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Interactive Influence of Turbidity and Light on Larval Bluegill (Lepomis macrochirus) Foraging

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In a series of in situ enclosure experiments with larval bluegill (*Lepomis macrochirus*), we demonstrate that turbidity from suspended sediments reduces bluegill consumption of crustacean zooplankton, primarily cyclopoid copepods and copepod nauplii. However, this reduction occurred only when light intensity in parts of enclosures fell below a threshold, estimated at <450 lx. Following recent studies demonstrating copepod diel vertical migration in response to predators, it appears that copepods in our experiments used low-light strata as a refuge. Without this apparent refuge present, larval bluegill consumption increased with increasing turbidity, but prey were smaller on average. Thus, prey biomass consumed by larval bluegill did not differ with turbidity in high-light conditions. We postulate that the shift to smaller prey across taxa at higher turbidity, when light intensity exceeded 450 lx, derives from increased prey–background contrast. In low-light conditions, larval bluegill consumed larger, but fewer, zooplankton with increasing turbidity, resulting in lower prey biomass consumed. Thus, we demonstrate the field conditions causing negative turbidity effects on larval fish foraging success, and thus growth and recruitment.

Dans une série d'expériences in situ (dans une enceinte) avec des larves de crapet arlequin (Lepomis macrochirus), nous avons démontré que la turbidité provoquée par les sédiments en suspension réduisait la consommation de crustacés zooplanctoniques, surtout des copépodes cyclopïdes et des nauplii de copépodes. Toutefois, cette réduction ne s'observait que lorsque l'intensité lumineuse dans certaines partie de l'enceinte était inférieure à une valeur seuil estimée à <450 lx. Des études de suivi récentes ayant montré la migration verticale nycthéméral des copépodes en réponse à l'action des prédateurs, il semble que dans nos expériences les copépodes aient utilisé les couches de faible intensité lumineuse comme refuge. Sans la présence de ce refuge apparent, la consommation des crapets larvaires augmentait avec l'accroissement de la turbidité, mais les proies étaient en moyenne plus petites. Ainsi, la biomasse des proies consommées par les crapets larvaires n'a pas changé avec la turbidité dans des conditions d'intensité lumineuse élevée. Nous avons postulé que le changement vers la consommation de proies plus petites dans l'ensemble des taxons en présence d'une turbidité plus élevée, lorsque l'intensité lumineuse dépasse 450 lx, est attribuable une augmentation du contraste entre les proies et le fond. Dans des conditions de faible luminosité, les crapets larvaires ont consommé du zooplancton moins nombreux, mais plus gros, au fur et à mesure qu'augmentait la turbidité, avec le résultat que la biomasse des proies consommées est plus petite. Ainsi, nous avons démontré les conditions qui, sur le terrain, sont à l'origine des effets négatifs liés à la turbidité sur l'alimentation des poissons larvaires et, par conséquent, sur la croissance et le recrutement.

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Survival of larval fish largely depends on size-selective predation (see reviews in Blaxter 1986; Miller et al. 1988). Therefore, any environmental factors affecting larval growth or exposure to predation should influence recruitment success (Miller et al. 1988; Margulies 1990). Turbidity from suspended sediments is a common feature of many lakes and reservoirs, particularly those in agricultural watersheds, and, along with sedimentation, is a major factor in rivers and lakes that can affect organisms throughout the aquatic community (see reviews in Muncy et al. 1979; Arruda et al. 1983; Wilber 1982; Bruton 1985; Goldsborough and Kemp 1988). Herein, Reçu le 2 juin 1992 Accepté le 26 octobre 1992

we quantify how turbidity from suspended sediments influences foraging success by larval fish, interpreting these results in the context of larval growth, predation risk, and recruitment.

For particulate feeding juvenile and adult fishes, turbidity reduces foraging ability by reducing reaction distance (i.e., maximum distance for detecting prey and predators, Vinyard and O'Brien 1976; Crowl 1989). However, studies with larval fishes have produced conflicting results; turbidity has decreased, increased, and had no effect on foraging success or growth rate (Johnston and Wildish 1982; Boehlert and Morgan 1985; Breitburg 1988; Chesney 1989).

In studies where foraging rate by larval fish declined with increasing turbidity, results were postulated to occur because of reduced reaction distances (Johnston and Wildish 1982; Breitburg 1988), as has been demonstrated for adult fishes (Vinyard and O'Brien 1976; Barrett et al. 1992). Conversely, when turbidity enhanced foraging success of larval fishes,

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increasing contrast between prey and background has been suggested as the mechanism (Boehlert and Morgan 1985). Finally, the lack of a turbidity effect on larval striped bass (*Morone saxatilis*) growth (Chesney 1989), as turbidity increased from 0 to 130 nephelometric turbidity units (NTU; APHA et al. 1985), a typical range in estuaries and turbid reservoirs, suggests that other interacting variables affect foraging success.

Light intensity reduces foraging rate and growth of larval fish at levels <100 lx, although the rate of decline and exact light conditions causing the decline vary among species (see review in Blaxter 1986). For striped bass larvae, growth in clear water declines exponentially as light intensity falls from 300 to 0.4 lx, while growth does not change above 450 lx (Chesney 1989). When turbidity negatively affects foraging (Johnston and Wildish 1982; Breitburg 1988), light intensity above laboratory containers has been <300 lx and as low as 5 lx. Positive effects of turbidity on larval fish feeding occurred when light intensity was >800 lx (Boehlert and Morgan 1985).

Hence, increased turbidity in lakes and rivers may influence larval fish feeding rates primarily through increased light attenuation, which may result in suboptimal light conditions for foraging. In addition, turbidity can alter prey-background contrast (Hinshaw 1985), as well as affect vertical distribution of fish larvae (Swenson and Matson 1976; Matthews 1984) and zooplankton (Zettler and Carter 1986). Given that light intensity declines with depth, results from experiments in shallow aquaria may not accurately reflect effects of turbidity on larval fish foraging in the field where predator and zooplankton prey can select optimal depths. Therefore, we designed field enclosure experiments using larval bluegill (*Lepomis macrochirus*), a common limnetic species that feeds on zooplankton (Werner 1967; Werner and Hall 1988; DeVries et al. 1991), to determine how turbidity influences its foraging in situ.

Methods

Foraging experiments were conducted on each of four consecutive days in Stonelick Lake, a 69-ha reservoir in southwest Ohio with an upstream-to-downstream turbidity gradient reflected by typical Secchi depths of about 20-120 cm, respectively. For these experiments, we placed two transparent polyethylene enclosures (1-m diameter) at each of three sites along the gradient. Depth was 1.7, 1.5, and 1.1 m at the low-, intermediate-, and high-turbidity sites, respectively; enclosures extended through the entire water column, but were closed at the bottom, excluding the mud substrate. Turbidity was ambient in the two lower turbidity treatments (i.e., about 10 and 20 NTU, Sargent-Welch turbidimeter). To attain the highturbidity treatment of about 30-40 NTU, we added small quantities of bottom sediments (i.e., from 50-cm sediment cores at the high-turbidity site) to enclosures the day before an experiment began. However, during one set of experiments, too much sediment was added to one enclosure resulting in a turbidity of 64 NTU.

Enclosures were filled 1 d before an experiment with filtered (60- μ m mesh) water pumped from a depth of 1 m. To generate similar zooplankton assemblages among turbidities, organisms for all enclosures were collected at the intermediate-turbidity site. Water was pumped from a depth of 1 m and two times the volume of a single enclosure filtered through a 60- μ m-mesh net. After thorough mixing, the zooplankton sample was split equally and half was introduced into each of the two enclosures within a turbidity treatment. Because of travel time between

turbidity treatment sites, we did not completely randomize inoculations of zooplankton among enclosures.

Larval bluegill (12.5 \pm 0.08 mm total length, $\bar{X} \pm$ SE) were collected from a nearby pond, held in the laboratory for at least 48 h, and fed *Artemia* sp. nauplii and a mixed culture of zoo-plankton (mostly copepods) daily. Larvae were then starved for about 18 h before release into enclosures. We added 15, 13, and 10 bluegill to low-, intermediate-, and high-turbidity enclosures, respectively; this resulted in a consistent density of 11 larvae \cdot m⁻³ among enclosures of different depths. Peak density of bluegill larvae in Stonelick Lake averages 2.2 fish \cdot m⁻³ (DeVries et al. 1991).

Bluegill larvae were introduced into 3-L containers (with 500- μ m-mesh sides), which were hung inside enclosures overnight. Before release of larvae into enclosures, water in each enclosure was mixed to resuspend sediments and zooplankton were sampled (i.e., after mixing). Water was not mixed during these short-term experiments because turbidity never declined sufficiently to cause turbidity in any treatments to overlap (i.e., maximum change at lowest and highest turbidity during experiments was about 2 and 10 NTU, respectively). Zooplankton were sampled with a 5.1-cm-diameter tube, collecting \geq 5 L of water from the entire water column. Water was filtered through a 60- μ m-mesh net and zooplankton were preserved in 10% formalin.

Bluegill larvae were released into enclosures at 9:00–10:00 and allowed to feed for 3.5–5.5 h, which ensured evacuation of initially ingested prey. Starved larvae select larger prey at first-feeding than are consumed during continuous feeding (Confer and O'Bryan 1989). We did not determine gut passage time explicitly for bluegill, but larvae of another centrarchid, largemouth bass (*Micropterus salmoides*), evacuate in about 2 h at 22–23°C (Laurence 1971). Water temperature in our enclosures was 24–27°C. Because evacuation rate increases with temperature, we believe that initially eaten prey had passed through larval guts within the duration of our experiments.

At experiment's end, turbidity was measured at the 0.5-m depth in all enclosures. In two sets, light intensity (measured as illuminance with a LICOR LI-188b photometer), dissolved oxygen, and temperature were measured at 0.5-m depth intervals in one enclosure for each turbidity treatment. Water was pumped out of enclosures (i.e., duration = ~ 10 min) and blue-gill larvae were filtered from this water. Not all larvae were retrieved intact because of pump damage; those collected were preserved in 10% formalin.

To quantify zooplankton abundance and composition, we identified at least 200 crustacean zooplankters from each sample. These organisms were obtained by adding the entire sample to a graduated cylinder and then removing vertically well-mixed subsamples with a large-bore (8-mm-diameter) pipette. The first 10 individuals identified of each taxon per sample were measured for length (nearest 0.034 mm, excluding spines and caudal rami); these lengths were converted to biomass using length – dry weight regressions (Dumont et al. 1975; G.G. Mittelbach, Kellogg Biological Station, Michigan State University, East Lansing, Mich., unpublished data).

We identified prey to the lowest taxon possible (86% of cladocerans to genus and 98% of adult copepods to suborder) in guts of 5–15 bluegill larvae from each enclosure. Up to 10 prey taxon⁻¹ fish⁻¹ were measured for length to estimate biomass consumed. We used Chesson's alpha (Chesson 1978, 1983), which compares the proportion of each taxon in the diet and in zooplankton samples, to quantify selection of prey by

TABLE 1. Light intensity (lx) across turbidities in enclosures with foraging bluegill larvae during low and high ambient light in Stonelick Lake, Ohio

Depth (m)	Low li	ight (set 3) a	at turbidity (High light (set 4) at turbidity (NTU):			
	11.5	18	43	64	10.5	19.5	31
0.0	7540	8400	4080	5020	63000	57000	65400
0.5	1960	1590	210	180	21400	10000	6980
1.0	457	314	14	5	5500	1200	479



FIG. 1. Zooplankon density in enclosures with varying turbidity in Stonelick Lake, Ohio, on four consecutive days. Enclosures were placed at low (L), intermediate (I), and high (H) turbidity sites within the lake. Standard errors are based on a single sample from each of two enclosures at each site.

larval bluegill. Because enclosures were the experimental units, proportions in the diet for alpha calculation were determined by pooling numbers consumed by all fish within each enclosure, rather than calculating alpha for individual fish and averaging for an enclosure (see Cochran (1977) for a discussion of averages of ratios). The formula for this index is

alpha =
$$\frac{r_i/p_i}{\sum (r_i/p_i)}$$

where the proportion of prey item i is p_i for the environment (enclosures) and r_i for the diet. The index ranges from zero to 1, with zero indicating complete avoidance of the prey taxon.

To determine if initial zooplankton densities in enclosures differed among turbidity treatments (sites) and sets (dates), we conducted two-way ANOVA of total zooplankton density (log transformed). There was significant interaction between treatments and sets (F = 3.3; df = 6,12; p = 0.037), suggesting patchy zooplankton distribution (Fig. 1). However, the range of zooplankton density was similar among turbidity treatments over the course of the experiment; therefore, we analyzed data for prey consumption as a function of turbidity, using multiple linear regression analysis, including zooplankton density as an independent variable.

Results

Light intensity was high for sets 1, 2, and 4 because there was little or no cloud cover. Overcast conditions prevailed during set 3 such that surface light intensity was reduced by 90% relative to set 4 (Table 1). Light intensity was not measured in sets 1 and 2, although weather conditions were similar to those in set 4. At 1-m depth in set 3, light intensity ranged between 5 and 460 lx from the highest (64 NTU) to lowest (11 NTU) turbidity (illuminance measured before an enclosure was drained) whereas in high-light conditions, light intensity remained >460 lx for all turbidities and depths (Table 1). Because low light (<100-300 lx) influences larval fish feeding and growth (Blaxter 1986; Chesney 1989), we analyzed high-light and low-light data separately.

Abundance in Diet

Number of zooplankton consumed by bluegill larvae was correlated with turbidity, but responses differed with light intensity (Fig. 2). At high light, larvae ate more zooplankton as turbidity increased ($r^2 = 0.35$; p = 0.006; N = 18), while fewer zooplankton were consumed with increasing turbidity at low light ($r^2 = 0.86$; p = 0.005; N = 6). Regardless of light intensity, neither zooplankton density nor the interaction between turbidity and density improved the fit of these models (p = 0.51 and 0.79, respectively, at high light and p = 0.48 and 0.37, respectively, at low light from multiple linear regressions). No zooplankton density effect was detectible, even though there was a wide range of density among enclosures (i.e., 125-800 organisms $\cdot L^{-1}$).

Among prey groups, cyclopoid copepods and copepod nauplii were selected by bluegill larvae, while copepod nauplii and the cladocerans *Bosmina longirostris* and *Ceriodaphnia* sp. were the most common prey in the environment (Fig. 3). Cyclopoid copepods and copepod nauplii consistently composed about 70% of prey consumed.

Because high and low light intensities occurred during different sets, differences attributed to light intensity – turbidity interactions may have derived from date effects. However, other variables potentially affecting larval bluegill foraging, such as dissolved oxygen concentration and temperature, were consistent during this 4-d experiment (Table 2). Hence, we conclude that larval bluegill foraging was influenced by light intensity as mediated through turbidity.

Prey Selection

To determine if turbidity affected prey selection, we regressed Chesson's alpha for each taxon against turbidity at high and low light. If turbidity failed to modify prey selection by larvae (*t*-test of slope, p > 0.05), we compared mean selectivity across turbidities to an expected value based on neutral

		Day						
	1	2	3	4				
Turbidity (NTU)								
Low	9, 10	8, 9	11, 11.5	10, 10.5				
Intermediate	16.5, 17	17.5, 18	18, 19.5	19, 19.5				
High	29, 29	39, 39.5	43,64	31, 31				
Temperature (°C)	25.5-27	24	25	24-26				
Oxygen (mg· L^{-1})	9.9-11.6	_		8.6-9.8				

TABLE 2. Physical conditions in enclosures during larval bluegill foraging trials in Stonelick Lake, Ohio, June 27–30, 1988. Turbidity, temperature, and dissolved oxygen data were taken at 0.5 m.



FtG. 2. Mean number of zooplankton consumed per larval bluegill from each enclosure in high-light (cloudless) and low-light (overcast) treatments in Stonelick Lake, Ohio. Each symbol represents a mean per enclosure (N = 5-15 per enclosure).

selection (i.e., the reciprocal of the number of prey taxa in the environment, which in this case was alpha = 0.14).

At high light, only selection for copepod nauplii and calanoid copepods changed as a function of turbidity (Table 3). Copepod nauplii were negatively selected at low turbidity (alpha = 0.02 \pm 0.01; $\bar{X} \pm$ SE). However, selection for copepod nauplii increased with increasing turbidity, resulting in positive selection at the highest turbidity (alpha = 0.33 \pm 0.10; Fig. 3). In contrast, as turbidity increased, selection for calanoid copepods declined, and was always less than neutral selection (alpha = 0.05 \pm 0.02 at about 10 NTU; Table 3). Although selection for other taxa was unaffected by turbidity at high light, cyclopoid copepods and *Diaphanosoma leuchtenbergianum* were consistently preferred by larval bluegill (alpha = 0.43 \pm 0.04 and 0.32 \pm 0.05, respectively); conversely, *B. longirostris*, *Ceriodaphnia* sp., and *Daphnia parvula* were negatively selected (alpha \leq 0.05; Table 3).

Low light occurred during only one set of experiments; thus, sample size (N = 6) and the power to detect changes in selection were low. Given this caveat, prey selection did not change across turbidity for any prey at low light intensity (p > 0.05; Table 3). As in high light, cyclopoid copepods were positively selected at all turbidities (alpha = $0.62 \pm 0.05;$ Table 3) whereas small cladocerans (i.e. *B. longirostris* and *Ceriodaphnia* sp.), calanoid copepods, and copepod nauplii were negatively selected (Table 3).



FIG. 3. Proportional abundance of zooplankton in enclosures and in diet of larval bluegill in Stonelick Lake, Ohio, at low (L), intermediate (I), and high (H) turbidity. CYC, cyclopoid copepods; CAL, calanoid copepods; NAU, copepod nauplii; DAP, *Daphnia parvula*; DIA, *Diaphanosoma leuchtenbergianum*; CER, *Ceriodaphnia* sp.; BOS, *Bosmina longirostris*.

Size Selection

Mean length (log transformed) of zooplankters consumed by larval bluegill declined as turbidity increased at high light intensity, corresponding to the increased selection for copepod nau-

TABLE 3. Regression analysis of prey selection (using Chesson's alpha, Chesson 1978, 1983) by larval bluegill in enclosures in Stonelick Lake, Ohio, as a function of turbidity at high-light (cloudless) and low-light (overcast) intensity. When slopes did not differ from zero (*t*-test, p < 0.05), mean alphas were compared with neutral selection (i.e., 0.14, reciprocal of the number of prey taxa in the environment). A dash indicates that the mean was not calculated because turbidity affected prey selection by larvae.

	High light $(N = 18)$						Low light $(N = 6)$					
	H_0 : slope = 0		H_0 : mean = 0.14			H_0 : slope = 0			H_0 : mean = 0.14			
Prey taxa	t	p > t	r^2	alpha ± SE	t	p > t	t	p > t	r ²	alpha ± SE	t	p > t
Copepods												
Nauplii	4.91	< 0.001	0.60				-1.79	0.15	0.45	0.04 ± 0.02	-5.37	< 0.01
Calanoid	-2.90	0.01	0.34				0.42	0.70	0.04	0.05 ± 0.01	-6.68	< 0.01
Cyclopoid	-0.79	0.44	0.04	0.43 ± 0.04	6.50	< 0.001	-0.31	0.77	0.02	0.62 ± 0.05	10.40	< 0.01
Cladocerans												
Bosmina												
longirostris	-0.42	0.68	0.01	0.01 ± 0.00	-51.13	< 0.001	-0.63	0.56	0.09	0.00 ± 0.00	- 156.3	< 0.001
Ceriodaphnia sp.	1.90	0.08	0.18	0.01 ± 0.00	- 85.71	< 0.001	1.80	0.15	0.45	0.02 ± 0.01	- 12.43	< 0.001
Daphnia parvula	-1.36	0.19	0.10	0.05 ± 0.01	-7.05	<0.001	-0.75	0.49	0.12	0.10 ± 0.03	-1.31	>0.2
Diaphanosoma												
leuchtenbergianum	-1.62	0.13	0.14	0.32 ± 0.05	3.62	<0.01	2.13	0.10	0.53	0.16 ± 0.03	0.58	>0.5



FIG. 4. Mean length of zooplankton in larval bluegill diets from enclosures along a turbidity gradient in Stonelick Lake, Ohio. See Table 1 for differences between light conditions. In high light, $r^2 = 0.43$ and p = 0.003; in low light, $r^2 = 0.83$ and p = 0.012.

plii ($r^2 = 0.43$; p = 0.003; N = 18; Fig. 4). At 10 NTU, predicted mean prey length was 0.28 mm whereas mean length declined 30% to 0.19 mm at 40 NTU. When we compared prey length within a taxon, smaller cyclopoid copepods and copepod nauplii (i.e., composing 59 and 22% of prey in the diet across all turbidities) were consumed by larval bluegill as turbidity increased at high light (F = 3.61; df = 4,14; p = 0.03; MAN-OVA using Wilk's criterion, SAS Institute, Inc., 1985).

At low light intensity, bluegill consumed larger prey as turbidity increased ($r^2 = 0.83$; p = 0.012; N = 6; Fig. 4). Mean prey length at low turbidity (11 NTU) was 0.28 mm, the same as at high light. However, at high turbidity (43 and 64 NTU), larvae consumed prey that averaged 0.35 mm long, representing a 75% increase in mean prey length over that consumed in high-light sets at high turbidity. Also, at low light, size of cyclopoid copepods and copepod nauplii consumed increased from low to high turbidity (F = 9.17; df = 4,4; p = 0.02; MANOVA). In summary, at high light, bluegill larvae tended to select smaller zooplankton with increasing turbidity whereas prey size increased when turbidity increased at low light.



FIG. 5. Biomass of prey recovered from larval bluegill guts as a function of turbidity at high and low light in Stonelick Lake, Ohio.

Biomass Consumed

Because number of prey consumed increased and size of prey decreased with turbidity at high light, and because reciprocal results were obtained at low light, we compared dry mass of zooplankton consumed across turbidity. At high light, the increase in number of prey consumed was offset by a reduction in prey size, resulting in no significant relationship between turbidity and biomass consumed $(r^2 = 0.00; p = 0.83, N = 18; Fig. 5)$. However, at low light, biomass consumed declined with increasing turbidity $(r^2 = 0.85; p = 0.009; N = 6; Fig. 5)$. Therefore, whereas turbidity influenced selection of prey taxa consumed by larval bluegill, change in prey biomass consumed was detected only for the low-light condition.

Discussion

Foraging by larval bluegill was influenced by interaction between turbidity and light. When light intensity throughout enclosures exceeded 450 lx, increasing turbidity caused larvae to select smaller prey both within and among taxa. However, biomass of prey consumed did not differ across turbidities because numbers of prey increased with turbidity. Conversely, when light intensity in large portions of enclosures fell below 300 lx during low ambient light conditions (i.e., set 3), both prey numbers and biomass consumed declined with increasing turbidity. These data provide insight into conflicting results in the literature for larval fish in which turbidity effects were measured at only one light intensity (i.e., either low or high light, Johnston and Wildish 1982; Boehlert and Morgan 1985; Breitburg 1988).

Johnston and Wildish (1982) found that larval Atlantic herring (*Clupea harengus*) consumed fewer prey as turbidity was increased from 0 to about 13 NTU at 300 lx (i.e., 20 mg estuarine sediments·L⁻¹ calculated from Breitburg 1988), while larval Pacific herring (*Clupea pallasi*) consumed more prey at 90 NTU than in clear water when light intensity was >800 lx (Boehlhert and Morgan 1985). Striped bass larvae also consumed fewer prey organisms from natural assemblages of copepods as turbidity increased at low light (i.e., 5 lx; Breitburg 1988). Our field results support these observations, in which turbidity has negative effects on foraging only at low light, and may enhance prey contrast at higher light intensity, increasing prey detection and consumption by larvae (see Hinshaw 1985 for discussion of contrast).

Larval fish responses to turbidity have been explained in terms of reduced reaction distance with increasing turbidity, as occurs with juvenile and adult particle-feeding planktivores (Howick and O'Brien 1983; O'Brien 1987). However, relative to adults and juveniles, larval bluegill have poor visual acuity. Although bluegill larvae rapidly develop a complex visual system as they grow (Kawamura and Mukai 1984), reaction distance (i.e., acuity) is probably little more than one body length at a size of 12 mm total length (TL) (review for other species in Blaxter 1986; Wanzenbock and Schiemer 1989). In contrast, when bluegill are 31 mm TL, they possess retinas with a high density of rods and cones (Hairston et al. 1982; Breck and Gitter 1983), enabling them to detect zooplankton as far as 30 cm away (Vinyard and O'Brien 1976). When reaction distance is short, there are fewer suspended particles between predator and prey to reflect light and interfere with detection (i.e., much as fog affects long-distance vision, but has little effect on detection of close objects). Thus, while juvenile and adult bluegill exhibit an exponential decline in reaction distance with increasing turbidity (Vinyard and O'Brien 1976; Miner 1990), there may be little negative effect on reaction distance of larvae with sufficient light (>300 lx).

In contrast with the results at high light, increasing turbidity in the low-light set appears to have reduced light intensity below a threshold necessary for maximizing foraging rates by larval bluegill. Because larvae possess a rudimentary visual system with relatively few rods necessary for vision at low light intensity, feeding and growth can be negatively affected at low light (<100-300 lx, Blaxter 1986; Chesney 1989). In our experiments at low ambient light, high- and intermediate-turbidity enclosures contained extensive areas in which light intensity was <300 lx and prey consumption was depressed. However, in these in situ enclosures, why would foraging decline if larvae could use areas with optimal lighting (i.e., top 0.5 m)? These results suggest that larval bluegill were either using the lowlight regions of enclosures causing foraging success to decline or zooplankton were concentrating in the low-light refuge, thus having the same effect on bluegill foraging.

Actively foraging larval fish can undertake relatively extensive diel vertical migrations (e.g., 10-30 m, Fortier and Leggett 1983; Wurtsbaugh and Neverman 1988; Munk et al. 1989) and appear to select optimal light levels. Increasing turbidity also tends to cause larval fish to migrate up in the water column, potentially to locate optimal light conditions for foraging (Swenson and Matson 1976; Matthews 1984; Boehlert and Morgan 1985). Because zooplankton also exhibit both diel vertical migration and migrate toward the surface as turbidity increases (Swenson and Matson 1976; Zettler and Carter 1986), the potential dissociation between prev concentrations and optimal search habitat for larval fish has not been considered. In experimental tanks, freshwater cyclopoid copepods tend to aggregate near the bottom even at night (Soto and Hurlbert 1991). Our data suggest that high turbidity and low light may create a refuge for zooplankton prey. The recent literature on diel vertical migration of zooplankton as a behavioral response to both invertebrate and vertebrate predators supports this refuge hypothesis (Ohman 1990; Ringelberg 1991; Bollens and Frost 1991).

Similar vertical distributions of larval herring and zooplankton, however, suggest synchronous habitat use (Fortier and Leggett 1983), although these observations were made over 10-m depth intervals. Because turbid conditions increase light attenuation rates, and thus compress light strata within shallow lakes (i.e., represented by our enclosures), zooplankton may concentrate within 50 cm of larval fish and still be within suboptimal habitat for fish foraging.

Zettler and Carter (1986) observed the progressive migration by zooplankters toward the surface along a turbidity gradient in a naturally turbid lake. In this deep lake (i.e., 50-60 m), they were able to show that many zooplankton concentrated in the top 5-10 m at high turbidity (14.5 NTU). However, using light attenuation rates generated from our system at 10.5 NTU in high light (see Table 1), light intensity at their high turbidity station was <300 lx at a depth of about 2.5 m. Thus, even with relatively fine-scale sampling (i.e., 5-m depth intervals in a system 60 m deep), in turbid systems it is difficult to determine if zooplankton concentrate within optimal habitat for larval fish foraging.

In high-light conditions, consumption of smaller zooplankton with increasing turbidity may be explained using predictions from optimal foraging theory (Stephens and Krebs 1986). Increasing turbidity at high light may increase prey contrast causing larvae to detect and consume smaller, and potentially more profitable, prey. Large prey are not always the most profitable in terms of net energy gain and growth (Bence and Murdoch 1986; Confer and Lake 1987) owing to increased handling time, poor capture efficiency, reduced assimilation efficiency, or increased gut passage time, as has been shown for early juvenile yellow perch (Perca flavescens, 22-62 mm TL) (Mills et al. 1986; Confer and O'Bryan 1989; Confer et al. 1990). Selection by yellow perch for cyclopoid and calanoid copepods and small *Daphnia* increases as prev density increases in high light whereas selection for large Daphnia declines. In fact, yellow perch grow faster feeding on small copepods than on Daphnia (Confer and Lake 1987). However, energy ingested per unit time does not differ between yellow perch feeding on copepods or Daphnia, in spite of the fact that foraging costs should be higher for copepods given their greater escape ability (Drenner et al. 1978). Given this, the observed increase in selection for smaller prey items (i.e., for copepod nauplii and smaller prey within taxa), and the fact that biomass consumed did not change at high light, suggests that turbidity effects on larval fish are complex.

Results from this experiment point to strong interactions between turbidity and light intensity on larval fish foraging and help to explain conflicting results from previous laboratory studies. Rather than directly reducing foraging ability of larval fish by decreasing reaction distance, turbidity appears to affect larval foraging through a reduction in light intensity and possibly prey contrast. At low light, increasing turbidity causes a reduction in biomass of prey consumed by larval bluegill; however, even at these low-light conditions, optimal light intensity (>450 lx) occurred in the upper 0.5 m of each enclosure. Thus, the precise mechanism explaining reduced consumption of prey needs to be determined by quantifying larval bluegill and zooplankton vertical distribution and prey consumption along the vertical, low-light gradient. However, previous laboratory studies and our field experiment clearly demonstrate that explanations for larval fish responses to perturbations (e.g., increasing turbidity in coral reef communities, Roberts 1991) cannot be directly extrapolated from studies of juvenile and adult fish. Because of rapid ontogenetic changes, larval fish cannot be considered miniature adults.

When turbidity reduces light intensity to threshold levels (<450 lx in our system), spatial overlap between predator and prey or prey-capture abilities appears to be reduced in shallow lakes, and larval bluegill growth may be inhibited. Because of highly size-dependent mortality from predation (Margulies 1990), even minor reductions in growth rate can have substantial effects on survival through this critical life stage.

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