (NE)²⁹ and by NFkB activation ³⁰. In this regard, increased SLPI levels were observed in acute respiratory distress syndrome ³¹, and in CF without any concurrent infection. However, SLPI is cleaved and down-regulated by NE in the presence of chronic bacterial infections ^{32,33}. Similarly to the WAP protein family, HE4 also displays a variety of functions. It acts as an anti-proteinase in the frame of epithelial host defenses of the respiratory tract ^{32,33}, and it is involved in sperm maturation ²². Although HE4 is overexpressed in the aforementioned tumors, it is also produced by other "normal" tissues (e.g., epididymis, salivary glands, prostate, etc.) at baseline levels ^{32,34}. The only report on a possible association of HE4 with CF utilized immunohistochemistry for the assessment of WFDC2 gene expression (hence HE4) in CF lung biopsy samples containing mainly tracheobronchial epithelial cells¹⁴. However, the authors did not analyze HE4 concentrations in blood specimens.

Here, we carried out a multicenter, analyst-blinded, case-control study of two unrelated CF populations of various age groups in which we analyzed serum HE4 concentrations. We also investigated the possible origin of abnormally high HE4 secretion into the blood via expression

analysis of the *WFDC2* gene in respiratory epithelium samples by measuring HE4-specific mRNA using RT-qPCR. Finally, we examined the level of miR-140-5p as one of the regulatory microRNAs of the *WFDC2* gene according to a miR database (<u>www.microrna.com</u>). To prove the overexpression of HE4 in connection with the CFTR-F508del genotype, supernatant culture medium samples from cystic fibrosis bronchial epithelial (CFBE) cells expressing wt- or F508del-CFTR were obtained for HE4 analysis as an *in vitro* complementation of the *ex vivo* measurements.

Material and Methods

Study participants

We summarized the main characteristics of all study groups (Table 1), and the demographic and laboratory parameters of CF study populations were presented in detail in the Supplementary Tables 1 and 2. Seventy-seven young CF patients were consecutively recruited from six Hungarian CF centers between October 2012 and February 2015. An independent adult cohort, comprising 57 adult CF patients was recruited from an adult CF centre in Prague. All CF subjects were diagnosed in accordance with the CF consensus diagnostic criteria [2] and exhibited the classic form of the disease. The overall clinical status was assessed by detailed clinical- and laboratory examination and "graded" by the Shwachman-Kulczycki (SK) score ⁸ in those centers where it was used, or based on the opinion of clinicians. Accordingly, we categorized the patients into these 3 subcohorts. CF individuals with available SK scores of 71-100 were classified as 'mild', those with 41-70 scores were in the 'moderate' category, and subjects with <40 scores were recruited into the 'severe' group. Serum samples were collected either during routine outpatient clinic visits (these CF patients had predominantly mild to moderate clinical course of CF) or during hospitalization due to e.g., acute pulmonary exacerbations

demonstrating increased cough with increased sputum production, difficulty in breathing, fatigue, and marked decline in lung function results; these criteria were based on Fuchs *et al* ³⁵. CRP concentrations were only analyzed in children with a severe disease course (n=29, 37.6%) and in all Czech adults. We also analyzed HE4 after medication at improved clinical states in 10 CF subjects who appeared at the Outpatient Clinic for a check-up after the recruitment into this study. These CF patients were treated by two types of intravenous antibiotics, mucolytic drugs, and other anti-inflammatory agents during hospitalization with an average period of 14 days. After discharge, these subjects took oral antibiotics for 3 weeks plus inhaled antibiotics for 3 months.

In order to assess the underlying non-specific effect of inflammation in other "non-CF" pulmonary diseases on HE4 serum concentrations, 64 patients with severe non-CF lung diseases (with chronic bronchitis, asthma or pneumonia). Furthermore, 12 other individuals who had non-CF bronchiectasis (n=2) or congenital disease (e.g. Werdnig-Hoffman-syndrome, Williams–Campbell syndrome, microcephaly, etc., n=7) or treatment resistant severe epilepsy (n=3), were also studied. These subjects suffered from severe recurrent lung infections leading to bronchiectasis due to repetitive aspiration and bronchial discharge retention. In addition, 18 non-CF adult subjects who had bronchiectasis or chronic obstructive pulmonary disease (COPD), were also recruited. These "clinical controls" were diagnosed using standard X-ray examinations and relevant laboratory tests. Exclusion criteria comprised a history of smoking, malignancy, immunodeficiency, chronic kidney, or liver disease. There was no significant difference in age between the CF and the clinical control cohorts, respectively; however, non-CF patients were younger than CF patients (P < 0.05) with a similar gender ratio (Table 1). Bronchial epithelial biopsies were obtained via diagnostic bronchoscopy from 3 Hungarian CF subjects (2 M/1 F; 3, 4, 9 years of age), and 3 other non-CF patients (2 M/1 F; 2, 4, 10 years of age) suffering from

repeated episodes of bronchitis (data not shown). Bronchoscopy was performed in the latter patients in order to reveal a background of wheezing and to exclude congenital stenosis of the main bronchi. Age- and sex-matched children and adult individuals without any apparent pulmonary disorders or chronic inflammatory processes were enrolled as random controls (Table 1). Finally, 12 apparently healthy parents of Hungarian CF patients (31 [25-37] years of age), who were all carriers of p.Phe508del mutation, were also examined for serum HE4 concentrations (data not shown).

Ethics statement

Written informed consent was obtained from the parent(s) or legal guardian of each child and adult carriers before the study. The study was approved by the Regional Ethics Committee of the University of Debrecen (DEOEC-RKEB/IKEB 3777-2012) and by the Local Ethics Committee of the Motol University Hospital, Prague.

Laboratory diagnosis of CF and the analysis of serum parameters

Sweat chloride values were analyzed using a Sweat Check Conductivity Analyzer (Wescor, USA). Genomic DNA isolation from blood leukocytes was performed with a QIAgen Blood Mini Kit (Qiagen, Germany). The 30 most common CFTR mutations in Hungary were tested using a Elucigene CF29v2 Kit (Tepnel-Diagnostics, UK) as well as an allele-specific PCR for the 'Slavic' CFTRdele2,3 (21kb) mutation, as previously described ⁶. The Czech adult patient cohort was genotyped by the Elucigene EU v3 assay (Tepnel-Diagnostics, UK) and the MLPA kit (MRC Holland, The Netherlands). Blood samples for HE4 were obtained by venous puncture, then centrifuged, and stored at -70 °C until analysis. Serum HE4 was determined through

chemiluminescent microparticle immunoassay (Architect i2000SR, Abbott, USA). Serum CRP was measured through electro-chemiluminescent immunoassay (Cobas e411, Roche, Germany).

Quantification of HE4 mRNA level in airway epithelium biopsies

Respiratory epithelial cells were kept in isotonic saline at 4 °C in sterile plastic tubes, and centrifuged at 1500 g for 5 min, and pellets were stored at -70 °C until analysis. RNA isolation was done using Trizol Reagent (Invitrogen, USA) according to the manufacturer's instructions. cDNA was synthesized using 200 ng of total RNA using a High Capacity cDNA Reverse Transcription Kit (Applied Biosystems, USA). RT-qPCR (Light Cycler 480, Roche) was used with the SYBR Green PCR Master Mix (Invitrogen) for quantification of the relative expression of the HE4 gene in airway epithelial samples. For further details, see Supplementary data.

Quantification of miR-140-5p expression in airway epithelium biopsies

MicroRNAs (miRNA) targeting WFDC2 (HE4) gene function were searched by using a prediction program (http://www.microrna.org). Since hsa-miR-140-5p was predicted by this algorithm as a putative regulator of HE4 gene, this miRNA was selected for further experiments. MiRNA was transcribed into cDNA and quantified via miRNA specific RT-PCR reaction using a TaqMan MicroRNA Reverse Transcription Kit (Life Technology, USA) according to the manufacturer's instructions. For further details, see Supplementary data.

HE4 analysis in the supernatants of CFBE cell cultures expressing wt or F508del CFTR

CFBE 41o- stably expressing either wt-CFTR or F508del-CFTR (a kind gift from Dr. JP Clancy, Dept. of Pediatrics, Cincinnati Children's Hospital, OH) were seeded in triplicate, either in plastic wells (250k cells per 3.8 cm² growth area) or on 12 mm collagen IV-coated Transwell permeable

supports (Corning Costar 3460: 250k cells per support) for monolayer formation and polarization, which was assessed by daily measurements of transepithelial resistance (TEER). Supernatant culture medium samples were taken when confluence (for unpolarized samples) or polarization (TEER > 600 Ω /cm²) was reached. The same cells were then treated with LPS (L2880, Sigma: 10 µg/ml, 24 h), following which supernatant was once more collected.

Statistical analysis

The Kolmogorov-Smirnov test was used for the evaluation of the normality of the data. Demographic and laboratory parameters were non-normally distributed thus being expressed as median (range), and were analyzed using the Mann-Whitney U test. Normally distributed data of RT-qPCR analyses were expressed as mean±SEM and were analyzed by Student's independent *t*test analysis. The Chi-square test was used to compare categorical variables. The Spearman's rho was calculated for correlation analysis. The discriminative power of HE4 and CRP was evaluated using receiver operating characteristics (ROC) curve analysis. P values < 0.05 were considered to be statistically significant. All analyses were performed using SPSS Statistics software, version 19.0 (IBM Corps., Armonk, NY, USA).

Results

Altered laboratory parameters of study populations

Sweat chloride levels were over 60 mmol/L in most CF individuals (median [range] 107 [90-120] mmol/L) except for 7 patients. Of note, this test was not performed in clinical controls and in the non-CF group since CF could be conveniently excluded using other tests. Serum CRP was increased (reference range: <4.6 mg/L) compared to normal in those young CF subjects with a severe clinical status at admission (n=29), in all CF adults and in the groups of non-CF controls

(Table 1). In addition, chronic bacterial colonization was detected in 54 young CF and in all adult CF individuals. Pancreatic insufficiency and CF-related diabetes mellitus (DM) were diagnosed in variable percentages of CF cohorts (Supplementary Tables 1 and 2).

Highly increased level of serum HE4 in CF regardless of genetic alteration compared to controls and non-CF subjects

Median serum HE4 levels were significantly (P<0.0001) elevated in the 77 CF children and 57 adult patients with CF compared to normal controls (Table 1 and Fig. 1A). Twenty-nine CF young subjects in severe condition had even higher HE4 concentrations (134.9 [124.5-275.0] pmol/L). In the adult CF cohort (n=57), there were 13 patients suffering from severe status (170.8 [146.8-182.9] pmol/L). Gender did not affect HE4 levels in either CF group: in children 95.4 [75-128.5] pmol/L in males vs. 101.5 [72.6-129.6] pmol/L in females; P=0.881, while in adulthood median HE4 was 119.4 [85.2-150.6] pmol/L in males; 114.8 [75.3-139.1] pmol/L in females; P=0.290 (data not shown). In terms of CF genetic background, we did not detect differences with variable genotypes among children: homozygous for p.Phe508del median HE4 was 99.5 [73.0-127.5] pmol/L vs. in the presence of other mutations 94.6 [66.4-127.8] pmol/L, and there was no change in HE4 of CF adults between those with p.Phe508del/p.Phe508del (123.1 [74.1-151.1] pmol/L) and others with a distinct genetic background (108.3 [83.9-140.0] pmol/L; P=0.348) (data not shown). We also analyzed HE4 concentrations in additional patients who suffered from severe non-CF lung diseases, such as bronchitis, asthma, pneumonia or bronchiectasis. Increased median HE4 level was measured in these cases (P<0.001) compared to normal controls, however, these data were much below the results observed in CF (Fig. 1A). More importantly, there was a statistically significant difference (P<0.001) in serum HE4 between CF and non-CF patients regardless of age. When we sub-grouped non-CF pulmonary young patients based on the types of lung disease, similar changes were seen in severe bronchitis or asthma (63.7 [54.1-79.5] pmol/L; n=42) and pneumonia (57.8 [51.2-74.2] pmol/L; n=22) (data not shown). Furthermore, we enrolled children suffered from non-CF bronchiectasis due to different clinical conditions showing similar HE4 values (80.6 [68.1-134.5] pmol/L) than CF; however, median HE4 level was much higher in CF patients. For CF adults, 18 non-CF patients were recruited with moderately elevated HE4 (59.2 [41.1-88.6] pmol/L). In contrast, the carriers of CF mutations had normal HE4 levels (33.9 [30.1-43.9] pmol/L; n=12). When we determined HE4 concentration in sputum from CF or other non-CF diseases, - similarly to sera - there were significantly higher HE4 concentrations in CF versus non-CF or healthy sputum specimens (see Supplementary data and Table 3).

Serum HE4 in CF is not affected by age and strongly correlates with disease severity

No significant difference in HE4 was observed among different age groups (P=0.164) (Fig. 1B). Age did not seem to have an impact on HE4 levels in these CF patients. In order to investigate if serum HE4 can be used as a prognostic disease marker in CF, the relationship between HE4 levels and disease severity was investigated. HE4 showed a positive trend that followed the degree of impairment in both disease cohorts (Figs. 2A and B). Significantly higher HE4 levels were seen in CF subjects with a moderate status (108.0 [95.8-133.0] pmol/L; P<0.001) compared to those with a mild status (60.8 [50.7-73.8] pmol/L). Even higher HE4 values were measured in those in severe condition with an acute exacerbation (134.9 [124.5-275.0] pmol/L; P<0.0001) (Fig. 2A). In adult CF, there was a positive correlation between HE4 and disease status: mild (72.5 [64.1-94.8] pmol/L), moderate (126.5 [104.3-148.3] pmol/L), and severe clinical symptoms (170.8 [146.8-182.9) pmol/L (Fig. 2B). As a preliminary evaluation of HE4 in the follow-up of CF, we analyzed HE4 after medication at improved clinical states in 10 CF subjects. Accordingly,

there were lower concentrations of serum HE4 (92.5 [87.5-106.2] pmol/L vs. 125.8 [111.8-153.9] pmol/L at baseline; P=0.105) after treatment, though results did not reach the range of normal controls (data not shown). Of special interest, we recognized a significant negative association between HE4 and FEV₁ in CF, which was used to evaluate residual lung function capacity (Spearman's rho=-0.522; P<0.0001) (Fig. 3A).

The association between serum HE4 and CRP in CF

We sought to analyze if changes in HE4, acting as an additional inflammatory parameter, was closely related to the increased levels of CRP. There was a significant and positive correlation between serum HE4 and CRP levels in CF children in severe condition plus all CF adults (Spearman's rho=0.595; P<0.001) (Fig. 3B); while no association was observed in patients with non-CF lung diseases (Spearman's rho=0.168; P=0.074) (data not shown). Based on these data, increased HE4 can signal massive pulmonary inflammation in CF similarly to CRP. When we further compared the laboratory characteristics of HE4 statistically to that of CRP in CF, HE4 showed a substantial AUC value of 0.724 (95% CI: 0.617-0.830) (P<0.0001), while CRP had an average AUC value of only 0.453 (95% CI: 0.347-0.559) (P=0.435). Overall, serum HE4 is able to track the development of clinically significant lung inflammation in CF with much higher accuracy than CRP.

The association of serum HE4 with chronic bacterial colonization and CF-related comorbidities

All adult CF participants had bacterial colonization, thus we could not analyze the relationship of HE4 with chronic bacterial infection, while children with positive microbiology tests had significantly higher levels of HE4 as compared to those without bacterial colonization (106.5 [75.8-133.3] pmol/L vs. 78.3 [61.4-96.9] pmol/L; P=0.001). In the presence or absence of CF-

related DM, no substantial alteration was seen in HE4 (DM: 102.6 [97.8-121.5] pmol/L vs. non-DM: 91.1 [64.7-128.9] pmol/L; P=0.645). In contrast, pancreatic insufficiency in adults was associated with higher serum HE4 values than those with normal pancreatic function (122.2 [84.1-149.6] pmol/L vs. 96.8 [73.8-108.8] pmol/L; P=0.037) (Supplementary Tables 1 and 2).

The diagnostic characteristics of HE4 in CF

We also studied the potential discriminative power of HE4 in the entire study population using ROC analyses. Based on the values of the entire CF group, the AUC for HE4 was 0.993 (95% CI: 0.986-0.999) (P<0.0001) at the 49.3 pmol/L cut-off value for differentiating CF patients from normal controls (Fig. 4). The sensitivity, specificity, positive predictive value (PPV), and negative predictive value (NPV) for HE4 were 97.0%, 96.6%, 66.5%, and 96.6%, respectively. When the AUC for HE4 was calculated for the discrimination of CF from other non-CF pulmonary diseases, this value was slightly lower at 0.778 (95% CI: 0.716-0.841) (P<0.0001) with a modest sensitivity of 56.0%, but a high specificity of 90.0% at the 98.4-pmol/L cut-off value with 64.6% PPV and 90.4% NPV (Fig. 4). In addition, we intended to observe if sweat chloride test results showed any relationship with serum HE4. Interestingly, a positive and significant correlation was seen between the actual level of serum HE4 and the sweat chloride values that were previously measured at the diagnosis of CF children (Spearman's rho=0.345; P=0.005) (data not shown). In a subgroup analysis, we analyzed HE4 in those patients (n=8) who had borderline sweat chloride test results (30-60 mmol/L) with predominantly mild/moderate clinical conditions. We found similar serum HE4 values in these cases (100.9 [88.2-115.0] pmol/L) as seen in the entire CF cohort. In summary, serum HE4 was already abnormal when the 'gold standard' sweat chloride test was only intermediate. Since this was a relatively low number of cases, further evaluation of HE4 is necessary in CF with borderline chloride results.

Increased level of HE4 mRNA was detected in respiratory epithelium biopsies of CF

Next, we wanted to investigate the origin of elevated HE4 in CF sera. The goal was to determine if HE4 was a consequence of the destruction of the airway epithelium causing an extensive release of HE4 due to the massive inflammation, or this protein was produced at a higher quantity in the airways. For this purpose, human bronchial epithelium biopsies were obtained via bronchoscopy from 3 CF and 3 non-CF patients for controls. HE4 mRNA level in CF was significantly upregulated compared to that of non-CF control subjects (HE4/36B4 ratio: (mean±SEM) 0.354±0.178 vs. 0.041±0.017; P=0.011; n=3 per group) suggesting that highly elevated serum HE4 concentrations in CF may result from its increased secretion by the airway epithelium (Fig. 5A).

The relationship of miR-140-5p with the expression of HE4 in CF

The level of miR-140-5p was analyzed in the same bronchial epithelial biopsies mentioned above from CF and non-CF individuals, and was normalized to RNU43. We observed that miR-140-5p expression in CF was significantly decreased compared to non-CF control subjects (miR-140-5p/RNU43 ratio: (mean±SEM) 0.033±0.008 vs. 0.443±0.076; n=3 per group; P=0.020) (Fig. 5B). In order to recognize if there was a strong association with HE4 mRNA and miR-140-5p expression, a Spearman's correlation was performed, which revealed a significant inverse correlation between miR-140-5p and HE4 mRNA levels (Spearman's rho: -0.744, P=0.006) (data not shown).

F508del-CFTR expressing CFBE cells produced markedly higher HE4 level than those expressing wt-CFTR

HE4 levels in the supernatants of unpolarized and polarized wt- and F508del-CFTR expressing CFBE cells were compared. We found that there were about 2-fold higher HE4 concentrations in

the supernatants of F508del-CFTR CFBE cells, and the difference was more pronounced in polarized cultures compared to unpolarized cells (Fig. 6). Moreover, these cultures were treated with LPS (10 μ g/mL) for 24 hours in order to analyze the direct effect of bacterial infection on HE4 production. We did not, however, observe any induction in HE4 secretion by this stimulation (data not shown).

Discussion

The lungs are usually most affected in CF, and the patient suffers from recurrent bacterial infections and severe inflammation causing destruction in the bronchi, which leads to a reduced life expectancy ⁹. The criteria for the diagnosis of this disease are a positive family history and/or abnormal newborn screening test results, typical clinical features, increased sweat chloride test, and the identification of 2 CFTR mutations in trans ⁴. However, there are challenging cases that have borderline sweat chloride values or when the genetic analysis reveals zero or only one CFTR mutation or mutation(s) with unknown clinical significance. To solve this situation, Sermet-Gaudelus *et al.* suggested the use of the nasal potential difference diagnostic score, which was found to be a useful tool for inconclusive CF patients ³⁶. Although there are several potential blood-based biomarkers for monitoring CF subjects and to identify those who are at a risk of acute pulmonary exacerbation, none of them, as yet, have been integrated into routine clinical practice ³⁷. Serum MPO, IL-1β, and CRP ¹⁵, plasma soluble CD14 ¹⁶, and plasma IL-8 ¹⁷ can predict pulmonary exacerbation effectively, but only at early time points. No significant association between CRP and the severity index of the disease was observed ³⁸.

HE4 was suggested as a tumor marker for epithelial ovarian cancer ²³. Since then, a large number of papers supported the superior diagnostic characteristics of HE4 versus CA 125 in this particular malignancy ^{24,39,40} with other possibilities we summarized in a recent review ⁴¹. In

addition, HE4 may be elevated in certain non-cancer diseases, such as chronic kidney disease ⁴², or in pulmonary tuberculosis where only moderately increased HE4 concentrations were reported ⁴³. HE4 was formerly investigated but only in regard to one aspect of CF, i.e., when high HE4 expression was described in CF lung biopsies by immunohistochemistry ¹⁴. No evidence regarding serum HE4 level changes relative to this disease was provided.

We found that serum HE4 levels were significantly elevated in both young and adult CF patients compared to normal controls. There was no difference in HE4 levels between males and females, similarly to sweat chloride values as previously described ⁴⁴. There were a number of increased HE4 values in non-CF patients especially in bronchiectasis that partially overlapped with a ratio of CF results measured in CF patients, however; the median HE4 concentration was much lower than that of CF (Fig. 1A). Within the non-CF group, no significant difference was observed when patients were further separated based on subtypes of lung disease. Since the non-CF children cohort consisted of younger patients as compared to CF children, we reanalyzed the difference in HE4 between CF children and a subgroup of age-matched non-CF children cohorts (7.5 [5.0-14.8] years of age), and there were significantly lower HE4 results (56.7 [46.4-71.6] pmol/L, P<0.0001) in this control group (n=49) in contrast to CF. Very importantly, heterozygous healthy parents of some of the studied CF patients showed similar HE4 levels to the controls, which is in good agreement with the recessive inheritance of the disease.

In the adult CF study group, CF patients showed the same HE4 results. We then investigated the effect of age on HE4 levels since age could have been a major factor for HE4 ^{41,45}. Among our CF patients, there was no substantial impact of age on HE4 results (Fig. 1B) in contrast to sweat chloride test results that undergo an age-dependent elevation, as exposed in an earlier study ⁴⁶.

Notably, serum HE4 level tightly correlated with bacterial colonization, and pancreatic insufficiency.

HE4 was able to act as a prognostic disease marker in CF when patients were categorized by variable degree of severity (Figs. 2A and B). HE4 showed a positive correlation with the degree of lung impairment. In this aspect, HE4 was proportionally elevated due to advanced inflammatory events. It is noteworthy to mention that this phenomenon is similar to pathologic stages in cancers ^{26,40}.

Previous reports discussed the potential role of CRP in the prediction and detection of pulmonary exacerbations with relatively good efficacy ^{15,17,38}. However, this parameter has yet to become a routinely used examination in CF. CRP results were abnormal but with a relatively modest change in most cases. As most adult patients were infected with *P. aeruginosa* or *B. cepacia complex*, higher CRP levels were expected even in the stable phase of the disease in these patients [35]. Then we compared the diagnostic characteristics of HE4 to CRP. Although HE4 showed a larger variation, a positive significant correlation was analyzed between serum HE4 and CRP levels in both children and adult study groups. Notably, HE4 in CF had a better diagnostic power according to their AUC values in contrast to CRP. In non-CF subjects, this HE4-CRP association could not be detected. Therefore, serum HE4 can be considered as another candidate biomarker for the indication of the accumulation or recurrence of pulmonary inflammation and pulmonary exacerbation of CF.

Treatment options for CF are limited; however, one drug, ivacaftor, has become clinically important and has been shown to be very beneficial in some mutations. In clinical studies, treatment efficacy was measured using FEV₁ and sweat chloride ⁴⁷. Testing would also benefit

from the use of a biomarker. We believe that measurement of HE4 might serve as a potential biomarker for the analysis of treatment in patient follow-up.

The diagnostic characteristics of HE4 were also evaluated using ROC analyses. The AUC for HE4 was 0.993 (95% CI: 0.986-0.999) at 49.3 pmol/L cut-off value for differentiating CF patients from controls with substantial values of sensitivity, specificity, PPV, and NPV. When the AUC for HE4 was calculated for the discrimination of CF from other non-CF pulmonary diseases, this value was slightly lower 0.778 (95% CI: 0.716-0.841). If one compares HE4 to sweat chloride analysis, the 'gold standard' test had a much higher PPV (77.1%), but worse NPV (54.8%) at 39 mEq/L based on Seia *et al.* ⁴⁴. We were curious about the relationship between sweat chloride test results and the concentrations of serum HE4. There was a positive significant correlation between these parameters in all CF subjects. Serum HE4 was already abnormal when the 'gold standard' sweat chloride test was only intermediate.

The origin of increased serum HE4 in CF and other non-cancer diseases has not been studied previously. Therefore, HE4 specific mRNA levels in bronchial epithelial biopsies from CF were analyzed. In CF, HE4 mRNA levels were significantly elevated compared to non-CF control samples; suggesting that high serum HE4 concentrations were a manifestation of increased production in the lungs. We recently found WFDC2 among the upregulated genes in a study of gene expression in native nasal epithelium of CF that supports the validity of this present data ⁴⁸. Similarly, there were several previous reports on modulated gene expression in mild and severe CF due to inflammation with or without bacterial infection or the presence of CFTR mutations ⁴⁹⁻⁵¹. Up-regulated genes in mild CF targeted lipid metabolism, G-protein coupled receptor expression and ion transports, while in severe CF altered genes were involved in oxidoreductase activity and the ubiquitin cycle ⁵⁰. In terms of classic CF mutations, the p.Phe508del mutation

was shown to have a minor effect on other gene expression profiles in cell cultures of CF airway epithelium cells compared to non-CF cells ⁵¹. However, these authors used very stringent cut-off statistics as compared to other studies to choose their gene list.

HE4 levels in the culture medium supernatants of unpolarized and polarized wt and F508del-CFTR CFBE cells were compared. We found that there were about 2-fold higher HE4 concentrations in the supernatant of F508del-CFTR CFBE cells in case of polarized cultures as compared to wt cells (Fig. 6). Moreover, these cultures were treated with LPS (10 µg/mL) for 24 hours in order to analyze the direct effect of bacterial infection on HE4 production. We did not observe any induction in HE4 secretion by this stimulation. This later data are in agreement with the results of Bingle *et al.* ¹⁴ who did not find an alteration in HE4 expression of LPS-treated tracheobronchial epithelial cells or type II pneumocytes. Overall, in CF endogenous CFTR deficiency may be the main factor for HE4 overexpression, while intrapulmonary inflammation/infection may be supplemental in HE4 secretion.

Certain cytokines such as IL-8 are present with increased concentrations in the BALF obtained from CF patients, while others (e.g. IL-10) are at relatively lower quantity versus non-CF samples ⁵². For instance, elevated levels of IL-8 have been shown to be selectively produced by airway epithelium and recruited neutrophils in CF ⁵³, with neutrophil elastase being a key mediator of its production ⁵⁴. A quantitative gene expression analysis was processed where IL-8 mRNA level was increased in *ex vivo* bronchial epithelial cells ⁴⁹. By measuring the expression of other genes, additional explanations for the development of CF phenotype could be revealed in connection with e.g. airway defense ⁵⁰. Recently, miRNAs with aberrant expression seem to be potential regulators of the disease pathomechanism ⁵⁵. Out of hundreds of miRNAs, 56 showed decreased levels, while 36 were up-regulated in CF versus non-CF samples ⁵⁵. Notably, miR-126 was found

to be down-regulated in CF airway epithelial cells that regulate the innate immune (TH₂) response via TOM1 expression in the TLR2/4 signaling pathway ⁵⁶. miR-155 was responsible for the overexpression of IL-8, and bacterial infection promoted the expression of other miRNAs such as miR-215 ⁵⁷.

In this study, we analyzed the level of miR-140-5p in connection with CF. This particular microRNA showing a high nucleotide complementarity to the HE4 specific mRNA target, has been recently studied in several types of malignancies, such as in non-small cell lung cancer ⁵⁸. Based on those data, the monocyte to macrophage differentiation-associated (MMD) gene was targeted by miR-140-5p that enhanced proliferation of lung tumor cells via ERK signaling ⁵⁸. According to a miRNA-related database, miR-140-5p was predicted to be associated with WFDC2/HE4 gene expression. We found that the level of miR-140-5p in respiratory epithelium biopsies was significantly decreased compared to non-CF control individuals, and was negatively associated with HE4 mRNA levels. Overall, lower levels of miR-140-5p - at least in part - may contribute to the exaggerated production of HE4 protein via higher HE4 mRNA expression in airway epithelium. Further investigations are needed to prove this relationship.

HE4 exhibited a proteinase inhibitory activity towards trypsin, elastase, matrix metallopeptidase 9, and the secretory proteinases from *Bacillus subtilis*⁵⁹. Accordingly, we suppose that HE4 as an immunological response to bacterial/inflammatory events may show an inhibitory effect in this disease. Moreover, HE4 was described as a fibroblast-derived mediator of kidney fibrosis⁶⁰. Accordingly, we speculate that increased HE4 may also contribute to fibrotic degradation of the lung in CF. Further experiments are required to support these theories.

The finding that HE4, a well-described tumor marker, is found up-regulated in CF, may be linked to a less differentiated epithelial state previously described in CF versus non-CF cells ⁶¹. Indeed,

our own meta-analysis of data of gene expression profile revealed that CF nasal cells' have a significant overlap with genes related to undifferentiated epithelial phenotype ⁶².

In conclusion, serum HE4 is elevated in CF and correlates with the severity of the disease, thus it has the potential to be utilized as a novel prognostic biomarker that detects pulmonary inflammation. However, further studies are needed to clarify if released HE4 plays any functional role relative to IL-8, NE or any metalloproteinases in the regulation of local inflammatory lung events of CF patients.

Acknowledgements

Author contributions. Béla Nagy Jr takes responsibility for the content of the manuscript, including the data and analysis. BNJr designed and performed experiments and wrote the manuscript, BN, LF, FG, OB, DN, RU, AS, AA, MM, and ZB provided human samples and analyzed data, ZF, LAC performed experiments, PAS designed experiments, HBP statistically analyzed data, JK, GB, MDA, MMJr, and IB provided overall direction, critical revisions for intellectual content, and providing final approval of the version to be published. The authors are grateful to Róza Földesi and Imréné Gém for their excellent technical assistance in RT-qPCR experiments and HE4 measurements, respectively. The authors thank Ines Pankonien for technical assistance with cell culture.

Financial disclosures: This research was supported by the European Union and the State of Hungary, co-financed by the European Social Fund in the framework of TÁMOP-4.2.4.A/2-11/1-2012-0001 'National Excellence Program', and the TÁMOP-4.2.2.A-11/1/KONV-2012-0045 project. Béla Nagy Jr was supported by a Lajos Szodoray Grant of the University of Debrecen. This study was supported by the Hungarian Research Fund (K109076 to IB) and the Czech

Ministry of Health Conceptual Development of Research Organization (00064203), European Regional Development Fund Prague (CZ.2.16/3.1.00/24022OPPK, NT/13770-4/2012, Norway Grants -NF-CZ11-PDP-3-003-2014 and COST-LD14073 to MM). The LAC/MDA laboratory was supported by UID/MULTI/04046/2013 centre grant (to BioISI) and the research grant (to MDA) FCT/MCTES (PTDC/BIM-MEC/2131/2014), Portugal.

Figure legends

Fig. 1. Serum HE4 levels in CF, non-CF, and normal controls (A). There were significantly increased HE4 concentrations in CF children compared to both controls and non-CF individuals; however, some elevation was also seen in non-CF subjects versus controls. Investigation of the effect of age on HE4 levels (B). The following HE4 concentrations were found in these subcohorts: 1-4 years of age (n=11): 122.7 [80.7-130.8] pmol/L; 5-9 years of age (n=19): 113.7 [82.6-190.3] pmol/L; 10-14 years of age (n=21): 91.1 [75.0-121.6] pmol/L; 15-18 years of age (n=26): 83.0 [64.4-112.1] pmol/L; 19-22 years of age (n=20): 107.6 [80.5-142.1] pmol/L; 23-29 years of age (n=24): 120.7 [81.1-120.7] pmol/L; 30-40 years of age (n=13): 115.7 [77.8-146.9] pmol/L. We did not find a significant difference in HE4 between these age groups.

Fig. 2. Correlation of HE4 with the clinical status of CF patients. CF children were categorized into 3 groups at the time of study recruitment showing mild (n=30), moderate (n=18), and severe (n=29) symptoms (A). Similar categorization was done in adults with CF with mild (n=22), moderate (n=22), and severe (n=13) states (B). Serum HE4 gradually increased in relative worsening conditions.

Fig. 3. The relationship between HE4 and FEV_1 (A) and serum CRP (B) values. There was a statistically significant inverse association (P<0.0001) between HE4 and FEV_1 values suggesting that HE4 was highly correlated with the degree of lung dysfunction (A). In contrast, HE4 was strongly associated wit CRP parameters in CF (B).

Fig. 4. ROC analysis of HE4 in the differentiation of CF from normal controls (with triangles) and CF from other non-CF lung diseases (with full circles). Substantial AUC values were calculated, and the 49.3 pmol/L cut-off value differentiated CF patients from normal controls, while the 98.4 pmol/L cut-off value was determined to distinguish CF from other pulmonary conditions.

Fig. 5. Analysis of the level of HE4 mRNA in bronchial epithelium biopsies obtained via bronchoscopy from 3 CF and 3 non-CF patients (A). RT-qPCR was assessed to quantify and compare HE4 mRNA expression. We found that HE4 mRNA concentrations were significantly up-regulated in CF versus non-CF control samples. HE4 mRNA level was normalized to the expression of the reference gene 36B4. Data are expressed in this ratio in mean±SEM. Analysis of relative miR-140-5p expression in respiratory epithelium biopsies obtained from CF and non-CF patients. The miRNA level was normalized to RNU43. We observed a significantly decreased level of miR-140-5p in CF compared to non-CF control samples. Data are expressed as a ratio to RNU43 in mean±SEM (B).

Fig. 6. The measurement of HE4 level in the supernatants of unpolarized and polarized wt and F508 deficient CFBE cells. There was about 2-fold higher HE4 level in the supernatant of F508del-CFTR cells in polarized cultures as compared to wt cells. n=3 per each type of sample.

References

- 1 Cohen-Cymberknoh M, Kerem E, Ferkol T, et al. Airway inflammation in cystic fibrosis: molecular mechanisms and clinical implications. *Thorax*. 2013;68(12):1157-1162
- 2 Rowe SM, Miller S, Sorscher EJ. Cystic fibrosis. N Engl J Med. 2005;352(19):1992-2001
- 3 Rosenstein BJ, Cutting GR. The diagnosis of cystic fibrosis: a consensus statement. Cystic Fibrosis Foundation Consensus Panel. *J Pediatr*. 1998;132(4):589-595
- 4 Voter KZ, Ren CL. Diagnosis of cystic fibrosis. *Clin Rev Allergy Immunol*. 2008;35(3):100-106
- 5 Ivády G, Koczok K, Madar L, et al. Molecular analysis of cystic fibrosis patients in Hungary – an update to the mutational spectrum. J Med Biochem. 2015;34(1):46-51
- 6 Ivady G, Madar L, Nagy B, et al. Distribution of CFTR mutations in Eastern Hungarians: relevance to genetic testing and to the introduction of newborn screening for cystic fibrosis. *J Cyst Fibros*. 2011;10(3):217-220
- 7 Krenkova P, Piskackova T, Holubova A, et al. Distribution of CFTR mutations in the Czech population: positive impact of integrated clinical and laboratory expertise, detection of novel/de novo alleles and relevance for related/derived populations. J Cyst Fibros. 2013;12(5):532-537
- 8 Bodnar R, Kadar L, Holics K, et al. Factors influencing quality of life and disease severity in Hungarian children and young adults with cystic fibrosis. *Ital J Pediatr*. 2014;4050
- 9 Bergin DA, Hurley K, Mehta A, et al. Airway inflammatory markers in individuals with cystic fibrosis and non-cystic fibrosis bronchiectasis. J Inflamm Res. 2013;61-11
- 10 Brazova J, Sediva A, Pospisilova D, et al. Differential cytokine profile in children with cystic fibrosis. *Clin Immunol*. 2005;115(2):210-215
- 11 Brazova J, Sismova K, Vavrova V, et al. Polymorphisms of TGF-beta1 in cystic fibrosis patients. *Clin Immunol*. 2006;121(3):350-357
- 12 Courtney JM, Ennis M, Elborn JS. Cytokines and inflammatory mediators in cystic fibrosis. J Cyst Fibros. 2004;3(4):223-231
- 13 Smyth AR, Bell SC, Bojcin S, et al. European Cystic Fibrosis Society Standards of Care: Best Practice guidelines. *J Cyst Fibros*. 2014;13 Suppl 1S23-42
- 14 Bingle L, Cross SS, High AS, et al. WFDC2 (HE4): a potential role in the innate immunity of the oral cavity and respiratory tract and the development of adenocarcinomas of the lung. *Respir Res.* 2006;761
- 15 Pereira LC, Moreira EA, Bennemann GD, et al. Influence of inflammatory response, infection, and pulmonary function in cystic fibrosis. *Life Sci.* 2014;109(1):30-36
- 16 Quon BS, Ngan DA, Wilcox PG, et al. Plasma sCD14 as a biomarker to predict pulmonary exacerbations in cystic fibrosis. *PLoS One*. 2014;9(2):e89341
- 17 Wojewodka G, De Sanctis JB, Bernier J, et al. Candidate markers associated with the probability of future pulmonary exacerbations in cystic fibrosis patients. *PLoS One*. 2014;9(2):e88567

- 18 Fantino E, Gangell CL, Hartl D, et al. Airway, but not serum or urinary, levels of YKL-40 reflect inflammation in early cystic fibrosis lung disease. BMC Pulm Med. 2014;1428
- 19 Colombo C, Costantini D, Rocchi A, et al. Cytokine levels in sputum of cystic fibrosis patients before and after antibiotic therapy. *Pediatr Pulmonol*. 2005;40(1):15-21
- 20 Gronowitz E, Pitkanen S, Kjellmer I, et al. Association between serum oncofetal antigens CA 19-9 and CA 125 and clinical status in patients with cystic fibrosis. *Acta Paediatr.* 2003;92(11):1267-1271
- 21 Augarten A, Berman H, Aviram M, et al. Serum CA 19-9 levels as a diagnostic marker in cystic fibrosis patients with borderline sweat tests. *Clin Exp Med*. 2003;3(2):119-123
- 22 Kirchhoff C, Habben I, Ivell R, et al. A major human epididymis-specific cDNA encodes a protein with sequence homology to extracellular proteinase inhibitors. *Biol Reprod.* 1991;45(2):350-357
- 23 Hellstrom I, Raycraft J, Hayden-Ledbetter M, et al. The HE4 (WFDC2) protein is a biomarker for ovarian carcinoma. *Cancer Res.* 2003;63(13):3695-3700
- 24 Moore RG, McMeekin DS, Brown AK, et al. A novel multiple marker bioassay utilizing HE4 and CA125 for the prediction of ovarian cancer in patients with a pelvic mass. *Gynecol Oncol*. 2009;112(1):40-46
- 25 Iwahori K, Suzuki H, Kishi Y, et al. Serum HE4 as a diagnostic and prognostic marker for lung cancer. *Tumour Biol*. 2012;33(4):1141-1149
- 26 Nagy B, Bhattoa HP, Steiber Z, et al. Serum human epididymis protein 4 (HE4) as a tumor marker in men with lung cancer. *Clin Chem Lab Med*. 2014;52(11):1639-1648
- 27 Bignotti E, Ragnoli M, Zanotti L, et al. Diagnostic and prognostic impact of serum HE4 detection in endometrial carcinoma patients. Br J Cancer. 2011;104(9):1418-1425
- 28 Clauss A, Lilja H, Lundwall A. A locus on human chromosome 20 contains several genes expressing protease inhibitor domains with homology to whey acidic protein. *Biochem J*. 2002;368(Pt 1):233-242
- 29 Schalkwijk J, Wiedow O, Hirose S. The trappin gene family: proteins defined by an N-terminal transglutaminase substrate domain and a C-terminal four-disulphide core. *Biochem J*. 1999;340 (Pt 3)569-577
- 30 Taggart CC, Greene CM, McElvaney NG, et al. Secretory leucoprotease inhibitor prevents lipopolysaccharide-induced IkappaBalpha degradation without affecting phosphorylation or ubiquitination. *J Biol Chem.* 2002;277(37):33648-33653
- 31 Sallenave JM, Donnelly SC, Grant IS, et al. Secretory leukocyte proteinase inhibitor is preferentially increased in patients with acute respiratory distress syndrome. *Eur Respir J.* 1999;13(5):1029-1036
- 32 Bingle L, Singleton V, Bingle CD. The putative ovarian tumour marker gene HE4 (WFDC2), is expressed in normal tissues and undergoes complex alternative splicing to yield multiple protein isoforms. *Oncogene*. 2002;21(17):2768-2773
- 33 Weldon S, McNally P, McElvaney NG, et al. Decreased levels of secretory leucoprotease inhibitor in the Pseudomonas-infected cystic fibrosis lung are due to neutrophil elastase degradation. *J Immunol*. 2009;183(12):8148-8156

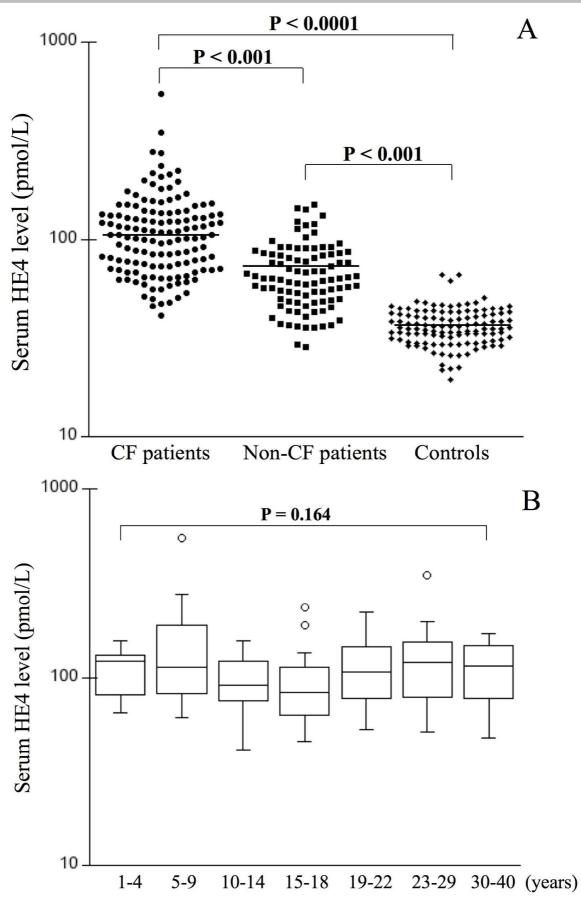
- 34 Galgano MT, Hampton GM, Frierson HF, Jr. Comprehensive analysis of HE4 expression in normal and malignant human tissues. *Mod Pathol.* 2006;19(6):847-853
- 35 Fuchs HJ, Borowitz DS, Christiansen DH, et al. Effect of aerosolized recombinant human DNase on exacerbations of respiratory symptoms and on pulmonary function in patients with cystic fibrosis. The Pulmozyme Study Group. *N Engl J Med.* 1994;331(10):637-642
- 36 Sermet-Gaudelus I, Girodon E, Sands D, et al. Clinical phenotype and genotype of children with borderline sweat test and abnormal nasal epithelial chloride transport. *Am J Respir Crit Care Med.* 2010;182(7):929-936
- 37 Shoki AH, Mayer-Hamblett N, Wilcox PG, et al. Systematic review of blood biomarkers in cystic fibrosis pulmonary exacerbations. *Chest.* 2013;144(5):1659-1670
- 38 Giron-Moreno RM, Justicia JL, Yamamoto S, et al. Role of C-reactive protein as a biomarker for prediction of the severity of pulmonary exacerbations in patients with cystic fibrosis. *BMC Pulm Med.* 2014;14(1):150
- 39 Lenhard M, Stieber P, Hertlein L, et al. The diagnostic accuracy of two human epididymis protein 4 (HE4) testing systems in combination with CA125 in the differential diagnosis of ovarian masses. *Clin Chem Lab Med*. 2011;49(12):2081-2088
- 40 Moore RG, Brown AK, Miller MC, et al. The use of multiple novel tumor biomarkers for the detection of ovarian carcinoma in patients with a pelvic mass. *Gynecol Oncol*. 2008;108(2):402-408
- 41 Kappelmayer J, Antal-Szalmas P, Nagy B, Jr. Human epididymis protein 4 (HE4) in laboratory medicine and an algorithm in renal disorders. *Clin Chim Acta*. 2014;438C35-42
- 42 Nagy B, Jr., Krasznai ZT, Balla H, et al. Elevated human epididymis protein 4 concentrations in chronic kidney disease. *Ann Clin Biochem*. 2012;49(Pt 4):377-380
- 43 Liu W, Yang J, Chi PD, et al. Evaluating the clinical significance of serum HE4 levels in lung cancer and pulmonary tuberculosis. *Int J Tuberc Lung Dis.* 2013;17(10):1346-1353
- 44 Seia M, Costantino L, Paracchini V, et al. Borderline sweat test: Utility and limits of genetic analysis for the diagnosis of cystic fibrosis. *Clin Biochem.* 2009;42(7-8):611-616
- 45 Moore RG, Miller MC, Eklund EE, et al. Serum levels of the ovarian cancer biomarker HE4 are decreased in pregnancy and increase with age. *Am J Obstet Gynecol.* 2012;206(4):349 e341-347
- 46 Mishra A, Greaves R, Smith K, et al. Diagnosis of cystic fibrosis by sweat testing: age-specific reference intervals. *J Pediatr*. 2008;153(6):758-763
- 47 Ramsey BW, Davies J, McElvaney NG, et al. A CFTR potentiator in patients with cystic fibrosis and the G551D mutation. *N Engl J Med*. 2011;365(18):1663-1672
- 48 Clarke LA, Sousa L, Barreto C, et al. Changes in transcriptome of native nasal epithelium expressing F508del-CFTR and intersecting data from comparable studies. *Respir Res.* 2013;1438

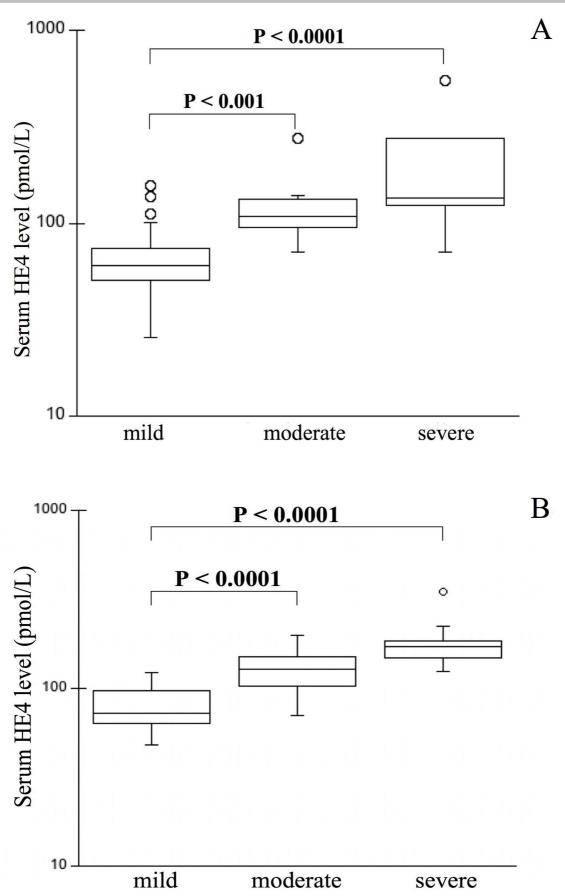
- 49 Muhlebach MS, Reed W, Noah TL. Quantitative cytokine gene expression in CF airway. *Pediatr Pulmonol*. 2004;37(5):393-399
- 50 Wright JM, Merlo CA, Reynolds JB, et al. Respiratory epithelial gene expression in patients with mild and severe cystic fibrosis lung disease. *Am J Respir Cell Mol Biol*. 2006;35(3):327-336
- 51 Zabner J, Scheetz TE, Almabrazi HG, et al. CFTR DeltaF508 mutation has minimal effect on the gene expression profile of differentiated human airway epithelia. *Am J Physiol Lung Cell Mol Physiol*. 2005;289(4):L545-553
- 52 Dosanjh AK, Elashoff D, Robbins RC. The bronchoalveolar lavage fluid of cystic fibrosis lung transplant recipients demonstrates increased interleukin-8 and elastase and decreased IL-10. *J Interferon Cytokine Res.* 1998;18(10):851-854
- 53 Inoue H, Massion PP, Ueki IF, et al. Pseudomonas stimulates interleukin-8 mRNA expression selectively in airway epithelium, in gland ducts, and in recruited neutrophils. *Am J Respir Cell Mol Biol.* 1994;11(6):651-663
- 54 Walsh DE, Greene CM, Carroll TP, et al. Interleukin-8 up-regulation by neutrophil elastase is mediated by MyD88/IRAK/TRAF-6 in human bronchial epithelium. *J Biol Chem.* 2001;276(38):35494-35499
- 55 Oglesby IK, McElvaney NG, Greene CM. MicroRNAs in inflammatory lung diseasemaster regulators or target practice? *Respir Res.* 2010;11148
- 56 Oglesby IK, Bray IM, Chotirmall SH, et al. miR-126 is downregulated in cystic fibrosis airway epithelial cells and regulates TOM1 expression. *J Immunol*. 2010;184(4):1702-1709
- 57 Tsuchiya M, Kumar P, Bhattacharyya S, et al. Differential regulation of inflammation by inflammatory mediators in cystic fibrosis lung epithelial cells. *J Interferon Cytokine Res.* 2013;33(3):121-129
- 58 Li W, He F. Monocyte to macrophage differentiation-associated (MMD) targeted by miR-140-5p regulates tumor growth in non-small cell lung cancer. *Biochem Biophys Res Commun.* 2014;450(1):844-850
- 59 Hua L, Liu Y, Zhen S, et al. Expression and biochemical characterization of recombinant human epididymis protein 4. *Protein Expr Purif*. 2014;10252-62
- 60 LeBleu VS, Teng Y, O'Connell, et al. Identification of human epididymis protein-4 as a fibroblast-derived mediator of fibrosis. *Nat Med.* 2013;19(2):227-231
- 61 Hajj R, Lesimple P, Nawrocki-Raby B, Birembaut P, Puchelle E, Coraux C. Human airway surface epithelial regeneration is delayed and abnormal in cystic fibrosis. *J Pathol.* 2007;211(3):340-350
- 62 Clarke LA, Botelho HM, Sousa L, Falcao AO, Amaral MD. Transcriptome metaanalysis reveals common differential and global gene expression profiles in cystic fibrosis and other respiratory disorders and identifies CFTR regulators. *Genomics*. 2015;106(5):268-277

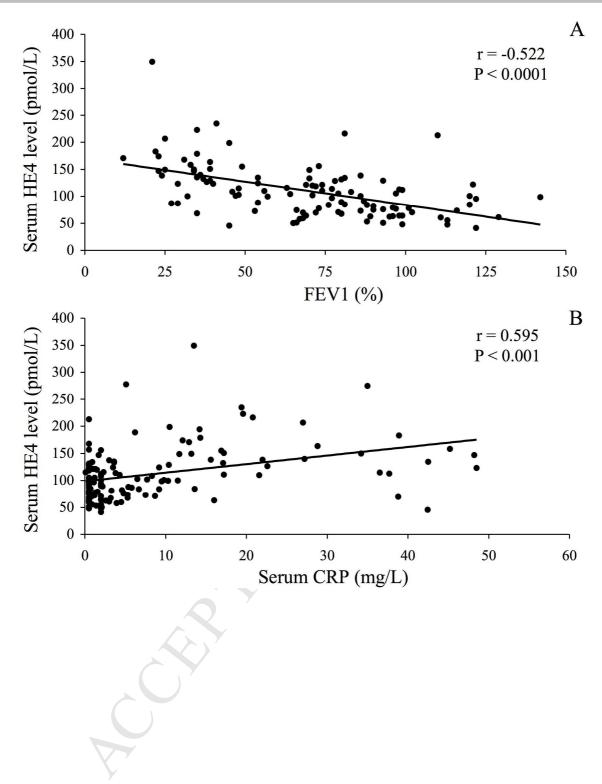
	Age (years)	Gender (m/f; n)	Serum HE4 (pmol/L)	Serum CRP (mg/L)
Young CF patients (n=77)	11 (7-17)	37/40	99.5 (73.1-128.9)	5.3 (2.0-9.3) [#]
Adult CF patients (n=57)	25 (21-29)	26/31	115.7 (77.8-148.7)	14.3 (12.9-27.0)
Non-CF young patients (n=76)	6 (3-14)	32/44	63.9 (54.3-83.4)	7.5 (1.1-18.2)
Non-CF adult patients (n=18)	26 (23-38)	7/11	59.2 (41.0-88.6)	8.9 (3.1-15.7)
Young controls (n=77)	13 (10-15)	33/44	36.3 (31.1-43.4)	nm
Adult controls (n=40)	25 (23-38)	17/23	33.8 (29.8-38.6)	nm

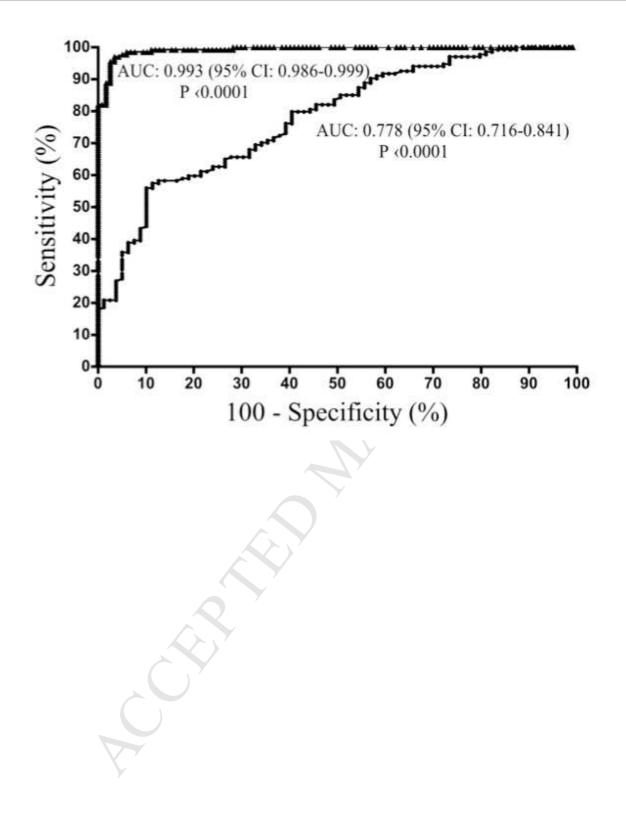
Table 1. Demographic and laboratory parameters of study populations. Data are expressed as median value (range). Serum CRP was measured in 29 individuals with severe clinical conditions out of 77 young CF-subjects marked with #. nm means 'not measured'.

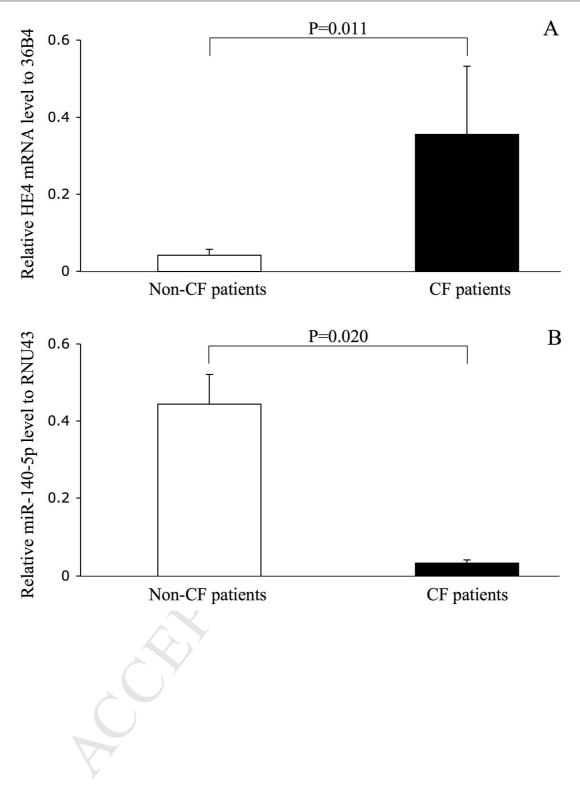
CERTE

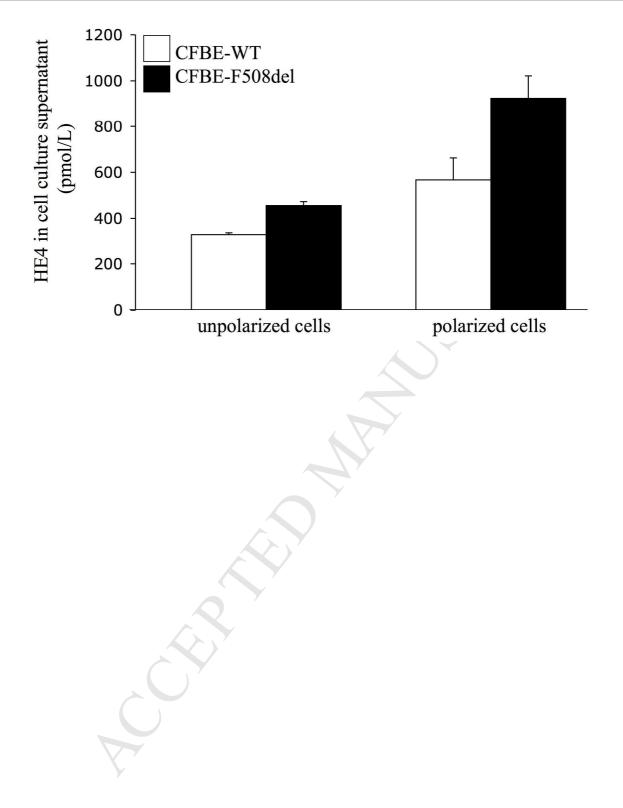












List of abbreviations: CF, cystic fibrosis; HE4, human epididymis protein 4; CFTR/*CFTR*, cystic fibrosis transmembrane conductance regulator/gene; CFBE, cystic fibrosis bronchial epithelial; CRP, C-reactive protein; BALF, bronchoalveolar lavage fluid; SK-score, Shwachman-Kulczycki score; FEV₁, forced expiratory volume in one second; NE, neutrophil elastase, TEER, transepithelial resistance