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1 **Whole-body photoreceptor networks are independent of ‘lenses’ in brittle stars**

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## 10 **Abstract**

11 Photoreception and vision are fundamental aspects of animal sensory biology and ecology,  
12 but important gaps remain in our understanding of these processes in many species. The  
13 colour-changing brittle star *Ophiocoma wendtii* is iconic in vision research, speculatively  
14 possessing a unique whole-body visual system that incorporates information from nerve  
15 bundles underlying thousands of crystalline ‘microlenses’. The hypothesis that these form a  
16 sophisticated compound eye-like system regulated by chromatophore movement has been  
17 extensively reiterated, with consequent investigations into biomimetic optics and similar  
18 ‘visual’ structures in living and fossil taxa. However, no photoreceptors or visual behaviours  
19 have ever been identified. We present the first evidence of photoreceptor networks in three  
20 *Ophiocoma* species, both with and without microlenses and colour-changing behaviour.  
21 High-resolution microscopy, immunohistochemistry and synchrotron tomography  
22 demonstrate that putative photoreceptors cover the animals’ oral, lateral, and aboral surfaces,  
23 but are absent at the hypothesised focal points of the microlenses. The structural optics of  
24 these crystal ‘lenses’ are an exaptation and do not fulfil any apparent visual role. This  
25 contradicts previous studies, yet the photoreceptor network in *Ophiocoma* appears even more  
26 widespread than previously anticipated, both taxonomically and anatomically.

27 **Keywords:** Extra-ocular photoreception, vision, ophiuroids, photoreceptors, sensory biology.

## 28 **Background**

29 The ability to sense light without eyes, extraocular photoreception (EOP), is being discovered  
30 across an increasingly diverse range of animal groups at an accelerating rate [1–4]. EOP  
31 generally confers behaviours such as circadian rhythms, phototaxis, reflexes, and colour  
32 change, but not spatial resolution [1,3,5]. Controversially, it has been proposed that some  
33 echinoderms may be able to consolidate extraocular information to facilitate image-forming

34 vision [6–9], placing them in a position of exceptional research interest [5]. Understanding  
35 the functional model and limits of integration in dispersed photoreceptor systems that may  
36 provide spatial resolution will have profound implications for neurobiology, visual evolution,  
37 and biomimetic design [1,10], but despite considerable research effort these remain elusive  
38 [5].

39 The brittle star *Ophiocoma wendtii* first attracted attention for its charismatic colour-changing  
40 behaviour and extreme sensitivity to illumination [11]. Animals undergo a striking  
41 transformation from black-brown during the day to beige-grey with dark bands at night,  
42 which can be artificially induced by changing their light environment, and strongly prefer  
43 shade to light exposure, including moonlight [11]. Morphological studies reported nerve  
44 bundles beneath expanded, highly regular calcite hemispheres on the dorsal arm plates  
45 (enlarged peripheral trabeculae, EPTs) [11,12]. The EPTs were speculatively interpreted as  
46 potential ‘microlenses’, proposed to focus light onto putative photoreceptors within or  
47 associated with the nerve bundles, with the passage of incoming light regulated by the  
48 activity of surrounding “pupillary” chromatophores [5,9,11–13]. This proposal remains  
49 unexplored and no photoreceptors have been identified to date; however, many subsequent  
50 studies interpreted new data in the context of this hypothesis being accepted. The  
51 architecture, distribution and optical properties of the arm plates in *Ophiocoma* are  
52 fundamental to the hypothesis that they focus light onto underlying photoreceptor elements  
53 [9,11,12], which has also contributed to interpretations of skeletal involvement in echinoid  
54 photoreception, yet the EPTs have only been presented in the literature from removed and  
55 chemically treated plates [9,12,14,15].

56 The repeated-unit nature and apparent optical sophistication of this system even led to the  
57 speculative suggestion of a compound eye-like function across the dorsal surface of the

58 animal, as has also been proposed in echinoids [7,16,17], facilitating its apparent ability to  
59 detect shadows and navigate towards dark shelters from a distance [7,9]. The hypothesis that  
60 the EPTs, chromatophores, and underlying nerves could form an advanced visual system has  
61 been extensively reiterated by other authors [1,5,7,15,18–26], with resultant investigations  
62 into biomimetic optics [9,10,19,20], and vision in both living [22,25] and fossil taxa [14,27].  
63 However, there is no morphological or behavioural evidence to support this idea, and no  
64 candidates for the necessary neural integration centres that might be required by such a  
65 system (though the precise nature of such centres remain unclear) [28].

66 Since the last morphological investigations of *O. wendtii*, numerous opsins – key components  
67 of most photosensitive pigments – were identified in the genome of the sea urchin  
68 *Strongylocentrotus purpuratus* [29]. This facilitated the discovery of the first opsin-  
69 expressing cells in urchins, brittle stars and sea stars, using antibodies subsequently raised  
70 against Sp-Op targets [16,26,30], as well as many more opsin sequences in other echinoderms  
71 [25,26,31]. Brittle stars, like other echinoderms, possess both rhabdomeric (r-) and ciliary (c-)  
72 visual opsins as well as multiple non-visual classes [25,26,32], but exhibit multiple  
73 duplications of the rhabdomeric class (closest to Sp-Op4) [26]. These are considered non-  
74 visual in most deuterostomes, but are strongly implicated in visual behaviour in both urchins  
75 and sea stars [16,33], and sequencing of arm transcriptomes in two brittle stars demonstrated  
76 detectable levels of expression of r-opsins similar to Sp-Op4, but not c-opsins, though these  
77 were detected at low levels by immunolabelling against Sp-Op1 [25].

78 We established multiple lines of evidence to investigate the presence and location of  
79 photoreceptors, determine their arrangement in relation to putative microlenses *in situ*, and  
80 compare *Ophiocoma wendtii* with two ecologically co-occurring congeners, one lacking  
81 EPTs and colour change behaviour [11]. Immunohistochemistry, scanning electron

82 microscopy (SEM), synchrotron tomography, and histology were supplemented with  
83 exploratory behavioural experiments (supplementary material) in order to finally locate  
84 putative photoreceptors and compare their distribution and structure across *Ophiocoma*.

## 85 **Materials and methods**

### 86 *Specimens*

87 Specimens of *Ophiocoma wendtii*, *O. echinata*, and *O. pumila* were collected from shallow  
88 reef rubble at Punta Hospital, Isla Solarte, Bocas del Toro, Panama (9°19'44.4"N,  
89 82°12'21.6"W, 0–3 m), and housed in outdoor flow-through unfiltered seawater aquaria under  
90 a natural 12:12 hr light:dark cycle at the Smithsonian Tropical Research Institute, Bocas del  
91 Toro, Panama. Animals were photographed, measured, and identified by disc diameter and  
92 longest arm length, and allowed three days recovery between collection and experiments.  
93 Animals that autotomised arms during or following collection were excluded from trials.  
94 Specimens were collected under ARAP permit 2014-52b and exported under ARAP export  
95 permit 2015-2.

### 96 *Synchrotron tomography*

97 Arm segments were fixed in 4% glutaraldehyde in a sodium cacodylate buffer (0.1M, pH 7.4)  
98 in their daylight state and stored in sodium cacodylate buffer. Segments were rinsed in buffer  
99 and serially dehydrated in acetone before drying with hexamethyldisilazane (HMDS) and  
100 mounting on stubs.

101 Three samples from *Ophiocoma wendtii* (three arm segments), *Ophiocoma pumila* (two arm  
102 segments and a pair of arm spines), and *Ophiocoma echinata* (two arm segments and one arm  
103 spine) were studied with non-destructive synchrotron tomography. Synchrotron radiation X-  
104 ray tomographic microscopy was performed at the TOMCAT beamline (Swiss Light Source,  
105 Paul Scherrer Institut, Villigen, Switzerland). Samples were scanned using an X-ray energy

106 of 20 keV, 1501 projections, and an exposure time of 250 ms. This gave tomographic datasets  
107 with a voxel size of 1.75  $\mu\text{m}$  (x, y and z), which were digitally reconstructed as three-  
108 dimensional virtual models (electronic supplementary material) using SPIERS [34] and  
109 AMIRA (FEI Visualization Science Group).

#### 110 *Histology and scanning electron microscopy*

111 Whole specimens and excised arm segments from *Ophiocoma wendtii*, *O. echinata*, and *O.*  
112 *pumila* were fixed in glutaraldehyde as above and stored in sodium cacodylate buffer (pH  
113 7.4). For histology, arm segments were post-fixed in 1% osmium tetroxide, decalcified in 2%  
114 ascorbic acid in 0.15 M sodium chloride solution for 72 hours [16] and dehydrated in an  
115 acetone series before embedding in Epon epoxy resin (Agar Scientific). Blocks were  
116 sectioned at 1  $\mu\text{m}$  on a Leica RM2255 automated microtome with a diamond knife  
117 (HistoJumbo, 8 mm, DiATOME, Switzerland) and stained with Richardson's solution.  
118 Sections were photographed using an Olympus E-600 digital camera mounted on an Olympus  
119 BX41 microscope.

120 For SEM, glutaraldehyde-fixed arm segments from *Ophiocoma wendtii* were washed in dilute  
121 cacodylate buffer, serially dehydrated in acetone, chemically dried overnight with HMDS,  
122 mounted on stubs and visualised on an FEI Quanta FEG scanning electron microscope at 15  
123 kV.

#### 124 *Immunohistochemistry*

125 Light-adapted arm segments from *Ophiocoma wendtii*, *O. echinata*, and *O. pumila* were  
126 tested for reactivity to sea urchin ciliary (Sp-Op1) and rhabdomeric (Sp-Op4) opsins [31].  
127 Segments were fixed in 4% paraformaldehyde in phosphate-buffered saline (PBS, pH 7.4) for  
128 30 minutes at room temperature before washing in PBS and decalcifying in 2% ascorbic acid  
129 in 0.15 M sodium chloride solution for 72 hours [adapted from 15]. Samples were rinsed in

130 PBS and stored in 0.05% sodium azide in PBS. Tissue used for sectioning was rinsed in PBS  
131 for 20 minutes before embedding in 4% agarose gel. Thick sections (150  $\mu$ m) were taken  
132 using a Leica VT 1200S vibratome. Arm segments and sections were washed in PBS and  
133 0.1% Triton X (PBS-T) and blocked in PBST and 0.5% normal goat serum (NGS) for one  
134 hour before incubation with anti-acetylated tubulin (1:200) and either anti-Sp-Opsin4 or anti-  
135 Sp-Opsin1 (*Strongylocentrotus purpuratus*, 1:50) [16] overnight, all at room temperature.  
136 These antibodies bind to and exhibit high sequence similarity to discovered homologs in  
137 brittle stars [25,26]. Specimens were then washed in PBST and incubated with either Alexa  
138 Fluor 633 goat anti-mouse (1:500) or Alexa Fluor 488 goat anti-rabbit (1:500) for at least  
139 three hours at room temperature, rinsed with PBST and visualised on a Leica TCS SPE  
140 confocal laser scanning microscope. Images and image stacks were captured using Leica  
141 Application Suite Advanced Fluorescence v.2.6.3 and prepared in Fiji [35].

## 142 **Results**

### 143 *Arm plate structure*

144 High-resolution synchrotron tomography and SEM visualised expanded peripheral trabeculae  
145 (EPTs, putative microlenses) *in situ* without disrupting soft tissue. Regular, near-  
146 hemispherical EPTs, 30–40  $\mu$ m in diameter, cover the dorsal (aboral) arm plates, but also the  
147 ventral (oral) arm plates and the dorsal and ventral margins of the lateral plates in *Ophiocoma*  
148 *wendtii* (Figure 1A,B,C), contrary to previous reports that they are restricted to the dorsal  
149 plates and dorsal margins of the lateral plates [9,12]. In cross-section, EPTs often appear to  
150 be at the distal face of an uninterrupted calcite core projecting through the plate (Figures 1D,  
151 S1, S4), leaving little or no room beneath the centre of the EPT for soft tissue. *In vivo*, the  
152 plates are covered by a fine dermal cuticle that is highly sensitive to chemical treatment [13]



153 (Figure 1A'). EPTs are interspersed by the projection of short ciliary tufts through the cuticle  
154 (Figure 1A') that may represent receptors described as *Stäbchen* [36,37].

155 *Ophiocoma echinata* and *O. pumila* are sympatric with *O. wendtii* and were included in the  
156 original study of colour change and light sensitivity in the latter [11]. Whereas *O. echinata*  
157 exhibits a similar day-to-night colour change to *O. wendtii*, *O. pumila* does not [11], and the  
158 EPTs found in both *O. wendtii* and *O. echinata* were apparently lacking in *O. pumila* [9,12].  
159 However, synchrotron scans of *Ophiocoma echinata* and *O. pumila* showed similarities  
160 between all three species. *Ophiocoma echinata* have slightly smaller (diameter 20–30  $\mu\text{m}$ )  
161 EPTs than *O. wendtii*, again present on the dorsal, ventral, and lateral arm plates and highly  
162 regular in shape (Figures 2A–E and S2). The dorsal, ventral, and dorso-ventral margins of the  
163 lateral arm plates in *O. pumila* also bear EPT-like structures, in contrast to previous findings  
164 from chemically treated plates [9,12] (Figure 2F–J). These structures are smaller (diameter  
165 20–25  $\mu\text{m}$ ), particularly on the ventral arm plates (diameter 15–20  $\mu\text{m}$ ), and more irregular  
166 yet anatomically similar to the EPTs observed in the other two species (Figures 1, 2, and S1–  
167 S3).

#### 168 *Nerves and opsin reactivity*

169 Immunohistochemistry allowed us to specifically target nerve fibres and cells reactive to sea  
170 urchin opsins, where photoreceptors have proved elusive using classical methods [12]. In all  
171 three *Ophiocoma* spp., a branching nerve net covers the proximal faces of the arm plates,  
172 extending laterally from the midline and emitting branching nerve bundles distally into the  
173 plate (Figures 3A,B, 4A, S5A). These originate in the radial nerve cord at the oral side and a  
174 smaller medial nerve at the aboral side (Figures 3B, 4A, S4, S5A). Crucially, the bundles  
175 innervating the arm plates do not terminate at the proposed focal point of the EPTs according  
176 to [9], instead projecting between them towards the outer surface of the arm (Figures 3B,D,

177 4C). Ovoid cells (soma approx. 10  $\mu\text{m}$ ) associated with these nerves surround the EPTs and  
178 react to r-opsin antibody Sp-Op4 (Figures 3A, 4D, S5B,C; see Figure S6 for controls). Cell  
179 bodies are located just above the midline of the EPTs, project towards the surface of the arm  
180 and bear rounded terminal expansions that react strongly to the r-opsin antibody (Figure 4D).  
181 These cells are notably absent at the putative focal point of the EPTs, where photoreceptors  
182 had been predicted [9,11,12]. They appear to lack specialised membrane structures and are  
183 reminiscent of the general receptors described in *Ophioderma longicauda* [38], though a  
184 short cilium is not always visible (e.g. Figure 4D); however, they do not resemble those  
185 reported in *Ophiura ophiura* [39], which are more akin to the *Stäbchen*. The opsin-reactive  
186 cells are regularly arranged over the aboral, lateral, and oral sides of the arm, as well as some  
187 at the surface of the spines, in *O. wendtii*, *O. echinata*, and *O. pumila*. They sometimes  
188 appear associated with ciliated cells potentially corresponding to those in *Ophionereis*  
189 *schayeri* [40]. Single and multiciliary tufts protrude between the EPTs (Figure 3).

190 There are also scattered Sp-Op1-reactive cells of similar size (Figure 4A), but these were less  
191 consistently observed and so are not further discussed here other than to highlight their  
192 presence. We also observed some reactivity to both opsins within the medial and lateral  
193 nerves and the radial nerve cord (Figures 3B, 4A and S5C), of which the latter has been  
194 reported to exhibit intrinsic photosensitivity and opsin expression [2,26,31].

195 Potential nerve connections between Sp-Op4-reactive cells, both laterally at the surface and  
196 in convergent innervating bundles (Figures 3A,B and S5C), could indicate integration or  
197 summation between them. However, we found no unusual or concentrated area of neuropil as  
198 might be expected for integrating visual information across such an expansive network.

## 199 Discussion

200 The putative photoreceptor system in *Ophiocoma wendtii*, *O. echinata* and *O. pumila* is  
201 extensive; our findings revealed a much larger network than previously posited, which is  
202 present across almost the complete body surface in all three species. The morphology,  
203 reactivity and arrangement of Sp-Op4-reactive cells support their candidacy as  
204 photoreceptors; past work indicates that r-opsins homologous to Sp-Op4 are involved in  
205 brittle star photoreception, and that they are likely expressed at higher levels than c-opsin  
206 homologs to Sp-Op1 [25,26], in line with our findings. Critically, the nerve bundles proposed  
207 to act as photoreceptors project past the EPTs towards the opsin-reactive cells. Contrary to  
208 expectations, these putative photoreceptors appear to be entirely independent of the EPTs;  
209 their anatomical configuration relative to the EPTs demonstrates no support for an optical  
210 role as ‘microlenses’ (Figures 3A,B and 4D).

211 The three *Ophiocoma* species possess vast networks of putative dermal photoreceptors  
212 covering their dorsal, ventral, and lateral arm plates. This is a considerable expansion on the  
213 system hypothesised to exist beneath the EPTs [9,12], both anatomically and taxonomically,  
214 and may represent one of the largest dispersed photoreceptor systems described to date,  
215 thanks to the ability to monitor expression of molecular markers. These findings complement  
216 proposed dermal photoreceptor networks in other echinoderms, most notably sea urchins  
217 [7,41], but turn the tables on previous theories about *Ophiocoma wendtii* [9,11]. We  
218 anticipate that future researchers will find similarly large extraocular systems in other taxa.

219 The optical involvement of the EPTs in a photoreceptor system is problematic for several  
220 reasons. The EPTs are present on the oral (ventral) and lateral surfaces (Figures 1 and 2) as  
221 well as the dorsal arm plates. The lateral plates would be a complex surface for integrated  
222 photoreception, let alone vision, and the oral surfaces would be largely redundant; although  
223 some brittle stars expose the ventral arm during feeding, *Ophiocoma* does not [42]. Second,

224 the sheer number of EPTs is enormous; we found an average of 510 EPTs per dorsal arm  
225 plate in *Ophiocoma wendtii*, with around 75 plates per arm (mean length 112 mm). Rough  
226 calculations indicate that an average-sized individual would possess over 300,000 EPTs.  
227 However, they apparently lack any further organisation of the photoreceptors into discrete  
228 units, as seen in other distributed visual systems [18,43,44], or a processing centre beyond the  
229 radial nerve cords, providing no indication of potential integration mechanisms for such an  
230 enormous network. Additionally, the acceptance angle of each receptor between the EPTs  
231 would be too large to enable high resolution. Indeed, *Ophiocoma wendtii* exhibits limited  
232 visual behaviour according to preliminary tests herein (Figure S7). As a third, independent,  
233 argument against an optical role for the EPTs, the cuticle, chromatophores, and other  
234 biological material also occlude their rounded shape and surface *in vivo* and may interfere  
235 with the passage of light (Figure 1). Expanded chromatophores cover the EPTs completely,  
236 with no aperture to indicate pupillary function [5,11,12] (Figure 3). Conversely, contracted  
237 chromatophores appear to lie beneath as well as between the EPTs [see 12], further shielding  
238 peripheral nerve elements from incoming light in dark-adapted animals.

239 Finally, and most importantly, the presence of photoreceptive elements is primarily detected  
240 in between and not beneath the EPTs. No opsin-reactive cells were observed at the reported  
241 focal point beneath the EPTs, and the nerve bundles that were implicated as primary  
242 photoreceptors [12] not only lack reactivity to the tested opsins, but project past the EPTs  
243 towards the plate surface. Visual photoreceptors in other taxa are not universally located at  
244 the optical focal point [43,45], but these opsin-reactive cells are within the dermal layer and  
245 apparently far from any potential optical effect of the EPTs; their projection and expansion  
246 above the EPTs also negate channelling or light-gathering roles. An identical pattern of anti-  
247 Sp-Op4 reactivity is present in *O. pumila*, which lacks highly regular EPTs and colour change

248 (Figure 3). The optical properties of the EPTs may be an exaptation relevant to materials  
249 science [9,10], but they do not appear to perform any optical role in *Ophiocoma*.

250 Although our findings contest the interpretation of the EPTs as microlenses in *Ophiocoma*,  
251 they are still compatible with the electrophysiological studies of Cobb and Hendler [13].  
252 They demonstrated increasing photosensitivity correlating with increasing loss of arm tissue,  
253 bleaching EPTs and dermal tissue, including chromatophores, until the nerve bundles beneath  
254 each EPT were affected. They argued that this demonstrated these nerve bundles are the  
255 primary photoreceptors. However, their findings that the receptors were located beneath the  
256 epidermis, regulated in their sensitivity by chromatophores, and became more sensitive with  
257 the removal of overlying tissue, are also compatible with the data presented here. The authors  
258 acknowledge that other unrecognised cell types could be responsible; given the resemblance  
259 of the r-opsin-reactive cells to generalised dermal receptors, it appears that they were indeed  
260 overlooked.

261 Of course, we too cannot eliminate the possibility that additional cells at the base of the EPTs  
262 were not detected in this (or any other) study, and echinoderms [46] including brittle stars  
263 [26] demonstrate high opsin diversity. Identifying a complete suite of opsin candidates in  
264 *Ophiocoma* will help detect other opsin-expressing (or cryptochrome-expressing [47]) tissues  
265 underlying the EPTs, if present, although transcriptomic studies in other brittle stars support a  
266 key role for Sp-Op4 homologs [25,26]. In addition, functions besides photoreception have  
267 now been described for several r-opsins in some arthropods and vertebrates [48]. However,  
268 the Sp-Op4-reactive cells we interpret as photoreceptor candidates conform to previous  
269 descriptions of receptor morphology and r-opsin expression in other ophiuroids, are  
270 positioned within the EPT-chromatophore layer in line with Hendler and Cobb [13], are  
271 highly numerous, and represent the only candidates identified in any study in over 30 years.

272 We propose it is highly likely that they are responsible for photosensitivity and corresponding  
273 behaviours in *Ophiocoma*.

274 Concerning visual ability, and especially the compound eye model suggested by several  
275 authors, we cannot support it based on our findings. *Ophiocoma wendtii* certainly exhibits  
276 high sensitivity to light [11] and strong shade-seeking responses (Supplementary material,  
277 Figure S7). Our preliminary behavioural experiments showed that *Ophiocoma wendtii* could  
278 be capable of basic image formation, as indicated by its ability to detect large, high-contrast  
279 targets (Figure S7). However, response to targets of  $35\text{--}57^\circ$  is coarse even in comparison to  
280 other echinoderms, including urchins using a dermal photoreceptor system where skeletal  
281 structures have also been implicated in spatial resolution [7,41]. The detection and location of  
282 large, dark, high-contrast targets from short distances also do not necessarily equate to spatial  
283 resolution rather than phototaxis (owing to lower overall light intensity in the region of the  
284 target), so we hesitate to unequivocally support visual capability. It is not yet clear precisely  
285 how the abilities of *O. echinata* and *O. pumila* compare to *O. wendtii* beyond their lesser  
286 sensitivity [11]; in light of their relatively distant phylogenetic positions in the genus [49],  
287 further comparisons will be of great interest in the context of wider photosensitivity in the  
288 taxon. A compound eye requires that each repeated optical unit represents, or scales to, a unit  
289 of resolution, a pixel. We find no evidence that the EPTs act as lenses in ommatidium-like  
290 optical units, so the photoreceptors could theoretically represent these themselves. If it acts as  
291 a compound eye *sensu stricto*, the vast photoreceptor network in *Ophiocoma* should confer  
292 fine resolution [50], but this is not supported by behavioural data (Figure S6).

293 Local signal integration and spatial summation could explain high sensitivity and low spatial  
294 resolution (if any; Figure S7) in *O. wendtii* [51]. However, the innervation networks do not  
295 show any organisational structure that would presumably be a prerequisite for complex signal  
296 integration in a compound-type eye, and synapses are known to be relatively rare in

297 ophiuroid nervous systems [28]. Photoresponsive behaviours may instead function through  
298 reflex activity within arms or arm segments. Thus, even basic directional light/dark  
299 perception could guide non-visual phototactic shelter-seeking behaviour in complex  
300 environments with high light intensity and low turbidity [52].

## 301 **Conclusions**

302 The correlation between increasing responsiveness, EPT distribution, and colour change  
303 formerly contributed a key piece of indirect evidence that EPTs are integral to photoreception  
304 [9,12]. The joint absence of EPTs and colour change in *Ophiocoma pumila* was interpreted as  
305 evidence for the involvement of the EPTs in light sensing [9,11], but it may still indicate their  
306 function. Colour change in *Ophiocoma* depends on the expansion and retraction of  
307 chromatophores over and around the EPTs [11]. Chromatophore activity is likely to be  
308 autonomous and does not appear to be associated with nervous or muscular accessories [12].  
309 We therefore propose that the large, regular EPTs found on the arm plates in *O. wendtii* and  
310 *O. echinata* could be a structural adaptation relating to chromatophore activity. By  
311 maximising separation of chromatophores in their contracted state, the distinction between  
312 contracted and expanded states is amplified, producing a more dramatic colour change. The  
313 chromatophore activity likely affects photoreceptor sensitivity by altering the amount of  
314 screening pigment surrounding them, in line with increased sensitivity in dark-adapted arms  
315 [13], but not by controlling the amount of light reaching the EPTs. Thus, the EPTs may have  
316 an accessory role in photoreception, through their potential role in colour change, but there is  
317 no optical focussing. This is dramatically at odds with the published literature and the popular  
318 status of *O. wendtii* as an advanced visual species [5,9].

319 Our findings also caution against interpretations of complex photoreceptor systems from  
320 skeletal evidence alone in living and fossil echinoderms [14,22,27]. For example, some

321 asteroids with visual optic cushions also have EPTs [22,33,53]; these skeletal structures that  
322 have optical properties (in the physical sense) are likely irrelevant to the organism's sensory  
323 biology. We propose that the placement, concentration, and connectivity of dermal  
324 photoreceptors confer high photosensitivity across the body, resulting in sensitive directional  
325 extraocular photoreception and not vision *per se* in *Ophiocoma wendtii*. This more accurate  
326 model, without requiring focussing lenses, marks a significant advance in understanding the  
327 capabilities of extraocular photoreception.

### 328 **Competing interests**

329 The authors have no competing interests.

### 330 **Author contributions**

331 LSR designed the study, collected animals, performed histology, SEM, and behavioural  
332 experiments, and analysed the data, assisted and supervised by JDS. LSR and EUL performed  
333 immunohistochemistry and interpreted results. IAR scanned specimens at the synchrotron,  
334 and LSR and IAR processed scan data. LSR and JDS wrote the manuscript, and all authors  
335 contributed editorial input and gave their approval for submission.

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350 **Supplementary material** is available online as files S1–S3, Figures S4–S8, and Table S9.

351 **Data availability**

352 All data are available on Dryad.

353 **References**

- 354 1. Ramirez MD, Speiser DI, Pankey SM, Oakley TH. 2011 Understanding the dermal  
355 light sense in the context of integrative photoreceptor cell biology. *Vis. Neurosci.* **28**,  
356 265–279.
- 357 2. Millott N, Yoshida M. 1959 The shadow reaction of *Diadema antillarum* Philippi I.  
358 The spine response and its relation to the stimulus. *J. Exp. Biol.* **37**, 363–375.
- 359 3. Yoshida M. 1979 Extraocular photoreception. In *Handbook of Sensory Physiology*.  
360 *Volume VII/6A: Vision in Invertebrates A: Invertebrate Photoreceptors* (ed H  
361 Autrum), pp. 581–640. Berlin, Heidelberg, New York: Springer-Verlag.
- 362 4. Bielecki J, Zaharoff AK, Leung NY, Garm A, Oakley TH. 2014 Ocular and  
363 extraocular expression of opsins in the rhopalium of *Tripedalia cystophora* (Cnidaria:  
364 Cubozoa). *PLoS One* **9**, e98870.
- 365 5. Hendler G. 2005 An echinoderm’s eye view of photoreception and vision. In  
366 *Echinoderms: München: Proceedings of the 11th International Echinoderm*

- 367 *Conference* (eds T Heinzeller, J Nebelsick), pp. 339–349. München: Taylor & Francis.
- 368 6. Blevins E, Johnsen S. 2004 Spatial vision in the echinoid genus *Echinometra*. *J. Exp.*  
369 *Biol.* **207**, 4249–53.
- 370 7. Yerramilli D, Johnsen S. 2010 Spatial vision in the purple sea urchin  
371 *Strongylocentrotus purpuratus* (Echinoidea). *J. Exp. Biol.* **213**, 249–55.
- 372 8. Jackson E, Johnsen S. 2011 Orientation to objects in the sea urchin *Strongylocentrotus*  
373 *purpuratus* depends on apparent and not actual object size. *Biol. Bull.* **220**, 86–88.
- 374 9. Aizenberg J, Tkachenko A, Weiner S, Addadi L, Hendler G. 2001 Calcitic microlenses  
375 as part of the photoreceptor system in brittlestars. *Nature* **412**, 819–22.
- 376 10. Aizenberg J, Hendler G. 2004 Designing efficient microlens arrays: lessons from  
377 Nature. *J. Mater. Chem.* **14**, 2066.
- 378 11. Hendler G. 1984 Brittlestar color-change and phototaxis (Echinodermata:  
379 Ophiuroidea: Ophiocomidae). *Mar. Ecol.* **5**, 379–401.
- 380 12. Hendler G, Byrne M. 1987 Fine structure of the dorsal arm plate of *Ophiocoma*  
381 *wendtii*: Evidence for a photoreceptor system (Echinodermata, Ophiuroidea).  
382 *Zoomorphology* **107**, 261–272.
- 383 13. Cobb JLS, Hendler G. 1990 Neurophysiological characterisation of the photoreceptor  
384 system in a brittlestar, *Ophiocoma wendtii* (Echinodermata: Ophiuroidea). *Comp.*  
385 *Biochem. Physiol. A* **97**, 329–333.
- 386 14. Gorzelak P, Salamon MA, Lach R, Loba M, Ferré B. 2014 Microlens arrays in the  
387 complex visual system of Cretaceous echinoderms. *Nat. Commun.* **5**, 3576, 6pp.

- 388 15. Polishchuk I *et al.* 2017 Coherently aligned nanoparticles within a biogenic single  
389 crystal: A biological prestressing strategy. *Science*. **358**, 1294–1298.
- 390 16. Ullrich-Lüter EM, Dupont S, Arboleda E, Hausen H, Arnone MI. 2011 Unique system  
391 of photoreceptors in sea urchin tube feet. *Proc. Natl. Acad. Sci. U. S. A.* **108**, 8367–72.
- 392 17. Woodley JD. 1982 Photosensitivity in *Diadema antillarum*: does it show scototaxis? In  
393 *Echinoderms: Tampa Bay: Proceedings of the International Echinoderm Conference*  
394 (ed JM Lawrence), p. 61. Rotterdam.
- 395 18. Speiser DI, Eernisse DJ, Johnsen S. 2011 A chiton uses aragonite lenses to form  
396 images. *Curr. Biol.* **21**, 665–70.
- 397 19. Yang S, Aizenberg J. 2005 Microlens arrays with integrated pores. *Nano Today*, 40–  
398 46.
- 399 20. Vukusic P, Sambles JR. 2003 Photonic structures in biology. *Nature* **424**, 852–855.
- 400 21. Mashanov V, Zueva O, Rubilar T, Epherra L, García-Arrarás JE. 2015 Echinodermata.  
401 In *Structure and Evolution of Invertebrate Nervous Systems*, pp. 665–688. Oxford:  
402 Oxford University Press.
- 403 22. Vinogradova E, Ruíz-Zepeda F, Plascencia-Villa G, José-Yacamán M. 2016 Calcitic  
404 microlens arrays in *Archaster typicus*: microstructural evidence for an advanced  
405 photoreception system in modern starfish. *Zoomorphology* **135**, 83–87.
- 406 23. Burke RD *et al.* 2006 A genomic view of the sea urchin nervous system. *Dev. Biol.*  
407 **300**, 434–460.
- 408 24. Rosenberg R, Lundberg L. 2004 Photoperiodic activity pattern in the brittle star

- 409 *Amphiura filiformis*. *Mar. Biol.* **145**, 651–656.
- 410 25. Delroisse J, Mallefet J, Flammang P. 2016 De novo adult transcriptomes of two  
411 European brittle stars: spotlight on opsin-based photoreception. *PLoS One* **11**,  
412 e0152988.
- 413 26. Delroisse J, Ullrich-Lüter E, Ortega-Martinez O, Dupont S, Arnone M-I, Mallefet J,  
414 Flammang P. 2014 High opsin diversity in a non-visual infaunal brittle star. *BMC*  
415 *Genomics* **15**, 1035.
- 416 27. Gorzelak P, Rahman IA, Zamora S, Gasinski A, Trzcinski J, Brachaniec T, Salamon  
417 MA. 2017 Towards a better understanding of the origins of microlens arrays in  
418 Mesozoic ophiuroids and asteroids. *Evol. Biol.* **44**, 339–346.
- 419 28. Cobb JLS, Moore A. 1989 Studies on the integration of sensory information by the  
420 nervous system of the brittlestar *Ophiura ophiura*. *Mar. Behav. Physiol.* **14**, 211–222.
- 421 29. Raible F, Tessmar-Raible K, Arboleda E, Kaller T, Bork P, Arendt D, Arnone MI.  
422 2006 Opsins and clusters of sensory G-protein-coupled receptors in the sea urchin  
423 genome. *Dev. Biol.* **300**, 461–475.
- 424 30. Lesser MP, Carleton KL, Böttger S A, Barry TM, Walker CW. 2011 Sea urchin tube  
425 feet are photosensory organs that express a rhabdomeric-like opsin and PAX6. *Proc.*  
426 *Biol. Sci.* **278**, 3371–9.
- 427 31. D’Aniello S *et al.* 2015 Opsin evolution in the Ambulacraria. *Mar. Genomics* **24**, 177–  
428 183.
- 429 32. Ramirez MD, Pairett AN, Pankey MS, Serb JM, Speiser DI, Swafford AJ, Oakley TH.  
430 2016 The last common ancestor of most bilaterian animals possessed at least nine

- 431 opsins. *Genome Biol. Evol.* **8**, 3640–3652.
- 432 33. Garm A, Nilsson D-E. 2014 Visual navigation in starfish: first evidence for the use of  
433 vision and eyes in starfish. *Proc. R. Soc. London. Ser. B, Biol. Sci.* **281**, 1–8.
- 434 34. Sutton MD, Garwood RJ, Siveter DJ, Siveter DJ. 2012 SPIERS and VAXML; A  
435 software toolkit for tomographic visualisation and a format for virtual specimen  
436 interchange. *Paleontol. Electron.* **15**, 1–15.
- 437 35. Schindelin J *et al.* 2012 Fiji - an Open Source platform for biological image analysis.  
438 *Nat. Methods* **9**, 676–682.
- 439 36. Whitfield PJ, Emson RH. 1983 Presumptive ciliated receptors associated with the  
440 fibrillar glands of the spines of the echinoderm *Amphipholis squamata*. *Cell Tissue*  
441 *Res.* **232**, 609–624.
- 442 37. Reichensperger A. 1908 Die Drüsengebilde der Ophiuren. *Zeitschrift für*  
443 *Wissenschaftliche Zool.* **91**, 304–350.
- 444 38. Märkel K, Röser U. 1985 Comparative morphology of echinoderm calcified tissues:  
445 Histology and ultrastructure of ophiuroid scales (Echinodermata, Ophiuroida).  
446 *Zoomorphology* **105**, 197–207.
- 447 39. Moore PA, Cobb JLS. 1986 Neurophysiological studies on the detection of mechanical  
448 stimuli by *Ophiura ophiura* (L.). *J. Exp. Mar. Bio. Ecol.* **104**, 125–141.
- 449 40. Byrne M. 1994 Ophiuroidea. In *Microscopic Anatomy of Invertebrates, Volume 14:*  
450 *Echinodermata* (eds FW Harrison, FS Chia), pp. 247–343.
- 451 41. Blevins E, Johnsen S. 2004 Spatial vision in the echinoid genus *Echinometra*. *J. Exp.*

- 452 *Biol.* **207**, 4249–53. (doi:10.1242/jeb.01286)
- 453 42. Sides EM, Woodley JD. 1985 Niche separation in three species of *Ophiocoma*  
454 (Echinodermata: Ophiuroidea) in Jamaica, West Indies. *Bull. Mar. Sci.* **36**, 701–715.
- 455 43. Land MF. 1965 Image formation by a concave reflector in the eye of the scallop,  
456 *Pecten maximus*. *J. Physiol.* **179**, 138–153.
- 457 44. Bok MJ, Capa M, Nilsson DE. 2016 Here, there and everywhere: the radiolar eyes of  
458 fan worms (Annelida, Sabellidae). *Integr. Comp. Biol.* **56**, 784–795.
- 459 45. Nilsson D-E, Gislen L, Coates MM, Skogh C, Garm A. 2005 Advanced optics in a  
460 jellyfish eye. *Nature* **435**, 201–205.
- 461 46. D’Aniello S *et al.* 2015 Opsin evolution in the Ambulacraria. *Mar. Genomics* **24**, 177–  
462 183.
- 463 47. Müller WEG, Wang X, Schröder HC, Korzhev M, Grebenjuk VA, Markl JS, Jochum  
464 KP, Pisignano D, Wiens M. 2010 A cryptochrome-based photosensory system in the  
465 siliceous sponge *Suberites domuncula* (Demospongiae). *FEBS J.* **277**, 1182–1201.
- 466 48. Leung NY, Montell C. 2017 Unconventional Roles of Opsins. *Annu. Rev. Cell Dev.*  
467 *Biol.* **33**, 241–264.
- 468 49. O’Hara TD, Hugall AF, Thuy B, Stöhr S, Martynov A V. 2017 Molecular  
469 phylogenetics and evolution restructuring higher taxonomy using broad-scale  
470 phylogenomics: The living Ophiuroidea. *Mol. Phylogenet. Evol.* **107**, 415–430.
- 471 50. Richter S *et al.* 2010 Invertebrate neurophylogeny: suggested terms and definitions for  
472 a neuroanatomical glossary. *Front. Zool.* **7**, 29.

- 473 51. Land MF, Nilsson D-E. 2012 *Animal Eyes*. Second. Oxford: Oxford University Press.
- 474 52. Nilsson D-E. 2009 The evolution of eyes and visually guided behaviour. *Philos. Trans.*  
475 *R. Soc. Lond. B. Biol. Sci.* **364**, 2833–2847.
- 476 53. Petie R, Garm A, Hall MR. 2016 Crown-of-thorns starfish have true image forming  
477 vision. *Front. Zool.* **13**, 41.
- 478
- 479

480 **Figure 1. Expanded peripheral trabeculae (EPTs), skeletal structures in *Ophiocoma***  
481 ***wendtii*.** Synchrotron X-ray tomography of arm segments. Hemispherical calcite structures  
482 previously characterised as lenses (dashed outlines) on the dorsal (A, A'), lateral (B) and, to a  
483 lesser extent, ventral (C) arm plates. *In vivo*, arm plates are covered by the cuticle, which  
484 obscures the regular form of the EPTs, and is interspersed by ciliary projections (arrowhead)  
485 (A'). In cross section (D), the continuous nature of the EPTs with the rest of the stereom is  
486 visible, particularly in the lateral regions (arrowhead). See supplementary materials (S1) for  
487 reconstructed model.

488 **Figure 2. Calcite elements on the arm plates in *Ophiocoma echinata* and *O. pumila***  
489 **visualised by synchrotron X-ray tomography.** *Ophiocoma echinata* (A–E) is covered with  
490 very regular, hemispherical EPTs on the dorsal arm plates (A, B, C), ventral arm plates (D),  
491 and the dorsal and ventral regions of the lateral (A, E) arm plates. The EPTs are surrounded  
492 by pigmented chromatophores giving a dark colour (B). *Ophiocoma pumila* (F–J) lacks  
493 chromatophores and appears much paler (G). The skeletal elements are less regular than the  
494 EPTs observed in *O. wendtii* (Figure 1) and *O. echinata* (A–E), but EPT-like hemispheres are  
495 present across the dorsal arm plates (F, H), margins of the lateral arm plates (I), and ventral  
496 arm plates (J). See supplementary materials (S2–3) for reconstructed models. Scale bars: A,  
497 F, 250 µm; B, G, 500 µm; C–E, H–J, 25 µm.

498 **Figure 3. Opsin-reactive cells are arranged between the EPTs in *Ophiocoma wendtii*.** A,  
499 A': Cells reactive to a sea urchin rhabdomeric opsin (Sp-Op4, red) and acetylated tubulin  
500 (green) are arranged around the distal part of the EPTs (dashed outlines) on the dorsal arm  
501 plate (DAP). Dorsal view of arm plate, with stack reaching slightly beneath plate surface. B,  
502 C, D: Stacked images of transverse sections through the DAP show the distal projection of  
503 nerves between EPTs towards the surface of the arm (B, D, arrowheads), originating from an



504 underlying lateral nerve (**B**) and terminating in multiciliary bundles at the surface (**C**).  
505 Proximal side of the plate is at the bottom of the image. Note that images in both planes show  
506 no opsin-reactive cells present at the focal point of the EPTs as predicted by [9]. Chr,  
507 chromatophore; EPT, expanded peripheral trabecula; ner, nerve.

508 **Figure 4. An expansive system of opsin-reactive cells and “lens”-like skeletal structures**  
509 **is also present in *Ophiocoma pumila*.** **A**, Horizontal section through dorsal arm plate (DAP,  
510 dashed outline) in *O. pumila* demonstrates the same innervation as *O. wendtii*, with a median  
511 nerve and paired, branching nerves (acetylated tubulin, green) extending laterally. Reactivity  
512 to the c-opsin Sp-Op1 is visible inconsistently across the plate surface and within the median  
513 nerve. Dorsal view. **B**, Surface of DAP reconstructed from synchrotron scan, with EPT-like  
514 structures (dashed outline) among more irregularly shaped stereom elements. Dorsal view. **C**,  
515 **D**, Transverse sections through the arm plate show projections from the lateral nerve  
516 (arrowheads) to opsin-reactive cells and ciliary tufts at the surface, between the EPT-like  
517 structures. Chr, chromatophore; EPT, expanded peripheral trabecula; lat ner, lateral nerve;  
518 med ner, median nerve; ner, nerve bundles.