

1 **Extrusion 3D printing of paracetamol tablets from a single formulation with**  
2 **tunable release profiles through control of tablet geometry**

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28 **Abstract**

29 An extrusion based 3D printer was used to fabricate paracetamol tablets with different geometries  
30 (mesh, ring, and solid) from a single paste-based formulation formed from standard pharmaceutical  
31 ingredients. The tablets demonstrate that tunable drug release profiles can be achieved from this single  
32 formulation even with high drug loading (>80% w/w). The tablets were evaluated for drug release using  
33 a USP dissolution testing type I apparatus. The tablets showed well-defined release profiles (from  
34 immediate to sustained release) controlled by their different geometries. The dissolution results showed  
35 dependency of drug release on the surface area/volume (SA/V) ratio and the SA of the different tablets.  
36 The tablets with larger SA/V ratios and SA had faster drug release. The 3D printed tablets were also  
37 evaluated for physical and mechanical properties including tablet dimension, drug content, weight  
38 variation, breaking force and were within acceptable range as defined by the international standards  
39 stated in the United States Pharmacopoeia. X-Ray Powder Diffraction, Differential Scanning  
40 Calorimetry, and Attenuated Total Reflectance Fourier Transform Infrared Spectroscopy were used to  
41 identify the physical form of the active and to assess possible drug-excipient interactions. These data  
42 again showed that the tablets meet USP requirement. These results clearly demonstrate the potential of  
43 3D printing to create unique pharmaceutical manufacturing, and potentially clinical, opportunities. The  
44 ability to use a single unmodified formulation to achieve defined release profiles could allow, for  
45 example, relatively straightforward personalization of medicines for individuals with different  
46 metabolism rates for certain drugs and hence could offer significant development and clinical  
47 opportunities.

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54 **1. Introduction**

55 Personalised medicine is defined as a customization of health care to individual patients through linking  
56 diagnostics and treatments with genetic testing and emerging technologies such as proteomics and  
57 metabolomics analysis (1). The main advantages of this approach, are to increase the effectiveness of  
58 the prescribed treatment regimen and to minimize their adverse effects such as those linked to  
59 overdosing of drugs with a narrow therapeutic index such as digoxin and anti-clotting agents (2). In the  
60 context of solid oral dosage forms, conventional large-scale tableting manufacturing methods are  
61 clearly unsuited to personalised medicine and in addition, provide restrictions on the complexity  
62 achievable in the dosage form in terms of, for example, tablet geometry, drug dosage, distribution and  
63 combinations. 3D printing offers the potential for the manufacture of bespoke solid oral dosage forms.  
64 3D printers also offer the possibility of reducing the number of manufacturing steps as currently used  
65 in traditional tablet production process, such as powder milling, wet granulation, dry granulation, tablet  
66 compression, and coating and the potential for rapid formulation development with limited quantities  
67 of active ingredients as available in early drug development (3, 4)

68 3D printing is hence a potentially significant platform that can produce viable solid dosage forms in  
69 complex geometries in a programmed, controlled manner and with accurate drug loading (5-8). Many  
70 believe, that 3D printers could play an important role in the development of personalised unit dose  
71 medication for targeting the specific needs of individual patients and treatments (5, 6, 9). In envisaging  
72 how such an approach could be taken to the practical manufacture of dosage forms it would clearly  
73 simplify matters greatly if the formulation (or 'ink' in 3D printer terms) could be kept as simple as  
74 possible, with little need for the use of multiple formulations that must be mixed precisely *in situ* within  
75 the 3D printer. Such a complex mixing approach would greatly complicate supply chains, increase  
76 quality control difficulties and subsequently raise regulatory barriers even higher than might be  
77 expected for such a new approach to manufacture. We propose, and demonstrate here, that the required  
78 need for personalization in terms of drug release profile can be achieved by the control of tablet  
79 geometry alone from a single formulation. Such an approach, we propose would significantly increase

80 the likelihood of 3D printing being adopted for the development and manufacture of personalised  
81 dosage forms.

82 Paracetamol is commercially available in many different dosage forms including; tablets, capsules,  
83 suspensions, suppositories, and intravenous solutions and is commonly used to treat mild to moderate  
84 pain caused by headaches, toothache, sprain, or strains (4). Here, paracetamol was chosen as a well-  
85 known freely available drug suitable for a proof of concept study. The common paracetamol doses  
86 available range from 300 to 500 mg, although 1000 mg is also available in some regions. Therefore,  
87 customizing of paracetamol effect/release (plasma peak levels) while prolonging its action by using  
88 different tablet geometries is potentially desirable (10). The effect of dosage form geometry on drug  
89 release for controlled release has been reported (10-12). Previously work has also been done on 3D  
90 printing of paracetamol formulations primarily using Fused Deposition Modelling (FDM) 3D printing  
91 (4, 13-18). However, the high extrusion temperature used in FDM ( $\geq 120$  °C) narrows the potential  
92 active ingredient library to include only heat stable actives (4). Other possible 3D printing methods like  
93 Stereolithography (SLA) and ink-jet printing currently use excipients that are not generally recognized  
94 as safe (GRAS) (13).

95 Different types of 3D printer are commercially available including the aforementioned FDM, Inkjet,  
96 Selective Laser Sintering (SLS) and SLA, and significant work has been done in the area of drug  
97 delivery using these approaches (7, 12-14, 19-23). Published research regarding 3D printing techniques  
98 to achieve controlled drug release include; Sadia and co-workers, who created multi-channelled tablets  
99 using FDM for a Biopharmaceutics Classification System (BCS) class IV drug, hydrochlorothiazide  
100 (24). Also Yang *et al.* used FDM to print tablets with differing internal scaffold structures to control  
101 ibuprofen release (25). SLS has been used by Fina *et al.* to create orally-disintegrating paracetamol  
102 tablets whose drug release depending upon the printing speed (17). We have also previously  
103 demonstrated the flexibility afforded by 3D extruding semi-solid formulations at ambient conditions  
104 using compendia grades available to form tablets to achieve controlled drug release (5, 6, 26). Whilst  
105 extrusion-based 3D printing avoids the heat stress associated with other techniques it has some  
106 disadvantages including; relatively low spatial resolution compared to other 3D printing approaches,

107 and that it may not be suitable for water-sensitive materials (degradation unless solvent or binder other  
108 than water is used). In this research the drying temperature was set at 80 °C to accelerate the drying time  
109 of the printed tablets (4). However, lower temperatures in a range of 40-60 °C can be employed, as is  
110 commonly used in drying oral solid dosage form but this leads to longer drying times. The aim of this  
111 work is to introduce extrusion-based 3D printer for the first time as a capable tool to print different  
112 geometries with meaningful drug loading that can be used to define drug release profiles.

## 113 **2. Materials and methods**

### 114 *2.1. Materials*

115 Paracetamol, and polyvinylpyrrolidone (PVP K25) were supplied by Sigma–Aldrich (Gillingham, UK).  
116 Croscarmellose sodium (NaCCS) (Primellose®) was kindly supplied as a gift from DFE Pharma. Starch  
117 was kindly supplied by Colorcon®. Milli-Q water (resistivity 18.2 MΩ cm) was used for all  
118 formulations and solutions. All other reagents were of either HPLC or analytical grade.

### 119 *2.2. Methods*

#### 120 *2.2.1. Design of paracetamol tablets*

121 A strategy of controlling the geometry to be generally oval shaped (easy to swallow) for the 3D printed  
122 tablets was chosen (Fig. 1). A tablets normal solid tablet geometry was altered to also produce an oval  
123 ring and an oval tablet with an internal mesh or lattice-like structure. The mesh tablets which were  
124 printed in 13 layers in an external oval ring (formed from two or three printed ovals) and an internal  
125 cross-lattice mesh format. There was an internal gap of 0.4mm between the two printed oval walls of  
126 layers 2-12 (Fig.1), with the top (layer 13) and bottom (layer 1) layers having three oval walls printed  
127 around the mesh structure with no gap between them to ensure tablet integrity. The ring tablets was  
128 simply produced by printing oval walls of different dimensions until the ring like structure was  
129 achieved. The outer dimensions of the designed oval tablets was 15 mm length × 8 mm width × 3.2 mm  
130 height for the solid tablets, 4.8 mm height for the ring tablets, and 5.2 mm height for the mesh tablets.  
131 The geometry of the tablets was designed using a 3D drawing package (BioCAD, regenHU Villaz-St-  
132 Pierre, Switzerland) with the aim of keeping the tablet weight constant across the three geometries.

133 2.2.2. 3D printing process of paracetamol tablets

134 Twelve grams of ground paracetamol and the required excipient powders (starch, PVP K25, and  
135 NaCCS) were mixed using a mortar and pestle for 10 min. 4.5 ml of Milli-Q water (resistivity 18.2 MΩ  
136 cm) was added and the powder was mixed to form a paste according to the formulae shown in Table I.

137 2.2.3. Extrusion based 3D printing process

138 A plastic 20 cm<sup>3</sup> syringe (Optimum® syringe barrels, Nordson EFD) was used to fill the paste into the  
139 syringe cartridge in the 3D printer (regenHU 3D). A stopper was fixed into Luer-Lock thread at the top  
140 end of the barrel after the filling process to avoid any unintentional leakage of paste from the cartridge  
141 showed in figure 2. Once ready for printing, the stopper was removed, and the required nozzle  
142 (Optimum® SmoothFlow™ tapered dispensing tips, 0.6 mm internal diameter (ID) Nordson EFD)  
143 installed. The filled cartridge was then installed into the printer head and the paste was extruded layer  
144 by layer until the desired tablet dimension was reached (Fig. 2). The 3D printed tablets were left on a  
145 heated printing platform (80 °C) overnight for complete drying. The tablets were stored in a sealed  
146 desiccator stored in a cool and dry location. The following printing parameters were used; tip diameter  
147 0.6 mm, printing speed = 6 mm/sec, printing pressure = 1.8 bar, number of printed layers = 13 for mesh  
148 tablets, 12 for ring tablets, and 8 for solid tablets. The tablet outer dimensions were kept the same but  
149 the geometries were varied using functions in BioCAD software. The tablet weights were kept constant  
150 within a measured range of  $308.01 \pm 4.52$  mg by adjusting the printed tablet height.

151 2.2.4. Dissolution studies

152 *In vitro* drug release studies of the paracetamol 3D printed tablets were performed using a USP Type I  
153 apparatus (rotation speed at 30 rpm, 900 ml phosphate buffer, pH 6.8 as the dissolution media at  $37 \pm$   
154  $0.5$  °C). 5.0 ml samples were withdrawn at 5, 10, 15, 30, 60, 120, 240, 360, 480, 600, and 720 min. The  
155 samples were centrifuged and 0.5 ml from the supernatant was drawn and diluted to 10 ml using the  
156 dissolution medium. The samples were analysed with a UV–vis spectrophotometer (Cary® 50 UV-vis  
157 spectrophotometer) at a  $\lambda$  max of 243 nm. Drug dissolution studies were conducted in sextuplicate and  
158 the average of percentage of cumulative drug release as a function of time was determined. Although  
159 the USP monograph specifications for paracetamol tablets dissolution testing state that the dissolution

160 rotation should be 50 rpm, a speed of 30 rpm was chosen to ensure that the tablet disintegration occurred  
161 mainly due to the effect of disintegrants rather than effects caused by basket rotation.

## 162 *2.3. Characterization techniques*

### 163 *2.3.1. X-Ray Powder Diffraction (XRPD)*

164 The XRPD patterns of pure paracetamol, excipients (PVP K25, NaCCS, and starch) and paracetamol  
165 formulation powder (powder mixture after tablet ground into powder) were obtained at room  
166 temperature using an X'Pert PRO (PANalytical, Almelo, Netherlands) setup in reflection mode using  
167 Cu K $\alpha$ 1 ( $\lambda = 1.54 \text{ \AA}$ ) operating in Bragg–Brentano geometry. The generator voltage was set to 40  
168 kV and the current to 40 mA and the samples were scanned over  $2\theta$  range of  $5^\circ$  until  $30^\circ$  in a step size  
169 of  $0.026^\circ$ .

### 170 *2.3.2. Attenuated Total Reflectance Fourier Transform Infrared Spectroscopy (ATR-FTIR)*

171 Infrared spectra of pure paracetamol, excipients powders (PVP K25, NaCCS, and starch) and  
172 paracetamol formulation powder (powder mixture after tablet ground into powder) were obtained using  
173 an ATR-FTIR (Agilent Cary 630 FTIR) spectrometer.

### 174 *2.3.3. Differential Scanning Calorimetry (DSC)*

175 The DSC measurements were performed on a TA Instruments' DSC Q2000 coupled to Universal  
176 Analysis 2000 with a thermal analyser. DSC analysis on such drug-excipient mixtures were obtained  
177 by grinding paracetamol tablets and sieving the powders ( $<150 \mu\text{m}$ ). Accurately weighed samples of 3-  
178 5 mg were placed and sealed in aluminium pans. The scans were performed under nitrogen flow (50  
179 mL/min) at a heating rate of  $10^\circ \text{C/min}$  from  $35^\circ \text{C}$  to  $200^\circ \text{C}$ . An empty sealed aluminium pan was  
180 used as a reference.

## 181 *2.4. Physical properties of paracetamol immediate release 3D printed tablets*

### 182 *2.4.1. Dimension of paracetamol 3D printed tablets*

183 To confirm the tablet size reproducibility, six tablets from each geometry were measured using Vernier  
184 callipers and their average values calculated.

185 2.4.2. *Weight variation and drug content in the final tablet*

186 Six paracetamol 3D printed tablets (from each geometry) were individually weighed and their average  
187 weight calculated. The individual tablet total weight deviation (%) was calculated. Paracetamol content  
188 in the final tablet was measured as follows; from each batch, 10 paracetamol tablets were weighed and  
189 crushed into powder. A quantity of paracetamol formulation powder equivalent to 0.25g of paracetamol  
190 was weighed and transferred into a 1000 ml volumetric flask. 900 ml of dissolution medium was added  
191 to the flask and placed on a stirrer for 4hrs. 5.0 ml of samples were withdrawn and centrifuged. 0.5 ml  
192 from the supernatant was drawn and diluted to 10 ml using the dissolution medium. The samples were  
193 analysed with a UV-vis spectrophotometer (Cary® 50 UV-vis spectrophotometer) at a  $\lambda$  max of 243  
194 nm. Content uniformity studies were conducted in triplicate and the average of percentage of  
195 paracetamol content was determined.

196 2.4.3. *Breaking force*

197 Six paracetamol 3D printed tablets (from each geometry) were randomly selected and tested for  
198 breaking force using a hardness tester (Hardness tester C50, I Holland Ltd., Holland). The breaking  
199 force values were recorded in N (Newton) units and the tensile strength values were calculated using  
200 equation 1 (27, 28). The tablet breaking force test was done parallel to the longest axis of the  
201 paracetamol tablets.

202  $\sigma_f = 3FL/2bd^2$  Eq. 1

203 Where  $\sigma_f$  is the tensile fracture strength of the tablet, F is the breaking force, L is the tablet length, b is  
204 the tablet width and d is the tablet thickness.

205 2.4.4. *Friability*

206 Ten paracetamol 3D printed tablets (from each geometry) were selected randomly and the tablets were  
207 accurately weighed (initial weight). The tablets were placed in a friability tester and rotated at a constant  
208 speed of 25 rpm for a period of 4 min in Erweka friabilator. The tablets were cleaned of any loose dust  
209 and reweighed (final weight) and the weight loss % (friability) calculated.

210 **3. Results and discussion**



211 *3.1. Tablet printing*

212 Batches of tablets were printed following the method outlined in Figure 2. Examples of printed tablets  
213 are shown in Figure 3.

214 *3.2. In vitro drug dissolution*

215 Dissolution data from the paracetamol tablets (Fig. 4) showed that the different tablets geometries with  
216 different height but similar dimension and total weight and dose (Tables IV and V) gave distinct release  
217 profiles. For the paracetamol mesh tablets, more than 70 % of the drug was released within the first 15  
218 minutes. In contrast, only 25 % and 12 % of the drug was released in the same period from the ring and  
219 the solid paracetamol tablets, respectively. This indicates that the tablet surface area showed an  
220 influence on drug release. Apart from surface area exposed to solution the drug release is also impacted  
221 by the inclusion of the disintegrant, NaCCS, which rapidly absorbs water and swells leading to rapid  
222 disintegration. For the mesh tablets with the increased surface means that water absorption takes place  
223 more rapidly than for the ring and solid tablets (Fig. 4).

224 The drug release from the 3D printed tablets correlates with the SA/V ratios, the higher the SA/V ratio  
225 value, the faster the drug release (Table II). This trend has also been reported by other researches (10,  
226 11, 29). Goyanes *et al.*, showed the effects of SA/V ratios of different geometries on paracetamol release  
227 from tablets prepared by hot melt extrusion (HME) (11). Also in the same study, the authors showed  
228 that the drug release was independent of the surface area (11). Research done by Yi *et al.*, demonstrated  
229 that the drug release from poly lactic-co-glycolic acid/ polycaprolactone/5-Fluorouracil (PLGA/PCL/5-  
230 FU) patches was dependent on the changes of SA produced by geometric modifications (12). The  
231 authors then concluded that the tendency of slowing drug release corresponded to a decrease in the  
232 SA/V ratio (12). Furthermore, Gökçe *et al.*, studied the influence of tablet SA/V ratio of two different  
233 geometries (cylinder and hexagonal) of the lipophilic matrix tablets of metronidazole prepared by  
234 Cutina HR (hydrogenated castor oil) (10). They found that the tablets with the highest release rates for  
235 both geometric shapes reflecting the highest surface area and the lowest SA/V ratio (10). Kyobula *et*  
236 *al.* showed that hot melt 3D inkjet printing can be used to manufacture complex and variable  
237 honeycomb geometry tablets for the controlled loading and release of the drug fenofibrate. In this case

238 the surface area and wettability of the tablet were shown to influence to the observed sustained drug  
239 release profiles (5). Hence, as can reasonably be expected, we can conclude that the tablet geometry  
240 and surface area generally have an effect on drug release behaviour and are parameters that can be  
241 manipulated to control drug release, even in formulations with additives such as a swellable  
242 disintegrant, as here. The higher the SA and SA/V ratio values the faster the drug release is from the  
243 3D printed tablets (Fig. 4 and Table II).

244 The demonstrated ability to use a single unmodified formulation to achieve defined release profiles  
245 presents opportunities to optimize or personalize medicines during formulation development and in  
246 clinical use. For example, relatively straightforward personalization of medicines would be possible for  
247 individuals with different metabolism rates due to their genetic makeup (26) for certain drugs and hence  
248 could address issues where people who metabolize drugs slowly may accumulate a toxic level of a drug  
249 in the body or in others who process a drug quickly and never have high enough drug concentrations to  
250 be effective.

### 251 3.3. Drug release kinetics

252 To further understand the drug release mechanisms displayed by the different geometries, the modes of  
253 release of paracetamol over 12 hours at a buffer pH 6.8 was modelled using Zero order, First order,  
254 Higuchi, and Korsmeyer–Peppas models (30, 31). According to fitted  $r^2$  values, the mesh and ring  
255 tablets were best fitted by the first order equation (i.e., log cumulative percentage of drug remaining is  
256 proportional to the time) (32) and the solid tablets were best fitted by the Higuchi model (i.e., cumulative  
257 percentage drug release versus square root of time) (32) with  $r^2$  values of 0.77, 0.97 and 0.99,  
258 respectively (Table III). The equation reveals  $n$  values (as in Eq. (2)) of 0.25 for mesh tablets, 0.44 for  
259 ring tablets and 0.56 for solid tablets.

$$260 \quad M_t/M_\infty = Kt^n \quad (2)$$

261 Where  $M_t/M_\infty$  is the fraction of drug released at time  $t$ ,  $K$  is the release rate constant and the release  
262 exponent (32, 33).

263 The above results suggest that the drug is released primarily by *Fickian diffusion* through a gel layer  
264 formed by the amylose in the added starch. Amylose is known to absorb water, swell and then form a

265 gel layer (34). The drug release from the mesh tablets was faster than the drug release from the other  
266 geometries (ring and solid). This is, we propose, related to the larger surface area (mesh>ring>solid)  
267 and the more easily disrupted geometry of the mesh tablets where the chance to form a stable gel layer  
268 and hence retard drug release is inhibited. The disintegrants (the amylopectin (insoluble component  
269 found in the starch that can absorb water, swell and act as disintegrant) and NaCCS)) work to weaken  
270 and disrupt the formed gel layer in the mesh tablets. In case of ring and solid tablets the geometry is  
271 more compact with a smaller surface area and less exposure to the dissolution medium than mesh tablets  
272 so the disintegration rate is reduced and there is an increased time to form a gel layer and hence  
273 retardation of drug release (solid>ring>mesh).

#### 274 3.4. XRPD

275 XRPD of the pure paracetamol, excipients (PVP K25, NaCCS, and starch) and paracetamol formulation  
276 powder (powder mixture after tablet ground into powder) was done to investigate any potential changes  
277 in physical form of the active on printing (Fig. 5 and 6). The Bragg peaks observed from the pure  
278 paracetamol (as received) match the Bragg peaks of paracetamol (calculated) reported in the Cambridge  
279 Structural Database (CSD) (Fig. 5).

280 The results in figure 6 show that the paracetamol (non-ground and ground powder) exhibited multiple  
281 sharp Bragg peaks in their XRPD patterns related to their crystalline nature. The post-printing XRPD  
282 data show the same Bragg peaks for the paracetamol. There was, therefore no evidence of a change in  
283 physical form (Form I) for the paracetamol in this formulation fabricated using extrusion based 3D  
284 printing. We believe that a portion of the paracetamol powder could have dissolved after addition a  
285 significant quantity of water (4.5 ml) into total paracetamol dry formulae (12 g) (paracetamol solubility  
286 12.78 g/l/20 °C) (34) as the whole mixture formed a paste, however this must have recrystallized back  
287 into form I if this had occurred. The XRPD data from figure 6 also did not show evidence of  
288 incompatibility between the active and the chosen additives (PVP K25 (10 % w/w), starch (8.33 %  
289 w/w) and NaCCS (0.63 % w/w)) in the 3D printed tablets.

#### 290 3.5. ATR-FTIR

291 Infrared spectral data show that the characteristic peaks positions remained unchanged from the  
292 paracetamol powder to the formulation, indicating that there were no detectable interactions between  
293 paracetamol (81 % w/w) and the chosen excipients (PVP K25 (10 % w/w), starch (8.33 % w/w) and  
294 NaCCS (0.63 % w/w)) in the tablets (Fig. 7).

### 295 3.6. DSC

296 DSC analysis was performed to explore potential incompatibility between the active and added  
297 excipients and the stability of drug crystallinity after the 3D printing process (grinding, mixing, paste  
298 formulation and drying process on a hot plate heated at 80 °C). The DSC data from figure 8 shows that  
299 the pure powder of paracetamol melts at 169.7 °C confirming the presence of form I (4, 35, 36) while  
300 the pure powder of PVP K25 shows a glass transition ( $T_g$ ) around 155 °C (4, 37). The same figure also  
301 shows clear evidence of an endothermic event (melting point) at 169.24 °C from the printed paracetamol  
302 formulation, indicating that the active is still in a crystalline form, specifically form I. From the above  
303 results and discussions, we found that DSC thermogram of paracetamol formulation powder after  
304 grinding, blending, printing, and post-printing processes with the excipients; starch, PVP K25 and  
305 NaCCS did not show significant changes in peak placement apart from the peak depression and  
306 reduction caused by the presence of the polymer in the formulation in comparison to the peak obtained  
307 from the pure paracetamol powder and again suggesting compatibility of the excipients.

### 308 3.7. Physical properties

309 The 3D printed tablets were evaluated for weight variation, content uniformity, breaking force, friability  
310 and tablet dimensions.

#### 311 3.7.1. Tablet's shape and dimension

312 Table IV confirms that the tablet dimensions were reproducible and comparable with the designed  
313 tablet's size and dimension and with the tablet size reported in the literature prepared by conventional  
314 tableting press machines (38-40).

#### 315 3.7.2. Weight variation

316 The paracetamol 3D printed tablets showed an acceptable percentage weight variation (table V) and,  
317 therefore, comply with the USP specification for uncoated tablets ( $\pm 7.5\%$  for average weight of tablets  
318 130 – 324 mg) (41, 42). The paracetamol content in the final tablets was also assessed and found to be  
319 103.2 %  $\pm 1.1$  for the mesh tablets, 104.0 %  $\pm 1.1$  for the ring tablets and 103.1 %  $\pm 1.5$  % for the solid  
320 tablets

### 321 *3.7.3. Breaking force*

322 Table VI shows the 3D printed tablets breaking forces (kg and N), and the tensile fracture strength.  
323 Tensile fracture strength of the paracetamol flat faced oval tablets were calculated (28). In a  
324 conventional tableting press compression forces can be used to control the physical properties of the  
325 final tablet, where a breaking force value of 4kg is the minimum satisfactory measurement (26, 43).  
326 Measured breaking force measurements were within the accepted range of 8.69-9.56 kg for the solid  
327 tablets but failed to reach the minimum satisfactory value for the mesh and ring tablets (table VI). It is  
328 clear that as compression force is not part of 3D printing process that the same opportunity to manipulate  
329 tablet hardness in this way does not exist and rather the formulation composition, solidification/drying  
330 process and the type of printer employed are critical factors. Clearly, further work beyond the scope of  
331 this paper is required in this area, however, from a subjective and qualitative assessment, the ring and  
332 mesh paracetamol 3D printed tablets appear to be quite robust and are able to tolerate a reasonable  
333 amount of rough handling. For example, they could be dropped onto a hard surface from a height of  
334 around 15 cm without observable damage. In addition, such tablets could be considered for manufacture  
335 close to the patient where traditional wear factors such as chipping, capping, and abrasion which  
336 normally occurred during manufacturing, packaging, and shipping processes are not relevant.

### 337 *Friability*

338 This is a USP test used to determine a tablets resistance to abrasion, capping, and chipping occurred  
339 during manufacturing, packaging, and shipping processes. All paracetamol 3D printed tablets of  
340 different geometries showed a satisfactory percentage of weight loss  $\leq 1$  % of the tablet weight (table  
341 VII) and, therefore, the tablets meet USP specifications (44).

342 **4. Conclusions**

343 Extrusion based 3D printing of different paracetamol tablet geometries with a high drug loading (81 %  
344 w/w) was successfully demonstrated. The mesh-geometry 3D printed tablets released more than 70 %  
345 of the active within 15 min achieving immediate release mesh shaped tablets. In contrast, only 25 %  
346 and 12 % of the drug was released in the same period from the ring and the solid paracetamol tablets,  
347 respectively, effectively demonstrating sustained release. Drug release from the tablets showed a clear  
348 dependency on the SA/V ratio. XRPD, FTIR and DSC data show that the paracetamol form was  
349 unaffected by the printing and that there were no detectable interactions between the paracetamol and  
350 the chosen excipients (Starch, PVP K25 and NaCCS). The 3D printed paracetamol tablets were also  
351 evaluated for weight variation, drug content in the final tablets, hardness, friability, and tablet  
352 dimensions and were within acceptable range as defined by the international standards stated in the  
353 USP. This work again validates that the extrusion-based 3D printing process is capable of producing  
354 viable tablets from materials having compendia grades available for pharmaceutical applications. More  
355 importantly this work demonstrates for the first time the application of extrusion-based printing for  
356 tailoring of drug release from a single formulation through control of only tablet geometry the first. We  
357 believe this is a significant step forward in the potential wider take up of 3D printing for the manufacture  
358 of medicines, particular in the areas of clinical development and personalised medicines. With this  
359 principal demonstrated, it becomes possible to envisage control of drug release and dose (through  
360 dosage form size) on an individual basis using a 3D printer, without the need for forming complex  
361 mixtures from different formulation ‘cartridges’. This would greatly simplify potential supply chains  
362 of formulation inks and the quality control of the printed product.

363 **Acknowledgments**

364 We gratefully acknowledge GSK for the funding of this work and Dr Jing Yang for access to the 3D  
365 printer. We also thank DFE Pharma for the complementary supply of NaCCS.

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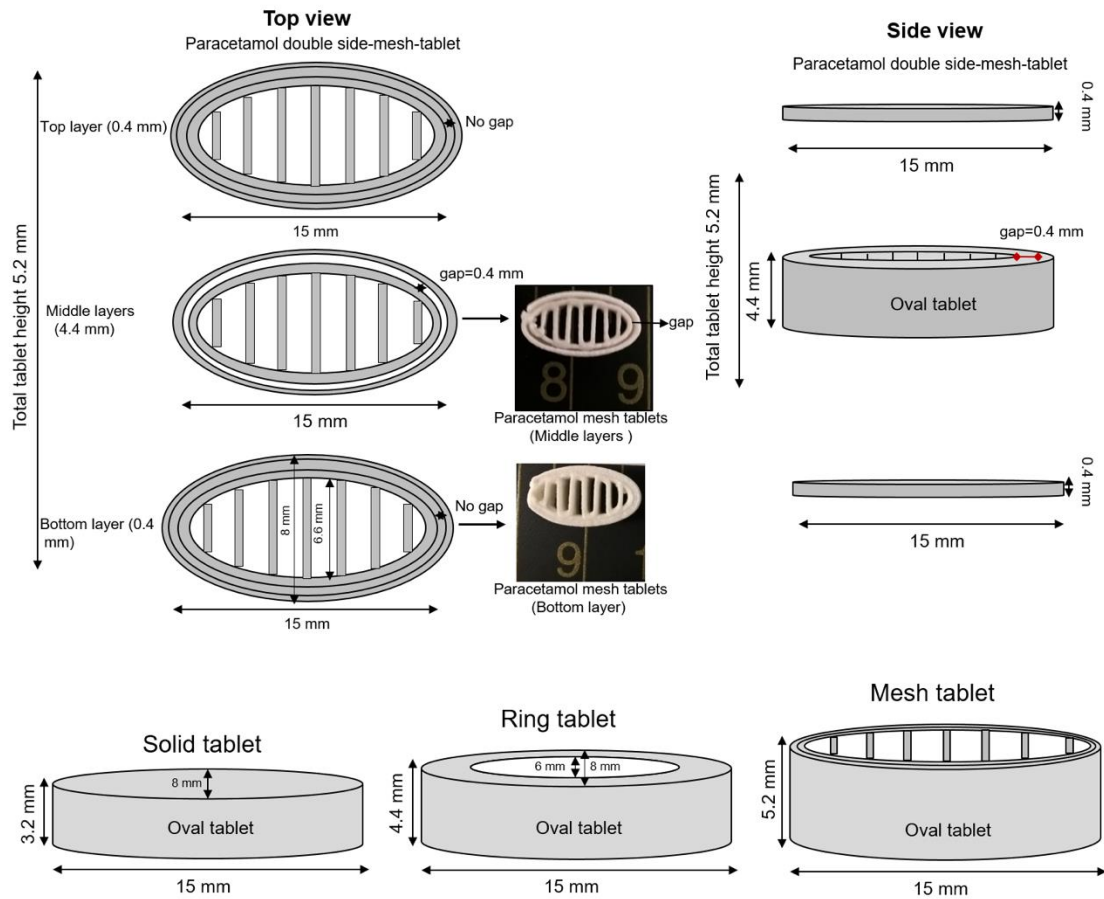
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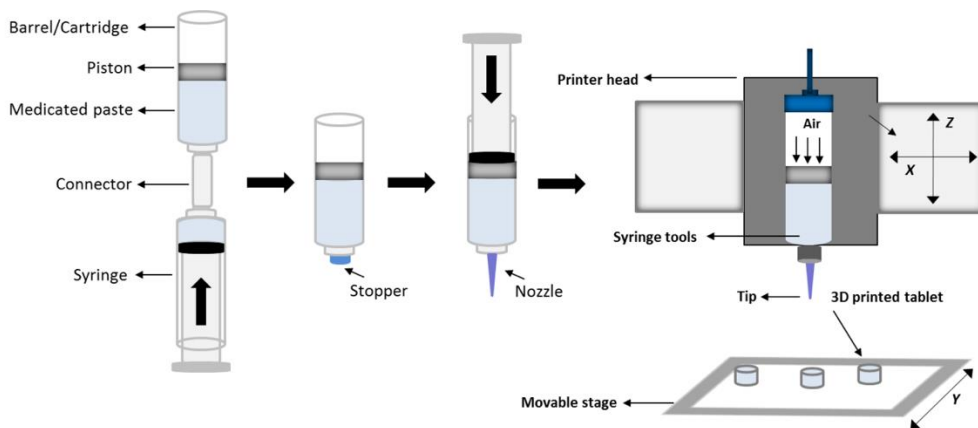
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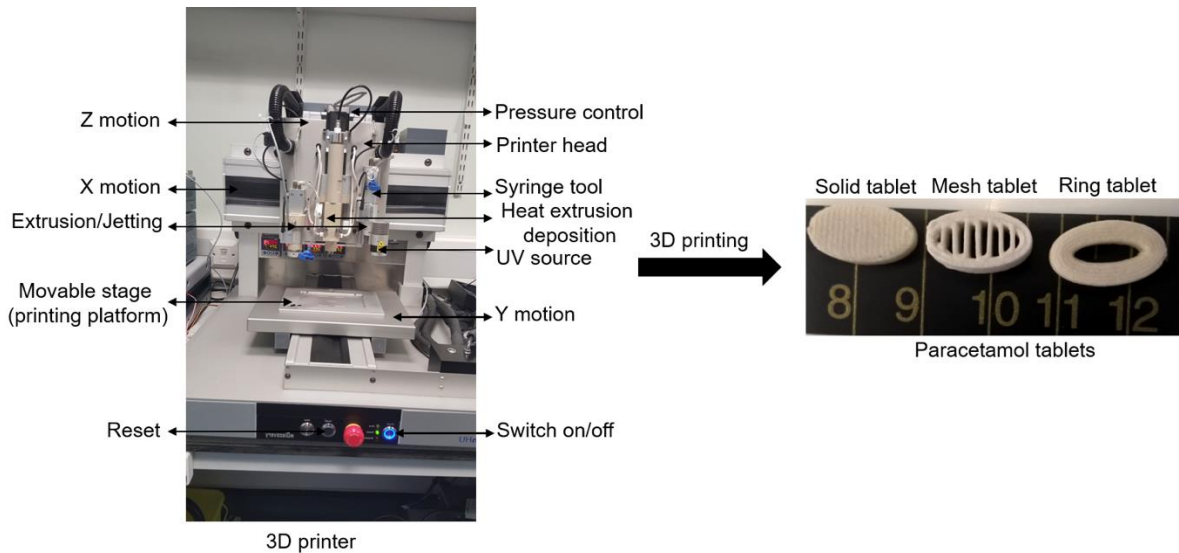
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524 **Fig. 1.** Schematic structural diagram of paracetamol 3D printed tablets with different geometric  
525 shapes; mesh, ring and solid tablets.



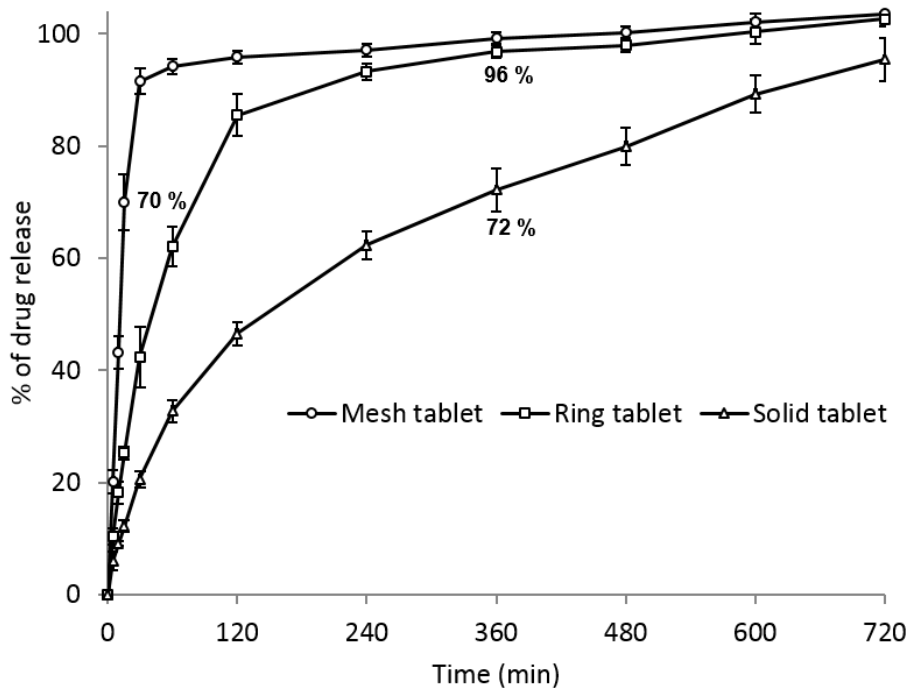
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527 **Fig. 2.** Schematic diagram of cartridge/barrel tool filling process.



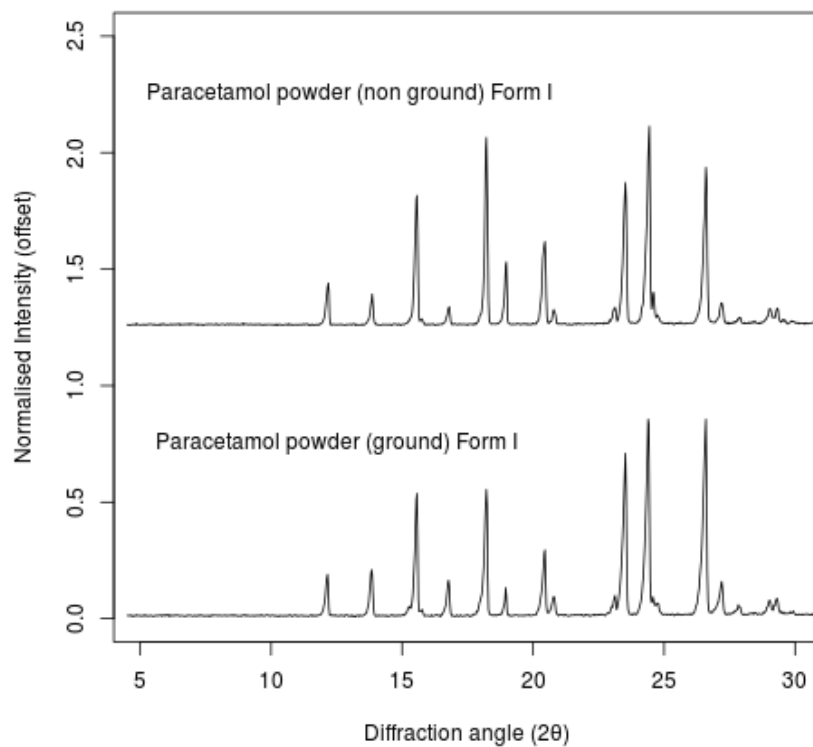
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529 **Fig. 3.** The regenHU 3D printer (left), and image of paracetamol tablets 15.35 mm length × 8.41 mm  
 530 width × 3.44 mm height for solid tablets, and 15.24 mm length × 8.41 mm width × 4.8 mm height for  
 531 ring tablets and 15.22 mm length × 8.48 mm width × 5.46 mm height for mesh tablets (average, n = 6)  
 532 (right).



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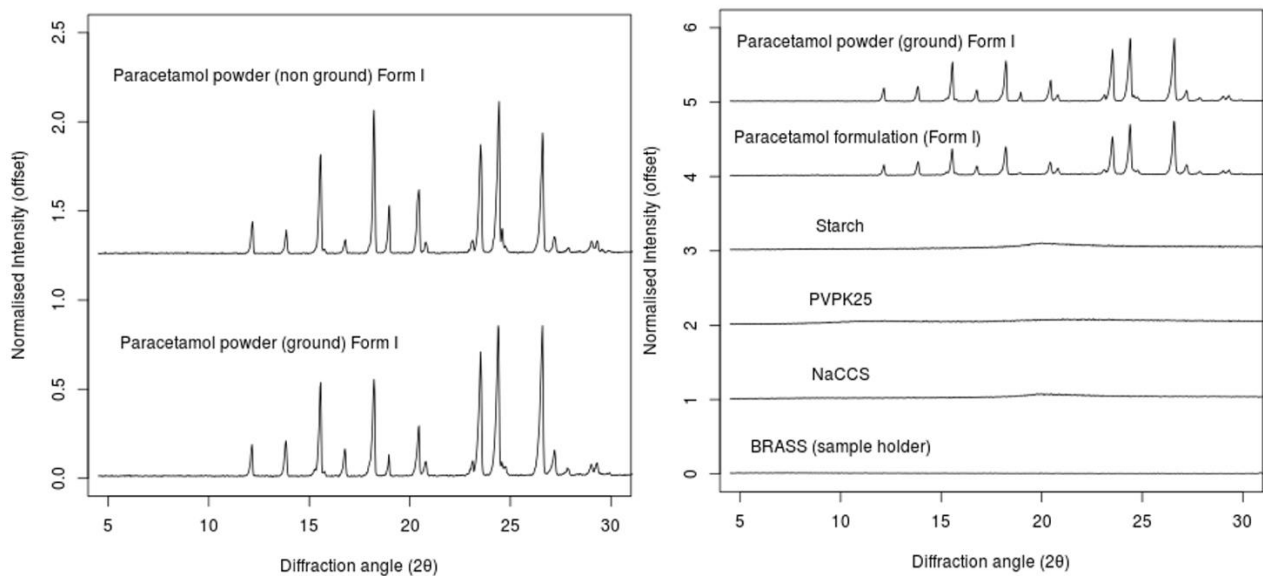
534 **Fig. 4.** In vitro cumulative paracetamol release profiles from three different geometries; mesh, ring  
535 and solid paracetamol tablets,  $n = 6$  (the printed tablets have different height but similar dimension  
536 and total weight and dose).



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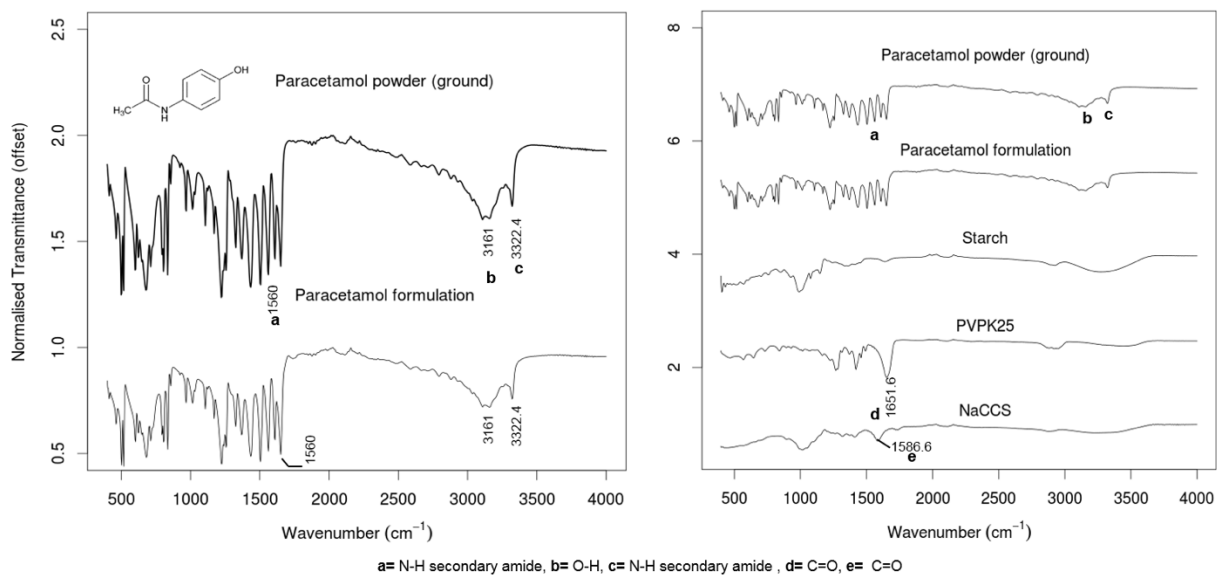
538 **Figure 5.** XRPD patterns of the calculated (top) and reference (measured) paracetamol.

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541 **Figure 6.** XRPD patterns of paracetamol powder (non-ground and ground Form I) (left), paracetamol  
 542 powder (ground Form I), paracetamol formulation, starch, PVP K25, NaCCS and Brass (sample  
 543 holder) (right).



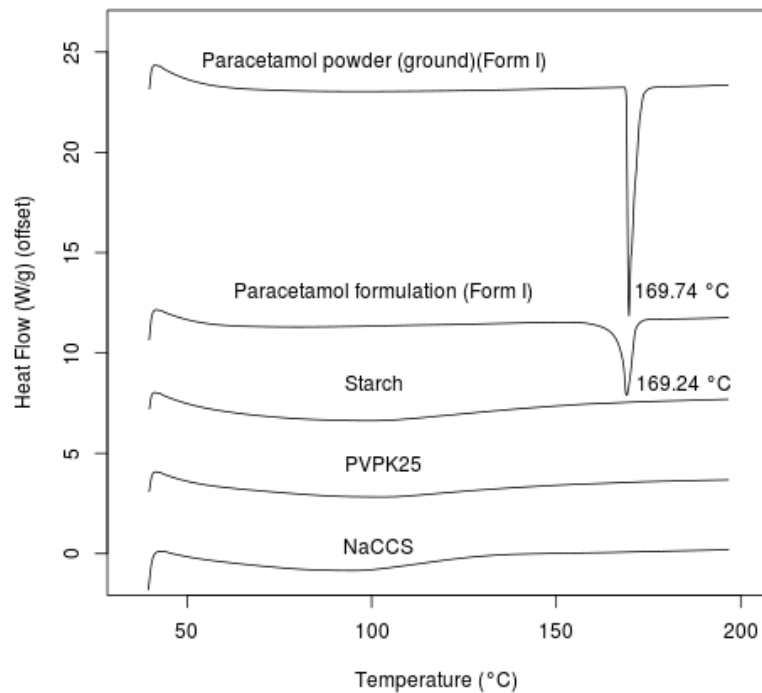
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a= N-H secondary amide, b= O-H, c= N-H secondary amide, d= C=O, e= C=O



546 **Figure 7.** FTIR spectra of paracetamol powder (ground Form I) and paracetamol formulation (left),  
 547 starch, PVP K25, NaCCS (right).



548

549 **Figure 8.** DSC thermograms of pure paracetamol, paracetamol formulation, starch, PVP K25, and  
 550 NaCCS.

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### List of tables

553 **Table I.** The percentage composition of various ingredients in paracetamol formulation feed stock.

Name of Material	Function	Total Formulae (mg)	Wt. % w/w (wet formulae)	Wt. % w/w (dry formulae)	Calc. drug weight (mg) (dry tablets)****
Paracetamol	API*	810.42	58.94	81.04	249.42
PVP**	Binder	100.00	7.27	10.00	30.78
Starch	Binder	83.33	6.06	8.33	25.64
CCS***	Disintegrant	6.25	0.45	0.63	1.94
Water	Binder	375.00	27.27	----	----
Total	----	1375.00	100.00	100.00	307.78

554  
 555 \*Active Pharmaceutical Ingredient, \*\*Polyvinylpyrrolidone \*\*\*Croscarmellose sodium \*\*\*\*Calculated from the  
 556 average of the total paracetamol tablet weight (307.78 mg,  $n = 6$ ).

557 **Table II.** Paracetamol 3D printed tablet's dimensions for different geometries of similar total weight  
 558 and increased surface area and SA/V ratios.

Geometry	Surface area (SA) (mm <sup>2</sup> )	Volume (V) (mm <sup>3</sup> )	SA/V ratio	Weight (mg)	Tablet dimension (mm)	Density (mg/mm <sup>3</sup> )
					*L×**H×***D	
Mesh	897±9.4	301±3.9	2.976±0.008	318±11.1	15.2±0.02×5.4±0.05×8.5±0.05	1.054±0.023
Ring	449.94±2.65	369.96±3.25	1.216±0.004	323.00±1.70	15.3±0.03×5.0±0.06×8.5±0.04	0.866±0.005
Solid	330.94±2.04	344.19±5.19	0.962±0.009	313.00±9.20	15.4±0.03×3.4±0.06×8.4±0.05	0.909±0.013

559 \*L=length, \*\*H=height, \*\*\*D=diameter

560 **Table III.** Fitting experimental release data, from the in vitro release of 3D printed paracetamol tablets  
 561 to Zero-order, First-order, Higuchi and Korsmeyer-Peppas kinetic equations at a buffer condition (pH  
 562 6.8-12 hrs).

Geometry	Zero order (r <sup>2</sup> )	First order (r <sup>2</sup> )	Higuchi (r <sup>2</sup> )	Korsmeyer-Peppas (r <sup>2</sup> )	<i>n</i> value
Mesh	0.38	<b>0.77</b>	0.53	0.64	0.25
Ring	0.67	<b>0.96</b>	0.84	0.91	0.44
Solid	0.91	0.98	<b>0.99</b>	0.98	0.56

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566 **Table IV.** Individual paracetamol 3D printed tablet's dimensions and their average, median, maximum,  
 567 minimum dimension, standard deviation.

Tablet no.	Mesh tablets (mm)			Ring tablets (mm)			Solid tablets (mm)		
	Length	Height	Width	Length	Height	Width	Length	Height	Width
1	15.24	5.42	8.50	15.01	5.13	8.30	15.39	3.50	8.35
2	15.21	5.51	8.47	15.33	5.08	8.47	15.40	3.46	8.47
3	15.20	5.40	8.38	15.38	4.94	8.50	15.34	3.36	8.46
4	15.22	5.46	8.51	15.30	5.07	8.40	15.36	3.52	8.33
5	15.26	5.45	8.53	15.26	5.09	8.42	15.26	3.38	8.42
6	15.19	5.49	8.47	15.16	5.06	8.38	15.37	3.42	8.45
Average	15.22	5.46	8.48	15.24	5.06	8.41	15.35	3.44	8.41
Median	15.22	5.46	8.49	15.28	5.08	8.41	15.37	3.44	8.44
Maximum	15.26	5.51	8.53	15.38	5.13	8.50	15.40	3.52	8.47

Minimum	15.19	5.40	8.38	15.01	4.94	8.30	15.26	3.36	8.33
SD	0.03	0.04	0.05	0.13	0.06	0.07	0.05	0.06	0.06

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569 **Table V.** Individual paracetamol 3D printed tablets weight, calculated paracetamol dose/tablet,  
570 percentage deviation, and their average, median, maximum, minimum weight and standard deviation.

Tablet no.	Ring-tablet			Mesh-Tablet			Solid-Tablet		
	Tablet weight (mg)	Calc. para. dose/tablet	Deviation %	Tablet weight (mg)	Calc. para. dose/tablet	Deviation %	Tablet weight (mg)	Calc. para. dose/tablet	Deviation %
1	312.90	253.57	0.78	308.80	250.25	0.33	307.10	248.87	0.44
2	318.80	258.36	2.68	300.70	243.69	-2.30	302.20	244.90	-1.16
3	309.90	251.14	-0.19	311.90	252.76	1.34	301.30	244.17	-1.46
4	307.70	249.36	-0.90	312.60	253.33	1.57	306.40	248.31	0.21
5	310.80	251.87	0.10	306.00	247.98	-0.58	306.60	248.47	0.28
6	302.80	245.39	-2.47	306.70	248.55	-0.35	310.90	251.95	1.68
Average	310.48	251.62	0.00	307.78	249.43	0.00	305.75	247.78	0.00
Median	310.35	251.51	-0.04	307.75	249.40	-0.01	306.50	248.39	0.25
Maximum	318.80	258.36	2.68	312.60	253.33	1.57	310.90	251.95	1.68
Minimum	302.80	245.39	-2.47	300.70	243.69	-2.30	301.30	244.17	-1.46
SD	5.33	4.32	1.72	4.38	3.55	1.42	3.52	2.85	1.15

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577 **Table VI.** Individual paracetamol 3D printed tablet's breaking force (kg and N), tensile fracture strength  
578 (MPa), and their average, median, maximum, minimum hardness and standard deviation.

Tablet no.	Mesh tablets			Ring tablets			Solid tablets		
	Breaking force (kg)	Breaking force (N)	Tensile strength (Mpa)	Breaking force (kg)	Breaking force (N)	Tensile strength (Mpa)	Breaking force (kg)	Breaking force (N)	Tensile strength (Mpa)
1	2.56	25.11	2.30	2.50	24.53	2.53	8.69	85.25	19.24
2	2.40	23.54	2.09	2.80	27.47	2.89	9.15	89.76	20.45
3	2.70	26.49	2.47	2.50	24.53	2.73	8.71	85.45	20.59
4	2.39	23.45	2.11	2.26	22.17	2.36	9.04	88.68	19.80
5	2.60	25.51	2.30	2.57	25.21	2.65	8.93	87.60	20.85
6	2.44	23.94	2.14	2.49	24.43	2.59	9.56	93.78	21.88

Average	2.52	24.67	2.24	2.52	24.72	2.63	9.01	88.42	20.47
Median	2.50	24.53	2.22	2.50	24.53	2.62	8.99	88.14	20.52
Maximum	2.70	26.49	2.47	2.80	27.47	2.89	9.56	93.78	21.88
Minimum	2.39	23.45	2.09	2.26	22.17	2.36	8.69	85.25	19.24
SD±	0.12	1.23	0.15	0.17	1.70	0.18	0.32	3.17	0.91

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580 **Table VII.** Friability of different paracetamol 3D printed geometries; mesh, ring, and solid tablets.

Tablet	Friability (%)	Comment
Mesh	0.65	Pass
Ring	0.62	Pass
Solid	0.59	Pass

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