

**RVC OPEN ACCESS REPOSITORY – COPYRIGHT NOTICE**

This author's accepted manuscript may be used for non-commercial purposes in accordance with [Wiley Terms and Conditions for Self-Archiving](#).

The full details of the published version of the article are as follows:

TITLE: Biomechanics of Two External Fixator Devices Used in Rat Femoral Fractures

AUTHORS: Osagie-Clouard, L; Kaufmann, J; Blunn, G; Coathup, M; Pendegrass, C; Meeson, R; Briggs, T; Moazen, M

JOURNAL: JOURNAL OF ORTHOPAEDIC RESEARCH

PUBLISHER: Wiley

PUBLICATION DATE: 4 May 2018 (online)

DOI: [10.1002/jor.24034](https://doi.org/10.1002/jor.24034)



## Biomechanics of Two External Fixator Devices Used in Rat Femoral Fractures

Journal:	<i>Journal of Orthopaedic Research</i>
Manuscript ID	JOR-17-0807
Wiley - Manuscript type:	Research Article (Member)
Date Submitted by the Author:	16-Nov-2017
Complete List of Authors:	osagie-clouard, liza; University College London Institute of Orthopaedics and Musculoskeletal Science, kaufmann, joshua; University College London Institute of Orthopaedics and Musculoskeletal Science Blunn, Gordon; UCL, IOMS; university of portsmouth Pendegrass, Catherine; University College London, Institute of Orthopaedics and Musculoskeletal Science Coathup, Melanie; university of central florida briggs, timothy; royal national orthopaedic hospital, stanmore meeson, richard; University College London Institute of Orthopaedics and Musculoskeletal Science Moazen, Mehran; UCL, mechanical engineering
Areas of Expertise:	finite element, fracture healing
Keywords:	Mechanics and Computational Modeling < Bone Fracture

SCHOLARONE™  
Manuscripts

1 **Biomechanics of Two External Fixator Devices Used in Rat Femoral Fractures**

2 Liza Osagie-Clouard<sup>1</sup>, Joshua Kaufmann<sup>1</sup>, Gordon Blunn<sup>1,2</sup>, Melanie Coathup<sup>3</sup>, Catherine Pendegrass<sup>1</sup>,  
3 Richard Meeson<sup>1</sup>, Timothy Briggs<sup>4</sup>, Mehran Moazen<sup>5</sup>

4 <sup>1</sup>Division of Surgery, University College London, Stanmore, UK; <sup>2</sup>University of Portsmouth,  
5 Portsmouth, UK; <sup>3</sup>University of Central Florida, USA; <sup>4</sup>Royal National Orthopaedic Hospital,  
6 Stanmore, UK; <sup>5</sup>Mechanical Engineering, University College London

7  
8 Corresponding Author

9 Liza Osagie-Clouard

10 IOMS,

11 Royal National Orthopaedic Hospital

12 Brockley Hill

13 Stanmore HA7 4LP

14 +447931809218

15 l.osagie@ucl.ac.uk

16

17 running title: external fixator biomechanics, and FEA

18

19 Author contributions: LOC, manuscript preparation, data collection, experimental design. JK

20 experimental design, data collection. GB, experimental design, manuscript preparation.

21 MC/CP/TB, experimental design and data analysis. RM, computational modelling and

22 manuscript preparation. MM, experimental design, data analysis, manuscript preparation.

23 all authors have read and approved the final submitted manuscript

24

25

26

**27 Abstract**

28 The use of external fixators allows for the direct investigation of newly formed  
29 intrafragmentary bone, and the radiographic evaluation of the fracture. We compared the in  
30 vitro stiffness' of two widely used external fixator devices used for in vivo analysis of fracture  
31 healing in rat femoral fractures with differing construction (Ti alloy ExFix1 and PEEK ExFix2)  
32 and correlated the results to a finite element (FE) model.

33

34 Rat femoral fracture fixation was modelled using two external fixators. For both constructs an  
35 osteotomy of 2.75mm was used, and offset maintained at 5mm. Tufnol, served as  
36 standardized substitutes for rat femora. Constructs were loaded under axial compression  
37 and torsion. Overall axial and torsional stiffness were compared between the in vitro models  
38 and FE results. FE models were also used to compare the fracture movement and overall  
39 pattern of von Mises stress across the external fixators.

40

41 In vitro axial stiffness of ExFix1 was  $29.26\text{N/mm} \pm 3.83$  compared to ExFix2  $6.31\text{N/mm} \pm 0.67$   
42 ( $p^* < 0.05$ ). Torsional stiffness of ExFix1 was  $47.5\text{Nmm}^\circ \pm 2.71$  compared to ExFix2 at  
43  $19.1\text{Nmm}^\circ \pm 1.18$  ( $p^* < 0.05$ ). FE results predicted similar comparative ratios between the  
44 ExFix1 and 2 as the in vitro studies. FE results predicted considerably larger intrafragmentary  
45 motion in the ExFix2 comparing to ExFix1.

46

47 We demonstrated significant differences in the stiffness' of the two external fixators; thus  
48 highlighting the large variations in the biomechanics of available external fixators and  
49 suggests that care must be taken when interpreting fracture healing outcomes; moreover,  
50 we also illustrate the utility of FEA modelling in this context.

51

52 **Keywords:** fracture fixation, finite element analysis, biomechanics

53

54

55

56 **1. Introduction**

57 Multiple physiological and mechanical factors govern the fracture healing process. Overall  
58 stiffness of the fracture fixation construct directly impacts the axial, torsional and shear  
59 intrafragmentary movement at the fracture site (1-3). These subsequently impact the healing  
60 process and as with physiological healing, rigid fixation will lead to intramembranous  
61 ossification, while those that are less rigid, allow for the creation of cartilaginous callus and  
62 endochondral ossification (4, 5).

63 Rodents have been widely used to investigate the fracture fixation. They are an invaluable  
64 animal model used to understand the fracture healing process and to develop new  
65 technologies and treatments to address complications such as non-union. A number of  
66 external fixators have been used to fix femoral fractures in rodents. These fixators, typically  
67 result in a combination of intramembranous and endochondral ossification with studies  
68 illustrating healing by various biological scenarios in different models (6, 7).

69 The literature comparing the biomechanical differences of existing external fixators in  
70 rodents is limited. Harrison et al. (8) reported no significant difference in axial stiffness  
71 between aluminium and titanium fixator bar materials. However pin material and thickness  
72 does have a large effect on torsional and axial stiffness. Mark et al. (9) reported a 50%  
73 decrease in axial stiffness and transverse stiffness of the fixator, when using a 1.0-mm  
74 compared to a 1.2-mm outer diameter pin. Willey et al (10) demonstrated significantly  
75 reduced stiffness at the fracture site of titanium alloy pins versus stainless steel in fixators of  
76 the same design, with similar effects of body material and offset on stiffness as previous  
77 studies. Glatt et al. (11) reported the development of a variable stiffness PEEK fixator where  
78 fracture rigidity can be altered during healing. This PEEK fixator is gaining favour for use in  
79 the investigation of rodent fracture healing as the four pin construct is lighter than traditional  
80 titanium and stainless steel fixators and has been shown to be well tolerated in vivo (12). In

81 contrast, the majority of studies utilise a more traditional unilateral fixator design such as the  
82 Harrison et al. titanium alloy fixator. Recently reported variations of the Harrison fixator utilise  
83 2 carbon fibre cross bars with four aluminium pins (13, 14); heavier than the Glatt fixator. No  
84 study to date has compared the effects of a variable stiffness fixator and a static fixator on  
85 the in vitro stabilisation of a rat femoral fracture model.

86 Studies investigating the effect of fixator construct on fracture stabilisation can be laborious,  
87 necessitating investigation of each design parameter-including crossbar number/size/ offset,  
88 pin size and each component material. Subsequently, the ability to utilise computational  
89 modelling to determine the mechanical characteristics of any fixator construct, is invaluable.  
90 So long as the models are validated using in vivo or in vitro experimental data finite element  
91 (FE) modelling provides a unique opportunity to model experimental scenarios  
92 computationally and accurately (15-17).

93 The aim of this study was to compare the biomechanics of two increasingly utilised rodent  
94 external fixators; a derivation of the Harrison et al titanium alloy fixator, and the Glatt/AO  
95 PEEK external fixator. We utilised a series of experimental in vitro testing and in silico  
96 computational models based on finite element method.

## 97 **2. Materials and Methods**

### 98 **2.1 External fixator designs**

99 The study compared two external fixator designs. The first (EXFix 1) has two graphite cross  
100 bars of 2x40mm, spaced 4mm apart, fixed between two titanium alloy (Ti6Al-4v) blocks.  
101 These blocks measured 8mm in height, 10mm in width and 7.2mm in depth. This design  
102 used 4 titanium alloy threaded pins of 0.8/1.0mm, fixed within the blocks with stainless steel  
103 grub screws. The second fixator (ExFix 2) was comprised of a single PEEK crossbar and  
104 again four stainless steel threaded pins. The crossbar measured 16.5mm long, 5mm wide  
105 and 2mm deep with four 1mm holes to locate the steel pins. A single 12.5mm long, 1mm  
106 wide rectangular opening runs parallel with the openings for the steel pins; again each pin

107 measured 0.8/1.0mm. The offset as measured from the free length of the pins beneath the  
108 crossbar to the upper surface of the bone, was kept constant at 5mm throughout testing.  
109 ExFix 1 weighed 6.23g (range 6.22-6.31g), and ExFix 2 3.11g (range 3.08-3.65g).

110 A hollowed homogenous rod of laminated Tufnol (Tufnol Composites, Birmingham, UK), of  
111 similar elastic modulus to adolescent rat femora (inner diameter 1.5mm, outer diameter  
112 4mm, length 35mm) served as standardised substitute for bone and fixed using ExFix1(n=5)  
113 and 2 (n=5). Fixation was carried out using custom drill guides of 0.8mm that allowed for the  
114 accurate predrilling of holes into the Tufnol, after which pins were manually screwed into  
115 position to breach both cortices by one thread. After the fixator was fixed to the Tufnol bone  
116 a fracture was created with a 2.75mm fracture gap maintained.

## 117 **2.2 In vitro testing**

118 The Tufnol specimens were tested non-destructively using a Zwick (Zwick-Roell, Germany)  
119 materials testing machine to determine axial and torsional stiffness. In compression, a  
120 maximum load of 40N was applied, with a preload of 0.5N at a rate of 0.5mm/min. Load was  
121 applied onto potted concave ends of the Tufnol via steel beads attached to the testing  
122 machine, and the loading-unloading process repeated three times for each sample.

123 In torsion both ends of the sample were fixed into titanium cylinders with grub screws to  
124 negate slipping during testing. One end of the Tufnol remained static, whilst a maximum  
125 vertical load of 40N was applied to the other end with a lever arm of 75mm, which led to a  
126 torsion of 3000 Nmm (26). Loading was repeated three times per specimen and torsional  
127 stiffness was calculated by dividing the applied torque by the degrees of rotation of the  
128 proximal end of the Tufnol.

## 129 **2.3 Finite element analysis**

130 Computer-aided design models of the bone and two external fixators were developed in  
131 CATIA V5 (Dassault Systèmes, Paris FR - Figure 1). Dimensions exactly reflected those of

132 the real-life fixator models and all parts assigned isotropic material properties; The Tufnol  
133 bone model has an elastic modulus of 6.5GPa and Poisson's ratio 0.4 (18-20). Titanium  
134 alloy blocks in the ExFix1 have an elastic modulus of 96GPa and a Poisson's ratio of 0.36.  
135 The Graphite rods have an elastic modulus of 4.1GPa and a Poisson's ratio of 0.17. The  
136 PEEK crossbar of the ExFix2 has an elastic modulus of 3.6GPa and a Poisson's ratio of  
137 0.38. Finally, stainless steel pins in both fixators were given the same mechanical properties:  
138 an elastic modulus of 193GPa and Poisson's ratio of 0.31. The effect of screw pull-out at the  
139 fixator-Tufnol interface was ameliorated by gluing these contacts during experimental  
140 testing; subsequently, the interface experienced minimal micro-motion upon loading in-vitro  
141 and allowed all pin-Tufnol interfaces to be modelled as "fully fixed".

142 Interfaces such as at the crossbar-pin interface had inherent micro-motion as they were  
143 either threaded into position or held with grub screws. Thus two simulations were created,  
144 one with all contacts "fully fixed" and a second with all grub screws and threaded contacts  
145 "relaxed" to account for this motion. The relaxed model used contact elements at the  
146 interfaces with a friction coefficient of 0.4 (15). The expectation being that the properties of  
147 each fixator would be between these two extreme models.

148 In order to replicate the boundary conditions of the test rigs, the constraints were applied  
149 within the concave housing of the Tufnol under axial loading conditions and along the  
150 outside face of the housing under torsional loading conditions. Additionally, the surface/node  
151 in which the load was applied was also constrained to translate in only the axis parallel to the  
152 line of loading.

153 Analyses were carried out in FE package ANSYS (Academic Research, Pennsylvania USA).  
154 Tetrahedral elements were used to mesh all components of the fixators and Tufnol.  
155 Convergence was tested on each fixator by increasing the number of elements from ca.  
156 5,000 to 2,000,000 incrementally. The solution for ExFix1 converged to within 5% at  
157 approximately 135,000 elements when measuring axial stiffness and approximately 260,000



158 elements when measuring torsional stiffness. For ExFix2, the solution converged for both  
159 quantities of interest at approximately 322,000 elements. Results converged substantially  
160 faster with the use of midside nodes, and as such they were used throughout.

161 In addition to axial and torsional stiffness, FEA was also used to evaluate fracture gap  
162 displacement as measured by nodes either side of the osteotomy. Von Mises stresses were  
163 calculated for each fixator and the points of maximal stress also determined. It must be  
164 noted that since in this study no detail validation of the strain pattern was carried out the  
165 stress results were analysed qualitatively.

## 166 **2.4 Statistical Analysis**

167 Statistical analysis was performed on the experimental data. The ANOVA assumption of  
168 normality was tested using the Shapiro–Wilks normality test. If the assumption was met, an  
169 ANOVA was performed, if not, a Mann Whitney *U* test was used. The data was analysed  
170 using Prism 4.03 (GraphPad Software Inc., San Diego, USA) and a significance level when  
171 comparing data was set at  $p < 0.05$ .

## 172 **3. Results**

### 173 **3.1 Axial stiffness:**

174 ExFix1 was  $29.26\text{N/mm} \pm 3.83$  compared to ExFix2  $6.31\text{N/mm} \pm 0.67$  ( $p^* < 0.05$ ). The fully  
175 restricted FEA model predicted axial values of  $79.95\text{N/mm}$  and  $31.57\text{N/mm}$  for ExFix1 and 2  
176 respectively. The model under secondary contact conditions produced axial values of  $46.12$   
177  $\text{N/mm}$  and  $7.52\text{N/mm}$  respectively (Figure 2A).

178

### 179 **3.2 Torsional stiffness:**

180 ExFix 1 was  $47.5\text{Nmm}^\circ \pm 2.71$  compared to ExFix 2 at  $19.1\text{Nmm}^\circ \pm 1.18$  ( $p^* < 0.05$ ). The  
181 fully restricted FEA model predicted torsional stiffness of  $98\text{Nmm}^\circ$  and  $50\text{Nmm}^\circ$  for ExFix 1

182 and 2 respectively. The model under secondary contact conditions produced torsional  
183 stiffness of 89.8Nmm/° and 27Nmm/° respectively (Figure 2B).

184

### 185 **3.3 Comparative ratios:**

186 The ratio of ExFix1: ExFix2, axial and torsional stiffness based on the in vitro experimental  
187 data was 4.6 and 2.5 respectively. The same ratio based on the FEA with fully fixed interface  
188 conditions were 2.5 (46% lower than the experimental data) and 2 (20% lower than the  
189 experimental data) for the axial and torsional stiffness respectively. The same ratio based  
190 on the FEA with relaxed interface were 5.1 (11% greater than experimental data) and 3.3  
191 (32% greater than experimental data) for the axial and torsional stiffness respectively (Figure  
192 3).

193

### 194 **3.4 Fracture movement:**

195 Total fracture movement as measured in the FE models, was greater for ExFix2 in all planes  
196 versus ExFix 1. Under 1mm of movement occurred with ExFix 1 at the maximal loading  
197 however, in the ExFix 2 the fragments come into contact leading to a fracture movement of  
198 about 2.7mm based on the relaxed interface model. Under axial loading ExFix 1 was found  
199 to have 0.54 and 0.91mm of movement with the fully fixed and relaxed models. Whereas  
200 ExFix 2 demonstrated 1.49 and 2.75mm of movement respectively. Under torsional  
201 conditions, ExFix1 showed 0.52 and 0.64mm of movement with the fully fixed and relaxed  
202 models. Versus ExFix2 with 2.20 and 2.74mm of movement respectively (Figure 4A and b).

203

### 204 **3.5 Stress pattern:**

205 The stress contour plots of the equivalent von Mises stresses for each fixator component are  
206 shown in Figure 5. In all components of the fixator ExFix1 experienced lower overall stress  
207 than ExFix2, in both axial and torsional loading. For all FE analysis maximum stress

208 occurred at the pin-Tufnol interface. In axial loading of both fixators, stress peaks in the pin  
209 closest to the point of loading was seen, whilst in torsion, maximum stress occurred in the  
210 pins either side of the fracture gap.

#### 211 **4. Discussion**

212 This study compared the mechanical characteristics of two commonly used external fixators  
213 in small animal fracture models. We used our in vitro findings to validate a series of finite  
214 element models based on axial and torsional stiffness data. Between the two fixators, we  
215 found significant differences in stiffness in both the axial and rotational planes, with ExFix1  
216 markedly more rigid in both planes. Throughout the study we maintained a constant offset,  
217 pin material and pin diameter, thus allowing the fixator design and crossbar material (Ti  
218 alloy/carbon fibre vs. PEEK) to be the dominating factors on overall stiffness. Previous  
219 studies have determined that pin size and material are the greatest determinants of fixator  
220 stiffness and intrafragmentary fracture movement (10, 21, 22), our data also suggests the  
221 significant impact that the fixator material properties and bar configuration have on the  
222 overall stiffness.

223 *In vitro* axial stiffness of both ExFix constructs were significantly less than those found with  
224 locked nailing techniques (23). ExFix1 was a third as stiff, and ExFix2 just over half as stiff  
225 as reported nailing data (23). Conversely rotational stiffness was greater for the external  
226 fixators than locked intramedullary nails, and indeed was greater than physiological numbers  
227 from intact bone (torsional stiffness 23Nmm/°). This greater stiffness in rotation, if related in  
228 vivo, will lead to reduced intrafragmentary movement in shear and as such will impact bone  
229 formation.

230 Our data suggests the FE model could predict the relative differences between the two  
231 external fixators. However, the FE models consistently predicted larger stiffness' then those  
232 found in vitro, this difference was considerably larger in the "fixed" model that did not  
233 account for any micro-motion at the pin-tufnol or the pin-fixator interfaces. When relaxing the

234 interfaces, the comparative ratios fell notably and were closer to the experimental *in vitro*  
235 data (see Fig 2). Again highlighting the fundamental role of micromotion at the interfaces in  
236 both the *in silico* and *in vitro* tests.

237 The difference in stiffness has a predictable effect on movement at the fracture gap, which  
238 has important implications on fracture healing. Intrafragmentary motion of between 0.2-1mm  
239 perpendicular to a diaphyseal fracture has been found to promote union, however, excessive  
240 axial and shear motion will result in delayed healing (1-3). Under axial conditions ExFix2  
241 experiences significant motion where bony fragments come into contact. ExFix1, however,  
242 restricts vertical motion under axial loading to under 1mm, within the desired envelope.  
243 Under torsion, this increases to a value equating to a rotation of up to 17 degrees. ExFix1  
244 limits rotation to less than half this amount at the same levels of loading. Under axial loading,  
245 translation and rotation at the fracture gap in ExFix1 is also negligible. Additionally, our  
246 findings are particularly relevant when investigating biological and pharmacological  
247 interventions where variability in stress across the gap will directly influence the efficacy of  
248 these factors (24-26).

249 The specific pin where the maximum stress occurs changes between loading conditions. In  
250 axial loading, maximum stress is located on the most proximal pin in both ExFix1 and ExFix2  
251 whereas under torsion, maximum stress occurred in the pin nearest the proximal end of the  
252 fracture. These changes are likely to be a function of the constraint of the tufnol bone  
253 creating higher stresses in the pins adjacent to the fracture site.

254 While the FE model could not exactly represent the *in vitro* assembly boundary conditions,  
255 the two conditions that were investigated can accurately predict upper and lower limits for *in vitro*  
256 results. Ultimately, we demonstrated considerable differences in the overall stiffness  
257 between the two fixators, which should be considered when comparing experimental *in vivo*  
258 data on fracture healing. Given a consistent fracture gap fractures stabilised using Exfix 2  
259 are more likely to heal though endochondral ossification or go onto a delayed or non union

260 compared to ExFix1. The *in silico* model where the threads are not fully bonded, predicted  
261 the comparative stiffness between the two fixators, as evidenced by the similar ratios. This  
262 data suggests that a computational protocol that includes the micro-motion present at the  
263 pin-bone interface, results in a reproducible model of experimental conditions. Further *in vivo*  
264 and computational work is required to demonstrate the effect of gap distance and fixator  
265 stiffness on the rate, type and quality of ossification and healing.

266

### 267 **Acknowledgments**

268 Funding was obtained from The Rosetrees and Gwen Fish Orthopaedic Trusts. Moazen was  
269 supported by Royal Academy of Engineering Research Fellowship.

### 270 **References**

- 271 1. Goodship AE, Kenwright J. The influence of induced micromovement upon the healing of  
272 experimental tibial fractures. *J Bone Joint Surg Br.* 1985;67:650–655.
- 273 2. Claes LE, Heigele CA, Neidlinger-Wilke C, Kaspar D, Seidl W, Margevicius KJ, Augat P.  
274 Effects of mechanical factors on the fracture healing process. *Clin Orthop Relat Res.*  
275 1998;355: S132–147.
- 276 3. Moazen M, Jones AC, Leonidou A, Jin Z, Wilcox RK, Tsiridis E. Rigid versus flexible plate  
277 fixation for periprosthetic femoral fracture - computer modelling of a clinical case. *Medical*  
278 *Engineering & Physics* 2012;34, 1041-8
- 279 4. McKibbin B. The biology of fracture healing in long bones. *J Bone Joint Surg.* 1978;60-  
280 B:150–162.
- 281 5. Mark H, Nilsson A, Nannmark U, Rydevik B. Effects of fracture fixation stability on  
282 ossification in healing fractures. *Clin Orthop Relat Res.* 2004;419,245-50

- 283 6. Histing T, Garcia P, Holstein JH, Klein M, Matthys R, Nuetzi R, Steck R, Laschke MW,  
284 Wehner T, Bindl R, Recknagel S, Stuermer EK, Vollmar B, Wildemann B, Lienau J, Willie B,  
285 Peters A, Ignatius A, Pohlemann T, Claes L, Menger MD. Small animal bone healing  
286 models: standards, tips, and pitfalls results of a consensus meeting. *Bone*. 2011;49:591-9
- 287 7. Holstein JH, Garcia P, Histing T, Kristen A, Scheuer C, Menger MD, Pohlemann T.  
288 Advances in the establishment of defined mouse models for the study of fracture healing and  
289 bone regeneration. *J Orthop Trauma*. 2009;23:S31-8
- 290 8. Harrison LJ, Cunningham JL, Strömberg L, Goodship AE. Controlled induction of a  
291 pseudoarthrosis: a study using a rodent model. *J Orthop Trauma*. 2003;17:11-21
- 292 9. Mark H, Bergholm J, Nilsson A. An external fixation method and device to study fracture  
293 healing in rats. *Acta Orthop Scand* 2003 74:476–482.
- 294 10. Willie B, Adkins K, Zheng X. Mechanical characterization of external fixator stiffness for a  
295 rat femoral fracture model. *J Orthop Res* 2009;27:687–93.
- 296 11. Glatt V, Evans C, Matthys R. Design, characterisation and in vivo testing of a new,  
297 adjustable stiffness, external fixator for the rat femur. *European Cells and Materials*. 2012  
298 12: 289-299.
- 299 12. Glatt V, Bartnikowski N, Quirk N, Schuetz M, Evans C. Reverse Dynamization: Influence  
300 of Fixator Stiffness on the Mode and Efficiency of Large-Bone-Defect Healing at Different  
301 Doses of rhBMP-2. *J Bone Joint Surg Am*. 2016;98:677-87
- 302 13 Ho CY, Sanghani A, Hua J, Coathup M, Blunn G. Mesenchymal stem cells with increased  
303 stromal derived factor 1 expression enhance fracture healing. *Tissue Eng Part A*. 2015;21,  
304 594-602
- 305 14 Lee O, Coathup M, Gooip A, Blunn G. Use of mesenchymal stem cells to facilitate bone  
306 regeneration in normal and chemotherapy treated rats. *Tissue Eng*. 2005;11, 1727-1735

- 307 15. Moazen M, Mak J, Jones AC et al. Evaluation of a new approach for modelling the  
308 screw-bone interface in a locking plate fixation: a corroboration study. *Journal of Engineering*  
309 *in Medicine*. 2013 227(7),746-756.
- 310 16. A. Macleod, P. Panka A. Simpson. Does screw-bone interface modelling matter in finite  
311 element analyses? *Journal of Biomechanics*. 2012;45,1712-6.
- 312 17. J. Wieding, R. Souffrant, A. Fritsche. Finite element analysis of osteosynthesis screw  
313 fixation in the bone stock: An appropriate method for automatic screw modelling. *PLOS One*.  
314 2012;7,1371.
- 315 18. Tufnol Composites Limited. 2016. Carp Brand Tufnol [online]:  
316 [http://www.tufnol.com/materials-full/fabric\\_laminates/carp-brand.aspx](http://www.tufnol.com/materials-full/fabric_laminates/carp-brand.aspx)
- 317 19. ANSYS® Academic Research, Release 16.2. Engineering Data, Material Properties,  
318 ANSYS, Inc.
- 319 20. AZO Materials. No date available. Supplier Data - Polyetheretherketone (PEEK)  
320 (Goodfellow) and Carbon - Graphite Materials [online].
- 321 21. Mark H, Nilsson A, Nannmark U. Effects of fracture fixation stability on ossification in  
322 healing fractures. *Clin Orthop Relat Res* 2004 419:245–250.
- 323 22. Mark H, Rydevik B. Torsional stiffness in healing fractures: influence of ossification: an  
324 experimental study in rats. *Acta Orthop* 2005 76:428–433.
- 325 23. Schoen M, Rotter R, Schattner S. Introduction of a new interlocked intramedullary nailing  
326 device for stabilization of critically sized femoral defects in the rat. *J Orthop Res* 2008  
327 26:184–189.

- 328 24. Sato M, Yasui N, Nakase T, Kawahata H, Sugimoto M, Hirota S, Kitamura Y, Nomura S,  
329 Ochi T. Expression of bone matrix proteins mRNA during distraction osteogenesis. J Bone  
330 Miner Res 1998, 13: 1221-1231.
- 331 25. Sato M, Ochi T, Nakase T, Hirota S, Kitamura Y, Nomura S, Yasui N. Mechanical  
332 tension-stress induces expression of bone morphogenetic protein (BMP)- 2 and BMP-4, but  
333 not BMP-6, BMP-7, and GDF-5 mRNA, during distraction osteogenesis. J Bone Miner Res  
334 1999, 14: 1084-1095.
- 335 26. Seebach C, Skripitz R, Andreassen TT, Aspenberg P. Intermittent parathyroid hormone  
336 (1-34) enhances mechanical strength and density of new bone after distraction osteogenesis  
337 in rats. J Orthop Res 2004, 22: 472-478.

338

### 339 **Figure Legends**

340 Figure 1. Computer aided designs of both external fixator models, with arrows demonstrating  
341 load constraint conditions.

342 Figure 2A and B. Demonstrating the torsional and axial stiffness' of both external fixators in  
343 vitro and in silico.

344 Figure 3. Demonstrating the comparative stiffness ratios in torsion and compression for in  
345 vitro and in silico testing.

346 Figure 4A and B. Demonstrating total fracture movement as found in silico under  
347 compression (A) and torsion (B).

348 Figure 5. Equivalent von-Mises stress contour plots on the crossbars of both fixator models.

349



350

351

352

353

354

355

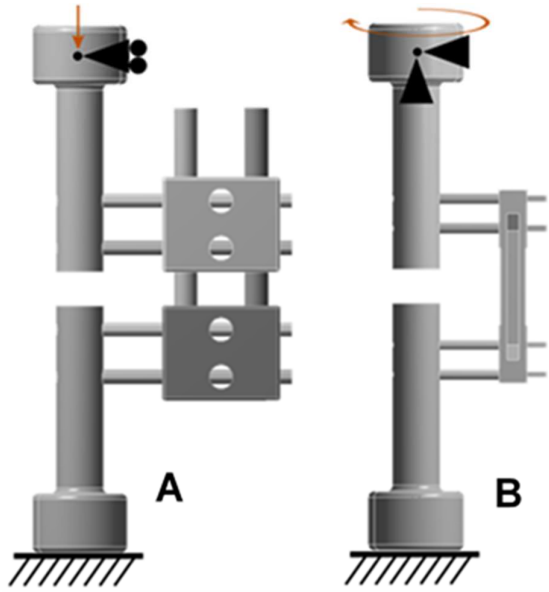
356

357

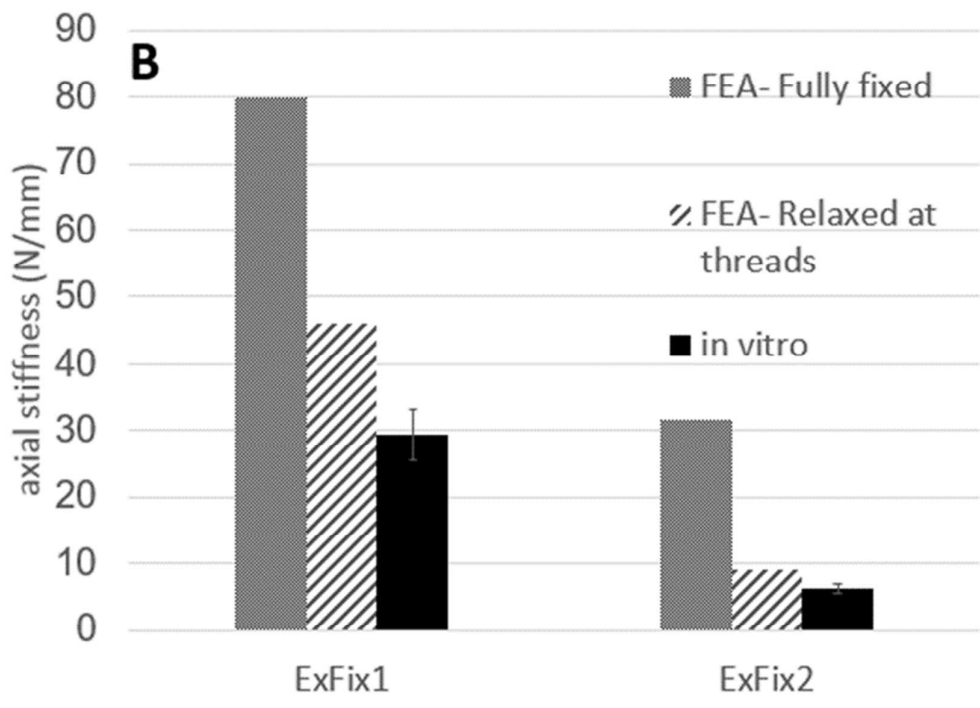
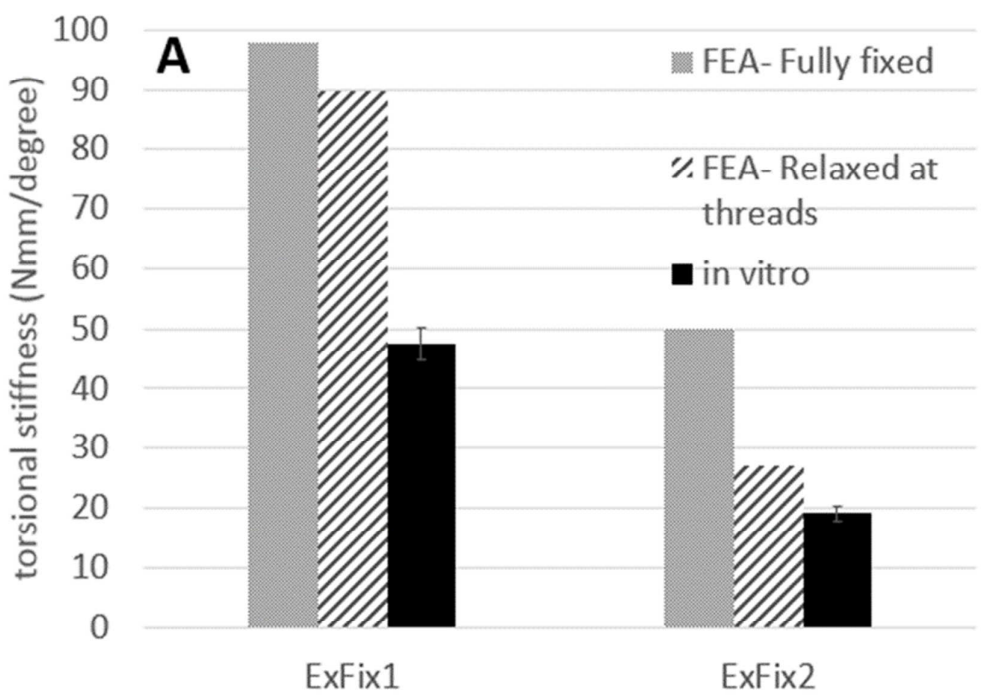
358

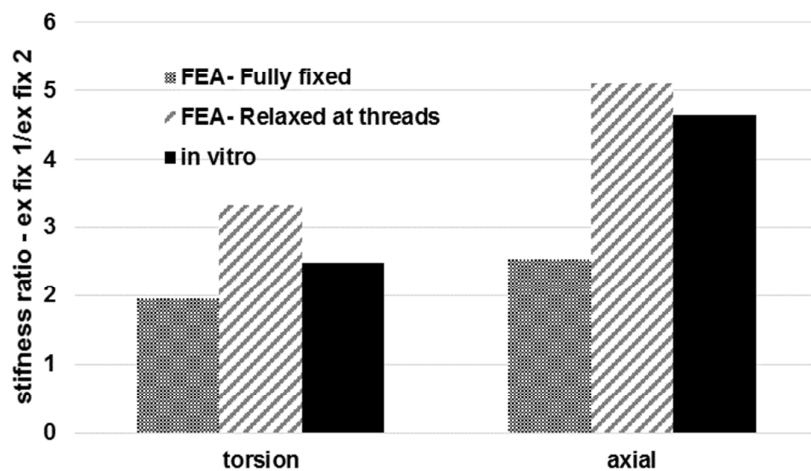
359

For Peer Review

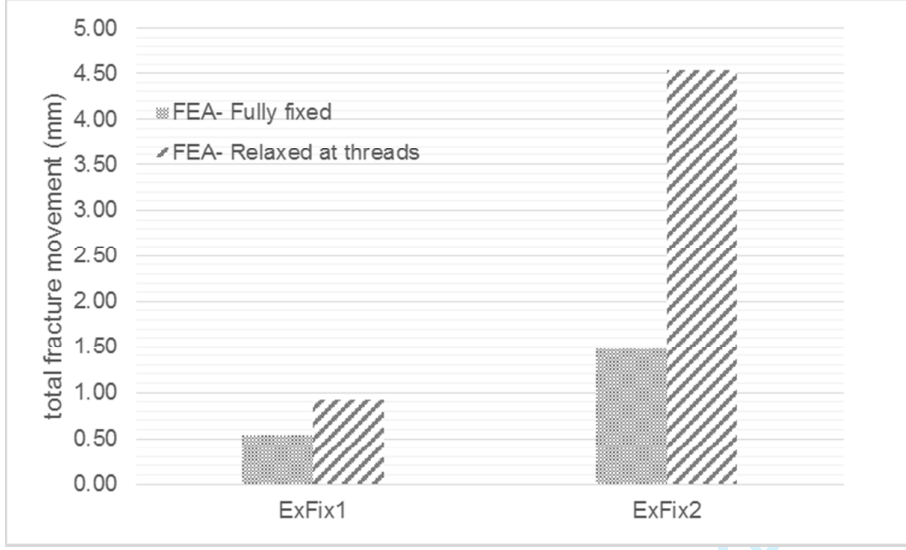
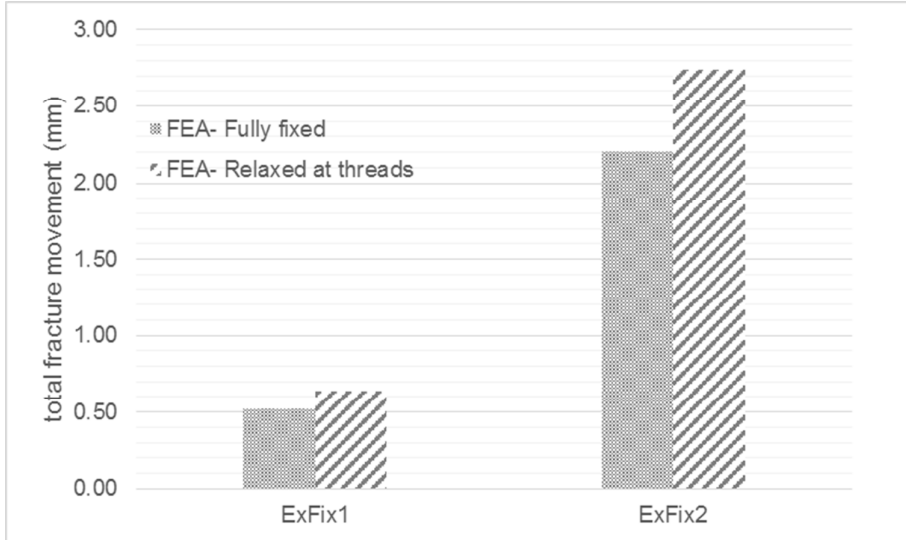


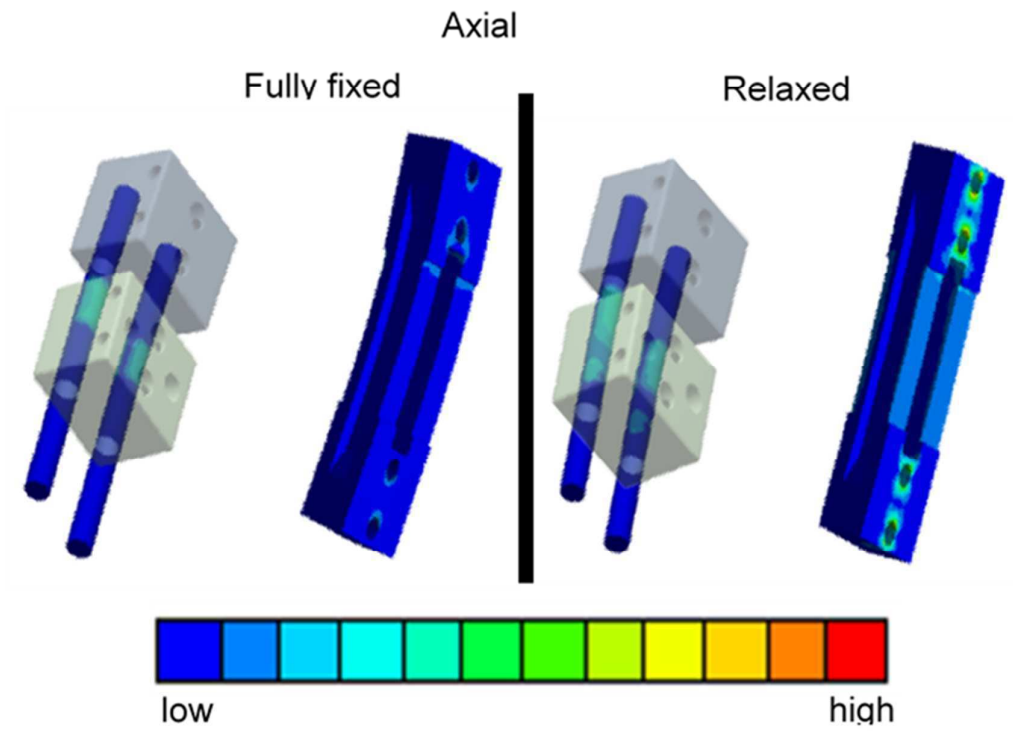
Or Peer Review





For Peer Review





Peer Review