From THE DEPARTMENT OF MEDICINE, SOLNA Karolinska Institutet, Stockholm, Sweden

PREDICTIVE BIOMARKERS IN RHEUMATOID ARTHRITIS

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Predictive biomarkers in rheumatoid arthritis

THESIS FOR DOCTORAL DEGREE (Ph.D)

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The public defence will take place on the 24th of August, 2018, at 9⁰⁰ am, at the Centre for Molecular Medicine (CMM) Lecture hall, L8:00, Karolinska University Hospital, Solna.

Those who have knowledge, don't predict. Those who predict, don't have knowledge

Lao Tzu 6th century BC, Chinese poet

ABSTRACT

Being a very heterogeneous disease, rheumatoid arthritis (RA) is challenging for treatment. On a group level, some therapy options might be superior compared with others, however, this does not mutually exclude the less effective option to be more suitable for some patients, if the superior therapy option does not help. In different patients the concentration of biomarkers may vary dramatically, however, translation of meaning of these variations for each patient is not feasible yet. Observations and comparisons of these biomarkers before start of therapy between patient groups with different outcome after therapy can help to understand their role and make individualised approach for the therapy choice.

Low or moderate levels of a multi-biomarker disease activity (MBDA) score measured at baseline or follow-up visits could identify RA patients at very low risk of radiographic progression (RP). Moreover, in patients with high MBDA score at the start of treatment escalation, those on infliximab+methotrexate (IFX+MTX) therapy had significantly less RP than patients on non-biological triple therapy (TT) (papers I and II).

In treatment-naive, early RA, patients who failed respond to MTX were randomized to IFX+MTX or non-biological TT. The categories of the MBDA score at the time of randomisation were differentially associated with treatment outcome after 1 year for these two therapies. Patients with low MBDA score benefited more from TT, while those with high MBDA score responded better to IFX (paper III).

Furthermore, when the 12 component biomarkers of the MBDA score were analysed at baseline, four of those (paper IV) as well as two of 177 proteins retrieved from an affinity proteomic study (paper V) were associated with treatment outcome: low disease activity (LDA) and EULAR good response after 3 months of MTX therapy. Combination of these biomarkers within each study also showed improved prediction of treatment outcome.

In patients failing on MTX monotherapy who were randomised to addition of biological TNF inhibitor IFX, very low serum IFX (sIFX) level and anti-drug antibody (ADA)-positivity were associated with poorer outcome. Among baseline parameters, female gender predicted very low sIFX level and ADA-positivity at follow-ups, with similar trend for RF-positivity (paper VI).

In summary, using combination of serum biomarkers helps to predict and identify preferential therapy option for subsets of patients. Further studies of these biomarkers, if validated, will facilitate personalised therapy approach for subsets of patients.

LIST OF SCIENTIFIC PAPERS

- Hambardzumyan K, Bolce R, Saevarsdottir S, Cruickshank SE, Sasso EH, Chernoff D, Forslind K, Petersson IF, Geborek P, van Vollenhoven RF. Pretreatment multi-biomarker disease activity score and radiographic progression in early RA: results from the SWEFOT trial. Annals of the rheumatic diseases. 2015 Jun;74(6):1102-9.
- II. Hambardzumyan K, Bolce RJ, Saevarsdottir S, Forslind K, Wallman JK, Cruickshank SE, Sasso EH, Chernoff D, van Vollenhoven RF. Association of a multibiomarker disease activity score at multiple time-points with radiographic progression in rheumatoid arthritis: results from the SWEFOT trial. RMD open. 2016;2(1):e000197.
- III. Hambardzumyan K, Saevarsdottir S, Forslind K, Petersson IF, Wallman JK, Ernestam S, Bolce RJ, van Vollenhoven RF. A Multi-Biomarker Disease Activity Score and the Choice of Second-Line Therapy in Early Rheumatoid Arthritis After Methotrexate Failure. Arthritis & rheumatology (Hoboken, NJ). 2017 May;69(5):953-63.
- IV. Hambardzumyan K, Bolce RJ, Wallman JK, van Vollenhoven RF, Saevarsdottir S.Serum biomarkers for prediction of response to methotrexate monotherapy in early rheumatoid arthritis: results from the SWEFOT trial. Manuscript
- V. Hambardzumyan K, Hamsten C, Idborg H, Lourido L, Saevarsdottir S, Nilsson P, van Vollenhoven RF, Jakobsson P-J. Association of serum protein levels at baseline with response to methotrexate at 3 months in patients with early rheumatoid arthritis. Manuscript
- VI. Hambardzumyan K, Hermanrud C, Marits P, Vivar N, Ernestam S, Wallman JK, van Vollenhoven RF, Fogdell-Hahn A, Saevarsdottir S. Association of female gender and positive RF with low serum infliximab and anti-drug antibodies, which relate to treatment failure in early RA. Manuscript

LIST OF ARTICLES NOT INCLUDED IN THE THESIS

- I. van Vollenhoven RF, Bolce RJ, Hambardzumyan K, Saevarsdottir S, Forslind K, Petersson IF, Sasso EH, Hwang CC, Segurado OG, Geborek P. Brief Report: Enhancement of Patient Recruitment in Rheumatoid Arthritis Clinical Trials Using a Multi-Biomarker Disease Activity Score as an Inclusion Criterion. Arthritis & rheumatology (Hoboken, NJ). 2015 Nov;67(11):2855-60
- II. Levitsky A, Brismar K, Hafstrom I, Hambardzumyan K, Lourdudoss C, van Vollenhoven RF, Saevarsdottir S. Obesity is a strong predictor of worse clinical outcomes and treatment responses in early rheumatoid arthritis: results from the SWEFOT trial. RMD open. 2017;3(2):e000458.

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LIST OF ABBREVIATIONS

- ACPA anti citrullinated protein antibodies
- ACR American College of Rheumatology
- ADA anti-drug andtibodies
- BMI body-mass index
- CCP cyclic citrullinated peptide
- CD cluster of differentiation
- CDAI clinical disease activity index
- CIA collagen-induced arthritis
- CNS central nervous system
- COMP cartilage oligomeric matrix protein
- CRP C-reactive protein
- CTLA-4 cytotoxic T-lymphocyte-associated protein 4
- CVD cardiovascular diseases
- DAS28 disease activity score based on 28 joints
- DMARD disease modifying anti-rheumatic drug
- EGF epidermal growth factor
- ELISA enzyme-linked immunosorbent assay
- ESR erythrocyte sedimentation rate
- ETN etanercept
- EULAR European League Against Rheumatism
- FGA alpha-chain of fibrinogen
- FOI fluorecent optical imaging
- GM-CSF granulocyte-macrophage colony-stimulating factor
- HAQ health assessment questioneare
- HCQ hydroxychloroquine
- IFX infliximab
- sIFX serum infliximab
- Ig immunoglobulin
- IL interleukin

- JAK Janus kinase
- JSN joint space narrowing
- JSW joint space width

LDA - low disease activity

- LOCF last observation carried forward
- MBDA multi-biomarker disease activity
- M-CSF macrophage colony-stimulating factor
- MFI median fluorecence intencity
- MMP matrix metalloproteinase
- MRI magnetic resonance imaging
- MTX methotrexate
- PatG patient global assessment of disease activity
- PhysG physician global assessment of disease activity
- RA rheumatoid arthritis
- RANKL receptor activator of nuclear factor kappa-B ligand
- RF rheumatoid factor
- ROC receiver operating characteristic
- ROS reactive oxygen species
- RP radiographic progression
- SAA serum amyloid A
- SDAI simplified disease activity index
- SHS modified van der Heijde score
- SJC swollen joint count
- SSZ sulfasalazine
- STAT signal transducer and activator of transcription proteins
- TCZ tocilizumab
- TJC tender joint count
- TNF tumour necrosis factor
- TNF-R tumour necrosis factor receptor
- TT triple therapy

VAS - visual analogue scale

VCAM-1 – vascular cell adhesion molecule

VEGF - vascular endothelial growth factor

YKL-40 – cartilage glykoprotein-39

1 INTRODUCTION

Rheumatoid arthritis (RA) is an autoimmune, chronic inflammatory disease characterised by painful joints and synovitis that can lead to a damage of cartilage and bone. It affects approximately 0.5-1% of total population (1) and occurs more often in women (60-86%) than men (2-4).

Aetiology and pathogenesis of RA are not completely understood. However, recent advances in research have shown important role of interaction of some genetic susceptibility factors (the strongest being carriage of *HLA-DRB1* and *PTPN22*), with environmental triggering factors such as smoking, infections and others (5-7). Better understanding the pathogenesis of RA would help to improve targeted treatment. However, the greatest challenge for this is the heterogeneity of the disease. Therefore, investigation of predictive and prognostic biomarkers is in high need, and identification of such factors could help discriminate subsets of patients with certain pathogenesis and predisposition of response to a specific therapy.

1.1 PATHOGENESIS OF RHEUMATOID ARTHRITIS

Rheumatoid arthritis is usually divided into two main subgroups in regard to auto-antibody status: seropositive and seronegative. Earlier, rheumatoid factor (RF) was the main autoantibody used for diagnosis and classification into sero-(RF) positive and seronegative disease. However, the commercial anti-cyclic citrullinated peptide (anti-CCP) test, used to measure anti-citrullinated protein antibodies (ACPA) that were discovered later than RF, show higher specificity for RA. The pathogenesis of seronegative RA is very unclear. The disease course of these patients is considered to be more heterogeneous compared with seropositive patients. In contrast, seropositive (anti-CCP positive) RA (2/3 of patients) has been studied in more detail and more clear picture has developed for the pathogenesis of anti-CCP positive RA. First, the discovery that anti-CCP antibodies can develop in non-symptomatic individuals several years before the onset of the disease attracted attention of many researchers to investigate the role of these autoantibodies in the pathogenesis. According to current theory (8, 9), development of ACPA starts from tissue located outside of the joints (for example, lungs or mucous membrane) leading to auto-antibody production. Shift of the process from extra-articular compartments to the joints is yet unclear. It is

believed that expression of citrullinated epitopes on precursor osteoclasts attracts ACPA and triggers activation of these cells leading eventually to bone erosion. They also start producing IL-8 which binds to nociceptors in the joints triggering pain. IL-8 also may attract neutrophils (which is commonly seen in early arthritis) giving a start of the inflammation process (activation and migration of other inflammatory cells, vasodilation and production of proinflammatory cytokines). Synovial membrane becomes infiltrated with macrophages, mast cells, T and B cells and plasma cells. These cells together with synovial fibroblasts produce pro-inflammatory cytokines (tumour necrosis factor (TNF), interleukin-6 (IL-6), IL-17, macrophage colony-stimulating factor (M-CSF), receptor activator of nuclear factor kappa-B ligand (RANKL)) and matrix enzymes that maintain and progress the inflammation in the joints, facilitate angiogenesis, cartilage destruction and osteoclastogenesis accelerating bone erosion (6). Histologically, RA synovium may have two phenotypes: diffuse, where lymphocytes are randomly infiltrated into synovial tissue without any structural formation and follicular, where T and B lymphocytes form clusters, so called germinal-like centres (10-12). It has been shown in RA patients that joints with follicular type of synovitis are at higher risk of destruction compared with diffuse synovitis (13).

1.2 CLINICAL CHARACTERISATION AND OUTCOME OF RHEUMATOID ARTHRITIS

1.2.1 Course and symptoms

The diagnosis of RA is complex and is based on physical examination as well as laboratory immunological analysis of the presence of auto-antibodies towards IgG (RF-positive) and/or anti-CCP (anti-CCP-positive). Numbers of swollen and tender joints detected by rheumatologist together with molecular components of inflammation such as C-reactive protein (CRP) and erythrocyte sedimentation rate (ESR) are other components of the diagnostic work-up.

The course of RA may vary among individuals from mild to more aggressive and destructing features. RA may affect almost any join, however, joints of hands and wrists as well as forefeet are most usually affected ones (14, 15). At the onset, morning stiffness on the joints is often present. Most common symptoms are presented in Table 1. RA may in addition have extra-articular manifestations (Table 1).

Joint pain
Joint stiffness
Weakness
Deformity
Fatigue
Malaise
Fever
Weight loss
Depression
Symmetrical
Distal
PIP
MCP
MTP
Wrist
Ankle
Skin (rheumatoid nodules)
Ocular (keratoconjuctivitis sicca, iritis)
Oral (salivaty inflammation)
Respiratory (pulmonary fibrosis, pleural effusion, cricoarytenoid inflammation)
Cardiac (pericarditis, valvular nodule formation, myocarditis)
Neurological (mononeuritis, nerve entrapment, cervical instability)
Hepatic (increased aminotransferase concentration)
Haematological (anaemia.
thrombocytosis, leukocytosis.
lymphadenopathy, slenomegaly.
thrombocytopenia
Vascular (vasculitis)

Table 1. Clinical features of rheumatoid arthritis*

MCP – metacarpophalangeal join, MTP – metatarsophalangeal joint, PIP – proxim interphalangeal join

* The table is adapted from Lee & Weinblatt, The Lancet 2001 and modified

1.2.2 Disease activity measures

Disease activity of RA is estimated using different clinical measures. Disease activity score based on examination of 28 joints and acute phase reactant (DAS28) is one of the routinely used outcome to monitor the disease (16). DAS28 is based on number of tender and swollen joints (TJC and SJC, respectively) of 28 examined joints (hands, wrists, elbows, shoulders and knees), patient's global health on visual analogue scale (VAS), and CRP or ESR (Table 2).

Table 2. Disease activity measures in rheumatoid arthritis				
	DAS28-CRP	DAS28-ESR	SDAI	CDAI
SJC	+	+	+	+
TJC	+	+	+	+
CRP	+	-	+	-
ESR	-	+	-	-
PatGH VAS	+	+	-	-
PatGDA VAS	-	-	+	+
PhysGDA VAS	-	-	+	+

CDAI - clinical disease activity index, CRP - C-reactive protein, DAS - disease activity score, ESR - erythrocyte sedimentation rate, PatGH - patint global health, PatGDA - patient global disease activity, PhysGDA - physitian global disease activity, SDAI - simplified disease activity index, SJC - swollen joint count, TIC - tender joint count, VAS - visual analogue scale.

Formulas for calculation of DAS28 based on CRP or ESR are expressed, respectively, as follows:

$$DAS28-CRP = 0.56\sqrt{(TJC28) + 0.28}\sqrt{(SJC28) + 0.36ln(CRP+1) + 0.014(GH) + 0.96}$$

$$DAS28-ESR = 0.56\sqrt{(TJC28)} + 0.28\sqrt{(SJC28)} + 0.70ln(ESR) + 0.014(GH)$$

The scale of DAS28 helps to determine patient disease activity (Table 3). Cut-off values used for determining disease activity categories by DAS28-CRP or DAS28-ESR are most often the same. However, it has been shown that DAS28-CRP indicates somewhat lower values compared with DAS28-ESR (17-19). Fleischmann et al illustrated that threshold values of DAS28-CRP of 2.4, 2.9 and 4.6 corresponding to remission, low and high disease activity, respectively were equivalent to established cut-offs for DAS28-ESR (2.6, 3.2 and 5.1, respectively) (20, 21).

Table 3. Disease activity categories					
Categories	DAS28	SDAI	CDAI		
High	>5.1	>26.0	>22.0		
Moderate	3.3-5.1	11.1-26.0	10.1-22.0		
Low	2.6-3.2	3.4-11.0	2.9-10.0		
Remissoin	<2.6	<3.4	<2.9		
Minimum value	0.49	0	0		
Maximum value	9.07	86	76		

CDAI - clinical disease activity index, DAS - disease activity score, SDAI - simplified disease activity index

Clinical or simplified disease activity indices (CDAI or SDAI) are other measures similar to DAS28 (Table 2), but easier to calculate without computer (22). Both include TJC and SJC based on 28 joints, patient global disease activity VAS and physician global disease activity VAS. SDAI includes in addition CRP, too. Both CDAI and SDAI are calculated by simple summing all the components together and have a range between 0-76 and 0.1-86, respectively.

1.2.3 Response criteria and functional assessment

Change and improvement of patients health is measured by the change of disease activity measures. For example, the European League Against Rheumatism (EULAR) has developed response criteria based on the change in DAS28 as well as the value of DAS28 at the follow-up (Table 4). According to the EULAR response criteria, patients are classified in no responders, moderate or good responders (23, 24). Similarly, the American College of Rheumatology (ACR) developed another criterion for evaluation of improvement during therapy. According to ACR criteria, patients achieving at least 20%, 50% or 70% of improvement in TJC and SJC, and in any 3 out of 5 other clinical parameters (pain VAS, patient's global assessment, physician global assessment, ESR or CRP and functional questionnaire), are classified as having respectively ACR20, ACR50 or ACR70 response to the therapy (25). To assess functional activity, health assessment questionnaire (HAQ) is used (26). The HAQ is self-assessing tool which reflects the ability of patients to perform daily activities. For each question patient may receive scores: 0 (without any difficulty to perform), 1 (with some difficulty), 2 (with much difficulty) and 3 (unable to perform). The total HAQ score is a mean of all obtained scores and therefore it has also the range between 0 and 3.

Table 4. EULAR response criteria*				
Improvement	DAS28 at end time-point			
of DAS28	>5.1	>3.2 - ≤5.1	≤3.2	
≤0.6	No	No	No	
>0.6 - ≤1.2	No	Moderate	Moderate	
>1.2	Moderate	Moderate	Good	

DAS - disease activity score, EULAR - European League Against Rheumatism

st The table is adapted from van Gestel et al., 1996. and modified

1.2.4 Imaging

Apart from symptomatic and laboratory data, patients are evaluated by imaging techniques, such as X-rays, ultrasound and magnetic resonance imaging (MRI). The most routinely used is X-ray examination of joints. It helps to detect pathological changes, such as joint space width (JSW), and bone erosion, which are not detectable by physical examination. The difference in JSW at earlier and later time-points is called joint space narrowing (JSN). Assessment of JSN and bone erosion allows to monitor the dynamic of these changes during a given time-period, so called radiographic progression of joint damage (RP). These detected pathological changes are due to processes with shift of bone and cartilage turnover towards degradation. There are different scoring systems developed for the evaluation of bone and cartilage damages and disease progression (27). In 1971 Sharp developed a method of assessment of hands and wrists (28), which was later modified in 1985 and is known as standard Sharp method (29). In 1989 van der Heijde modified the standard Sharp scoring method of evaluation of hands and wrists (van der Heijde modified Sharp score), which includes also evaluation of feet and is based on bone erosions and JSN (30-32). Maximum erosion scores of hands and feet (per joint) are 5 and 10, respectively, and maximum JSN score is 4 per joint. The total van der Heijde modified Sharp score has a range between 0 and 448 per patient. Another radiographic evaluation method of joints was introduced by Larsen in 1974 (33). This score also has been modified several times. Initially it contained also soft tissue swelling and periarticular osteoporosis which was removed during later modifications (34). The x-ray technique, however, has some limitations. For example, alteration in erosion size compared with previous picture might be due to slightly different position of joints and it indicates past events in patients, but no conclusion can be done for the current situation.

While radiography gives the possibility to detect pathological changes occurring on surfaces of hard tissue (bone and cartilage) and reflects the past of the patient, other methods such as ultrasound and MRI are able, in addition, to scan soft tissues of the inflamed joints. These tools can measure and illustrate pathological thickening of synovial membrane, tendons sheaths, ligaments and bursae, which indicates the degree of ongoing inflammation (35, 36). With the help of power Doppler and colour Doppler tools, ultrasound may also illustrate increased blood flow in the joints. In addition, MRI may detect pathological alterations in the bone, so called bone marrow oedema. Another useful aspect for both ultrasound and MRI is that they have been shown to identify subclinical inflammation, therefore are considered as important tools for early detection of pathological changes as well as monitoring patients in remission.

A new method for imaging, fluorescent optical imaging (FOI), is being tested for its utility in early diagnosis and monitoring of patients with RA (37, 38). The method is based on a contrast media which is injected intravenously followed by scanning of hands for detection of the fluorescence intensity. The hyperaemic tissues give higher level of fluorescence intensity compared with normal (uninflamed). The method showed high correlation and agreement with other imaging techniques in one study (38), and weaker findings were in another study (39). A good agreement of FOI with ultrasound for detecting clinically silent synovitis was reported by Kisten et al (40). Higher sensitivity but lower specificity of FOI in RA patients compared with MRI or power Doppler ultrasound was observed by Krohn et al (41).

1.3 THERAPY OF RHEUMATOID ARTHRITIS

Since the aetiology and pathogenesis of RA is not fully understood, the treatment is challenging and is directed towards suppression of immune system by using corticosteroids and disease modifying anti-rheumatic drugs (DMARDs). Even introduction of biological DMARDs (bDMARDs) in treatment of RA did not result in 100% response of patients. The inconsistency of clinical outcome during therapies and presence of such a variation in therapy options highlights the heterogeneity of RA and a need of prognostic biomarker research.

Based on results of many studies and current international guidelines, methotrexate (MTX) is the most recommended non-biological conventional DMARD (cDMARD) for first-line therapy (42, 43). MTX is a folate antagonist leading to reduction of pyrimidine and purine synthesis and stop in DNA replication; therefore it is being used in treatment of oncological diseases. In RA, the beneficial effect of lower doses of MTX is not completely clear (44, 45). However, it is believed that its anti-proliferative action on lymphocytes leads to immune suppression. MTX can also inhibit JAK/STAT pathway (46), which is also a target of recently developed therapies. Taking into account that majority of pro-inflammatory cytokines are synthetized through this pathway, this property of MTX may explain its anti-inflammatory effect on RA. MTX was also found to lead to apoptosis of activated lymphocytes via increasing reactive oxygen species (ROS) (47-49). Another hypothesis of beneficial effect of MTX on RA patients is its ability to release endogenous adenosine, which binds its A2a receptor leading to immunosuppression (50, 51).

In clinical practice of RA therapy approximately 30-40% of patients responds and improves their symptoms and lab outcome. According to EULAR and ACR recommendations, for those patients who do not achieve low disease activity or remission, the MTX therapy is intensified by adding other non-biological cDMARDs, such as sulfasalazine (SSZ) and hydroxychloroquine (HCQ) or biological DMARDs (bDMARDs) (42, 43). It has been proven by many investigations that the use of combinational cDMARD therapy is significantly effective than MTX monotherapy. For example, O'Dell and colleagues demonstrated that addition of SSZ and HCQ to MTX led to significantly better outcome compared with MTX monotherapy (52, 53). Similar result was observed by de Jong et al from tREACH trial (54).

Biological therapy of RA became possible after Köhler and Milstein developed a method of monoclonal antibody production from hybridoma fused cells 4 decades ago (55). Today, there are 9 biological DMARDs used in RA care, of which 5 are TNF inhibitors (infliximab, etanercept, adalimumab, certolizumab and golimumab), one is against B lymphocytes (anti-CD20; rituximab), one against T lymphocytes (abatacept), one against interleukin-6 receptor (IL-6 inhibitor; tocilizumab) and one IL-1 receptor inhibitor (anakinra) (56). The first study on biological therapy (anti-TNF) of RA was published in 1994 illustrating superiority of both higher and lower doses compared with placebo (57). The results from another study confirmed efficacy of addition of infliximab (IFX) to MTX monotherapy (58). In several randomized controlled trials significant effectiveness of etanercept was also proven (compared with placebo groups) on cDMARD-non-responders (59-61). The application of adalimumamb and its efficacy compared with MTX+placebo have been shown in numerous of trials (62-65). On 36 patients from a British double-blinded study, single intravenous infusion of certolizumab showed improvement by ACR20 and ACR50 in a dose-dependent manner (66). Other study applied sub-cutaneous administration of certolizumab, showing that

400 mg dosing resulted in the best improvement compared with placebo or lower doses (67). Keystone and colleagues have demonstrated that certolizumab in combination with MTX was significantly more effective therapy option than MTX monotherapy (68). In an American study, RA patients with inadequate response to MTX received in addition double-blinded subcutaneously golimumab at two different doses or placebo (69). After 16 weeks of the initiation of the therapy, significantly higher proportion of patients achieved ACR20 response from golimumab group, compared with placebo. Similar results were reported from GO-MORE clinical trial, where majority of patients who did not respond to cDMARD therapy, receiving subcutaneously golimumab achieved EULAR good/moderate response (70). Rituximab was developed for treatment of lymphocytic leukemia. In 2001 first results were published on the application of rituximab in few patients with RA (71). All of them achieved response according to ACR70 criteria during therapy. Later the effectiveness of rituximab was shown in other RA patietns (72-77). Anakinra have also been shown to be significantly superior to MTX+placebo in improving clinical and radiological outcomes (78-81). First background evidence that CTLA-4 pathway may be a reasonable target for therapy in RA was demonstrated in 1995, when symptoms of collagen-induced arthritis in rats were prevented by CTLA-4-Ig (82). In 2002 Moreland and colleagues published first evidence of CTLA-4-Ig safety and effectiveness on patients who failed DMARD therapy (83). Later came more studies confirming effectiveness of anti-T cell therapy (abatacept) in cDMARD and bDMARD non-responders (84-89). The high levels of IL-6 in autoimmune diseases led to more investigations on it ending up with development of antibodies targeting IL-6 receptor (90). First evidence of tocilizumab on RA patients was illustrated on a Japanese study by Nishimoto et al, where they demonstrated that patients with persistent symptoms achieved ACR20 response in dose-dependent manner (91). This was followed by a European multicenter study where MTX inadequate responders were randomized to different doses of tocilizumab with or without MTX and MTX+placebo (92). Patients on tocilizumab monotherapy achieved better outcome than placebo group, but those on combination were the best.

The relative effectiveness of combined cDMARD versus bDMARD therapy has been studied in several trials. However, it is not so clear which of these alternatives is better option. In the double-blinded randomized trials TEAR and RACAT, the edition of etanercept was generally not more effective than addition of sulfasalazine and hydroxychloroquine (triple therapy), establishing formal non-inferiority in the latter trial but identifying some clinical outcomes that differed between the treatments (93, 94). Addition of infliximab for 6 months to triple therapy showed a clinically beneficial trend at 2 years compared with triple therapy+placebo group of patients from the NEO-RACo clinical trial (95). Following further yearly examination, however, the slope between the two arms merged closer – resulting in disappearance of the trend at 5 years. In the open-label randomized control trial SWEFOT, the addition of infliximab to MTX was significantly more effective after one year than escalation to triple therapy, but the effectiveness was no longer significant after two years (96).

1.4 PREDICTIVE BIOMARKERS

All these results were obtained on group level. However, it was shown in RACAT trial that some patients who failed to respond to combined cDMARDs did well after switching to bDMARD therapy, and some bDMARD-non-responders did better after they switched to combined cDMARD therapy (94). This suggests that cases are different and an individual approach is needed for more optimal therapy choice for each individual case. For this reason, predictive biomarkers, including demographic and molecular parameters have been studied extensively.

1.4.1 Clinical and demographic factors as predictive tools for disease outcome

Studies on different patient populations show that presence of several factors, including some of the risk factors for aetiology of RA, is indicator of more severe disease and poorer prognosis in RA. For example, smoking and female sex have been shown by many researchers to be poor prognostic factors for disease outcome in patients with RA on different therapies (97-99). Baseline DAS28, on which the treatment outcome is based, is considered one of the strongest and consistently illustrated predictors of response. Results from other baseline clinical and demographic factors are less consistent.

1.4.2 Molecular biomarkers

1.4.2.1 Auto-antibodies

Autoantibodies are one of the earliest components of the disease. They arise several years before disease onset. Most commonly known and used are RF and anti-CCP antibodies, which

have diagnostic role. RF is found in many other autoimmune diseases, but anti-CCP is more specific for RA (100). Being positive for these autoantibodies (but more often for anti-CCP) have been shown in most of studies, relatively consistently, to predict higher risk of radiographic progression (100-106).

Data regarding the association of RF and anti-CCP with subsequent clinical response or remission is more controversial. For example, in predominantly male US veterans with established RA, both RF- and anti-CCP-positivity (but more RF-positivity) were predictive for not achieving remission (107). In an Irish study, anti-CCP-positivity was associated with more lymphocytic infiltration in the synovium, lower proportion of EULAR response and higher proportion of radiographic progression (108). In a Thai study, being double-positive was an indicator of lower likelihood for achievement of remission at 1 year follow-up, compared with double-positive patients (109). Being positive for either of the auto-antibodies and having low likelihood of achievement of remission was shown by other researchers too (110-112). In contrast, several researchers found no association with auto-antibody status at baseline and subsequent clinical outcome (113-116). In addition, higher titers of anti-CCP antibodies have been shown to be predictors of better clinical outcome for some biological therapies (117, 118). The last observation makes anti-CCP antibody interesting possible tool for personalised therapy.

1.4.2.2 Acute phase reactants

C-reactive protein (CRP) is an inflammatory marker produced mainly by hepatocytes during inflammation. Secondary sources might be also monocytes, lymphocytes, adipocytes and some other cells (119, 120). Not being specific for RA, nevertheless, it has a diagnostic role for RA, but also is used to monitor patient inflammation intensity during therapy. Apart from being an inflammatory marker, CRP contributes in maintaining the inflammatory process, by affecting endothelial cells (upregulates VCAM-1 and E-selectin expression) and facilitating migration of leukocytes from blood stream to tissues (121).

CRP is correlated with many symptoms of RA patients and is associated with subsequent radiographic progression (122-125). CRP might have a direct involvement in the joint damage process via increasing RANKL leading to osteoclatogenesis (126).

Similarly to CRP, *erythrocyte sedimentation rate (ESR)* is an indicator of inflammation, correlates with CRP and symptoms of patients with RA, and its high level is associated with

radiographic progression (127-129). It measures rate of sedimentation of erythrocytes in mm per hour which depends on concentration of CRP and other inflammatory molecules in the blood. Apart from CRP, ESR is sex-specific and the threshold values for high category for male versus female is different (>20 versus >30 mm/hr, respectively). In a study, lower baseline levels of CRP (<3 mg/L), but not low ESR (<20 mm/hr) was independently associated with higher likelihood of remission and LDA during anti-TNF treatment (130). In contrast, another study demonstrated higher levels of both CRP and ESR (>10 mg/L and >30 mm/hr, respectively) to be predictive for remission to TCZ (131). Similarly controversial results were observed by Navarro-Compan et al in their systematic literature review (132).

Serum amyloid A (SAA) is another acute phase protein synthesised mainly by hepatocytes and is correlated with ESR, CRP and disease activity in patients with RA (133, 134). Extrahepatic sources, such as synoviocytes and chondrocytes of RA patients, have also been demonstrated (135, 136). In 1983, Chambers et al showed in 185 RA patients that all those with high SAA also had high CRP, however, 40% of patients with low CRP still had high SAA and they were similar clinically with those with high CRP (137). Similar result regarding SAA having higher sensitivity than CRP and ESR have been observed by others (138, 139). This may be caused by the method used (probably, SAA was measured with more sensitive method compared with CRP method). But if the discrepancy is true, both may be useful discriminative biomarkers since in the liver, CRP synthesis is driven by IL-6 and synthesis of SAA by IL-1 (140). SAA has been shown to correlate with 28SJC, and induce expression of MMPs and cell adhesion molecules by synovial fibroblasts and endothelial cells, and facilitate leukocyte migration and angiogenesis (141-145). SAA but not CRP or ESR was also shown to correlate with cartilage turnover markers and predict 1 year radiographic progression (145). However, the pro-inflammatory property of SAA shown *in-vitro* previously was detracted by Björkman et al showing that natural SAA behaves differently from recombinant one (146). The reducing effect of anti-TNF therapy on SAA has been shown by Charles et al, however, not at that magnitude as on IL-6 and CRP (147). Tofacitinib was also shown to decrease both IL-6 and SAA (148). Decrease of SAA by golimumab+MTX during first 4 weeks might be predictive for response at 16 weeks in RA (149). SAA was higher in anti-CCP-positive RA patients compared with negative ones. Its better correlation with DAS28 compared with anti-CCP autoantibodies might make it better biomarker for monitoring RA patients (150). Higher levels of SAA were shown to be associated with a risk of CVDs and renal involvement in

patients with RA (151). SAA was more decreased by ETN+MTX than by conventional DMARDs (152).

1.4.2.3 Inflammatory cytokines, metalloproteinases, growth factors and cell adhesion molecules

In RA patients, there is a communication between immune cells and joint synoviocytes. One of the way of these communications occurs via cytokines. Cytokines are molecules that are released by some cells and affect the same or other cells via their specific receptors. The cytokines may have pro-inflammatory or anti-inflammatory role. In RA the equilibrium is shifted towards predominantly pro-inflammatory cytokines.

Interleukin-1 (IL-1) is an inflammatory cytokine that is commonly produced by different cell types, including synovial fibroblasts in RA patients, and induces production of other inflammatory mediators including TNF- α . In RA, being produced by synovial macrophages and fibroblasts, IL-1 facilitates migration of the inflammatory cells to the joints. By this and inducing proteoglycan degradation, IL-1 plays a fundamental role in joint damage (153). However, Eklund et al has demonstrated that the association of baseline high IL-1 β with the presence of erosions does not predict radiographic progression after initiation of DMARD therapy (154). Natural regulator of IL-1 is IL-1 receptor antagonist (IL-1Ra). In RA, the balance between production of IL-1 and IL-1Ra is altered (155-157). Treatment of RA patients with recombinant IL-1Ra (anakinra) have been shown to drop clinical disease activity and slow down radiographic progression (78, 79, 158). IL-1 was also shown to be associated with anaemia in RA patients (159). In 48 RA patients treated with MTX, having high levels of baseline IL-1ß AND lower levels of IL-8 was associated with better clinical outcome (based on Ritchie-index, duration of morning stiffness and CRP) (160). Low level of IL-1 β expression was observed in synovial tissue of patient with diffuse lymphoid infiltrates, which was associated with seronegative mild form of the disease course, while high expression of IL-1 β was seen in patient with granulomatous synovial tissue – associated with extra-articular manifestation with nodules formation (10). Seitz et al showed that in patients with good ACR response (over ACR50) to 6 months of MTX, the ratio of IL-1Ra/IL-1β produced by PBMCs was significantly lower compared with non-responders (<ACR20) (161). Polish researchers showed that MTX+prednisone decreased serum levels of IL-1β and IL-6 but there was no association between the change and clinical response to the treatment (162). In 15 patients with RA and temporomandibular joint pain, high pre-treatment level of plasma IL-1 β was a poor prognostic factor for reduction of the pain after treatment with IFX (163). In another study on 51 RA patients, pre-treatment synovial expression of IL-1 β did not differ between ACR20 responders and non-responders to IFX (164). Kayakabe et al, on the other hand demonstrated in white blood cells collected at baseline that their LPS-stimulation caused significantly higher production of IL-1 β in 6-month post-anti-TNF treated responders (EULAR good/moderate) than non-responders (165). Baseline low or early (after 4 weeks) achievement of low serum IL-1 β levels could predict response to tocilizumab after 1 year of therapy in advanced RA patients (166).

IL-6, together with TNF- α , are considered as disease driving main molecules. IL-6 is a proinflammatory cytokine that is produced by variety of cells including lymphocytes, fibroblasts, osteoblasts and endothelial cells. Its action occurs via both membrane bound IL-6 receptor (IL-6R) and soluble receptor (sIL-6R) affecting different cell types. In RA, IL-6 is increased in synovium and blood, and its level correlates with clinical disease activity and joint damage. Its proliferative effect on endothelial cells via increase of VEGF (167) suggests that IL-6 may contribute to angiogenesis in inflamed synovium of patients with RA. T- and B-cell responses are also require IL-6 stimulation. For example, IL-6 helps in differentiation of B cells into plasma cells and antibody production (168, 169), an event that is considered as a start point in the pathogenesis of RA. Moreover, a correlation was observed between IL-6 and RF titres both in synovial fluid and serum of seropositive patients (170, 171). IL-6 together with IL-1ß also differentiate naïve T-cells into Th17 subset (172), which has been shown to have a fundamental role in autoimmune diseases. Expression of IL-6R on neutrophils suggests that IL-6 may play a role in neutrophil infiltration of inflamed synovium seen in RA. Indeed, increased IL-6, caused by co-culturing of RA fibroblasts with endothelial cells, and enhanced adherence of neutrophils to endothelium have been reported (173). Moreover, biopsies of synovial tissue from knee joints of 51 RA patients showed positive correlation between synovial IL-6 levels and infiltration of inflammatory cells in synovium (174). IL-6 transsignalling (interaction of IL-6 with sIL-6R) facilitates osteoclastogenesis via RANKL pathway, leading to bone resorption and joint destruction in RA while the osteoclastogenesis was inhibited by anti-IL-6R antibodies (175, 176). In collaboration with IL-1, IL-6 induces production of MMPs by synovial cells, leading to cartilage degradation (177). Via direct stimulation of hepatocytes, IL-6 may enhance acute phase response (178, 179) and production of hepcidin (180) – a peptide disturbing iron metabolism and causing anaemia, which is commonly seen in RA. Fatigue, which is a common accompanying symptom in RA was also shown to be achieved in healthy volunteers after IL-6 injection (181).

Methotrexate and other conventional DMARD treatment of patients with RA decreased levels of IL-6 (182, 183). Reduction of IL-6 level during first year of DMARD therapy was shown to correlate with clinical and radiographic outcome at 3 years (184). However, a study on 33 RA patients treated with etanercept and 46 RA patients treated with rituximab did not show statistical difference in baseline serum IL-6 levels between responders and non-responders at 3 months (185, 186). Similarly, a post-hoc analysis of 2 randomised control trials (CIMESRTA, N=150 and OPERA, N=180) failed to illustrate predictive capacity of baseline serum IL-6 level for clinical remission of radiographic progression during 2 and 5 years of follow-up (187). Surprisingly, another study of 73 established RA patients showed better clinical outcome from ETN therapy among those, who had higher baseline IL-6 level (188). Contrary, a study on incomplete responders to conventional combination DMARD or anti-TNF therapy also illustrated association of low IL-6 (<15 pg/ml) at start of RTX with higher likelihood of good EULAR response at 6 months of follow-up (189). Japanese researchers demonstrated in biologic-naïve RA patients that lower pre-treatment serum IL-6 levels were associated with response to TCZ (190). Moreover, they reported that patients with low, but not high pre-treatment IL-6 had more benefit from TCZ than from IFX therapy. Wang and colleagues, on the other hand demonstrated that baseline serum IL-6 levels was not predictive for remission to TCZ therapy based on data from five clinical trials (191). Nevertheless, another Japanese study in DMARD-incomplete responders demonstrated that patients with low serum IL-6 levels (<30 pg/ml) at 3 and 6 months after TCZ start had significantly higher proportion achieving CDAI- and DAS28-based remission at 1 year (192). The association was independent of age and sex. Results from a Japanese study on 33 RA patients with incomplete response to MTX and starting IFX therapy demonstrated poorer clinical outcome in those patients who had higher pre-IFX treatment serum IL-6 level (>5.45 pg/ml) (193). Another Japanese study of 25 RA patients with incomplete response to IFX (3 mg/kg treatment dose) that underwent dose escalation, demonstrated that patients with pre-escalation serum IL-6 level of <5.16 pg/ml experienced more increase in sIFX level with more response compared with patients with ≥ 5.16 pg/ml level of serum IL-6 (194). This led to better 1-year drug survival after escalation in patients with low compared with high serum IL-6 level. A study on 253 eRA patients showed that serum IL-6 levels were associated with radiographic progression during 4 years of follow-up, independently from BMI (195). However, the association became a non-significant trend when anti-CCP was included in the model. Gottenberg and colleagues, on the other hand, observed an independent from anti-CCP status association of baseline IL-6 with 1 year radiographic progression in 578 DMARD-naïve eRA patients (196). In another study of 173 RA patients urinary IL-6 showed independent prediction of RP at 1 and 3 years follow-ups (197). Patients with high levels of IL-6 had approximately 3 times higher risk of RP. This association remained even when looking in patients with low ESR only. In 88 newly diagnosed untreated RA patients pre-treatment higher IL-6 levels (>7.6 pg/ml, based on ROC curve) were associated with MRI-detected bone erosion during 1 year of follow-up (198). The association was independent from, and even stronger than seropositivity. Baseline low IL-6 levels, on the other hand, predicted MRI-detected erosion repair.

As mentioned earlier, TNF is another key, and mostly studied cytokine in pathogenesis of RA. Similar to IL-6 it affects many different cell types (pleiotropic) and is produced mainly by monocytes, macrophages, some T cells and other type of cells such as synoviocytes. Being a classical pro-inflammatory cytokine it can also express anti-inflammatory effects. Its action is expressed via binding one of the two receptors: TNF-RI (p55) and TNF-RII (p75). TNF-RI is expressed on many cell types, while TNF-RII mainly on immune and endothelial cells, as well as synoviocytes. TNF-RI has death domain and can activate caspase system leading to cell apoptosis, while TNF-RII lacks this domain. Both receptors may also induce cell proliferation and inflammation (199). A natural way of regulation of TNF-signalling is proteolytic cleavage of the membrane bound receptors, which still can bind TNF but do not induce intracellular signalling (decoy receptors) (200). Disability of cleavage of TNF-RI caused by mutation, has been shown to induce auto-inflammatory conditions (201). Signalling from TNF leads to production and secretion of many other inflammatory mediators such as IL-1, IL-6, GM-CSF and IL-8 (202-205). TNF facilitates also osteoclastogenesis and bone erosion through RANKL/RANK and other pathways, as well as cartilage degradation, angiogenesis and attraction of neutrophils (204). Blocking TNF in RA patients and mice with arthritis lead to decrease in nociceptive activity in the CNS detected by MRI already 24 hours after first injection of the anti-TNF drug (206). Notably, clinical and serological inflammatory signs were still unaffected. This suggests that TNF also contributes to pain feeling via directly affecting the CNS in patients with RA.

TNF has been shown to be associated with histologically different disease phenotypes. For example, a study on 25 RA patients showed that subjects with follicular synovitis had

significantly higher levels of serum TNF compared with patients with diffuse infiltrates (207). A similar study on 47 RA patients by the same investigators showed similar results: higher serum levels of TNF, TNF-RI and TNF-RII in patients with follicular synovitis compared with diffuse histological type (208). In 50 RA patients, higher baseline production of TNF-RI from PBMC and higher serum levels of TNF-RII were observed in better responders to 6 months of MTX therapy, when compared among 4 response groups (>ACR70 and ACR70-50, ACR20-50 and <ACR20) (161). However, no significant association was observed for TNF. In an American study, pre-treatment sTNF levels were significantly higher in RA patients who did not respond to IFX therapy (assessed by Richie score index) compared with those who responded (209). In non-responders, increasing frequency of administration improved better clinical outcome than increasing the dose of IFX. A study in 60 RA patients demonstrated in multivariate analysis that synovial tissue mRNA level of TNF (among other cytokines) was predictive for joint damage progression (210). Contrary, a study of 99 serum biomarkers (including TNF) in 152 RA patients from GO-BEFORE trial did not reveal significant correlation between baseline levels of TNF or in the change of TNF at week 4 and radiographic damage progression at week 28 or 52 (211). Another study of 44 RA patients did not reveal any association of peripheral blood mRNA level at baseline with subsequent response to IFX at 22 weeks (212). Immunohistochemical examination of pre-treatment synovial biopsies of 143 RA patients indicated association of higher expression level of TNF with subsequent response (delta-DAS28>1.2) to IFX at week 16 (213). Responders also had higher number of infiltrated macrophages and T cells (potential sources of the TNF). In 33 RA patients treated with ETN, serum level of TNF at baseline was not significantly different among responders (improvement in 3 of 4 ACR criteria) and non-responders at 3 months (185). In contrast, a study on 93 early RA patients from Japan, Sweden and USA demonstrated predictive capacity of pre-treatment serum TNF for response to ETN (higher in responders) (214). In 42 RA patients treated with non-biological DMARDs, baseline serum TNF was significantly lower in ACR20 responders than non-responders at 6 months (215). A Japanese study done on 327 RA patients from a double-blinded randomized control trial (RISING), (patients started IFX after MTX failure), illustrated a clear association of baseline plasma TNF levels with response (EULAR good or DAS28 remission or ACR-based) at 54 weeks (216). In patients receiving the highest dose of IFX (10 mg/kg) significantly higher proportion of patients responded if they had lower baseline TNF levels (dose-response relationship among patients with low, intermediate and high levels of plasma TNF levels). It was also observed that patients with high baseline levels of TNF had lowest level of serum IFX at 54 weeks, which was also associated with poorer outcome.

IL-17, another cytokine, relatively recently have been shown to be involved in pathogenesis of autoimmune diseases. It is mainly produced by Th17 subset of CD4+ T cells and can be considered as opposing balance to Treg cells (217). IL-17 acts via its dimerised receptor leading to production of other inflammatory cytokines and chemokines, which could partially explain migration of neutrophils and monocytes (218, 219). Increased level of IL-17 has been reported in synovial fluid and serum of patients with RA (220, 221). Surprisingly, a nested case-control study has demonstrated that levels of IL-17 in pre-RA individuals (subjects without any symptoms and complains, who developed later RA) was significantly higher compared with the levels at the diagnosis (222). It can also induce osteoclastogenesis via RANKL pathway. The inflammatory and bone resorptive effect of IL-17 is synergised by the presence of TNF (219). In RA patients, serum IL-17 levels are highly correlated with disease activity, serum levels of TNF and CRP, and synovitis and bone erosions detected by MRI (221, 223). Nevertheless, IL-17 seems to be more important for pathogenesis of other inflammatory diseases such as psoriasis, since blockade of IL-17 or its receptor has been shown to have the best effect and were approved for psoriasis (219).

In 60 RA patients from the DAMAGE study cohort, expression level of synovial tissue mRNA of IL-17 in combination with synovial mRNA of TNF or disease duration was shown to predict joint damage progression detected by MRI at 2 years of follow up (210). In 20 active RA patients (10 treated with MTX and 10 - with adalimumab), Th17 cell levels in peripheral blood at baseline were associated with poor clinical outcome at 3 months (224). Levels of IL-17 were decreased in 25 RA patients after 30 weeks of treatment with MTX+IFX, but not in patients on MTX monotherapy (n=20) (225). A study in 48 RA patients treated with anti-TNF (adalimumab or IFX) demonstrated that baseline IL-17 levels were significantly higher in EULAR non-responders compared to moderate/good responders after 6 months (226). In addition, among other Th17-related cytokines only IL-17 increased during the therapy in non-responders, while others decreased. A multivariate logistic regression analysis indicated IL-17 the only predictor of poor response. Similarly, Alzabin et al demonstrated in 2 groups of RA patients (n=24 and n=19) treated with anti-TNF, that in nonresponders there was significant increase of serum IL-17, while in responders the level was stable (227). Also there was an inverse relationship between proportion of Th17 cells at baseline and DAS28 at 4 week after the therapy. A double-blinded randomised control trial of TCZ therapy in patients with inadequate response to DMARD therapy, demonstrated that baseline low serum level of IL-17 was associated with achievement of remission at 12 weeks (228). Of patients with low IL-17 levels, 48% achieved remission compared with only 17% of those with high IL-17 levels.

In the joints of RA patients, a release of *matrix metalloproteinases (MMPs)* occur, that leads to digestion and destruction of bone and cartilage tissue. There are 23 different types of MMPs in the humans (from MMP-1 to MMP-28, where MMP-4, MMP-5, MMP-6 and MMP-22 do not exist in the nomenclature) (229). Physiologically, the MMPs are proteases with Ca^{2+} or Zn^{2+} active sites. They are playing important role in degradation of components of extracellular matrix, such as collagen, during tissue remodelling at different stages of development of the organisms. Typical source of MMPs are cells of connective tissues of organs, fibroblast, for example, but also other cells: neutrophils, monocytes, macrophages and endothelial cells. Expression of MMPs may be increased by inflammatory cytokines, hormones and growth factors.

In patients with RA, main matrix degrading members of MMPs are considered MMP-1 (responsible for degradation of collagen type I, II, III, VII and X), MMP-3 (degrades collagen type III, IV, IX and X), MMP-7 (degrades collagen type IV and X), MMP-9 (degrades collagen type IV, V, VII and X) and MMP-13 (degrades collagen type I, II, III and IV) (230, 231). It has been shown that MMPs, for example, MMP-1 and MMP-3, reflect clinical disease activity and structural damage of the joints in patients with RA (232-236). Their levels can also decrease during therapy, which is accompanied by clinical improvements (237, 238). MMP-7 has been shown to be linked with tissue remodelling in the lungs in patients with interstitial lung disease. In RA patients with interstitial lung disease, circulating levels of MMP-7 are significantly higher compared to patients without the complication (239). In a Japanese study of early RA patients, higher serum MMP-3 levels were associated with increased risk of joint damage (based on Larsen score) in the future (240). A study in RA patients revealed MMP-1 level more that MMP-3 level to be associated with development of erosions and stronger association of MMP-3 compared with MMP-1 with CRP (241). Early changes in MMP-3 caused by treatment using MTX or tenidap were predictive for later clinical response evaluated by DAS (242). Another study in 98 early untreated RA patients demonstrated that baseline serum MMP-1 and MMP-3 levels correlated with clinical improvements and joint damage progression (delta Larsen score) (243). Even patients with low CRP but high MMP-1 and MMP-3 showed higher risk of erosive disease during subsequent 1 year. In patients without erosion at baseline, high level of MMP-3 was predictive for progression of Larsen score. A study in RA patients from ASPIRE cohort showed that baseline serum MMP-3 level as well as its reduction during first 6 weeks of therapy were associated with response (based on ACR criteria) at 54 weeks (238). Among other baseline markers, in a multivariate logistic regression model, only MMP-3 was predictive for clinical and radiographic outcome at 1 year. Similar predictive results of MMP-3 (especially for radiographic progression) were seen by other researchers (244-248). However, a study of 45 RA patients treated with etanercept showed that reduction of MMP-3 during 6 weeks of therapy did not differ between responders (patients in remission, n=10) and non-responders (n=35), while the decrease of another cartilage turnover biomarker, cartilage oligomeric matrix protein (COMP) was significantly greater in responders than nonresponders (249). Another controversial result was reported by Japanese researchers illustrating that baseline MMP-3 levels had a trend of higher level among progressors than non-progressors, while RF- and ACPA-positivity were significantly associated with subsequent radiographic progression (250). In contrast to this, a study in 118 RA patients that were followed up during 8 years, even though in a univariate analysis many baseline markers (including COMP and MMP-3) were associated with RP at 8 years of follow-up, a multivariate analysis revealed only MMP-3, anti-CCP-positivity and baseline radiographic damage of joints as independent predictors of radiographic progression (251). Significantly greater reduction in serum MMP-9 was observed in responders compared to non-responders to golimumab add-on therapy in MTX inadequate responders (149). In 114 patients with RA, an improvement of MMP-3 during first 4 weeks of therapy with adalimumab was independently from DAS28-CRP associated with remission at 1 year (252).

Human cartilage glycoprotein-39 (YKL-40) is a protein produced by chondrocytes, synovial fibroblasts, macrophages and neutrophils. It is considered as a chitinase enzyme with lost enzymatic activity towards chitin. Increased levels of serum YKL-40 were linked with inflammation and tissue remodelling, however, still the physiological role of YKL-40 is unclear (253). In synovium of patients with RA, compared with healthy controls YKL-40 is also elevated. Even though serum levels are significantly lower than that in synovium, there is a clear correlation between them. Moreover, both synovial and serum levels reflect disease severity and degree of joint damage in RA (253, 254).

In a study of 191 early RA patients treated with DMARDs (MTX or SSZ), baseline serum levels of YKL-40 were not associated with radiographic progression during subsequent 3

years of follow-up (255). Similar non-association of baseline YKL-40 with 2 years radiographic progression was observed in another RA group treated with adalimumab, even though it was correlated with CRP, MMP-1 and MMP-3 (236). Similarly, a Finish study confirmed unpredictability of baseline serum YKL-40 levels for subsequent radiographic progression, however, correlation with inflammatory markers was obvious (256). A study in 136 RA patients also showed no predictive value of baseline YKL-40 levels for RP during subsequent 5-10 years (257). And finally, a study in patients from CIMESTRA trial (n=150) confirmed non-predictive ability of serum YKL-40 for RP, but also for achievement of clinical remission (187). In the same study, the authors confirmed these results in a validation cohort from OPERA trial (n=180).

Leptin is a lipid hormone produced by white adipose tissue and is known as a regulator of appetite and energy expenditure via acting on hypothalamus and inhibiting production of neuropeptide Y. Its production is positively correlated with body weight and decreases appetite in order to avoid obesity (258). Leptin has been found to be related also to inflammation and immune response. This relationship is two-sided: pro-inflammatory cytokines increase leptin level, as well as leptin induces production of pro-inflammatory cytokines and activates phagocytosis (259). Leptin's pro-inflammatory role via impact on innate and adaptive (mostly T cells) immune systems has been shown in many studies (260). In RA, synovial leptin level is significantly lower than in serum, and RA patients have significantly higher level of leptin than healthy controls (261). However, the data about leptin in RA is quite controversial among different studies, as discussed in Tian et al review article (260). Some studies showed protective role of leptin for radiographic progression, while others failed to illustrate that. Similarly, a controversial results about effect of anti-TNF therapy on serum leptin is discussed in the same review, indicating no change in the majority of studies, however, some studies observed increase in leptin level. Probably, its high dependence on proportion of white adipose tissue makes it difficult for any conclusion. In a group of consecutive RA patients treated with non-biological DMARDs, baseline leptin was associated with higher DAS28 at 6 months (262). In patients with normal BMI, the association hold also for 1 and 2 years outcome. The complexity of results regarding leptin as reflector or predictor of disease activity could be explained by its dependence on other parameters, such as proportion of white adipose tissue. This in its turn is affected by body weight and gender. Smoking should also be considered when investigating leptin in RA patient, since it suppresses leptin production (263).

Another adipokine, *resistin*, which was initially thought also to be produced mainly by adipocytes, and to cause insulin resistance, later has been shown to be produced by peripheral blood mononuclear cells and more to be involved in subclinical inflammation processes and autoimmune diseases (264). In RA, levels of resistin in both synovial tissue and serum is elevated compared to healthy controls and even compared to patients with osteoarthritis and spondyloarthropathy (265). Resistin is produced by different immune cells on response to inflammatory cytokines and induces homing of endothelial progenitor cells to the joints, and overexpression of VEGF (266). Consequently, resistin might be involved in the process of angiogenesis in RA. Resistin was also shown to correlate with disease activity markers such as serum TNF and CRP, RF anti-CCP and joint radiographic data (267). Anti-TNF treatment decreases serum level of resistin in RA patients which is associated with post-treatment reduction of CRP and TNF (268). However, as usual with the majority of biomarkers, there are some studies not confirming above mentioned results for resistin. For example, Otero et al showed that leptin, adiponectin and visfatin levels but not resistin levels were significantly different in 31 RA patients compared to 17 healthy controls, and that the mentioned three biomarkers but not resistin correlated with CRP (269).

For prediction of radiographic progression over 4 years, baseline resistin levels had no value (195, 270). Regarding clinical disease activity only correlations have been published between resistin and different disease activity measures from the same time-points, but no predictive results have been reported.

Some of characteristic features of RA are hyperplasia of synovial fibroblasts leading to invasion of the synovial tissue, hypoxia and neovascularisation, leading to pannus formation. *Epidermal growth factor (EGF)* is a known marker for tumour tissue since it induces neoplastic alteration. It also is responsible for angiogenesis. Similarly to tumour, in RA, EGF is upregulated. Both synovial and serum concentrations are higher compared to healthy individuals, but even compared with osteoarthritic patients (271-273). Hyperplasia of synovial fibroblast may partially be explained by increased EGF, since upregulation of EGFR expression on these cells is reported in RA patients. A significant correlation with neovascularisation also indicates its role in the latter. A mouse model of collagen induced arthritis demonstrated that inhibition of EGFR reduces symptoms of the arthritis, and proliferation of synovial fibroblasts and osteoclasts, indicating indirectly that EGF and its signalling may play an important role in the pathogenesis of RA (273). These results were replicated by another research group (274).
Early changes in serum EGF levels were found to be positively correlated with later changes in total van der Heijde-Sharp radiographic score in early RA patients on MTX monotherapy, but not in patients on MTX+golimumab (211).

Vascular endothelial growth factor (VEGF) is the main angiogenic factor that has been considered in RA patients. It is produced by macrophages, synovial fibroblasts, platelets and endothelial cells (275). Similar to EGF, levels of VEGF have been shown to be higher in synovial tissue, fluid and sera of RA patients compared with healthy people or osteoarthritis patients (276). Fast growth of fibroblast-like synoviocytes causes hypoxic environment in the arthritic joints which activates angiogenesis via upregulation of VEGF (277). Blocking VEGFR on the other hand improved symptoms in collagen induced arthritis in mice (278). In RA, serum levels of VEGF have been shown to correlate with inflammatory markers and clinical disease activity symptoms such as CRP, tender joint count and pain. Pro-inflammatory cytokines such as TNF, IL-1, IL6 and others, induce production of VEGF and therapy of patients with RA (both non-biological and biological DMARDs) decreases the serum level of VEGF and reduce newly built capillaries, but not normal blood vessels (275, 277, 279).

Increased levels of serum VEGF is associated with radiographic progression at 1 year of follow-up and reduction of sVRGF levels – with clinical improvements (280). Another study in two different cohorts of RA patients: CIMESTRA (N=150) and OPERA (N=180), showed correlation of VEGF with DAS28, but could illustrate predictive ability of VEGF for radiographic progression at 2 years of follow-up only for OPERA cohort, and for neither of the cohorts was VEGF predictive for clinical remission (187).

Migration of the leukocytes from blood stream through endothelial layer to the affected tissue is one of necessary and indispensable processes for inflammatory response. The cytokines that are released from macrophages and other cells in the damaged tissue play a crucial role in recruiting the leucocytes. The process of migration of the inflammatory cells occurs due to changes on the surface of both the leukocytes and endothelial cells, which enables the former to become more adherent to the later. In this way the leukocytes start rolling on the blood vessel walls before the migration occurs. Stickiness of the cells is ensured by adhesion molecules that are overexpressed on the membrane, induced by inflammatory cytokines. Here, *vascular cell adhesion molecule-1 (VCAM-1)*, among other is known to have an important role for binding integrins on the surface of the leukocytes. VCAM-1 is mainly

expressed on endothelial cells, but in inflamed environment its expression is detected on other cells such as tissue macrophages, dendritic cells, bone marrow fibroblasts and others (281).

The importance of VCAM-1 in RA has been shown by many researchers, which is discussed in a review article by Kong et al (281). Serum level of soluble VCAM-1 is higher compared with healthy controls. Treatment with DMARD or biological medications decreases its level. Blocking VCAM-1 with monoclonal antibodies dramatically reduces signs of arthritis in the CIA moue models. However, when studying expression of different molecules on synovial membrane in 40 RA patients treated with non-biological DMARDs, an association was found between decrease in expression of other cell adhesion molecules, but not VCAM-1 on the synovial biopsies and improvement of clinical outcome (282). Macias et al demonstrated in RA patients that after MTX monotherapy, responders had significantly lower serum VCAM-1 compared with incomplete responders (283). In 143 RA patients failing two DMARD therapies and starting IFX, expression of VCAM-1 in joint tissue sections, detected by IHC just before IFX start, did not differ between subsequent responders and non-responders at week 16 of IFX therapy (213).

1.4.2.4 Multi-biomarker disease activity score

RA is a very heterogeneous disease. In addition to this, different clinical trials address different questions, using different inclusion and exclusion criteria, which make patients to differ significantly between studies. And finally, the stage of disease could affect variations in pathological processes and treatment responses. These are important facts that one needs to consider when trying to replicate biomarker research on different RA cohorts. The abundance of discrepant results for the same biomarker could be explained by these factors. Therefore using one biomarker would probably always lead to failure in validation of results in a different cohort.

A multi-biomarker disease activity (MBDA) score (Vectra® DA) is a blood-based test that has been developed as a tool for monitoring of disease activity in complement to existing disease activity measures such as DAS28 and radiographic imaging of the joints (284, 285). It has been developed by a multi-step process, starting from identification of 130 candidate molecules from literature searches, and narrowing down during the further prioritisation studies to 24 candidate biomarkers (286). The 24-biomarker-based disease activity score already correlated with DAS28 and radiographic progression. At the next stage of development, some biomarkers were eliminated because of their high dependence on the conditions of the analyses, resulting in a high variability, and other reasons. After the final optimisation and algorithm training, 12 biomarkers remained in the model. The algorithm of the final MBDA score is calculated from serum concentrations of the following 12 biomarkers: VCAM-1, EGF, VEGF, IL-6, TNF-RI, MMP-1, MMP-3, YKL-40, leptin, resistin, SAA and CRP. The MBDA score is well correlated with DAS28 and other disease activity measures, and has a scale between 1 and 100 indicating from the lowest to the highest disease activity, respectively (284, 287, 288). Its validated cut-offs generate categories of patients in LDA (MBDA score <30), moderate disease activity (MBDA score 30-44) and high disease activity (MBDA score >44). Being correlated with disease activity and inflammatory markers, the MBDA score seems to complement them as shown in the study by Lee et al (289). In their study it was shown that many RA patients with CRP<1.0 mg/dl, had moderate/high MBDA score and there was a linear association between SJC and the MBDA score but not CRP. The dynamic of the MBDA score also reflects the change of disease activity measured by other clinical methods. These associations were independent of a type of anti-TNF treatment (IFX, adalimumab or etanercept) (290). However, assessment of the MBDA score in RA patients treated with tocilizumab should be interpreted cautiously, since tocilizumab causes increase of serum IL-6 leading to increase in the MBDA score, while decreasing other disease activity indices (291).

Predictive studies for the MBDA score have also been conducted by many researchers. For example, the MBDA score predicted progression of joint damage detected by radiographs, ultrasound or MRI in different RA studies (292-296). However, Bakker et al demonstrated that even though the MBDA score was well associated with disease activity, it could not predict subsequent radiographic progression (287). It was also shown that the MBDA score predicting relapse in RA patients that were in sustained remission (297). The prediction was independent from ACPA and their combination increased associations: in patients with low MBDA score and positive ACPA status only 13% relapsed, while among those with higher MBDA score and positive ACPA status 76% relapsed. In the DRESS study, in patients in remission and tapering anti-TNF therapy the MBDA score did not predict flare or successful tapering/discontinuation, however, did predict flares in usual care patients (298). In contrast to these associations mentioned above, another Dutch study (POET) has demonstrated that patients in LDA and tapering anti-TNF treatment, high baseline MBDA score predicted flare at 12 months (299). Another controversial finding was reported in a study of RA patients

treated with abatacept or adalimumab, where the MBDA score was correlated with DAS28-CRP and CDAI only at baseline, but not at follow-ups (300). Moreover, the researchers reported that CDAI was better associated with radiographic progression outcome than the MBDA score. To summarise, it is worth to mention that for interpretation of the MBDA score, consideration of the age, and BMI of the patients have an important role (301).

1.4.2.5 Serum level of therapeutic bDMARD and anti-drug antibodies

Prediction of response to bDMARDs should have been more clear compared to nonbiological DMARDs, because of a clearly defined target. However, the prediction is complicated by the immunogenicity of the therapeutic drugs. The structure of the bDMARD molecules triggers the immune system of the patients to respond and generate anti-drug antibodies (ADA). Most of the cases these antibodies are neutralising (binding to the targetbinding region of the drug molecule). But even non-neutralising antibodies may create immune complexes leading to accelerated clearance of the bDMARD from the organism of the patient. In any of the cases the immunogenicity will lead to decrease of therapeutic drug concentration in the blood consequently leading to loss of efficiency. Infliximab is considered to be the most immunogenic (302) because of its not fully humanised molecular structure (TNF-binding site is from mouse).

According to different studies the proportion of RA patients developing ADA to IFX fluctuates between 12% and 54% (303-310). Low drug levels in the blood of patients was associated with ADA-positivity (305, 307, 311). In addition, both low drug level and ADA-positivity have been associated with poorer clinical outcome and worse drug survival compared with patients with high drug level or ADA-negative patients (303, 312-315). These observations suggest that monitoring of patient bDMARD level in the blood as well as analysis for ADA may provide essential information to the rheumatologist for optimisation of biological therapy in patients with RA. Garces and colleagues, for example, have suggested an algorithm of treatment of RA patients with bDMARDs based on the availability of information about current disease activity, serum trough level of the therapeutic bDMARD and ADA status (316). Such studies need validation on different RA cohorts using the same methods of analysis for serum drug levels and ADA status. Confirmation of the importance of the blood drug monitoring and ADA status for achievement of good clinical outcome would improve dramatically the healthcare in rheumatology.

In conclusion, RA, as a very heterogeneous disease with unclear aetiology and pathogenesis, is challenging for investigation. The optimal therapy is different for individuals; therefore conducting biomarker studies has an important role in discovery of individuals with different mechanisms of disease. In biomarker discovery a huge role plays the method and patients inclusion criteria since different methods of measurement of biomarkers, serum drug levels and anti-drug antibodies may yield very different results, likewise whether patients were on different therapies during several years or they are newly diagnosed, treatment-naïve early RA patients. Considering these factors in the investigations will provide more reliable data for description of predictors in RA populations.

2 AIM OF THE THESIS

2.1 OVERARCHING AIM

In the presence of challenge of optimal therapy choice and to predict disease course and outcome, the general aim of this PhD thesis was to investigate different blood proteins related to inflammation and immune system as predictors of disease outcome in patients with early RA. The exploration of predictors will contribute in identifying patient phenotypes with certain characteristics, which will enable the best personalised approach for each individual patient.

2.2 SPECIFIC AIM

The study-specific aims of this PhD projects were:

- 1) For Study I
 - a) Using the MBDA score, to identify patients at high versus low risk of subsequent joint radiographic progression.
- 2) For Study II:
 - a) Using the MBDA score at the time of treatment escalation, to identify patients at higher chance of response to IFX or conventional triple therapy (TT), after failure to MTX monotherapy.
- 3) For Study III:
 - a) Using individual biomarkers that comprise the MBDA score, to identify patients at higher versus lower chance of response to MTX monotherapy in early RA.
 - b) Using affinity proteomics analyses to explore serum biomarkers that could identify patients at higher versus lower chance of response to MTX monotherapy in early RA.
- 4) For Study IV:
 - a) In early RA patients randomised to IFX+MTX therapy, to study relationship between serum IFX concentration, anti-drug antibodies (ADA) and clinical outcome. We also aimed to investigate baseline predictors for low serum IFX level and development of ADA.

3 MATERIALS AND METHODS

3.1 PATIENT POPULATION

For all study projects materials from Swedish Farmacotherapy (SWEFOT) trial were used (96, 317). The SWEFOT is a randomised-controlled clinical trial of DMARD-naïve, early RA patients (N=493). Newly diagnosed RA patients with active disease (DAS28 >3.2), symptom duration <1 year and age >18 years were included to the trial and started MTX monotherapy for 3 months duration. Patients achieving low disease activity (DAS28 \leq 3.2) after 3 months continued the MTX therapy and those patients with DAS28 \geq 3.2 were randomised to conventional triple therapy (MTX-sulfasalazine+hydroxychloroquine) or to biological therapy (MTX+IFX). The patients were followed up 2 years from the inclusion into the trial (Figure 1).

Schematic illustration of the SWEFOT trial design





This thesis is composed of four Study projects (Figure 2) and for each of the project a subset of patients was selected based on availability of data needed for the specific study aim. For the MBDA score projects, totally serum samples of 302 patients were analysed at baseline, of whom 290 were analysed at 3 months and 190 (only randomised patients) at 1 year. However, each of the substudies done on the MBDA score data, different number of these 302 patients were selected (see below for details in each individual project).



Figure 2. Plan of PhD projects

3.2 STUDY I

In Study I the MBDA score was investigated for prediction of subsequent radiographic progression. In the first paper of the Study I (paper I), the MBDA score of 235 patients at baseline were analysed for prediction of RP at 1 year. For the second paper (paper II) the MBDA scores at baseline, 3 months and 1 year were analysed in 220, 205 and 133 patients respectively for prediction of RP between baseline and 1 or 2 years and between 1 and 2 years. For the RP, van der Heijde modified Sharp scores (SHS) were used. An increase of the SHS >5 was considered as RP. Cut-offs of >3 and >0 were also compared. The Proportion of patients with RP were compared among categories of the MBDA score (low: <30, moderate: 30-44 and high: >44) and other disease activity measures (CRP, ESR and DAS28). In the second paper (paper II) not only MBDA score at a certain time-point, but also the dynamic of it from baseline to 3 or 12 months and from 3 to 12 months was assessed for prediction of RP from baseline to 1 or 2 years and from 1 to 2 years.

3.3 STUDY II

In the Study II, which was published in paper III, the association of the MBDA score at 3 months with second-line treatment outcome at 1 year was assessed. As treatment outcome measures, low disease activity and EULAR good response were used. For patients with missing clinical data at 1 year, last observation carried forward (LOCF) was applied. In this study 157 patients who failed to achieve LDA on MTX monotherapy were included. Of the 157 patients, 75 were randomised to conventional TT and 82, to IFX therapy. Apart from validated categories for the MBDA score (low, moderate and high), new cut-offs were also applied based on receiver operating characteristic (ROC) curve analysis for dichotomisation of patients into high and low MBDA score categories. The proportion of patients achieving LDA or EULAR good response between groups with low versus high MBDA categories were compared for each therapy group separately and then the results were compared between the therapy groups. As reference predictors CRP, ESR and DAS28 at 3 months were compared.

3.4 STUDY III

In the Study III protein biomarkers at baseline were analysed for association with treatment outcome (LDA and EULAR good response) at 3 months in RA patients treated with MTX monotherapy.

3.4.1 Study IIIa

Here we investigated the MBDA score and comprising it 12 individual biomarkers at baseline as predictors of achievement of LDA or EULAR good response at 3 months. Two hundred and ninety-eight patients were included of whom, 104 achieved LDA and 101, EULAR good response. Four biomarkers that were significantly different between patients who achieved and did not achieve LDA. They were analysed by ROC curve analysis for dichotomisation into low and high categories. Then the proportion of patients achieving treatment outcome was assessed. The four biomarkers were also combined into a *combined biomarker score* resulting in a scale between 0 and 4 and indicating respectively, the association with lowest to highest risk for not achieving the treatment outcome. The combined biomarker score was also added to a predictive matrix together with previously established demographic predictors such as sex and age.

3.4.2 Study IIIb

In this study 177 target proteins were assessed at baseline in 135 selected RA patients as predictors of outcome of MTX monotherapy at 3 months. In univariate analyses, eight proteins differed between patients who achieved (n=50) and did not achieve (n=85) LDA at a level of p-value <0.001. In a multivariate analysis, only two were significantly different. These two proteins were analysed by ROC curve and were dichotomised into low and high categories with subsequent comparison of proportion of patients achieving treatment outcomes. The two proteins were combined then generating four categories: low/low, low/high, high/low and high/high, and the proportions of patients achieving treatment outcome were assessed between these categories. In this study, an attempt of reproducing the results for one of the proteins was done in another RA cohort (COMBINE, N=74).

3.5 STUDY IV

In the Study IV, 101 patients who failed MTX therapy and were randomised to MTX+IFX treatment were included for investigation of serum level of IFX (sIFX), anti-drug antibodies (ADA) and their relation with treatment outcome (LDA or remission), as well as to identify baseline predictors of low sIFX level and ADA development. The serum samples were analysed at 3 (missing n=8), 9 (missing n=6) and 21 months after initiation of IFX treatment. All 289 serum samples were analysed for sIFX level, but only samples with sIFX concentration <0.2 μ g/ml (n=64) were analysed for ADA-positivity (Figure 3).



Figure 3. Schematic illustration of serum samples analyses for IFX and ADA in 101 patients from the SWEFOT trial

Patients who were ADA-positive at least ones during the follow-up, were considered as ever ADA-positive and their baseline parameters were compared with parameters of never ADA-positive patients.

3.6 SERUM SAMPLE ANALYSES

Serum samples for all study projects were analysed out of our facility by different laboratory personals.

3.6.1 Analyses of 12 proteins and the MBDA score generation

The serum samples were shipped to Crescendo Bioscience, South San-Francisco, CA, USA for the analysis. The individual 12 biomarkers were analysed by an electrochemiluminescence-based multiplex immunoassay on the Meso Scale Discovery Multi-Array platform. The MBDA score was calculated based on concentration of the 12 biomarkers using Vectra® DA trained algorithm, which is a patent of Crescendo Bioscience and is undisclosed.

3.6.2 Analyses of serum proteins for the affinity proteomics project

Serum samples were sent to Science of Life Laboratory (SciLifeLab) for the analysis of 177 proteins, which was done in Prof. Peter Nilsson's lab. The 177 target proteins were selected based on previous studies in inflammatory and autoimmune diseases. These proteins were analysed using 380 antibodies from Human Protein Atlas. In addition four controls were also used: anti-human IgG and anti-albumin as positive controls, and rabbit IgG and beads without any proteins as negative controls. The levels of the proteins were expressed in median fluorescent intensity (MFI).

3.6.3 Analysis of sIFX levels and ADA

Both sIFX and ADA were measured using in-house validated ELISA methods that are used at Swedish University Hospitals. For sIFX, plates coated with TNF were used. For detection of ADA, only samples with <0.2 μ g/ml sIFX levels were analysed, since IFX interferes with the analyts and gives false-positive results. For detection of ADA, competitive ELISA was used: the analyte, which is also IFX, is incubated with TNF-coated plate followed by adding serum, whose unbound ADA competes and displaces with TNF and binds to the analyte.

3.7 STATISTICAL ANALYSES

For non-parametric variables Mann-Whitney U test was used for all study projects. In all projects also proportion of patients were compared between two different groups, for which Chi-squared or Fisher's Exact test were used. In Study III, the homogeneity of odds ratio was tested by Breslow-Day test. For prediction analyses, uni- and multivariate logistic regression analyses were used (Study I, III and IV). In Study IIIb, of 380 variables (for 177 proteins), those with p-value <0.001 in the univariate analyses were included in multivariate logistic regression model. For cut-of determination, ROC-curve analysis was applied in Studies II and III. The cut-of levels were based on values corresponding to the highest sum of sensitivity and specificity.

4 RESULTS AND DISCUSSION

4.1 STUDY I (PAPERS I AND II)

In Study I, we investigated the MBDA score as a predictor of RP. The selected patients for this study did not differ significantly by their baseline parameters from the entire SWEFOT population. As it was expected, the MBDA score correlated with disease activity markers such as DAS28 and CRP (Figure 4).



Figure 4. Correlation of the MBDA score with DAS28 (A) and CRP (B) at baseline.

4.1.1 Discordance between the MBDA score and other disease activity markers

The categories of these markers, however, had some discrepancies (Figure 5). For example, of 235 patients 5, 29 and 201 had low, moderate and high MBDA scores, however, of 71 patients with low CRP 42 had high and 24 had moderate MBDA scores and about quarter of patients with low CRP but high MBDA had RP at 1 year (Figure 5C). Patients with low MBDA score (n=5), on the other hand, had low CRP and none of them progressed. No patient had low DAS28 at baseline, since it was an exclusion criteria for the SWEFOT trial. In total, low/moderate versus high categories of the MBDA score at baseline discriminated patients at very low versus high risk of subsequent RP at 1 year (3% and 21%, respectively, p=0.012; Figure 5D).

Baseline DAS28-ESR Baseline DAS28-CRP Low Moderate High Low Moderate High **Baseline MBDA** (<2.7) (>2.7 - 4.1) (>4.1) Total Baseline MBDA (≤3.2) (>3.2 - 5.1) (>5.1) Total 0 0/5 Low (<30) 0/1 0/4 0 Low (<30) 0/3 0/2 0/5 Moderate (30 - 44) 0/15 0 1/14 1/29 Moderate (30 - 44) 0 1/5 0/24 1/29 High (>44) 0 9/51 33/150 42/201 High (>44) 2/16 40/185 42/201 0 Total 0 10/68 33/167 43/235 Total 0 3/22 40/213 43/235

в



Figure 5. Cross-tabulation of all analysed patients and subset with rapid radiographic progression over one year, by baseline disease activity measures. The denominator in each cell represents the number of patients cross-classified by baseline MBDA and DAS28-ESR (A), baseline MBDA and DAS28-CRP (B) and baseline MBDA and CRP (C) disease activity scores. The numerator in each cell represents the number of patients with radiographic progression at one year. Figure D illustrates radiographic progressors for MBDA low, moderate and high score groups (%). Radiographic progression at one year is defined by increase in SHS > 5 compared to baseline.

4.1.2 The MBDA score in prediction of RP

In bivariate analyses, the MBDA score was independently of other risk factors such as RF, ACPA and sex, predictive for radiographic outcome of the joint damage during 1 year of follow-up (Table 5). In multivariate logistic regression analysis, high MBDA score at baseline compared with low/moderate, was 3.9 times more associated with higher risk of RA at 1 year (adjusted for sex, symptom duration, baseline erosions, current smoking and HAQ score). This was similar with results from the BeSt study, however, for a univariate analysis (for high

A

MBDA score compared with low/moderate at baseline, RR=3.7) (293). Few more studies, including the BeSt study illustrated predictive ability of the MBDA score for RP (292-295), even though two of these studies (293, 294) used slightly different cut-offs for RP (Δ SHS>0 or >3).

	Odds Ratio ⁺	95% CI	P-Value [‡]
Univariate Analyses:			
Baseline MBDA score	1.05	(1.02, 1.08)	< 0.001
Baseline DAS28-ESR	1.31	(0.94, 1.81)	0.107
Baseline DAS28-CRP	1.22	(0.88, 1.71)	0.237
Baseline CRP (mg/L)	1.10	(1.02, 1.18)	0.018
Bivariate Models:			
Baseline MBDA adjusted for DAS28-ESR	1.05	(1.02, 1.08)	< 0.001
Baseline MBDA adjusted for DAS28-CRP	1.05	(1.02, 1.09)	< 0.001
Baseline MBDA adjusted for CRP	1.06	(1.02, 1.10)	0.002
Baseline MBDA adjusted for ESR	1.04	(1.01, 1.07)	0.021
Baseline MBDA adjusted for Rheumatoid Factor	1.05	(1.02, 1.08)	< 0.001
Baseline MBDA adjusted for CCP Status	1.05	(1.03, 1.08)	< 0.001
Baseline MBDA adjusted for Total Swollen Joint Count	1.05	(1.02, 1.08)	< 0.001
Baseline MBDA adjusted for Total Tender Joint Count	1.05	(1.02, 1.08)	< 0.001
Baseline MBDA adjusted for Global Assessment of Disease Activity	1.05	(1.02, 1.08)	< 0.001
Baseline MBDA adjusted for SHS	1.05	(1.02, 1.08)	< 0.001
Baseline MBDA adjusted for Symptom Duration	1.05	(1.02, 1.08)	< 0.001
Baseline MBDA adjusted for Sex	1.05	(1.02, 1.08)	<0.001
Multivariate Model*:			
Baseline MBDA adjusted for Sex, Symptom Duration, baseline erosions, current smoking status, HAQ score	1.05	(1.02, 1.08)	< 0.001
High (>44) baseline MBDA score adjusted for Sex. Symptom Duration, baseline erosions, current smoking status, HAQ score	3.86	(1.04.14.26)	0.04

† The odds ratio was estimated from a logistic regression model. The logistic model is estimating the probability of radiographic progression at year 1.

For the univariate model, the odds of radiographic progression increases by 5% for every one-unit increase in the baseline MBDA score. When accounting for other disease activity measures individually (bivariate models), the odds of radiographic progression increase in a cumulative manner, approximately 4-6% for every one-unit increase in the baseline MBDA score.

‡ P-value was calculated using Wald's chi-square test.
*Multivariate model adjusted for significant univariate predictors of one year radiographic progression (n=207), as in Saevarsdottir S et al, Currentsmoking status is a strong predictor of radiographic progression in early rheumatoid arthritis: results from the SWEFOT trial, ARD in press.

4.1.3 Association of the MBDA score at follow-ups with subsequent RP

In the paper II, we demonstrated that the MBDA score at follow-ups was also informative for RP during subsequent year (Figure 6). None of the patients achieving low MBDA score from moderate (n=11) and only one patient of those who dropped from high category to low (n=17)during 3 months of MTX monotherapy, had RP at 1 year. In contrast, patients who were persistently high at 3 months (n=88), 25% had RP. Similar associations were observed when looking at conversion of the MBDA category from baseline to 1 year and RP during the second year of the follow-up (Figure 6B). CRP and ESR in contrast, could not perform the identification or risk for RP similarly well.



Figure 6. Association of radiographic progression with change in disease activity categories according to the MBDA score, CRP and ESR. Change in categories according to MBDA score, CRP or ESR from BL to Month 3 and radiographic progression from BL to Year 1 (A); and change in categories according to MBDA score, CRP or ESR from BL to Year 1 and radiographic progression from Year 1 to Year 2 (B).

In the second paper, we also observed differential association of the MBDA score at baseline or 3 months with RP at 2 years, for each of the randomised treatment arm. Among patients with high MBDA score, those on TT had significantly higher proportion of RP compared with patients on biological therapy (Figure 7A: 45% vs 25% and 57% vs 32%, respectively; p<0.05 for both comparisons).



Figure 7. Radiographic progression among 3 therapy groups (triple therapy group, anti-TNF treatment group and MTX-responders) stratified by MBDA categories at multiple time-points. Proportion of patients with radiographic progression defined as Δ SHS>5 (A) and Δ SHS>3 (B). Left and middle bar-graphs represent proportion of patients with 2-year radiographic progression stratified by the MBDA score at BL and Month 3, respectively. Right bar-graph shows radiographic progression from Year 1 to Year 2 among patients stratified by the MBDA score at Year 1.

4.2 STUDY II (PAPER III)

4.2.1 Association of validated categories of the MBDA score with treatment outcome

It has been shown previously that of patients randomised to MTX+IFX therapy, significantly more proportion achieved EULAR good response than from patients on TT (39% vs 25%, respectively; p=0.016) (96). Even being statistically significant, the difference is not very large from clinical point of view. In MTX-incomplete responders, the MBDA score categories at the time of treatment randomisation yielded clinically meaningful difference and moreover, it identified a subgroup that had a higher chance of achievement of LDA on TT compared with patients on IFX (Figure 8). Namely, of patients with low MBDA score, more patients

achieved LDA if treated with TT than with IFX (88% vs 18%, p=0.006) and reverse was true for patients with high MBDA score (35% vs 58%, p=0.040).



Figure 8. Proportion of patients with response (DAS28≤3.2) to second-line therapy at Year 1 stratified by conventional cut-offs of MBDA score at the start of treatment intensification. Proportion of responders at Year 1 to triple therapy (grey bars) or anti-TNF therapy (white bars) stratified by low (<30), moderate (30-44), and high (>44) MBDA categories at Month 3.

4.2.2 New threshold of the MBDA score for prediction of treatment outcome

ROC curve analysis yielded a cut-off of 38 for dichotomisation of patients into higher and lower categories, which illustrated similar associations (79% vs 44%, respectively, p=0.019 and 36% vs 58%, respectively, p=0.018; Figure 9). Other inflammatory markers (CRP, ESR and DAS28) at 3 months did not illustrate or showed a weaker association.

Our findings support the results from the RACAT trial demonstrating that of patients not responding to MTX+etanercept, some switchers to TT responded, while some of non-responders on TT who switched to MTX+etanercept also achieved better clinical outcome (94). The MBDA score compared with DAS28, ESR and CRP is a multi-component molecular marker which probably indicates changes much earlier than the clinical or mono-

component molecular markers can do. Thus, validation of these results could lead to reduction of unnecessary use of biological drugs for a subgroup of patients, while indicator for their successful use for another subset of patients.



Proportion of responders (DAS28≤3.2) at Year 1

Figure 9. Proportion of clinical responders (DAS28<3.2) to second-line therapy at Year 1 stratified by ROC-based cut-offs of disease activity measures at Month 3. Proportion of responders at Year 1 to triple therapy (grey bars) or anti-TNF therapy (white bars) stratified by MBDA score (A), CRP (B), ESR (C) and DAS28 (D). The overall p-values for 4 groups are calculated by Breslow-Day test, and for triple therapy versus anti-TNF therapy groups – by χ^2 test, unless otherwise stated. †p-value was calculated using Fisher's Exact test.

4.3 STUDY III (PAPER IV AND V)

In early RA patients treated with MTX, baseline serum levels of biomarkers were analysed for associations with achievement of treatment outcome (LDA or EULAR good response) at 3 months follow-up.

4.3.1 Study IIIa (paper IV)

In the paper IV, the 12 component proteins of the MBDA score were analysed as potential predictors of response to MTX.

4.3.1.1 Identification of potential predictors

Of 298 analysed patients 104 achieved LDA. Among baseline parameters, as it has been previously published (98), sex, age and some clinical parameters were significantly different between patients achieving and not achieving LDA at 3 months (Table 6).

Baseline Characteristics	Stratified by disease activity (DAS28) at 3 months follow-up			
Median (±IQR)	DAS28 ≤ 3.2 (N=104) ^a	DAS28 > 3.2 (N=194) ^b	P-value	
Female: N (%)	60 (58)	154 (79)	< 0.001	
Age: years	61 (51-69)	55 (44-62)	0.001	
Smoking: N (%)	19 (24)	47 (26)	0.780	
Symptom Duration: months	5 (3-8)	5 (4-8)	0.283	
Anti-CCP Positive: N (%)	63 (64)	107 (58)	0.292	
RF Positive: N (%)	73 (71)	124 (65)	0.274	
Prednisolone use: N (%)	15 (14)	20 (10)	0.293	
28 Swollen Joint Count	9 (7-13)	11 (7-15)	0.087	
28 Tender Joint Count	6 (4-10)	10 (6-15)	< 0.001	
ESR (mm/h)	28 (15-42)	36 (24-62)	0.001	
CRP (mg/L)	15 (9-33)	20 (9-53)	0.033	
PatG (VAS 0-100mm)	50 (29-67)	63 (43-77)	0.001	
Pain (VAS 0-100mm)	49 (33-64)	62 (46-74)	< 0.001	
HAQ	1.0 (0.50-1.38)	1.25 (0.88-1.75)	< 0.001	
DAS28	5.2 (4.6-5.9)	5.9 (5.3-6.4)	< 0.001	

Table 6. Baseline characteristics of early RA patients from the SWEFOT trial

IQR – interquartile range, anti-CCP - anti-cyclic citrullinated peptide, RF – rheumatoid factor, ESR – erythrocyte sedimentation rate, CRP – Creactive protein, PatG – Patient's Global Assessment of Disease Activity score, VAS – visual analogue scale, HAQ, - health assessment questionnaire, DAS – disease activity score.

[°] Missing patients from DAS28≤3.2 column: Smoking (n=25), Anti-CCP (n=6) and RF (n=1).

 $^{\circ}$ Missing patients from DAS28>3.2 column: Smoking (n=11), Anti-CCP (n=9), RF, PatG and Pain (n=2), HAQ (n=4) and DAS28 (n=3).

The MBDA score did not differ between these 2 groups of patients. In CAMERA trial, on the other hand, the MBDA score was associated with DAS28-CRP and discriminated patients between remission/low and moderate/high disease activity levels (287). In a univariate analysis, four of the 12 proteins differed at a level of p<0.05 (Table 7).

Baseline Biomarkers	Stratified by disease activity (DAS28) at 3 months follow-up			
Median (±IQR)	DAS28 ≤ 3.2 (N=104) [°]	DAS28 > 3.2 (N=194) ^b	P-value	
VCAM-1 (mg/L)	0.70 (0.60-0.86)	0.64 (0.56-0.77)	0.005	
TNF-RI (μg/L)	1.9 (1.6-2.4)	1.7 (1.4-2.2)	0.005	
IL-6 (ng/L)	49 (22-97)	67 (29-150)	0.044	
CRP (mg/L)	20 (8-43)	27 (10-110)	0.045	
MMP-1 (μg/L)	9.35 (5.87-15)	11 (6.48-18)	0.067	
MMP-3 (μg/L)	48 (28-82)	56 (29-123)	0.105	
Leptin (µg/L)	9.25 (3.58-14)	10.50 (4.60-20.25)	0.108	
VEFG (ng/L)	405 (265-600)	390 (250-590)	0.719	
Resistin (µg/L)	6.60 (5.35-8.78)	6.80 (5.10-8.68)	0.857	
YLK-40 (μg/L)	85 (54-120)	79 (57-130)	0.862	
EGF (ng/L)	160 (103-258)	170 (100-253)	0.903	
SAA (mg/L)	20 (7-46)	17 (4-73)	0.990	
MBDA score	59 (47-66)	59 (49-74)	0.143	

Table 7. Baseline biomarkers of early RA patients from the SWEFOT trial

IQR – interquartile range, DAS – disease activity score, VCAM-1 - vascular cell adhesion molecule-1, TNF-RI - tumour nectosis factor receptor-1, IL-6 - interleukin-6, CRP – C-reactive protein, MMP - matrix metalloproteinase, VEGF - vascular endothelial growth factor, YKL-40 - human cartilage glycoprotein-39, EGF - epidermal growth factor, SAA - serum amyloid A, MBDA – multi-biomarker disease activity.

Biomarkers that had p-value <0.2 (n=7), were entered into a multivariate logistic regression model for identification of LDA at 3 months as a dependent outcome. Four of these biomarkers were independently associated with LDA (increase in concentrations of VCAM-1 and TNF-RI, as well as decrease in CRP and leptin were associated with LDA, Table 8). There is no clear data regarding these four biomarkers illustrating predictive capacity for response to MTX. Controversial results are published by different researchers regarding CRP, some illustrating association of its low level with response (110) and some failing to do so (114, 318-320). The association of high baseline leptin levels with high DAS28 at 6 months in RA patients treated with non-biological DMARDs was illustrated by a research group (262). Being associated with BMI, we could not find any relation of BMI with treatment outcome in our study (data not shown). Increased sTNF-RI might indicate increased sTNF, since cleavage of membrane-bond TNF-RI is considered as negative regulation of increased production of TNF and inflammation (decoy receptor) (200, 201). Considering this fact and since MTX is known to decrease TNF level (321, 322), higher levels of TNF-RI could mean a disease

driven by TNF, where MTX could be a proper option for treatment. Indeed, a study has demonstrated that patients with high sTNF-RI had better response to MTX (161).

baseline for prediction of low DAS28 at 3 months			
Biomarkers	OR	95% CI	
VCAM-1 (per mg/L increase)	6.70	1.17-38.62	
TNF-RI (per μg/L increase)	3.10	1.70-5.71	
IL-6 (per ng/L increase)	0.99	0.99-1.00	
CRP (per mg/L increase)	0.99	0.98-0.99	
MMP-1 (per μg/L increase)	0.98	0.94-1.01	
MMP-3 (per μg/L increase)	1.00	0.99-1.00	
Leptin (per µg/L increase)	0.97	0.95-0.99	
Results after step-wise logistic regression analysis			
VCAM-1 (per mg/L increase)	8.19	1.47-45.65	
TNF-RI (per μg/L increase)	2.48	1.43-4.31	
CRP (per mg/L increase)	0.99	0.98-0.99	
Leptin (per μg/L increase)	0.97	0.95-0.99	

Table 8. Multivariate logistic regression of protein biomarkers at baseline for prediction of low DAS28 at 3 months

VCAM - vascular cell adhesion molecule, TNF-RI - tumour necrosis factor receptor 1, IL-6 - interleukin-6, CRP - C-reactive protein, MMP - matrix metalloproteinase, OR - odds ratio, CI - confidence interval

4.3.1.2 Dichotomisation of biomarker levels

ROC curve analyses identified cut-off levels based on highest sum of sensitivity and specificity generating low and high categories for these biomarkers: CRP: \leq 51.5 and >51.5 mg/ml, leptin: \leq 14.5 and >14.5 µg/ml, TNF-RI: \leq 1.75 and >1.75 µg/ml and VCAM-1: \leq 0.605 and >0.605 mg/ml. There were significantly higher proportions of patients achieving LDA among subjects with low CRP and leptin, and high TNF-RI and VCAM-1 (Figure 10).



Figure 10. Proportions of eRA patients achieving low disease activity after 3 months of MTX monotherapy, stratified for different biomarkers. Proportions of patients with low DAS28 in patients dichotomised according to CRP (A), leptin (B), TNF-RI (C) and VCAM-1 (D). DAS28 – disease activity score based on 28 joints, CRP – C-reactive protein, DAS28 – 28-joint disease activity score, MTX – methotrexate, TNF-RI – tumour necrosis factor receptor-1, VCAM-1 – vascular cell adhesion molecule-1.

4.3.1.3 Combined biomarker score

The combined biomarker score that was based on the four proteins, was independently associated with treatment outcome (odds ratio adjusted for RF, ACPA, sex, age and current smoking OR=0.44, 95CI=0.30-0.65). There was a gradual decrease in proportion of patients in LDA or EULAR good response with increase of the combined biomarker score (Figure 11).



Figure 11. Proportions of early RA patients achieving low DAS28 or EULAR good response after 3 months of MTX monotherapy, stratified for the combined biomarker score. Proportions of patients with low DAS28 (A) or EULAR good response (B) within subsets based on the combined biomarker score: combined score = 0 (green bars), combined score = 1 (blue bars), combined score = 2 (orange bars), combined score = 3 (red bars) and combined score = 4 (black bars).

The biomarker score was tested in combination with previously published (98) predictors (sex and age) for identification of patients achieving treatment outcome. Even though one of the components of the combined biomarker score has a co-linearity with sex (females have higher leptin level compared with males), the combined prediction matrix identified subset of patients with very high predictability of response. For example, among older male patients with the lowest combined biomarker score (n=14), 13 achieved LDA and 12 EULAR good response, while among younger female patients with the combined biomarker score of 3 and 4 (n=22), only one achieved LDA or EULAR good response (Figure 12).



Figure 12. Matrix of prediction of response to MTX in early RA patients, based on age, gender and combined biomarker score. Proportion of patients achieving low DAS28 (A) or EULAR good response (B).

The main limitation of the study is the limited number of patients in some subgroups which emphasises the need for validation in other study populations. The SWEFOT trial, on the other hand, is designed with low bias, due to the few exclusion criteria and routine-care based recruitment, and all patients were treated with MTX, which is the recomendet first-line therapy today.

4.3.2 Study IIIb (paper V)

4.3.2.1 Identification of predictive proteins

The 135 patients included in this study had more often RF than the remaining patients from the SWEFOT population (72% and 58%, respectively, p=0.002). Of screened 177 serum proteins at baseline, in univariate analysis, eight had different level of MFI (at the level of p<0.001) among patients achieving and not achieving LDA at 3 months. Of the eight candidate biomarkers, two: MMP-7 and α -chain of fibrinogen (FGA) were independently associated with treatment outcome. The ROC curve analysis resulted in area under the curve of 0.692 and 0.699 for MMP-7 and FGA, respectively (p<0.001, Figure 13A). Low levels of the two biomarkers indicated significantly higher proportion of patients in LDA at 3 months (Figure 13B).



Figure 13. Assessment of baseline levels of MMP-7 and FGA as predictors of LDA at 3 months. Receiver operating characteristic curve analysis and area under the curve (A) of MMP-7 (green line), FGA (red line); Proportion of patients achieving low DAS28 at 3 months among groups dichotomised by MMP-7 or FGA (B) low levels – purple bars and high levels – yellow bars.

When considering both biomarkers simultaneously, of patients with low levels for both biomarkers, 79% had LDA and EULAR good response, while of those with high levels for both biomarkers, 18% and 15%, respectively achieved LDA or EULAR good response (p<0.001, Figure 14).



Figure 14. Assessment of combined baseline levels of MMP-7 and FGA as predictors of outcome. Proportion of patients in LDA (A) and EULAR good response (B) among patients with low/low (purple bar), high/high (yellow bar), low/high (green bar) and high/low (blue bar) categories of MMP-7/FGA.

4.3.2.2 An attempt of replication of the results for MMP-7 in the COMBINE cohort

Neither MMP-7 nor FGA have been shown to predict response to MTX. Validation of results for MMP-7 in another RA cohort (COMBINE, N=74) did not confirm the finding. We found three differences between the SWEFOT and COMBINE cohorts: method of measuring MMP-7, significantly lower glucocorticoid use and significantly higher DAS28 at baseline and at 3 months among patients from SWEFOT compared with COMBINE cohort. When considering only patients treated without glucocorticoids, we still could not confirm the association of baseline low MMP-7 with LDA at 3 months in COMBINE cohort. Therefore, we assume that method of measurement of MMP-7 and baseline DAS28 could contribute to the bias.

Confirmation of these results in another similar RA cohort with untreated early RA patients using the same method of measurement, might increase efficiency of MTX monotherapy choice in treatment of RA.

4.4 STUDY IV (PAPER VI)

4.4.1 Distribution of data

Immunogenicity of biological medications has been established as a challenge for achievement of optimal treatment outcome (310, 323). Of 289 available serum samples (from all 3 time-points together), we observed a very low sIFX level ($<0.2 \mu g/ml$) in 64 samples, and of those, 47 (73.4%) were ADA-positive. Of 101 patients analysed in this study, 34 patients were ever ADA-positive. There were no baseline parameters significantly different between ever and never ADA-positive patients. However, there were more women among ever (85%) compared with never (67%) ADA-positive patients, respectively (p=0.052, Table 9) with similar trend for RF-positivity (79% vs 62%, p=0.0079).

Patient Characteris,	MTX-incomplete responders randomized to IFX therapy (N=101)			
Median (IQR) ^a	All (N=101) ⁶	Ever ADA-positive (N=34) [°]	Never ADA- positive (N=67) ^d	P-value [®]
Female: N (%)	74 (73)	29 (85)	45 (67)	0.052
Age (years)	55 (43-62)	50 (42-59)	55 (44-63)	0.246
BMI	24.2 (22.2-26.4)	23.8 (22.2-27.0)	24.4 (22.3-26.2)	0.740
Current Smokers: N (%)	27 (27)	10 (29)	17 (25)	0.665
Symptom Duration (months)	6 (4-8)	6 (3-9)	5 (4-8)	0.610
Anti-CCP Positive: N (%)	62 (67)	22 (71)	40 (64)	0.534
RF Positive: N (%)	68 (68)	27 (79)	41 (62)	0.079
28 Swollen Joint Count	6 (3-10)	4 (3-9)	6(3-10)	0.483
28 Tender Joint Count	6 (4-10)	7 (4-12)	6 (3-9)	0.134
ESR (mm/h)	23 (12-37)	24 (14-37)	21 (11-34)	0.446
CRP Level (mg/L)	9 (4-17)	9 (4-15)	9 (4-18)	0.853
PatG (VAS 0-100mm) Score	49 (35-67)	50 (36-67)	49 (34-68)	0.979
Pain VAS	47 (30-64)	44 (30-66)	49 (30-63)	0.740
PhysG	2 (2-2)	2 (2-2)	2 (2-2)	0.392
HAQ	0.9 (0.63-1.38)	0.9 (0.38-1.25)	1.0 (0.63-1.38)	0.081
DAS28	4.9 (4.1-5.6)	5.0 (4.3-5.8)	4.8 (4.1-5.5)	0.328

 Table 9. Characteristics of SWEFOT participants at the time of randomization to IFX stratified into ever and never anti-drug antibody positive patients

ADA – anti-drug antibody, IQR – interquartile range, IFX – infliximan, BMI – body mass index, anti-CCP - anti-cyclic citrullinated peptide, RF – rheumatoid factor, ESR – erythrocyte sedimentation rate, CRP – C-reactive protein, PatG – Patient's Global Assessment of Disease Activity, VAS – visual analogue scale, PhysG – Physician Global Assessment of Disease Activity, HAQ – health assessment questionnaire, DAS – disease activity score.

a Data for Female, Age, Current Smoking, Symptom Duration, Anti-CCP and RF are presented from diagnosis and trial recruitment date.

b Number of missing patients for "101" column: BMI (n=27), Anti-CCP (n=8), RF (n=1), 28 Swollen Joint Count, 28 Tender Joint Count, CRP, PatG VAS (n=3), ESR, Pain VAS, PhysG and HAQ (n=4).

c Number of missing patients for IFX-Responders column: BMI (n=7), Anti-CCP (n=3), 28 Swollen Joint Count, 28 Tender Joint Count, ESR, CRP, PatG VAS, Pain VAS, PhysG and HAQ (n=1). d Number of missing patients for IFX-non-responders column: BMI (n=20), RF (n=1), Anti-CCP (n=5), 28 Tender Joint Count, 28 Swollen Joint Count, CRP and PatG VAS (n=2), ESR, Pain VAS, PhysG and HAQ (n=3). e Comparing ever and never ADA-positive patients.

4.4.2 Association of sIFX levels and ADA status with treatment outcome

Higher proportion of patients in LDA or remission was observed among patients with sIFX $>0.2 \mu g/ml$ compared with ADA-positive patients, and the difference was significant at the study cessation (at 21 months, Figure 15A and B). Similarly higher sIFX level was associated with better clinical outcome (Figure 15C and D). These results are in concordance with previous studies (303, 305, 310, 324).





4.4.3 Prediction of low sIFX or ADA

Gender and RF status among other baseline parameters showed borderline significnat associations with ever ADA-positivity. In multivariate analyses, female gender and RF-positivity remained borderline significantly associated with development of ADA. Higher frequency of ADA-positivity among women compared with men could be explained by the fact that B cells in females have higher capacity for antibody production (325). However, there are controversial data regarding associations of RF and gender with ADA status (324, 326). Unlike SWEFOT, patients from these studies have significantly higher disease duration (6-14 years) and lower incidence of ADA-positive patients, which indicates more suppressed immune system by long-lasting DMARD therapy, leading to reduced capacity of antibody production. In addition, RF may interfere with ELISA kit for ADA measurement and give false-positive results. However, we also observed a trend of RF-positivity with low sIFX level, and the trend became stronger at 21 months (34% vs 16%, p=0.059, Figure 16). Similar trend was observed when comparing female versus male patients with significant difference at 21 months (35% vs 7%, respectively, p=0.006, Figure 16).





It is important to note that sIFX levels analysed in this study were from samples taken at follow-up visits and not just before next infusion (i.e. trough levels), which is the biggest limitation. However, confirmation of associations of the sIFX levels and ADA-positivity with treatment outcome at later time-point, found by others researchers at earlier time-points indicates that the trend associations observed in our study could be strengthened if trough levels are used. Therefore, further investigations of prediction of response to IFX using sIFX levels, ADA and other baseline parameters might help identify patients at higher risk and improve decision-making for the switch of biological therapy.

5 CONCLUSION

In the presence of different treatment options for RA and the heterogeneity of the disease, there is a huge need for predictive tools to help chose the optimal treatment for each individual patient. This thesis project overall, tried to address this question via exploratory analyses of serum proteins, as potential predictors of treatment outcome.

In paper I and II we showed predictive capacity of the MBDA score at baseline and followups for RP during subsequent one or two years. Apart from confirming the association of low MBDA score with very low risk of RP and superiority of the MBDA score compared to CRP, ESR and DAS28, we also showed for the first time that patients with high MBDA score would benefit more from MTX+IFX therapy than from TT to lower RP.

In paper III, the MBDA score identified a subset of patients that benefited significantly more from TT compared with biological IFX treatment, a finding that yielded much attention since TT is much cheaper, and has now been supported by results from O'Dell et al (94).

Paper IV and V highlighted some protein biomarkers at baseline for prediction of response to MTX monotherapy. As in all biomarker studies, those findings need to be validated since these molecules can be potential key players in the pathology of some RA patients.

In paper VI we confirmed previously published results on association of low sIFX levels and ADA-positivity with poorer treatment response to IFX, but also found that RF and female gender might be risk factors for immunogenicity and low sIFX levels.

In summary, through investigations of serum proteins related to inflammation we identified potential predictors of RP and clinical outcome, which may help to understand pathology behind RA and aid therapy choice. For biological treatment, studies of immunogenicity and blood trough level of the drug support routine monitoring of sIFX and ADA in the clinic and serve as basis for development of an algorithm when to switch to other treatment options.

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7 REFERENCES

- Smolen JS, Aletaha D, McInnes IB. Rheumatoid arthritis. Lancet. 2016 Oct 22;388(10055):2023-38.
- Biver E, Beague V, Verloop D, Mollet D, Lajugie D, Baudens G, et al. Low and stable prevalence of rheumatoid arthritis in northern France. Joint, bone, spine : revue du rhumatisme. 2009 Oct;76(5):497-500.
- 3. Li R, Sun J, Ren LM, Wang HY, Liu WH, Zhang XW, et al. Epidemiology of eight common rheumatic diseases in China: a large-scale cross-sectional survey in Beijing. Rheumatology. 2012 Apr;51(4):721-9. PubMed
- 4. Myasoedova E, Crowson CS, Turesson C, Gabriel SE, Matteson EL. Incidence of extraarticular rheumatoid arthritis in Olmsted County, Minnesota, in 1995-2007 versus 1985-1994: a population-based study. The Journal of rheumatology. 2011 Jun;38(6):983-9.
- Klareskog L, Stolt P, Lundberg K, Kallberg H, Bengtsson C, Grunewald J, et al. A new model for an etiology of rheumatoid arthritis: smoking may trigger HLA-DR (shared epitope)-restricted immune reactions to autoantigens modified by citrullination. Arthritis and rheumatism. 2006 Jan;54(1):38-46.
- 6. McInnes IB, Schett G. The pathogenesis of rheumatoid arthritis. The New England journal of medicine. 2011 Dec 8;365(23):2205-19.
- 7. Klareskog L, Catrina AI, Paget S. Rheumatoid arthritis. Lancet. 2009 Feb 21;373(9664):659-72.
- 8. Malmstrom V, Catrina AI, Klareskog L. The immunopathogenesis of seropositive rheumatoid arthritis: from triggering to targeting. Nature reviews Immunology. 2017 Jan;17(1):60-75.
- Catrina AI, Svensson CI, Malmstrom V, Schett G, Klareskog L. Mechanisms leading from systemic autoimmunity to joint-specific disease in rheumatoid arthritis. Nature reviews Rheumatology. 2017 Feb;13(2):79-86.
- Klimiuk PA, Goronzy JJ, Bjor nsson J, Beckenbaugh RD, Weyand CM. Tissue cytokine patterns distinguish variants of rheumatoid synovitis. The American journal of pathology. 1997 Nov;151(5):1311-9.
- 11. Yanni G, Whelan A, Feighery C, Quinlan W, Symons J, Duff G, et al. Contrasting levels of in vitro cytokine production by rheumatoid synovial tissues demonstrating different patterns of mononuclear cell infiltration. Clinical and experimental immunology. 1993 Sep;93(3):387-95.
- 12. Young CL, Adamson TC, 3rd, Vaughan JH, Fox RI. Immunohistologic characterization of synovial membrane lymphocytes in rheumatoid arthritis. Arthritis and rheumatism. 1984 Jan;27(1):32-9.
- 13. Klimiuk PA, Sierakowski S, Latosiewicz R, Skowronski J, Cylwik JP, Cylwik B, et al. Histological patterns of synovitis and serum chemokines in patients with rheumatoid arthritis. The Journal of rheumatology. 2005 Sep;32(9):1666-72.
- 14. Grassi W, De Angelis R, Lamanna G, Cervini C. The clinical features of rheumatoid arthritis. European journal of radiology. 1998 May;27 Suppl 1:S18-24.
- 15. Lee DM, Weinblatt ME. Rheumatoid arthritis. Lancet. 2001 Sep 15;358(9285):903-11.
- Prevoo ML, van 't Hof MA, Kuper HH, van Leeuwen MA, van de Putte LB, van Riel PL. Modified disease activity scores that include twenty-eight-joint counts. Development and validation in a prospective longitudinal study of patients with rheumatoid arthritis. Arthritis and rheumatism. 1995 Jan;38(1):44-8.
- Inoue E, Yamanaka H, Hara M, Tomatsu T, Kamatani N. Comparison of Disease Activity Score (DAS)28- erythrocyte sedimentation rate and DAS28- C-reactive protein threshold values. Annals of the rheumatic diseases. 2007 Mar;66(3):407-9.
- Matsui T, Kuga Y, Kaneko A, Nishino J, Eto Y, Chiba N, et al. Disease Activity Score 28 (DAS28) using C-reactive protein underestimates disease activity and overestimates EULAR response criteria compared with DAS28 using erythrocyte sedimentation rate in a large observational

cohort of rheumatoid arthritis patients in Japan. Annals of the rheumatic diseases. 2007 Sep;66(9):1221-6.

- 19. Tamhane A, Redden DT, McGwin G, Jr., Brown EE, Westfall AO, Reynolds RJt, et al. Comparison of the disease activity score using erythrocyte sedimentation rate and C-reactive protein in African Americans with rheumatoid arthritis. The Journal of rheumatology. 2013 Nov;40(11):1812-22.
- Fleischmann R, van der Heijde D, Koenig AS, Pedersen R, Szumski A, Marshall L, et al. How much does Disease Activity Score in 28 joints ESR and CRP calculations underestimate disease activity compared with the Simplified Disease Activity Index? Annals of the rheumatic diseases. 2015 Jun;74(6):1132-7.
- 21. Fleischmann RM, van der Heijde D, Gardiner PV, Szumski A, Marshall L, Bananis E. DAS28-CRP and DAS28-ESR cut-offs for high disease activity in rheumatoid arthritis are not interchangeable. RMD open. 2017;3(1):e000382.
- 22. Aletaha D, Smolen J. The Simplified Disease Activity Index (SDAI) and the Clinical Disease Activity Index (CDAI): a review of their usefulness and validity in rheumatoid arthritis. Clinical and experimental rheumatology. 2005 Sep-Oct;23(5 Suppl 39):S100-8.
- 23. van Gestel AM, Prevoo ML, van 't Hof MA, van Rijswijk MH, van de Putte LB, van Riel PL. Development and validation of the European League Against Rheumatism response criteria for rheumatoid arthritis. Comparison with the preliminary American College of Rheumatology and the World Health Organization/International League Against Rheumatism Criteria. Arthritis and rheumatism. 1996 Jan;39(1):34-40.
- 24. van Gestel AM, Haagsma CJ, van Riel PL. Validation of rheumatoid arthritis improvement criteria that include simplified joint counts. Arthritis and rheumatism. 1998 Oct;41(10):1845-50.
- 25. Felson DT, Anderson JJ, Boers M, Bombardier C, Furst D, Goldsmith C, et al. American College of Rheumatology. Preliminary definition of improvement in rheumatoid arthritis. Arthritis and rheumatism. 1995 Jun;38(6):727-35.
- 26. Siegert CE, Vleming LJ, Vandenbroucke JP, Cats A. Measurement of disability in Dutch rheumatoid arthritis patients. Clinical rheumatology. 1984 Sep;3(3):305-9.
- 27. Boini S, Guillemin F. Radiographic scoring methods as outcome measures in rheumatoid arthritis: properties and advantages. Annals of the rheumatic diseases. 2001 Sep;60(9):817-27.
- Sharp JT, Lidsky MD, Collins LC, Moreland J. Methods of scoring the progression of radiologic changes in rheumatoid arthritis. Correlation of radiologic, clinical and laboratory abnormalities. Arthritis and rheumatism. 1971 Nov-Dec;14(6):706-20.
- 29. Sharp JT, Young DY, Bluhm GB, Brook A, Brower AC, Corbett M, et al. How many joints in the hands and wrists should be included in a score of radiologic abnormalities used to assess rheumatoid arthritis? Arthritis and rheumatism. 1985 Dec;28(12):1326-35.
- van der Heijde DM, van Riel PL, Nuver-Zwart IH, Gribnau FW, vad de Putte LB. Effects of hydroxychloroquine and sulphasalazine on progression of joint damage in rheumatoid arthritis. Lancet. 1989 May 13;1(8646):1036-8.
- 31. van der Heijde DM, van Leeuwen MA, van Riel PL, Koster AM, van 't Hof MA, van Rijswijk MH, et al. Biannual radiographic assessments of hands and feet in a three-year prospective followup of patients with early rheumatoid arthritis. Arthritis and rheumatism. 1992 Jan;35(1):26-34.
- 32. van der Heijde D. How to read radiographs according to the Sharp/van der Heijde method. The Journal of rheumatology. 2000 Jan;27(1):261-3.
- 33. Larsen A. A RADIOLOGICAL METHOD FOR GRADING THE SEVERITY OF RHEUMATOID ARTHRITIS ABSTRACT. Scandinavian journal of rheumatology. 1975 1975;4(4):225-33.
- 34. Larsen A. How to apply Larsen score in evaluating radiographs of rheumatoid arthritis in long-term studies. The Journal of rheumatology. 1995 Oct;22(10):1974-5.
- 35. Sudol-Szopinska I, Jans L, Teh J. Rheumatoid arthritis: what do MRI and ultrasound show. Journal of ultrasonography. 2017 Mar;17(68):5-16.
- 36. Boutry N, Morel M, Flipo RM, Demondion X, Cotten A. Early rheumatoid arthritis: a review of MRI and sonographic findings. AJR American journal of roentgenology. 2007 Dec;189(6):1502-9.

- Fischer T, Ebert B, Voigt J, Macdonald R, Schneider U, Thomas A, et al. Detection of rheumatoid arthritis using non-specific contrast enhanced fluorescence imaging. Academic radiology. 2010 Mar;17(3):375-81.
- Werner SG, Langer HE, Ohrndorf S, Bahner M, Schott P, Schwenke C, et al. Inflammation assessment in patients with arthritis using a novel in vivo fluorescence optical imaging technology. Annals of the rheumatic diseases. 2012 Apr;71(4):504-10.
- 39. Meier R, Thurmel K, Moog P, Noel PB, Ahari C, Sievert M, et al. Detection of synovitis in the hands of patients with rheumatologic disorders: diagnostic performance of optical imaging in comparison with magnetic resonance imaging. Arthritis and rheumatism. 2012 Aug;64(8):2489-98.
- 40. Kisten Y, Gyori N, Af Klint E, Rezaei H, Levitsky A, Karlsson A, et al. Detection of clinically manifest and silent synovitis in the hands and wrists by fluorescence optical imaging. RMD open. 2015;1(1):e000106.
- 41. Krohn M, Ohrndorf S, Werner SG, Schicke B, Burmester GR, Hamm B, et al. Near-infrared Fluorescence Optical Imaging in Early Rheumatoid Arthritis: A Comparison to Magnetic Resonance Imaging and Ultrasonography. The Journal of rheumatology. 2015 Jul;42(7):1112-8.
- 42. Smolen JS, Landewe R, Breedveld FC, Buch M, Burmester G, Dougados M, et al. EULAR recommendations for the management of rheumatoid arthritis with synthetic and biological disease-modifying antirheumatic drugs: 2013 update. Annals of the rheumatic diseases. 2014 Mar;73(3):492-509.
- 43. Singh JA, Saag KG, Bridges SL, Jr., Akl EA, Bannuru RR, Sullivan MC, et al. 2015 American College of Rheumatology Guideline for the Treatment of Rheumatoid Arthritis. Arthritis & rheumatology (Hoboken, NJ). 2016 Jan;68(1):1-26.
- 44. Bannwarth B, Labat L, Moride Y, Schaeverbeke T. Methotrexate in rheumatoid arthritis. An update. Drugs. 1994 Jan;47(1):25-50.
- 45. Cronstein BN. Going with the flow: methotrexate, adenosine, and blood flow. Annals of the rheumatic diseases. 2006 Apr;65(4):421-2.
- 46. Thomas S, Fisher KH, Snowden JA, Danson SJ, Brown S, Zeidler MP. Methotrexate Is a JAK/STAT Pathway Inhibitor. PloS one. 2015;10(7):e0130078.
- 47. Phillips DC, Woollard KJ, Griffiths HR. The anti-inflammatory actions of methotrexate are critically dependent upon the production of reactive oxygen species. British journal of pharmacology. 2003 Feb;138(3):501-11.
- 48. Herman S, Zurgil N, Langevitz P, Ehrenfeld M, Deutsch M. The induction of apoptosis by methotrexate in activated lymphocytes as indicated by fluorescence hyperpolarization: a possible model for predicting methotrexate therapy for rheumatoid arthritis patients. Cell structure and function. 2003 Apr;28(2):113-22.
- 49. Hsu PC, Hour TC, Liao YF, Hung YC, Liu CC, Chang WH, et al. Increasing ornithine decarboxylase activity is another way of prolactin preventing methotrexate-induced apoptosis: crosstalk between ODC and BCL-2. Apoptosis : an international journal on programmed cell death. 2006 Mar;11(3):389-99.
- 50. Hasko G, Cronstein BN. Adenosine: an endogenous regulator of innate immunity. Trends in immunology. 2004 Jan;25(1):33-9.
- 51. Wessels JA, Huizinga TW, Guchelaar HJ. Recent insights in the pharmacological actions of methotrexate in the treatment of rheumatoid arthritis. Rheumatology. 2008 Mar;47(3):249-55.
- 52. ODell JR, Haire CE, Erikson N, Drymalski W, Palmer W, Eckhoff PJ, et al. Treatment of rheumatoid arthritis with methotrexate alone, sulfasalazine and hydroxychloroquine, or a combination of all three medications. New Engl J Med. 1996 May 16;334(20):1287-91.
- 53. O'Dell JR, Leff R, Paulsen G, Haire C, Mallek J, Eckhoff PJ, et al. Treatment of rheumatoid arthritis with methotrexate and hydroxychloroquine, methotrexate and sulfasalazine, or a combination of the three medications: results of a two-year, randomized, double-blind, placebo-controlled trial. Arthritis and rheumatism. 2002 May;46(5):1164-70.
- 54. de Jong PH, Hazes JM, Han HK, Huisman M, van Zeben D, van der Lubbe PA, et al. Randomised comparison of initial triple DMARD therapy with methotrexate monotherapy in combination with low-dose glucocorticoid bridging therapy; 1-year data of the tREACH trial. Annals of the rheumatic diseases. 2014 Jul;73(7):1331-9.
- 55. Kohler G, Milstein C. Continuous cultures of fused cells secreting antibody of predefined specificity. Nature. 1975 Aug 7;256(5517):495-7.
- 56. Malaviya AP, Ostor AJ. Rheumatoid arthritis and the era of biologic therapy. Inflammopharmacology. 2012 Apr;20(2):59-69.
- 57. Elliott MJ, Maini RN, Feldmann M, Kalden JR, Antoni C, Smolen JS, et al. Randomised doubleblind comparison of chimeric monoclonal antibody to tumour necrosis factor alpha (cA2) versus placebo in rheumatoid arthritis. Lancet. 1994 Oct 22;344(8930):1105-10.
- 58. Maini R, St Clair EW, Breedveld F, Furst D, Kalden J, Weisman M, et al. Infliximab (chimeric antitumour necrosis factor α monoclonal antibody) versus placebo in rheumatoid arthritis patients receiving concomitant methotrexate: a randomised phase III trial. The Lancet. 1999 12/4/;354(9194):1932-9.
- 59. Moreland LW, Baumgartner SW, Schiff MH, Tindall EA, Fleischmann RM, Weaver AL, et al. Treatment of rheumatoid arthritis with a recombinant human tumor necrosis factor receptor (p75)-Fc fusion protein. The New England journal of medicine. 1997 Jul 17;337(3):141-7.
- Weinblatt ME, Kremer JM, Bankhurst AD, Bulpitt KJ, Fleischmann RM, Fox RI, et al. A trial of etanercept, a recombinant tumor necrosis factor receptor:Fc fusion protein, in patients with rheumatoid arthritis receiving methotrexate. The New England journal of medicine. 1999 Jan 28;340(4):253-9.
- 61. Moreland LW, Schiff MH, Baumgartner SW, Tindall EA, Fleischmann RM, Bulpitt KJ, et al. Etanercept therapy in rheumatoid arthritis. A randomized, controlled trial. Annals of internal medicine. 1999 Mar 16;130(6):478-86.
- 62. Kempeni J. Preliminary results of early clinical trials with the fully human anti-TNFalpha monoclonal antibody D2E7. Annals of the rheumatic diseases. 1999 Nov;58 Suppl 1:I70-2.
- 63. Weinblatt ME, Keystone EC, Furst DE, Moreland LW, Weisman MH, Birbara CA, et al. Adalimumab, a fully human anti-tumor necrosis factor alpha monoclonal antibody, for the treatment of rheumatoid arthritis in patients taking concomitant methotrexate: the ARMADA trial. Arthritis and rheumatism. 2003 Jan;48(1):35-45.
- 64. van de Putte LB, Rau R, Breedveld FC, Kalden JR, Malaise MG, van Riel PL, et al. Efficacy and safety of the fully human anti-tumour necrosis factor alpha monoclonal antibody adalimumab (D2E7) in DMARD refractory patients with rheumatoid arthritis: a 12 week, phase II study. Annals of the rheumatic diseases. 2003 Dec;62(12):1168-77.
- 65. Rau R, Simianer S, van Riel PL, van de Putte LB, Kruger K, Schattenkirchner M, et al. Rapid alleviation of signs and symptoms of rheumatoid arthritis with intravenous or subcutaneous administration of adalimumab in combination with methotrexate. Scandinavian journal of rheumatology. 2004;33(3):145-53.
- 66. Choy EH, Hazleman B, Smith M, Moss K, Lisi L, Scott DG, et al. Efficacy of a novel PEGylated humanized anti-TNF fragment (CDP870) in patients with rheumatoid arthritis: a phase II double-blinded, randomized, dose-escalating trial. Rheumatology. 2002 Oct;41(10):1133-7.
- 67. Kaushik VV, Moots RJ. CDP-870 (certolizumab) in rheumatoid arthritis. Expert opinion on biological therapy. 2005 Apr;5(4):601-6.
- 68. Keystone E, Mason D, Combe B. The anti-tnf certolizumab pegol in combination with methotrexate is significantly more effective than methotrexate alone in the treatment of patients with active rheumatoid arthritis: Preliminary results from the rapid 1 study. Annals of the rheumatic diseases. 2007 Jul;66:55-.
- 69. Kay J, Matteson EL, Dasgupta B, Nash P, Durez P, Hall S, et al. Golimumab in patients with active rheumatoid arthritis despite treatment with methotrexate: a randomized, double-blind, placebo-controlled, dose-ranging study. Arthritis and rheumatism. 2008 Apr;58(4):964-75.

- Combe B, Dasgupta B, Louw I, Pal S, Wollenhaupt J, Zerbini CA, et al. Efficacy and safety of golimumab as add-on therapy to disease-modifying antirheumatic drugs: results of the GO-MORE study. Annals of the rheumatic diseases. 2014 Aug;73(8):1477-86.
- 71. Edwards JC, Cambridge G. Sustained improvement in rheumatoid arthritis following a protocol designed to deplete B lymphocytes. Rheumatology. 2001 Feb;40(2):205-11.
- 72. De Vita S, Zaja F, Sacco S, De Candia A, Fanin R, Ferraccioli G. Efficacy of selective B cell blockade in the treatment of rheumatoid arthritis Evidence for a pathogenetic role of B cells. Arthritis and rheumatism. 2002 Aug;46(8):2029-33.
- 73. Leandro MJ, Edwards JC, Cambridge G. Clinical outcome in 22 patients with rheumatoid arthritis treated with B lymphocyte depletion. Annals of the rheumatic diseases. 2002 Oct;61(10):883-8.
- 74. Kneitz C, Wilhelm M, Tony HP. Improvement of refractory rheumatoid arthritis after depletion of B cells. Scandinavian journal of rheumatology. 2004;33(2):82-6.
- 75. Edwards JC, Leandro MJ, Cambridge G. B lymphocyte depletion in rheumatoid arthritis: targeting of CD20. Current directions in autoimmunity. 2005;8:175-92.
- 76. Emery P, Fleischmann R, Filipowicz-Sosnowska A, Schechtman J, Szczepanski L, Kavanaugh A, et al. The efficacy and safety of rituximab in patients with active rheumatoid arthritis despite methotrexate treatment: results of a phase IIB randomized, double-blind, placebo-controlled, dose-ranging trial. Arthritis and rheumatism. 2006 May;54(5):1390-400.
- 77. Wendler J, Burmester GR, Sorensen H, Krause A, Richter C, Tony HP, et al. Rituximab in patients with rheumatoid arthritis in routine practice (GERINIS): six-year results from a prospective, multicentre, non-interventional study in 2,484 patients. Arthritis research & therapy. 2014;16(2):R80.
- 78. Bresnihan B, Alvaro-Gracia JM, Cobby M, Doherty M, Domljan Z, Emery P, et al. Treatment of rheumatoid arthritis with recombinant human interleukin-1 receptor antagonist. Arthritis and rheumatism. 1998 Dec;41(12):2196-204.
- 79. Jiang Y, Genant HK, Watt I, Cobby M, Bresnihan B, Aitchison R, et al. A multicenter, double-blind, dose-ranging, randomized, placebo-controlled study of recombinant human interleukin-1 receptor antagonist in patients with rheumatoid arthritis: radiologic progression and correlation of Genant and Larsen scores. Arthritis and rheumatism. 2000 May;43(5):1001-9.
- Watt I, Cobby M. Treatment of rheumatoid arthritis patients with interleukin-1 receptor antagonist: radiologic assessment. Seminars in arthritis and rheumatism. 2001 Apr;30(5 Suppl 2):21-5.
- 81. Cohen S, Hurd E, Cush J, Schiff M, Weinblatt ME, Moreland LW, et al. Treatment of rheumatoid arthritis with anakinra, a recombinant human interleukin-1 receptor antagonist, in combination with methotrexate: results of a twenty-four-week, multicenter, randomized, double-blind, placebo-controlled trial. Arthritis and rheumatism. 2002 Mar;46(3):614-24.
- Knoerzer DB, Karr RW, Schwartz BD, Mengle-Gaw LJ. Collagen-induced arthritis in the BB rat. Prevention of disease by treatment with CTLA-4-Ig. The Journal of clinical investigation. 1995 Aug;96(2):987-93.
- 83. Moreland LW, Alten R, Van den Bosch F, Appelboom T, Leon M, Emery P, et al. Costimulatory blockade in patients with rheumatoid arthritis: a pilot, dose-finding, double-blind, placebocontrolled clinical trial evaluating CTLA-4lg and LEA29Y eighty-five days after the first infusion. Arthritis and rheumatism. 2002 Jun;46(6):1470-9.
- Kremer JM, Westhovens R, Leon M, Di Giorgio E, Alten R, Steinfeld S, et al. Treatment of rheumatoid arthritis by selective inhibition of T-cell activation with fusion protein CTLA4Ig. The New England journal of medicine. 2003 Nov 13;349(20):1907-15.
- Kremer JM, Dougados M, Emery P, Durez P, Sibilia J, Shergy W, et al. Treatment of rheumatoid arthritis with the selective costimulation modulator abatacept: twelve-month results of a phase iib, double-blind, randomized, placebo-controlled trial. Arthritis and rheumatism. 2005 Aug;52(8):2263-71.

- Genovese MC, Becker JC, Schiff M, Luggen M, Sherrer Y, Kremer J, et al. Abatacept for rheumatoid arthritis refractory to tumor necrosis factor alpha inhibition. The New England journal of medicine. 2005 Sep 15;353(11):1114-23.
- Westhovens R, Cole JC, Li T, Martin M, Maclean R, Lin P, et al. Improved health-related quality of life for rheumatoid arthritis patients treated with abatacept who have inadequate response to anti-TNF therapy in a double-blind, placebo-controlled, multicentre randomized clinical trial. Rheumatology. 2006 Oct;45(10):1238-46.
- 88. Kremer JM, Genant HK, Moreland LW, Russell AS, Emery P, Abud-Mendoza C, et al. Effects of abatacept in patients with methotrexate-resistant active rheumatoid arthritis: a randomized trial. Annals of internal medicine. 2006 Jun 20;144(12):865-76.
- 89. Genovese MC, Schiff M, Luggen M, Becker JC, Aranda R, Teng J, et al. Efficacy and safety of the selective co-stimulation modulator abatacept following 2 years of treatment in patients with rheumatoid arthritis and an inadequate response to anti-tumour necrosis factor therapy. Annals of the rheumatic diseases. 2008 Apr;67(4):547-54.
- 90. Sato K, Tsuchiya M, Saldanha J, Koishihara Y, Ohsugi Y, Kishimoto T, et al. Humanization of a mouse anti-human interleukin-6 receptor antibody comparing two methods for selecting human framework regions. Molecular immunology. 1994 Apr;31(5):371-81.
- 91. Nishimoto N, Yoshizaki K, Miyasaka N, Yamamoto K, Kawai S, Takeuchi T, et al. Treatment of rheumatoid arthritis with humanized anti-interleukin-6 receptor antibody: a multicenter, double-blind, placebo-controlled trial. Arthritis and rheumatism. 2004 Jun;50(6):1761-9.
- 92. Maini RN, Taylor PC, Szechinski J, Pavelka K, Broll J, Balint G, et al. Double-blind randomized controlled clinical trial of the interleukin-6 receptor antagonist, tocilizumab, in European patients with rheumatoid arthritis who had an incomplete response to methotrexate. Arthritis and rheumatism. 2006 Sep;54(9):2817-29.
- 93. Moreland LW, O'Dell JR, Paulus HE, Curtis JR, Bathon JM, St Clair EW, et al. A randomized comparative effectiveness study of oral triple therapy versus etanercept plus methotrexate in early aggressive rheumatoid arthritis: the treatment of Early Aggressive Rheumatoid Arthritis Trial. Arthritis and rheumatism. 2012 Sep;64(9):2824-35.
- 94. O'Dell JR, Mikuls TR, Taylor TH, Ahluwalia V, Brophy M, Warren SR, et al. Therapies for active rheumatoid arthritis after methotrexate failure. The New England journal of medicine. 2013 Jul 25;369(4):307-18.
- 95. Rantalaiho V, Kautiainen H, Korpela M, Hannonen P, Kaipiainen-Seppanen O, Mottonen T, et al. Targeted treatment with a combination of traditional DMARDs produces excellent clinical and radiographic long-term outcomes in early rheumatoid arthritis regardless of initial infliximab. The 5-year follow-up results of a randomised clinical trial, the NEO-RACo trial. Annals of the rheumatic diseases. 2014 Nov;73(11):1954-61.
- 96. van Vollenhoven RF, Ernestam S, Geborek P, Petersson IF, Coster L, Waltbrand E, et al. Addition of infliximab compared with addition of sulfasalazine and hydroxychloroquine to methotrexate in patients with early rheumatoid arthritis (Swefot trial): 1-year results of a randomised trial. Lancet. 2009 Aug 8;374(9688):459-66.
- 97. Saevarsdottir S, Rezaei H, Geborek P, Petersson I, Ernestam S, Albertsson K, et al. Current smoking status is a strong predictor of radiographic progression in early rheumatoid arthritis: results from the SWEFOT trial. Annals of the rheumatic diseases. 2014 Apr 4.
- Saevarsdottir S, Wallin H, Seddighzadeh M, Ernestam S, Geborek P, Petersson IF, et al. Predictors of response to methotrexate in early DMARD naive rheumatoid arthritis: results from the initial open-label phase of the SWEFOT trial. Annals of the rheumatic diseases. 2011 Mar;70(3):469-75.
- 99. Saevarsdottir S, Wedren S, Seddighzadeh M, Bengtsson C, Wesley A, Lindblad S, et al. Patients with early rheumatoid arthritis who smoke are less likely to respond to treatment with methotrexate and tumor necrosis factor inhibitors: observations from the Epidemiological Investigation of Rheumatoid Arthritis and the Swedish Rheumatology Register cohorts. Arthritis and rheumatism. 2011 Jan;63(1):26-36.

- 100. Nishimura K, Sugiyama D, Kogata Y, Tsuji G, Nakazawa T, Kawano S, et al. Meta-analysis: diagnostic accuracy of anti-cyclic citrullinated peptide antibody and rheumatoid factor for rheumatoid arthritis. Annals of internal medicine. 2007 Jun 5;146(11):797-808.
- 101. Bukhari M, Thomson W, Naseem H, Bunn D, Silman A, Symmons D, et al. The performance of anti-cyclic citrullinated peptide antibodies in predicting the severity of radiologic damage in inflammatory polyarthritis: results from the Norfolk Arthritis Register. Arthritis and rheumatism. 2007 Sep;56(9):2929-35.
- 102. Jilani AA, Mackworth-Young CG. The role of citrullinated protein antibodies in predicting erosive disease in rheumatoid arthritis: a systematic literature review and meta-analysis. International journal of rheumatology. 2015;2015:728610.
- 103. Koga T, Okada A, Fukuda T, Hidaka T, Ishii T, Ueki Y, et al. Anti-citrullinated peptide antibodies are the strongest predictor of clinically relevant radiographic progression in rheumatoid arthritis patients achieving remission or low disease activity: A post hoc analysis of a nationwide cohort in Japan. PloS one. 2017;12(5):e0175281.
- 104. Visser K, Goekoop-Ruiterman YP, de Vries-Bouwstra JK, Ronday HK, Seys PE, Kerstens PJ, et al. A matrix risk model for the prediction of rapid radiographic progression in patients with rheumatoid arthritis receiving different dynamic treatment strategies: post hoc analyses from the BeSt study. Annals of the rheumatic diseases. 2010 Jul;69(7):1333-7.
- 105. Aletaha D, Alasti F, Smolen JS. Rheumatoid factor determines structural progression of rheumatoid arthritis dependent and independent of disease activity. Annals of the rheumatic diseases. 2013 Jun;72(6):875-80.
- 106. Carpenter L, Norton S, Nikiphorou E, Jayakumar K, McWilliams DF, Rennie KL, et al. Reductions in Radiographic Progression in Early Rheumatoid Arthritis Over Twenty-Five Years: Changing Contribution From Rheumatoid Factor in Two Multicenter UK Inception Cohorts. Arthritis care & research. 2017 Dec;69(12):1809-17.
- 107. Miriovsky BJ, Michaud K, Thiele GM, O'Dell JR, Cannon GW, Kerr G, et al. Anti-CCP antibody and rheumatoid factor concentrations predict greater disease burden in U.S. veterans with rheumatoid arthritis. Annals of the rheumatic diseases. 2010 05/03;69(7):1292-7.
- 108. Orr C, Najm A, Biniecka M, McGarry T, Ng CT, Young F, et al. Synovial Immunophenotype and Anti-Citrullinated Peptide Antibodies in Rheumatoid Arthritis Patients: Relationship to Treatment Response and Radiologic Prognosis. Arthritis & rheumatology (Hoboken, NJ). 2017 Nov;69(11):2114-23.
- 109. Katchamart W, Koolvisoot A, Aromdee E, Chiowchanwesawakit P, Muengchan C. Associations of rheumatoid factor and anti-citrullinated peptide antibody with disease progression and treatment outcomes in patients with rheumatoid arthritis. Rheumatology international. 2015 Oct;35(10):1693-9.
- 110. Gossec L, Dougados M, Goupille P, Cantagrel A, Sibilia J, Meyer O, et al. Prognostic factors for remission in early rheumatoid arthritis: a multiparameter prospective study. Annals of the rheumatic diseases. 2004 Jun;63(6):675-80.
- 111. Mancarella L, Bobbio-Pallavicini F, Ceccarelli F, Falappone PC, Ferrante A, Malesci D, et al. Good clinical response, remission, and predictors of remission in rheumatoid arthritis patients treated with tumor necrosis factor-alpha blockers: the GISEA study. The Journal of rheumatology. 2007 Aug;34(8):1670-3.
- 112. Bas S, Genevay S, Meyer O, Gabay C. Anti-cyclic citrullinated peptide antibodies, IgM and IgA rheumatoid factors in the diagnosis and prognosis of rheumatoid arthritis. Rheumatology. 2003 May;42(5):677-80.
- 113. Boire G, Cossette P, de Brum-Fernandes AJ, Liang P, Niyonsenga T, Zhou ZJ, et al. Anti-Sa antibodies and antibodies against cyclic citrullinated peptide are not equivalent as predictors of severe outcomes in patients with recent-onset polyarthritis. Arthritis research & therapy. 2005;7(3):R592-603.

- 114. Vazquez I, Graell E, Gratacos J, Canete JD, Vinas O, Ercilla MG, et al. Prognostic markers of clinical remission in early rheumatoid arthritis after two years of DMARDs in a clinical setting. Clinical and experimental rheumatology. 2007 Mar-Apr;25(2):231-8.
- 115. Verschueren P, Esselens G, Westhovens R. Predictors of remission, normalized physical function, and changes in the working situation during follow-up of patients with early rheumatoid arthritis: an observational study. Scandinavian journal of rheumatology. 2009 May-Jun;38(3):166-72.
- 116. da Mota LM, Dos Santos Neto LL, de Carvalho JF, Pereira IA, Burlingame R, Menard HA, et al. The presence of anti-citrullinated protein antibodies (ACPA) and rheumatoid factor on patients with rheumatoid arthritis (RA) does not interfere with the chance of clinical remission in a follow-up of 3 years. Rheumatology international. 2012 Dec;32(12):3807-12.
- 117. Gottenberg JE, Ravaud P, Cantagrel A, Combe B, Flipo RM, Schaeverbeke T, et al. Positivity for anti-cyclic citrullinated peptide is associated with a better response to abatacept: data from the 'Orencia and Rheumatoid Arthritis' registry. Annals of the rheumatic diseases. 2012 Nov;71(11):1815-9.
- 118. Gardette A, Ottaviani S, Tubach F, Roy C, Nicaise-Roland P, Palazzo E, et al. High anti-CCP antibody titres predict good response to rituximab in patients with active rheumatoid arthritis. Joint, bone, spine : revue du rhumatisme. 2014 Oct;81(5):416-20.
- 119. Calabro P, Chang DW, Willerson JT, Yeh ET. Release of C-reactive protein in response to inflammatory cytokines by human adipocytes: linking obesity to vascular inflammation. Journal of the American College of Cardiology. 2005 Sep 20;46(6):1112-3.
- 120. Kuta AE, Baum LL. C-reactive protein is produced by a small number of normal human peripheral blood lymphocytes. The Journal of experimental medicine. 1986 Jul 1;164(1):321-6.
- 121. Pasceri V, Willerson JT, Yeh ET. Direct proinflammatory effect of C-reactive protein on human endothelial cells. Circulation. 2000 Oct 31;102(18):2165-8.
- 122. Mallya RK, de Beer FC, Berry H, Hamilton ED, Mace BE, Pepys MB. Correlation of clinical parameters of disease activity in rheumatoid arthritis with serum concentration of C-reactive protein and erythrocyte sedimentation rate. The Journal of rheumatology. 1982 Mar-Apr;9(2):224-8.
- 123. Plant MJ, Williams AL, O'Sullivan MM, Lewis PA, Coles EC, Jessop JD. Relationship between timeintegrated C-reactive protein levels and radiologic progression in patients with rheumatoid arthritis. Arthritis and rheumatism. 2000 Jul;43(7):1473-7.
- 124. van Leeuwen MA, van der Heijde DM, van Rijswijk MH, Houtman PM, van Riel PL, van de Putte LB, et al. Interrelationship of outcome measures and process variables in early rheumatoid arthritis. A comparison of radiologic damage, physical disability, joint counts, and acute phase reactants. The Journal of rheumatology. 1994 Mar;21(3):425-9.
- 125. Wolfe F. Comparative usefulness of C-reactive protein and erythrocyte sedimentation rate in patients with rheumatoid arthritis. The Journal of rheumatology. 1997 Aug;24(8):1477-85.
- 126. Kim K-W, Kim B-M, Moon H-W, Lee S-H, Kim H-R. Role of C-reactive protein in osteoclastogenesis in rheumatoid arthritis. Arthritis research & therapy. 2015 March 4;17(1):41.
- 127. Jansen LM, van der Horst-Bruinsma IE, van Schaardenburg D, Bezemer PD, Dijkmans BA. Predictors of radiographic joint damage in patients with early rheumatoid arthritis. Annals of the rheumatic diseases. 2001 Oct;60(10):924-7.
- 128. Sanmarti R, Gomez-Centeno A, Ercilla G, Larrosa M, Vinas O, Vazquez I, et al. Prognostic factors of radiographic progression in early rheumatoid arthritis: a two year prospective study after a structured therapeutic strategy using DMARDs and very low doses of glucocorticoids. Clinical rheumatology. 2007 Jul;26(7):1111-8.
- 129. Vastesaeger N, Xu S, Aletaha D, St Clair EW, Smolen JS. A pilot risk model for the prediction of rapid radiographic progression in rheumatoid arthritis. Rheumatology. 2009 Sep;48(9):1114-21.
- 130. Conigliaro P, Triggianese P, Sole Chimenti M, Tonelli M, Sunzini F, Kroegler B, et al. Factors Predicting 2 Years of Remission and Low Disease Activity in Rheumatoid Arthritis Patients

Treated with TNF-inhibitors. The Israel Medical Association journal : IMAJ. 2017 Aug;19(8):467-72.

- 131. Narvaez J, Magallares B, Diaz Torne C, Hernandez MV, Reina D, Corominas H, et al. Predictive factors for induction of remission in patients with active rheumatoid arthritis treated with tocilizumab in clinical practice. Seminars in arthritis and rheumatism. 2016 Feb;45(4):386-90.
- 132. Navarro-Compan V, Gherghe AM, Smolen JS, Aletaha D, Landewe R, van der Heijde D. Relationship between disease activity indices and their individual components and radiographic progression in RA: a systematic literature review. Rheumatology. 2015 Jun;54(6):994-1007.
- 133. Benson MD, Cohen AS. Serum amyloid A protein in amyloidosis, rheumatic, and enoplastic diseases. Arthritis and rheumatism. 1979 Jan;22(1):36-42.
- 134. Maury CP, Teppo AM, Wegelius O. Relationship between urinary sialylated saccharides, serum amyloid A protein, and C-reactive protein in rheumatoid arthritis and systemic lupus erythematosus. Annals of the rheumatic diseases. 1982 Jun;41(3):268-71.
- 135. O'Hara R, Murphy EP, Whitehead AS, FitzGerald O, Bresnihan B. Acute-phase serum amyloid A production by rheumatoid arthritis synovial tissue. Arthritis research. 2000;2(2):142-4.
- 136. Momohara S, Okamoto H, Yamanaka H. Chondrocyte of rheumatoid arthritis serve as a source of intra-articular acute-phase serum amyloid A protein. Clinica chimica acta; international journal of clinical chemistry. 2008 Dec;398(1-2):155-6.
- 137. Chambers RE, MacFarlane DG, Whicher JT, Dieppe PA. Serum amyloid-A protein concentration in rheumatoid arthritis and its role in monitoring disease activity. Annals of the rheumatic diseases. 1983 Dec;42(6):665-7.
- 138. Grindulis KA, Scott DL, Robinson MW, Bacon PA, McConkey B. Serum amyloid A protein during the treatment of rheumatoid arthritis with second-line drugs. British journal of rheumatology. 1985 May;24(2):158-63.
- 139. Cunnane G, Grehan S, Geoghegan S, McCormack C, Shields D, Whitehead AS, et al. Serum amyloid A in the assessment of early inflammatory arthritis. The Journal of rheumatology. 2000 Jan;27(1):58-63.
- 140. Pepys MB. Rheumatoid arthritis: the role of acute-phase proteins. British journal of rheumatology. 1993 Jun;32 Suppl 3:1-2.
- 141. Migita K, Kawabe Y, Tominaga M, Origuchi T, Aoyagi T, Eguchi K. Serum amyloid A protein induces production of matrix metalloproteinases by human synovial fibroblasts. Laboratory investigation; a journal of technical methods and pathology. 1998 May;78(5):535-9.
- 142. O'Hara R, Murphy EP, Whitehead AS, FitzGerald O, Bresnihan B. Local expression of the serum amyloid A and formyl peptide receptor-like 1 genes in synovial tissue is associated with matrix metalloproteinase production in patients with inflammatory arthritis. Arthritis and rheumatism. 2004 Jun;50(6):1788-99.
- 143. Mullan RH, Bresnihan B, Golden-Mason L, Markham T, O'Hara R, FitzGerald O, et al. Acute-phase serum amyloid A stimulation of angiogenesis, leukocyte recruitment, and matrix degradation in rheumatoid arthritis through an NF-kappaB-dependent signal transduction pathway. Arthritis and rheumatism. 2006 Jan;54(1):105-14.
- 144. Lee MS, Yoo SA, Cho CS, Suh PG, Kim WU, Ryu SH. Serum amyloid A binding to formyl peptide receptor-like 1 induces synovial hyperplasia and angiogenesis. J Immunol. 2006 Oct 15;177(8):5585-94.
- 145. Connolly M, Mullan RH, McCormick J, Matthews C, Sullivan O, Kennedy A, et al. Acute-phase serum amyloid A regulates tumor necrosis factor alpha and matrix turnover and predicts disease progression in patients with inflammatory arthritis before and after biologic therapy. Arthritis and rheumatism. 2012 Apr;64(4):1035-45.
- 146. Bjorkman L, Raynes JG, Shah C, Karlsson A, Dahlgren C, Bylund J. The proinflammatory activity of recombinant serum amyloid A is not shared by the endogenous protein in the circulation. Arthritis and rheumatism. 2010 Jun;62(6):1660-5.

- 147. Charles P, Elliott MJ, Davis D, Potter A, Kalden JR, Antoni C, et al. Regulation of cytokines, cytokine inhibitors, and acute-phase proteins following anti-TNF-alpha therapy in rheumatoid arthritis. J Immunol. 1999 Aug 1;163(3):1521-8.
- 148. Migita K, Izumi Y, Jiuchi Y, Kozuru H, Kawahara C, Izumi M, et al. Effects of Janus kinase inhibitor tofacitinib on circulating serum amyloid A and interleukin-6 during treatment for rheumatoid arthritis. Clinical and experimental immunology. 2014 Feb;175(2):208-14.
- 149. Visvanathan S, Wagner C, Rojas J, Kay J, Dasgupta B, Matteson EL, et al. E-selectin, interleukin 18, serum amyloid a, and matrix metalloproteinase 9 are associated with clinical response to golimumab plus methotrexate in patients with active rheumatoid arthritis despite methotrexate therapy. The Journal of rheumatology. 2009 Jul;36(7):1371-9.
- 150. Shen C, Sun XG, Liu N, Mu Y, Hong CC, Wei W, et al. Increased serum amyloid A and its association with autoantibodies, acute phase reactants and disease activity in patients with rheumatoid arthritis. Molecular medicine reports. 2015 Feb;11(2):1528-34.
- 151. Targonska-Stepniak B, Majdan M. Serum amyloid A as a marker of persistent inflammation and an indicator of cardiovascular and renal involvement in patients with rheumatoid arthritis. Mediators of inflammation. 2014;2014:793628.
- 152. Hwang YG, Balasubramani GK, Metes ID, Levesque MC, Bridges SL, Jr., Moreland LW. Differential response of serum amyloid A to different therapies in early rheumatoid arthritis and its potential value as a disease activity biomarker. Arthritis research & therapy. 2016 May 17;18(1):108.
- 153. Smeets TJ, Barg EC, Kraan MC, Smith MD, Breedveld FC, Tak PP. Analysis of the cell infiltrate and expression of proinflammatory cytokines and matrix metalloproteinases in arthroscopic synovial biopsies: comparison with synovial samples from patients with end stage, destructive rheumatoid arthritis. Annals of the rheumatic diseases. 2003 Jul;62(7):635-8.
- 154. Eklund KK, Leirisalo-Repo M, Ranta P, Maki T, Kautiainen H, Hannonen P, et al. Serum IL-1beta levels are associated with the presence of erosions in recent onset rheumatoid arthritis. Clinical and experimental rheumatology. 2007 Sep-Oct;25(5):684-9.
- 155. Schiff MH. Role of interleukin 1 and interleukin 1 receptor antagonist in the mediation of rheumatoid arthritis. Annals of the rheumatic diseases. 2000 Nov;59 Suppl 1:i103-8.
- 156. Harris ED, Jr. Rheumatoid arthritis. Pathophysiology and implications for therapy. The New England journal of medicine. 1990 May 3;322(18):1277-89.
- 157. Arend WP. Cytokine imbalance in the pathogenesis of rheumatoid arthritis: the role of interleukin-1 receptor antagonist. Seminars in arthritis and rheumatism. 2001 Apr;30(5 Suppl 2):1-6.
- 158. Campion GV, Lebsack ME, Lookabaugh J, Gordon G, Catalano M. Dose-range and dose-frequency study of recombinant human interleukin-1 receptor antagonist in patients with rheumatoid arthritis. The IL-1Ra Arthritis Study Group. Arthritis and rheumatism. 1996 Jul;39(7):1092-101.
- 159. Maury CP, Andersson LC, Teppo AM, Partanen S, Juvonen E. Mechanism of anaemia in rheumatoid arthritis: demonstration of raised interleukin 1 beta concentrations in anaemic patients and of interleukin 1 mediated suppression of normal erythropoiesis and proliferation of human erythroleukaemia (HEL) cells in vitro. Annals of the rheumatic diseases. 1988 Dec;47(12):972-8.
- 160. Kolarz G, Mayrhofer F, Peichl P, Posch E, Scherak O, Singer F, et al. Prognostic factors for the outcome of methotrexate treatment in rheumatoid arthritis. Clinical rheumatology. 1995 Sep;14(5):515-8.
- 161. Seitz M, Zwicker M, Villiger PM. Pretreatment cytokine profiles of peripheral blood mononuclear cells and serum from patients with rheumatoid arthritis in different american college of rheumatology response groups to methotrexate. The Journal of rheumatology. 2003 Jan;30(1):28-35.
- 162. Nowak D, Lewandowicz J, Dbkowska B, Marczak J. Combination of methotrexate and prednizone decreases circulating concentrations of interleukin 1 beta and Interleukin 6 in patients with rheumatoid arthritis. Poor correlation of cytokine suppression with clinical improvement. International journal of immunopathology and pharmacology. 1999 Jan-Apr;12(1):13-21.

- 163. Kopp S, Alstergren P, Ernestam S, Nordahl S, Bratt J. Interleukin-1beta influences the effect of infliximab on temporomandibular joint pain in rheumatoid arthritis. Scandinavian journal of rheumatology. 2006 May-Jun;35(3):182-8.
- 164. Buch MH, Reece RJ, Quinn MA, English A, Cunnane G, Henshaw K, et al. The value of synovial cytokine expression in predicting the clinical response to TNF antagonist therapy (infliximab). Rheumatology. 2008 Oct;47(10):1469-75.
- 165. Kayakabe K, Kuroiwa T, Sakurai N, Ikeuchi H, Kadiombo AT, Sakairi T, et al. Interleukin-1beta measurement in stimulated whole blood cultures is useful to predict response to anti-TNF therapies in rheumatoid arthritis. Rheumatology. 2012 Sep;51(9):1639-43.
- 166. Okano T, Inui K, Tada M, Sugioka Y, Mamoto K, Wakitani S, et al. Levels of interleukin-1 beta can predict response to tocilizumab therapy in rheumatoid arthritis: the PETITE (predictors of effectiveness of tocilizumab therapy) study. Rheumatology international. 2016 Mar;36(3):349-57.
- 167. Nakahara H, Song J, Sugimoto M, Hagihara K, Kishimoto T, Yoshizaki K, et al. Anti-interleukin-6 receptor antibody therapy reduces vascular endothelial growth factor production in rheumatoid arthritis. Arthritis and rheumatism. 2003 Jun;48(6):1521-9.
- 168. Jego G, Bataille R, Pellat-Deceunynck C. Interleukin-6 is a growth factor for nonmalignant human plasmablasts. Blood. 2001 Mar 15;97(6):1817-22.
- 169. Dienz O, Eaton SM, Bond JP, Neveu W, Moquin D, Noubade R, et al. The induction of antibody production by IL-6 is indirectly mediated by IL-21 produced by CD4+ T cells. The Journal of experimental medicine. 2009 Jan 16;206(1):69-78.
- 170. Dasgupta B, Corkill M, Kirkham B, Gibson T, Panayi G. Serial estimation of interleukin 6 as a measure of systemic disease in rheumatoid arthritis. The Journal of rheumatology. 1992 Jan;19(1):22-5.
- 171. Sawada T, Hirohata S, Inoue T, Ito K. Correlation between rheumatoid factor and IL-6 activity in synovial fluids from patients with rheumatoid arthritis. Clinical and experimental rheumatology. 1991 Jul-Aug;9(4):363-8.
- 172. Acosta-Rodriguez EV, Napolitani G, Lanzavecchia A, Sallusto F. Interleukins 1beta and 6 but not transforming growth factor-beta are essential for the differentiation of interleukin 17-producing human T helper cells. Nature immunology. 2007 Sep;8(9):942-9.
- 173. Lally F, Smith E, Filer A, Stone MA, Shaw JS, Nash GB, et al. A novel mechanism of neutrophil recruitment in a coculture model of the rheumatoid synovium. Arthritis and rheumatism. 2005 Nov;52(11):3460-9.
- 174. Matsumoto T, Tsurumoto T, Shindo H. Interleukin-6 levels in synovial fluids of patients with rheumatoid arthritis correlated with the infiltration of inflammatory cells in synovial membrane. Rheumatology international. 2006 Oct;26(12):1096-100.
- 175. Palmqvist P, Persson E, Conaway HH, Lerner UH. IL-6, leukemia inhibitory factor, and oncostatin M stimulate bone resorption and regulate the expression of receptor activator of NF-kappa B ligand, osteoprotegerin, and receptor activator of NF-kappa B in mouse calvariae. J Immunol. 2002 Sep 15;169(6):3353-62.
- 176. Kotake S, Sato K, Kim KJ, Takahashi N, Udagawa N, Nakamura I, et al. Interleukin-6 and soluble interleukin-6 receptors in the synovial fluids from rheumatoid arthritis patients are responsible for osteoclast-like cell formation. Journal of bone and mineral research : the official journal of the American Society for Bone and Mineral Research. 1996 Jan;11(1):88-95.
- 177. Suzuki M, Hashizume M, Yoshida H, Shiina M, Mihara M. IL-6 and IL-1 synergistically enhanced the production of MMPs from synovial cells by up-regulating IL-6 production and IL-1 receptor I expression. Cytokine. 2010 Aug;51(2):178-83. PubMed PMID: 20403707. Epub 2010/04/21. eng.
- 178. Castell JV, Gomez-Lechon MJ, David M, Andus T, Geiger T, Trullenque R, et al. Interleukin-6 is the major regulator of acute phase protein synthesis in adult human hepatocytes. FEBS Lett. 1989 Jan 2;242(2):237-9.
- 179. Heinrich PC, Castell JV, Andus T. Interleukin-6 and the acute phase response. The Biochemical journal. 1990 Feb 1;265(3):621-36.

- 180. Nemeth E, Rivera S, Gabayan V, Keller C, Taudorf S, Pedersen BK, et al. IL-6 mediates hypoferremia of inflammation by inducing the synthesis of the iron regulatory hormone hepcidin. The Journal of clinical investigation. 2004 May;113(9):1271-6.
- 181. Spath-Schwalbe E, Hansen K, Schmidt F, Schrezenmeier H, Marshall L, Burger K, et al. Acute effects of recombinant human interleukin-6 on endocrine and central nervous sleep functions in healthy men. The Journal of clinical endocrinology and metabolism. 1998 May;83(5):1573-9.
- 182. Barrera P, Haagsma CJ, Boerbooms AM, Van Riel PL, Borm GF, Van de Putte LB, et al. Effect of methotrexate alone or in combination with sulphasalazine on the production and circulating concentrations of cytokines and their antagonists. Longitudinal evaluation in patients with rheumatoid arthritis. British journal of rheumatology. 1995 Aug;34(8):747-55.
- 183. Hobl EL, Mader RM, Erlacher L, Duhm B, Mustak M, Broll H, et al. The influence of methotrexate on the gene expression of the pro-inflammatory cytokine IL-12A in the therapy of rheumatoid arthritis. Clinical and experimental rheumatology. 2011 Nov-Dec;29(6):963-9.
- 184. Straub RH, Muller-Ladner U, Lichtinger T, Scholmerich J, Menninger H, Lang B. Decrease of interleukin 6 during the first 12 months is a prognostic marker for clinical outcome during 36 months treatment with disease-modifying anti-rheumatic drugs. British journal of rheumatology. 1997 Dec;36(12):1298-303.
- 185. Fabre S, Dupuy AM, Dossat N, Guisset C, Cohen JD, Cristol JP, et al. Protein biochip array technology for cytokine profiling predicts etanercept responsiveness in rheumatoid arthritis. Clinical and experimental immunology. 2008 Aug;153(2):188-95.
- 186. Fabre S, Guisset C, Tatem L, Dossat N, Dupuy AM, Cohen JD, et al. Protein biochip array technology to monitor rituximab in rheumatoid arthritis. Clinical and experimental immunology. 2009 Mar;155(3):395-402.
- 187. Brahe CH, Dehlendorff C, Ostergaard M, Johansen JS, Ornbjerg LM, Horslev-Petersen K, et al. Circulating serum interleukin-6, serum chitinase-3-like protein-1, and plasma vascular endothelial growth factor are not predictive for remission and radiographic progression in patients with early rheumatoid arthritis: post-hoc explorative and validation studies based on the CIMESTRA and OPERA trials. Scandinavian journal of rheumatology. 2018 Jan 16:1-11.
- 188. Shi R, Chen M, Litifu B. Serum interleukin-6 and survivin levels predict clinical response to etanercept treatment in patients with established rheumatoid arthritis. Modern rheumatology / the Japan Rheumatism Association. 2018 Jan;28(1):126-32.
- 189. Ferraccioli G, Tolusso B, Bobbio-Pallavicini F, Gremese E, Ravagnani V, Benucci M, et al. Biomarkers of good EULAR response to the B cell depletion therapy in all seropositive rheumatoid arthritis patients: clues for the pathogenesis. PloS one. 2012;7(7):e40362.
- 190. Shimamoto K, Ito T, Ozaki Y, Amuro H, Tanaka A, Nishizawa T, et al. Serum interleukin 6 before and after therapy with tocilizumab is a principal biomarker in patients with rheumatoid arthritis. The Journal of rheumatology. 2013 Jul;40(7):1074-81.
- 191. Wang J, Devenport J, Low JM, Yu D, Hitraya E. Relationship Between Baseline and Early Changes in C-Reactive Protein and Interleukin-6 Levels and Clinical Response to Tocilizumab in Rheumatoid Arthritis. Arthritis care & research. 2016 Jun;68(6):882-5.
- 192. Aizu M, Mizushima I, Nakazaki S, Nakashima A, Kato T, Murayama T, et al. Changes in serum interleukin-6 levels as possible predictor of efficacy of tocilizumab treatment in rheumatoid arthritis. Modern rheumatology / the Japan Rheumatism Association. 2017 Sep 12:1-7.
- 193. Hayashi S, Suzuki K, Yoshimoto K, Takeshita M, Kurasawa T, Yamaoka K, et al. Early Prognostic Factors Associated with the Efficacy of Infliximab Treatment for Patients with Rheumatoid Arthritis with Inadequate Response to Methotrexate. Rheumatology and therapy. 2016 Jun;3(1):155-66.
- 194. Takasugi K, Nishida K, Natsumeda M, Yamashita M, Yamamoto W, Ezawa K. IL-6 is an independent predictive factor of drug survival after dose escalation of infliximab in patients with rheumatoid arthritis. Modern rheumatology / the Japan Rheumatism Association. 2017 Aug 22:1-9.

- 195. Klein-Wieringa IR, van der Linden MP, Knevel R, Kwekkeboom JC, van Beelen E, Huizinga TW, et al. Baseline serum adipokine levels predict radiographic progression in early rheumatoid arthritis. Arthritis and rheumatism. 2011 Sep;63(9):2567-74.
- 196. Gottenberg JE, Dayer JM, Lukas C, Ducot B, Chiocchia G, Cantagrel A, et al. Serum IL-6 and IL-21 are associated with markers of B cell activation and structural progression in early rheumatoid arthritis: results from the ESPOIR cohort. Annals of the rheumatic diseases. 2012 Jul;71(7):1243-8.
- 197. Park YJ, Yoo SA, Kim GR, Cho CS, Kim WU. Urinary interleukin-6 as a predictor of radiographic progression in rheumatoid arthritis: A 3-year evaluation. Scientific reports. 2016 Oct 12;6:35242.
- 198. Kondo Y, Kaneko Y, Sugiura H, Matsumoto S, Nishina N, Kuwana M, et al. Pre-treatment interleukin-6 levels strongly affect bone erosion progression and repair detected by magnetic resonance imaging in rheumatoid arthritis patients. Rheumatology. 2017 Jul 1;56(7):1089-94.
- 199. Noack M, Miossec P. Selected cytokine pathways in rheumatoid arthritis. Seminars in immunopathology. 2017 Jun;39(4):365-83.
- 200. Bartsch JW, Wildeboer D, Koller G, Naus S, Rittger A, Moss ML, et al. Tumor necrosis factor-alpha (TNF-alpha) regulates shedding of TNF-alpha receptor 1 by the metalloprotease-disintegrin ADAM8: evidence for a protease-regulated feedback loop in neuroprotection. The Journal of neuroscience : the official journal of the Society for Neuroscience. 2010 Sep 8;30(36):12210-8.
- 201. McDermott MF, Aksentijevich I, Galon J, McDermott EM, Ogunkolade BW, Centola M, et al. Germline mutations in the extracellular domains of the 55 kDa TNF receptor, TNFR1, define a family of dominantly inherited autoinflammatory syndromes. Cell. 1999 Apr 2;97(1):133-44.
- 202. Brennan FM, Chantry D, Jackson A, Maini R, Feldmann M. Inhibitory effect of TNF alpha antibodies on synovial cell interleukin-1 production in rheumatoid arthritis. Lancet. 1989 Jul 29;2(8657):244-7.
- 203. Namba S, Nakano R, Kitanaka T, Kitanaka N, Nakayama T, Sugiya H. ERK2 and JNK1 contribute to TNF-alpha-induced IL-8 expression in synovial fibroblasts. PloS one. 2017;12(8):e0182923.
- 204. Moelants EAV, Mortier A, Van Damme J, Proost P. Regulation of TNF-α with a focus on rheumatoid arthritis. Immunol Cell Biol. 2013;91(6):393-401.
- 205. Haworth C, Brennan FM, Chantry D, Turner M, Maini RN, Feldmann M. Expression of granulocyte-macrophage colony-stimulating factor in rheumatoid arthritis: regulation by tumor necrosis factor-alpha. European journal of immunology. 1991 Oct;21(10):2575-9.
- 206. Hess A, Axmann R, Rech J, Finzel S, Heindl C, Kreitz S, et al. Blockade of TNF-alpha rapidly inhibits pain responses in the central nervous system. Proceedings of the National Academy of Sciences of the United States of America. 2011 Mar 1;108(9):3731-6.
- 207. Klimiuk PA, Sierakowski S, Latosiewicz R, Cylwik B, Skowronski J, Chwiecko J. Serum cytokines in different histological variants of rheumatoid arthritis. The Journal of rheumatology. 2001 Jun;28(6):1211-7.
- 208. Klimiuk PA, Sierakowski S, Latosiewicz R, Cylwik JP, Cylwik B, Skowronski J, et al. Circulating tumour necrosis factor alpha and soluble tumour necrosis factor receptors in patients with different patterns of rheumatoid synovitis. Annals of the rheumatic diseases. 2003 May;62(5):472-5.
- 209. Edrees AF, Misra SN, Abdou NI. Anti-tumor necrosis factor (TNF) therapy in rheumatoid arthritis: correlation of TNF-alpha serum level with clinical response and benefit from changing dose or frequency of infliximab infusions. Clinical and experimental rheumatology. 2005 Jul-Aug;23(4):469-74.
- 210. Kirkham BW, Lassere MN, Edmonds JP, Juhasz KM, Bird PA, Lee CS, et al. Synovial membrane cytokine expression is predictive of joint damage progression in rheumatoid arthritis: a two-year prospective study (the DAMAGE study cohort). Arthritis and rheumatism. 2006 Apr;54(4):1122-31.
- 211. Wagner C, Chen D, Fan H, Hsia EC, Mack M, Emery P, et al. Evaluation of serum biomarkers associated with radiographic progression in methotrexate-naive rheumatoid arthritis patients treated with methotrexate or golimumab. The Journal of rheumatology. 2013 May;40(5):590-8.

- 212. Pachot A, Arnaud B, Marrote H, Cazalis MA, Diasparra J, Gouraud A, et al. Increased tumor necrosis factor-alpha mRNA expression in whole blood from patients with rheumatoid arthritis: reduction after infliximab treatment does not predict response. The Journal of rheumatology. 2007 Nov;34(11):2158-61.
- 213. Wijbrandts CA, Dijkgraaf MG, Kraan MC, Vinkenoog M, Smeets TJ, Dinant H, et al. The clinical response to infliximab in rheumatoid arthritis is in part dependent on pretreatment tumour necrosis factor alpha expression in the synovium. Annals of the rheumatic diseases. 2008 Aug;67(8):1139-44.
- 214. Hueber W, Tomooka BH, Batliwalla F, Li W, Monach PA, Tibshirani RJ, et al. Blood autoantibody and cytokine profiles predict response to anti-tumor necrosis factor therapy in rheumatoid arthritis. Arthritis research & therapy. 2009;11(3):R76.
- 215. Maillefert JF, Puechal X, Falgarone G, Lizard G, Ornetti P, Solau E, et al. Prediction of response to disease modifying antirheumatic drugs in rheumatoid arthritis. Joint, bone, spine : revue du rhumatisme. 2010 Dec;77(6):558-63.
- 216. Takeuchi T, Miyasaka N, Tatsuki Y, Yano T, Yoshinari T, Abe T, et al. Baseline tumour necrosis factor alpha levels predict the necessity for dose escalation of infliximab therapy in patients with rheumatoid arthritis. Annals of the rheumatic diseases. 2011 Jul;70(7):1208-15.
- 217. Gaffen SL. The role of interleukin-17 in the pathogenesis of rheumatoid arthritis. Current rheumatology reports. 2009 Oct;11(5):365-70.
- 218. van Hamburg JP, Asmawidjaja PS, Davelaar N, Mus AM, Colin EM, Hazes JM, et al. Th17 cells, but not Th1 cells, from patients with early rheumatoid arthritis are potent inducers of matrix metalloproteinases and proinflammatory cytokines upon synovial fibroblast interaction, including autocrine interleukin-17A production. Arthritis and rheumatism. 2011 Jan;63(1):73-83.
- 219. Miossec P. Update on interleukin-17: a role in the pathogenesis of inflammatory arthritis and implication for clinical practice. RMD open. 2017;3(1):e000284.
- 220. Shahrara S, Pickens SR, Dorfleutner A, Pope RM. IL-17 induces monocyte migration in rheumatoid arthritis. J Immunol. 2009 Mar 15;182(6):3884-91.
- 221. Al-Saadany HM, Hussein MS, Gaber RA, Zaytoun HA. Th-17 cells and serum IL-17 in rheumatoid arthritis patients: Correlation with disease activity and severity. Egyptian Rheumatologist. 2016 Jan;38(1):1-7.
- 222. Kokkonen H, Soderstrom I, Rocklov J, Hallmans G, Lejon K, Rantapaa Dahlqvist S. Up-regulation of cytokines and chemokines predates the onset of rheumatoid arthritis. Arthritis and rheumatism. 2010 Feb;62(2):383-91.
- 223. Pavlovic V, Dimic A, Milenkovic S, Krtinic D. Serum levels of IL-17, IL-4, and INFgamma in Serbian patients with early rheumatoid arthritis. Journal of research in medical sciences : the official journal of Isfahan University of Medical Sciences. 2014 Jan;19(1):18-22.
- 224. Yue C, You X, Zhao L, Wang H, Tang F, Zhang F, et al. The effects of adalimumab and methotrexate treatment on peripheral Th17 cells and IL-17/IL-6 secretion in rheumatoid arthritis patients. Rheumatology international. 2010 Nov;30(12):1553-7.
- 225. Shen H, Xia L, Lu J, Xiao W. Infliximab reduces the frequency of interleukin 17-producing cells and the amounts of interleukin 17 in patients with rheumatoid arthritis. Journal of investigative medicine : the official publication of the American Federation for Clinical Research. 2010 Oct;58(7):905-8.
- 226. Chen DY, Chen YM, Chen HH, Hsieh CW, Lin CC, Lan JL. Increasing levels of circulating Th17 cells and interleukin-17 in rheumatoid arthritis patients with an inadequate response to anti-TNFalpha therapy. Arthritis research & therapy. 2011 Jul 30;13(4):R126.
- 227. Alzabin S, Abraham SM, Taher TE, Palfreeman A, Hull D, McNamee K, et al. Incomplete response of inflammatory arthritis to TNFalpha blockade is associated with the Th17 pathway. Annals of the rheumatic diseases. 2012 Oct;71(10):1741-8.
- 228. Lee SJ, Park W, Park SH, Shim SC, Baek HJ, Yoo DH, et al. Low baseline interleukin-17A levels are associated with better treatment response at 12 weeks to tocilizumab therapy in rheumatoid arthritis patients. Journal of immunology research. 2015;2015:487230.

- 229. Jablonska-Trypuc A, Matejczyk M, Rosochacki S. Matrix metalloproteinases (MMPs), the main extracellular matrix (ECM) enzymes in collagen degradation, as a target for anticancer drugs. Journal of enzyme inhibition and medicinal chemistry. 2016;31(sup1):177-83.
- 230. Malemud CJ. Matrix Metalloproteinases and Synovial Joint Pathology. Progress in molecular biology and translational science. 2017;148:305-25.
- 231. Murphy G, Knauper V, Atkinson S, Butler G, English W, Hutton M, et al. Matrix metalloproteinases in arthritic disease. Arthritis research. 2002;4 Suppl 3:S39-49.
- 232. Skacelova M, Hermanova Z, Horak P, Ahmed K, Langova K. Higher levels of matrix metalloproteinase-3 in patients with RA reflect disease activity and structural damage. Biomedical papers of the Medical Faculty of the University Palacky, Olomouc, Czechoslovakia. 2017 Sep;161(3):296-302.
- 233. Uemura Y, Hayashi H, Takahashi T, Saitho T, Umeda R, Ichise Y, et al. [MMP-3 as a Biomarker of Disease Activity of Rheumatoid Arthritis]. Rinsho byori The Japanese journal of clinical pathology. 2015 Dec;63(12):1357-64.
- 234. Ma J, Wang X, Mo Y, Chen L, Zheng D, Wei X, et al. [Value of serum matrix metalloproteinase-3 in the assessment of active disease in patients with rheumatoid arthritis]. Zhonghua yi xue za zhi. 2015 Dec 15;95(47):3823-8.
- 235. Mahmoud RK, El-Ansary AK, El-Eishi HH, Kamal HM, El-Saeed NH. Matrix metalloproteinases MMP-3 and MMP-1 levels in sera and synovial fluids in patients with rheumatoid arthritis and osteoarthritis. The Italian journal of biochemistry. 2005 Sep-Dec;54(3-4):248-57.
- 236. den Broeder AA, Joosten LA, Saxne T, Heinegard D, Fenner H, Miltenburg AM, et al. Long term anti-tumour necrosis factor alpha monotherapy in rheumatoid arthritis: effect on radiological course and prognostic value of markers of cartilage turnover and endothelial activation. Annals of the rheumatic diseases. 2002 Apr;61(4):311-8.
- 237. Fiedorczyk M, Klimiuk PA, Sierakowski S, Gindzienska-Sieskiewicz E, Chwiecko J. Serum matrix metalloproteinases and tissue inhibitors of metalloproteinases in patients with early rheumatoid arthritis. The Journal of rheumatology. 2006 Aug;33(8):1523-9.
- 238. Visvanathan S, Marini JC, Smolen JS, Clair EW, Pritchard C, Shergy W, et al. Changes in biomarkers of inflammation and bone turnover and associations with clinical efficacy following infliximab plus methotrexate therapy in patients with early rheumatoid arthritis. The Journal of rheumatology. 2007 Jul;34(7):1465-74.
- 239. Chen J, Doyle TJ, Liu Y, Aggarwal R, Wang X, Shi Y, et al. Biomarkers of rheumatoid arthritisassociated interstitial lung disease. Arthritis & rheumatology (Hoboken, NJ). 2015 Jan;67(1):28-38.
- 240. Yamanaka H, Matsuda Y, Tanaka M, Sendo W, Nakajima H, Taniguchi A, et al. Serum matrix metalloproteinase 3 as a predictor of the degree of joint destruction during the six months after measurement, in patients with early rheumatoid arthritis. Arthritis and rheumatism. 2000 Apr;43(4):852-8.
- 241. Cunnane G, Fitzgerald O, Beeton C, Cawston TE, Bresnihan B. Early joint erosions and serum levels of matrix metalloproteinase 1, matrix metalloproteinase 3, and tissue inhibitor of metalloproteinases 1 in rheumatoid arthritis. Arthritis and rheumatism. 2001 Oct;44(10):2263-74.
- 242. Ribbens C, Andre B, Jaspar JM, Kaye O, Kaiser MJ, De Groote D, et al. Matrix metalloproteinase-3 serum levels are correlated with disease activity and predict clinical response in rheumatoid arthritis. The Journal of rheumatology. 2000 Apr;27(4):888-93.
- 243. Green MJ, Gough AK, Devlin J, Smith J, Astin P, Taylor D, et al. Serum MMP-3 and MMP-1 and progression of joint damage in early rheumatoid arthritis. Rheumatology. 2003 Jan;42(1):83-8.
- 244. Young-Min S, Cawston T, Marshall N, Coady D, Christgau S, Saxne T, et al. Biomarkers predict radiographic progression in early rheumatoid arthritis and perform well compared with traditional markers. Arthritis and rheumatism. 2007 Oct;56(10):3236-47.
- 245. Shinozaki M, Inoue E, Nakajima A, Hara M, Tomatsu T, Kamatani N, et al. Elevation of serum matrix metalloproteinase-3 as a predictive marker for the long-term disability of rheumatoid

arthritis patients in a prospective observational cohort IORRA. Modern rheumatology / the Japan Rheumatism Association. 2007;17(5):403-8.

- 246. Courvoisier N, Dougados M, Cantagrel A, Goupille P, Meyer O, Sibilia J, et al. Prognostic factors of 10-year radiographic outcome in early rheumatoid arthritis: a prospective study. Arthritis research & therapy. 2008;10(5):R106.
- 247. Galil SM, El-Shafey AM, Hagrass HA, Fawzy F, Sammak AE. Baseline serum level of matrix metalloproteinase-3 as a biomarker of progressive joint damage in rheumatoid arthritis patients. International journal of rheumatic diseases. 2016 Apr;19(4):377-84.
- 248. Shiozawa K, Yamane T, Murata M, Yoshihara R, Tsumiyama K, Imura S, et al. MMP-3 as a predictor for structural remission in RA patients treated with MTX monotherapy. Arthritis research & therapy. 2016 Feb 27;18:55.
- 249. Kawashiri SY, Kawakami A, Ueki Y, Imazato T, Iwamoto N, Fujikawa K, et al. Decrement of serum cartilage oligomeric matrix protein (COMP) in rheumatoid arthritis (RA) patients achieving remission after 6 months of etanercept treatment: comparison with CRP, IgM-RF, MMP-3 and anti-CCP Ab. Joint, bone, spine : revue du rhumatisme. 2010 Oct;77(5):418-20.
- 250. Mamehara A, Sugimoto T, Sugiyama D, Morinobu S, Tsuji G, Kawano S, et al. Serum matrix metalloproteinase-3 as predictor of joint destruction in rheumatoid arthritis, treated with nonbiological disease modifying anti-rheumatic drugs. The Kobe journal of medical sciences. 2010 Sep 30;56(3):E98-107.
- 251. Houseman M, Potter C, Marshall N, Lakey R, Cawston T, Griffiths I, et al. Baseline serum MMP-3 levels in patients with Rheumatoid Arthritis are still independently predictive of radiographic progression in a longitudinal observational cohort at 8 years follow up. Arthritis research & therapy. 2012 Feb 7;14(1):R30.
- 252. Hattori Y, Kojima T, Kaneko A, Kida D, Hirano Y, Fujibayashi T, et al. High rate of improvement in serum matrix metalloproteinase-3 levels at 4 weeks predicts remission at 52 weeks in RA patients treated with adalimumab. Modern rheumatology / the Japan Rheumatism Association. 2018 Jan;28(1):119-25.
- 253. Lee CG, Da Silva CA, Dela Cruz CS, Ahangari F, Ma B, Kang MJ, et al. Role of chitin and chitinase/chitinase-like proteins in inflammation, tissue remodeling, and injury. Annual review of physiology. 2011;73:479-501.
- 254. Vaananen T, Vuolteenaho K, Kautiainen H, Nieminen R, Mottonen T, Hannonen P, et al. Glycoprotein YKL-40: A potential biomarker of disease activity in rheumatoid arthritis during intensive treatment with csDMARDs and infliximab. Evidence from the randomised controlled NEO-RACo trial. PloS one. 2017;12(8):e0183294.
- 255. Combe B, Dougados M, Goupille P, Cantagrel A, Eliaou JF, Sibilia J, et al. Prognostic factors for radiographic damage in early rheumatoid arthritis: a multiparameter prospective study. Arthritis and rheumatism. 2001 Aug;44(8):1736-43.
- 256. Peltomaa R, Paimela L, Harvey S, Helve T, Leirisalo-Repo M. Increased level of YKL-40 in sera from patients with early rheumatoid arthritis: a new marker for disease activity. Rheumatology international. 2001 Jul;20(5):192-6.
- 257. Syversen SW, Goll GL, van der Heijde D, Landewe R, Gaarder PI, Odegard S, et al. Cartilage and bone biomarkers in rheumatoid arthritis: prediction of 10-year radiographic progression. The Journal of rheumatology. 2009 Feb;36(2):266-72.
- 258. Wolf G. Leptin: the weight-reducing plasma protein encoded by the obese gene. Nutrition reviews. 1996 Mar;54(3):91-3.
- 259. Gaillard RC, Spinedi E, Chautard T, Pralong FP. Cytokines, leptin, and the hypothalamo-pituitaryadrenal axis. Annals of the New York Academy of Sciences. 2000;917:647-57.
- 260. Tian G, Liang JN, Wang ZY, Zhou D. Emerging role of leptin in rheumatoid arthritis. Clinical and experimental immunology. 2014 Sep;177(3):557-70.
- 261. Bokarewa M, Bokarew D, Hultgren O, Tarkowski A. Leptin consumption in the inflamed joints of patients with rheumatoid arthritis. Annals of the rheumatic diseases. 2003 Oct;62(10):952-6.

- 262. Xibille-Friedmann DX, Ortiz-Panozo E, Bustos Rivera-Bahena C, Sandoval-Rios M, Hernandez-Gongora SE, Dominguez-Hernandez L, et al. Leptin and adiponectin as predictors of disease activity in rheumatoid arthritis. Clinical and experimental rheumatology. 2015 Jul-Aug;33(4):471-7.
- 263. Erlandsson MC, Doria Medina R, Toyra Silfversward S, Bokarewa MI. Smoking Functions as a Negative Regulator of IGF1 and Impairs Adipokine Network in Patients with Rheumatoid Arthritis. Mediators of inflammation. 2016;2016:3082820.
- 264. Migita K, Maeda Y, Miyashita T, Kimura H, Nakamura M, Ishibashi H, et al. The serum levels of resistin in rheumatoid arthritis patients. Clinical and experimental rheumatology. 2006 Nov-Dec;24(6):698-701.
- 265. Senolt L, Housa D, Vernerova Z, Jirasek T, Svobodova R, Veigl D, et al. Resistin in rheumatoid arthritis synovial tissue, synovial fluid and serum. Annals of the rheumatic diseases. 2007 Apr;66(4):458-63.
- 266. Su CM, Huang CY, Tang CH. Characteristics of resistin in rheumatoid arthritis angiogenesis. Biomarkers in medicine. 2016 Jun;10(6):651-60.
- 267. Fadda SM, Gamal SM, Elsaid NY, Mohy AM. Resistin in inflammatory and degenerative rheumatologic diseases. Relationship between resistin and rheumatoid arthritis disease progression. Z Rheumatol. 2013 Aug;72(6):594-600.
- 268. Gonzalez-Gay MA, Garcia-Unzueta MT, Gonzalez-Juanatey C, Miranda-Filloy JA, Vazquez-Rodriguez TR, De Matias JM, et al. Anti-TNF-alpha therapy modulates resistin in patients with rheumatoid arthritis. Clinical and experimental rheumatology. 2008 Mar-Apr;26(2):311-6.
- 269. Otero M, Lago R, Gomez R, Lago F, Dieguez C, Gomez-Reino JJ, et al. Changes in plasma levels of fat-derived hormones adiponectin, leptin, resistin and visfatin in patients with rheumatoid arthritis. Annals of the rheumatic diseases. 2006 Sep;65(9):1198-201.
- 270. Giles JT, van der Heijde DM, Bathon JM. Association of circulating adiponectin levels with progression of radiographic joint destruction in rheumatoid arthritis. Annals of the rheumatic diseases. 2011 Sep;70(9):1562-8.
- 271. Shiozawa S, Shiozawa K, Tanaka Y, Morimoto I, Uchihashi M, Fujita T, et al. Human epidermal growth factor for the stratification of synovial lining layer and neovascularisation in rheumatoid arthritis. Annals of the rheumatic diseases. 1989 Oct;48(10):820-8.
- 272. Farahat MN, Yanni G, Poston R, Panayi GS. Cytokine expression in synovial membranes of patients with rheumatoid arthritis and osteoarthritis. Annals of the rheumatic diseases. 1993 Dec;52(12):870-5.
- 273. Swanson CD, Akama-Garren EH, Stein EA, Petralia JD, Ruiz PJ, Edalati A, et al. Inhibition of epidermal growth factor receptor tyrosine kinase ameliorates collagen-induced arthritis. J Immunol. 2012 Apr 1;188(7):3513-21.
- 274. Chen SY, Shiau AL, Wu CL, Wang CR. Amelioration of experimental arthritis by intra-articular injection of an epidermal growth factor receptor tyrosine kinase inhibitor. Clinical and experimental rheumatology. 2015 Nov-Dec;33(6):839-43.
- 275. Maruotti N, Cantatore FP, Crivellato E, Vacca A, Ribatti D. Angiogenesis in rheumatoid arthritis. Histology and histopathology. 2006 May;21(5):557-66.
- 276. Lee SS, Joo YS, Kim WU, Min DJ, Min JK, Park SH, et al. Vascular endothelial growth factor levels in the serum and synovial fluid of patients with rheumatoid arthritis. Clinical and experimental rheumatology. 2001 May-Jun;19(3):321-4.
- 277. Szekanecz Z, Besenyei T, Paragh G, Koch AE. Angiogenesis in rheumatoid arthritis. Autoimmunity. 2009 Nov;42(7):563-73.
- 278. Choi ST, Kim JH, Seok JY, Park YB, Lee SK. Therapeutic effect of anti-vascular endothelial growth factor receptor I antibody in the established collagen-induced arthritis mouse model. Clinical rheumatology. 2009 Mar;28(3):333-7.
- 279. Marrelli A, Cipriani P, Liakouli V, Carubbi F, Perricone C, Perricone R, et al. Angiogenesis in rheumatoid arthritis: a disease specific process or a common response to chronic inflammation? Autoimmunity reviews. 2011 Aug;10(10):595-8.

- 280. Ballara S, Taylor PC, Reusch P, Marme D, Feldmann M, Maini RN, et al. Raised serum vascular endothelial growth factor levels are associated with destructive change in inflammatory arthritis. Arthritis and rheumatism. 2001 Sep;44(9):2055-64.
- 281. Kong DH, Kim YK, Kim MR, Jang JH, Lee S. Emerging Roles of Vascular Cell Adhesion Molecule-1 (VCAM-1) in Immunological Disorders and Cancer. International journal of molecular sciences. 2018 Apr 2;19(4).
- 282. Smith MD, Slavotinek J, Au V, Weedon H, Parker A, Coleman M, et al. Successful treatment of rheumatoid arthritis is associated with a reduction in synovial membrane cytokines and cell adhesion molecule expression. Rheumatology. 2001 Sep;40(9):965-77.
- 283. Macias I, Garcia-Perez S, Ruiz-Tudela M, Medina F, Chozas N, Giron-Gonzalez JA. Modification of pro- and antiinflammatory cytokines and vascular-related molecules by tumor necrosis factor-a blockade in patients with rheumatoid arthritis. The Journal of rheumatology. 2005 Nov;32(11):2102-8.
- 284. Curtis JR, van der Helm-van Mil AH, Knevel R, Huizinga TW, Haney DJ, Shen Y, et al. Validation of a novel multibiomarker test to assess rheumatoid arthritis disease activity. Arthritis care & research. 2012 Dec;64(12):1794-803.
- 285. Eastman PS, Manning WC, Qureshi F, Haney D, Cavet G, Alexander C, et al. Characterization of a multiplex, 12-biomarker test for rheumatoid arthritis. Journal of pharmaceutical and biomedical analysis. 2012 Nov;70:415-24.
- 286. Centola M, Cavet G, Shen Y, Ramanujan S, Knowlton N, Swan KA, et al. Development of a multibiomarker disease activity test for rheumatoid arthritis. PloS one. 2013;8(4):e60635.
- 287. Bakker MF, Cavet G, Jacobs JW, Bijlsma JW, Haney DJ, Shen Y, et al. Performance of a multibiomarker score measuring rheumatoid arthritis disease activity in the CAMERA tight control study. Annals of the rheumatic diseases. 2012 Oct;71(10):1692-7.
- 288. Hirata S, Dirven L, Shen Y, Centola M, Cavet G, Lems WF, et al. A multi-biomarker score measures rheumatoid arthritis disease activity in the BeSt study. Rheumatology. 2013 Jul;52(7):1202-7.
- 289. Lee YC, Hackett J, Frits M, Iannaccone CK, Shadick NA, Weinblatt ME, et al. Multibiomarker disease activity score and C-reactive protein in a cross-sectional observational study of patients with rheumatoid arthritis with and without concomitant fibromyalgia. Rheumatology. 2016 Apr;55(4):640-8.
- 290. Hirata S, Li W, Defranoux N, Cavet G, Bolce R, Yamaoka K, et al. A multi-biomarker disease activity score tracks clinical response consistently in patients with rheumatoid arthritis treated with different anti-tumor necrosis factor therapies: A retrospective observational study. Modern rheumatology / the Japan Rheumatism Association. 2014 Oct 8:1-6.
- 291. Reiss WG, Devenport JN, Low JM, Wu G, Sasso EH. Interpreting the multi-biomarker disease activity score in the context of tocilizumab treatment for patients with rheumatoid arthritis. Rheumatology international. 2016 Feb;36(2):295-300.
- 292. van der Helm-van Mil AH, Knevel R, Cavet G, Huizinga TW, Haney DJ. An evaluation of molecular and clinical remission in rheumatoid arthritis by assessing radiographic progression. Rheumatology. 2013 May;52(5):839-46.
- 293. Markusse IM, Dirven L, van den Broek M, Bijkerk C, Han KH, Ronday HK, et al. A Multibiomarker Disease Activity Score for Rheumatoid Arthritis Predicts Radiographic Joint Damage in the BeSt Study. Journal of Rheumatology. 2014 Nov;41(11):2114-9.
- 294. Li WY, Sasso EH, van der Helm-van Mil AHM, Huizinga TWJ. Relationship of multi-biomarker disease activity score and other risk factors with radiographic progression in an observational study of patients with rheumatoid arthritis. Rheumatology. 2016 Feb;55(2):357-66.
- 295. Hirata S, Li W, Kubo S, Fukuyo S, Mizuno Y, Hanami K, et al. Association of the multi-biomarker disease activity score with joint destruction in patients with rheumatoid arthritis receiving tumor necrosis factor-alpha inhibitor treatment in clinical practice. Modern rheumatology / the Japan Rheumatism Association. 2016 Nov;26(6):850-6.

- 296. Krabbe S, Bolce R, Brahe CH, Dohn UM, Ejbjerg BJ, Hetland ML, et al. Investigation of a multibiomarker disease activity score in rheumatoid arthritis by comparison with magnetic resonance imaging, computed tomography, ultrasonography, and radiography parameters of inflammation and damage. Scandinavian journal of rheumatology. 2017 Sep;46(5):353-8.
- 297. Rech J, Hueber AJ, Finzel S, Englbrecht M, Haschka J, Manger B, et al. Prediction of disease relapses by multibiomarker disease activity and autoantibody status in patients with rheumatoid arthritis on tapering DMARD treatment. Annals of the rheumatic diseases. 2016 Sep;75(9):1637-44.
- 298. Bouman CAM, van der Maas A, van Herwaarden N, Sasso EH, van den Hoogen FHJ, den Broeder AA. A multi-biomarker score measuring disease activity in rheumatoid arthritis patients tapering adalimumab or etanercept: predictive value for clinical and radiographic outcomes. Rheumatology. 2017 Jun 1;56(6):973-80.
- 299. Ghiti Moghadam M, Lamers-Karnebeek FBG, Vonkeman HE, Ten Klooster PM, Tekstra J, Schilder AM, et al. Multi-biomarker disease activity score as a predictor of disease relapse in patients with rheumatoid arthritis stopping TNF inhibitor treatment. PloS one. 2018;13(5):e0192425.
- 300. Fleischmann R, Connolly SE, Maldonado MA, Schiff M. Brief Report: Estimating Disease Activity Using Multi-Biomarker Disease Activity Scores in Rheumatoid Arthritis Patients Treated With Abatacept or Adalimumab. Arthritis & rheumatology (Hoboken, NJ). 2016 Sep;68(9):2083-9.
- 301. Curtis JR, Greenberg JD, Harrold LR, Kremer JM, Palmer JL. Influence of obesity, age, and comorbidities on the multi-biomarker disease activity test in rheumatoid arthritis. Seminars in arthritis and rheumatism. 2018 Feb;47(4):472-7.
- 302. Anderson PJ. Tumor necrosis factor inhibitors: clinical implications of their different immunogenicity profiles. Seminars in arthritis and rheumatism. 2005 Apr;34(5 Suppl1):19-22.
- 303. Bendtzen K, Geborek P, Svenson M, Larsson L, Kapetanovic MC, Saxne T. Individualized monitoring of drug bioavailability and immunogenicity in rheumatoid arthritis patients treated with the tumor necrosis factor α inhibitor infliximab. Arthritis & Rheumatism. 2006;54(12):3782-9.
- 304. Haraoui B, Cameron L, Ouellet M, White B. Anti-infliximab antibodies in patients with rheumatoid arthritis who require higher doses of infliximab to achieve or maintain a clinical response. The Journal of rheumatology. 2006 Jan;33(1):31-6.
- 305. Krintel SB, Grunert VP, Hetland ML, Johansen JS, Rothfuss M, Palermo G, et al. The frequency of anti-infliximab antibodies in patients with rheumatoid arthritis treated in routine care and the associations with adverse drug reactions and treatment failure. Rheumatology. 2013 Jul;52(7):1245-53.
- 306. Pascual-Salcedo D, Plasencia C, Ramiro S, Nuno L, Bonilla G, Nagore D, et al. Influence of immunogenicity on the efficacy of long-term treatment with infliximab in rheumatoid arthritis. Rheumatology. 2011 Aug;50(8):1445-52.
- 307. Svenson M, Geborek P, Saxne T, Bendtzen K. Monitoring patients treated with anti-TNF-α biopharmaceuticals: assessing serum infliximab and anti-infliximab antibodies. Rheumatology. 2007;46(12):1828-34.
- 308. van den Bemt BJ, den Broeder AA, Snijders GF, Hekster YA, van Riel PL, Benraad B, et al. Sustained effect after lowering high-dose infliximab in patients with rheumatoid arthritis: a prospective dose titration study. Annals of the rheumatic diseases. 2008 Dec;67(12):1697-701.
- 309. van der Bijl AE, Breedveld FC, Antoni CE, Kalden JR, Kary S, Burmester GR, et al. An open-label pilot study of the effectiveness of adalimumab in patients with rheumatoid arthritis and previous infliximab treatment: relationship to reasons for failure and anti-infliximab antibody status. Clinical rheumatology. 2008 Aug;27(8):1021-8.
- 310. Wolbink GJ, Vis M, Lems W, Voskuyl AE, de Groot E, Nurmohamed MT, et al. Development of antiinfliximab antibodies and relationship to clinical response in patients with rheumatoid arthritis. Arthritis and rheumatism. 2006 Mar;54(3):711-5.

- 311. Bartelds GM, Wijbrandts CA, Nurmohamed MT, Stapel S, Lems WF, Aarden L, et al. Clinical response to adalimumab: relationship to anti-adalimumab antibodies and serum adalimumab concentrations in rheumatoid arthritis. Annals of the rheumatic diseases. 2007 Jul;66(7):921-6.
- 312. Radstake TRDJ, Svenson M, Eijsbouts AM, van den Hoogen FHJ, Enevold C, van Riel PLCM, et al. Formation of antibodies against infliximab and adalimumab strongly correlates with functional drug levels and clinical responses in rheumatoid arthritis. Annals of the rheumatic diseases. 2009;68(11):1739-45.
- 313. Rahman MU, Strusberg I, Geusens P, Berman A, Yocum D, Baker D, et al. Double-blinded infliximab dose escalation in patients with rheumatoid arthritis. Annals of the rheumatic diseases. 2007;66(9):1233-8.
- 314. St.Clair EW, Wagner CL, Fasanmade AA, Wang B, Schaible T, Kavanaugh A, et al. The relationship of serum infliximab concentrations to clinical improvement in rheumatoid arthritis: Results from ATTRACT, a multicenter, randomized, double-blind, placebo-controlled trial. Arthritis & Rheumatism. 2002;46(6):1451-9.
- 315. Wolbink GJ, Voskuyl AE, Lems WF, de Groot E, Nurmohamed MT, Tak PP, et al. Relationship between serum trough infliximab levels, pretreatment C reactive protein levels, and clinical response to infliximab treatment in patients with rheumatoid arthritis. Annals of the rheumatic diseases. 2005 May;64(5):704-7.
- 316. Garces S, Antunes M, Benito-Garcia E, da Silva JC, Aarden L, Demengeot J. A preliminary algorithm introducing immunogenicity assessment in the management of patients with RA receiving tumour necrosis factor inhibitor therapies. Annals of the rheumatic diseases. 2014 Jun;73(6):1138-43.
- 317. van Vollenhoven RF, Geborek P, Forslind K, Albertsson K, Ernestam S, Petersson IF, et al. Conventional combination treatment versus biological treatment in methotrexate-refractory early rheumatoid arthritis: 2 year follow-up of the randomised, non-blinded, parallel-group Swefot trial. Lancet. 2012 May 5;379(9827):1712-20.
- 318. Hider SL, Silman AJ, Thomson W, Lunt M, Bunn D, Symmons DP. Can clinical factors at presentation be used to predict outcome of treatment with methotrexate in patients with early inflammatory polyarthritis? Annals of the rheumatic diseases. 2009 Jan;68(1):57-62.
- 319. Patro PS, Singh A, Misra R, Aggarwal A. Myeloid-related Protein 8/14 Levels in Rheumatoid Arthritis: Marker of Disease Activity and Response to Methotrexate. The Journal of rheumatology. 2016 Apr;43(4):731-7.
- 320. Wessels JA, van der Kooij SM, le Cessie S, Kievit W, Barerra P, Allaart CF, et al. A clinical pharmacogenetic model to predict the efficacy of methotrexate monotherapy in recent-onset rheumatoid arthritis. Arthritis and rheumatism. 2007 Jun;56(6):1765-75.
- 321. Becker C, Barbulescu K, Hildner K, Meyer zum Buschenfelde KH, Neurath MF. Activation and methotrexate-mediated suppression of the TNF alpha promoter in T cells and macrophages. Annals of the New York Academy of Sciences. 1998 Nov 17;859:311-4.
- 322. Majumdar S, Aggarwal BB. Methotrexate suppresses NF-kappaB activation through inhibition of IkappaBalpha phosphorylation and degradation. J Immunol. 2001 Sep 01;167(5):2911-20.
- 323. Wadhwa M, Knezevic I, Kang HN, Thorpe R. Immunogenicity assessment of biotherapeutic products: An overview of assays and their utility. Biologicals : journal of the International Association of Biological Standardization. 2015 Sep;43(5):298-306.
- 324. Siljehult F, Arlestig L, Eriksson C, Rantapaa-Dahlqvist S. Concentrations of infliximab and antidrug antibodies in relation to clinical response in patients with rheumatoid arthritis. Scandinavian journal of rheumatology. 2018 Apr 27:1-6.
- 325. Ngo ST, Steyn FJ, McCombe PA. Gender differences in autoimmune disease. Frontiers in neuroendocrinology. 2014 Aug;35(3):347-69.
- 326. Eng GP, Bouchelouche P, Bartels EM, Bliddal H, Bendtzen K, Stoltenberg M. Anti-Drug Antibodies, Drug Levels, Interleukin-6 and Soluble TNF Receptors in Rheumatoid Arthritis

Patients during the First 6 Months of Treatment with Adalimumab or Infliximab: A Descriptive Cohort Study. PloS one. 2016;11(9):e0162316.