

Structures and Activities of Tiahuramides A-C, Cyclic Depsipeptides
from a Tahitian Collection of the Marine Cyanobacterium *Lyngbya majuscula*

Annabel Levert,^{†,□} Rebeca Alvariño,[‡] Louis Bornancin,[†] Eliane Abou Mansour,^{†,▽} Adam M. Burja,^{§,○}
Anne-Marie Genevière,[±] Isabelle Bonnard,^{†,||} Eva Alonso,[‡] Luis Botana[‡] and Bernard Banaigs^{*†,||}

[†] CRIOBE, USR CNRS-EPHE-UPVD 3278, Université de Perpignan, 66860 Perpignan, France.

[‡] Departamento de Farmacología, Facultad de Veterinaria, Universidad de Santiago de Compostela, Lugo 27003, Spain.

[§] Heriot-Watt University, Edinburgh, Scotland EH14 4 AS.

[±] Sorbonne Universités, UPMC Univ Paris 06, CNRS, Biologie Intégrative des Organismes Marins (BIOM), Observatoire Océanologique, F-66650, Banyuls/Mer, France.

^{||} Laboratoire d'Excellence "CORAIL", France.

- * To whom correspondence should be addressed. Tel: +33 4 68 662074. Fax: +33 4 68 662223.
E-mail: banaigs@univ-perp.fr.
□ Present address: SAS AkiNaO, Perpignan, France.
▽ Present address: Université de Neuchâtel, Switzerland.
○ Present address: DSM Nutritional Products, 6480 Dobbin Road, Columbia, MD, 21045, USA.

A.L., R.A. and L.B. contributed equally to this work.

SUPPORTING INFORMATION

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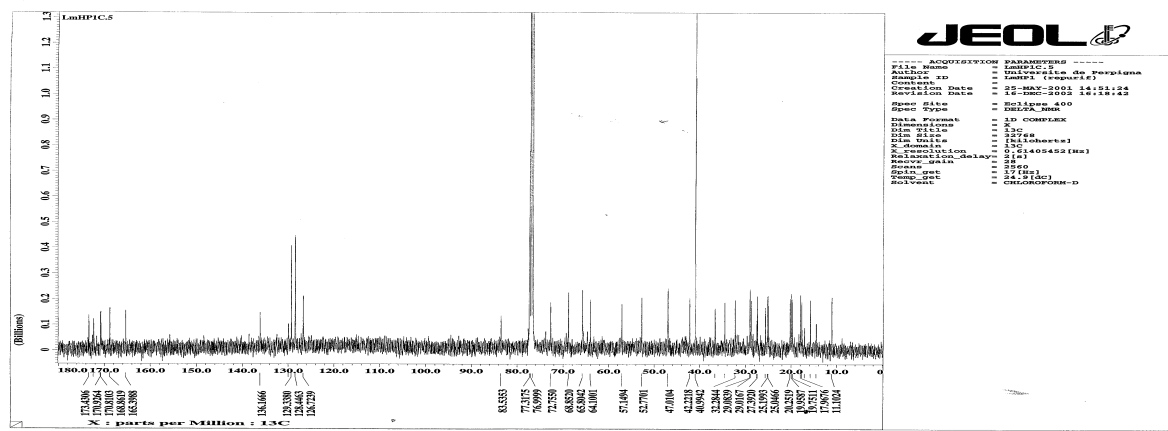
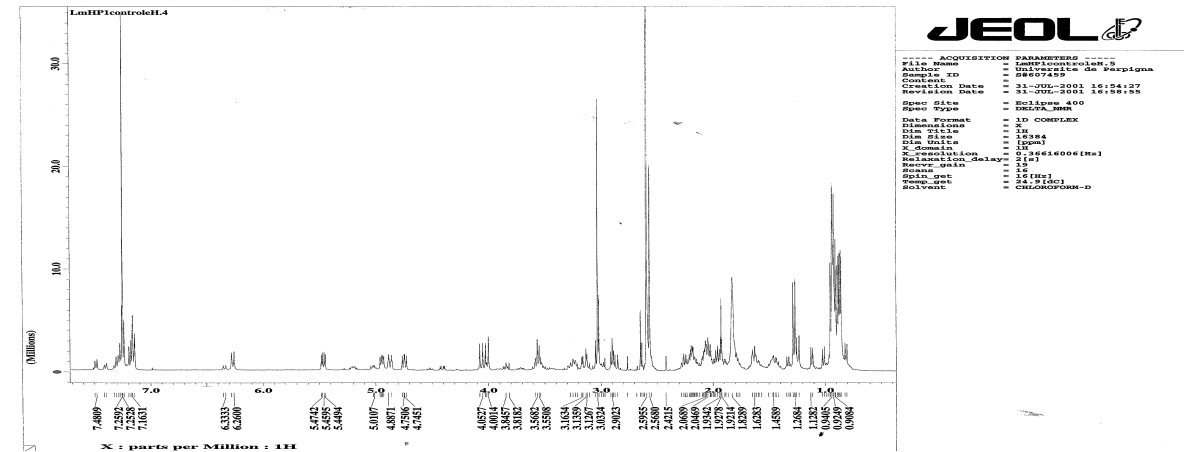
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- **NMR Measurement Conditions.** All spectra were obtained with a NM-40TH5 dual ^1H , ^{13}C probe in a JEOL EX400 operating at 400 MHz for proton and 100.53 MHz for carbon-13 at 298 K. ^1H and ^{13}C NMR chemical shifts are referenced to solvent peaks: δ_{H} 7.24 (residual CHCl_3), δ_{C} 77.0 for CDCl_3 . Tiahuramide A (7 mg), tiahuramide B (4 mg) and tiahuramide C (3.5 mg) were dissolved in a 5 mm tube in 0.75 ml of CDCl_3 .

Two-dimensional (2D) homonuclear correlated experiments DQF-COSY, HOHAHA and ROESY were all acquired using standard procedures with a spectral width of approximately 4000 Hz in both columns F_1 and F_2 . HOHAHA and ROESY were acquired in the phase sensitive mode. The time domain matrix consisted of 256 points in t_1 and 1024-2048 points in t_2 with 64-128 acquisitions for 256 experiments in t_1 . Data sets were zero-filled to 512 points in t_1 prior to Fourier transformation to obtain a frequency domain matrix of 512 x 1024-2048 real data points. Squared sine bell apodization functions were used. The HOHAHA spectra were recorded with a mixing time of 100 ms. ROESY spectra were measured with mixing times of 150, 250 and 350 ms. Heteronuclear correlated experiments were performed in ^1H -detected mode using the standard pulse programs HSQC and HMBC with a spectral width of approximately 20,000 Hz in F_1 and 4,000 Hz in F_2 . The time domain matrix consisted of 256 points in t_1 and 2048 points in t_2 with 128 acquisitions for 256 experiments in t_1 . Data sets were zero-filled to 512 points in t_1 prior to Fourier transformation to obtain a frequency domain matrix of 512 x 2048 real data points. The evolution delay was set to optimize 140 Hz couplings for HSQC and 8 Hz couplings for HMBC. Squared sine bell apodization functions were used.

- Figure S1. ¹H, ¹³C, DEPT NMR spectra of tiahuramide A (1).

Tiahuramide A /¹H & ¹³C



Tiahuramide A / DEPT

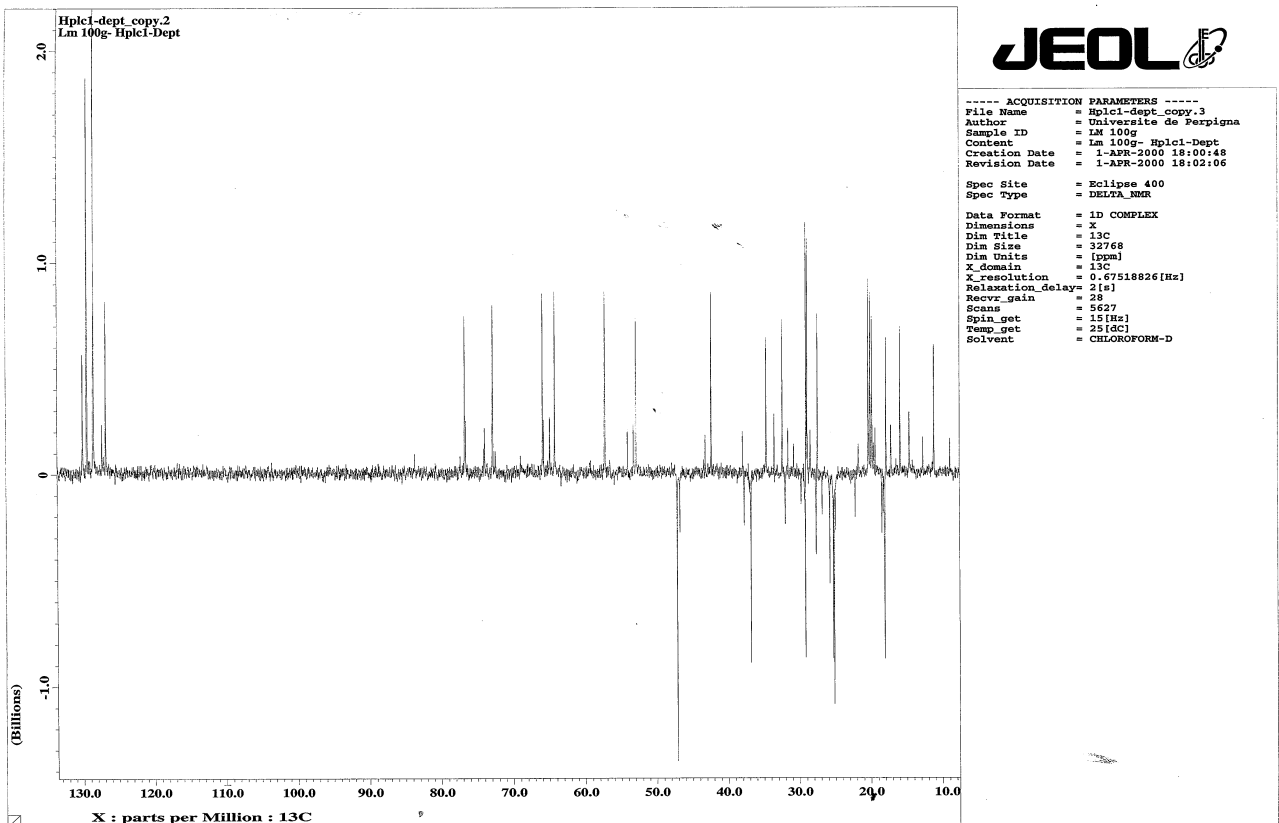
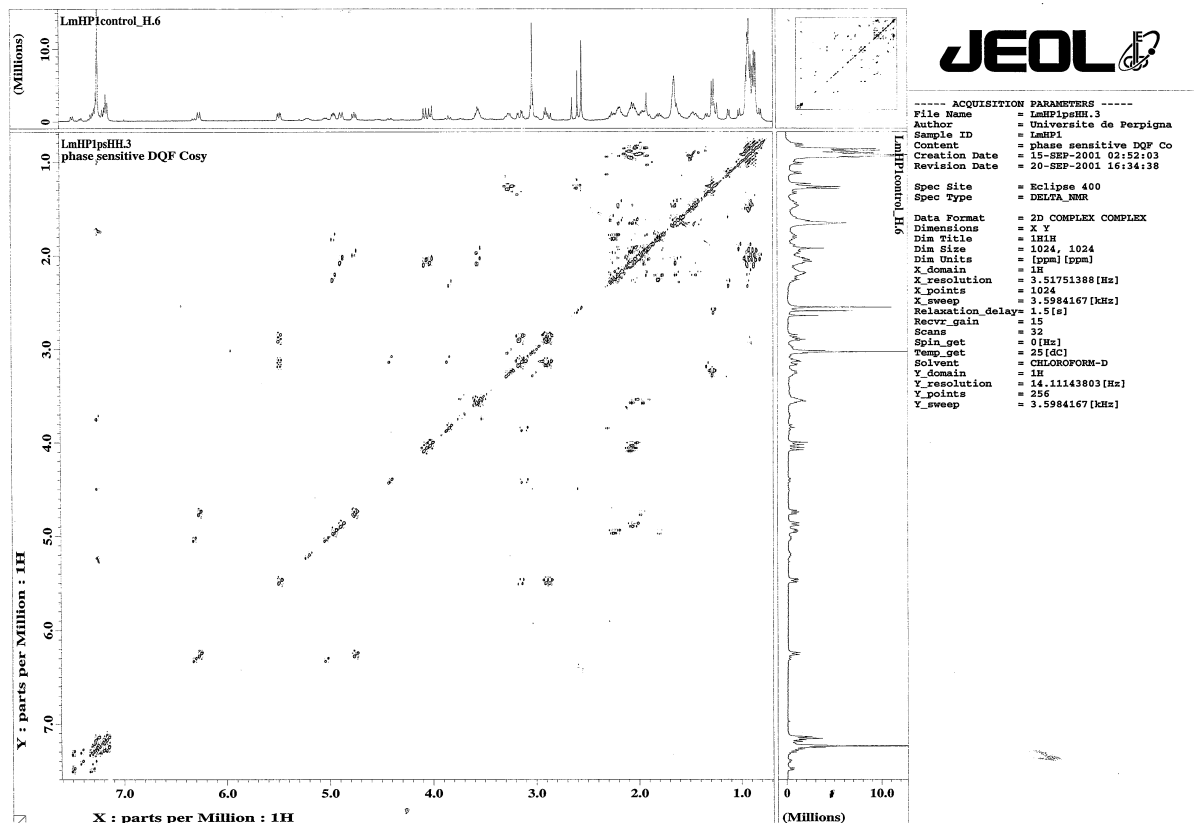


Figure S2. DQF COSY and HOHAHA NMR spectra of tiahuramide A (1).

Tiahuramide A / DQF COSY



Tiahuramide A / HOHAHA 50 ms

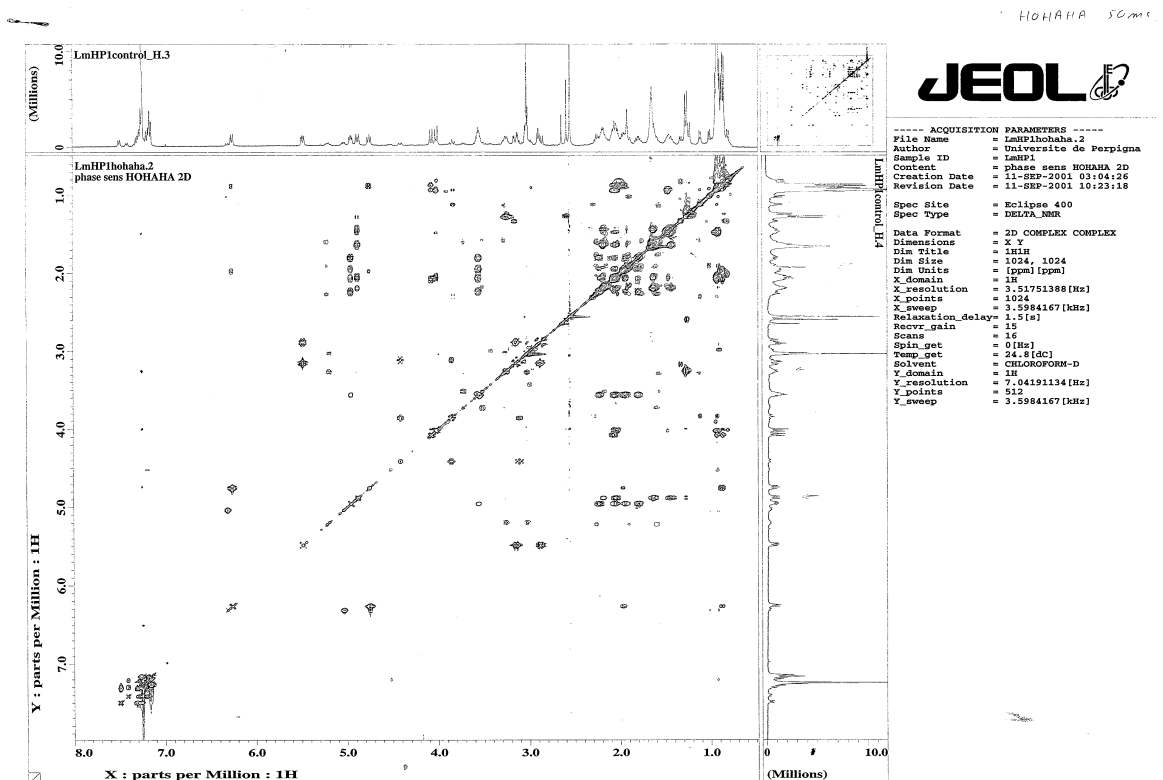
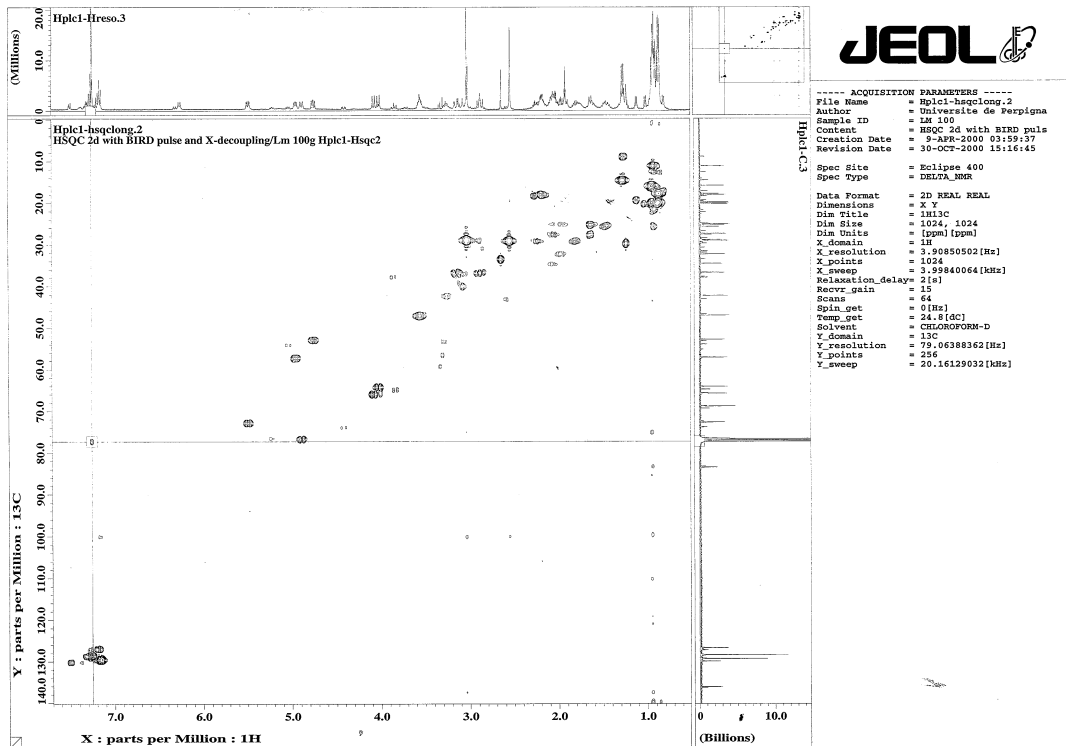
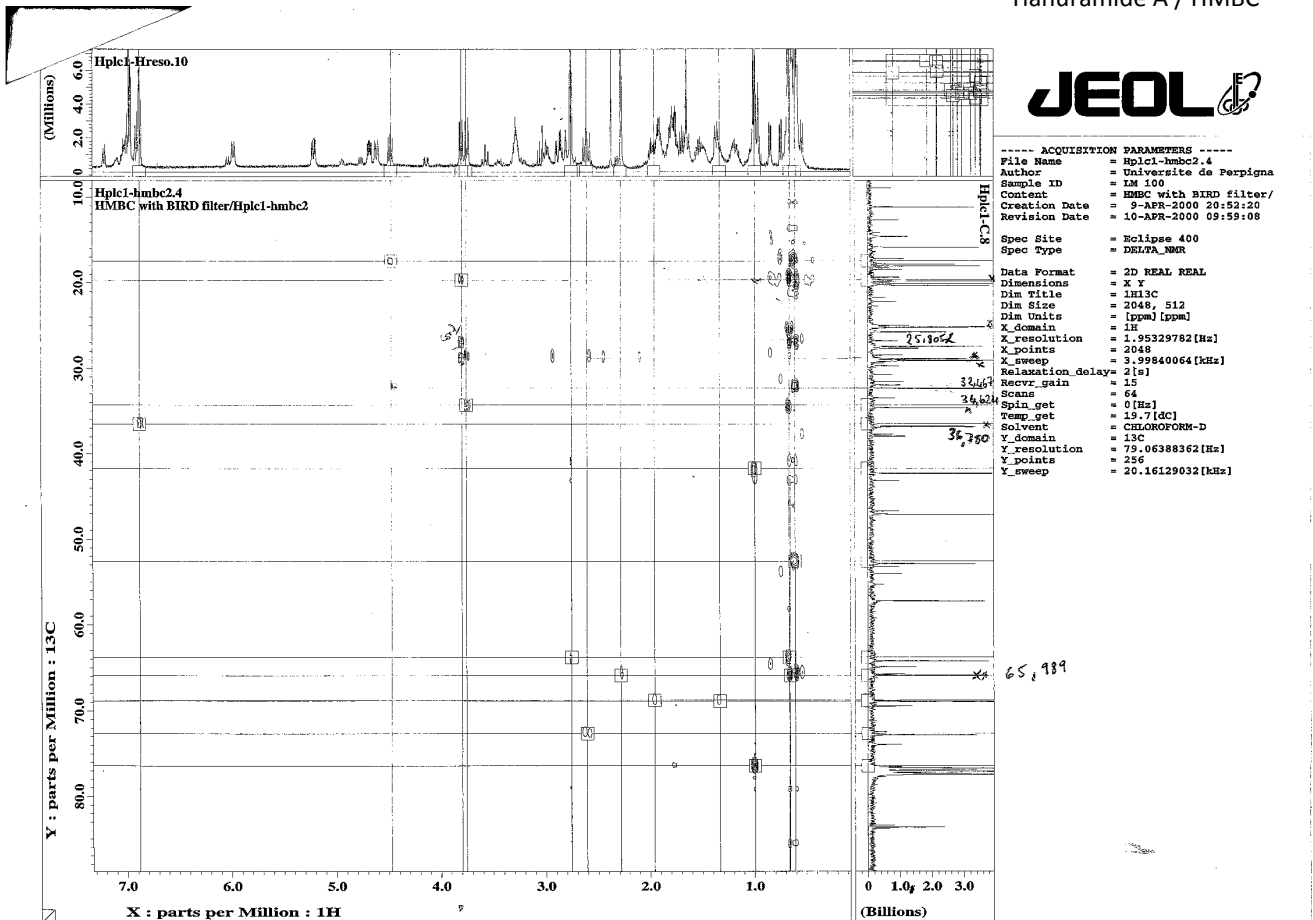


Figure S3. HSQC and HMBC NMR spectra of tiahuramide A (1).

Tiahuramide A / HSQC

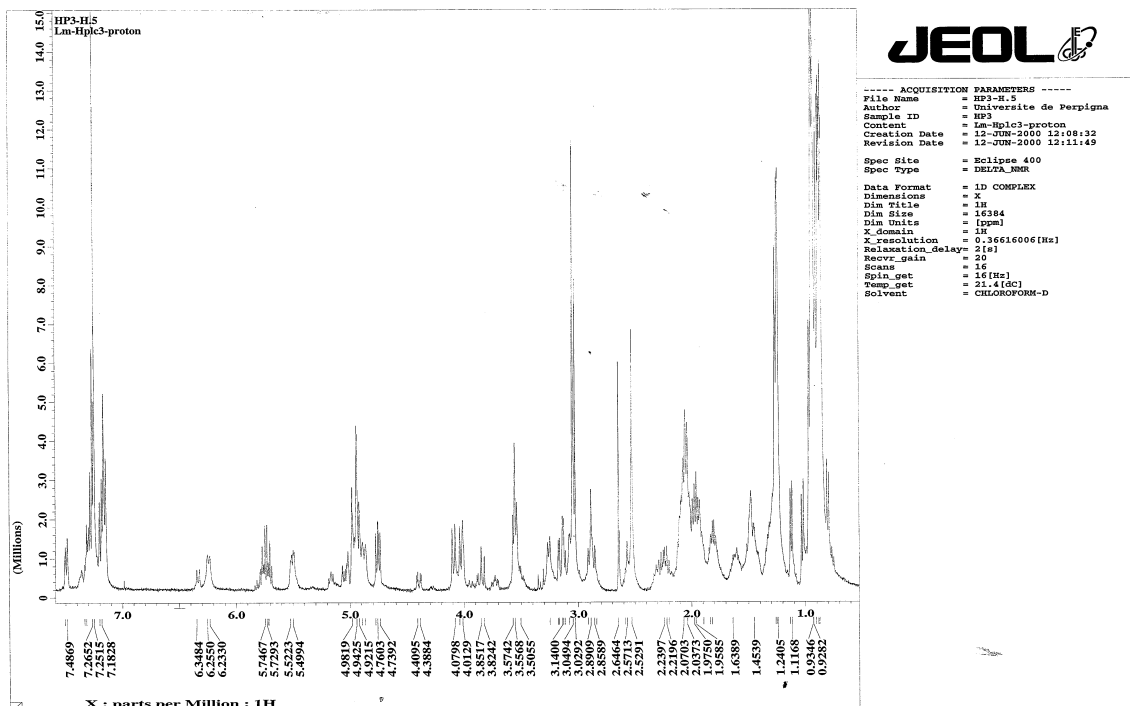


Tiahuramide A / HMBC



- Figure S4. ¹H spectra of tiahuramides B (2) and C (3).

Tiahuramide B / ¹H



Tiahuramide C / ¹H

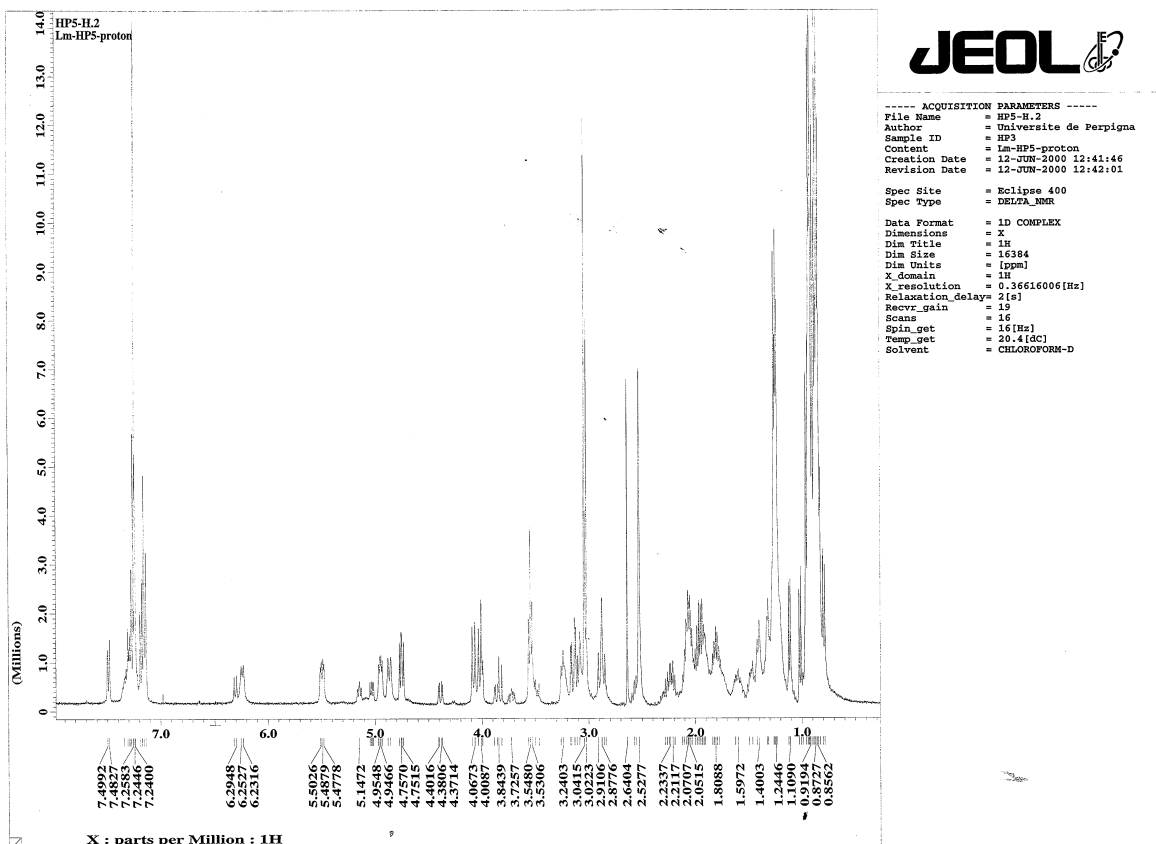
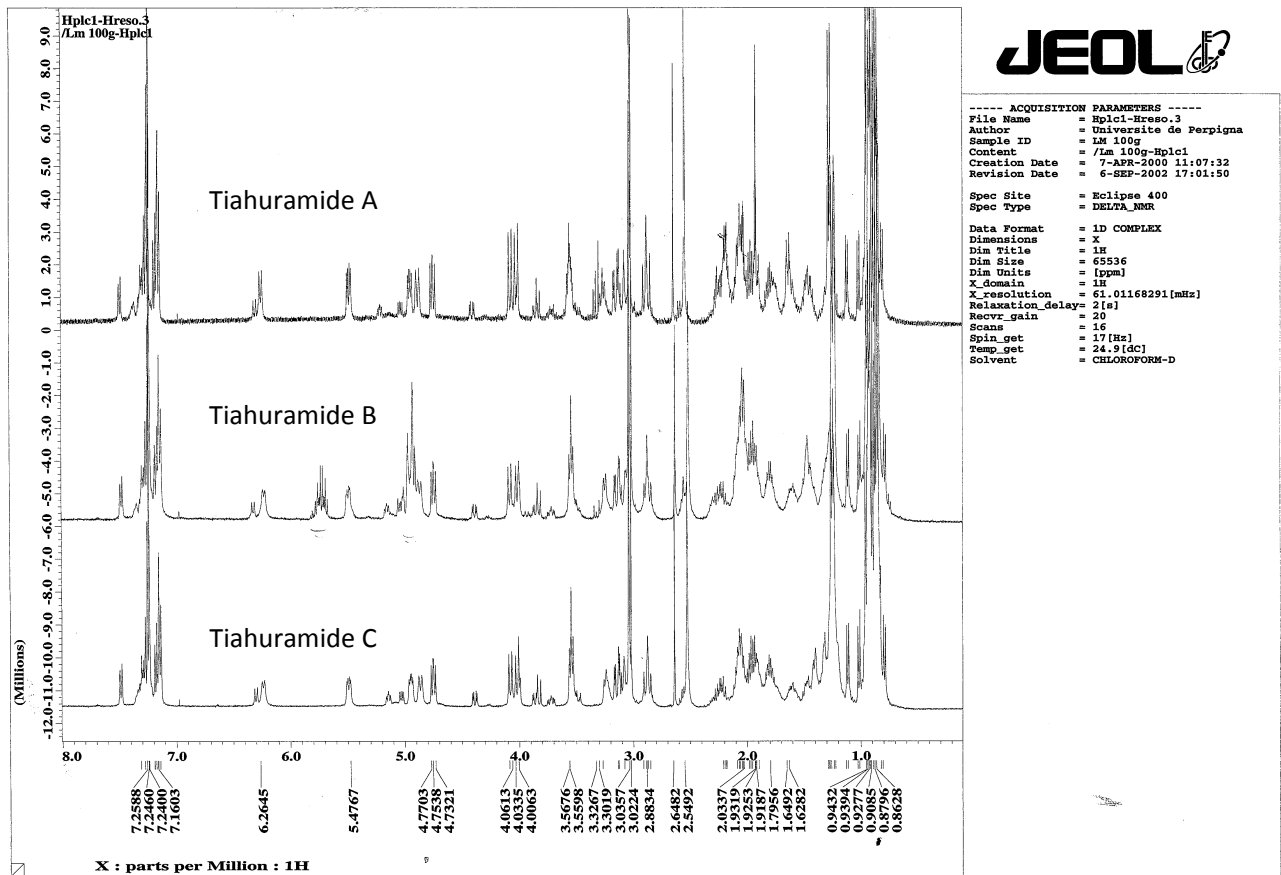


Figure S5. Comparison of ^1H and ^{13}C NMR spectra of tiahuramides A-C (1-3).

Tiahuramides A-C / ^1H comparison



Tiahuramides A-C / ^{13}C comparison

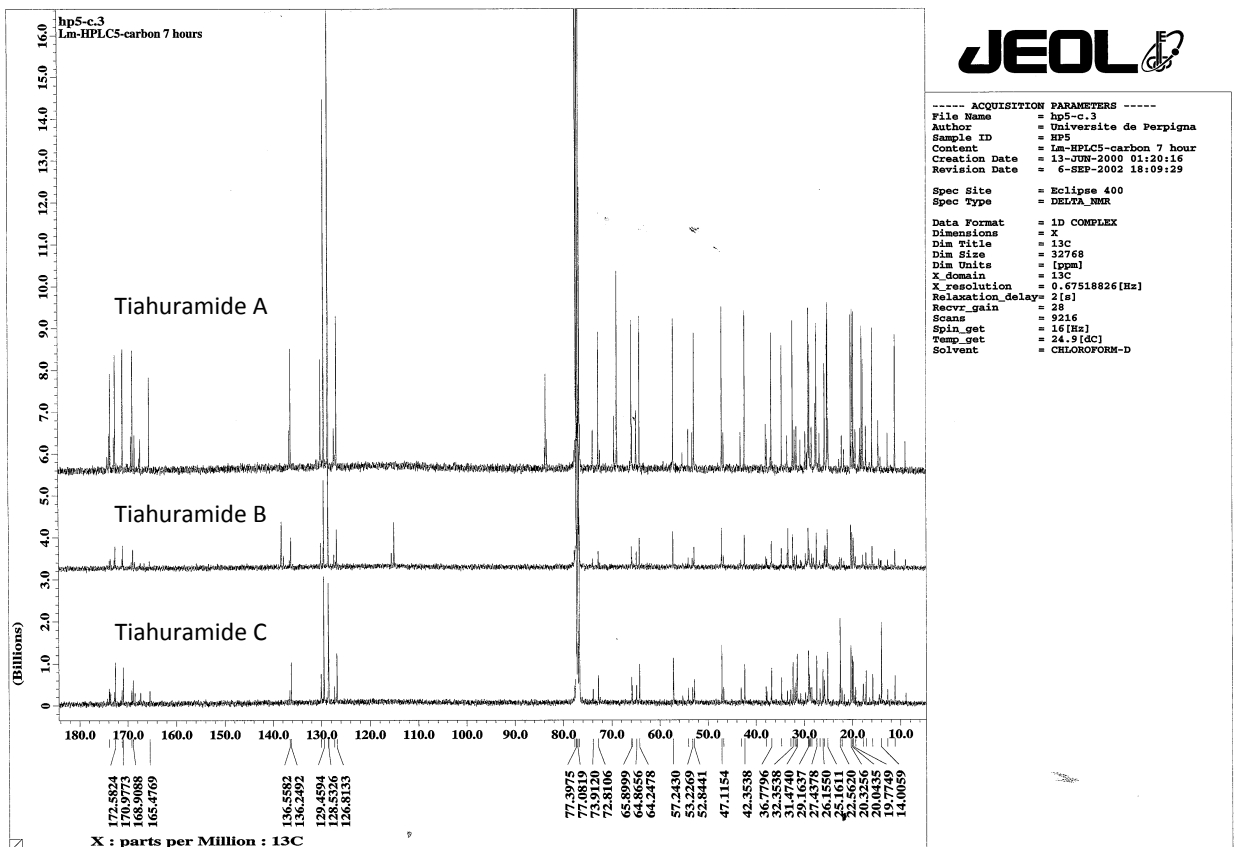


Figure S6. HPLC chromatograms of the Marfey's analysis.

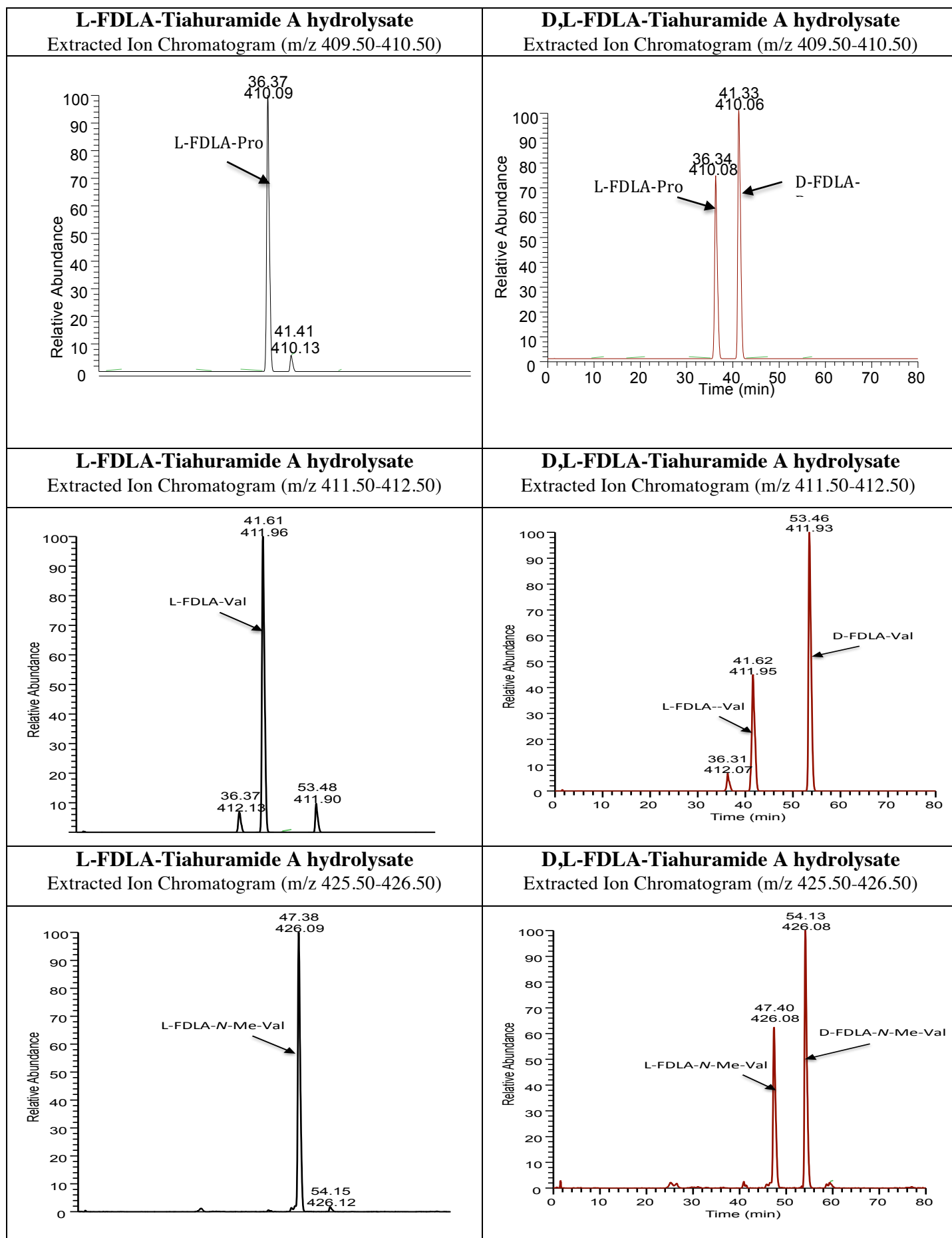


Table S1: Retention times (min) of standard and natural Marfey's *N*-Me-Ile derivatives

	Standard <i>N</i> -Me-Ile		<i>N</i> -Me-Ile from Tiahuramide A hydrolysate
	<i>N</i> -Me-L- <i>allo</i> -Ile (2 <i>S</i> ,3 <i>R</i>)-Ile	<i>N</i> -Me-L-Ile (2 <i>S</i> ,3 <i>S</i>)-Ile	<i>N</i> -Me-L-Ile (2 <i>S</i> ,3 <i>S</i>)-Ile
L-FDLA	52.12	51.78	51.78
D-FDLA	59.26	59.08	59.10

Gradient : from 10% CH₃CN-90% 0.01 M formic acid to 50%-50% at 0.3 mL/min over 70 min, then to 80%-20% over 10 min

Figure S7. HPLC chromatograms of the Marfey's analysis of standard and natural Marfey's *N*-Me-Ile derivatives

