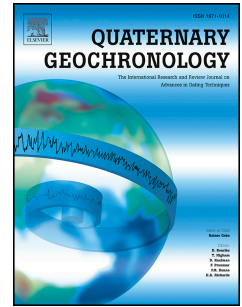


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1 **Protein diagenesis in archaeological gastropod shells and the suitability of this material**
2 **for amino acid racemisation dating: *Phorcus lineatus* (da Costa, 1778)**

3

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12

13 **Abstract**

14

15 The inter- and intra-crystalline fractions of the topshell *Phorcus lineatus* recovered from
16 modern specimens and shells from archaeological sites in Northern Spain covering Neolithic,
17 Mesolithic, and Upper Magdalenian periods were examined for amino acid composition and
18 racemisation over time. The main loss of proteins from the inter-crystalline fraction occurred
19 within the first 6,000 years after the death of the organism. In contrast, the intra-crystalline
20 fraction isolated by bleaching—with a different protein composition to that of the inter-
21 crystalline fraction—appeared to behave like a closed system for at least 12.6 ka, as reflected
22 by the lack of a significant decrease in amino acid content. However, changes in the relative

1

23 composition of the amino acids present in these shells occurred during this period. The
24 concentration of aspartic acid remained almost constant with age within the intra-crystalline
25 fraction and its contribution to the total amino acid content also remained the same. Good
26 correspondence was obtained between Asx D/L values in unbleached and bleached samples
27 and age, thereby allowing the dating of archaeological sites and the determination of
28 chronometric age.

29

30 **Key-words:** *Phorcus lineatus*; inter- and intra-crystalline proteins; amino acids;
31 microstructure; archaeology

32

33 **Highlights:**

34

- 35 - Inter- and intra-crystalline protein fractions of *P. lineatus* shells differ (amino acid
36 proportions).
- 37 - The main loss of proteins (85-90%) from the inter-crystalline fraction occurs within
38 the first 6 ka.
- 39 - The intra-crystalline protein fraction behaves like a closed-system.
- 40 - Asx D/L in unbleached and bleached *P. lineatus* specimens can be used for
41 chronological purposes over ~ 13 ka.
- 42 - The percentage of aspartic acid remained constant in intra-crystalline proteins for over
43 ca. 13 ka.

44

45 **1. Introduction**

46

47

48 Dating archaeological sites is crucial for interpreting changes in past human behaviour and
49 for reconstructing environmental conditions. In recent decades, radiocarbon dating has
50 become the most common approach for the chronological assessment of archaeological sites
51 (Taylor, 1987; Stuiver and Braziunas, 1993; Bronck Ramsey et al., 2004; Reimer et al.,
52 2013). However, the limitations of the method (e.g. expense, time-constraints) makes it
53 difficult to date large numbers of samples. In addition, radiocarbon dating is not suitable for
54 dating samples older than 50 ka (Walker, 2005; Chiu et al., 2007; Reimer et al., 2013). In this
55 context, it is therefore necessary to develop cheaper and faster methods for the chronological
56 analysis of archaeological deposits. Amino acid racemisation (AAR) is one of the most used
57 alternative methods to radiocarbon dating, as it is a faster and less expensive technique,
58 allowing the dating of archaeological sites (Masters and Bada, 1977; Wehmiller, 1977).
59 While AAR dating goes beyond the time range of the radiocarbon method, it has also been
60 employed for dating Holocene sites (e.g. Bateman, 2008; Ortiz et al., 2009, 2015; Demarchi
61 et al., 2011). Moreover, many studies have demonstrated that AAR is a satisfactory tool for
62 dating a range of material from palaeontological and archaeological sites, such as teeth and
63 shells (Helfman and Bada, 1976; Wehmiller, 1977; Julg et al., 1987; Bateman, 2008; Torres
64 et al., 2013, among others).

65 Shell middens are unique archaeological deposits composed of large amounts of shells that
66 were discarded by humans in the past after use or consumption of their content (Waselkov,
67 1987; Stein, 1992; Colonese et al., 2011; Gutiérrez-Zugasti et al., 2011). Understanding shell
68 midden formation/transformation/erosion, as well as changes in subsistence strategies and

3

69 settlement patterns of human groups, usually requires a large number of dates to be obtained.
70 In this regard, AAR can be helpful for chronological purposes in this type of context, as a
71 large number of samples can be analysed from a single horizon, thus facilitating the
72 identification of time-averaging and the time over which a certain site formed (Kowalewski
73 et al., 1998), or potential anthropogenic heating (Demarchi et al., 2011).

74 In Atlantic Europe, previous studies have centered on the use of the limpet *Patella vulgata*
75 Linnaeus, 1758 for dating Palaeolithic, Mesolithic and Neolithic shell middens (Bateman,
76 2008; Ortiz et al., 2009, 2015; Demarchi et al., 2011). Recent studies of modern and
77 archaeological *P. vulgata* in northern Spain have shown the potential of inter- and intra-
78 crystalline proteins in the shells of this limpet for AAR geochronology (Ortiz et al., 2009,
79 2015; Demarchi et al., 2013a,b). In the studies of Demarchi et al. (2013a, b), artificial
80 diagenesis was induced in the whole protein content (inter- and intra-crystalline proteins) and
81 in the isolated intra-crystalline protein fraction (IcP) of modern *Patella* shells. The extent and
82 racemisation of various amino acids provided data on protein diagenesis in modern limpets,
83 showing that the IcP fraction behaves like a closed system and is thus suitable for
84 geochronological purposes. Ortiz et al (2015) revealed the patterns of protein degradation in
85 fossil *P. vulgata* representatives collected from several archaeological sites of diverse ages
86 (ca. 34 ka cal BP to 5.8 ka cal BP) in Northern Spain, by examining the amino acid content
87 and D/L values in the whole protein content and the IcP fraction separately. The main protein
88 leaching from the inter-crystalline fraction was observed to occur within the first 6 ka after
89 the death of the organism. In contrast, the IcP fraction, which has a distinct protein
90 composition to that of the inter-crystalline fraction, appeared to behave as a closed system for
91 at least 34 ka. Notwithstanding, Asx D/L values appeared to be suitable for geochronological
92 purposes even when considering the whole protein fraction, likely to be due to rapid initial
93 leaching of the inter-crystalline matrix (Ortiz et al., 2015).

94 In contrast, the inter-crystalline fraction of *Glycymeris* sp. shells does not seem to behave as a
95 closed system, with the inter- and intra-crystalline proteins probably being similar (Demarchi
96 et al., 2015, Ortiz et al., 2017). In spite of the high intra-sample variability, the extent of
97 racemisation in unbleached *Glycymeris* sp. shells should be used with caution for AAR dating
98 (Torres et al., 2014; Demarchi et al., 2015; Ortiz et al., 2017).

99 In a recent study, *Phorcus turbinatus* shells were subjected to AAR analysis in Ksâr 'Akil site
100 (Lebanon). IcPs provided a robust fraction for AAR dating, showing closed-system behaviour
101 (Bosch et al., 2015). However, the poor resolution of the D/L values obtained on multiple
102 amino acids hampered the usefulness of AAR for chronological applications within this site,
103 at least between 43 to 30 ka BP.

104 Therefore, further research is required to clarify the processes of protein preservation and
105 degradation and the concomitant success of AAR for dating archaeological localities using
106 molluscs. The quality of the archaeological record, as well as the range of species available
107 and their abundance, makes northern Spain an excellent area to test dating methods such as
108 AAR. Although the limpet *P. vulgata* is one of the most common species in archaeological
109 sites in northern Iberia, other molluscs are also present, including the topshell *Phorcus*
110 *lineatus* (da Costa, 1778) (syn. *Osilinus lineatus*). This mollusc, also known as toothed or
111 thick topshell, is commonly found in archaeological sites of a range of ages in northern Spain
112 (Table 1; González-Morales, 1982; Bailey and Craighead, 2003; Gutiérrez-Zugasti, 2009,
113 2011; Álvarez-Fernández, 2011), thereby allowing the analysis of long-term chronological
114 sequences. Previous mineralogical studies (by SEM and X-ray diffraction) of *P. lineatus*
115 shells (Fig. 2) have shown that they have a very thin calcite outer layer (with foliated and
116 prismatic structures) and an inner nacreous aragonite layer (Mannino et al., 2003; Mannino
117 and Thomas, 2007; Gutiérrez-Zugasti et al., 2015). These studies showed that the inner
118 aragonitic structures of archaeological *P. lineatus* shells remained unaltered and well

119 preserved over 8,000 yr time range, probably not undergoing recrystallization or post-
120 depositional isotopic exchange (Mannino, 2000; Mannino et al., 2003; Mannino and Thomas,
121 2007), therefore likely to be suitable for AAR dating.

122

123 Therefore, here we provide the background to the successful application of AAR of *P.*
124 *lineatus* for geochronological purposes. We report the systematic study of the behaviour of
125 the whole protein content (inter- and intra-crystalline proteins) and the IcP fraction (bleaching
126 tests) separately within this species. To this end, we did the following:

127 - Tested the patterns of diagenetic reactions and robustness of whole protein content and IcP
128 fraction during artificial diagenesis (leaching tests at high temperature).

129 - Compared the diagenetic patterns in archaeological representatives within the IcP fraction
130 and the whole-shell proteins. Shell specimens from archaeological sites of known ages (Fig. 1,
131 Table 1) covering the Upper Magdalenian (16.3-13.5 ka cal BP), Azilian (13.5-10.7 ka cal
132 BP), Mesolithic (10.7-6.3 ka cal BP), and Neolithic (ca. 6.3-5.7 ka cal BP) periods were
133 selected for analysis

134 - Evaluated the potential for artificial diagenesis at high temperature in order to mimic
135 diagenesis in archaeological sites, by comparing results from heated and archaeological
136 shells.

137 - Tested the suitability of AAR for dating purposes.

138

139 **2. Material and methods**

140

141 A total of 101 shell samples of *P. lineatus* were selected from 19 stratigraphic levels
142 belonging to 8 archaeological sites located in the regions of Asturias and Cantabria (Northern
143 Spain) and radiocarbon-dated to the Upper Palaeolithic, Mesolithic and Neolithic periods
144 (Fig. 1; Table 1). Shells were stored at the Museum of Archaeology of Asturias (MAA) and
145 the Museum of Prehistory and Archaeology of Cantabria (MUPAC). For comparative
146 purposes, 5 modern specimens (collected alive) were recovered from Cue beach (Asturias),
147 located close to the archaeological sites (Fig. 1).

148 Between 5 and 8 *P. lineatus* shells (analytical samples) from each archaeological level were
149 analysed for amino acid content (Table 2). In the laboratory, shells were carefully sonicated
150 and cleaned with water to remove sediment. Peripheral parts of the shells, approximately 20–
151 30%, were removed after chemical cleaning of the sample with 2 M HCl.

152 For all shell samples, we drilled a small disc in the aperture—a procedure that has been
153 shown to reduce intra-shell variability (cf. Murray-Wallace, 1995). Approximately 5–20 mg
154 of carbonate was extracted from each shell and subjected to AAR analysis of total protein
155 content (inter- and intra-crystalline proteins) and the isolation of IcP through bleaching
156 (Penkman et al., 2008; Demarchi et al., 2013a). Samples from the aperture of modern
157 specimens were also used to measure the amino acids in the total protein fraction and in the
158 IcP fraction after leaching (heating at 140°C over a range of time intervals).

159

160 **2.1 Leaching**

161

162 Leaching was performed following the protocol described in Canoira et al (2003) and Torres
163 et al (2017). A set of 20 modern shell samples were placed in borosilicate glass ampoules,
164 together with 2 g of quartz sand (deeply pre-cleaned by oven baking at 600°C for 6 h). Next,
165 120 ml of ultraclean water (HPLC-grade) was added with a syringe. The top of the ampoule
166 was fitted into rubber tubing connected to a vacuum-N₂ line, being alternately exposed to
167 vacuum and N₂, a procedure repeated three to four times to flush out all the air, following
168 Kriausakul and Mitterer (1978), Goodfriend and Meyer (1991) and Canoira et al. (2003). The
169 ampoule was later sealed under nitrogen. The ampoules were placed in a rack and put in an
170 oven at 140°C.

171 Two ampoules with quartz sand were removed at the following intervals: 1, 2, 4, 6, 8, 24, 48,
172 72, and 240 h. The ampoules were opened and dried. Shell samples were separated, washed
173 with distilled water, sonicated, and vacuum-dried. They were then analysed for total amino
174 acid content and IcP fraction after bleaching.

175 After heating, 100 mL of the supernatant water was also analysed for the amino acids leached
176 into the water (THAAw).

177

178 **2.2 Bleaching**

179

180 Powdered shell samples (from archaeological levels and leaching experiment) were used to
181 isolate IcPs. The shell particles measured less than 500 µm, following Demarchi et al.
182 (2013a, p. 151), a size for which bleaching is likely to be most effective. We exposed these
183 samples to 12% sodium hypochlorite (NaOCl) for 48 h—a time reported to be the optimal
184 bleaching period for some molluscs (Penkman et al., 2008; Demarchi et al., 2013a).

185 For each fraction, 50 μ L of NaOCl per mg of powdered shell was added to accurately
186 weighed subsamples at room temperature. To ensure the complete penetration of the
187 oxidising agent, the vials containing the powders and the bleach were shaken every 24 h. The
188 bleach was then removed, and the powders were rinsed five times in ultrapure water and once
189 in HPLC-grade methanol, with centrifugation for 4 min between each rinse to minimise the
190 removal of powder. Finally, the samples were air-dried overnight.

191

192 **2.3 Amino acid analysis**

193

194 Amino acid concentrations and racemisation/epimerisation ratios were quantified using a
195 HPLC, following the sample preparation protocol described in Kaufman and Manley (1998)
196 and Kaufman (2000). This procedure involves hydrolysis, which was performed under an N₂
197 atmosphere in 20 μ L/mg of 7 M HCl for 20 h at 100°C. The hydrolysates were evaporated to
198 dryness *in vacuo* and then rehydrated in 10 μ L/mg of 0.01 M HCl with 1.5 mM sodium azide
199 and 0.03 mM L-homo-arginine (internal standard).

200 Samples were injected into an Agilent HPLC-1100 equipped with a fluorescence detector.

201 Excitation and emission wavelengths were programmed at 230 nm and 445, respectively. A

202 Hypersil BDS C18 reverse-phase column (5 μ m; 250 x 4 mm i.d.) was used for the analysis.

203 Derivatisation was achieved before injection by mixing the sample (2 μ L) with the pre-

204 column derivatisation reagent (2.2 μ L), which comprised 260 mM isobutyryl-L-cysteine

205 (chiral thiol) and 170 mM o-phthalaldehyde, dissolved in a 1.0 M potassium borate buffer

206 solution at pH 10.4. Eluent A consisted of 23 mM sodium acetate with 1.5 mM sodium azide

207 and 1.3 mM EDTA, adjusted to pH 6.00 with 10 M sodium hydroxide and 10% acetic acid.

208 Eluent B was HPLC-grade methanol, and eluent C consisted of HPLC-grade acetonitrile. A

209 linear gradient was performed at 1.0 mL/min and 25°C, from 95% eluent A and 5% eluent B
210 upon injection to 76.6% eluent A, 23% eluent B, and 0.4% eluent C at min 31; and then with
211 a progressive gradient at 1.07 mL/min and the following percentages: 46.2% eluent A, 48.8%
212 eluent B, and 5.0% eluent C at min 95. As a laboratory routine, we separated glycine (Gly)
213 and the D and L peaks of the following amino acids (Fig. 1-Supplementary Data): aspartic
214 acid and asparagine (Asx); glutamic acid and glutamine (Glx); serine (Ser); alanine (Ala);
215 valine (Val); phenylalanine (Phe); isoleucine (Ile); leucine (Leu); threonine (Thr); arginine
216 (Arg); and tyrosine (Tyr).

217

218 **2.4 Data screening of the AAR analyses**

219

220 A total of 108 powdered samples taken from the aperture of archaeological *P. lineatus* shells
221 were analysed for amino acid content. The same 108 samples were also used for the
222 bleaching experiment. Of these samples, 14 results (12.9% of the data- 3 in El Penicial, 3 in
223 Bricia-A, 3 in Bricia-C, 1 in El Mazo-101, 1 in El Toral III-21, 2 in Mazaculos II-A2, and 1
224 in level 24 of La Riera) were excluded because Asx and Glx D/L values fell off the
225 covariance trend (cf. Kaufman, 2003, 2006; Laabs and Kaufman, 2003) (Supplementary
226 Data) and/or because of abnormally high D/L values, characterised by Asx D/L and Glx D/L
227 values falling outside the 2σ range of the group (cf. Hearty et al., 2004; Kosnik and Kaufman,
228 2008). These samples also showed a low amino acid content. A similar percentage of the data
229 set from bleached samples was also excluded following the same rejection criteria exposed
230 above, coinciding in most cases with the outliers identified for unbleached samples. Most of
231 the samples with high D/L values may have been subjected to anthropogenic heating (3 in El

232 Penicial, 1 in Bricia-A, 1 in El Mazo-101, 1 in El Toral III-21, and 1 in level 24 of La Riera),
233 as they showed a similar pattern to that of heated ostrich eggshells (Brooks et al., 1991;
234 Crisp, 2013) and suspected burned *P. vulgata* shells (Demarchi et al, 2011), i.e., the sum of
235 total amino acid concentrations were considerably lower, especially for Asx, Ser, Thr and
236 Arg. Each result and the samples rejected are shown in the Supplementary Data. The data
237 used in the following sections are only from the screened samples and do not include outliers.
238 None of the 20 powdered samples of modern *P. lineatus* shells that were analysed for total
239 amino acid content or the same 20 samples that were bleached for the isolation of the IcP
240 fraction were rejected.

241

242 **3 Modern *P. lineatus* shells**

243

244 ***3.1 Amino acid concentration and composition***

245

246 The total concentration of amino acids in modern shells was ca. 386 nmol/mg, whereas in the
247 IcP fraction registered ca. 16 nmol/mg (Table 3). Thus, the latter accounted for around 4.2%
248 of the total proteins in these shells (Fig. 3A).

249 The amino acid composition of the inter- and intra-crystalline proteins in modern shells also
250 differed, as the percentage of the individual concentration of Asx ([Asx]) was higher in
251 bleached (40%) than in unbleached samples (14%) (Table 3), the relative proportion of Ala
252 and Gly being higher in the latter (Fig. 3C).

253

254 ***3.2 Amino acid D/L values***

255

256 The mean Asx, Glx, Ala, Ser D/L values and D-allo/L-Ile values in bleached and unbleached
257 modern *P. lineatus* shells are shown in Fig. 3B. We selected these amino acids because they
258 account for a considerable percentage of the amino acid content in modern shells (Fig. 3C).
259 Asx, Glx, and Ala were the amino acids with highest D/L values in unbleached samples. In
260 contrast, Asx, Glx, Ala, and Ser D/L and D-allo/L-Ile values were higher in bleached samples
261 than in unbleached ones, the differences between Ser and Asx being significant.

262

263 **3.3 Discussion**

264

265 The IcPs in modern *P. lineatus* shells accounted for a small fraction with respect to the total
266 protein content (ca. 4.2%) (Fig. 3A). Acidic amino acids were not abundant in the whole
267 shell, representing only 22%. The relatively low percentages of acidic amino acids found in
268 *P. lineatus* may be explained by the aragonitic composition of the shell (Mannino et al., 2003;
269 Mannino and Thomas, 2007; Gutiérrez-Zugasti et al., 2015), as the presence of acidic and
270 Asp-rich proteins is usually linked to calcitic structures (Gotliv et al., 2005; Marin et al.,
271 2012). In the aragonite-dominated shells of *Margaritifera falcata* and *Bithynia tentaculata*,
272 Asx and Glx account for a low percentage (ca. 15-25%) of the amino acid content, although
273 in *Corbicula fluminalis* is ca. 45% (Penkman et al., 2008).

274 In contrast, acidic amino acids accounted for 45-50% of the IcP fraction (Fig. 3C). Similarly,
275 in some other mollusc shells, the percentage of Asx has been reported to increase after
276 bleaching (Penkman et al., 2008). This may be explained by the strong binding of acidic
277 amino acids to the mineral matrix observed in ostrich eggshell (Demarchi et al., 2016).

278 Moreover, we observed that the relative composition of other amino acids (Ala, Gly, Leu,
279 Val) in inter- and intra-crystalline fractions was dissimilar. This observation indicates that the

280 inter- and intra-crystalline protein compositions differ, thus potentially affecting the AAR
281 rates (Fig. 3B) (Penkman et al., 2008; Crisp, 2013; Demarchi et al., 2013a).

282 It is worth noting that, coinciding with previous studies, the D/L values were higher in the IcP
283 fraction of modern *P. lineatus* than in the whole shell. The differences found in the
284 concentration of amino acids and D/L values between the inter- and intra-crystalline proteins
285 were in agreement with the findings of Sykes et al. (1995) and Penkman et al. (2007, 2008),
286 who observed distinct racemisation rates in these fractions in a variety of mollusc shells.
287 These results could be due to the removal of certain proteins and amino acids (mainly free
288 amino acids-FAA) from the inter-crystalline matrix of the shells during bleaching (cf.
289 Penkman et al., 2007, 2008). According to Penkman et al. (2008), the loss of FAA—which
290 tend to be more highly racemised than the total hydrolysable amino acids (THAA) (Mitterer
291 and Kriausakul, 1984)—in the inter-crystalline during diagenesis produces a
292 decrease in D/L values for the THAA of the whole shell. In this regard, the higher
293 concentration of free amino acids (which are the most highly racemised) within the IcP
294 fraction (Fig. 3A) may explain the higher D/L values obtained in the intra-crystalline fraction
295 compared to whole shell (Fig. 3B). Other processes should also be taken into account, such as
296 the different contribution of the distinct amino acids to the proteins entrapped within the
297 biomineral, as some amino acids may be more tightly bound to the mineral (Demarchi et al.,
298 2016). Also, the position of each amino acid in the protein chains can cause a variation in
299 degradation rates (Kriausakul and Mitterer, 1980; Wehmiller, 1980, 1993; Mitterer and
300 Kriausakul, 1984), i.e., the rate of racemisation differs depending on the sequence position of
301 amino acids, with most amino acids racemising only when in a terminal position.

302

303 **4 Behaviour of shell proteins during artificial diagenesis**

304

305 **4.1 Amino acid concentrations vs. heating time**

306

307 The total concentration of amino acids decreased with heating time in unbleached samples
308 (Fig. 4). In contrast, the total concentration of amino acids remained similar during artificial
309 diagenesis in bleached *P. lineatus* shells.

310 After 6 h leaching at 140°C, approximately 60% of total amino acid content of unbleached
311 shells remained, while almost all of amino acids in bleached shells were retained after 240 h
312 of leaching at 140°C (Table 2).

313

314 **4.2 Amino acid composition vs. heating time**

315

316 • **Unbleached**

317

318 The proportion of Asx remained similar over time in the 140°C experiment (Fig. 5A, Table 5
319 Supplementary Information), as did that of Phe, Ile, Leu, and Thr. Only after heating at 140°C
320 for 240 h did the percentages of Glx and Val increase, while that of Arg decreased; this
321 amino acid is very labile. The percentages of Ser and Gly decreased with heating time after 1
322 h, whereas that of Ala increased.

323

324 • **Bleached**

325

326 The variation of the percentages of each amino acid in the IcP fraction with heating time did
327 not precisely reproduce what was observed in unbleached shells (Fig. 5B, Table 6
328 Supplementary Information), although in most cases the pattern was the same, i.e., the

329 percentage of Ala, Glx, and Val increased, while that of Ser decreased. In contrast to
330 unbleached samples, the percentage of Gly increased, while that of Thr decreased.

331

332 **4.3 D/L values vs. heating time**

333

334 The D/L values increased with time in unbleached and bleached shells (Fig. 6). As expected,
335 the highest D/L values for all amino acids were observed after heating for 240 h, thereby
336 indicating that rates of racemisation/epimerisation in *P. lineatus* shells are regulated by
337 temperature and time.

338

- 339 • **Unbleached shells**

340

341 Asx showed the most rapid racemisation rate, followed by Phe, Glx, and Ala (Fig. 6).

342

- 343 • **Bleached shells**

344

345 The IcP fraction in modern *P. lineatus* shells showed higher D/L values than the inter-
346 crystalline one (Fig. 6). In this case, Ala and Phe showed the most rapid racemisation rates,
347 followed by Asx and Glx, although Asx showed the highest D/L values before 8 h.

348

349 **4.4 Discussion**

350

351 The heating experiment at 140°C confirmed the distinct protein composition of the inter- and
352 intra-crystalline fractions, as the percentage of each amino acid in unbleached and bleached
353 samples differed (Figs. 5A, 5B). Moreover, contrary to the unbleached samples, the IcP

354 fraction remained almost constant, indicating that this fraction remained closed over time
355 during isothermal heating. In this regard, according to Demarchi et al. (2013a, p. 154), the
356 total concentration of amino acids in unbleached samples would eventually reach the
357 concentration levels detected in bleached powders, as prolonged leaching would isolate the
358 IcP fraction. However, this was not observed here, i.e. after 240 h heating at 140°C the amino
359 acid concentration in unbleached samples was still 14 times higher than that in the IcP
360 fraction (Table 3). Nevertheless, it is likely that leaching would occur over geological
361 timescales (Miller and Hare, 1980; Collins and Riley, 2000; Bright and Kaufman, 2011); after
362 ca. 6 ka cal B.P., the amino acid concentration in unbleached samples was reduced by around
363 85-90% (see Section 5).

364 The leaching experiment also showed that, upon isothermal heating, 40% of proteins from the
365 unbleached samples were lost (Table 3). However, only a small amount of the total amino
366 acids lost in the unbleached samples were found in the water (4.9%). Therefore, the main loss
367 of amino acids from the inter-crystalline fraction is likely to be due to decomposition (the
368 processes leading to the chemical degradation of the molecular structure of the amino acids),
369 either within the shell or once leached into the water.

370

371 **5 Amino acids in archaeological shells**

372

373 ***5.1 Amino acid composition of the whole shell***

374

375 The total amino acid concentration of unbleached *P. lineatus* shells (representing the amino
376 acids that comprise inter- and intra-crystalline proteins) was higher in modern specimens than

377 in archaeological ones (Fig. 7), and more variable; the total amino acid content decreased by
378 around 85-90% from modern to archaeological shells. However, the concentrations were
379 similar in archaeological *P. lineatus* shells of diverse ages (Neolithic, Mesolithic, and Upper
380 Magdalenian), even in the oldest samples analysed in this study. It is worth noting that [Asx]
381 and [Glx] were higher in modern *P. lineatus* shells, while archaeological samples showed
382 similar concentrations of these amino acids (Asx and Glx).

383 Similar percentages for all amino acids (considering [Asx], [Glx], [Ser], [Ile], [Leu], [Phe],
384 [Val], [Ala], Gly, [Arg] and [Thr]) were obtained in archaeological levels of different ages.
385 However, the percentage of each respective amino acid varied in a different way with respect
386 to that of the modern shells (Fig. 8A). The percentage of Asx increased sharply with age (Fig.
387 8A), i.e. for modern specimens it was around 14%, whereas for the Neolithic ones
388 (Mazaculos II-A2) it was 35%, remaining similar for Mesolithic and Upper Magdalenian
389 ones. The percentage of Glx, Val and Leu also increased from modern to archaeological
390 samples. In this regard, samples older than ca. 6 ka cal BP (Neolithic) showed similar
391 proportions of Asx. In contrast, the percentage of Ser, Ala, Phe, Gly and Arg showed a rapid
392 decrease in *P. lineatus* shells from modern to Neolithic age, after which the percentage
393 remained almost constant. It should be noted that the percentage of Ile and Thr remained
394 almost the same.

395

396 **5.2 Amino acid composition of the IcP fraction**

397

398 In contrast to that observed in unbleached samples, the concentration of amino acids in
399 bleached *P. lineatus* shells (representing the amino acids that comprise only IcPs) was similar

400 for modern and archaeological representatives of distinct ages (Fig. 7). The same results were
401 obtained for [Asx].

402 Similar percentages were obtained for Asx, Glx, Ile, Leu, and Phe in modern and
403 archaeological shells (Table 2). In contrast, the percentage of Ser decreased sharply with age
404 (Fig. 8B), from modern (ca. 15%) to Neolithic (ca. 8%) specimens, remaining similar for
405 Mesolithic and Upper Magdalenian ones. The percentage of Arg and Thr also decreased from
406 modern to archaeological samples. The percentage composition of Ala, Val, and Gly showed
407 a rapid increase from modern to Neolithic *P. lineatus* shells, after which the percentage of
408 these amino acids remained almost constant.

409

410 **5.3 Interpretation of amino acid concentration trends**

411

412 **5.3.1 Whole shell amino acids**

413

414 Significant protein leaching is likely to have occurred from the inter-crystalline fraction
415 during the ca. 6,000 yr cal BP after the death of *P. lineatus*, as the total amino acid content
416 decreased ca. 85-90% over this time and then stabilised. After this decrease, the amino acid
417 content in *P. lineatus* shells of Mesolithic and Upper Magdalenian ages (up to ca. 12.6 ka cal
418 BP) remained almost the same (Fig. 4), whereas the contribution of each amino acid to the
419 total content differed (Fig. 8A, B). Thus, there was an increase in the relative proportion of
420 Asx, Glx, Val, and Leu with age, while the relative composition of other amino acids such as
421 Ser, Ala, Phe, Gly and [Arg] decreased with age.

422 Of note is the decrease in the percentage of Ala with age in the whole shell (Fig. 8A), as an
423 increase in the relative concentration of this amino acid is commonly observed upon artificial
424 diagenesis in molluscs (Penkman et al., 2008; Demarchi et al., 2011, 2013b) and eggshells
425 (Miller et al., 2000; Crisp, 2013). In contrast, the percentage of Ala increased in the IcP
426 fraction of *P. lineatus* (Fig. 8B). This increase is assumed to be caused by the decomposition
427 of other amino acids, such as Asx and mainly Ser, into Ala (Bada and Miller, 1970; Bada et
428 al., 1978; Bada and Man, 1980; Walton, 1998).

429 We also observed a decrease in [Ser]/[Ala] values in unbleached *P. lineatus* samples, thereby
430 indicating a general pattern of increased protein degradation with age, as also interpreted
431 from the increase in D/L values (Fig. 9A) and a decrease in amino acid concentrations with
432 age (Fig. 7).

433 Therefore, the different behaviour of Ala in unbleached *P. lineatus* shells may be explained
434 by the loss of free Ala from the inter-crystalline matrix. Also, a different decomposition rate
435 of amino acids in this species compared to other taxa cannot be ruled out. In fact, Ala
436 accounted for ca. 25% of the total amino acid content in unbleached *P. lineatus*, whereas in
437 other molluscs and eggshells it represents less than 15% (Penkman et al., 2008; Demarchi et
438 al., 2011, 2013b, Crisp, 2013), thus indicating differences in protein composition. Moreover,
439 the total amino acid content reduced considerably from modern to archaeological shells (ca.
440 85-90%), thereby revealing a considerable loss of amino acids. In this regard, the biomineral
441 might also play an important role due to differential mineral binding of amino acids
442 (Demarchi et al., 2016).

443

444 **5.3.2 IcP amino acids**

445

446 The IcPs accounted for around 4% of the total protein content of modern shells (Fig. 4). This
447 percentage increased sharply with age (up to 60–70% over 6 ka), with apparently limited
448 degradation of the proteins in this fraction (the concentration of amino acids remained
449 constant with age in bleached samples), thereby indicating that there was an important
450 preferential break-down and loss of inter-crystalline proteins. Similarly, [Asx] remained
451 constant with age.

452 In contrast, the percentage of the distinct amino acids in bleached samples did not vary with
453 age in a similar way to unbleached samples. This observation confirms that the composition
454 of the intra- and inter-crystalline fractions differed.

455 Moreover, the amino acid percentages in unbleached samples showed a different pattern after
456 leaching (Fig. 5A) and in archaeological sites (Fig. 8A). These observations can be attributed
457 to the IcP fraction becoming more representative in archaeological *P. lineatus* shells (60-70%
458 with respect to the total proteins) after 6 ka due to amino acid loss from the inter-crystalline
459 fraction by leaching and decomposition. This finding coincides with reports by Penkman et
460 al. (2008), who observed that the proportion of intra-crystalline amino acids within the whole
461 shell increases as the sample ages.

462 The differences found in the concentration and composition of amino acids and D/L values
463 between the inter- and intra-crystalline proteins are in agreement with Sykes et al. (1995) and
464 Penkman et al. (2007, 2008), who observed distinct racemisation rates in these fractions in a
465 variety of mollusc shells. In leaching experiments (140°C for 24 h to 240 h) on unbleached
466 and bleached *B. tentaculata* and *P. vulgata* shells, Penkman et al. (2008) and Demarchi et al.
467 (2013a) reported that only a small percentage (1–4%) of the total amino acid content leached

468 from the IcP fraction, in contrast to a higher percentage (ca. 40%) from unbleached shells
469 under the same conditions. While inter-crystalline proteins are more susceptible to
470 decomposition or leaching, the IcP fraction has been found to behave like a closed system in
471 various mollusc shells, including those of *P. lineatus*. Similarly, Bosch et al. (2015)
472 concluded that the IcP fraction of the topshell *P. turbinatus* approximates a closed-system.
473 The results observed in unbleached and bleached archaeological *P. lineatus* shells confirmed
474 that the inter- and intra-crystalline fractions of this species differ in protein profiles, thus
475 showing distinct racemisation rates and compositions. IcPs seemed to remain in a closed
476 system, as the total concentration of amino acids remained similar with age, although
477 percentages of some amino acids varied. It should be highlighted that the concentration and
478 percentage of Asx (which is the amino acid commonly used for dating recent samples)
479 remained constant with age in bleached samples.

480

481 **6 Aminochronology**

482

483 To establish the aminochronology of the archaeological samples here, we used only Asx,
484 because it is the amino acid that shows the fastest racemisation. Furthermore, due to their low
485 D/L values, other amino acids were not suitable to discriminate between archaeological sites
486 of these ages.

487

488 **6.1 Asx D/L values vs. age -unbleached**

489

490 In general, topshells from archaeological sites showed Asx D/L values consistent with their
491 age (Table 4; Fig. 9A), i.e. in the Neolithic site (Mazaculos II-A2) shells had the lowest Asx
492 D/L values, followed by those belonging to the Mesolithic (shell midden and level 29 of La
493 Riera, level 1.3 of Mazaculos II, El Penical, Bricia-A, La Trecha, Arenillas, El Mazo and
494 Toral), Azilian/Magdalenian (level 27 of La Riera), and Magdalenian (level 24 of La Riera,
495 Bricia-C) periods. To select the best fit for the amino-age estimation algorithm, we compared
496 the correlation coefficients (r^2) for various approaches. We used the relationship between D/L
497 Asx values vs. age because it provided the highest correlation coefficient.

498

499 **6.2 Asx D/L values vs. age -bleached**

500

501 As with the Asx D/L values of unbleached samples, D/Ls also increased with age in the
502 bleached fraction (Table 4; Fig. 9B). Asx D/L values in the bleached samples showed a
503 strong correspondence ($r^2 = 0.88$) with age.

504 Of note, the Asx D/L values were similar in unbleached and bleached samples, although in
505 unheated modern specimens they were lower in the former (Table 4).

506

507 **6.3 Aminochronological considerations**

508

509 The mean Asx D/L values of 106 bleached and unbleached *P. lineatus* shells from the
510 archaeological levels (after the rejection of samples with abnormally high D/L values)
511 increased with age (Figs. 9A and 9B). Asx D/L values were similar in unbleached and
512 bleached samples. This observation could be attributable to the considerable loss of amino

513 acids in the inter-crystalline fraction over time, thereby producing a significant contribution
514 of the IcP fraction to the whole protein content. Thus, good correlations were obtained for
515 Asx D/L values in the inter- and intra-crystalline fractions *versus* age.

516 However, the extent of racemisation of some amino acids (Asx, Glx, Ala, Val) within the IcP
517 fraction of *P. turbinatus* specimens from Ksâr 'Akil (Lebanon) revealed intralayer variability
518 of the D/L values comparable with intra-horizon variability. Therefore D/L values could not
519 be used to resolve the chronology within the site at the timescale relevant in that study,
520 between 30 and 43 ka BP (Bosch et al., 2015). The low coefficients of variation for Asx D/L
521 values in both bleached and unbleached *P. lineatus* samples (> 7% in most of cases) can
522 explain the good resolution observed for discriminating the chronology of archaeological
523 sites.

524 A general increase in Asx D/L values with radiocarbon age was observed (Figs. 9A and 9B)
525 up to 13 ka cal BP. We propose that the palaeoclimatic variations that occurred after the
526 accumulation of the archaeological remains did not significantly affect the amino acid
527 racemisation rate of *P. lineatus* shells, in contrast to that observed from limpet shells of the
528 Solutrean and Gravettian periods (Ortiz et al., 2015). The sites studied here were formed after
529 the Last Glacial Maximum (LGM), when climate amelioration occurred and was maintained
530 throughout the Holocene with no significant temperature variations during the last 13 ka cal
531 BP, with the exception of Younger Dryas (Bard, 2002; Peck et al., 2008).

532 This study indicates that Asx D/L values of unbleached and bleached *P. lineatus* shells are
533 useful for dating *P. lineatus* shells from archaeological levels in this region (Figs. 9A and
534 9B).

535

536 **7. Conclusions**

537

538 In summary:

539 1.-The protein composition of the inter- and intra-crystalline fractions of the topshell *P.*
540 *lineatus* differs, as shown by differences in the percentages of amino acids present. IcP amino
541 acids accounted for 4% of the modern total amino acid content, but this percentage increased
542 to 60-70% after ca. 6 ka BP.

543 2.-The percentage of Asx remained constant with age (in archaeological samples over ca. 13
544 ka cal BP) within IcPs. However, in inter-crystalline proteins, the percentage of this amino
545 acid increased sharply in the first ca. 6 ka after the death of *P. lineatus* and then stabilised.
546 The relative composition of other amino acids in the inter-crystalline fraction decreased with
547 age (Ser, Ala, Phe, Gly, and Arg), whereas the percentage of Glx, Val and Leu increased. In
548 contrast, within IcPs, Ala and Gly increased. The different protein composition of the inter-
549 and intra-crystalline fractions, the closed system behaviour of the IcP fraction, and the
550 differential mineral binding of amino acids may explain these differences.

551 3.-The IcP fraction behaved like a closed system, as the concentration of amino acids did not
552 vary significantly after heating at 140°C. The main loss of inter-crystalline proteins occurred
553 through decomposition, and only a small fraction was leached into water (ca. 4.9%).

554 4.-The main leaching of inter-crystalline proteins in *P. lineatus* shells occurred within at least
555 the first 6,000 yr cal BP after the death of the organism. This is evidenced by the considerable
556 decrease (85-90%) in the total amino acid content in archaeological samples with respect to
557 modern representatives. However, the total amount of amino acids in the IcP fraction

558 remained virtually intact for at least 12.6 ka, thereby confirming that this fraction
559 approximates a closed system.

560 5.-Differences in the amino acid content of inter- and intra-crystalline proteins, which
561 undergo racemisation at different rates, may be produced because the products of diagenesis
562 are likely to remain in the IcP fraction. Likewise, the preferential removal of certain proteins
563 and amino acids from the inter-crystalline matrix over time might cause this fraction to
564 degrade faster than the intra-crystalline one. Although Asx D/L values were higher in
565 unbleached samples, there was good correspondence between the Asx D/L values in inter-
566 and intra-crystalline proteins. However, other amino acids, such as Glx, showed lower levels
567 of racemisation in the inter-crystalline proteins. We consider that the age of archaeological
568 levels can be established through analysing unbleached samples; however, bleaching
569 provides important information and complements the interpretation of the results obtained
570 from the inter-crystalline fraction.

571 6.-Asx D/L values in unbleached and bleached *P. lineatus* shells were comparable and
572 showed a good correspondence with age. They can both therefore be used for the age
573 calculation of archaeological localities.

574 7.-In brief, AAR is a satisfactory tool for dating *P. lineatus* from archaeological sites
575 covering the Upper Magdalenian to Neolithic periods, i.e. from ca. 13 to 5.5 ka cal BP.

576

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578

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589

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870 **Figure captions**

871

872 Figure 1. Geographical location of the caves studied. 1-La Trecha, 2-Arenillas, 3-Mazaculos
873 II, 4-El Mazo, 5-El Toral III, 6-La Riera, 7-Bricia, and 8-El Penicial. Cue beach is also
874 plotted.

875

876 Figure 2. A) Photograph of an archaeological *P. lineatus* shell from level 108 of El Mazo. B)
877 Cross-section of a shell showing the different layers, and the sampling area.

878

879 Figure 3. A) Concentration (nmol/mg) of Asx, Glx, Gly, Ala, Ser, Val and Ile in unbleached
880 and bleached samples of modern *P. lineatus* shells (errors are shown in Table 1
881 Supplementary Information). B) D/L values of Ala, Ile, Asx, Glx, and Ser in unbleached and
882 bleached samples of modern *P. lineatus* shells (errors are shown in Table 2 Supplementary
883 Information). C) Relative amino acid composition of unbleached and bleached *P. lineatus*
884 shells (errors are shown in Table 3 Supplementary Information).

885

886 Figure 4. Variation of total amino acid concentration (nmol/mg) in unbleached and bleached
887 samples of modern *P. lineatus* shells in response to heating at 140°C.

888

889 Figure 5. Percentage of each amino acid in unbleached (A) and bleached (B) samples of *P.*
890 *lineatus* shells after heating at 140°C (Tables 5 and 6 Supplementary Information).

891

892 Figure 6. Asx, Glx, Ala and Phe D/L values in unbleached and bleached samples of *P.*
893 *lineatus* shells heated at 140°C versus heating time (h).

894

895 Figure 7. A) Total amino acid concentration of the unbleached and bleached modern and
896 fossil *P. lineatus* shells.

897

898 Figure 8. Percentage of each amino acid in unbleached (A) and bleached (B) samples of *P.*
899 *lineatus* shells from modern and archaeological sites (Tables 7 and 8 Supplementary
900 Information). The same colour code was used for all the levels of the same period, and
901 localities are plotted in the age order indicated in the legend.

902

903 Figure 9. Best-fit relation between Asx D/L values obtained in (A) unbleached and (B)
904 bleached samples of *P. lineatus* shells versus age.

905

906 **Tables**

907 **Table 1. Archaeological levels studied and the periods assigned. Calibrated ages (yr cal)**
 908 **were converted using the Radiocarbon Calibration Program 7.1 (CALIB 7.1) (Stuiver et**
 909 **al., 2017) with the calibration dataset IntCal13 (Reimer et al., 2013). Reservoir effect**
 910 **was corrected using data from Monge Soares et al. (2016).**

Cave	Archaeological level	Age (¹⁴ C yr cal BP)
Cue beach	-	Modern
Arenillas (ARE)	Shell midden	Mesolithic [1] 7,975±23 (OxA-X-2488) 8,227±58 (OxA-27154)
Bricia (BRI)	Shell midden (Level A) Level C	Mesolithic [2,3] 7,680±150 (GaK 2908) Upper Magdalenian [2] aar 13,934 ± 1,949 [5]
Mazaculos II (MAZ)	Level A2 Shell midden Level 1.3	Neolithic [4] 5798±121 [4] Mesolithic [4] 8,490±40 (UGAM-9081) 8,529±49 (OxA-26953)
La Riera (RIE)	Shell midden Level 29 Level 27upper Level 24	Mesolithic [6] 7,375±185 (GaK-3046) Mesolithic [6] 9,722±379 (GaK-2909) Azilian/Magdalenian[6] 12,510±195 (BM-1494); Upper Magdalenian [6] 12,660±545 (GaK-6982)
El Mazo (EMA)	101 113 120 105 108	Mesolithic [1] 7,927±42 (OxA-30780) 8,112±52 (OxA-30806) Mesolithic [1] 8,032±43 (OxA-28403) 8,385±18 (OxA-28404) Mesolithic [1] 8,255±50 (OxA-28405) 8,436±38 (OxA-30976) Mesolithic [1] 8,209±86 (OxA-30535) 8,402±19 (OxA-30977) Mesolithic [1] 8,899±91 (OxA-28411) 9,193±63 (OxA-26954)
El Penical (PEN)	Surface shell midden	Mesolithic [2,7] 9,760±250 (GaK 2906)
La Trecha (LTR)	Level 1	Mesolithic [8] 8,303±72(URU0083)
El Toral (TOR)	Level 17 Level 21 Level 13C	Mesolithic [9] 7370±40(UGAMS 5403) Mesolithic [9] 7510±40(UGAMS 5400) 7620±30(UGAMS 5401) Mesolithic [9] 9530±20(UGAMS 5404)

911 1.-Monge Soares et al. (2016); 2.-Clark (1976); 3.-Jordá (1957, 1958); 4.-González-Morales (1982); 5.-
912 Ortiz et al. (2009); 6.-Straus et al. (1978); Straus and Clark (1986); 7.-Vega del Sella (1914); 8.-González-
913 Morales et al. (2002), 9.-Rigaud and Gutiérrez-Zugasti (2016).

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917 **Table 2. Percentage of Asx and Glx content with respect to the total amino acid content**
 918 **of unbleached and bleached samples of modern and archaeological *P. lineatus* shells.**

Period	Localities	N	%Asx	%Asx	%Glx	%Glx
			unbleached	bleached	unbleached	bleached
	Modern	5	14.3 ± 0.4	40.4 ± 2.1	7.5 ± 0.8	8.4 ± 1.0
N	MAZ-A2	5	31.6 ± 1.9	37.6 ± 0.9	8.6 ± 0.5	9.1 ± 0.8
M	TOR-17	5	31.0 ± 2.7	35.2 ± 2.5	8.9 ± 0.7	9.2 ± 0.5
	RIE-SM	5	32.3 ± 2.2	37.1 ± 1.9	9.8 ± 3.8	8.6 ± 0.5
	TOR-21	5	32.4 ± 3.2	35.1 ± 2.4	9.8 ± 0.4	9.7 ± 1.6
	BRI-A	5	30.7 ± 4.9	38.5 ± 0.4	8.1 ± 0.2	9.1 ± 0.5
	ARE	5	30.0 ± 0.2	36.6 ± 1.0	8.3 ± 0.2	8.9 ± 0.5
	EMA-101	5	31.2 ± 1.6	35.6 ± 0.5	9.7 ± 1.6	9.0 ± 0.8
	EMA-113	5	30.1 ± 1.7	35.2 ± 1.1	8.5 ± 0.4	8.8 ± 0.3
	LTR	5	29.6 ± 4.4	37.1 ± 1.4	9.2 ± 0.7	9.6 ± 0.7
	EMA-105	5	30.1 ± 2.1	36.5 ± 0.9	9.0 ± 0.9	8.9 ± 0.1
	EMA-120	5	32.4 ± 1.7	35.1 ± 1.6	8.3 ± 0.4	8.8 ± 0.2
	MAZ-1.3	5	32.0 ± 0.2	37.3 ± 0.8	8.5 ± 0.2	9.0 ± 0.2
	EMA-108	5	31.8 ± 2.8	35.2 ± 3.6	8.5 ± 1.4	9.4 ± 0.9
	TOR-13C	5	31.3 ± 1.5	37.5 ± 1.1	9.6 ± 0.4	8.9 ± 0.3
	RIE-29	5	30.5 ± 5.2	36.5 ± 1.1	8.3 ± 0.3	8.4 ± 0.3
	PEN	8	30.3 ± 0.7	37.2 ± 0.8	8.4 ± 0.2	9.7 ± 0.6
AZ/UM	RIE-27	5	28.9 ± 1.9	35.4 ± 1.5	8.5 ± 0.3	8.7 ± 0.2
UM	RIE-24	5	30.3 ± 1.4	35.3 ± 1.5	8.6 ± 1.8	9.4 ± 1.0
	BRI-C	8	32.5 ± 0.3	36.9 ± 1.1	8.5 ± 0.3	9.5 ± 0.9

919 N:Neolithic; M: Mesolithic (Asturian); Az: Azilian; UM: Upper Magdalenian.

920

921

922 **Table 3. Loss of amino acids from bulk unbleached and bleached *P. lineatus* powders**
 923 **(experimental samples) after 24h of heating at 140°C (n= number of samples). Total**
 924 **concentrations (nmol/mg) were calculated using [Asx], [Glx], [Ser], [Ala], [Val], [Ile],**
 925 **[Leu], [Phe], [Gly], [Arg] and [Thr].**

926

Loss of amino acids after 24h heating at 140°C	Unbleached	Bleached
Initial [total] concentration in shell unheated (pmol/mg) (n=5)	385.733	16.130
[total] THAA after heating (pmol/mg) (n=2) for 24h	231.393	16.092
[total] THAA in water, heated (pmol/mg equiv.) (n=2)	18.903	-
Overall loss in shell (%) from the original	40.0	0.23
Loss into water by leaching (%) from the original	4.9	-
Loss by decomposition (%) from the original	35.1	-

927

928 **Table 4. Asx D/L values in unbleached and bleached samples of modern and**
 929 **archaeological *P. lineatus* shells.**

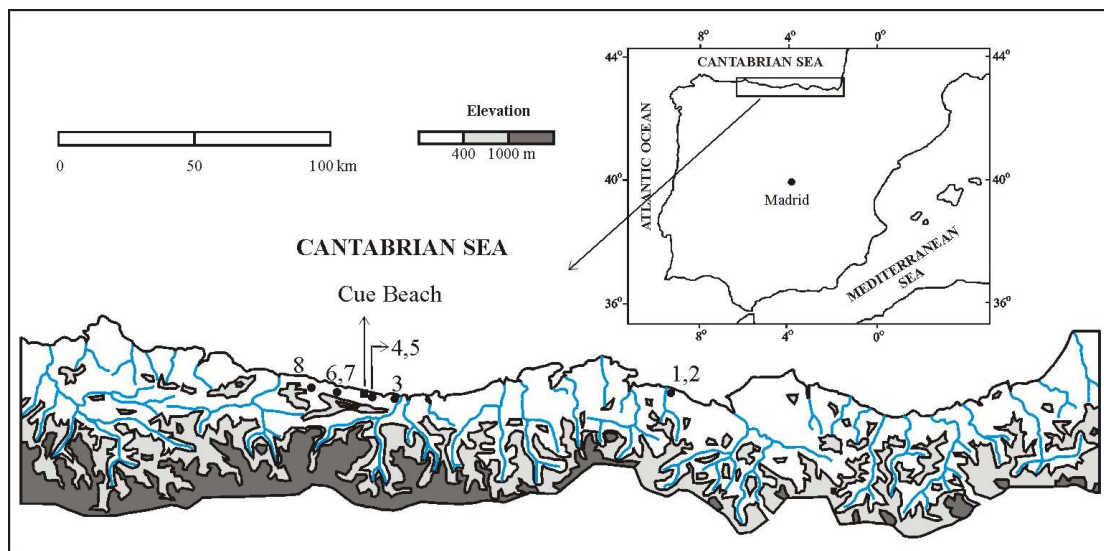
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Period	Localities	D/L Asx unbleached	D/L Asx bleached
	Cue beach	0.048±0.001	0.084±0.004
N	MAZ-A2	0.177±0.022	0.180±0.012
M	TOR-17	0.225±0.011	0.208±0.009
	RIE-SM	0.223±0.021	0.228±0.015
	TOR-21	0.219±0.019	0.199±0.009
	BRI-A	0.237±0.018	0.240±0.023
	ARE	0.224±0.011	0.208±0.019
	EMA-101	0.235±0.009	0.227±0.033
	EMA-113	0.227±0.009	0.197±0.010
	LTR	0.235±0.013	0.235±0.011
	EMA-105	0.219±0.016	0.207±0.011
	EMA-120	0.228±0.008	0.199±0.006
	MAZ-1.3	0.218±0.021	0.201±0.007
	EMA-108	0.233±0.010	0.216±0.020
	TOR-13C	0.236±0.017	0.239±0.001
	RIE-29	0.251±0.020	0.257±0.017
	PEN	0.244±0.019	0.246±0.018
AZ/UM	RIE-27	0.268±0.013	0.268±0.028
UM	RIE-24	0.285±0.002	0.284±0.017
	BRI-C	0.299±0.030	0.294±0.027

931 N:Neolithic; M: Mesolithic (Asturian); Az: Azilian; UM: Upper Magdalenian.

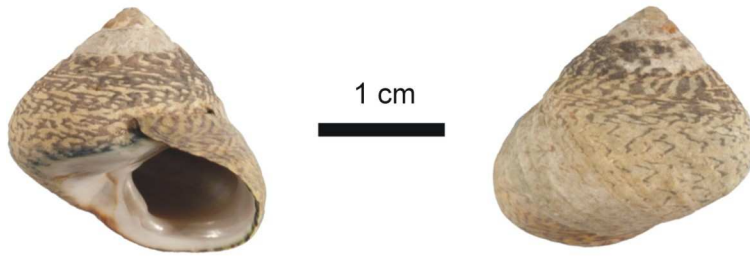
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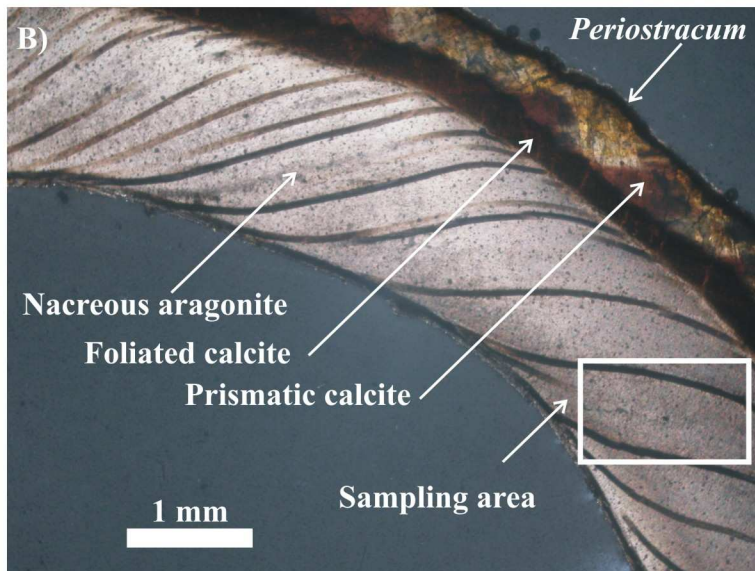


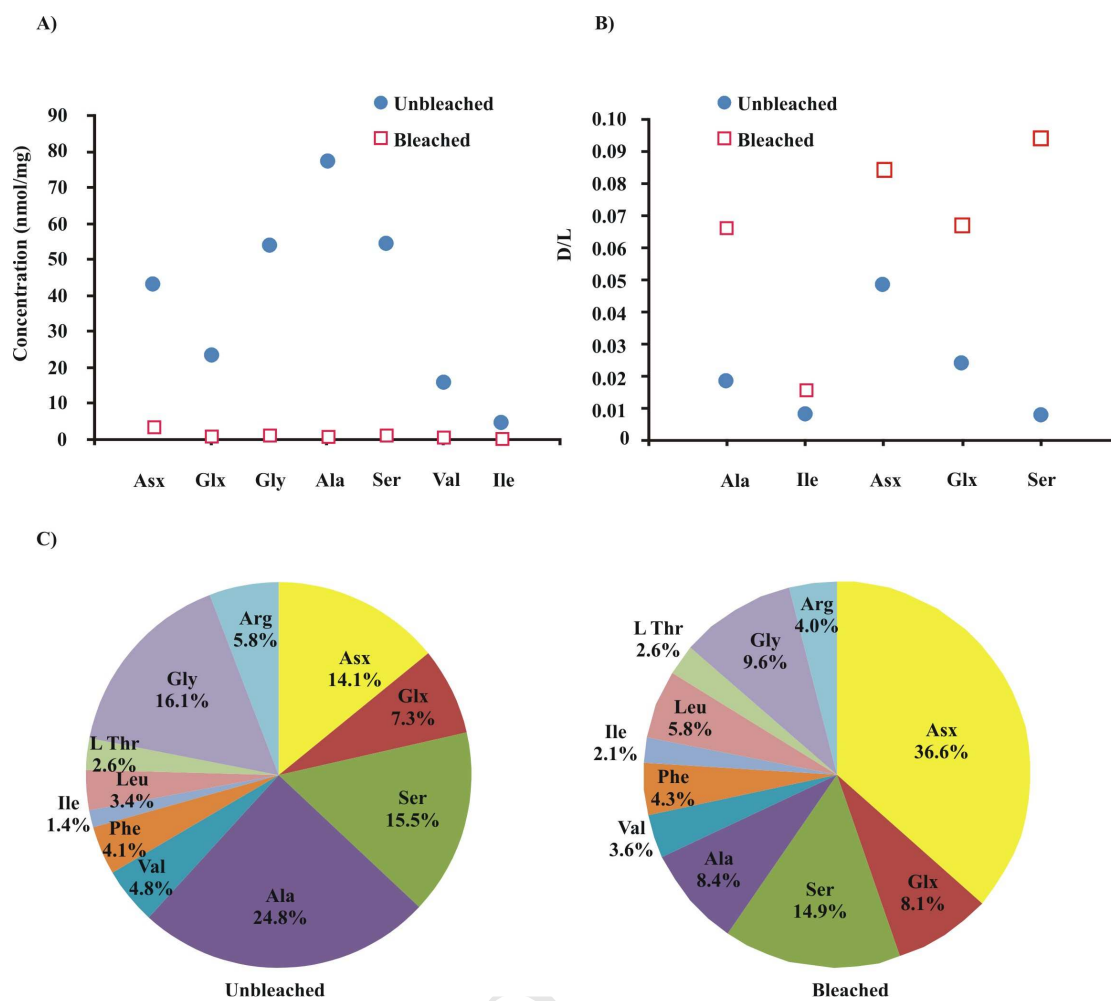
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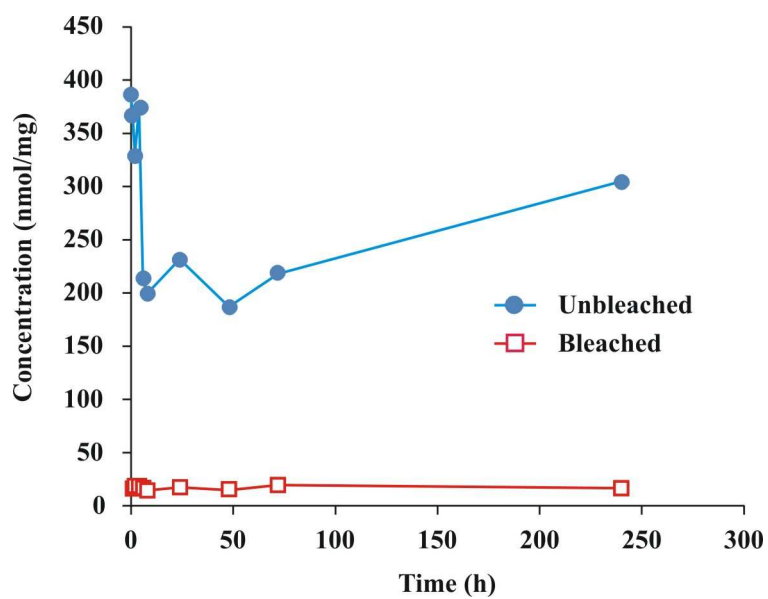
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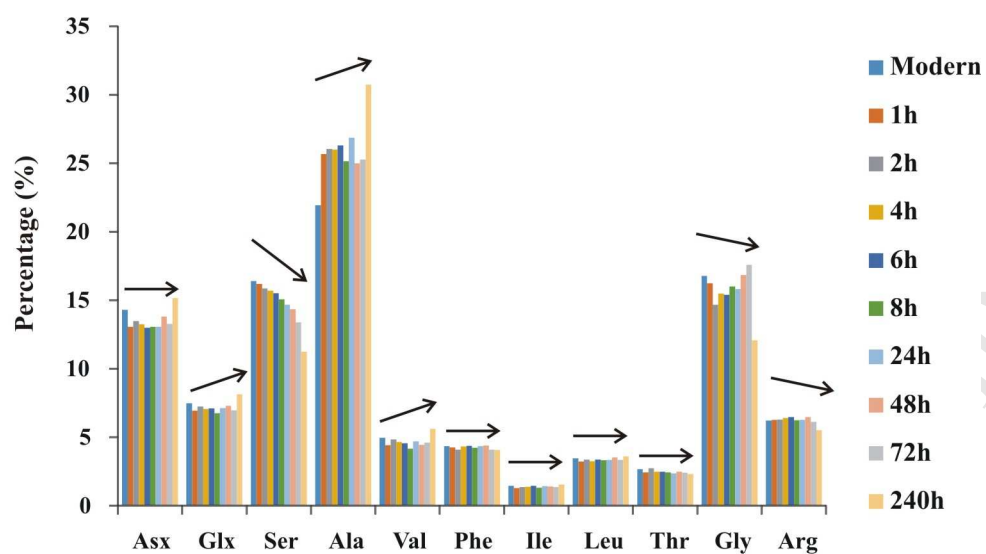
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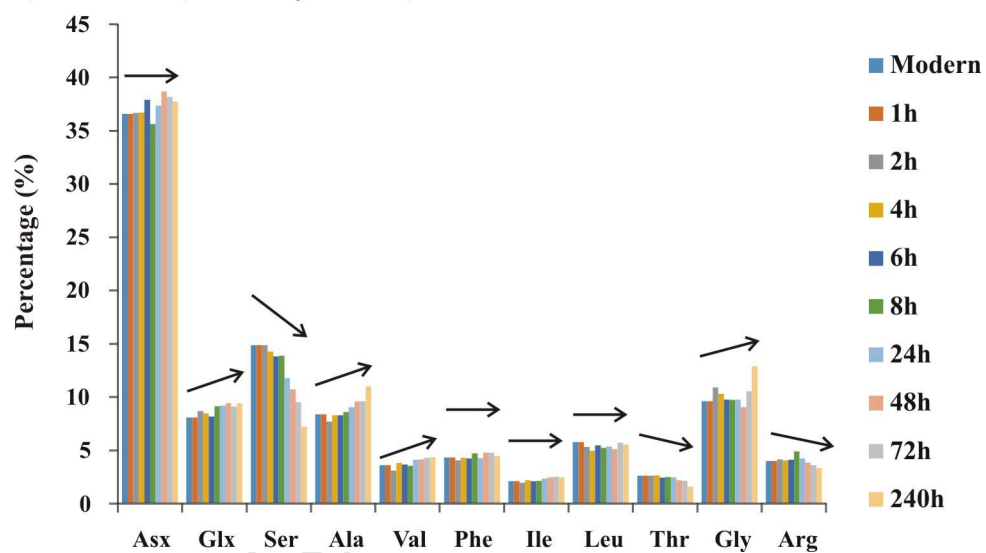


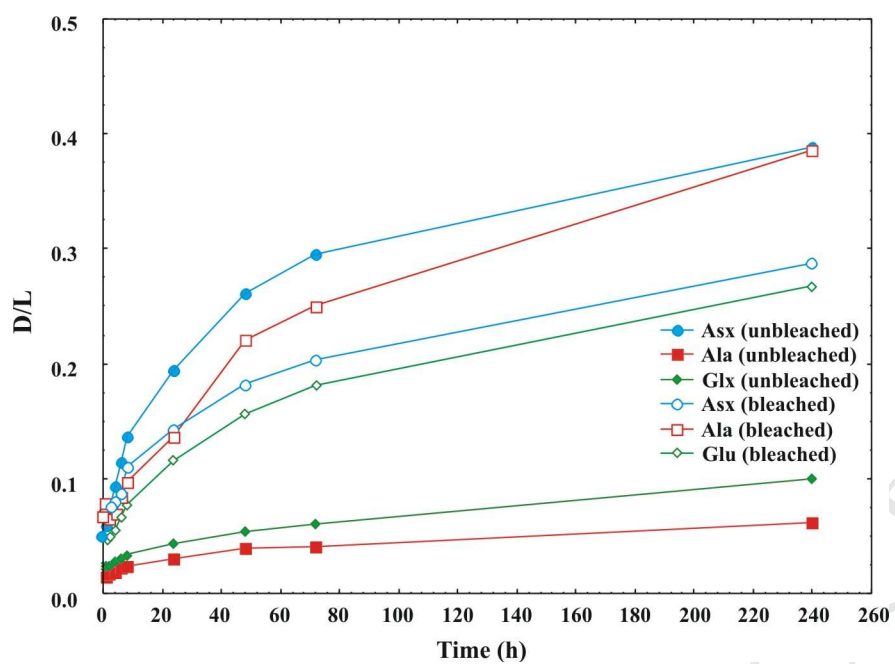


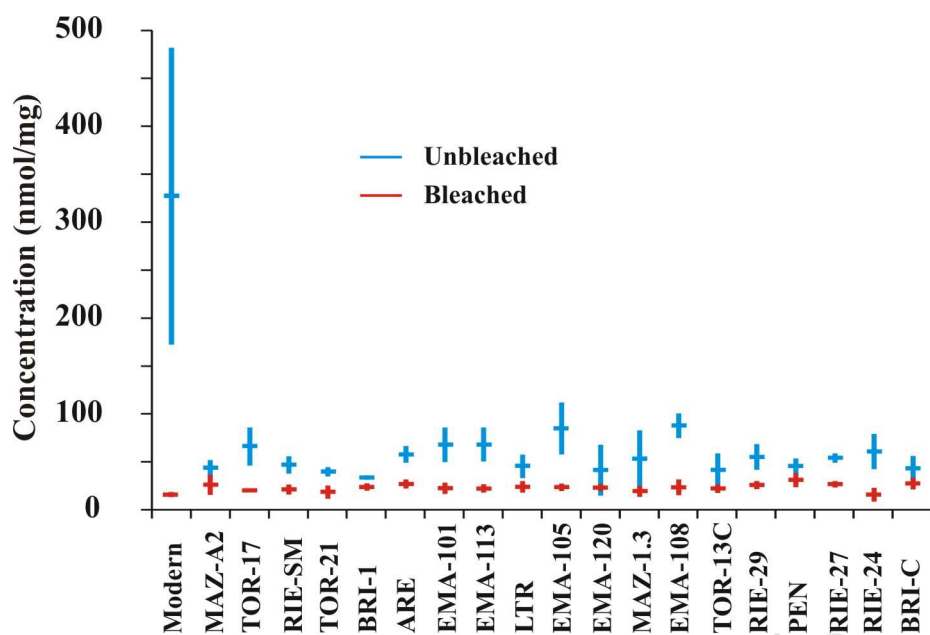
A) unbleached (whole shell)



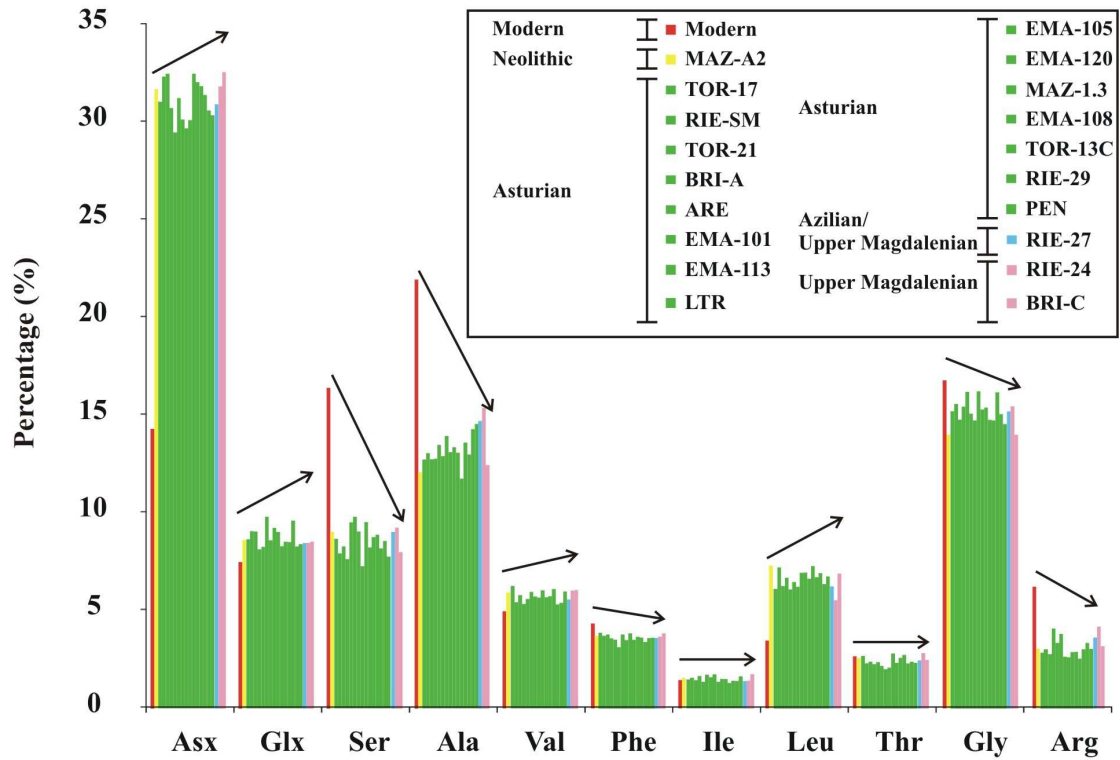
B) bleached (intra-crystalline)



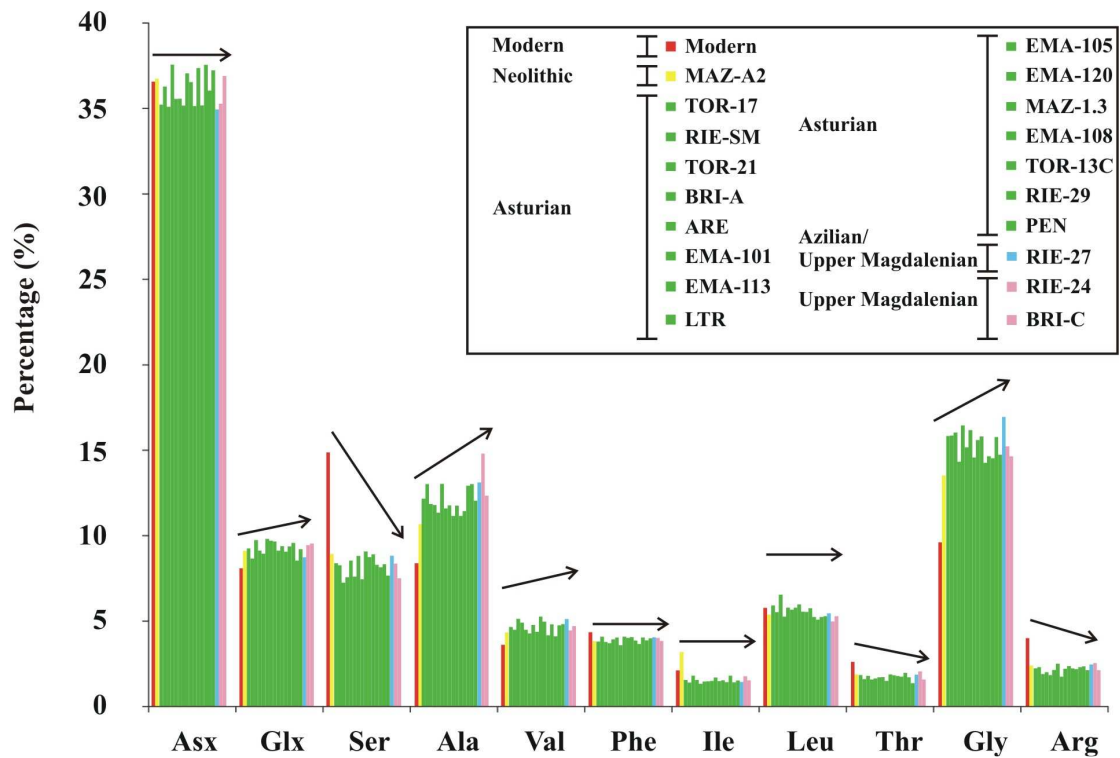


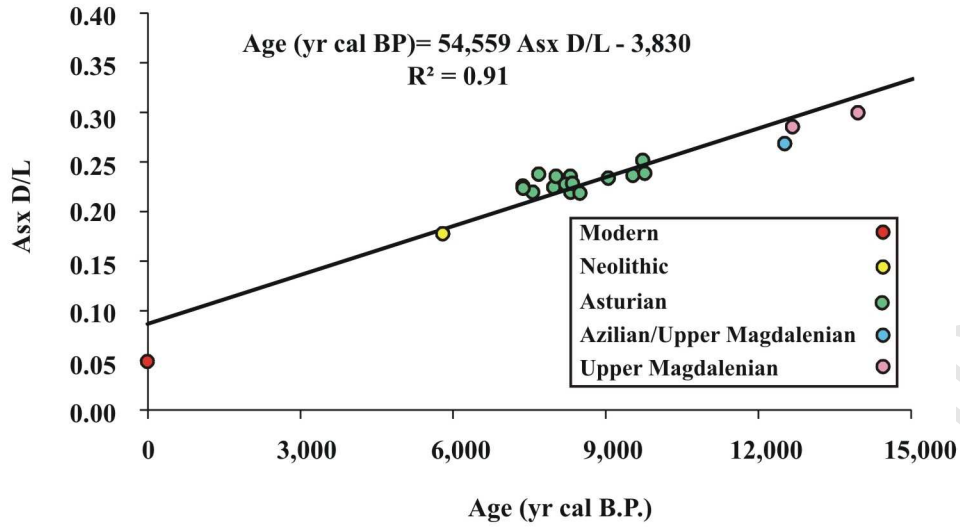


A) unbleached (whole shell)



B) bleached (intra-crystalline)



A) unbleached (whole shell)**B) bleached (intra-crystalline)**