



# THE UNIVERSITY *of* EDINBURGH

## Edinburgh Research Explorer

### **Invited review: Genomic selection for small ruminants in developed countries: how applicable for the rest of the world?**

**Citation for published version:**

Mrode, R, Tarekegn, GM, Mwacharo, JM & Djikeng, A 2018, 'Invited review: Genomic selection for small ruminants in developed countries: how applicable for the rest of the world?' *Animal*. DOI: 10.1017/S1751731117003688

**Digital Object Identifier (DOI):**

[10.1017/S1751731117003688](https://doi.org/10.1017/S1751731117003688)

**Link:**

[Link to publication record in Edinburgh Research Explorer](#)

**Document Version:**

Peer reviewed version

**Published In:**

Animal

**General rights**

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

**Take down policy**

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact [openaccess@ed.ac.uk](mailto:openaccess@ed.ac.uk) providing details, and we will remove access to the work immediately and investigate your claim.



1 **Invited Review: Genomic selection for small ruminants in Europe and other**  
2 **developed countries: How applicable for the rest of the world?**

3  
4 R. Mrode<sup>1,2</sup>, G. Mekuriaw Tarekegn<sup>3,a</sup>, J.M. Mwacharo<sup>4</sup> and A. Djikeng<sup>5,b</sup>

5  
6 <sup>1</sup>*Animal Biosciences, International Livestock Research Institute (ILRI), P.O. Box 30197 Nairobi*  
7 *00100, Kenya*

8 <sup>2</sup>*Scotland Rural College, Peter Wilson Building, Kings Buildings, West Mains Road, Edinburgh,*  
9 *EH9 3JG.*

10 <sup>3</sup>*Department of Animal Production and Technology, Biotechnology Research Institute, Bahir Dar*  
11 *University, P.O. Box 79 Ethiopia, Addis Ababa*

12 <sup>4</sup>*Small Ruminant Genetics and Genomics Group, International Centre for Agricultural Research*  
13 *in the Dry Areas (ICARDA), P.O. Box 5689 Addis Ababa Ethiopia*

14 <sup>5</sup>*Biosciences Eastern and Central Africa-International Livestock Research Institute (BecA-ILRI*  
15 *Hub), P.O. Box 30197 Nairobi 00100, Kenya*

16  
17 <sup>a</sup>*Present address: Swedish University of Agricultural Sciences, Uppsala Sweden*

18 <sup>b</sup>*Present address: Centre for Tropical Livestock Genetics and Health. The Roslin Institute &*  
19 *Royal (Dick) School of Veterinary Studies. Easter Bush, Midlothian, EH25 9RG, Scotland*

20  
21 Corresponding author: Raphael Mrode. Email: [R.Mrode@cgiar.org](mailto:R.Mrode@cgiar.org)

22  
23 **Abstract**

24 Improved management and use of estimated breeding values in breeding programs,  
25 have resulted in rapid genetic progress for small ruminants (SR) in Europe and other  
26 developed countries. The development of SNP (single nucleotide polymorphisms) Chips  
27 opened opportunities for genomic selection (GS) in SR in these countries. Initially  
28 focused on production traits (growth and milk), GS has been extended to functional  
29 traits (reproductive performance, disease resistance and meat quality). The GS systems  
30 have been characterized by smaller reference populations compared with those of dairy  
31 cattle and consisting mostly of cross- or multi-breed populations. Molecular information  
32 has resulted in gains in accuracy of between 0.05 and 0.27 and proved useful in  
33 parentage verification and the identification of QTLs for economically important traits.  
34 Except for a few established breeds with some degree of infrastructure, the basic

35 building blocks to support conventional breeding programs in small holder systems are  
36 lacking in most developing countries. In these systems, molecular data could offer quick  
37 wins in undertaking parentage verification and genetic evaluations using **G** matrix, and  
38 determination of breed composition. The development of next-generation molecular  
39 tools has prompted investigations on genome-wide signatures of selection for mainly  
40 adaptive and reproduction traits in SR in developing countries. Here, the relevance of  
41 the developments and application of GS and other molecular tools in developed  
42 countries to developing countries context is examined. Worth noting is that in the latter,  
43 the application of GS in SR will not be a “one-size fits all” scenario. For breeds with  
44 some degree of conventional genetic improvement, classical GS may be feasible. In  
45 smallholder systems, where production is key, community based breeding programs  
46 can provide the framework to implement GS. However, in fragile growth systems, e.g.  
47 those found in marginal environments, innovative GS to maximize adaptive diversity will  
48 be required. A cost-benefit analysis should accompany any strategy of implementing  
49 GS in these systems.

50

51 **Key words:** Small ruminants, genomic selection, signatures of selection, QTL

52

### 53 **Implication**

54 The basic building blocks for conventional breeding programs for small ruminants in  
55 most developing countries are lacking. However, genomic data offers unique  
56 opportunities to circumvent some of the limitations through parent verification, genetic  
57 evaluations using the G matrix and understanding the molecular basis of adaptation

58 through GWAS. The application of genomic selection may however need to be tailored  
59 to the conditions of specific production environments e.g. smallholder verses pastoral  
60 systems.

61

## 62 **Introduction: Role of small ruminants (SR) in developing countries**

63 Globally, the largest number of SR occur in Asia (49.70%), followed by Africa (27.90%)  
64 and then Europe (8.70 %), summing up to 86.3% of world total (FAOSTAT 2013). SR  
65 meat and milk production represents 4.8% and 3.4% of the total meat and milk  
66 produced, respectively, in the world. These percentages are comparatively smaller in  
67 developed (3.0% and 1.6%) than in developing countries (6.2% and 6.1%, respectively),  
68 emphasizing the significant role of SR in developing countries. In addition, SR offer a  
69 wide range of products in developing countries including skins, manure and  
70 (mo)hair/pelts, and play critical socio-cultural roles in many communities (Kosgey and  
71 Okeyo, 2007). They also represent a large repository of genetic diversity that is well  
72 adapted to diverse agro-ecologies and are critical to the poor in marginal areas where  
73 arable agriculture is too risky or rearing cattle is not feasible (Devendra, 2002). The  
74 production systems in Europe are based mostly on improved management and well  
75 defined and structured breeding programs, while about 70 to 85% of SR products are  
76 derived from the smallholder and pastoral systems in developing countries. The  
77 smallholder and pastoral systems are low-input, characterized by small flock sizes, lack  
78 of infrastructure and animals of unimproved genotypes. Most often, higher productivity  
79 is not usually the goal trait, especially when production risks are high (Amer *et al.*,  
80 1998).

81

82 Current advances in molecular biology has resulted in the discovery of unprecedented  
83 levels of genomic variation as a result of sequencing efforts, and consequently, the  
84 development of various single nucleotide polymorphisms (SNP) chips for genotyping  
85 purposes. The reduction in genotyping costs and advances in statistical methods  
86 (Meuwissen, *et al.*, 2001), has made it possible to incorporate molecular information in  
87 SR breeding programs in many European and developed countries to accelerate the  
88 rate of genetic progress in production and somewhat difficult to measure traits. The  
89 question that arises therefore is how applicable are the molecular based methods  
90 including genomic selection (GS) to the rest of the world especially in developing  
91 countries. This review presents an overview of GS and other molecular based methods  
92 in the improvement of SR in the developed countries and then examines their potential  
93 and feasibility for application in the developing countries.

94

### 95 **Systems for conventional breeding programs in developed countries for SR**

96 Fundamental to the implementation of GS is the existence of an already established  
97 system of genetic evaluation based on efficient performance and pedigree recording. In  
98 an attempt to increase the efficiency of the productivity of SR, many developed  
99 countries have implemented breeding programs based on estimated breeding values  
100 (EBVs) using performance and pedigree data. The maturity in mixed model approaches  
101 (Henderson, 1949) has resulted in more accurate estimates of EBVs accelerating the  
102 rate of genetic progress and the profitability of SR enterprises. For instance, in the New  
103 Zealand sheep industry there was an 83% increase in kg of lamb produced per ewe and

104 up to 28% overall in carcass weight from 1990 to 2012 (Beef and Lamb NZ, 2012).  
105 Examples of established well-structured genetic evaluation systems that underpin such  
106 genetic improvement programs for SR include Basco database for sheep and beef  
107 improvement in the United Kingdom (<http://www.basco.org/sheep>), French genetics for  
108 cattle, sheep and goats (<http://en.france-genetique-elevage.org>), Sheep Improvement  
109 Limited - SIL in New Zealand (<https://www.sil.co.nz/>), and Canadian dairy goat breeding  
110 program (<http://www.goatgenetics.ca/>).

111  
112 These improvement programs for SR are mostly focused on meat, wool, and dairy  
113 production, and more recently, breeding objectives have also included other functional  
114 traits such as reproductive performance and disease resistance/tolerance but little  
115 emphasis on carcass and meat quality traits (Pannier *et al.*, 2014). While rapid rates of  
116 genetic progress for growth-related or milk traits have been achieved in these programs,  
117 a relatively lower rate of progress is possible for traits that are measured later in the life  
118 of females, such as reproductive ability, breeding seasonality and longevity (Rupp *et al.*,  
119 2016) due to the longer generation interval or, in carcass composition traits which are  
120 recorded on the relatives of selection candidates and require animals to be sacrificed  
121 (Daetwyler *et al.*, 2012).

122

### 123 **Overview of GS and molecular approaches in developed countries for SR**

124 The advent of GS and genome wide association studies (GWAS) opened new  
125 opportunities for breeding programs in SR especially for traits measured late in life and  
126 carcass traits. These opportunities in GS and GWAS resulted from the development of

127 next-generation sequencing technologies which allowed de novo sequencing of sheep  
128 and goat genomes; and the subsequent development of dense SNP Chips such as the  
129 Illumina Goat SNP50 BeadChip (Tosser-Klopp *et al.*, 2014), the Ovine SNP50  
130 BeadChip (Kijas *et al.*, 2009) and recently the Ovine 600K SNP BeadChip (Anderson *et*  
131 *al.*, 2014). Recently, a low density panel with 16301 SNPs for sheep has been  
132 developed by the International Sheep Genomics Consortium (Larroque *et al.*, 2017)

133  
134 The basic principle undergirding GS is that SNPs are assumed to be at LD with QTLs in  
135 the genome. Therefore the use of SNPs as markers enables all QTLs in the genome to  
136 be identified through the mapping of chromosome segments defined by adjacent SNPs.  
137 The implementation of GS usually involves estimating the SNP effects in a reference  
138 population which consists of individuals with phenotypic records and genotypes. This is  
139 then followed by prediction of genomic estimated breeding values (GEBV) for selection  
140 candidates with no phenotypes of their own (Meuwissen, *et al.*, 2001). Details of the  
141 design of actual GS in SR have been described by Rupp *et al.* (2016).

142  
143 Genomic predictions and selection in SR for developed countries have either been  
144 successfully implemented or their feasibility demonstrated on a number of standard  
145 production traits such as wool, growth traits, muscle and fat depth in New Zealand  
146 (Auvray *et al.*, 2014), Australia (Daetwyler *et al.*, 2010), in dairy sheep and goats in  
147 France (Carillier *et al.*, 2014) and in dairy goats in the UK (Mucha *et al.*, 2015). Recently  
148 GS in SR has been applied to breed for disease resistance such as parasite and fly-  
149 strike resistance (Pickering *et al.*, 2015) and facial eczema (Phua *et al.*, 2014). There is

150 also on-going work on genomic prediction for traits such as feeding efficiency and  
151 methane emissions (Pickering *et al.*, 2015).

152  
153 The characteristics of these genomic prediction systems for SR include reference  
154 populations of smaller sizes compared to dairy cattle and consisting of mostly cross-  
155 breeds or multi-breed populations. In summary, the reference populations ranged from  
156 1,900 for Western Pyrenees dairy sheep breeds to 8,000 multi-breed Australian meat  
157 sheep (Rupp *et al.*, 2016). The gains in accuracy provided by molecular information are  
158 rather lower (range from 0.05 to 0.27) given the small size of the reference populations.  
159 Details of accuracies from studies on genomic predictions for SR are outlined by Rupp,  
160 *et al.* (2016).

161  
162 Genomics has offered the opportunity to identify and include major genes (QTLs)  
163 associated with reproductive, disease, or production traits. A comprehensive list of such  
164 QTLs is outlined by Rupp, *et al.* (2016) including some of the genes that are already  
165 being used in breeding programs, such as *PrP*, *FecL* or the  $\alpha$ -s1 casein (French goats),  
166 to pre-select candidates for progeny testing. In recent times, genomic approaches have  
167 been used to identify novel mutations influencing functional traits. For instance, Demars  
168 *et al.* (2013) used GWAS and identified new mutations associated with prolificacy in  
169 sheep. The discovery of actual genes and causative mutations underlying prolificacy  
170 has been a subject of intense investigation in sheep in developed countries. The  
171 findings have paved the way for the development of commercial DNA assays/tests/Kits,  
172 which require no parental information, to identify breeding stock with high prolificacy.



173 Such tests have been developed for the Inverdale (*FecX<sup>l</sup>*) and Boorola (*FecB<sup>B</sup>*)  
174 mutations and are commercially available in Australian and New Zealand sheep  
175 industry where rams are tested to breed heterozygous progenies (Davis 2005;  
176 Walkden-Brown *et al.* 2009).

177  
178 In addition to genomic prediction, the use of genotypic information plays an important  
179 role in parentage verification and assignment in SR in developed countries. In breeding  
180 schemes for SR, parentage identification is an issue due to the limited use of artificial  
181 insemination and use of natural mating, involving most likely multiple sires, in extensive  
182 systems. In these natural mating schemes parentage is either unknown or incomplete  
183 and the use of genetic markers, initially microsatellites and currently SNPs, have proved  
184 useful to detect misidentified and unknown parents. For details of the various SNP chips  
185 available for parentage verification, the reader should see Rupp *et al.* (2016). In  
186 addition to parentage identification, genotypic information is useful for assessing genetic  
187 diversity and structure of local sheep and goat breeds. Genotypic data gives more  
188 accurate estimates of relationship between individuals than pedigree records and  
189 therefore offers better opportunities for more accurate estimation of co-ancestry, mate  
190 assignment, and inbreeding coefficients (Rupp *et al.* (2016).

191  
192 **Summary of some breed improvement programs for SR in developing countries**

193 The existence of well-established conventional genetic evaluation and selection  
194 programs provide the necessary platform for the implementation of GS. In most  
195 developing countries, genetic improvement programs for SR are scarce. The major

196 constraints include lack of performance and pedigree information and the non-existence  
197 of institutional frameworks and infrastructure including inadequate farmers'  
198 organizations at the village level to effectively participate in breeding schemes (Kosgey  
199 and Okeyo, 2007). Most of the production occurs in small holder systems which are  
200 characterized by small flock sizes, uncontrolled mating and lack of pedigree recording  
201 and therefore the difficulty of defining adequate contemporary groups. However in a few  
202 countries, breeding improvement programs for SR have been implemented, and these  
203 are briefly summarized.

204

205 *(i) Kenya Dual Purpose Goat Development (KDPG) Project*

206 The KDPG breeding program was started in 1980 as part of the Small-Ruminant  
207 Collaborative Research Support Program (SR-CRSP) funded by the United States  
208 Agency for International Development (USAID) and implemented by Kenya's Ministry of  
209 Livestock Development. The overall objective was to develop a synthetic breed of goat  
210 that combined the adaptability of the indigenous East African and Galla goats and the  
211 growth and milk producing abilities of the Toggenburg and Anglo-Nubian breeds. Ojango  
212 et al (2010) provides a detailed summary of the breeding program for the KDGP goat.

213

214 The foundation flock consisted of 250 Small East African (E) goats from across Kenya  
215 and 200 Galla (G) goats sourced from the dry Northeastern province of Kenya. With no  
216 production data available, these animals were selected based on phenotypic  
217 characteristics such as large and sound udders and teats and the local "milk line" claim,  
218 a distinctive black stripe along the back of some Galla goats. These were initially mated

219 to different Toggenburg and Anglo-Nubian bucks, and later insemination was done  
220 using semen from the USA. A nucleus breed was established at Ol-Magogo Estate of  
221 the National Animal Research Centre, Naivasha (Mwandotto *et al.* 1992), where  
222 productivity and pedigree recording was undertaken by enumerators. An  
223 interdisciplinary farming systems approach was used (Ojango *et al.* 2010) to develop  
224 and test the breeding program (Semenye *et al.* 1989). KDPG development occurred at  
225 Ol-Magogo Estate, while breeding animals were provided to a station in Maseno  
226 Western Kenya, which was closer to the target farmers. On-farm testing of the KDPG  
227 was carried out by smallholder farmers from contrasting socio-cultural and  
228 environmental backgrounds. Each farmer received 2-4 breeding does and breeding  
229 bucks were rotated amongst groups of farmers. The project developed the KDPG  
230 breed. On-farm the KDGP reached their milk peak after one week of kidding, producing  
231 600ml per day for household use while On-station, it reached peak milk production,  
232 three weeks after kidding, producing 1500ml per day (Onim 1992). In on-farm trials, the  
233 KDPG produced on average 0.49 litres per day with a range of between 0.05 and 2.70  
234 litres per day (Semenye *et al.*, 1989). The local does at the station peaked after six  
235 weeks of kidding, producing a daily milk production of 400 ml (Onim 1992). At the peak  
236 of its operation, a breeding flock of the KDPG established at Ol-Magogo Estate stood at  
237 1800 animals. By 2005, the population was less than 400 animals (Bett, 2005) due to  
238 the termination of the breeding and farmer development program for the KDPG and the  
239 SR-CRSP project (Ojango *et al.* 2010). Within the last decade, there has been renewed  
240 interest in the KDPG and a re-evaluation of its breeding strategies (Ojango *et al.*, 2010).  
241

242

243

244 *(ii) Community Based Breeding Programs (CBBPs) for Sheep and goats in Ethiopia*

245 The International Center for Agricultural Research in the Dry Areas (ICARDA), the

246 International Livestock Research Institute (ILRI), and Austria's University of Natural

247 Resources and Life Sciences, in partnership with the Ethiopian National Agricultural

248 Research System (ENARS), have designed and implemented community-based SR

249 breeding programs in Ethiopia since 2009 (Haile *et al.*, 2014). Similar CBBP for

250 indigenous goats of Ethiopia and Cameroon were also implemented by Biosciences

251 Eastern and Central Africa (BecA-ILRI) Hub in 2013 for three production systems (arid

252 agro-pastoral, semi-arid agro-pastoral and highland mixed crop-livestock systems;

253 Woldu *et al.*, 2016). The CBBP are designed to take into account farmers' needs, views,

254 decisions, and active participation, from inception to implementation, and their success

255 is based upon proper consideration of farmers' breeding objectives, infrastructure,

256 participation, and ownership (Wurzinger *et al.* 2011). The goal of CBBPs is to improve

257 the productivity and income of small-scale resource-poor SR producers by providing

258 access to improved animals that respond to improved feeding and management, and

259 facilitating the targeting of specific market opportunities.

260

261 There is a governmental rural organization associated with each of the sites where the

262 CBBPs are in operation. Local enumerators are recruited for each site to assist the

263 research system in animal identification and recording. Indigenous knowledge of the

264 community is considered at each phase of the project. For example, the community

265 decides how rams are managed and how they are shared and used. The aim is to get  
266 community members to work as a team in selecting, managing and using rams. Two  
267 stages of selection are applied, initial screening when first sales of young rams occur  
268 (4–6 months) and final selection for admission to breeding at 12 months of age.  
269 Selection at the first stage is based on 6 months weight and ewe lambing interval.  
270 Yearling weights and body conformation are considered in the final selection.

271

272 There are currently 23 CBBPs across Ethiopia operating in 15 sheep and 8 goat sites.  
273 Each CBBP involves an average of 60 households per site and 600 flocks with an  
274 average flock size of 10 animals. To automate the recording and ensure real-time  
275 archiving, an online database “DREMS” (Data Recording and Management System)  
276 was developed (jointly by EMBRAPPA-Brazil and ICARDA). In DREMS, data can be  
277 keyed-in offline from a mobile device (tablet, computer, mobile phone etc.) and updated  
278 once online. The information is archived in a server maintained at EMBRAPPA-Brazil.

279

### 280 *(iii) Goat improvement programs in South Africa*

281 The South Africa Boer Goat Breeders’ Association was formed in 1959 but development  
282 of the Boer goat as a meat breed dates back to 1918. The National Performance  
283 Testing Scheme however, commenced in 1970. Two other dairy breeds were further  
284 developed from the Boer goat. These were the White Savanna, which was initiated in  
285 1957 and a breed society formed in 1993 and the Kalahari Red which started in 1990.  
286 Genetic improvement of these dairy goats is still based on the convectional hand and  
287 eye method (Casey and Webb, 2010) and the South African Studbook Association and

288 Milch Goat Breeders Society handles records of goat breeds and milk production.

289 Genetic progress is rather slow, but substantial amount of genetic improvement has  
290 been realized in the past especially in the meat goat sector.

291

292 Animal recording in the mohair producing Angora goats, was piloted in 1983, with the  
293 approval of the Angora Stud Breeders' Society. This was followed by the closure of the  
294 Angora herd book in 1984 and in 1999 animal recording for the Angora goat was  
295 operationalized within the National Small Stock Information Scheme of South Africa.

296 The breeding program for the Angora goat in the South African mohair production  
297 systems was designed on the basis of the study by Snyman and Olivier (1999). The  
298 initial selection index was based on fibre diameter, fleece weight and body weight.

299 Intensive selection for increased mohair production from the early 1970s until 1990 with  
300 no selection directed towards weaning weight, resulted in unthrifty animals with an  
301 inability to survive sub-optimal conditions (Visser and Van Marle Köster, 2014). The  
302 selection strategy was re-evaluated in 2002 and it was concluded that selection for  
303 decreased fibre diameter, while maintaining or increasing body weight and fleece weight  
304 seems optimum for the breed. In addition, molecular research has been undertaken with  
305 a view of including molecular information in the breeding program. A microsatellite  
306 marker panel consisting of 14 markers has been developed and utilized for parentage  
307 verification in the Breed (Visser *et al.*, 2011a). Similarly eighteen QTLs for mohair traits  
308 including fleece weight, fibre diameter and other related traits have been identified on  
309 thirteen chromosomes (Visser *et al.*, 2011b).

310

311 (iv) Goat and Sheep improvement programs in India  
312 A goat improvement program involving 34 villages was initiated by the Nimbkar  
313 Agricultural Research Institute in 1991 in South-Central Maharashtra of India with the  
314 aim of improving goat productivity through cross-breeding (Nimbka, 1991). Thirty-four  
315 villages within a 15 km radius of Phaltan town in South-Central Maharashtra province  
316 formed the target area for the cross-breeding project. It involved 13 Sirohi bucks  
317 selected on their individual growth rates and their mothers' milk yields, ten Alpine x  
318 Sirohi and ten Toggenburg x Sirohi bucks which were bought and introduced into the  
319 project. The improved bucks were placed in the villages for cross-breeding of local  
320 goats and no efforts were made for the dissemination of cross-bred males and females  
321 generated in the course of the project. The project was supported by veterinarians, who  
322 visited each project village once a week. The project ran for four years but collapsed  
323 due to lack of funds. The author concluded that it provided a framework for an effective  
324 breeding program when individual units are small and spread out over a large area.  
325 Similar cross breeding program to improve the fecundity of Deccani sheep of  
326 Maharashtra was summarized by Nimbka *et al.*, (2002), which involved the  
327 introgression of the Booroola gene from the Indian Garole breed into the Deccani and a  
328 composite breed.

329 **The relevance of developments in GS and other molecular approaches in**  
330 **developed countries for the rest of the world**

331 *Parentage and breed composition verification*

332 One the possible quick wins from the developments in molecular based approaches and  
333 the utilization of genotypic information in SR breeding in developing countries includes

334 parent verification and breed composition of cross bred animals. The rather extensive  
335 systems for the management of SR in small holder systems and the lack of  
336 infrastructure to capture pedigree information has resulted in the inability to undertake  
337 genetic evaluation in these systems or control breeding. Therefore the availability of  
338 genotypic information will reduce the need for accurate pedigree recording as genomic  
339 relationships can be computed to undertake genetic evaluation, estimate inbreeding and  
340 undertake parentage verification. However most of the initial work on parentage  
341 verification so far in these systems are based on microsatellites. A microsatellite marker  
342 panel consisting of 14 markers has been developed and utilized for parentage  
343 verification in the Angora Breed (Visser *et al.*, 2011a). Similarly, genotypic data can  
344 easily be used through admixture analysis to determine breed composition in cases  
345 where crossbreeding and uncontrolled mating is practiced and therefore be utilized to  
346 match appropriate genotypes to the relevant management systems. However the  
347 utilization of genomic information in the small holder systems for SR is rather slow  
348 compared to dairy cattle where genetic predictions have been undertaken using the **G**  
349 matrix computed using SNP data (Brown *et al.* 2016). In the ILRI led Africa Dairy  
350 Genetic Gains (ADGG) project, a small chip of about 400 SNPs for parentage  
351 verification and breed composition for dairy cattle is being developed (Gibson & Mwai,  
352 personal communication).

353

354 *Detection of signatures of selection and use of molecular markers in breeding in small*  
355 *ruminants*



356 In contrast to SR in Europe and the developed world, SR in the developing countries  
357 remain nondescript in genotype and phenotype, the consequence of modest  
358 anthropological selection. The analysis of microsatellites and recently SNP genotype  
359 and full genome sequence data in SR in the developing world has revealed high genetic  
360 diversity that mirrors their extensive phenotypic diversity as well as the diversity in their  
361 production environments and historical migration and admixture patterns. Analysis of  
362 signatures of selection have revealed candidate regions in the genome harbouring  
363 genes with demonstrated roles in phenotypic variation including fat and thin tail, horn  
364 size and polledness, body morphology, limbs and skeleton development, pigmentation  
365 etc. (Fariello *et al.*, 2014). In a GWAS study, Gholizadeh *et al.* (2015) identified  
366 significant association between a single SNP located in the *SYNE1* gene on  
367 chromosome 8 with yearling weight in Baluchi sheep found across southwest Pakistan,  
368 eastern Iran and southern Afghanistan. Using different approaches, multiple selection  
369 sweep regions spanning several candidate genes relating to various traits (immunity,  
370 nervous and endocrine system development, metabolism, thermo-regulation,  
371 reproduction etc.), metabolic pathways and biological processes driving adaptation to  
372 local environments have been revealed in Black, Draa and Northern goat populations of  
373 Morocco (Benjelloun *et al.*, 2015), various breeds of sheep in Africa, Asia and South  
374 West Asia (Fariello *et al.*, 2014), Barki sheep and goats from Egypt (Kim *et al.*, 2016)  
375 and the indigenous goats in South Africa (Mdladla, 2016). Kim *et al.* (2016) further  
376 identified one selection sweep region that was common to both the Barki sheep and  
377 goats from Egypt providing possible evidence for a common region under selection in a  
378 common environment in the two species. Gouveia *et al.*(2017) identified genomic

379 regions under selection which overlap genes influencing traits associated with  
380 ecological adaptation, phenotypic and production differences amongst three Brazilian  
381 locally adapted sheep breeds (Brazilian Creole, Morada Nova and Santa Ines). It is  
382 worth noting that in most of these studies, the identified selection sweeps and their  
383 genomic distribution differ between populations/breeds but reflect, to a large extent, the  
384 outcomes of local adaptation. This suggests that artificial selection seems to play a  
385 minor role in driving genome evolution in SR in developing countries and natural  
386 selection tends to favour adaptive diversity.

387

388 The characterization of the indigenous goats in Ethiopia and Cameroon populations  
389 using mitochondrial DNA and 50k SNP chip array was also undertaken by Getinet,  
390 (2016). A high level of genetic diversity but weak genetic structure was found among the  
391 goat populations in both countries. However, the Keffa goat, reared in highly tsetse  
392 infested area, and Abergelle goat, also known with its drought tolerance, were found to  
393 have relatively maintained their pure genetic background. Coding regions of the  
394 kisspeptin gene were found in Gondar and Woyto-Guji goats in Ethiopia and the  
395 genotypes detected were associated with multiple births in these goat populations  
396 (Getinet *et al.*, 2016).

397

398 With the exception of Brazil and possibly India, the use of molecular markers in SR  
399 breeding lags behind in most developing countries. In Brazil use of molecular markers in  
400 animal breeding has concentrated on two fronts: those controlled by many genes of  
401 small effect ( meat and milk production, Lôboa, *et al.*, 2010), on which classical

402 breeding is based, or traits controlled by few genes of large effect. The latter have  
403 several examples in sheep such as those linked to prolificacy (booroola, inverdale or  
404 galway), muscle mass (callipyge) or resistance/susceptibility to scrapie (PNRP). Castro  
405 *et al.* (2006) identified a mutation linked to prolificacy specific to naturalized Brazilian  
406 breeds.

407

#### 408 *Potential for the application of genomic selection in developing countries*

409 The production system for SR in developing countries can be considered to occur along  
410 a trajectory in terms of management systems with one end of the spectrum consisting of  
411 breeds operating mostly on a commercial scale basis, having some degree of genetic  
412 improvement and investment on infrastructure while on the end is the fragile growth  
413 systems, mostly the pastoralists/nomads in arid environments. (Smith, *et al.*, 2013).  
414 Opportunities for the application of GS will therefore not be "one size fits all" but very  
415 much dependent on the intersection of the spectrum being considered. Commencing at  
416 one end of the spectrum where production is at a commercial scale, with some  
417 organizational structures (breed societies) and some investments in IT infrastructure,  
418 such as the Boer, Savanna, Kalahari and Angora goat breeds in South Africa, huge  
419 opportunities exist for GS. In this production system, the emphasis is mostly on  
420 productivity traits with less weight put on adaptive traits. The existing structures such as  
421 the progeny testing scheme (Snyman and Olivier, 1999), the availability of performance  
422 and pedigree records imply that classic GS (GBLUP or Single step) could be applied.  
423 Potential improvements in this setting is to translate to the use of digital systems (mobile  
424 phones or tablets) to collect performance data (Mrode *et al.*, 2017), as it avoids or

425 reduces the huge organizational infrastructure and high costs associated with recording  
426 systems. Additional benefit from genomics in this setting is the reduction for the need to  
427 accurately record pedigrees as genomic relationships can be computed and parentage  
428 verification implemented using SNP genotype data.

429

430 In the middle of the spectrum are the small holder systems which account for most (70-  
431 80%) of the outputs from SR. While the emphasis here is still on production traits,  
432 adaptive traits play very significant role, therefore innovative GS will be needed that  
433 ensures adequate balance between production and adaptive traits. However,  
434 community based breeding programs, such as the FARM-Africa Meru and Tharaka-Nithi  
435 Districts dairy goat and animal healthcare project in Kenya among other initiatives,  
436 seem to be the best approaches for implementing GS as the rotational use of selected  
437 males provide opportunity to select superior males using SNP genotype data, given that  
438 performance recording pooled from several flocks by digital means can be initiated or is  
439 already in place. The within breed selection implemented in these projects ensures that  
440 adaptability of local breeds can be monitored overtime and mating can be controlled.  
441 However several scenarios need to be evaluated considering different genotyping  
442 strategies and the economic aspects to determine the best approach for implementing  
443 GS in this setting. In addition, production and adaptive traits can be optimized in the  
444 context of CBBP via gene/genome/haplotype block editing (Jenko *et al.*, 2015) utilizing  
445 the genomic regions identified in combination with GS in developing appropriate  
446 synthetic breeds.

447

448 At the other end of the spectrum is the fragile growth systems consisting mostly of very  
449 extensive systems of the pastoralists and nomads in arid and semi-arid environments,  
450 where adaptive traits are key and the main goal will be to maximize adaptive diversity.  
451 The implementation of GS in this system possess major challenges. However the  
452 widespread usage of the mobile phone in these systems for other purposes, such as  
453 money transfers, imply that digital data capture and recording could be possible with  
454 adequate farmer training. The use of communal grazing lands, watering points and  
455 other services could innovatively be used to introduce recording of basic performance  
456 data, initiate sampling for genotyping of animals and also introduce the use of superior  
457 males. The initial use of such data could be the application of GWAS and investigating  
458 signatures of selection to identify genomic regions associated with various aspects of  
459 adaptability (disease and drought for instance). As more data accumulates, genomic  
460 data will allow for a better understanding of genetic diversity in the fragile growth sector  
461 and how to select for it: for instance, the use of weighted GBLUP or Bayesian methods  
462 to optimize various aspects of adaptability. In the long term, usage of gene editing in  
463 addition to GS to increase and optimize the frequency of favourable alleles associated  
464 with different aspects of adaptability (Jenko *et al.*, 2015) could be a possibility.

465

466

### 467 **Economic aspects of genomic selection in developing countries**

468 Given the significant role that SR play in the livelihood of farmers, the implementation of  
469 GS in the various management systems described especially the low-input small holder  
470 system and the fragile growth sectors should be accompanied by a cost-benefit

471 analysis. The bottom line is that the introduction of GS should financially be beneficial to  
472 farmers and produce animals that are able to fulfil the other socio-cultural roles played  
473 by SR in the community. The relatively high economic efficiency of GS in the dairy cattle  
474 is derived mostly from the large reduction in generation interval (König et al 2008). In  
475 small ruminants, the reduction in generation interval is not as large (Larroque et al,  
476 2017) and the relatively higher cost of genotyping limits the cost-effectiveness of GS.  
477 Shumbusho et al (2016) found that GS alone was not more beneficial in a French meat  
478 sheep breed compared with classical selection except when compared with some early  
479 measured phenotypes. However the introduction of the low density chip (16k) for sheep  
480 increases the prospects of higher economic returns from GS. Larroque *et al.*,(2017)  
481 demonstrated very high accuracy of imputation of the 16K chip to the 50k Chip and  
482 concluded that it increases the cost-effectiveness of genomic selection for French  
483 sheep breeds. Prior to implementing GS, some aspects to consider include product  
484 management and marketing issues that may accompany improved productivity and the  
485 prevailing socio-economic status of farmers, and flock structures and dynamics within  
486 the smallholder system. However cost can be reduced by sharing facilities such as  
487 databases or analytic platforms which may already be in existence for other livestock  
488 species such as cattle. The availability of the LD chip for sheep increases the prospect  
489 of long term genomic selection in small ruminants in developing countries.

490

## 491 **Conclusions**

492 Unique genotypes of several goat and sheep breeds found in developing countries and  
493 especially in Africa present a good opportunity for understanding genetic diversity,

494 structure and adaptation. The availability of molecular tools and approaches have  
495 enabled the understanding of the genetic basis for this diversity and adaptation, initially  
496 through the use of microsatellites and more recently SNP genotype and full genome  
497 sequence data. This information is foundational in terms of its incorporation in future  
498 breeding programs for SR in developing countries. In the long term, the use of  
499 gene/haplotype editing and other emerging breeding strategies could play a role in  
500 incorporating these into breeding programs for increased productivity.

501

502 The basic building blocks for conventional breeding are lacking in most of the small  
503 holder systems in developing countries apart from a few of the established breeds with  
504 some degree of supporting infrastructure. Genotypic data offers quick wins in terms of  
505 parentage verification, breed composition determination (admixture) and genetic  
506 evaluation using the **G** matrix.

507

508 Genomic selection in SR in developing countries will not be a scenario of “one-size fits  
509 all” but it will depend on the type of production system. Classic GS is feasible in breeds  
510 with some degree of conventional genetic improvement already in place. The CBBP  
511 provides a good framework for the implementation of GS in small holder systems and  
512 Innovative GS will be needed in fragile growth systems where adaptation is an  
513 important trait. Identifying regions of the genome associated with various aspects of  
514 adaptability and maximizing diversity of adaptation in animals reared will be essential.  
515 Adequate cost-benefit analysis should be part of any strategy adopted in implementing  
516 GS in these production systems.

517

518 **References**

519 Anderson R, McEwan J, Brauning R, Kijas J, Dalrymple J, Worley K, Daetwyler H, Van

520 Stijn T, Clarke S, Baird H and Khan A 2014. Development of a high density (600K)

521 Illumina ovine SNP chip and its use to fine map the yellow fat locus. Retrieved on

522 28, March, 2017 from:

523 <https://pag.confex.com/pag/xxii/webprogram/Paper10725.html>

524 Amer PR, Mpofu N, Bondoc O 1998. Definition of breeding objectives for sustainable

525 production systems. In Proceedings of the Sixth World Congress on Genetics

526 Applied to Livestock Production, 11–16 January 1998, Armidale, NSW,

527 Australia, 28, pp.97–104.

528 Auvray B, McEwan J, Newman SA, Lee M, Dodds K 2014. Genomic prediction of

529 breeding values in the New Zealand sheep industry using a 50K SNP chip.

530 Journal of Animal science 92, 4375-4389.

531 Barillet F, Astruc JM, Baloche G, Buisson D, Lagriffoul G 2014. Genomic selection in

532 French dairy sheep: main results and design to implement genomic breeding schemes.

533 39th ICAR meeting, Session S2 Dairy Sheep and Goats.

534 23 May 2014. Berlin, Germany

535 Beef and Lamb New Zealand, 2012. Domestic Trends and Measuring Progress against

536 the Red Meat Sector Strategy. Presentation to the Red Meat Sector Conference

537 16 July, 2012. Queestown. Retrieved on 28 March 2017 from

538 [http://www.mia.co.nz/docs/mia\\_conference/2012/Rob%20Davidson.pdf](http://www.mia.co.nz/docs/mia_conference/2012/Rob%20Davidson.pdf)



539 Benjelloun B, Alberto FJ, Streeter I, Boyer F, Coissac E, Stucki S, BenBati M,  
540 Ibelbachyr M, Chentouf M, Bechchari A, Leempoel K, Alberti A, Engelen S,  
541 Chikhi A, Clarke L, Flicek P, Joost S, Taberlet P, Pompanon F, NetGen  
542 Consortium 2015. Characterizing neutral genomic diversity and selection  
543 signatures in indigenous populations of Moroccan goats (*Capra hircus*) using  
544 WGS data. *Frontiers in Genetics* 6, 107. doi: 10.3389/fgene.2015.00107.

545 Bett CR, 2005. Developing breeding strategies for the Kenya dual purpose goat. *Animal*  
546 *Sciences*, Egerton University: Egerton University.

547 Brown A, Ojango J, Gibson J, Coffey M, Okeyo M, Mrode R 2016 Genomic  
548 selection in a crossbred cattle population using data from the Dairy Genetics  
549 Project for East Africa. *Journal of Dairy Science*, 99:7308-7312

550 Campbell, Q 2003. The origin and description of southern Africa's indigenous goats.  
551 South Africa. *Journal of Animal Science* 33, 18–22.

552 Carillier C, Larroque H and Robert-Granié C 2014. Comparison of joint versus  
553 purebred genomic evaluation in the French multi-breed dairy goat population.  
554 *Genetics Selection Evolution* 46, 67. doi:10.1186/s12711-014-0067-3.

555 Casey NH and Webb EC 2010. Managing goat production for meat quality. *Small*  
556 *Ruminant Research* 89, 218–224.

557 Castro EA, Lopez IMR and Lim A 2006. Characterization of a new SNP in the growth  
558 and differentiation factor 9 (GDF- 9) gene, specific for the Brazilian Santa Inês  
559 sheep. *Proceedings of the 8th World Congress on Genetics Applied to Livestock,*  
560 *Production*, 13-18 August 2006, Belo Horizontal, Brazil, pp. 22-25.

561 Davis GH 2005. Major genes affecting ovulation rate in sheep. *Genetics Selection*

562 Evolution 37, (Suppl. 1) S11-S23. doi:10.1051/gse:2004026.

563 Devendra C 2002. Potential productivity from small ruminants and contribution to  
564 improved livelihoods in developing countries. In: Batista, A.M.V., Barbosa, S.B.P.,  
565 do Santos, M.V.F., Ferrira, L.M.C. (Eds.), Proceedings of the Thirty Ninth  
566 Reuniao Anual, Sociedade Brasilia de Zootecnia. 29 July–1 August 2002,  
567 Recife, Brazil, Secretaria Executiva, Sociedade Brasileira de Zootecnia, Brasilia,  
568 Brazil, pp. 246–269.

569 Daetwyler HD 2014. Using Genomics to Improve Reproduction Traits in Sheep. 10th  
570 World Congress on Genetics Applied to Livestock Production. Asas.

571 Daetwyler HD, Pong-Wong, R, Villanueva, B, Woolliams, JA 2010 The impact of  
572 genetic architecture on genome-wide evaluation methods. Genetics 185, 1021-  
573 1031.

574 Daetwyler HD, Swan AA, van der Werf JH and Hayes BJ 2012. Accuracy of  
575 pedigree and genomic predictions of carcass and novel meat quality traits in  
576 multi-breed sheep data assessed by cross-validation. Genetics Selection,  
577 Evolution, 44:33.

578 Demars J, Fabre S, Sarry J, Rossetti R, Gilbert R, Persani L, Tosser-Klopp G, Mulsant  
579 P, Nowak Z, Drobik W, Martyniuk E, Bodin L 2013. Genome-wide association  
580 studies identify two novel BMP15 mutations responsible for an atypical  
581 hyperprolificacy phenotype in sheep. PLoS Genetics 9, e1003482.

582 FAOSTAT, 2013. FAO statistical yearbook 2913. World food and agriculture.  
583 <http://www.fao.org/docrep/018/i3107e/i3107e.PDF>

584 Fariello M-I, Servin B, Tosser-Klopp G, Rupp R, Moreno C, ISGC, San Cristobal M,  
585 Boitard S. 2014. Selection signatures in Worldwide sheep populations. PLoS  
586 ONE 9(8), e103813.

587 Getinet M 2016. Molecular characterization of Ethiopian indigenous goat populations:  
588 genetic diversity, demographic dynamics and kisspeptin gene polymorphism.  
589 PhD dissertation. Addis Ababa Ethiopia.

590 Getinet M, Kassahun T, Tadelle D, Mwai O, Djikeng A, Osama S, Alayu K and,  
591 Solomon A 2016. Analysis of Kisspeptin (KISS1) gene and its association with  
592 litter size in goats. Accepted: African journal of Biotechnology.

593 Gholizadeh M, Rahimi-Mianji G, Nejati-Javaremi A 2015. Genomewide association  
594 study of body weight traits in Baluchi sheep. Journal of Genetics 94, 143-146.

595 Gouveia JJS, Paiva SR, McManus CM, Caetano AR, Kijas JW, Faco O, Azevedo HC,  
596 Araujo AM, Souza CJH, Yamagishi MEB, Carneiro PLS, Lobo RNB, Oliveira  
597 SMP, Silva MVGB 2017. Genome-wide search for signatures of selection in three  
598 major Brazilian locally adapted sheep breeds. Livestock Science 197, 36-45.

599 Haile A, Dessie T, Rischkowsky B 2014. Performance of indigenous sheep breeds  
600 managed under community based breeding programs in the highlands of  
601 Ethiopia: Preliminary results. Addis Ababa: ICARDA.  
602 <http://hdl.handle.net/10568/35466> .

603 Jenko J, Gorjanc G, Cleveland MA, Varshney KR, Whitelaw BA, Woolliams JA and  
604 Hickey JM 2015. Potential of promotion of alleles by genome editing to improve  
605 quantitative traits in livestock breeding programs. Genetics Selection  
606 Evolution, 47:55, DOI 10.1186/s12711-015-0135-3

607 Kijas JW, Lenstra JA, Hayes B, Boitard S, Porto Neto LR, San Cristobal M 2012.  
608         Genome-wide analysis of the world's sheep breeds reveals high levels of  
609         historic mixture and strong recent selection. PLoS biology 10.

610 Kim E-S, Elbeltagy AR, Aboul-Naga AM, Rischkowsky B, Sayre B, Mwacharo JM,  
611         Rothschild MF 2016. Multiple genomic signatures of selection in goats and sheep  
612         indigenous to a hot arid environment. Heredity 116, 255-264

613 König S, Simianer H and Willam A 2008. Economic evaluation of genomic breeding  
614         programs. Journal Dairy Science 92, 382–391

615 Kosgey, IS and Okeyo, AM 2007. Genetic improvement of small ruminants in low  
616         input, smallholder Technical and infrastructural issues. Small Ruminant Research  
617         70,76–88

618 Larroque H, Chassier M, Saintilan R, Astruc JM 2017. Imputation accuracy from a low  
619         density SNP panel in 5 dairy sheep breeds in France. In: Book of Abstracts of the  
620         68th Annual Meeting of the European Federation of Animal Science (p. 151-151).

621 Lôboa, R.N.B., Facóá, O, Lôbob, AM.B.O and Villela, L.C V. 2010. Brazilian goat  
622         breeding programs. Small Ruminant Research 89, 149–154Mdladla K 2016.  
623         Landscape genomic approach to investigate genetic adaptation in South African  
624         indigenous goat populations. PhD Thesis, University of KwaZulu Natal,  
625         Pietermaritzburg, South Africa, 220 pages.

626 Meuwissen TH, Hayes BJ, Goddard ME 2001. Prediction of total genetic value using  
627         genome-wide dense marker maps. Genetics 157, 1819-1829.

628 Mrode R, Han J, Mwacharo JA and De Koning DJ 2016. Novel tools to inform animal  
629 breeding programs. International Livestock Research Institute brief no 14.  
630 <https://livestockfish.cgiar.org/2017/01/27/lfbrief14/>

631 Mucha S, Mrode R, MacLaren-Lee I, Coffey M, and Conington J 2015. Estimation of  
632 genomic breeding values for milk yield in UK dairy goats. *Journal Dairy  
633 Science*, 98(11):8201–8208.

634 Mueller JP 1991. Transferencia de tecnología a pequeños productores de caprinos en  
635 la Argentina. II Reunión de la Red de Rumiantes Menores. Santiago de Chile, 5  
636 al 8 de noviembre. Comunicación Técnica INTA Bariloche Nro. PA 184.

637 Mwandotto BAJ, Ruvuna F, Taylor JF, Cartwright TC 1992. Breeding strategies for  
638 genetic improvement. In *On-farm research and technology for dual-purpose  
639 goats* (Semenye PP and Hutchcroft Eds). Small Ruminant Collaborative  
640 Research Support Program, Kenya. National Printing Press Ltd. Kisumu Kenya.

641 Nimbka C 1999. A village goat cross-breeding project in Maharashtra, India. Workshop  
642 for developing breeding strategies for lower input Animal Production  
643 environments. Bella, Italy, September, 1999. ICAR Technical Series 3, 435 –443

644 Nimbkar C, Ghalsasi PM, Walkden-Brown SW and Kahn LP 2002. Breeding program  
645 for the genetic improvement of the Deccani sheep of Maharashtra, India. 7th  
646 World Congress on Genetics Applied to Livestock Production, August 19-23,  
647 2002, Montpellier, France. Session 25. Developing sustainable breeding  
648 strategies in medium- to low-input systems Communication N° 25-11

649 Ojango JMK, Okeyo AM, Rege JEO 2010. The Kenya dual purpose goat development  
650 project. Animal Genetics Training Resource (AGTR), An ILRI/SLU Project.

651 [http://agtr.ilri.cgiar.org/agtrweb/index.php?option=com\\_content&view=article&id](http://agtr.ilri.cgiar.org/agtrweb/index.php?option=com_content&view=article&id)  
652 [203&Itemid=240](http://agtr.ilri.cgiar.org/agtrweb/index.php?option=com_content&view=article&id).

653 Onim JFM (1992). Dual-purpose goat research in western Kenya. In Kategile JA and  
654 Mubi S (eds). Future of Livestock industries in East and southern Africa.  
655 Proceedings of a workshop held at Kadoma Ranch Hotel, Zimbabwe, 20-23 July  
656 1992. International Livestock Centre for Africa. Addis Ababa, Ethiopia  
657 229pp.

658

659 Pannier L, Pethick DW, Geesink GH, Ball AJ, Jacob RH, Gardner GE 2014.  
660 Intramuscular fat in the longissimus muscle is reduced in lambs from sires  
661 selected for leanness. *Meat science* 96, 1068-1075.

662 Phua S, Hyndman D, Baird H, Auvray B, McEwan J, Lee M, Dodds K 2014.  
663 Towards genomic selection for facial eczema disease tolerance in the New  
664 Zealand sheep industry. *Animal Genetics* 45, 559-564.

665 Pickering NK, Oddy VH, Basarab J, Cammack K, Hayes B, Hegarty RS, Lassen J,  
666 McEwan JC, Miller S, Pinares-Patiño, CS and Haas de Y 2015. Invited review:  
667 Genetic possibilities to reduce enteric methane emissions from  
668 ruminants. *Animal* 9,1431–1440.

669 Rupp R, Mucha S, Larroque H, McEwan J and Conington J 2016. Genomic  
670 application in sheep and goat breeding. *Animal Frontiers* 6, 1:39 – 44.

671 Semenye PP. Onim JFM, Conelly WT, Fitzhugh HA 1989. On-farm evaluation of dual-  
672 purpose goat production systems in Kenya. *Journal of Animal Science* 67, 3096-3102.

673

674 Shumbusho F, Raoul J, Astruc JM, Palhiere I, Lemarié S, Fugeray-Scarbel A, Elsen  
675 JM 2016. Economic evaluation of genomic selection in small ruminants: a sheep meat  
676 breeding program. *Animal* 10, 1033-1041.

677 Smith JW, Tarawali S, Grace D and Sones K 2013. Feeding the world in 2050: Trade-  
678 offs, synergies and tough choices for the livestock sector. *Tropical Grasslands -*  
679 *Forrajes Tropicales* 1, 125-136.

680 Snyman MA, Olivier JJ 1999. Repeatability and heritability of objective and subjective  
681 fleece traits and body weight in South African Angora goats. *Small Ruminant*  
682 *Research*, 34 103-109.

683 Tosser-Klopp G, Bardou P, Bouchez O, Cabau C, Crooijmans R, Dong Y, Donnadiou-  
684 Tonon C, Eggen A, Heuven HC, Jamli S, Jiken AJ, Klopp C, Lawley CT, McEwan J,  
685 Martin P, Moreno CR, Mulsant P, Nabihoudine I, Pailhoux E, Palhiere I, Rupp R,  
686 Sarry J, Sayre BL, Tircazes A, Jun W, Wang  
687 W, Zhang W 2014. Design and characterization of a 52K SNP chip for goats.  
688 *PloS one* 9, e86227.

689 Visser C, Van Marle Köster E 2014. Strategies for the Genetic Improvement of South  
690 African Angora goats. *Small Ruminant Research*, 121: 89–95.

691 Visser C, Van Marle-Köster E, Friedrich H 2011a. Parentage verification of South  
692 African Angora goats, using microsatellite markers. *South Africa Journal of*  
693 *Animal Science*, 41(3): 250-255.

694 Visser C, Crooijmans RPMA, Bovenhuis H, Van Marle-Köster E 2011b. QTL for  
695 mohair traits in South African Angora goats. *Small Ruminant Research* 100,  
696 8-14.

697 Walberg L (2011). Milk production in dairy cows and goats – a case study in the Nyando  
698 district in South-Western Kenya. Degree Project, Swedish University of Agricultural  
699 Sciences (SLU), 29pp.  
700

701 Walkden-Brown, S.W., J.H.J. van der Werf, C. Nimbkar, V.S. Gupta (eds). 2009. Use of  
702 the *FecB* (Booroola) gene in sheep-breeding programs. Proc. Helen Newton  
703 Turner Memorial International Workshop Pune, Maharashtra, India, 10–12  
704 November 2008. ACIAR Proceedings No. 133. Australian Centre for International  
705 Agricultural Research: Canberra.

706 Woldu T, Markemann, A, Reiber, C, Mutha, PC, Zárate, AV 2016. Optimizing  
707 contributions of goat farming to household economic success and food security  
708 in three production systems in Ethiopia. Journal of Agriculture and Rural  
709 Development in the Tropics and Subtropics 117, 73–85.

710 Wurzinger M, Sölkner J, Iñiguez L 2011. Important aspects and limitations in  
711 considering community-based breeding programs for low-input smallholder  
712 livestock systems. Small Ruminant Research 98, 170-175.