

MM3 Public Access

Author manuscript

Curr Osteoporos Rep. Author manuscript; available in PMC 2018 April 01.

Published in final edited form as:

Curr Osteoporos Rep. 2017 April; 15(2): 76–87. doi:10.1007/s11914-017-0354-3.

Bone Pain and Muscle Weakness in Cancer Patients

Daniel P. Milgrom, MD, Neha L. Lad, MD^* , Leonidas G. Koniaris, MD, and Teresa A. Zimmers, PhD

Department of Surgery, Indiana University School of Medicine, Indianapolis, IN

Abstract

Purpose of review—In this article we will discuss the current understanding of bone pain and muscle weakness in cancer patients. We will describe the underlying physiology and mechanisms of cancer-induced bone pain (CIBP) and cancier-induced muscle wasting (CIMW), as well as current methods of diagnosis and treatment. We will discuss future therapies and research directions to help patients with these problems.

Recent Findings—There are several pharmacologic therapies that are currenly in pre-clinical and clinical testing that appear to be promising adjuncts to current CIBP and CIMW therapies. Such therapies include resiniferitoxin, which is a targeted inhibitor of nociptive nerve fibers, and selective androgen receptor modulators, which show promise in increasing lean mass.

Summary—CIBP and CIMW are a significant causes of morbidity in affected patients. Current management is mostly palliative; however, targeted therapies are poised to revolutionize how these problems are treated.

Keywords

Cancer-induced bone	pain; muscle	weakness; b	one metastesis;	cachexia	

INTRODUCTION

With the rising incidence of cancer worldwide and advances in treatment, there has been an increase in the number of patients living with debilitating complications of chronic cancer. The most common cause of cancer-induced pain arises from bone metastases.[1] Of advanced cancer suffers, 60–84% are estimated to experience varying degrees of bone pain. [2] Cancer-induced bone pain (CIBP) involves both neuropathic and inflammatory pain pathways, associated with tumor, stroma, and adjacent tissues, including peripheral and central nerves.[1]

Compliance with Ethical Guidelines

Conflict of Interest

Leonidas Koniaris, Daniel Milgrom, Neha Lad, and Teresa Zimmers declare no conflict of interest.

Human and Animal Rights and Informed Consent

This article does not contain any studies with human or animal subjects performed by any of the authors.

Correspondence: Teresa A. Zimmers, PhD, Department of Surgery, R3-C518, Indiana University School of Medicine, Indianapolis, IN 46202, USA, zimmerst@iu.edu.

*Co-first author

The establishment of animal models have helped to elucidate the relationship between tumor, pain and neuronal interactions.[3] This in turn has helped in our understanding of this disease process and is helping to develop new targetted therapies to treat CIBP. Evaluation of patients with CIBP requires a comprehensive assessment of their current health status including the development of a trusting relationship, obtaining a thorough history of the pain, understanding the doses and durations of pain medications used to date, evaluation of psychological status, performing a thorough physical exam including neurologic exam, and reviewing diagnostic studies and laboratory findings. The ultimate goal is to develop an individualized treatment plan to obtain an acceptable quality of life.[2]

1.0 BONE PAIN IN CANCER

Myeloid leukemia, prostate, lung, and breast cancers are the malignancies most commonly associated with bone metastases. Although methods such as magnetic resonance imaging (MRI), computed tomography (CT) scan and 18-fluorodeoxyglucose positron emission tomography (FDG-PET/CT) may provide early diagnosis of bone metastases, the current treatment options remain mostly palliative and thus are generally reserved for patients once they become symptomatic. The importance of addressing symptoms must not be understated, as pain will drastically decrease quality of life (QoL), and furthermore, there is mounting evidence that survival for cancer patients is linked to symptom control.[4].

1.1 CLINICAL PRESENTATION AND ASSESSMENT OF CIBP

CIBP is one of the leading causes of significant morbidity in cancer patients. Early diagnosis and therapy are important to improve QoL. CIBP may present with symptoms that range from dull, vague, persistent pain to intermittent, sharp, severe pain and is generally exacerbated by physical activity. A careful history is required in making the diagnosis, whereas physical examination aided by various diagnostic modalities helps in confirmation of the pain's etiology. CIBP usually presents gradually and is progressive. It is usually related to weight bearing or movement, and develops into shooting neuropathic pain and pathologic fractures. Commonly involved sites are vertebrae, pelvis, femur, ribs and skull.[2] Patients also describe bouts of severe, intermittent pain despite analgesic intervention, called breakthrough pain, which is a sign of inadequate pain management.[5]

1.2 DIAGNOSTIC APPROACH TO BONE METASTASES

Biochemical findings, like elevated serum calcium, decreased renal function, increased urine calcium and urine hydroxyproline (an indirect measure of increased bone turn over), serum alkaline phosphatase level, and decreased osteocalcin (especially in multiple myeloma) aid in the diagnosis of bone involvement in cancer patients. Also, electrocardiography may demonstrate a shortened QT interval secondary to hypercalcemia. Diagnostic imaging with plain films (x-ray), bone scintigraphy (BS), MRI, CT scan and FDG-PET/CT are commonly used techniques. The diagnostic strategy is greatly influenced by pathology, available imaging modalities, and location of skeletal metastasis. A recent study from Denmark compared the diagnostic accuracy of the above modalities and with pathologic reports of bone biopsies. The sensitivity of MRI and FDG-PET/CT was better than CT, whereas CT

had higher specificity than FDG-PET/CT. For osteolytic and mixed lesions MRI and FDG-PET/CT were more sensitive as compared to CT and vice versa for osteosclerotic lesions. For spinal lesions, MRI had the highest sensitivity (92%) and specificity (80%); whereas for non-spinal lesions, FDG-PET/CT had the highest sensitivity (97%) and specificity (69%), but was not significantly different from MRI or CT. X-ray and BS were found to be inferior in diagnostic accuracy when compared to the other modalities.[6] In the case of an equivocal bone lesion in patients with hepatocellular carcinoma, single-photon emission computed tomography (SPECT) combined with spiral CT is found to be more accurate.[7]. 18F-NaF/FDG-PET/CT was found to be superior to whole body MRI and BS for evaluation skeletal disease in breast and prostate cancer, since it detects extra-skeletal disease which can significantly alter disease management.[8, 9].

There are inherent biologic and physical factors limiting the effectiveness of imaging technologies. Specificity is diminished by an inability to distinguish between metastatic tumor burden versus joint degeneration. Flare from increased radiotracer uptake in previously diagnosed, new, or undetected lesions after initiating therapy may also make image interpretation difficult. Scans are used to assess disease progression and response to therapy. However, by monitoring bone activity, a pleotropic drug which affects bone remodeling rather than cancer cells may lead to misinterpretation of results.[10] Scan duration, resolution and artifactual uptake are challenges which can be overcome by more disease-specific targeted imaging techniques.[10]

1.3 PATHOPHYSIOLOGY

Understanding the molecular aspects of the pathogenesis of bone metastases and subsequent complications associated with their development underlies the basis of developing targeted therapies. The development of tumor metastases involves sequential steps, including progressive tumor growth, vascularization, invasion, detachment, embolization, survival in circulation, arrest at site of metastasis, extravasation, evasion of host defense, and progressive growth.[11] There is disruption of the fundamental balance between osteoclasts, osteoblasts, and signaling pathways involved in controlling bone density. Osteoclasts and precursor osteoclasts express receptor activator of nuclear factor kappa β (RANK), ligand of RANK (RANKL) - the key stimulator of bone resorption, and cytokine osteoprotegerin (OPG), which inhibits bone resorption.[12, 13]

a. Osteolytic Metastases

Osteolytic metastases are more common than osteoblastic metasteses and are seen with breast and lung tumors and multiple myeloma. Metastatic cells produce many factors, such as parathyroid hormone related peptide (PTHrP), TGF- β , interleukins (ILs)- IL-1, IL-6, colony stimulating factor-1 (CSF-1), insulin-like growth factor-1 (IGF-1), prostaglandins, CXCR4, which interact with osteoblasts to modulate the RANK-RANKL pathway to stimulate osteoclast precursors and alter the microenvironment, starting the vicious cycle of osteolysis.[14]

PTHrP is known to be one of the most critical mediators of osteoclastic activation. It works by binding PTH-receptor 1 (PTH-R1), inducing RANKL expression and OPG

downregulation in osteoblasts.[12, 14-18] Deleted in Liver Cancer 1 (DLC1), a metastasis suppressor gene, acts through its RhoGTPase activating protein (RhoGAP) activity, which inhibits RhoA, RhoB, RhoC and cell division cycle 42 (cdc42) via hydrolysis of GTPase bound GTP.[19] DLC1-Rho signaling regulates osteoclastogenesis by blocking TGF-βinduced PTHrP secretion, and thus regulates metastatic colonization of circulating breast cancer cells. Experiments in mice have demonstrated enhanced bone metastasis in breast cancer cells lacking DLC1.[20] Data suggest that chemokine receptor CXCR4/CXCR7 and its ligand CXCL12/stromal derived factor- 1α (SDF- 1α) are highly expressed in skeletal metastases, especially in breast cancer cells. [15, 16, 21] Hypoxic microenvironments in bone (pO2 1–7%) stimulate hypoxia-inducible factor- 1α (HIF- 1α). HIF- 1α stimulates hypoxia response elements (HRE) and multiple factors, such as vascular endothelial growth factor (VEGF)[22], IGF-2, and CXCR4, have been implicated in metastatic bone colonization.[15] Hypoxia and constitutively active HIF-1a in MDA-MB-231 human breast cancer cells was found to be associated with increased osteoclast formation and decreased osteoblast differentiation, thereby promoting progression of bone metastasis. [23] Tumor hypoxia enhanced expression of connective tissue growth factor (CTGF) and IL-11, which initiate invasive angiogenesis and expression of hypoxia-associated genes, have been shown to contribute to the development of bone metastasis in hepatocellular carcinoma. [24]

b. Osteoblastic Metastases

Endothelin-1 (ET-1) growth factor induces osteoblastic proliferation via the ET-1A receptor (ETAR), as well as enhances expression of bone specific proteins osteocalcin, osteonectin, and alkaline phosphatase. Osteoblastic metastases from prostate and breast carcinoma are found to high levels of ET-1 and ETAR. The ETAR antagonist ABT-627 (Atrasentan) has been shown to block development of macroscopically evident osteoblastic metastasis.[15, 25, 26]

The Wnt family are cysteine-rich glycoproteins that mediate bone emybronic bone development and promote adult bone formation. They have autocrine and paracrine effects, enhancing proliferation and induction of osteoblastic activity in prostate cancer bone metastasis. Metastatic prostate cancer cells express the Wnt inhibitor dickkopf-1 (DKK-1) early in the development of skeletal metastasis. As disease progresses, DKK-1 expression decreases and unmasks Wnt osteoblastic activity, leading to osteosclerosis at metastatic sites. The initial osteolytic phase, mediated by DKK-1, RANKL, and PTHrP, causes an altered tumor environment. This leads to hypoxia and production of HIF-1α, VEGF and ET-1, thereby promoting osteoblastic activity.[27] A recent study showed significant inhibition of ERα signaling in prostate cancer cells *in vivo* leading to inhibition of osteoblastic lesions and formation of lung metastases.[28]

c. Hypercalcemia

Hypercalcemia is commonly seen in advanced stages of cancer.[29] PTHrP, which is secreted by various cells, mediates nearly 80% of malignancy-related hypercalcemia.[30] PTHrP acts on the same receptors as PTH in the bone, kidneys and intestine, increases bone resorption via RANKL, and increases calcium absorption in the intestine and reabsorption in the kidneys, leading to hypercalcemia.[31] Excessive calcium release from bone coupled

with abnormal retention of calcium in circulation and osteolytic metastases accounts for approximately 20% of malignancy-related hypercalcemia. [29, 30] Studies also demonstrate that in lymphomas and some other ovarian germ cell tumors, increased activity of 1α -hydroxylase and formation of 1,25-dihydroxycholecalciferol contributes to hypercalcemia. [29, 30]

d. Mechanism of Pain

While a multitude of factors contribute to pain caused by bone metastases, the exact mechanisms still remain unclear. The mechanisms hypothesized to cause bone pain are: a. stimulation of endosteal nerve endings resulting in destruction of bone tissue and release of chemical agents such as prostaglandins, bradykinin, substance P, and histamine; b. increasing stretch of the periosteum by enlarging tumors; c. fractures; and d. growth of tumor into surrounding tissues, especially nerves.[32]

Cancer cells promote proliferation and activity of bone-destroying osteoclasts via activation of the RANKL/RANK pathway. Osteoclast-mediated resorption of bone occurs through formation highly acidic resorption 'pits' between the osteoclasts and bone, stimulating the TRPV1 and ASIC3 channels expressed by a significant population of nerve fibers that drive bone cancer pain.[33–35] Mineralized bone undergoes loss of mechanical strength and stability due to the action of osteolytic and osteoblastic tumors. Extensive remodeling due to these effects can result in distortion from what would otherwise be an innocuous mechanical stress, activating the mechanosensitive nerve fibers of the bone.[33]

Cancer cells and surrounding stromal cells secrete a variety of factors (for example: bradykinin, endothelins, IL-6, nerve growth factor (NGF), and proteases), which sensitize or directly excite primary afferent neurons.[33] Studies have shown NGF to activate Trk-A-expressing sensory neurons directly, sensitizing TRPV1. The retrograde transport of NGF/TrkA complexes into nociceptor neurons induces and increases synthesis of the neurotransmitters substance P and calcitonin gene-related peptide, transcription factors (ATF3), and sodium channels, thus modulating supporting cells in dorsal root ganglia (DRG) and peripheral nerves.[33, 36–38]

Murine models of sarcoma, breast, and prostate derived bone cancer have shown active and pathological sprouting and neuroma formation by sensory and sympathetic nerve fibers that innervate the skeleton.[33] This sprouting requires NGF and sustained administration of anti-NGF or Pan Trk (Trk A, Trk B, Trk C). Inhibition of pathological sprouting and neuroma-like structure formation in sensory nerve fibers significantly inhibits pain generation.[33, 39–42] Several animal studies have shown sensitization of the spinal cord innervating the tumor bearing tissues by modification in levels of dynorphin, ATF3, astrocytes, microglia, c-Fos expression and substance P internalization.[33, 43]

1.4 THERAPEUTIC MANAGEMENT OF CIBP

The approach towards management of CIBP involves gradual escalation from conservative to interventional techniques based on response and severity of symptoms.

I. NON-PHARMACOLOGICAL APPROACH

The International Association for the Study of Pain defines pain as "an unpleasant sensory and emotional experience associated with actual or potential tissue damage, or described in terms of such damage." As such, cancer patients pain is highly influenced by various psychological and social factors. Interventions such as meditation, relaxation techniques, guided imagery, hypnosis, cognitive behavioral coping skills, therapist contact[44] form an essential part of a conjoint palliative approach.[45]

II. PHARMACOTHERAPY

Most physicians treat CIBP according to the World Health Organization's three-step ladder for cancer pain relief. This entails treating pain initially with a non-opiod medications, then escalating to opiods of increasing strength, and adding adjuvant therapies as needed as the patient's pain increases.[45]

- **a. Nonsteroidal Anti-inflammatory Drugs (NSAIDs)**—NSAID mechanism of action is via the inhibition of the cyclooxygenase [COX (COX1 & COX2)] enzymes, which catalyze formation of prostaglandins from arachidonic acid as a critical step in the inflammatory response.[3] COX-2 is highly expressed in tumor cells and peripheral macrophages around tumor cells, where it is involved in tumor cell invasion, migration and metastasis. COX-2 has been shown to reduce tumor burden in sarcoma-bearing bones in addition to reducing pain and bone destruction in an *in vivo* murine model.[46] However, various phase II and III trials found increased cardiovascular events from COX-2 inhibition, thereby tempering the enthusiasm for use of COX-2 inhibitors in cancer patients.[47, 48]
- **b. Opioids**—Opioids are one of the most frequently used analgesics for CIBP. They act on opioid G-protein coupled receptors (μ , κ , and δ), inhibiting substance P release in the dorsal horn.[47] Opiods are used by nearly 80% of cancer patients for pain control. Sustained and on-demand formulations are used in conjuction to provide pain suppression and breakthrough pain relief respectively. Opiods have limiting side effects of nausea, itching constipation, tolerance, development of addiction, and respiratory depression.[3, 45] Furthermore, a study in a murine model demonstrated prolonged exposure to opiods might worsen CIBP, accelerate bone loss, and increase incidence of spontaneous fractures.[49] Opioids synergize with NSAIDs, benzodiazapines, and anti-depressant therapies, which may improve pain control and limit the need for opioids. The concomitant use with benzodiazepines risks exacerbating some of the deleterious side effects of opioids, particularly respiratory depression. Increasingly, combination of low dose pregabalinantidepressants with opioids has been found to be effective in the management of neuropathic CIBP.[50]
- **c. Bisphosphonates**—Older generation bisphosphonates (i.e. clodronate and etidronate) were metabolized by osteoclasts into cytotoxic ATP analogs, interfering with mitochondrial membrane, potentially leading to osteoclast apoptosis. Newer generation nitrogen-containeing intravenous bisphosphonates (i.e. pamidronate, ibandronate and zoledronate) are internalized by osteoclasts and inhibit the farnesyl pyrophosphate (FPP) synthase enzyme, active in prenylation of several GTPases involved in bone resorption. This causes

accumulation of isopentyl pyrophosphate and adenosine monophosphate, which conjugate to form an endogenous ATP analogue that inhibits mitochondrial adenine nucleotide translocase (ANT) and leads to osteoclast apoptosis.[51] Multiple studies have demonstrated efficacy of bisphosphonates in reducing skeletal complications and CIBP.[52] Zoledronate, in addition to reducing skeletal morbidity, has been reported to have direct antitumor properties: induction of tumor cell apoptosis, inhibition of cancer cell invasion[53], limiting metastatic outgrowth in visceral tissues[51, 54], and causing decrease in VEGF levels, thereby potentially slowing bone disease.[55, 56] While bisphosphonates are useful adjuvants in the treatment of CIBP, they do not themselves block pain, and must be used in conjuction with other therapies.[57] Intravenous bisphosphante therapy requires dental evaluation and follow-up to monitor for osteonecrosis of the jaw as well as renal monitoring and caution for patients with kidney disease or who are taking other nephrotoxic agents.[51]

d. Novel Targeted Therapies

Denosumab: Denosumab is a monoclonal antibody against RANKL. It is a potent inhibitor of osteoclast-mediated bone resorption. Multiple phase III trials have shown an increase in bone mineral density, a decreased risk of fractures, and a delay in skeletal-related events (SRE) with use of denosumab [15]; however, data regarding overall survival remains controversial.[58] Denosumab was superior to zolendronate in preventing SRE in patients with advanced disease regardless of performance status and disease extent.[59]

Atrasentan: ET-1 receptor sensitization and/or activation has been associated with hyperalgesia of CIBP. The ET1A receptor antagonist atrasentan is under phase II trials for bone metastates in renal carcinoma patients.[60] However, a meta-analysis of its use in prostate cancer showed a significant decrease in CIBP and SRE with a delayed rise in PSA and bone alkaline phosphatase.[61]

Osteoprotegerin: OPG combines with RANKLto inhibit activation of RANK on osteoclasts, thereby preventing bone destruction induced by tumor cells.[3, 62] OPG has been shown to have inhibitory potential in breast cancer-induced bone destruction.[63]

<u>Dasatinib</u>: Src is a prototypic member of a nonreceptor tyrosine kinase family that is involved in various critical cellular functions, including cell morphology, cell growth, proliferation, differentiation, adhesion, migration and survival.[15] Dasatinib, a Src inhibitor, has been show to reduce metastatic potential and induce apoptosis in preclinical studies of pancreas, head and neck, and lung cancers.[15] Also, *in vivo* and *in vitro* studies show suppression of Src causes inhibition of breast cancer cells and reduced incidence of metastasis.[64]

<u>Anti-NGF</u>: New potential therapies in clinical phases of development target molecules like NGF, a molecule which is integrally involved in the upregulation, sensitization and disinhibition of neurotransmitters in the primary afferent nerves. Anti-NGF therapy could be effective in blocking CIBP due to NGF.[51]

Resiniferatoxin (RTX): RTX is an ultrapotent agonist of the transient receptor potential vanilloid 1 (TRPV1) receptor. RTX acts on TRPV1 to allow for prolonged calcium influx, inducing cytotoxicity and death selectively in nociceptive fibres expressing TRPV1. Treatment with RTX has been shown to significantly improveme pain control in dogs with CIBP compared to standard of care analgesic therapy.[65]

Other therapies: Preclinical studies show TRPV1 and cannabinoid 2 receptor agonists could be used as adjuncts to ameliorate opioid side effects.[51] CXCL12/CXCR4 is found to play a central role in cancer cell proliferation, invasion and dissemination in various malignancies and is a potential drug-target in cancer management.[66]

III. RADIATION THERAPY (RT)

- a. External Beam Radiation Therapy (EBRT)—EBRT is the most common form of RT used for palliation of CIBP. EBRT is produced in a linear accelerator which projects electrons onto a tungsten target, producing megavoltage photons directed towards bone lesions. The treatment usually takes 10–15 minutes per dose and relief can be acheived in 50–80% patients. The acute side effects of radiation therapy are generally self-limiting and consist mainly of fatigue. Late side-effects in this patient group are relatively uncommon given the short life expectancy.[67] A systematic review of 24 randomized control trials (RCT) showed that single fraction administration of 8 Gy was statistically superior in pain response with minimal iatrogenic toxicity.[68] Evaluation of QoL following RT for patients with CIBP found improvement in symptoms and function using the Brief Pain Inventory score in all 17 studies included in the analysis.[69] Post-operative RT after surgical stabilization of metastatic bone disease has been found to be effective in local disease control. Along with bisphosphonates it might have the additional effect of delaying local progression.[70] Dexamethasone has been shown to reduce radiation-induced pain flare in the treatment of painful bone metastases in a double-blind randomized control trial.[71]
- **b. Stereotactic Body Radiation Therapy (SBRT)**—SBRT uses image-guidance technology to deliver single or multiple fractions of high dose RT and can deliver nearly 2 to 7-times the standard palliative dose.[72] A systematic review showed SBRT provided excellent local control with lower toxicity in patients with metastatic renal cell carcinoma. [73] Although pain relief is higher in in SBRT, cost effectiveness of SBRT in comparison with EBRT in patients with a shorter expected survival (<11 months) remains contested.[74]
- **c. Role of Re-irradiation**—Re-irradiation is effective and comparable to initial RT and should be recommended to patients suffering from ongoing CIBP irrespective of initial response to RT.[75]
- **d. Radioisotopes**—*Strontium-89* (S^{89}) is a beta-emitting radioisotope with a half-life of 50.5 days. Osteoblastic bone metastases have higher uptake than surrounding bone. Therefore S^{89} is used for the treatment of metastatic prostate or breast cancer with significant pain relief in 60–92%.[76]

Samarium-153 (Sm^{153}) is a beta-emitter with a half-life of 1.9 days. It is chelated to ethylene diamine tetramethylene phosphate (EDTMP) which targets bone matrix as

pyrophosphate. It is used in various primary tumors and confers a superior survival in breast cancer patients at a higher dose.[76]

Other isotopes like Tin-117m (Sn^{117m}), Radium-223 (Ra²²³), and Rhenium-186(Re¹⁸⁶) are being tested in various clinical trials for CIBP in prostate and breast cancer. All of these therapies have distinct advantages over EBRT, which requires large areas of RT and is limited by toxicity.

e. MRI-guided Focused Ultrasound Ablation—Ultrasound ablation is a promising alternative therapy that was first tested in uterine fibroids. It uses non-invasive, non-ionizing ultrasound for pain palliation and tumor control. The interaction between the ultrasound beam and tissues results in a rise in cell temperature, leading to coagulative necrosis at a thermal range of 65–85 degrees C, which is limited to focal tissue volumes of 0.2–5mm³ and has negligible effect on surrounding tissue. The major advantages of this technology are the ability to be performed in an outpatient setting with three-dimensional MRI visualization for precise planning, continuous temperature mapping with MR thermometry, and immediate post-treatment assessment.[77, 78]

IV. INVASIVE PROCEDURES

a. Surgical Management—There is a significant palliative role for surgery in patients with CIBP in conjunction with other modalities. Surgical intervention is usually indicated for impending pathologic fractures, spine instability that threatens spinal cord function, or the development of nerve deficits. Based on pathology and patient prognosis, interventions may range from conservative measures to fracture stabilization with internal fixation or arthroplasty.[79] A systematic review assessed pain and functional outcomes following surgical management of metastases to the humerus, femur and pelvis and found pain relief in 93, 91, and 93% of subjects respectively and improved function in 94, 89 and 94% of subjects respectively. In this study, there was a also substantial risk of perioperative complications (17%) and mortality (4%).[80]

Vertebroplasty is a technique involving fluoroscopic, percutaneous injection of polymethylmethacrylate and bone cement into the vertebral body for stabilization and pain relief in patients with compression fractures. *Kyphoplasty* involves placement of an inflatable balloon into the vertebral body with subsequent injection of bone cement. Both procedures can be performed under local or general anesthesia and are shown to provide effective and safe reduction in pain and improvement in mobility.[81] For palliative treatment of spinal metastases, the increasing use of minimally invasive techniques of tumor resection and decompression of neurologic elements have resulted in improved recovery with minimal morbidity and mortality.[82]

b. Intra-thecal analgesia—For patients requiring higher doses of opioids with unacceptable systemic side-effects, intra-thecal therapy may be a good alternative. Various intra-thecal analgesics including morphine sulphate, hydromorphone, and bupivacaine have proven efficacity.[83]

c. Laser-induced thermotherapy—The use of Nd:YAG laser has been reported in a small case series as a treatment of treat spinal metastasis under CT—guidance. A total of 1400–2600J energy delivered over 60–90 minutes has yielded 30–45% reductions in CIBP without complications.[84]

2.0 MUSCLE WEAKNESS IN CANCER

Cancer cachexia is a complex metabolic condition characterized by skeletal muscle wasting (with or without fat loss), anemia, reduced caloric intake, and altered immune function, which contributes to increased disability, fatigue, diminished QoL, and reduced survival.[85, 86] Skeletal muscle wasting and resultant functional impairment significantly affect QoL. Cancer-related muscle loss is multifactorial, resulting in asthenia and functional impairment similar to that seen in patients with age-related sarcopenia as well that manifested by active muscle break-down.[87, 88] The common metabolic abnormalities to cancer cachexia and sarcopenia include altered hormone levels, elevated cytokines, increased insulin resistance, increased muscle proteolysis, elevated acute phase proteins, and altered nutrient utilization. [87] Many experts believe, however, that muscle loss in cancer is a more active process, mediated by a number of pro-inflammatory cytokines, as well as members of the $TGF\beta$ -superfamily including activins[89] and myostatin.[90, 91]

2.1 CLINICAL PRESENTATION AND ASSESSMENT OF CANCER INDUCED MUSCLE WEAKNESS (CIMW)

CIMW is one of the major symptoms of cancer cachexia. CT and DXA imagine can be used to quantify sarcopenia which correlates with clinical asthenia, fatigue, reduced tolerance to treatments, impaired QoL and reduced survival.[88]

2.2 PATHOPHYSIOLOGY OF CIMW

The skeletal muscle loss due to cachexia results from decreased protein anabolism, increased proteolysis, or a combination of both. The four major proteolytic pathways in skeletal muscle are:

- The lysosomal system, which includes the cysteine proteases and cathepsins B, H, and, L, as well as aspartate protease cathepsin D, mainly degrades extracellular proteins and cell receptors.
- **2.** The calcium-activated calpains I and II, which mainly cause tissue injury, necrosis and autolysis.
- 3. The ATP-dependent ubiquitin proteasome proteolytic pathway, which works with the calpain system to degrade myofilaments. This pathway plays a predominant role in degradation of myofibrillar proteins particularly in patients with a weight loss of >10%.[92]
- 4. The STAT3 pathway, which directly induces myocyte atrophy.[93, 94]. It induces muscle-specific E3 ubiquitin ligases [e.g. muscle atrophy F box (encoded by

MAFbx/atrogin-1) and muscle RING finger 1 (MuRF1)], which cause polyubiquitation of proteins targeted for degradation.[95]

Cachexia is known to feature tumor-induced activation of the host immune system and elevated proinflammatory cytokines IL-1 β , IL-6, interferon(IFN- γ), TNF- α , and proteolysis inducing factor (PIF), all of which may primarily stimulate a catabolic state in skeletal muscle.[85, 96, 97] On a subcellular level, skeletal muscle weakness in cancer is due to a decrease in the number of strongly bound cross-bridges and a reduction in myosin-actin cross-bridge kinetics characterized by an increased myosin attachment time.[98] Chemotherapeutic agents like doxorubicin cause increased oxidative stress via formation of reactive oxygen species and activate caspases leading to loss of muscle mass and atrophy via the E3 ubiquitin-ligase proteasome pathway.[99] The mechanism of muscle wasting involves multiple host and tumor factors, decreased levels of testosterone and IGF-1, and decreased food intake contributing to both antianabolic and procatabolic processes.[85]

2.3 CURRENT THERAPEUTIC APPROACH TO CIMW

Unlike starvation, cancer cachexia does not respond to nutritional supplementation. Although caloric replacement up to 1.5mg/kg has shown some benefit in stabilizing weight, [100] benefits of nutritional supplementation may be limited.[85] Essential amino acid (EAA) supplements, including ~2.5g of leucine, HMB supplements and vitamin D may improve muscle mass and function parameters.[101] Exercise therapy can help maintain or slow the loss of physical function.[101]

2.4 CURRENT AND POTENTIAL PHARMACOLOGIC THERAPIES FOR CIMW

5-HT3 antagonists

Mirtazapine and olanzapine provide 24-hour nausea control and increased appetite in cancer patients. They have the added benefit of controlling anxiety and aiding with better sleep.[96]

Megesterol acetate

This progetagen, combined with *thalidomide*, an anti-TNFa agent, has shown to significantly increase appetite with consequent improvement in body weight and QoL due to anti-inflammatory properties.[102] Megesterol carries an increased risk of thromboembolism, while thalidomide is known to cause birth defects in pregnant patients.

Enobosam

This nonsteroidal, selective androgen receptor modulators (SARMs), is in a phase III trial. Treatment with this medication has demonstrated increased lean body mass and is promising as an agent for the prevention and treatment of skeletal muscle wasting.[103] This medication prevents the need for non-selective systemic steroids, which carry significant side effects.

Ghrelin analogues

Anamorelin, is an oral ghrelin-receptor agonist with appetite-enhancing and anabolic effects, which has shown promising results in phase III trials.[104]

Myostatin antagonists

Using a soluble receptor antagonist of myostatin (sActRIIB) in cachectic tumor-bearing animals has shown improvement in muscle weight and force through myostatin blockade. [105]

β2 adrenoreceptor-selective agonist

Formoterol, promotes muscle growth and skeletal muscle hypertrophy in animal models. Espindolol, a nonspecific $\beta 1$ and $\beta 2$ adrenoreceptor antagonist with intrinsic sympathomimetic activity at the $\beta 2$ adrenoreceptor has a novel anabolic-catabolic transforming property. These are prospective new drugs particularly beneficial for patients suffering from cancer cachexia with declined cardiac function.[106] Combination of sActRIIB with formoterol appears to be very promising in animal studies.[107]

3.0 ALTERATION OF BONE AND MUSCLE PHYSIOLOGY IN CANCER

CIMW is a major clinical problem in advanced stage cancer, and is usually associated with bone pain, fractures, hypercalcemia and nerve compression. [87] Bone and muscle function are interdependent physiologically. However, in cancer patients, accelerated bone resorption due to metastases increases "osteokines," which significantly alter muscle function. Similarly, factors released from muscle can further exacerbate bone's role in muscle dysfunction. [87] Normal excitation-contraction (E-C) coupling in skeletal muscle involves release of sequestered calcium from the sarcoplasmic reticulum into the cytoplasm via the activated ryanodine receptor/calcium release channel (RyR1), leading to calcium-dependent actin-myosin cross-bridging and muscle contraction. [108] Modifications to RyR1 from chronic oxidative stress causes disruption of RyR1 and its stabilizing subunit calstabim1, resulting in leaky calcium channels. In addition, $TGF\beta$, a critical bone remodeling factor can mediate oxidative stress, and thereby further contribute to muscle dysfunction. [87]

3.1 Muscle Dysfunction Associated with Bone Metastasis in Cancer

Muscle secretes many factors, collectively called "myokines," which affect other tissues. They include bone active molecules like IGF-1 and FGF-2, myostatin (also called growth differentiation factor 8 [GDF8])[109], and IL-6.[110–113] IGF-1 and FGF-2 stimulate bone formation,[114, 115] and myostatin deficiency increases bone density.[116, 117] Conversely, Indian hedgehog (Ihh) promotes myoblast survival and myogenesis in mouse and chick embryos.[87, 118] Preclinical mice model data show that predominantly osteolytic MDA-MB-231 breast cancer, A549 lung cancer, PC3 prostate cancer and JJN3 multiple myeloma or mixed osteolytic/osteoblastic bone metastases result in lower muscle specific force, lower muscle strength, and RyR1 modifications consistent with leaky calcium channels regardless of weight loss or lower muscle mass as compared to non-tumor bearubg mice.[119] This suggets that there is a relationship between tumor-induced osteolysis-linked alterations in the bone microenvironment and skeletal muscle dysfunction. The RyR1 calcium release channel stabilizer Rycal (S107) improves the function of the leaky RyR1 channels by inhibiting oxidation-induced depletion of channel stabilizing subunit catstabin1 from the RyR1 complex, thereby stabilizing the closed state of the channels and preventing aberrant

calcium leakage. Experiments of S107 function have shown improved forelimb grip strength in mice with breast cancer bone metastases. However, S107 does not affect development or progression of bone metastasis, tumor burden, body weight, muscle mass, or distribution of fat and lean mass compared to vehicle treated mice.[119] Thus, S107 treatment suggests that there is no direct correlation between bone destruction and reduced muscle function. Further studies are needed to assess the potential role of S107 in clinical practice to improve CIMW.

3.2 Bone Derived Factor(s) causing Muscle Dysfunction

Bone matrix stores many growth factors known to affect muscle, such as Activin A, $TGF\beta$, IGF-1, and bone morphogenic protein 2 (BMP-2).[111] The high affinity Activin type 2 receptor, ActRIIB, mediates signaling of a small group of $TGF\beta$ family members (Activin A, myostatin, GDF-11) and plays major role in regulating muscle mass.[113, 120, 121] In a murine model of cachexia, ActRIIB blockade prevents muscle wasting, induces muscle satellite cell mobilization and differentiation, and significantly prolongs survival.[122] However, it remains unclear from these studies if the effect is due to blockade of Activin A, myostatin or GDF-11 signaling due to receptor overlap.[111, 123]

During osteoclastic resorption, TGFβ is released from mineralized bone. In MDA-MB-231 mice with bone metastases, TGFβ was shown to induce more SMAD3 phosphorylation in skeletal muscle compared to mice without metastases.[119, 124] TGFβ-1 receptor kinase inhibitor (SD-208), bisphosphonates (e.g. zoledronate - which inhibit osteoclastic resorption thereby lowering the release of TGF β), and a pan-TGF β neutralizing antibody (clone 1D11), have all shown a decrease in TGF\$\beta\$ in various experimental models, either in combination or alone. This in turn lowered skeletal muscle SMAD3 phosphorylation and preserved calstabin 1 binding to RyR1 complex, resulting in improved muscle function. Combination therapy showed additional benefit by lowering tumor burden and number of osteoclasts. TGFB inhibition improves muscle function, and bone-derived TGFβ contributes to CIMW, at least in part by inducing oxidation of RyR1.[119] TGFβ released from the bone matrix due to increased catabolism upregulates membrane protein Nox4 in the sarcoplasmic reticulum. Nox4 oxidizes the RyR1 channel and causes a calcium leak, lowering tetanic calcium, impairing muscle force production and contributing to muscle weakness in cancer with bone metastases. GKT137831m, a Nox1/Nox4 inhibitor, prevents skeletal muscle oxidation and nitrosylation of RyR1, restores calstabin binding, and improves extensor digitorum longus force in mice with MDA-MB-231 bone metastases as compared to vehicle treated mice; however, it did not block upstream TGFβ signaling and SMAD3 phosphorylation. In addition, it has no effect on osteolytic lesion size, muscle mass, body weight, or grip strength. Thus, targeting skeletal muscle weakness caused by the TGFβ-Nox4-RyR1 axis represents a novel therapeutic approach for patients.[125]

IGF-1 stimulates myogenic cell proliferation and differentiation[126, 127], while BMP-2 signaling leads to muscle hypertrophy and thereby regulates muscle mass.[111] These are potential targets for restoring muscle mass. Additionally, vitamin D repletion may help functional status, as vitamin D deficiency studied in rodent models using vitamin D receptor knock-out (VDRKO) mice resulted in an increase in sinking episodes in a forced swim test. [111, 128]

MicroRNAs (miRNA) *in vivo* inhibit osteoclast activity and reduce osteolytic bone metastasis. Serum levels of soluble intracellular adhesion molecule (sICAM1) correlate with bone metastasis burden. These levels are affected by activation of NFκB signaling by bone metastatic cancer cells[129], and two osteoclast mRNAs, miR-16 and miR-378[130], which are elevated in osteoclast differentiation,. Hence, miRNAs could be potential therapeutic targets and clinical biomarkers of bone metastases.[130, 131]

CONCLUSION

The closely interrelated bone and muscle physiology is altered in cancer patients. The myokines secreted by skeletal muscle cells significantly impact the surrounding bone. Likewise, bone releases multiple growth factors during physiologic remodeling and affects muscle function. The metastasis of tumor cells to bone causes disruption between osteoclasts and osteoblasts along with various signaling pathways. The alteration of the microenvironment due to increased proinflammatory cytokines released from osteolytic bone resorption accelerates myofibrillar degradation and apoptosis. Clinically this manifests as a spectrum ranging from muscle weakness and fatigue to cachexia in skeletal muscle accompanied by bone pain, fractures, and neuropathy. Diagnosis is mainly clinical, while imaging and biochemical studies may aid in cases of challenging cases. Although the primary approach remains conservative, various therapeutic interventions have been formulated based on factors involving the metabolism of bone and skeletal muscle. Novel therapeutic agents targeting the molecular mechanism appear to be promising. Further studies are needed identify the exact mechanisms of the different cancers that metastasize to bone and interplay between bone and muscle to help develop effective targeted therapies.

References

Papers of particular interest, published recently, have been highlighted as:

- •Of importance
- ••Of major importance
- 1. Falk S, Bannister K, Dickenson AH. Cancer pain physiology. Br J Pain. 2014; 8(4):154–62. [PubMed: 26516549]
- 2. Mercadante S. Malignant bone pain: pathophysiology and treatment. Pain. 1997; 69(1–2):1–18. [PubMed: 9060007]
- 3. Zhu XC, et al. Advances in cancer pain from bone metastasis. Drug Des Devel Ther. 2015; 9:4239–45.
- 4. Irwin KE, et al. Early palliative care and metastatic non-small cell lung cancer: potential mechanisms of prolonged survival. Chron Respir Dis. 2013; 10(1):35–47. [PubMed: 23355404]
- 5. Mercadante S, Arcuri E. Breakthrough pain in cancer patients: pathophysiology and treatment. Cancer Treat Rev. 1998; 24(6):425–32. [PubMed: 10189409]
- Lange MB, et al. Diagnostic accuracy of imaging methods for the diagnosis of skeletal malignancies: A retrospective analysis against a pathology-proven reference. Eur J Radiol. 2016; 85(1):61–7. [PubMed: 26724650]
- 7. Zhang Y, et al. The added value of SPECT/spiral CT in patients with equivocal bony metastasis from hepatocellular carcinoma. Nuklearmedizin. 2015; 54(6):255–61. [PubMed: 26615876]

8. Minamimoto R, et al. Prospective Comparison of 99mTc-MDP Scintigraphy, Combined 18F-NaF and 18F-FDG PET/CT, and Whole-Body MRI in Patients with Breast and Prostate Cancer. J Nucl Med. 2015; 56(12):1862–8. [PubMed: 26405167]

- 9. Iagaru A, et al. Prospective evaluation of (99m)Tc MDP scintigraphy, (18)F NaF PET/CT, and (18)F FDG PET/CT for detection of skeletal metastases. Mol Imaging Biol. 2012; 14(2):252–9. [PubMed: 21479710]
- Ulmert D, Solnes L, Thorek D. Contemporary approaches for imaging skeletal metastasis. Bone Res. 2015; 3:15024. [PubMed: 26273541]
- 11. Fidler IJ, Radinsky R. Genetic control of cancer metastasis. J Natl Cancer Inst. 1990; 82(3):166–8. [PubMed: 2296042]
- 12. Middlemiss T, Laird BJ, Fallon MT. Mechanisms of cancer-induced bone pain. Clin Oncol (R Coll Radiol). 2011; 23(6):387–92. [PubMed: 21683564]
- 13. Roodman GD. Biology of osteoclast activation in cancer. J Clin Oncol. 2001; 19(15):3562–71. [PubMed: 11481364]
- 14. Kakonen SM, Mundy GR. Mechanisms of osteolytic bone metastases in breast carcinoma. Cancer. 2003; 97(3 Suppl):834–9. [PubMed: 12548583]
- 15. Papachristou DJ, Basdra EK, Papavassiliou AG. Bone metastases: molecular mechanisms and novel therapeutic interventions. Med Res Rev. 2012; 32(3):611–36. [PubMed: 20818675]
- 16. Kozlow W, Guise TA. Breast cancer metastasis to bone: mechanisms of osteolysis and implications for therapy. J Mammary Gland Biol Neoplasia. 2005; 10(2):169–80. [PubMed: 16025223]
- 17. Guise TA, et al. Basic mechanisms responsible for osteolytic and osteoblastic bone metastases. Clin Cancer Res. 2006; 12(20 Pt 2):6213s–6216s. [PubMed: 17062703]
- 18. Schramek D, Sigl V, Penninger JM. RANKL and RANK in sex hormone-induced breast cancer and breast cancer metastasis. Trends Endocrinol Metab. 2011; 22(5):188–94. [PubMed: 21470874]
- 19. Weidle UH, et al. Molecular Mechanisms of Bone Metastasis. Cancer Genomics Proteomics. 2016; 13(1):1–12. [PubMed: 26708594]
- 20. Wang Y, et al. DLC1-dependent parathyroid hormone-like hormone inhibition suppresses breast cancer bone metastasis. J Clin Invest. 2014; 124(4):1646–59. [PubMed: 24590291]
- 21. Wu W, et al. Prognostic significance of CXCL12, CXCR4, and CXCR7 in patients with breast cancer. Int J Clin Exp Pathol. 2015; 8(10):13217–24. [PubMed: 26722521]
- 22. Min Y, et al. C/EBP-delta regulates VEGF-C autocrine signaling in lymphangiogenesis and metastasis of lung cancer through HIF-1alpha. Oncogene. 2011; 30(49):4901–9. [PubMed: 21666710]
- 23. Hiraga T, et al. Hypoxia and hypoxia-inducible factor-1 expression enhance osteolytic bone metastases of breast cancer. Cancer Res. 2007; 67(9):4157–63. [PubMed: 17483326]
- 24. Gao YB, et al. Enhanced production of CTGF and IL-11 from highly metastatic hepatoma cells under hypoxic conditions: an implication of hepatocellular carcinoma metastasis to bone. J Cancer Res Clin Oncol. 2013; 139(4):669–79. [PubMed: 23307318]
- 25. Mohammad KS, Guise TA. Mechanisms of osteoblastic metastases: role of endothelin-1. Clin Orthop Relat Res. 2003; (415 Suppl):S67–74. [PubMed: 14600594]
- 26. Thakkar SG, Choueiri TK, Garcia JA. Endothelin receptor antagonists: rationale, clinical development, and role in prostate cancer therapeutics. Curr Oncol Rep. 2006; 8(2):108–13. [PubMed: 16507220]
- 27. Hall CL, et al. Role of Wnts in prostate cancer bone metastases. J Cell Biochem. 2006; 97(4):661–72. [PubMed: 16447163]
- 28. Mishra S, et al. Estrogen and estrogen receptor alpha promotes malignancy and osteoblastic tumorigenesis in prostate cancer. Oncotarget. 2015
- 29. Mirrakhimov AE. Hypercalcemia of Malignancy: An Update on Pathogenesis and Management. N Am J Med Sci. 2015; 7(11):483–93. [PubMed: 26713296]
- 30. Stewart AF. Clinical practice Hypercalcemia associated with cancer. N Engl J Med. 2005; 352(4): 373–9. [PubMed: 15673803]
- 31. Wysolmerski JJ. Parathyroid hormone-related protein: an update. J Clin Endocrinol Metab. 2012; 97(9):2947–56. [PubMed: 22745236]

32. Nielsen OS, Munro AJ, Tannock IF. Bone metastases: pathophysiology and management policy. J Clin Oncol. 1991; 9(3):509–24. [PubMed: 1705581]

- 33. Mantyh PW. Bone cancer pain: from mechanism to therapy. Curr Opin Support Palliat Care. 2014; 8(2):83–90. [PubMed: 24792411]
- 34. Honore P, Mantyh PW. Bone cancer pain: from mechanism to model to therapy. Pain Med. 2000; 1(4):303–9. [PubMed: 15101876]
- 35. Clohisy DR, Perkins SL, Ramnaraine ML. Review of cellular mechanisms of tumor osteolysis. Clin Orthop Relat Res. 2000; (373):104–14.
- 36. Gould HJ 3rd, et al. A possible role for nerve growth factor in the augmentation of sodium channels in models of chronic pain. Brain Res. 2000; 854(1–2):19–29. [PubMed: 10784102]
- 37. Ji RR, et al. p38 MAPK activation by NGF in primary sensory neurons after inflammation increases TRPV1 levels and maintains heat hyperalgesia. Neuron. 2002; 36(1):57–68. [PubMed: 12367506]
- 38. Obata K, et al. Expression of neurotrophic factors in the dorsal root ganglion in a rat model of lumbar disc herniation. Pain. 2002; 99(1–2):121–32. [PubMed: 12237190]
- 39. Mantyh WG, et al. Blockade of nerve sprouting and neuroma formation markedly attenuates the development of late stage cancer pain. Neuroscience. 2010; 171(2):588–98. [PubMed: 20851743]
- 40. Ghilardi JR, et al. Neuroplasticity of sensory and sympathetic nerve fibers in a mouse model of a painful arthritic joint. Arthritis Rheum. 2012; 64(7):2223–32. [PubMed: 22246649]
- 41. Bloom AP, et al. Breast cancer-induced bone remodeling, skeletal pain, and sprouting of sensory nerve fibers. J Pain. 2011; 12(6):698–711. [PubMed: 21497141]
- 42. Jimenez-Andrade JM, et al. Pathological sprouting of adult nociceptors in chronic prostate cancer-induced bone pain. J Neurosci. 2010; 30(44):14649–56. [PubMed: 21048122]
- 43. Schwei MJ, et al. Neurochemical and cellular reorganization of the spinal cord in a murine model of bone cancer pain. J Neurosci. 1999; 19(24):10886–97. [PubMed: 10594070]
- 44. Syrjala KL, Cummings C, Donaldson GW. Hypnosis or cognitive behavioral training for the reduction of pain and nausea during cancer treatment: a controlled clinical trial. Pain. 1992; 48(2): 137–46. [PubMed: 1350338]
- 45. Schneider G, Voltz R, Gaertner J. Cancer Pain Management and Bone Metastases: An Update for the Clinician. Breast Care (Basel). 2012; 7(2):113–120. [PubMed: 22740797]
- 46. Sabino MA, et al. Simultaneous reduction in cancer pain, bone destruction, and tumor growth by selective inhibition of cyclooxygenase-2. Cancer Res. 2002; 62(24):7343–9. [PubMed: 12499278]
- 47. Gough N, Miah AB, Linch M. Nonsurgical oncological management of cancer pain. Curr Opin Support Palliat Care. 2014; 8(2):102–11. [PubMed: 24675403]
- 48. Antman EM, et al. Use of nonsteroidal antiinflammatory drugs: an update for clinicians: a scientific statement from the American Heart Association. Circulation. 2007; 115(12):1634–42. [PubMed: 17325246]
- 49. King T, et al. Morphine treatment accelerates sarcoma-induced bone pain, bone loss, and spontaneous fracture in a murine model of bone cancer. Pain. 2007; 132(1–2):154–68. [PubMed: 17706870]
- 50. Nishihara M, et al. Combinations of low-dose antidepressants and low-dose pregabalin as useful adjuvants to opioids for intractable, painful bone metastases. Pain Physician. 2013; 16(5):E547–52. [PubMed: 24077205]
- 51. Pantano F, et al. New targets, new drugs for metastatic bone pain: a new philosophy. Expert Opin Emerg Drugs. 2011; 16(3):403–5. [PubMed: 21623686]
- 52. Santini D, et al. Zoledronic acid in the management of metastatic bone disease. Expert Opin Biol Ther. 2006; 6(12):1333–48. [PubMed: 17223741]
- 53. Boissier S, et al. Bisphosphonates inhibit breast and prostate carcinoma cell invasion, an early event in the formation of bone metastases. Cancer Res. 2000; 60(11):2949–54. [PubMed: 10850442]
- 54. Hiraga T, et al. Zoledronic acid inhibits visceral metastases in the 4T1/luc mouse breast cancer model. Clin Cancer Res. 2004; 10(13):4559–67. [PubMed: 15240548]

55. Santini D, et al. Repeated intermittent low-dose therapy with zoledronic acid induces an early, sustained, and long-lasting decrease of peripheral vascular endothelial growth factor levels in cancer patients. Clin Cancer Res. 2007; 13(15 Pt 1):4482–6. [PubMed: 17671133]

- Vincenzi B, et al. Zoledronic acid-related angiogenesis modifications and survival in advanced breast cancer patients. J Interferon Cytokine Res. 2005; 25(3):144–51. [PubMed: 15767788]
- 57. Coleman RE. Impact of Bone-Targeted Treatments on Skeletal Morbidity and Survival in Breast Cancer. Oncology (Williston Park). 2016; 30(8)
- 58. Raje N, et al. Evaluating results from the multiple myeloma patient subset treated with denosumab or zoledronic acid in a randomized phase 3 trial. Blood Cancer J. 2016; 6:e378. [PubMed: 26745852]
- 59. Lipton A, et al. Effect of denosumab versus zoledronic acid in preventing skeletal-related events in patients with bone metastases by baseline characteristics. Eur J Cancer. 2015; 53:75–83. [PubMed: 26693901]
- 60. Carducci MA, et al. Atrasentan in Patients With Advanced Renal Cell Carcinoma: A Phase 2 Trial of the ECOG-ACRIN Cancer Research Group (E6800). Clin Genitourin Cancer. 2015; 13(6):531–539.e1. [PubMed: 26272427]
- 61. Qiao L, et al. Endothelin-A receptor antagonists in prostate cancer treatment-a meta-analysis. Int J Clin Exp Med. 2015; 8(3):3465–73. [PubMed: 26064237]
- 62. Bougioukli S, et al. Combination therapy with BMP-2 and a systemic RANKL inhibitor enhances bone healing in a mouse critical-sized femoral defect. Bone. 2015
- 63. Lee SK, et al. Isoliquiritigenin Inhibits Metastatic Breast Cancer Cell-induced Receptor Activator of Nuclear Factor Kappa-B Ligand/Osteoprotegerin Ratio in Human Osteoblastic Cells. J Cancer Prev. 2015; 20(4):281–6. [PubMed: 26734591]
- 64. Rucci N, et al. Inhibition of protein kinase c-Src reduces the incidence of breast cancer metastases and increases survival in mice: implications for therapy. J Pharmacol Exp Ther. 2006; 318(1):161–72. [PubMed: 16627750]
- 65•. Brown DC, Agnello K, Iadarola MJ. Intrathecal resiniferatoxin in a dog model: efficacy in bone cancer pain. Pain. 2015; 156(6):1018–24. This paper proposes a promising novel therapy for CIBP. [PubMed: 25659068]
- 66. Cojoc M, et al. Emerging targets in cancer management: role of the CXCL12/CXCR4 axis. Onco Targets Ther. 2013; 6:1347–61. [PubMed: 24124379]
- 67. Lutz S. The role of radiation therapy in controlling painful bone metastases. Curr Pain Headache Rep. 2012; 16(4):300–6. [PubMed: 22576786]
- 68. Dennis K, et al. Single fraction conventional external beam radiation therapy for bone metastases: a systematic review of randomised controlled trials. Radiother Oncol. 2013; 106(1):5–14. [PubMed: 23321492]
- 69. McDonald R, et al. Quality of life after palliative radiotherapy in bone metastases: A literature review. J Bone Oncol. 2015; 4(1):24–31. [PubMed: 26579481]
- Wolanczyk MJ, Fakhrian K, Adamietz IA. Radiotherapy, Bisphosphonates and Surgical Stabilization of Complete or Impending Pathologic Fractures in Patients with Metastatic Bone Disease. J Cancer. 2016; 7(1):121–4. [PubMed: 26722368]
- 71. Chow E, et al. Dexamethasone in the prophylaxis of radiation-induced pain flare after palliative radiotherapy for bone metastases: a double-blind, randomised placebo-controlled, phase 3 trial. Lancet Oncol. 2015; 16(15):1463–72. [PubMed: 26489389]
- 72. Yu HH, Hoffe SE. Beyond the conventional role of external-beam radiation therapy for skeletal metastases: new technologies and stereotactic directions. Cancer Control. 2012; 19(2):129–36. [PubMed: 22487975]
- 73. Kothari G, et al. Outcomes of stereotactic radiotherapy for cranial and extracranial metastatic renal cell carcinoma: a systematic review. Acta Oncol. 2015; 54(2):148–57. [PubMed: 25140860]
- 74. Kim H, et al. Cost-effectiveness analysis of single fraction of stereotactic body radiation therapy compared with single fraction of external beam radiation therapy for palliation of vertebral bone metastases. Int J Radiat Oncol Biol Phys. 2015; 91(3):556–63. [PubMed: 25680599]
- 75. Wong E, et al. Re-irradiation for painful bone metastases a systematic review. Radiother Oncol. 2014; 110(1):61–70. [PubMed: 24094630]

76. Tomblyn M. The role of bone-seeking radionuclides in the palliative treatment of patients with painful osteoblastic skeletal metastases. Cancer Control. 2012; 19(2):137–44. [PubMed: 22487976]

- 77. Napoli A, et al. MR imaging-guided focused ultrasound for treatment of bone metastasis. Radiographics. 2013; 33(6):1555–68. [PubMed: 24108551]
- 78. Kobus T, McDannold N. Update on Clinical Magnetic Resonance-Guided Focused Ultrasound Applications. Magn Reson Imaging Clin N Am. 2015; 23(4):657–67. [PubMed: 26499282]
- 79. Quinn RH, et al. Contemporary management of metastatic bone disease: tips and tools of the trade for general practitioners. J Bone Joint Surg Am. 2013; 95(20):1887–95. [PubMed: 24288805]
- 80. Wood TJ, et al. Surgical management of bone metastases: quality of evidence and systematic review. Ann Surg Oncol. 2014; 21(13):4081–9. [PubMed: 25223925]
- 81. Hameed A, et al. Bone disease in multiple myeloma: pathophysiology and management. Cancer Growth Metastasis. 2014; 7:33–42. [PubMed: 25187738]
- 82. Rao PJ, et al. Minimally invasive percutaneous fixation techniques for metastatic spinal disease. Orthop Surg. 2014; 6(3):187–95. [PubMed: 25179352]
- 83. Smith HS. Painful boney metastases. Ann Palliat Med. 2012; 1(1):14–31. [PubMed: 25841427]
- 84. Foster RC, Stavas JM. Bone and soft tissue ablation. Semin Intervent Radiol. 2014; 31(2):167–79. [PubMed: 25053865]
- 85. Dodson S, et al. Muscle wasting in cancer cachexia: clinical implications, diagnosis, and emerging treatment strategies. Annu Rev Med. 2011; 62:265–79. [PubMed: 20731602]
- 86. Sjoblom B, et al. Skeletal muscle radiodensity is prognostic for survival in patients with advanced non-small cell lung cancer. Clin Nutr. 2016
- 87. Waning DL, Guise TA. Cancer-associated muscle weakness: What's bone got to do with it? Bonekey Rep. 2015; 4:691. [PubMed: 25992285]
- 88. Ryan AM, et al. Cancer-associated malnutrition, cachexia and sarcopenia: the skeleton in the hospital closet 40 years later. Proc Nutr Soc. 2016; 75(2):199–211. [PubMed: 26786393]
- 89. Marino FE, Risbridger G, Gold E. Activin-betaC modulates cachexia by repressing the ubiquitin-proteasome and autophagic degradation pathways. J Cachexia Sarcopenia Muscle. 2015; 6(4):365–80. [PubMed: 26673867]
- 90. Han HQ, et al. Myostatin/activin pathway antagonism: molecular basis and therapeutic potential. Int J Biochem Cell Biol. 2013; 45(10):2333–47. [PubMed: 23721881]
- Laurent MR, et al. Muscle-bone interactions: From experimental models to the clinic? A critical update. Mol Cell Endocrinol. 2015
- 92. Tisdale MJ. Mechanisms of cancer cachexia. Physiol Rev. 2009; 89(2):381–410. [PubMed: 19342610]
- 93. Sala D, Sacco A. Signal transducer and activator of transcription 3 signaling as a potential target to treat muscle wasting diseases. Curr Opin Clin Nutr Metab Care. 2016; 19(3):171–6. [PubMed: 27023048]
- Zimmers TA, Fishel ML, Bonetto A. STAT3 in the systemic inflammation of cancer cachexia.
 Semin Cell Dev Biol. 2016
- 95. Rom O, Reznick AZ. The role of E3 ubiquitin-ligases MuRF-1 and MAFbx in loss of skeletal muscle mass. Free Radic Biol Med. 2015
- 96. Baracos VE. Cancer-associated cachexia and underlying biological mechanisms. Annu Rev Nutr. 2006; 26:435–61. [PubMed: 16602932]
- 97. Argiles JM, et al. Molecular mechanisms involved in muscle wasting in cancer and ageing: cachexia versus sarcopenia. Int J Biochem Cell Biol. 2005; 37(5):1084–104. [PubMed: 15743680]
- 98. Toth MJ, et al. Molecular mechanisms underlying skeletal muscle weakness in human cancer: reduced myosin-actin cross-bridge formation and kinetics. J Appl Physiol (1985). 2013; 114(7): 858–68. [PubMed: 23412895]
- 99. Gilliam LA, St Clair DK. Chemotherapy-induced weakness and fatigue in skeletal muscle: the role of oxidative stress. Antioxid Redox Signal. 2011; 15(9):2543–63. [PubMed: 21457105]
- 100. Kumar NB, et al. Cancer cachexia: traditional therapies and novel molecular mechanism-based approaches to treatment. Curr Treat Options Oncol. 2010; 11(3–4):107–17. [PubMed: 21128029]

101. Argiles JM, et al. Cachexia and sarcopenia: mechanisms and potential targets for intervention. Curr Opin Pharmacol. 2015; 22:100–6. [PubMed: 25974750]

- 102. Wen HS, et al. Clinical studies on the treatment of cancer cachexia with megestrol acetate plus thalidomide. Chemotherapy. 2012; 58(6):461–7. [PubMed: 23406994]
- 103. Dobs AS, et al. Effects of enobosarm on muscle wasting and physical function in patients with cancer: a double-blind, randomised controlled phase 2 trial. Lancet Oncol. 2013; 14(4):335–45. [PubMed: 23499390]
- 104•. Temel JS, et al. Anamorelin in patients with non-small-cell lung cancer and cachexia (ROMANA 1 and ROMANA 2): results from two randomised, double-blind, phase 3 trials. Lancet Oncol. 2016 This paper gives the results of two phase 3 trials, in which anamorelin had success in restoring lean muscle mass in patients with cachexia.
- 105. Busquets S, et al. Myostatin blockage using actRIIB antagonism in mice bearing the Lewis lung carcinoma results in the improvement of muscle wasting and physical performance. J Cachexia Sarcopenia Muscle. 2012; 3(1):37–43. [PubMed: 22450815]
- 106. Ebner N, et al. Highlights from the 7th Cachexia Conference: muscle wasting pathophysiological detection and novel treatment strategies. J Cachexia Sarcopenia Muscle. 2014; 5(1):27–34. [PubMed: 24595460]
- 107. Toledo M, et al. Complete reversal of muscle wasting in experimental cancer cachexia: Additive effects of activin type II receptor inhibition and beta-2 agonist. Int J Cancer. 2016; 138(8):2021–9. [PubMed: 26595367]
- 108. Bellinger AM, et al. Remodeling of ryanodine receptor complex causes "leaky" channels: a molecular mechanism for decreased exercise capacity. Proc Natl Acad Sci U S A. 2008; 105(6): 2198–202. [PubMed: 18268335]
- 109. Smith RC, et al. Myostatin Neutralization Results in Preservation of Muscle Mass and Strength in Preclinical Models of Tumor-Induced Muscle Wasting. Mol Cancer Ther. 2015; 14(7):1661–70. [PubMed: 25908685]
- 110. DiGirolamo DJ, Kiel DP, Esser KA. Bone and skeletal muscle: neighbors with close ties. J Bone Miner Res. 2013; 28(7):1509–18. [PubMed: 23630111]
- 111. Waning DL, Guise TA. Molecular mechanisms of bone metastasis and associated muscle weakness. Clin Cancer Res. 2014; 20(12):3071–7. [PubMed: 24677373]
- 112. Bonetto A, et al. JAK/STAT3 pathway inhibition blocks skeletal muscle wasting downstream of IL-6 and in experimental cancer cachexia. Am J Physiol Endocrinol Metab. 2012; 303(3):E410–21. [PubMed: 22669242]
- 113. Lee YS, Lee SJ. Regulation of GDF-11 and myostatin activity by GASP-1 and GASP-2. Proc Natl Acad Sci U S A. 2013; 110(39):E3713–22. [PubMed: 24019467]
- 114. Ohuchi H, Noji S. Fibroblast-growth-factor-induced additional limbs in the study of initiation of limb formation, limb identity, myogenesis, and innervation. Cell Tissue Res. 1999; 296(1):45–56. [PubMed: 10199964]
- 115. Yakar S, et al. Circulating levels of IGF-1 directly regulate bone growth and density. J Clin Invest. 2002; 110(6):771–81. [PubMed: 12235108]
- 116. Hamrick MW, et al. Increased muscle mass with myostatin deficiency improves gains in bone strength with exercise. J Bone Miner Res. 2006; 21(3):477–83. [PubMed: 16491296]
- 117. Elkasrawy MN, Hamrick MW. Myostatin (GDF-8) as a key factor linking muscle mass and bone structure. J Musculoskelet Neuronal Interact. 2010; 10(1):56–63. [PubMed: 20190380]
- 118. Bren-Mattison Y, Hausburg M, Olwin BB. Growth of limb muscle is dependent on skeletal-derived Indian hedgehog. Dev Biol. 2011; 356(2):486–95. [PubMed: 21683695]
- 119. Waning DL, et al. Excess TGF-beta mediates muscle weakness associated with bone metastases in mice. Nat Med. 2015; 21(11):1262–71. [PubMed: 26457758]
- 120. Lee SJ, et al. Regulation of muscle growth by multiple ligands signaling through activin type II receptors. Proc Natl Acad Sci U S A. 2005; 102(50):18117–22. [PubMed: 16330774]
- 121. Chen JL, et al. Elevated expression of activins promotes muscle wasting and cachexia. Faseb j. 2014; 28(4):1711–23. [PubMed: 24378873]
- 122. Zhou X, et al. Reversal of cancer cachexia and muscle wasting by ActRIIB antagonism leads to prolonged survival. Cell. 2010; 142(4):531–43. [PubMed: 20723755]

123. Bowser M, et al. Effects of the activin A-myostatin-follistatin system on aging bone and muscle progenitor cells. Exp Gerontol. 2013; 48(2):290–7. [PubMed: 23178301]

- 124. Korpal M, et al. Imaging transforming growth factor-beta signaling dynamics and therapeutic response in breast cancer bone metastasis. Nat Med. 2009; 15(8):960–6. [PubMed: 19597504]
- 125. Hubackova S, et al. IL1- and TGFbeta-Nox4 signaling, oxidative stress and DNA damage response are shared features of replicative, oncogene-induced, and drug-induced paracrine 'bystander senescence'. Aging (Albany NY). 2012; 4(12):932–51. [PubMed: 23385065]
- 126. Schiaffino S, et al. Mechanisms regulating skeletal muscle growth and atrophy. FEBS J. 2013; 280(17):4294–314. [PubMed: 23517348]
- 127. Kagiya T. MicroRNAs and Osteolytic Bone Metastasis: The Roles of MicroRNAs in Tumor-Induced Osteoclast Differentiation. J Clin Med. 2015; 4(9):1741–52. [PubMed: 26343739]
- 128. Burne TH, et al. Swimming behaviour and post-swimming activity in Vitamin D receptor knockout mice. Brain Res Bull. 2006; 69(1):74–8. [PubMed: 16464687]
- 129. Mongre RK, et al. A New Paradigm to Mitigate Osteosarcoma by Regulation of MicroRNAs and Suppression of the NF-kappaB Signaling Cascade. Dev Reprod. 2014; 18(4):197–212. [PubMed: 25949190]
- 130. Ell B, et al. Tumor-induced osteoclast miRNA changes as regulators and biomarkers of osteolytic bone metastasis. Cancer Cell. 2013; 24(4):542–56. [PubMed: 24135284]
- 131. Pollari S, et al. Identification of microRNAs inhibiting TGF-beta-induced IL-11 production in bone metastatic breast cancer cells. PLoS One. 2012; 7(5):e37361. [PubMed: 22629385]