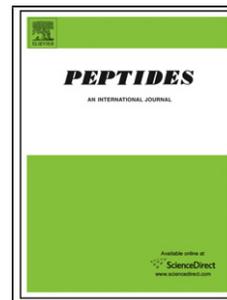


Accepted Manuscript

Title: PACAP and VIP signalling in chondrogenesis and osteogenesis

Author: Tamás Juhász Solveig Lind Helgadottir Andrea
Tamás Dóra Reglődi Róza Zákány



PII: S0196-9781(15)00040-6
DOI: <http://dx.doi.org/doi:10.1016/j.peptides.2015.02.001>
Reference: PEP 69410

To appear in: *Peptides*

Received date: 4-11-2014
Revised date: 16-1-2015
Accepted date: 20-1-2015

Please cite this article as: Juhász T, Helgadottir SL, Tamás A, Reglődi D, Zákány R, PACAP and VIP signalling in chondrogenesis and osteogenesis, *Peptides* (2015), <http://dx.doi.org/10.1016/j.peptides.2015.02.001>

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

1 PACAP and VIP signalling in chondrogenesis and osteogenesis

2 Tamás Juhász ^{a,*}, Solveig Lind Helgadóttir ^a, Andrea Tamás ^b, Dóra Reglódi ^b, Róza Zákány ^a

3

4 ^aDepartment of Anatomy, Histology and Embryology, University of Debrecen, Faculty of
5 Medicine, Nagyerdei krt. 98, H-4032, Debrecen, Hungary

6 ^b Department of Anatomy PTE-MTA “Lendület” PACAP Research Team, University of Pécs,
7 Medical School, Szigeti út 12, H-7624, Pécs, Hungary

8 * *Corresponding author.* Address: Department of Anatomy, Histology and Embryology,
9 University of Debrecen, Faculty of Medicine, Nagyerdei krt. 98, H-4032, Debrecen, Hungary.
10 Tel.: +36-52-255-567; fax: +36-52-255-115. *Email address:* juhaszt@anat.med.unideb.hu

11

12

13 **Main findings presented in this Manuscript are as follows:**

14

- 15 • Elements of VIP and PACAP signalling are present in cartilage and bone cells.
- 16 • Exogenous PACAP exerts a positive effect on *in vitro* cartilage and bone formation.
- 17 • PACAP plays a chondroprotective role under oxidative stress.

18

19

20 Abstract

21 Skeletal development is a complex process regulated by multifactorial signalling cascades that
22 govern proper tissue specific cell differentiation and matrix production. The influence of
23 certain regulatory peptides on cartilage or bone development can be predicted but are not

24 widely studied. In this review, we aimed to assemble and overview those signalling pathways
25 which are modulated by PACAP and VIP neuropeptides and are involved in cartilage and
26 bone formation. We discuss recent experimental data suggesting broad spectrum functions of
27 these neuropeptides in osteogenic and chondrogenic differentiation, including the canonical
28 downstream targets of PACAP and VIP receptors, PKA or MAPK pathways, which are key
29 regulators of chondro- or osteogenesis. Recent experimental data support the hypothesis that
30 PACAP is a positive regulator of chondrogenesis, while VIP has been reported playing an
31 important role in the inflammatory reactions of surrounding joint tissues. Regulatory function
32 of PACAP and VIP in bone development has also been proved, however the source of the
33 peptides is not obvious. Crosstalk and collateral connections of the discussed signalling
34 mechanisms make the system complicated and may obscure the pure effects of VIP and
35 PACAP. Chondro-protective properties of PACAP during oxidative stress observed in our
36 experiments indicate a possible therapeutic application of this neuropeptide.

37

38

39 **Keywords**

40 PKA; CREB; hedgehog; BMP; Runx2

41

41

42 Abbreviations

43 ALP, alkaline phosphatase; BMP, bone morphogenetic protein; cAMP, cyclic adenosine
44 monophosphate; CREB, cAMP response element-binding protein; ECM, extracellular matrix;
45 HH, hedgehog; IHH, Indian Hedgehog; MAPK, mitogen-activated protein kinase; NFAT,
46 nuclear factor of activated T cells; PAC1, pituitary adenylate cyclase-activating polypeptide
47 type I receptor; PACAP, pituitary adenylate cyclase polypeptide; PKA, protein kinase A;
48 PKC, protein kinase C; PP2A, protein phosphatase 2A; PP2B, protein phosphatase 2B;
49 PTHrP, parathyroid hormone related peptide; Runx2, Runt-related transcription factor 2;
50 SHH, Sonic Hedgehog; TGF β , transforming growth factor- β ; VIP, vasoactive intestinal
51 peptide; VPAC, vasoactive intestinal peptide receptor

52

53 Development of skeletal elements is influenced by several regulatory peptides, which may
54 derive from the evolving tissue or the surrounding nerve terminals. Production of proper long
55 bone architecture requires a cartilage template and involves time and growth factor dependent
56 activation of precisely defined regulating mechanisms and signalling cascade systems [1].

57 Hyaline cartilage is an avascular and aneural tissue [2] with a uniquely organized extracellular
58 matrix. Parallel with the bone formation, vessels and nerves penetrate the cartilage template
59 and release various regulatory factors, which can be responsible for remodelling of cartilage
60 and initiation of bone matrix production by osteoblasts. During the last decade several
61 theories have emerged regarding the regulation of the formation of these tissues by different
62 autocrine and paracrine mechanisms, with presumed involvement of various regulatory
63 peptides [3-6].

64

65 1. Pituitary Adenylate Cyclase Activating Polypeptide (PACAP) and Vasoactive
66 intestinal peptide (VIP)

67 Vasoactive intestinal peptide (VIP) and pituitary adenylate cyclase activating
68 polypeptide (PACAP) are neurohormones and members of the VIP–secretin–GHRH–
69 glucagon superfamily. Originally, both of these short neuropeptides were demonstrated
70 predominantly released in specific area of central nervous system [7]. VIP consists of 28
71 aminoacids and is produced by a variety of cells and tissues in addition to neuronal cells.
72 Among others, specific cells of the intestinal system can produce VIP along with some
73 immune and endocrine cells. Among its diverse physiological effects, VIP has important
74 functions in neuronal development and both in innate and acquired immunity [8].

75 PACAP was originally isolated from ovine hypothalamus extracts and later two
76 bioactive forms were identified: a shorter, 27 amino acid (PACAP 27) and a longer 38 amino

77 acid (PACAP38) form [9]. The N-terminal region of the polypeptide is evolutionary
78 conserved and shows a high homology with that of VIP [7]. PACAP is a pleiotropic
79 neuropeptide with various effects in the central nervous system, including trophic effects
80 during neuronal development and protective effects in neuronal regeneration. This protective
81 effect is one of its most promising features for therapeutic use, even if considering the short
82 half-life *in vivo* [10,11]. In the last decade, increasing amount of evidence has emerged
83 regarding the important roles of PACAP in peripheral organs such as uterus [12], ovary [13],
84 testis [14], moreover its presence has been proved in human milk [15]. Nonetheless, only
85 sporadic data exist about its function in skeletal elements [16-18].

86 PACAP and VIP can be ligands of three main receptors; PAC1, VPAC1 and VPAC2.
87 PACAP binds to PAC1 with the highest affinity, while the latter two attract PACAP and VIP
88 with equal affinity [19]. All of the three receptors are well characterized G protein coupled
89 receptors, the activation of which induces elevation of intracellular cAMP levels activating
90 protein kinase A (PKA) [7]. The so called “canonic” signalling activation may lead to the
91 nuclear translocation of CREB transcription factor and consequent activation of the
92 expression of various genes. PACAP binding is also able to control the MAPK pathways,
93 such as ERK and p38 kinases [7]. The versatility of PACAP/VIP receptor induced signal
94 transduction indicates its multifactorial regulation, implying a vast array of signalling
95 connections. This includes, for example, activation of IP₃ receptors inducing the release of
96 Ca²⁺ from endoplasmic reticulum (ER) [20]. The elevation of ic. Ca²⁺ concentration activates
97 various Ca²⁺ dependent signalling molecules such as classical PKCs, MAPK [21] or protein
98 phosphatases like PP2B [22]. The diversity of the developmental function is also hallmarked
99 by the fact that PACAP receptor activation may crosstalk with other signalling pathways such
100 as TGFβ [23], BMP [24], Hedgehog [25] and Notch signalisation [26]. Moreover, the general

101 protective and regenerative effects of PACAP originate from its antiapoptotic function [27]
102 and its ability to decrease inflammatory reactions [28].

103

104 2. Regulation of chondrogenesis focused on VIP and PACAP

105 As articular cartilage has very poor regeneration capacity, the exploration of new
106 strategies to improve replacement or reconstruction of cartilage is very important. Currently,
107 no effective or curative treatment is available for degenerative cartilage diseases such as
108 osteoarthritis. The signalling pathways of proper cartilage development are still under
109 investigation since plenty of the molecular signalling puzzles have neither been solved nor
110 locked in their adequate positions.

111 Chondrogenic differentiation is a multistep process involving rapid proliferation and
112 condensation of chondroprogenitor cells. Formation of chondrogenic nodules and cartilage
113 specific extracellular matrix production both are required for proper hyaline cartilage
114 development [29]. Transcription factors of the SoxE family such as Sox5, Sox6 and Sox9 are
115 essential for the induction of mRNA expression of cartilage matrix-specific proteins (e.g.
116 COL2A1, aggrecan core protein). Sox9 is one of the pivotal signalling elements of
117 chondrogenesis, therefore, its regulation by reversible phosphorylation can be a key
118 momentum of the proper differentiation cycle. Sox9 promoter is known to be regulated by the
119 CREB that binds to a CRE site upstream of Sox9 [30]. We have demonstrated that Sox9 and
120 CREB transcription factors are phosphorylated by PKA during cartilage formation [31,32].
121 Moreover, a quite complex regulatory mechanism and synergism between Sox9 function and
122 the cAMP–PKA–CREB pathway was published in both mature and differentiating
123 chondrocytes which includes BMP pathway connections [33]. Finally, we have shown that
124 the activation of signalling elements phosphorylated by PKA can be equilibrated by a few
125 Ser/Thr protein phosphatases such as PP2A and PP2B [34,35]. Since the regulation of these

126 cartilage specific signalling pathways are cAMP or Ca^{2+} dependent it could be a question of
127 interest whether PACAP/VIP neuropeptides have any signalisation connection with proper
128 hyaline cartilage formation.

129 Only sporadic data exist on the functions of regulatory peptides in chondrogenesis.
130 Role of various regulatory peptides such as VIP are well known in inflammatory diseases;
131 moreover, VIP is a promising agent in the therapeutic treatment of rheumatoid arthritis [11].
132 Although the articular cartilage is aneural, the surrounding synovial membrane is rich in nerve
133 endings, which may release VIP into the synovial cavity and subsequently induce anti-
134 inflammatory processes [36]. About the functions of PACAP in the adult joints we still have
135 exiguous knowledge despite the fact that PACAP-positive nerve endings have been described
136 in cartilage canals of porcine epiphyseal cartilage more than 15 years ago [37]. Our laboratory
137 was the first to demonstrate that the mRNAs of preproPACAP as well as PAC1, VPAC1 and
138 VPAC2 receptors are expressed in chicken “high density” chondrogenic cell cultures.
139 Furthermore, we have shown the expression of the PAC1 receptor protein in
140 chondroprogenitor cells [17] and increased extracellular matrix synthesis was detected during
141 PACAP administration suggesting the positive effect of this neuropeptide in cartilage
142 development. Our findings suggested the presence of PACAP-related autocrine and/or
143 paracrine effects in cartilage itself, reflecting on a possible new signalling mechanism in the
144 regeneration of hyaline cartilage [38,39]. Although the receptors of VIP were expressed by
145 chondrogenic cells in our experiments, others found that this neuropeptide did not influence
146 the matrix production of chondrocytes and synovial cells [40] suggesting certain tissue
147 specific effects of these neuropeptides. Classical downstream targets of PAC1 receptor
148 activation such as PKA, PKC and MAPK signalling cascades play essential role in
149 chondrogenesis [32,35,41]. It has been published that PKA phosphorylates CREB and Sox9
150 transcription factors [32], the latter one being a key regulator of chondrogenesis [42]. PACAP

151 administration into the medium of chondrogenic cell cultures increased the phosphorylation
152 both of Sox9 and CREB, and enhanced matrix production of the differentiating cells was also
153 observed [17] (Fig 1.). PAC1 receptor activation can be responsible for the elevation of
154 intracellular Ca^{2+} concentration via regulating Ca^{2+} dependent phosphatases such as PP2B
155 (also known as calcineurin). This enzyme is one of the positive regulators of *in vitro*
156 chondrogenesis [35,41,43]. Therefore, we investigated the involvement of this Ser/Thr
157 phosphatase in PACAP signalling pathways and connection between PP2B activity and
158 PACAP signalling was proved [17] (Fig 1.), similarly to chromaffin cells [44]. These *in vitro*
159 results indicated that the presence of PACAP is essential for proper cartilage formation,
160 however the phenotype of PACAP KO mice [45] did not show any dramatic macroscopical
161 morphological alteration of skeleton. Although the analysis of the genetically modified
162 animals has not been completed yet, our initial observations suggested alterations in the
163 composition of the cartilage extracellular matrix and in the expression of various signalling
164 molecules in the knee joints of PACAP KO mice (our unpublished data). In the reproductive
165 organ system of these mice, the lack of PACAP gene resulted in reduced fertility and altered
166 mating behaviour of females [46], moreover the maturation [47] and the morphology [48] of
167 gonadal cells showed notable differences. The complex phenotypic changes raise the
168 possibility of multiple crosstalk of PACAP signalling with developmental pathways
169 connected to various morphogens, as well as certain compensatory mechanisms of PACAP
170 signalling cascades. For instance MAPK and Wnt signalling both play important roles in the
171 proper cartilage formation and tissue patterning [49] and a PACAP-independent PAC1
172 receptor activation has been directly linked to the regulation of Wnt/ β -catenin pathways [50].
173 Notch signalling activation plays a crucial role in chondrogenesis [51] and exerts modulatory
174 function in osteoarthritis [52] Recently, crosstalk of G protein coupled receptors and Notch
175 signalling has been reported in bacterial LPS induced macrophages [53]. SHH pathway is

176 another essential positive chondroregulatory pathway [54] and it can be inhibited by PACAP
177 activation [55].

178 Recently we have demonstrated a chondro-protective effect of PACAP in chondrogenic
179 cell cultures where the administration of the neuropeptide compensated the harmful effects of
180 oxidative stress. It has been shown that PACAP can prevent the harmful effects of cerebral
181 ischemia or oxidative stress induced apoptosis in the central nervous system [56]. PACAP
182 deficient mice showed higher sensitivity to injury during retinal ischemic conditions, axonal
183 lesion, intestinal inflammation or oxidative stress of the kidneys [57]. The presence of
184 PACAP/VIP had preventing role in rheumatoid arthritis [58,59], and cardioprotective effects
185 of these peptides have also been demonstrated [60]. In the light of these data, the cartilage
186 protecting effect of PACAP was predictable; however the exploration of the molecular
187 background of this phenomenon has only started yet. In chicken chondrogenic cells, the
188 addition of PACAP 1-38 during oxidative stress prevented the inhibition of cartilage matrix
189 production by free oxygen radicals and the increased activity of PKA seemed to take part in
190 this compensatory effect [17]. The addition of the neuropeptide also exerted effect on matrix
191 metalloproteinase (MMP) expression in chondrogenic cell cultures in the presence of reactive
192 oxygen species (our unpublished data). Similar results have been published in alveolar cells
193 where both VIP and PACAP were able to decrease the expression of certain MMPs and
194 reduced the activation and expression of caspase3 [61]. VIP and its receptors are expressed in
195 synovial fibroblasts [62] and it enables the release of inflammatory factors either by these
196 cells or immunocompetent cells residing in the surrounding synovial tissues [63]. Finally,
197 PACAP has been shown to have modulatory effects on inflammatory processes of rheumatoid
198 arthritis [64]. These data all strongly suggest that PACAP is a promising future therapeutic
199 agent in inflammatory and degenerative joint diseases [65].

200

201 3. VIP and PACAP in osteogenic signalling cascades

202 Similarly to chondrogenic differentiation, proper osteogenesis requires high spatial and
203 temporary organization supported by complex bone specific developing mechanisms and
204 signalling. Development of this skeletal tissue involves differentiation of osteoblasts from
205 osteoprogenitors. It is followed by an initial deposition of a bone specific organic ECM
206 abundant in collagen type I completed with certain bone specific matrix components such as
207 osteocalcin or osteonectin. This osteoid undergoes calcification then meaning deposition of
208 calcium hydroxyapatite crystals in the bone matrix with active contribution of osteoblasts.
209 Differentiation of osteoblast is regulated by three main signalling cascades such as BMP,
210 WNT and Hedgehog cascades [66-68]. BMPR activation subsequently induces the
211 phosphorylation of Smad1/5 and with the help of Smad4 the complex is translocated into the
212 nuclei of osteogenic cells and initiates expression of bone specific genes such as the
213 transcription factor osterix, alkaline phosphatase (ALP) or collagen type I [69,70]. The
214 expression of BMPs is regulated by CREB transcription factor activated via PKA signalling
215 pathways [70]. On the other hand a well balanced expression of hedgehog signalling elements
216 governed by another bone specific transcription factor, Runx2 is also essential for proper long
217 bone formation [71]. Runx2 can be directly phosphorylated by PKA [72] and subsequently
218 activates the expression of bone specific signalling elements or ECM components. This
219 complex signalisation involves broad spectrum crosstalk opportunities with the PACAP/VIP
220 signalisation, further highlighting the significance of neuropeptide signalling in bone
221 formation and regeneration.

222 During endochondral ossification, after the invasion of vessels and nerves into the cartilage
223 template osteoprogenitor cells start to migrate into the diaphysis of the developing long bone
224 and differentiate into osteoblasts. This process can also be regulated by neuropeptides [73].

225 During the elongation of long bones PACAP positive nerve fibers penetrate the bone matrix

226 [37]. VIP positive sympathetic nerve endings were also identified releasing these
227 neuropeptides [74]. As an interesting observation, receptor composition and effects of VIP
228 exhibited differences in cells of bones developed in different ways (i.e. membranous or
229 endochondral). Moreover, the direct communication of sympathetic nerve fibers with
230 osteoblasts showed an embryonic origin dependent response and signalisation, suggesting that
231 the innervation of periosteum by peptidergic fibers plays important function both in bone
232 regeneration and formation [75]. The role of PACAP and VIP in osteogenesis was further
233 supported by the observations where MC3T3 E1 mouse calvaria derived osteoblast cell line
234 [76] and UMR-106 cells isolated from rat osteosarcoma [16] were shown both expressing the
235 receptors for these neuropeptides. Accumulation of cAMP in osteoblasts is proved to be as a
236 result of combined activation of PACAP and VIP and regulates diverse signalling pathways
237 influencing osteoblast differentiation. In line with this, presence of certain neuropeptides was
238 shown to be elevated after bone fracture, indicating their importance in successful
239 regeneration [77]. A recent report demonstrated release of various neuropeptides from
240 periosteal nerve endings resulting in enhancement of intercellular communication and
241 increased metabolic activity of osteoblasts [78]. As it was described above, osteogenic
242 transformation, bone matrix production and mineralization are regulated by multiple
243 signalling cascades [79], where the activation of MAPK and PKA plays essential roles. Runx2
244 is one of the key transcription factors which governs osteoblast differentiation [80] and it is
245 regulated by PKA signalling pathways [81]. We have demonstrated that the administration of
246 PACAP into the medium of UMR-106 cell line enhanced the nuclear translocation of Runx2
247 and increased expression of collagen type I, ALP and osterix genes was observed (Fig. 2.).
248 Interestingly, the phosphorylation of CREB by PKA was not remarkably increased after
249 PACAP addition in this osteosarcoma derived cell line [16] (Fig 2.). BMP signalling pathway
250 is another fundamental regulator of osteogenesis and crosstalk with Runx2 has been reported

251 [83]. Moreover, the TGF β /BMP pathways are activated by PACAP or VIP [24]. Indeed, the
252 administration of PACAP increased the expression of BMPs in UMR-106 cells and
253 expression of BMPR1, one of its major receptors, became also elevated. As a consequence of
254 BMPR activation, a pronounced elevation of the nuclear presence of Smad1 transcription
255 factor was detected under the effect of PACAP administration [16] (Fig 2.). VIP can also be
256 regulated by TGF β /BMP signalling pathways as Smads may activate VIP expression [85]
257 suggesting a complex reciprocal signalling with numerous compensatory escape routes during
258 bone development [16].

259 PACAP and VIP may directly activate ERK1/2 e.g. during adipogenesis [86] or in osteoblast
260 cells [87], furthermore CREB phosphorylation is regulated by the MAPK system in MC3T3
261 cells [88]. Additionally, intracellular Ca²⁺ concentration can be elevated by PACAP [89] or
262 VIP [90], resulting in an activation of classical PKCs and ERK both influencing osteoblast
263 differentiation [91]. Nonetheless, PACAP treatment of UMR-106 cells did not alter the Ca²⁺
264 concentration of these osteoblast cells, and activation of classical PKCs was not detected, in
265 our experiments [16] (Fig 2.). Ca²⁺ influx can be evoked by PACAP [92] and the presence of
266 PACAP and VIP is able to decrease the Ca²⁺ entry via L- and N-type calcium channels in
267 neurons [93]. It is known that the administration of PACAP affects Ca²⁺ oscillation [94] and
268 alters the Ca²⁺ related vesicular transport of chromaffin cells [95]. Besides this dynamic
269 alteration of intracellular Ca-homeostasis, PACAP also exerts effects on matrix
270 mineralisation. We found that addition of PACAP elevated the deposition of inorganic matrix
271 components in the ECM of UMR-106 cells [16]. Moreover, an altered mineralisation was
272 detected during tooth formation of PACAP deficient mice [96], suggesting a yet unknown
273 connection between PACAP and Ca²⁺ release of osteoblasts, ameloblasts and/or odontoblasts.
274 As a possible mechanism for PACAP induced extracellular Ca²⁺ accumulation during
275 osteogenesis, calcitonin gene-related protein was proved to effect on osteoclast function [97]

276 and the presence of PACAP decreased the matrix-resorption and consequent Ca-release by
277 these cells [95,96].

278 Hedgehog signalling is of key importance amongst the regulatory mechanisms of bone
279 and cartilage development [71]. A well defined balance between Indian Hedgehog (IHH) and
280 Parathyroid Hormone Related Peptide (PTHrP) is essential for proper long bone formation,
281 regulation of proliferation and matrix production of osteoblasts via the activation of Runx2
282 transcription factor [98]. PTHrP directly communicates with PKA signalling inducing the
283 activation of CREB and NFAT factors in osteoblasts [99]. In UMR-106 cells the application
284 of PACAP elevated the expression of PTHrP without altering the IHH expression [16]. Sonic
285 Hedgehog (SHH) pathway is known to be regulated by PACAP signalling [55] and the
286 activation of PKA downregulates the function of Gli1, which consequently decreases the
287 proliferation [25]. In PACAP KO mice, enhanced SHH signalling was detected during tooth
288 development [94]. On the contrary, exogenous administration of PACAP elevated the
289 expression of SHH and a more pronounced nuclear presence of Gli1 was found in rat UMR-
290 106 cells [16]. This contradiction may stem from the osteosarcoma origin of UMR cells, as
291 malignant cells can exhibit alterations of various signalling mechanisms. Although we do not
292 have data about the possible function of VIP in osteogenesis, previous results suggest that
293 multifactorial signalling pathways of these regulatory peptides exert modulatory effect on
294 matrix production and differentiation in bone development [100].

295

296 Conclusion

297 Regulatory pathways of PACAP and VIP form a complex signalling network indicating the
298 communication of a huge variety of signalling cascades accomplishing and supporting the
299 diverse functions of these regulatory peptides. Different compensatory mechanisms can
300 switch on or off upon activation or inactivation of certain signalling cascades in the

301 interconnected system, which can obscure the physiological function of PACAP and/or VIP
302 during chondrogenesis and osteogenesis. Better understanding of the functions of these
303 neurohormones during skeletal development may help us to find possibilities for their
304 therapeutic application in various skeletal diseases.

305 Acknowledgements

306 The authors are grateful for Mrs. Krisztina Bíró for excellent technical assistance during the
307 studies. This work was supported by grants from Akira Arimura Foundation Research Grant,
308 the Hungarian Science Research Fund (OTKA CNK80709 and OTKA K 104984), Bolyai
309 Scholarship and the Hungarian Ministry of Education (TÁMOP 4.2.1.B-10/2/KONV-2010-
310 002, PTE-MTA “Lendület” Program) and from the New Széchenyi Plan (TÁMOP-4.2.2.A-
311 11/1/KONV-2012-0053, TÁMOP-4.2.2.A-11/1/KONV-2012-0024,; The project is co-
312 financed by the European Union and the European Social Fund). This research and T.J. was
313 supported by Szodoray Lajos Fund and by the European Union and the State of Hungary, co-
314 financed by the European Social Fund in the framework of TÁMOP 4.2.4. A/2-11-1-2012-
315 0001 ‘National Excellence Program’. T.J. and R.Z. are supported by GOP-1.1.1-11-2012-
316 0197 financed by the Hungarian government and the EU.

317

318

319

319

320

Reference List

321

322 1. Paiva KB, Granjeiro JM. Bone tissue remodeling and development: Focus on matrix
323 metalloproteinase functions. *Arch.Biochem.Biophys.* 2014;561C, 74-87.

324 2. Pacifici M, Koyama E, Iwamoto M. Mechanisms of synovial joint and articular
325 cartilage formation: recent advances, but many lingering mysteries. *Birth Defects*
326 *Res.C.Embryo.Today* 2005;75, 237-248.

327 3. Bach FC, Rutten K, Hendriks K, Riemers FM, Cornelissen P, de Bruin A, Arkesteijn
328 GJ, Wubbolts R, Horton WA, Penning LC, Tryfonidou MA. The paracrine feedback
329 loop between vitamin D(3) (1,25(OH)(2)D(3)) and PTHrP in prehypertrophic
330 chondrocytes. *J.Cell Physiol* 2014;229, 1999-2014.

331 4. Lai JH, Kajiyama G, Smith RL, Maloney W, Yang F. Stem cells catalyze cartilage
332 formation by neonatal articular chondrocytes in 3D biomimetic hydrogels. *Sci.Rep.*
333 2013;3, 3553.

334 5. Liu Y, Olsen BR. Distinct VEGF functions during bone development and
335 homeostasis. *Arch.Immunol.Ther.Exp.(Warsz.)* 2014;62, 363-368.

336 6. Xu L, Wang Q, Xu F, Ye Z, Zhou Y, Tan WS. Mesenchymal stem cells downregulate
337 articular chondrocyte differentiation in noncontact coculture systems: implications in
338 cartilage tissue regeneration. *Stem Cells Dev.* 2013;22, 1657-1669.

339 7. Vaudry D, Falluel-Morel A, Bourgault S, Basille M, Burel D, Wurtz O, Fournier A,
340 Chow BK, Hashimoto H, Galas L, Vaudry H. Pituitary adenylate cyclase-activating

- 341 polypeptide and its receptors: 20 years after the discovery. *Pharmacol.Rev.* 2009;61,
342 283-357.
- 343 8. Moody TW, Hill JM, Jensen RT. VIP as a trophic factor in the CNS and cancer cells.
344 *Peptides* 2003;24, 163-177.
- 345 9. Miyata A, Arimura A, Dahl RR, Minamino N, Uehara A, Jiang L, Culler MD, Coy
346 DH. Isolation of a novel 38 residue-hypothalamic polypeptide which stimulates
347 adenylate cyclase in pituitary cells. *Biochem.Biophys.Res.Comm.* 1989;164, 567-
348 574.
- 349 10. Bourgault S, Vaudry D, Botia B, Couvineau A, Laburthe M, Vaudry H, Fournier A.
350 Novel stable PACAP analogs with potent activity towards the PAC1 receptor.
351 *Peptides* 2008;29, 919-932.
- 352 11. Sethi V, Rubinstein I, Kuzmis A, Kastrissios H, Artwohl J, Onyuksel H. Novel,
353 biocompatible, and disease modifying VIP nanomedicine for rheumatoid arthritis.
354 *Mol.Pharm.* 2013;10, 728-738.
- 355 12. Reglodi D, Tamas A, Koppan M, Szogyi D, Welke L. Role of PACAP in Female
356 Fertility and Reproduction at Gonadal Level - Recent Advances. *Front*
357 *Endocrinol.(Lausanne)* 2012;3, 155.
- 358 13. Koppan M, Varnagy A, Reglodi D, Brubel R, Nemeth J, Tamas A, Mark L, Bodis J.
359 Correlation between oocyte number and follicular fluid concentration of pituitary
360 adenylate cyclase-activating polypeptide (PACAP) in women after superovulation
361 treatment. *J.Mol.Neurosci.* 2012;48, 617-622.

- 362 14. Nakamura K, Nakamachi T, Endo K, Ito K, Machida T, Oka T, Hori M, Ishizaka K,
363 Shioda S. Distribution of pituitary adenylate cyclase-activating polypeptide (PACAP)
364 in the human testis and in testicular germ cell tumors. *Andrologia* 2014;46, 465-471.
- 365 15. Csanaky K, Reglodi D, Banki E, Tarcai I, Mark L, Helyes Z, Ertl T, Gyarmati J,
366 Horvath K, Santik L, Tamas A. Examination of PACAP38-like immunoreactivity in
367 different milk and infant formula samples. *Acta Physiol Hung.* 2013;100, 28-36.
- 368 16. Juhasz T, Matta C, Katona E, Somogyi C, Takacs R, Hajdu T, Helgadottir SL, Fodor
369 J, Csernoch L, Toth G, Bako E, Reglodi D, Tamas A, Zakany R. Pituitary Adenylate
370 Cyclase-Activating Polypeptide (PACAP) Signalling Enhances Osteogenesis in UMR-
371 106 Cell Line. *J.Mol.Neurosci.* 2014;doi10.1007/s12031-014-0389-1
- 372 17. Juhasz T, Matta C, Katona E, Somogyi C, Takacs R, Gergely P, Csernoch L, Panyi G,
373 Toth G, Reglodi D, Tamas A, Zakany R. Pituitary adenylate cyclase activating
374 polypeptide (PACAP) signalling exerts chondrogenesis promoting and protecting
375 effects: implication of calcineurin as a downstream target. *PLoS.One.* 2014;9, e91541.
- 376 18. Kovacs CS, Chik CL, Li B, Karpinski E, Ho AK. Pituitary adenylate cyclase-
377 activating peptide stimulates cyclic AMP accumulation in UMR 106 osteoblast-like
378 cells. *J.Endocrinol.* 1996;149, 287-295.
- 379 19. Gourlet P, Vandermeers A, Vertongen P, Rathe J, De Neef P, Cnudde J, Waelbroeck
380 M, Robberecht P. Development of high affinity selective VIP1 receptor agonists.
381 *Peptides* 1997;18, 1539-1545.
- 382 20. Tanaka K, Shibuya I, Uezono Y, Ueta Y, Toyohira Y, Yanagihara N, Izumi F, Kanno
383 T, Yamashita H. Pituitary adenylate cyclase-activating polypeptide causes Ca²⁺
384 release from ryanodine/caffeine stores through a novel pathway independent of both

- 385 inositol trisphosphates and cyclic AMP in bovine adrenal medullary cells.
386 J.Neurochem. 1998;70, 1652-1661.
- 387 21. Szabo A, Danyadi B, Bogнар E, Szabadfi K, Fabian E, Kiss P, Mester L, Manavalan S,
388 Atlasz T, Gabriel R, Toth G, Tamas A, Reglodi D, Kovacs K. Effect of PACAP on
389 MAP kinases, Akt and cytokine expressions in rat retinal hypoperfusion.
390 Neurosci.Lett. 2012;523, 93-98.
- 391 22. Schuhmann K, Romanin C, Baumgartner W, Groschner K. Intracellular Ca²⁺ inhibits
392 smooth muscle L-type Ca²⁺ channels by activation of protein phosphatase type 2B
393 and by direct interaction with the channel. J.Gen.Physiol 1997;110, 503-513.
- 394 23. Oka H, Jin L, Kulig E, Scheithauer BW, Lloyd RV. Pituitary adenylate cyclase-
395 activating polypeptide inhibits transforming growth factor-beta1-induced apoptosis in
396 a human pituitary adenoma cell line. Am.J.Pathol. 1999;155, 1893-1900.
- 397 24. Pavelock KA, Girard BM, Schutz KC, Braas KM, May V. Bone morphogenetic
398 protein down-regulation of neuronal pituitary adenylate cyclase-activating polypeptide
399 and reciprocal effects on vasoactive intestinal peptide expression. J.Neurochem.
400 2007;100, 603-616.
- 401 25. Niewiadomski P, Zhujiang A, Youssef M, Waschek JA. Interaction of PACAP with
402 Sonic hedgehog reveals complex regulation of the hedgehog pathway by PKA. Cell
403 Signal. 2013;25, 2222-2230.
- 404 26. Lu Q, Tong B, Luo Y, Sha L, Chou G, Wang Z, Xia Y, Dai Y. Norisoboldine
405 suppresses VEGF-induced endothelial cell migration via the cAMP-PKA-NF-
406 kappaB/Notch1 pathway. *PLoS.One*. 2013;8, e81220.

- 407 27. Szabadfi K, Szabo A, Kiss P, Reglodi D, Setalo G Jr, Kovacs K, Tamas A, Toth G,
408 Gabriel R. PACAP promotes neuron survival in early experimental diabetic
409 retinopathy. *Neurochem.Int.* 2014;64, 84-91.
- 410 28. Heimesaat MM, Dunay IR, Schulze S, Fischer A, Grundmann U, Alutis M, Kuhl AA,
411 Tamas A, Toth G, Dunay MP, Gobel UB, Reglodi D, Bereswill S. Pituitary adenylate
412 cyclase-activating polypeptide ameliorates experimental acute ileitis and extra-
413 intestinal sequelae. *PLoS.One.* 2014;9, e108389.
- 414 29. Goldring MB, Tsuchimochi K, Ijiri K. The control of chondrogenesis. *J.Cell Biochem.*
415 2006;97, 33-44.
- 416 30. Piera-Velazquez S, Hawkins DF, Whitecavage MK, Colter DC, Stokes DG, Jimenez
417 SA. Regulation of the human SOX9 promoter by Sp1 and CREB. *Exp.Cell Res.*
418 2007;313, 1069-1079.
- 419 31. Juhasz T, Matta C, Somogyi C, Katona E, Takacs R, Soha RF, Szabo IA, Cserhati C,
420 Szody R, Karacsonyi Z, Bako E, Gergely P, Zakany R. Mechanical loading stimulates
421 chondrogenesis via the PKA/CREB-Sox9 and PP2A pathways in chicken micromass
422 cultures. *Cell Signal.* 2014;26, 468-482.
- 423 32. Zakany R, Szucs K, Bako E, Felszeghy S, Czifra G, Biro T, Modis L, Gergely P.
424 Protein phosphatase 2A is involved in the regulation of protein kinase A signaling
425 pathway during in vitro chondrogenesis. *Exp.Cell Res.* 2002;275, 1-8.
- 426 33. Zhao L, Li G, Zhou GQ. SOX9 directly binds CREB as a novel synergism with the
427 PKA pathway in BMP-2-induced osteochondrogenic differentiation. *J.Bone*
428 *Miner.Res.* 2009;24, 826-836.

- 429 34. Zakany R, Bako E, Felszeghy S, Hollo K, Balazs M, Bardos H, Gergely P, Modis L.
430 Okadaic acid-induced inhibition of protein phosphatase 2A enhances chondrogenesis
431 in chicken limb bud micromass cell cultures. *Anat.Embryol.(Berl)* 2001;203, 23-34.
- 432 35. Zakany R, Szijgyarto Z, Matta C, Juhasz T, Csontos C, Szucs K, Czifra G, Biro T,
433 Modis L, Gergely P. Hydrogen peroxide inhibits formation of cartilage in chicken
434 micromass cultures and decreases the activity of calcineurin: implication of ERK1/2
435 and Sox9 pathways. *Exp.Cell Res.* 2005;305, 190-199.
- 436 36. Konttinen YT, Tiainen VM, Gomez-Barrena E, Hukkanen M, Salo J. Innervation of
437 the joint and role of neuropeptides. *Ann.N.Y.Acad.Sci.* 2006;1069, 149-154.
- 438 37. Strange-Vognsen HH, Arnbjerg J, Hannibal J. Immunocytochemical demonstration of
439 pituitary adenylate cyclase activating polypeptide (PACAP) in the porcine epiphyseal
440 cartilage canals. *Neuropeptides* 1997;31, 137-141.
- 441 38. Ahmed N, Dreier R, Gopferich A, Grifka J, Grassel S. Soluble signalling factors
442 derived from differentiated cartilage tissue affect chondrogenic differentiation of rat
443 adult marrow stromal cells. *Cell Physiol Biochem.* 2007;20, 665-678.
- 444 39. Gelse K, Brem M, Klinger P, Hess A, Swoboda B, Hennig F, Olk A. Paracrine effect
445 of transplanted rib chondrocyte spheroids supports formation of secondary cartilage
446 repair tissue. *J.Orthop.Res.* 2009;27, 1216-1225.
- 447 40. Rahman S, Dobson PR, Bunning RA, Russell RG, Brown BL. The regulation of
448 connective tissue metabolism by vasoactive intestinal polypeptide. *Regul.Pept.*
449 1992;37, 111-121.

- 450 41. Juhasz T, Matta C, Mészár Z, Nagy G, Szigyarto Z, Molnar Z, Kolozsvari B, Bako E,
451 Zakany R Optimized transient transfection of chondrogenic primary cell cultures.
452 CEJB. 2010;5: 572-584.
- 453 42. de Crombrughe B, Lefebvre V, Behringer RR, Bi W, Murakami S, Huang W.
454 Transcriptional mechanisms of chondrocyte differentiation. *Matrix Biol.* 2000;19,
455 389-394.
- 456 43. Matta C, Mobasher A, Gergely P, Zakany R. Ser/Thr-phosphoprotein phosphatases in
457 chondrogenesis: neglected components of a two-player game. *Cell Signal.* 2014;26,
458 2175-2185.
- 459 44. Lee HW, Hahm SH, Hsu CM, Eiden LE. Pituitary adenylate cyclase-activating
460 polypeptide regulation of vasoactive intestinal polypeptide transcription requires Ca²⁺
461 influx and activation of the serine/threonine phosphatase calcineurin. *J.Neurochem.*
462 1999;73, 1769-1772.
- 463 45. Hattori S, Takao K, Tanda K, Toyama K, Shintani N, Baba A, Hashimoto H,
464 Miyakawa T. Comprehensive behavioral analysis of pituitary adenylate cyclase-
465 activating polypeptide (PACAP) knockout mice. *Front Behav.Neurosci.* 2012;6, 58.
- 466 46. Shintani N, Mori W, Hashimoto H, Imai M, Tanaka K, Tomimoto S, Hirose
467 M, Kawaguchi C, Baba A. Defects in reproductive functions in PACAP-deficient
468 female mice. *Regul Pept.* 2002;109:45-8
- 469 47. Barberi M, Di Paolo V, Latini S, Guglielmo MC, Cecconi S, Canipari R. Expression
470 and functional activity of PACAP and its receptors on cumulus cells: effects on oocyte
471 maturation. *Mol Cell Endocrinol.* 2013; 375(1-2):79-88.

- 472 48. Brubel R, Kiss P, Vincze A, Varga A, Varnagy A, Bodis J, Mark L, Jambor E, Maasz
473 G, Hashimoto H, Helyes Z, Toth G, Tamas A, Koppan M, Reglodi D. Effects
474 of pituitary adenylate cyclase activating polypeptide on human sperm motility. *J Mol*
475 *Neurosci.* 2012; 48(3):623-30.
- 476 49. Zhang Y, Pizzute T, Pei M. A review of crosstalk between MAPK and Wnt signals
477 and its impact on cartilage regeneration. *Cell Tissue Res.* 2014;358(3):633-49.
- 478 50. Yu R, Cui Z, Li M, Yang Y, Zhong J. Dimer-Dependent Intrinsic/Basal Activity of the
479 Class B G Protein-Coupled Receptor PAC1 Promotes Cellular Anti-Apoptotic
480 Activity through Wnt/ β -Catenin Pathways that Are Associated with Dimer
481 Endocytosis. *PLoS One.* 2014 ;9(11):e113913.
- 482 51. Serrano MJ, So S, Hinton RJ. Roles of notch signalling in mandibular
483 condylar cartilage. *Arch Oral Biol.* 2014;59(7):735-40.
- 484 52. Sassi N, Gadgadi N, Laadhar L, Allouche M, Mourali S, Zandieh-Doulabi
485 B, Hamdoun M, Nulend JK, Makni S, Sellami S. Notch signaling is involved in
486 human articular chondrocytes de-differentiation during osteoarthritis. *J Recept Signal*
487 *Transduct Res.* 2014;34(1):48-57.
- 488 53. Sangphech N, Osborne BA, Palaga T. Notch signaling regulates the phosphorylation
489 of Akt and survival of lipopolysaccharide-activated macrophages via regulator of G
490 protein signaling 19 (RGS19). *Immunobiology.* 2014;219(9):653-60.
- 491 54. Kwon HJ. ATP oscillations mediate inductive action of FGF and Shh signalling on
492 prechondrogenic condensation. *Cell Biochem.Funct.* 2013;31, 75-81.

- 493 55. Waschek JA, Cicco-Bloom E, Nicot A, Lelievre V. Hedgehog signaling: new targets
494 for GPCRs coupled to cAMP and protein kinase A. *Ann.N.Y.Acad.Sci.* 2006;1070,
495 120-128.
- 496 56. Sanchez A, Chiriva-Internati M, Grammas P. Transduction of PACAP38 protects
497 primary cortical neurons from neurotoxic injury. *Neurosci.Lett.* 2008;448, 52-55.
- 498 57. Reglodi D, Kiss P, Szabadfi K, Atlasz T, Gabriel R, Horvath G, Szakaly P, Sandor
499 B, Lubics A, Laszlo E, Farkas J, Matkovits A, Brubel R, Hashimoto H, Ferencz
500 A, Vincze A, Helyes Z, Welke L, Lakatos A, Tamas A. PACAP is an endogenous
501 protective factor-insights from PACAP-deficient mice. *J Mol*
502 *Neurosci.* 2012;48(3):482-92.
- 503 58. Hernanz A, Medina S, de Miguel E, Martín-Mola E. Effect of calcitonin gene-related
504 peptide, neuropeptide Y, substance P, and vasoactive intestinal peptide on interleukin-
505 1beta, interleukin-6 and tumor necrosis factor-alpha production by peripheral whole
506 blood cells from rheumatoid arthritis and osteoarthritis patients. *Regul Pept.* 2003;115,
507 19-24.
- 508 59. Pulsatelli L, Dolzani P, Silvestri T, De Giorgio R, Salvarani C, Macchioni P, Frizziero
509 L, Meliconi R. Synovial expression of vasoactive intestinal peptide in polymyalgia
510 rheumatica. *Mol Pharm.* 2013;10(2):728-38.
- 511 60. Dvoráková MC. Cardioprotective role of the VIP signaling system. *Drug News*
512 *Perspect.* 2005 Jul-Aug;18(6):387-91.
- 513 61. Onoue S, Ohmori Y, Endo K, Yamada S, Kimura R, Yajima T. Vasoactive intestinal
514 peptide and pituitary adenylate cyclase-activating polypeptide attenuate the cigarette

- 515 smoke extract-induced apoptotic death of rat alveolar L2 cells. *Eur.J.Biochem.*
516 2004;271, 1757-1767.
- 517 62. Juarranz Y, Gutierrez-Canas I, Santiago B, Carrion M, Pablos JL, Gomariz RP.
518 Differential expression of vasoactive intestinal peptide and its functional receptors in
519 human osteoarthritic and rheumatoid synovial fibroblasts. *Arthritis Rheum.* 2008;58,
520 1086-1095.
- 521 63. Carrion M, Perez-Garcia S, Jimeno R, Juarranz Y, Gonzalez-Alvaro I, Pablos JL,
522 Gutierrez-Canas I, Gomariz RP. Inflammatory mediators alter interleukin-17 receptor,
523 interleukin-12 and -23 expression in human osteoarthritic and rheumatoid arthritis
524 synovial fibroblasts: immunomodulation by vasoactive intestinal Peptide.
525 *Neuroimmunomodulation.* 2013;20, 274-284.
- 526 64. Botz B, Bolcskei K, Kereskai L, Kovacs M, Nemeth T, Szigeti K, Horvath I, Mathe D,
527 Kovacs N, Hashimoto H, Reglodi D, Szolcsanyi J, Pinter E, Mocsai A, Helyes Z.
528 Differential regulatory role of pituitary adenylate cyclase-activating polypeptide in the
529 serum-transfer arthritis model. *Arthritis Rheumatol.* 2014;66, 2739-2750.
- 530 65. Mobasheri A. The future of osteoarthritis therapeutics: emerging biological therapy.
531 *Curr.Rheumatol.Rep.* 2013;15, 385.
- 532 66. Chen G, Deng C, Li YP. TGF-beta and BMP signaling in osteoblast differentiation
533 and bone formation. *Int.J.Biol.Sci.* 2012;8, 272-288.
- 534 67. Kim JH, Liu X, Wang J, Chen X, Zhang H, Kim SH, Cui J, Li R, Zhang W, Kong Y,
535 Zhang J, Shui W, Lamplot J, Rogers MR, Zhao C, Wang N, Rajan P, Tomal J, Statz J,
536 Wu N, Luu HH, Haydon RC, He TC. Wnt signaling in bone formation and its
537 therapeutic potential for bone diseases. *Ther.Adv.Musculoskelet.Dis.* 2013;5, 13-31.

- 538 68. Pan A, Chang L, Nguyen A, James AW. A review of hedgehog signaling in cranial
539 bone development. *Front Physiol* 2013;4, 61.
- 540 69. Wang L, Park P, La MF, Than K, Rahman S, Lin CY. Bone formation induced by
541 BMP-2 in human osteosarcoma cells. *Int.J.Oncol.* 2013;43, 1095-1102.
- 542 70. Zhang R, Edwards JR, Ko SY, Dong S, Liu H, Oyajobi BO, Papasian C, Deng HW,
543 Zhao M. Transcriptional regulation of BMP2 expression by the PTH-CREB signaling
544 pathway in osteoblasts. *PLoS.One.* 2011;6, e20780.
- 545 71. Ehlen HW, Buelens LA, Vortkamp A. Hedgehog signaling in skeletal development.
546 *Birth Defects Res.C.Embryo.Today* 2006;78, 267-279.
- 547 72. Jonason JH, Xiao G, Zhang M, Xing L, Chen D. Post-translational Regulation of
548 Runx2 in Bone and Cartilage. *J.Dent.Res.* 2009;88, 693-703.
- 549 73. Lerner UH, Persson E. Osteotropic effects by the neuropeptides calcitonin gene-
550 related peptide, substance P and vasoactive intestinal peptide.
551 *J.Musculoskelet.Neural.Interact.* 2008;8, 154-165.
- 552 74. Hohmann EL, Elde RP, Rysavy JA, Einzig S, Gebhard RL. Innervation of periosteum
553 and bone by sympathetic vasoactive intestinal peptide-containing nerve fibers. *Science*
554 1986;232, 868-871.
- 555 75. Bataille C, Mauprivez C, Hay E, Baroukh B, Brun A, Chaussain C, Marie PJ, Saffar
556 JL, Cherruau M. Different sympathetic pathways control the metabolism of distinct bone
557 envelopes. *Bone.* 2012 May;50(5):1162-72

- 558 76. Suzuki A, Kotoyori J, Oiso Y, Kozawa O. Pituitary adenylate cyclase-activating
559 polypeptide induces cAMP production independently from vasoactive intestinal
560 polypeptide in osteoblast-like cells. *Cell Signal*. 1994;6, 11-16.
- 561 77. Onuoha GN. Circulating sensory peptide levels within 24 h of human bone fracture.
562 *Peptides*. 2001;22:1107-10.
- 563 78. Ma W, Zhang X, Shi S, Zhang Y. Neuropeptides stimulate human osteoblast activity
564 and promote gap junctional intercellular communication.
565 *Neuropeptides*. 2013;47(3):179-86.
- 566 79. Lundberg P, Boström I, Mukohyama H, Bjurholm A, Smans K, Lerner UH. Neuro-
567 hormonal control of bone metabolism: vasoactive intestinal peptide stimulates alkaline
568 phosphatase activity and mRNA expression in mouse calvarial osteoblasts as well as
569 calcium accumulation mineralized bone nodules. *Regul Pept*. 1999;85, 47-58.
- 570 80. Okura H, Sato S, Kishikawa S, Kaneto S, Nakashima T, Yoshida N, Takayanagi H,
571 Kiyono H. Runx2-I isoform contributes to fetal bone formation even in the absence of
572 specific N-terminal amino acids. *PLoS.One*. 2014;9, e108294.
- 573 81. Li TF, Dong Y, Ionescu AM, Rosier RN, Zuscik MJ, Schwarz EM, O'Keefe RJ, Drissi
574 H. Parathyroid hormone-related peptide (PTHrP) inhibits Runx2 expression through
575 the PKA signaling pathway. *Exp.Cell Res*. 2004;299, 128-136.
- 576 82. Zhang X, Akech J, Browne G, Russell S, Wixted JJ, Stein JL, Stein GS, Lian JB.
577 Runx2-smad signaling impacts the progression of tumor-induced bone disease.
578 *Int.J.Cancer*. 2014; doi: 10.1002/ijc.29094.
- 579 83. Pitts RL, Wang S, Jones EA, Symes AJ. Transforming growth factor-beta and ciliary
580 neurotrophic factor synergistically induce vasoactive intestinal peptide gene

- 581 expression through the cooperation of Smad, STAT, and AP-1 sites. *J.Biol.Chem.*
582 2001;276, 19966-19973.
- 583 84. Arsenijevic T, Gregoire F, Chiadak J, Courtequisse E, Bolaky N, Perret J, Delporte C.
584 Pituitary adenylate cyclase activating peptide (PACAP) participates in adipogenesis by
585 activating ERK signaling pathway. *PLoS.One.* 2013;8, e72607.
- 586 85. Persson E, Lerner UH. The neuropeptide VIP regulates the expression of
587 osteoclastogenic factors in osteoblasts. *J.Cell Biochem.* 2011;112, 3732-3741.
- 588 86. Maeda Y, Sekiguchi F, Yamanaka R, Sugimoto R, Yamasoba D, Tomita S, Nishikawa
589 H, Kawabata A. Mechanisms for proteinase-activated receptor 1-triggered
590 prostaglandin E2 generation in mouse osteoblastic MC3T3-E1 cells. *Biol.Chem.* 2014;
591 doi: 10.1515/hsz-2014-0148
- 592 87. Mustafa T, Grimaldi M, Eiden LE. The hop cassette of the PAC1 receptor confers
593 coupling to Ca²⁺ elevation required for pituitary adenylate cyclase-activating
594 polypeptide-evoked neurosecretion. *J.Biol.Chem.* 2007;282, 8079-8091.
- 595 88. Li D, Jiao J, Shatos MA, Hodges RR, Dartt DA. Effect of VIP on intracellular [Ca²⁺],
596 extracellular regulated kinase 1/2, and secretion in cultured rat conjunctival goblet
597 cells. *Invest Ophthalmol.Vis.Sci.* 2013;54, 2872-2884.
- 598 89. Miraoui H, Oudina K, Petite H, Tanimoto Y, Moriyama K, Marie PJ. Fibroblast
599 growth factor receptor 2 promotes osteogenic differentiation in mesenchymal cells via
600 ERK1/2 and protein kinase C signaling. *J.Biol.Chem.* 2009;284, 4897-4904.

- 601 90. May V, Clason TA, Buttolph TR, Girard B.M, Parsons RL. Calcium Influx, But Not
602 Intracellular Calcium Release, Supports PACAP-Mediated ERK Activation in HEK
603 PAC1 Receptor Cells. *J.Mol.Neurosci.* 2014
- 604 91. Hayashi K, Endoh T, Shibukawa Y, Yamamoto T, Suzuki T. VIP and PACAP inhibit
605 L-, N- and P/Q-type Ca²⁺ channels of parasympathetic neurons in a voltage
606 independent manner. *Bull.Tokyo Dent.Coll.* 2002;43, 31-39.
- 607 92. Harfi I, Sariban E. Mechanisms and modulation of pituitary adenylate cyclase-
608 activating protein-induced calcium mobilization in human neutrophils.
609 *Ann.N.Y.Acad.Sci.* 2006;1070, 322-329.
- 610 93. Payet MD, Bilodeau L, Breault L, Fournier A, Yon L, Vaudry H, Gallo-Payet N.
611 PAC1 receptor activation by PACAP-38 mediates Ca²⁺ release from a cAMP-
612 dependent pool in human fetal adrenal gland chromaffin cells. *J.Biol.Chem.* 2003;278,
613 1663-1670.
- 614 94. Sandor B, Fintor K, Felszeghy S, Juhasz T, Reglodi D, Mark L, Kiss P, Jungling A,
615 Fulop BD, Nagy AD, Hashimoto H, Zakany R, Nagy A, Tamas A. Structural and
616 Morphometric Comparison of the Molar Teeth in Pre-eruptive Developmental Stage
617 of PACAP-Deficient and Wild-Type Mice. *J.Mol.Neurosci.* 2014; doi
618 10.1007/s12031-014-0392-6
- 619 95. Lundberg P, Lundgren I, Mukohyama H, Lehenkari PP, Horton MA, Lerner UH.
620 Vasoactive intestinal peptide (VIP)/pituitary adenylate cyclase-activating peptide
621 receptor subtypes in mouse calvarial osteoblasts: presence of VIP-2 receptors and
622 differentiation-induced expression of VIP-1 receptors. *Endocrinology* 2001;142, 339-
623 347.

- 624 96. Winding B, Wiltink A, Foged NT. Pituitary adenylyl cyclase-activating polypeptides
625 and vasoactive intestinal peptide inhibit bone resorption by isolated rabbit osteoclasts.
626 *Exp.Physiol* 1997;82, 871-886.
- 627 97. Akopian A, Demulder A, Ouriaghli F, Corazza F, Fondu P, Bergmann P. Effects of
628 CGRP on human osteoclast-like cell formation: a possible connection with the bone
629 loss in neurological disorders? *Peptides*. 2000;21:559-64.
- 630 98. Franceschi RT, Xiao G. Regulation of the osteoblast-specific transcription factor,
631 Runx2: responsiveness to multiple signal transduction pathways. *J.Cell Biochem*.
632 2003;88, 446-454.
- 633 99. Park HJ, Baek K, Baek JH, Kim HR. The Cooperation of CREB and NFAT is
634 Required for PTHrP-Induced RANKL Expression in Mouse Osteoblastic Cells. *J.Cell*
635 *Physiol*. 2014; doi: 10.1002/jcp.24790.
- 636 100. Yoo YM, Kwag JH, Kim KH, Kim CH. Effects of neuropeptides and mechanical
637 loading on bone cell resorption in vitro. *Int J Mol Sci*. 2014;15(4):5874-83.
- 638

638

639 Figure 1. Signalling pathways of PACAP induced chondrogenesis. The increased
640 concentration of cAMP level elevates PKA activity. Phosphorylated form of the downstream
641 targets of PKA such as CREB and Sox9 translocate into the nucleus of chondrogenic cells and
642 induce the gene expression of collagen type II., aggrecan and various GAG such as hyaluronic
643 acid. Activation of PAC1 receptor can also elevate the intracellular Ca^{2+} concentration leading
644 to increased PP2B, PKC or MAPK signalling activity. The elevated expression and nuclear
645 presence of PP2B regulated NFAT4 are also responsible for the augmented matrix production.

646

647 Figure 2. Multiple regulation connections' of PACAP signalling pathways in osteogenic
648 differentiation. PACAP binding to its receptors elevates the intracellular cAMP concentration
649 and activates PKA in osteoblast cells. CREB, the canonical downstream target of the kinase is
650 not significantly activated (arrows crossed by red lines) but the nuclear localisation of Runx2
651 is elevated. Although the cAMP regulated pathway is active the presence of the neuropeptide
652 does not result in a Ca^{2+} concentration increase, subsequently the Ca^{2+} dependent signalling
653 pathways are not activated (arrows crossed by red lines). PACAP also induces the expression
654 of BMPs which may crosstalk via the nuclear activity of Smad1 with Runx2 transcription
655 factor. SHH binding to PTCH1 receptor can induce the nuclear translocation of Gli1
656 transcription factor which is suppressed by the increased activation of PKA.

657

658

659

660

