

Research Paper

Circulating ectosomes: Determination of angiogenic microRNAs in type 2 diabetes

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Abstract

Ectosomes (*Ects*) are a subpopulation of extracellular vesicles formed by the process of plasma membrane shedding. In the present study, we profiled ectosome-specific microRNAs (miRNAs) in patients with type 2 diabetes mellitus (T2DM) and analyzed their pro- and anti-angiogenic potential.

Methods: We used different approaches for detecting and enumerating *Ects*, including atomic force microscopy, cryogenic transmission electron microscopy, and nanoparticle tracking analysis. Furthermore, we used bioinformatics tools to analyze functional data obtained from specific miRNA enrichment signatures during angiogenesis and vasculature development.

Results: Levels of miR-193b-3p, miR-199a-3p, miR-20a-3p, miR-26b-5p, miR-30b-5p, miR-30c-5p, miR-374a-5p, miR-409-3p, and miR-95-3p were significantly different between *Ects* obtained from patients with T2DM and those obtained from healthy controls.

Conclusion: Our results showed differences in the abundance of pro- and anti-angiogenic miRNAs in *Ects* of patients with T2DM, and are suggestive of mechanisms underlying the development of vascular complications due to impaired angiogenesis in such patients.

Key words: angiogenesis, diabetes, extracellular vesicles, microRNA, systems biology

Introduction

According to a global report on diabetes by the World Health Organization (2016), the global prevalence of diabetes mellitus (DM) has exceeded 400 million patients. Prevalence of DM is highest in the Southeast Asia and Western Pacific regions, with age-standardized mortality rates of 129.1 and 67.8 deaths per 100,000 people, respectively [1]. In these

regions, the majority of patients with diabetes have type 2 diabetes mellitus (T2DM). Despite substantial progress in the diagnosis and medical care of patients with diabetes, the life expectancy of these patients is significantly shorter than that of the general population [2]. Diabetes imposes a large financial burden on the global health-care system and human

capital. Vascular complications are the major cause of increased mortality and morbidity associated with DM. Functions of the cardiovascular system, including angiogenesis and disease states such as atherosclerosis, are seriously altered in hyperglycemic conditions, which is typically observed in patients with poorly controlled DM. Cardiovascular alterations often manifest as macroangiopathies, like coronary artery disease (CAD) and stroke, or as microangiopathies, like non-healing foot ulcers, retinopathy and diabetic nephropathy.

Of the different circulating extracellular vesicles (EVs), submicron-sized ectosomes (*Ects*) have not attracted as much interest as nano-sized exosomes have [3, 4]. However, after a study by Wolf (1967) describing the origin of *Ect*-like particles from platelets, numerous studies have investigated the possible mechanisms underlying *Ect* formation [5]. *Ects* belong to a large family of membrane vesicles or EVs, also referred to as microparticles (MPs), that are released in an evolutionarily-conserved manner in both prokaryotes and eukaryotes [6].

Ects retain many features of their parental cells, including specific surface antigens and receptors [7]. Therefore, characteristics of *Ects* are based on their size and their specific molecular signature, with some surface proteins and phosphatidylserine (PS) being the key identifiers of *Ects* [6]. PS-positive *Ects* can be easily distinguished from other circulating EVs through annexin V or lactadherin binding [8, 9].

In the blood, platelets and endothelial cells are the main source of circulating *Ects*; however, under stress conditions such as hypoxia, inflammation, and diabetes, other blood cells, including macrophages and neutrophils, release several EVs [10, 11]. Endothelial cell- and platelet-derived *Ects* substantially affect the cardiovascular system, especially endothelial cells and hemostasis [12-14]. Numerous studies suggest that endothelial cell-derived *Ects* exert an anti-vasorelaxing effect on other endothelial cells, promote hemostasis, and exert procoagulant effects [12, 15]. In contrast, platelet-derived *Ects* increase the adhesion of hematopoietic stem/progenitor cells to the endothelium, significantly improving their engraftment [16], and regulate cell migration and neovascularization [17, 18].

Changes in EV levels and heterogeneity have been observed in the circulating blood of patients with type 1 diabetes mellitus (T1DM) and T2DM. Although patients with T1DM and T2DM show an increase in overall *Ect* levels, patients with T1DM have a higher number of platelet-derived *Ects* than patients with T2DM. Moreover, the procoagulant activity of *Ects* is correlated with the levels of glycated hemoglobin (HbA1C) in patients with T1DM [19]. In

contrast, the procoagulant activity of *Ects* is lower in patients with T2DM having diabetic retinopathy than in those having CAD and diabetic foot syndrome [20].

Pioneering studies on EVs performed over a decade ago indicate that both exosomes and *Ects* contain RNA, including microRNAs (miRNAs); this finding has substantially renewed interest in EVs as mediators of cell-to-cell communication [21]. miRNAs regulate the transcription and translation of several genes involved in cell metabolism and various canonical and non-canonical pathways in all living cells [22]. In DM, modulation of miRNA expression is suggested to affect the production of insulin and occurrence of diabetic complications [23].

Accumulating evidence indicates that most circulating miRNAs are carried by AGO protein complexes or plasma proteins (lipoproteins) [24, 25]. EVs are suggested to function as alternative carriers that protect extracellular RNA from RNase degradation, indicating that they are efficient tools for transport in the human body [26, 27].

The present study examined miRNAs isolated from the plasma of patients with T2DM. In addition, this study determined miRNA signatures in *Ect*-enriched and *Ect*-depleted plasma fractions and analyzed the pro- and anti-angiogenic potential of these miRNAs by using bioinformatics and system biology tools.

Methods

Patients and sample collection

This study involved 15 patients with T2DM and 15 control subjects, both groups comprising male and female subjects aged between 68 and 82 years (mean age, 67.9 ± 5.23 years). Clinical characteristics of the study subjects are listed in **Table 1**. Inclusion criteria for diabetic patients and control subjects were similar to those described previously [28]. Diabetes was defined according to Polish Diabetes Society guidelines, based on oral glucose tolerance test (OGTT). All T2DM patients were undergoing diabetes treatment. Obesity and insulin resistance were not criteria for exclusion. Both patients and controls with acute coronary syndrome (ACS), acute ischemic stroke (IS) or critical limb ischemia were excluded from this study if they occurred within 6 months prior to the study enrollment. Other criteria for exclusion were patients with a history of cancer, renal and liver failure, and past or present systemic inflammation as defined by high-sensitivity C-reactive protein (hs-CRP) levels above 10 mg/L.

Blood samples from all the subjects were obtained at the same time of the day (between 08:00 and 10:00 A.M.) under fasting conditions by

performing venipuncture of the antecubital vein with a >21 gauge needle and by applying a light tourniquet. Approximately 2–3 mL blood was initially withdrawn but not included in the analysis. Blood for miRNA profiling was collected and mixed with an anticoagulant, sodium citrate, centrifuged twice at $2500 \times g$ for 15 min to obtain platelet-poor plasma (PPP). The blood samples were subsequently treated with an anticoagulant, EDTA, for performing hematological analysis and determining HbA1C levels. Biochemical and biomarker analyses were performed by collecting the blood samples in serum separator tubes. Plasma and serum samples were aliquoted, and frozen at $-80\text{ }^{\circ}\text{C}$ for further analyses. A schematic description of the procedure is presented in **Figure 1**.

Table 1. Characteristics of study groups

Variable	Type 2 diabetes (n=15)	Control group (n=15)	P
Gender: Female/Male (N)	1/14	0/15	NS
Age (years)	67 (65-71)	65 (63-71)	0.337
BMI (kg/m ²)	30.25 ± 3.76	25.23 ± 1.22	<0.0001
Current smokers (N/%)	1/6.7	0/0	0.34
Former smokers (N/%)	4/26.7	3/20	0.73
Alcohol consumption (N/%)	3/20	5/33.3	0.53
Duration of diabetes (years)	6 (3-12)	NA	-
Hypertension (N/%)	8/53.3	0/0	0.01
Other cardiac diseases (N/%)	1/6.7	0/0	0.34
Peripheral atherosclerosis (N/%)	2/13.3	1/6.7	0.58
Retinopathy (N/%)	1/6.7	0/0	0.34
Glucose (mM)	8.96 ± 3.17	6.11 ± 0.97	0.003
HbA1C (%)	7.5 (7.1-7.8)	NA	-
hs-CRP (mg/L)	0.81 ± 0.67	0.64 ± 0.87	0.56
TC (mM)	4.11 ± 0.87	4.89 ± 0.55	0.007
CHOL LDL (mM)	1.94 ± 0.64	2.95 ± 0.45	<0.0001
CHOL HDL (mM)	1.13 (1.05-1.26)	1.23 (1.15-1.27)	0.169
TG (mM)	2.17 (1.40-2.59)	1.36 (1.09-1.73)	0.022
Medications			
Aspirin (N/%)	5/33.3	2/13.3	0.31
Insulin (N/%)	5/33.3	NA	-
Metformin (N/%)	9/60	NA	-
Sulphonylurea (N/%)	6/40	NA	-
Hypotensive treatment (N/%)	9/60	NA	-
Statins (N/%)	5/33.3	1/6.66	0.13
Fenofibrates (N/%)	1/6.7	0/0	0.34

BMI: body-mass index; CHOL HDL: cholesterol high-density lipoprotein fraction; CHOL LDL: cholesterol low-density lipoprotein fraction; HbA1C: glycated haemoglobin A1C fraction; hs-CRP: high sensitivity C-reactive protein; TC: total cholesterol; TG: triglycerides.

Values are given as a mean ± standard deviation (SD), a median with an interquartile interval (Q1-Q3) or the number and percentage (%). Bold means statistically significant difference between the two groups at $p < 0.05$

Ethics committee approval and patient consent

The study complied with the Declaration of Helsinki, and the study protocol was approved by the Jagiellonian University Bioethics Committee (permission no. KBET/206/B/2013, extended until December 31, 2017) [28]. Informed consent was obtained from all the study subjects.

Determination of biochemical parameters

Biochemical parameters, including triglyceride, LDL-C, and HDL-C levels, were determined using MaxMat analyzer with ELITech Clinical Systems tests (Puteaux, France), with a limit of detection (LoD) of 0.06 mM. hs-CRP was determined using APTEC Ultra-Sensitive CRP test (APTEC Diagnostics NV, Belgium), with a LoD of 0.13 mg/L. HbA1C level was measured by performing HPLC with D-10 Hemoglobin Testing System (Bio-Rad Laboratories, Inc., Hercules, California, USA), certified by the National Glycohemoglobin Standardization Program (NGSP) organization.

Ect preparation from plasma samples

PPP was thawed in a water bath at $37\text{ }^{\circ}\text{C}$ to prevent cryoprecipitation, mixed, and centrifuged at $16,000 \times g$ and $4\text{ }^{\circ}\text{C}$ for 90 min to obtain *Ect*-enriched and *Ect*-depleted plasma fractions, according to Zheng et al., with some modifications including long $16,000 \times g$ centrifugation [29]. The *Ect*-enriched plasma fraction was frozen at $-80\text{ }^{\circ}\text{C}$ for performing subsequent analyses (**Figure 1**).

Extracellular vesicle (Ect) detection

Atomic force microscopy

Atomic force microscopy (AFM) images of the *Ect*-enriched and *Ect*-depleted plasma fractions were obtained using NanoWizard (JPK Instruments AG, Berlin, Germany). All experiments were performed in distilled water using a commercial liquid cell (JPK Instruments, Berlin). V-shape gold-coated cantilevers (MLCT cantilever; Bruker Nano Inc., Billerica, MA, USA) with nominal spring constants of 0.01 N/m were used for scanning in fluid. Surface images (256×256 pixels) were obtained from a scan size of $10 \times 10\text{ }\mu\text{m}$ and a scan rate of 0.4 Hz. Approximately 10–15 images of the surface were obtained for each sample. The size, cross-section, height, and diameter of all the EVs, including *Ects*, were calculated using JPK software (JPK Instruments).

Cryogenic transmission electron microscopy

Cryogenic transmission electron microscopy (Cryo-TEM) of the *Ect*-enriched plasma fraction was performed using a Tecnai F20 TWIN microscope (FEI Company, USA) equipped with a field emission gun and operated at an acceleration voltage of 200 kV. Micrographs were recorded using an Eagle 4k HS camera (FEI Company) and processed using TIA software (FEI Company). Specimens were prepared by vitrifying a particle aqueous solution on grids with a holey carbon film (Quantifoil R 2/2; Quantifoil Micro Tools GmbH, Jena, Germany). The grids were activated for 15 s in oxygen/argon plasma by using

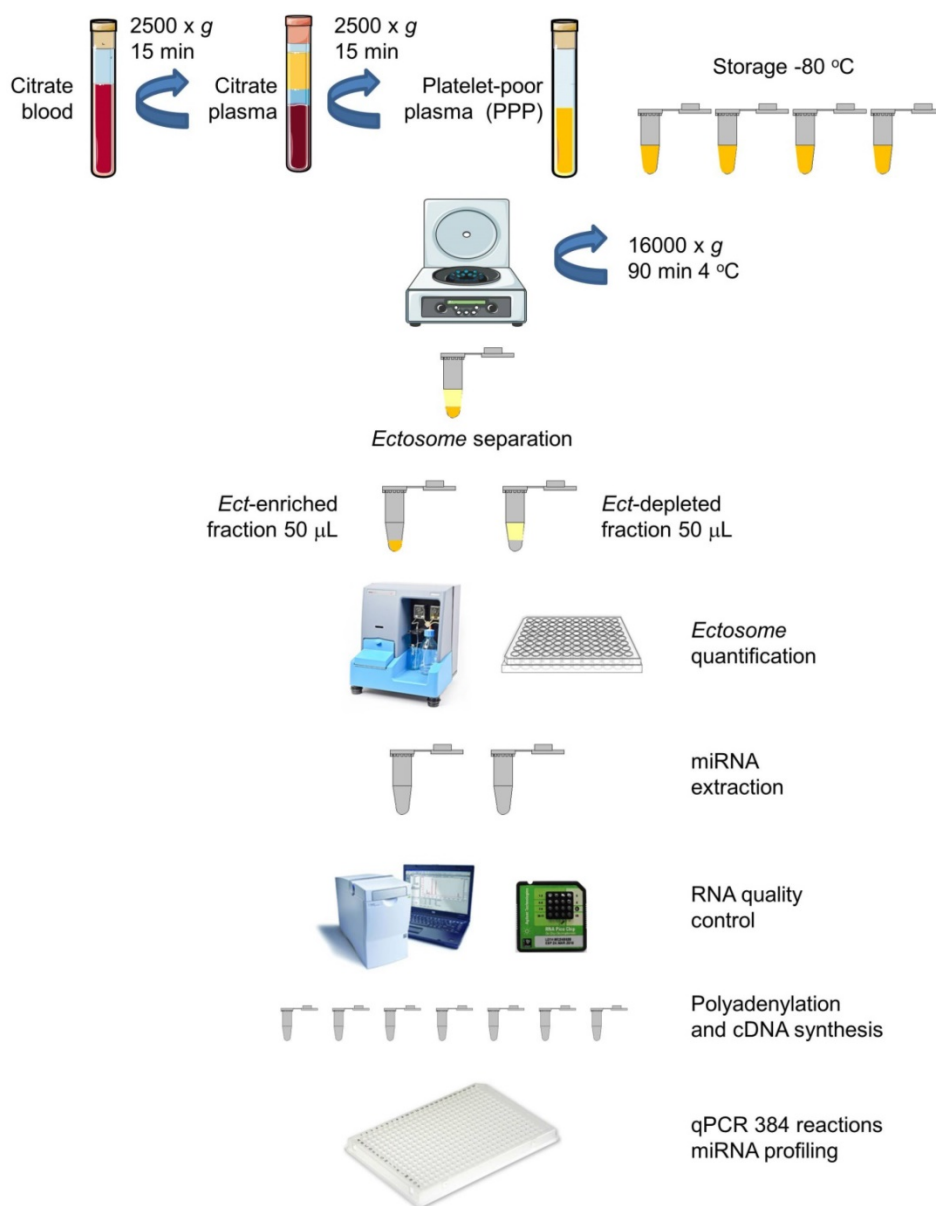


Figure 1. Circulating Ects and plasma supernatant: workflow for miRNA extraction and analysis. Platelet-poor plasma (PPP) was harvested from blood samples of patients by centrifugation. The plasma was aliquoted in Eppendorf tubes (350 µL) and was stored at -80°C . Ects were separated by centrifugation, and two plasma fractions, namely, *Ect*-enriched and *Ect*-depleted plasma fractions, were used for further analyses (adapted from [30]).

Fischione 1020 plasma cleaner (E.A. Fischione Instruments, Inc., Export, PA, USA). Samples were prepared by applying a droplet ($2\ \mu\text{L}$) of the solution onto the grid, blotting with a filter paper for 2 s, and immediately freezing in liquid ethane using a completely automated blotting device (Vitrobot Mark IV; FEI Company, Hillsboro, OR, USA). The vitrified specimens were kept in liquid nitrogen until they were inserted into a Cryo-TEM holder (Gatan 626; Gatan Inc., Pleasanton, CA, USA) and analyzed with a TEM at -178°C .

Transmission electron microscopy

The *Ect*-enriched plasma fraction was washed with PBS and centrifuged at $18,000 \times g$ (20 min, 4°C). Supernatant was discarded and pelleted ectosomes

were fixed with 2.5% glutaraldehyde in 0.1 M cacodylic buffer (CB) overnight at 4°C . Then, samples were washed four times for 15 min with 0.1 M CB and then postfixed for 1 h in 1% osmium tetroxide/0.1 M CB solution. Subsequently, samples were dehydrated by passing through a graded ethanol series (50%, 70%, 80%, 90%, and 100%, 5 min each), embedded in PolyBed 812 (epoxy resin) and polymerized for 48 h at 60°C . Ultrathin sections were prepared with an EM UC7 Leica ultramicrotome and collected on a copper mesh; the latter was covered with formvar film. Finally, the sections were contrasted with the use of uranyl acetate and lead citrate, and viewed on a JEOL JEM 2100HT TEM at 80 kV accelerating voltage.

Extracellular vesicle (Ect) enumeration

Nanoparticle tracking analysis

The samples were thawed on ice and were vortexed immediately before performing nanoparticle tracking analysis (NTA). To prevent EV aggregation, all the samples were diluted to 100× in HEPES buffer (10 mM HEPES/NaOH, 140 mM NaCl, and 2.5 mM CaCl₂, pH 7.4). NTA was performed using a NanoSight NS500 (Malvern Instruments Ltd., Malvern, United Kingdom) equipped with a sample chamber and a 405 nm laser. The assay was performed at room temperature (23.3 °C ± 0.1 °C), and particles present in the samples were visualized based on light scattered by them when illuminated by the laser light. Data were captured and analyzed using NTA 2.3 Build 0025 software (NanoSight NS500, Malvern Instruments Ltd., Malvern, United Kingdom). All the samples were analyzed for 30 s, with manual shutter and gain adjustments in advanced settings. Mean squared displacement of each particle tracked was measured simultaneously to determine the particle diffusion coefficient (D_t), sphere-equivalent, and hydrodynamic radius (R) using the Stokes–Einstein equation $D_t = K_b T / 6\eta R\pi$, where K_b is Boltzmann's constant, T is temperature, η is solvent viscosity, and R is the hydrodynamic radius of the particle. Thus, NTA helped in measuring both the size and concentration of EVs.

Analysis of the procoagulant activity of Ects

Ect concentration was determined by performing Zymuphen MP Activity assay based on the procoagulant activity of the human plasma (through thrombin generation) by using a manual method (cat. no. 521096; Hyphen BioMed SAS, Neuville sur Oise, France), and was expressed as MP concentration (in nM). The assay was developed in an end-point mode. Absorbance was measured at 405 nm, with a reference wavelength of 690 nm, by using Infinite F200 Pro plate

reader (Tecan Group Ltd., Männedorf, Switzerland) at 3 min after substrate introduction at 37 °C and reaction termination. The detection threshold was set at <0.05 nM, and intra-assay variability was set at <8%.

miRNA profiling

miRNA extraction

miRNAs were extracted from the *Ect*-enriched and *Ect*-depleted plasma fractions by using miRNeasy Serum/Plasma Kit (Qiagen) and Trizol LS Reagent (Invitrogen). RNA yield and concentration were determined by performing capillary electrophoresis with Agilent Bioanalyzer 2100 and Eukaryote Total RNA PicoChip (Agilent Technologies, Inc., Santa Clara, CA, USA). An average of 144 ± 89 and 42.6 ± 45.9 pg/μL total RNA was recovered from 50 μL of *Ect*-enriched and *Ect*-depleted plasma fractions, respectively. Electropherograms of miRNAs present in *Ect*-enriched and *Ect*-depleted plasma fractions are presented in **Figure 2**.

miRNA analysis by quantitative PCR

Extracted miRNA from the *Ect*-enriched fraction and supernatants was reverse transcribed using a polyadenylation and cDNA synthesis kit (Exiqon). Before cDNA synthesis, RNAs were spiked with a synthetic miRNA (UniSp6, Exiqon) that served as a control for cDNA synthesis. Screening for differentially-expressed miRNAs in ectosomes was performed by using a low-density qPCR array, Serum/Plasma miRCURY LNA PCR Panel V1 (Exiqon) (plate layout shown in **Table S1**) in a Vii7 real-time PCR system (Thermo Fisher Scientific, Waltham, MA, USA) equipped with a 384-well reaction plate. qPCR was performed using SYBR Green Master Mix Universal RT (Exiqon) under the amplification conditions recommended by the manufacturer. Amplification data were processed by

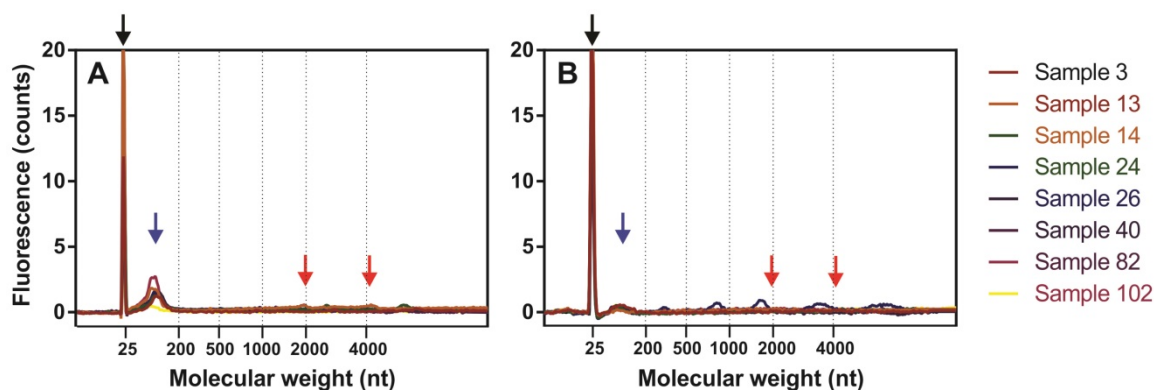


Figure 2. Fluorescence profiles of isolated miRNAs determined by capillary electrophoresis using Agilent 2100 Bioanalyzer PicoChip. Fluorograms were merged together according to a nucleotide size axis. (A) *Ect*-enriched and (B) *Ect*-depleted plasma fractions. For comparison, miRNAs were isolated from the same samples by using the miRNeasy Serum/Plasma Kit. Black arrow indicate marker dye, blue arrows indicate short RNA fragments (miRNAs), and red arrows indicate 18S and 28S rRNAs.

the ΔC_T method with a global normalization approach using an open source DataAssist 3.0 software (Thermo Fisher Scientific). Fold changes were calculated by the $\Delta\Delta C_T$ method, and statistically significant variations among different groups were determined by calculating adjusted p-values using the Benjamini-Hochberg false discovery rate procedure. Specific candidate miRNAs were chosen from screening results and individually validated in an additional cohort of patients by qPCR using specific LNA primers (Exiqon) along with seven additional pro- and anti-angiogenic miRNAs from previously described studies (Table 2). For individual qPCR validation, we performed three technical replicates per sample. Amplification data were also processed using the ΔC_T method using the synthetically spiked miRNA as a normalization control.

Analysis of miRNA targets and functional interactions

Validated miRNA-target relationships were determined using miRNet database [31]. Functional interactions between proteins were predicted using STRING database by considering a global interaction score of >0.9 [32]. A graphical network showing significant miRNAs and functionally corresponding genes was constructed using NAViGaTOR software [33].

Statistical analysis

Statistical analyses were performed using Statistica 12.0 package (StatSoft Polska, Kraków, Poland). Distribution of variables was examined using Kolmogorov-Smirnov normality test. Quantitative variables were characterized using descriptive statistics, i.e., mean and standard deviation (SD) for normally distributed data or

median and interquartile interval (Q1-Q3) for non-normally distributed data. Qualitative variables were compared using two-sided Chi-square and Fisher's exact tests. Student's *t*-test (normally distributed data) or Mann-Whitney *U* test (non-normally distributed data) was used to determine significant differences between variables. Statistical significance was set at $p < 0.05$. For the PCR-array analysis (volcano plots and heat maps), a cut-off for level of fold-change of >2 and p -value < 0.05 were established. Additionally, to reveal the role of classical cofounders in diabetes (age, BMI, arterial hypertension and diabetes duration) the authors applied a logistic regression model in miRNA analysis based on the median miRNA expression value (1 for higher, 0 for lower values).

Results

Characterization of plasma-derived Ects

AFM and Cryo-TEM detected *Ects* and smaller EVs (exosomes) in the *Ect*-enriched and *Ect*-depleted plasma fractions (Figure 3 and Figure 4). AFM showed the abundance of large vesicles such as *Ects* (mean height, ~200 nm) in the *Ect*-enriched fraction obtained by performing high-speed centrifugation (Figure 3). The average height of the identified vesicles in the *Ect*-depleted fraction was lower than that in the *Ect*-enriched fraction (Figure 3C). Cryo-TEM detected *Ects* in the *Ect*-enriched plasma fraction obtained from patients with uncontrolled (Figure 4B) and controlled diabetes (Figure 4C). *Ects* were detected as typical multilayered vesicles with a dense cargo.

Table 2. Target sequences and nomenclature of selected miRNAs analyzed in the study. Accession sequences and names are presented according to miRBase 21.

miR name (human)	microRNA target sequence	Sequence reference	LNA™ microRNA PCR primer set
hsa-miR-126-3p	UCGUACCGUGAGUAAUAAUGCG	MIMAT0000445	EQ-204227
hsa-miR-126-5p	CAUUUUUUUUUUUGGUACGCG	MIMAT0000444	EQ-206010
hsa-miR-193b-3p	AACUGGCCUCAAAGUCCCGCU	MIMAT0002819	EQ-204226
hsa-miR-199a-3p	ACAGUAGUCUGCACAUUGGUUA	MIMAT0000232	EQ-204536
hsa-miR-20a-3p	ACUGCAUUUAUGAGCAUAAAAG	MIMAT0004493	EQ-204052
hsa-miR-221-3p	AGCUACAUUGUCUGCGGGUUUC	MIMAT0000278	EQ-204532
hsa-miR-23b-3p	AUCACAUUGCCAGGGAUUACC	MIMAT0000418	EQ-204790
hsa-miR-26a-5p	UUCAAGUAAUCCAGGAUAGGCU	MIMAT0000082	EQ-206023
hsa-miR-26b-5p	UUUAUUUAACAACCGAUUAGUG	MIMAT0000083	EQ-204172
hsa-miR-29a-5p	ACUGAUUUUUUUUGGUGUUCAG	MIMAT0004503	EQ-204430
hsa-miR-30b-5p	UGUAAACAUCUACACUCAGCU	MIMAT0000420	EQ-204765
hsa-miR-30c-5p	UGUAAACAUCUACACUCAGCU	MIMAT0000244	EQ-204783
hsa-miR-374a-5p	UUUAUUUAACAACCGAUUAGUG	MIMAT0000727	EQ-204758
hsa-miR-409-3p	GAAUGUUGCUCGGUGAACCCCU	MIMAT0001639	EQ-204358
hsa-miR-495-3p	AAACAAACAUGGUGCACUUCUU	MIMAT0002817	EQ-206015
hsa-miR-95-3p	UUCAACGGGUUUUUUUGAGCA	MIMAT0000094	EQ-204288
hsa-let-7i-5p	UGAGGUAGUAGUUUGUGUGUU	MIMAT0000415	EQ-204394

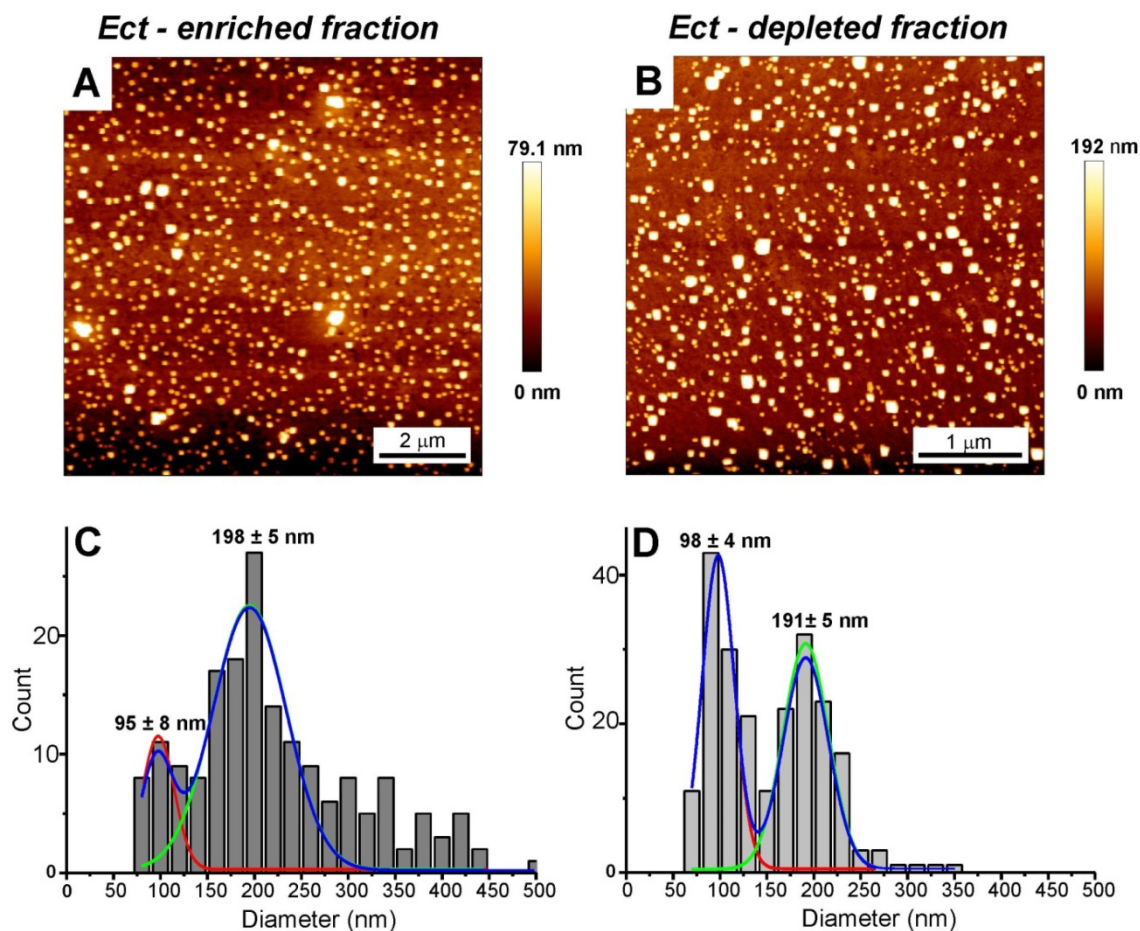


Figure 3. AFM and analysis of EVs in the Ect-depleted (left) and Ect-enriched (right) plasma fractions from a representative patient with T2DM. Samples were immobilized on poly-L-lysine-coated slides. AFM images of EVs. Presence of large vesicles was confirmed in the Ect-enriched (A) and depleted plasma fractions (B). (C-D) The mean value of the height of the observed structures calculated based on AFM images. Appropriate size distribution histograms of immobilized EVs in both the plasma fractions showing the presence of Ect populations. Diameters are expressed as mean \pm standard error.

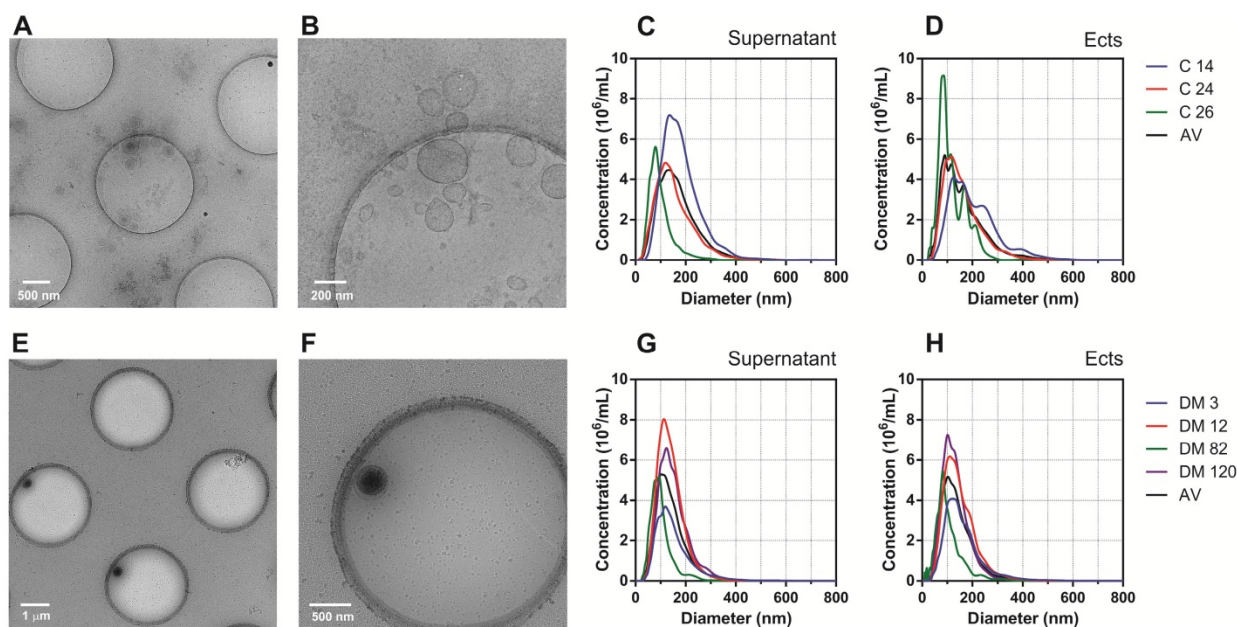


Figure 4. Characteristics of Ects present in the Ect-enriched and Ect-depleted plasma fractions. Representative wide-field and magnified cryo-TEM images of Ects isolated from a control subject (A-B) and a patient with diabetes (glucose level: 12.02 mM; HbA1C level: 9.2%) (E-F). Representative plots of NTA size distribution of EVs in Ect-enriched and Ect-depleted plasma fractions from the control group (C-D) and diabetic patients (G-H). Samples were analyzed in 1000 \times dilutions.

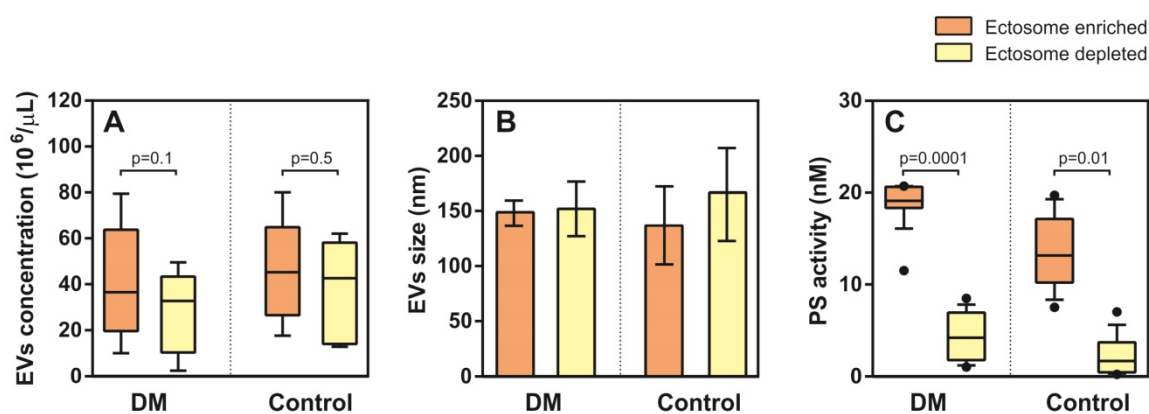


Figure 5. Enumeration and characteristics of Ects present in the Ect-enriched and Ect-depleted plasma fractions. (A) Number of microvesicles present in the Ect-enriched and Ect-depleted plasma fractions obtained from diabetic patients (DM) compared with that in the Ect-enriched and Ect-depleted plasma fractions obtained from healthy controls (Control), as determined by performing nanoparticle tracking analysis (NTA). **(B)** Ects particle size as determined by NTA in both groups of samples. **(C)** PS (phosphatidylserine) procoagulant activity exposed on EVs as quantified by Zymuphen MP assay. Whisker plots in (A) and (C) represent data as median (box) with interquartile interval (10-90) showing the outliers. Depicted p-values in (A) and (C) between the studied groups of patients were calculated by a two-tailed non-parametric Mann-Whitney test.

NTA analysis did not show significant differences in EV density (concentration) between T2DM and controls in the Ect-enriched fraction: 36.5 (19.7-63.7) 10⁶/μL vs. 45.2 (29.5-59.8) 10⁶/μL ($p = 0.8$); or in the Ect-depleted fraction: 33.7 (10.3-43.3) 10⁶/μL vs. 42.7 (14.2-20.8) 10⁶/μL ($p = 0.2$) (Figure 5A). Similarly, differences in EVs were detected: 148 (144-152) nm vs. 137 (130-191) nm ($p = 0.6$); and 152 (143-160) nm vs. 165 (100-183) nm ($p = 0.9$) (Figure 5B). Significant difference in PS-exposed (procoagulant) concentrations between T2DM and controls were observed in the Ect-enriched fraction: 19.10 (18.34-20.59) nM vs. 13.15 (10.31-17.01) nM ($p = 0.0001$); as well as in the Ect-depleted fraction: 4.21 (2.05-6.81) nM vs. 1.68 (0.48-3.72) nM ($p = 0.01$) (Figure 5C).

Ectosomes from patients with T2DM showed a specific miRNA enrichment signature

miRNA content in Ects obtained from patients with T2DM and healthy controls was determined by performing low-density qPCR array with LNA primers against 179 circulating miRNAs (Table S1). miRNA levels were compared with those in corresponding paired supernatants obtained during Ect purification by using the same approach (Figure 6). We identified 25 Ect-specific miRNAs in samples obtained from healthy controls and 28 Ect-specific miRNAs in samples obtained from patients with T2DM. Of the 28 miRNAs identified in samples obtained from patients with T2DM, 10 miRNAs, namely, hsa-miR-20a-3p, hsa-miR-26a-5p, hsa-miR-26b-5p, hsa-miR-29a-5p, hsa-miR-374a-5p, hsa-miR-30b-5p, hsa-miR-30c-5p, hsa-let-7i-5p, hsa-miR-199a-3p, and hsa-miR-221-3p, were exclusively present in Ects isolated from patients with T2DM (Figure 7A).

To determine the possible functional role of miRNAs present in Ects obtained from patients with

T2DM, we performed a validated target analysis followed by graphical network integration, including protein-protein interactions (Figure 7B). Our results showed that the examined miRNAs targeted genes involved in angiogenesis and vasculature development. Moreover, the examined miRNAs exerted a cooperative regulatory effect on several angiogenesis-related mRNA transcripts that were simultaneously targeted by two or more of the Ect-specific miRNAs. Four Ect-specific miRNAs, namely, hsa-miR-26a-5p, hsa-miR-26b-5p, hsa-miR-30b-5p, and hsa-miR-30c-5p, simultaneously targeted the mRNA transcripts of two genes involved in angiogenesis, namely, *MTDH* and *PDCD10*, in patients with T2DM. Interestingly, pro-angiogenic hsa-miR-30 family miRNAs and anti-angiogenic hsa-miR-26 family miRNAs simultaneously targeted *MTDH* and *PDCD10* mRNA transcripts. Three Ect-specific miRNAs targeted seven mRNA transcripts in patients with DM. Of these seven mRNA transcripts, *DDAH* and *GNA13* mRNA transcripts were regulated by pro-angiogenic miRNAs and *PRX*, *CYR61*, *NUS1*, *PTEN*, and *WNT5A* mRNA transcripts were regulated by anti-angiogenic miRNAs. Determination of Ect-specific miRNA signatures in patients with T2DM identified several previously characterized pro- and anti-angiogenic miRNAs involved in diverse physiological processes (Table 3).

Validation of Ect miRNA levels in patients with DM

We used two different approaches to validate Ect-specific miRNAs. The first approach involved a literature search to identify miRNAs involved in the pathogenesis of diabetes and its cardiovascular complications [23, 27, 34, 35], and the second approach involved analysis of qPCR data.

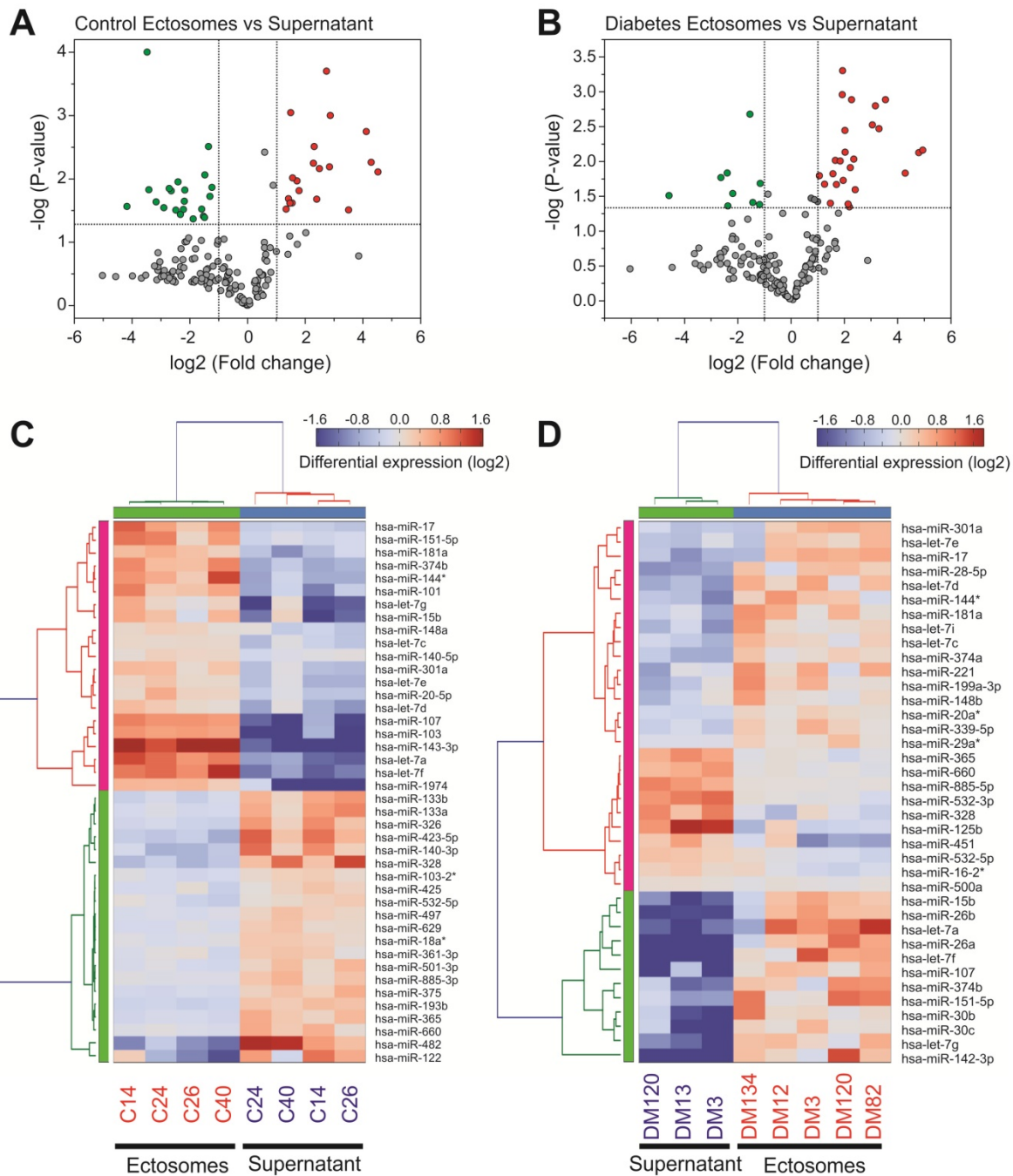


Figure 6. qPCR screening of enriched and depleted miRNAs present in *Ects* compared with those present in corresponding supernatants. Volcano plots of specific miRNAs present in the *Ect*-enriched plasma fraction compared with those present in corresponding supernatants obtained from (A) healthy controls and (B) patients with T2DM. Green and red dots represent significantly down- and upregulated miRNAs, respectively, taking into account the following thresholds: p-value < 0.05 and fold-change > 2. Heatmap for the differentially enriched miRNAs in *Ects* obtained from (C) healthy control and (D) patients with T2DM.

Table 3. Functional data of *Ect*-specific miRNAs involved in angiogenesis and vasculature development in patients with T2DM.

miRNAs	Function	Comments	References
hsa-miR-20a-3p	Nonrelated	The miRNA hsa-miR-20a-5p generated from the same pre-miRNA loop is anti-angiogenic	[67]
hsa-miR-26a-5p	Anti-angiogenic	miR-26a targets a SMAD1-Id1-p21 ^{WAF/CIP1} /p27 signaling axis and inhibits angiogenesis in endothelial cells. In contrast, pro-angiogenic stimuli such as VEGF or TNF-α reduced miR-26a expression in endothelial cells.	[68]
hsa-miR-26b-5p	Anti-angiogenic	Both in vitro and in vivo studies showed that miR-26b-5p could suppress vascular mimicry and angiogenesis by down-regulating the expression of VE-cadherin, Snail and MMP2 and could inhibit the apoptosis of hepatocellular carcinoma cells	[69]
hsa-miR-29a-5p	Nonrelated	The miRNA hsa-miR-29a-3p generated from the same pre-miRNA loop is anti-angiogenic	[70]

miRNAs	Function	Comments	References
hsa-miR-374a-5p	Nonrelated	The isomer miRNA hsa-miR-374b is anti-angiogenic	[71]
hsa-miR-30b-5p hsa-miR-30c-5p	Pro-angiogenic	Overexpression of hsa-miR-30 family in endothelial cells led to increased vessel number and length in an in vitro model of sprouting angiogenesis	[72]
hsa-let-7i-5p	Hypertension	Let-7 family targets the FGF5 transcript in microvascular endothelial cells	[73]
hsa-miR-199a-3p	Pro-angiogenic	MiR-199a-5p, promotes metastatic invasion, angiogenesis, and colonization in melanoma by targeting ApoE and the heat-shock factor DNAJA4.	[74]
hsa-miR-221-3p	Anti-angiogenic	Levels of miR-221/222 were significantly higher in coronary artery disease (CAD). MiR-221-c-kit pathway may play an important role in diabetes-associated vascular dysfunction	[75, 76]

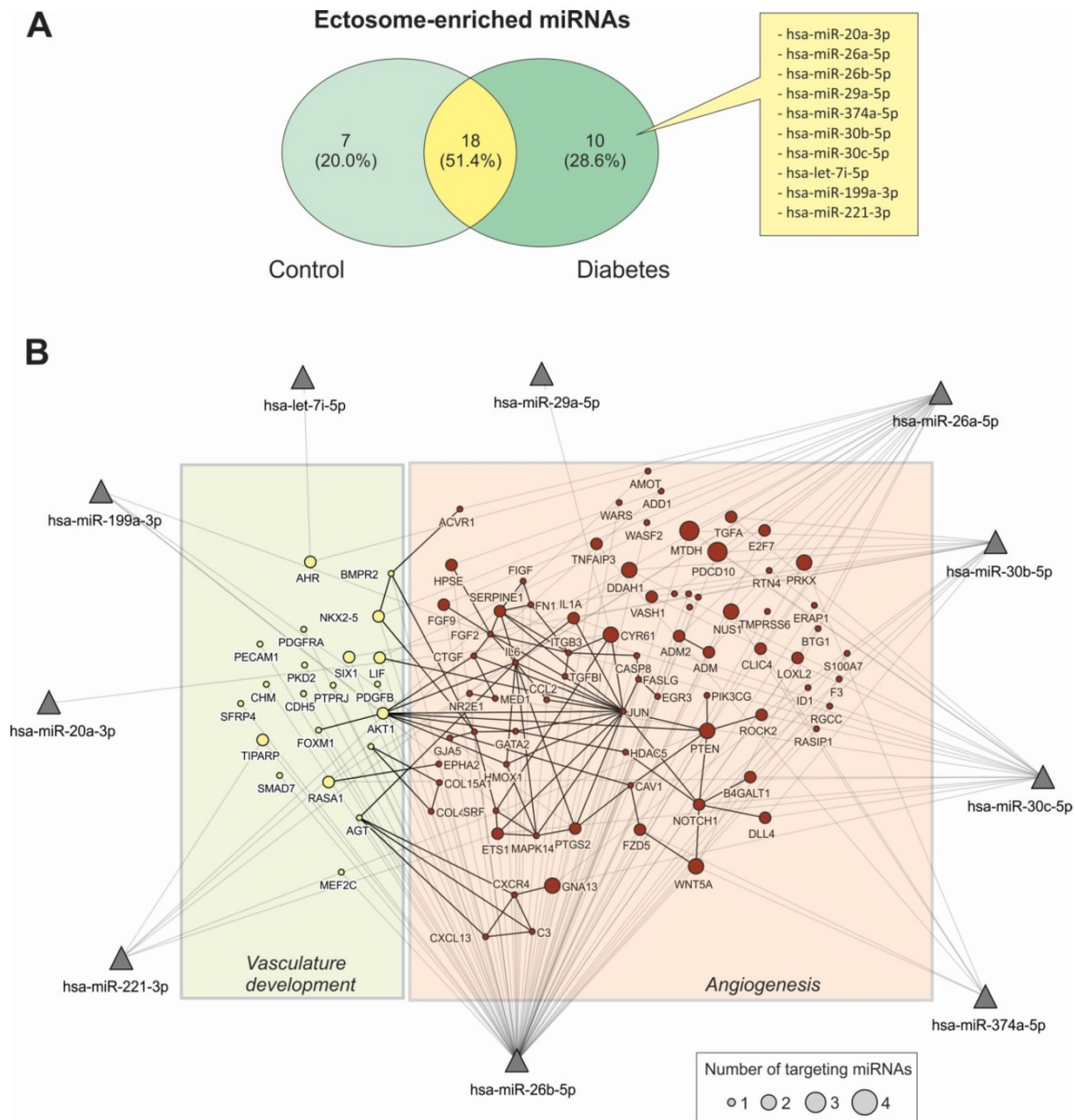


Figure 7. Functional characterization of Ect-enriched miRNAs in patients with T2DM. (A) Venn diagram representation of Ect-enriched miRNAs compared with those present in corresponding supernatants obtained from patients with T2DM and healthy controls. Patients with T2DM showed a specific miRNA enrichment pattern, which included 10 unique miRNAs not detected in healthy controls. (B) Integrative analysis of validated targets involved in vasculature development and angiogenesis for the 10 unique miRNAs enriched in Ect from T2DM patients. miRNAs are represented as filled triangles, and validated target genes are represented as circles, with a size proportional to the number of miRNAs regulating a particular mRNA. miRNA–target gene relationships obtained from the miRNet database [31] are indicated using grey lines. Black lines indicate functional interactions between proteins, as determined using the STRING database [32] by using a global interaction score of >0.9. The graphical network representation was constructed using the NAViGaTOR software [33].

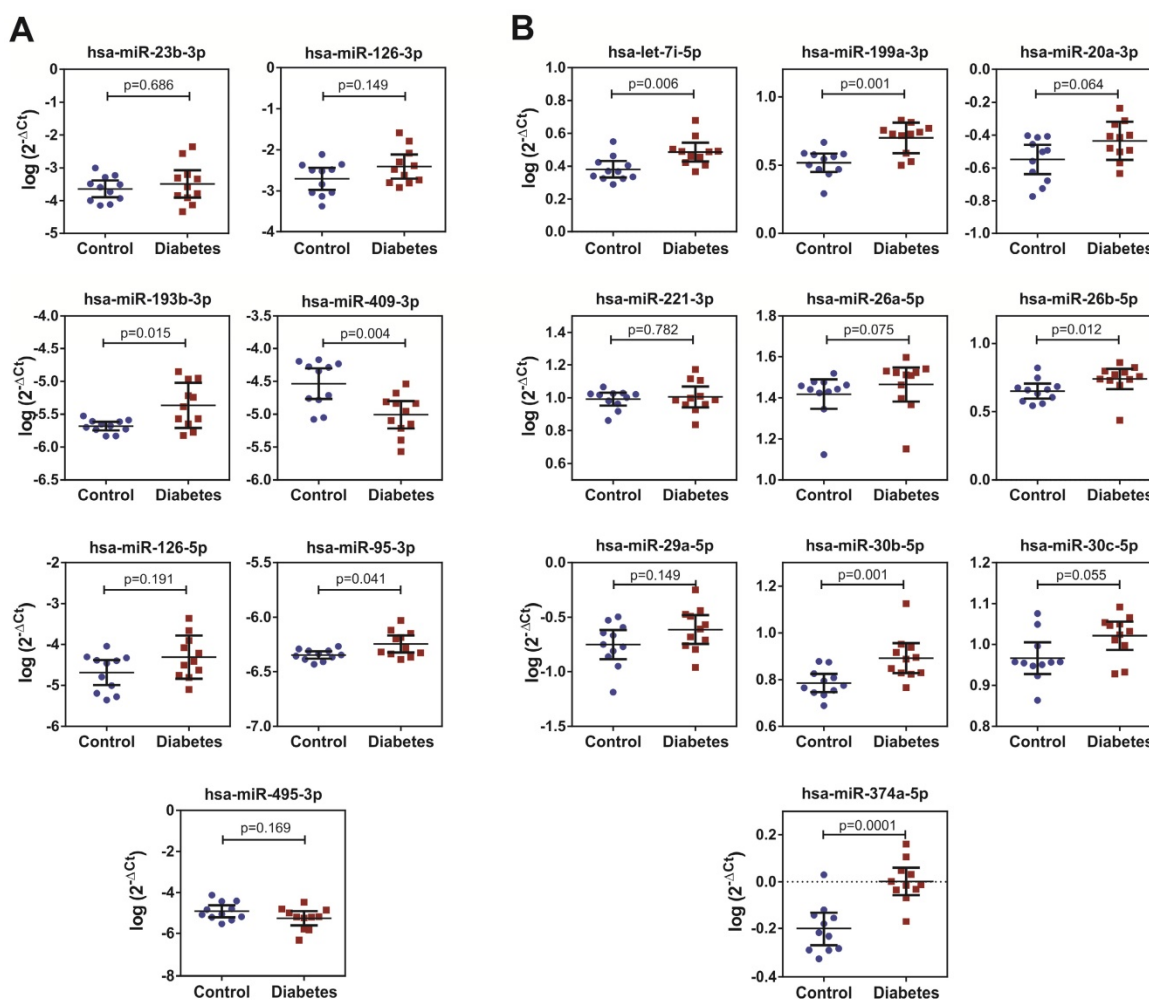


Figure 8. Levels of the selected *Ect*-specific miRNAs in 11 patients with T2DM and 11 healthy controls, as determined by performing qPCR with specific LNA primers. (A) Pro- and anti-angiogenic miRNAs selected by performing literature mining. (B) miRNAs exclusively enriched in *Ect*s obtained from patients with T2DM and selected by performing qPCR screening. Depicted p-values between the studied groups of patients were calculated by a two-tailed non-parametric Mann-Whitney test.

We performed qPCR to validate the levels selected by miRNA screening in the *Ect*-enriched plasma fractions obtained from 11 healthy controls and 11 patients with DM. Results of qPCR-based validation are shown in Figure 8. Of the reference miRNAs identified by performing a literature search, hsa-miR-193b-3p and hsa-miR-95-3p were significantly enriched in the *Ect*-enriched plasma fraction obtained from patients with T2DM ($p = 0.015$ and $p = 0.041$, respectively; Figure 8A). Interestingly, hsa-miR-409-3p was significantly downregulated in the *Ect*-enriched plasma fraction obtained from patients with T2DM ($p = 0.004$). Of the *Ect*-enriched miRNAs selected by performing miRNA screening, levels of five among them were significantly increased, namely, has-let-7i-5p ($p = 0.006$), hsa-miR-199a-3-5p ($p = 0.001$), hsa-miR-26b-5p ($p = 0.012$), hsa-miR-30b-5p ($p = 0.01$) and hsa-miR-374a-5p ($p = 0.0001$). Another three of the selected miRNAs had a tendency to increase: has-miR-20a-3p ($p = 0.064$), has-miR-26a-5p ($p = 0.075$) and

hsa-miR-30c-5p ($p = 0.055$) in the *Ect*-enriched plasma fraction obtained from patients with T2DM (Figure 8B).

The role of confounding factors on miRNA levels in the *Ect*-enriched fraction

Age and hypertension are the most confounding factors in D2TM. To reveal the role of confounding factors in diabetes incidence, the logistic regression model was applied. We found that none of the classical risk factors interfered significantly with analyzed miRNA levels in diabetic patients (Table S2).

Discussion

In the present study, we characterized the differences in miRNA cargo between *Ect*s isolated from T2DM patients and healthy controls by various methods. To show their molecular profile and physical properties, technical and experimental requirements were used to attribute their miRNA

cargo [3]. The novelty of the present study is the use of the array approach to quantify and discriminate *Ects*-enriched miRNAs in diabetic patients.

Origin and characteristics of circulating *Ects* in T2DM

Most *Ects*, especially those released by platelets and endothelial cells, are spherical in nature. *Ects* are not formed through exocytosis but are formed by outward membrane budding due to the loss of calcium-dependent membrane phospholipid asymmetry, cytoskeletal rearrangement, and cell surface shedding [36]. Cryo-TEM allowed us to detect very small structures ranging from ~5 to 200–300 nm in size, even in aqueous samples in a virtually artifact-free manner. In the present study, plasma samples were placed on a grid, and the small liquid film was rapidly frozen to 90 K to prevent the formation of ice crystals and maintain the transparency of the frozen water film in an amorphous state. We confirmed the multilayered structure of large microvesicles (*Ects*) encapsulating small vesicles, which was not abundant in the control sample. This finding concurs with previous observations by Isseman *et al.* in their Cryo-TEM study [37]. Additionally, we did a TEM imaging analysis that allowed us to characterize average EV size within a similar range to that in the cryo-TEM study. Nevertheless, the direct measurement of diameter by means of TEM is unreliable after the additional washing steps during sample preparation, which has been discussed elsewhere [6]. In the present study, we performed additional TEM imaging to visualize circulating *Ect* morphology in the plasma (**Figure S1**).

In the plasma, platelets are considered as the main source of EVs [10]. Cytometric analysis of plasma obtained from patients with T2DM showed that EVs originate from other cellular sources, of which most have been previously characterized as being CD36 positive [38]. Both *Ects* and apoptotic bodies are released by T lymphocytes under stress conditions, and these vesicles induce vascular hyperactivity in response to vasoconstrictors [39]. Endothelial cell-derived *Ects* impair endothelial function *in vitro* by decreasing acetylcholine-induced vasorelaxation and nitric oxide production [12]. Results of a Matrigel assay performed in a previous study showed that endothelial cell-derived *Ects* affect the formation of vascular networks by decreasing total capillary length, mesh number, and branching points, and increasing mesh area, thus confirming their anti-angiogenic role [40]. In contrast, platelet-derived *Ects* strongly activate angiogenesis by promoting the proliferation, survival, migration, and

tube formation by human umbilical vein endothelial cells [17]. Increasing evidence indicates that EVs (mainly exosomes) derived from the platelet-rich plasma promote the re-epithelization of chronic cutaneous wounds by effectively inducing proliferation and migration of endothelial cells and fibroblasts in a diabetic rat model [41].

We can conclude that differences in *Ects* characteristics and biological effects result from not only their different origins but also their pathology.

Specific miRNA signature in circulating *Ects*

The choice of the high-speed centrifugation technique for EV purification facilitates the separation of two fractions of PPP: *Ect*-enriched and *Ect*-depleted fractions. This is a method of choice used previously in a number of studies with different modifications [13, 29, 42]. In the *Ect*-enriched fraction, two types of ectosomes are usually abundant: those of platelet origin, and those of endothelial origin.

Platelet-derived *Ects* deliver various extracellular miRNAs to different vascular cells [27]. Activated platelets and platelet-derived microvesicles contain miRNAs and other non-coding RNAs [43]. Of these RNAs, two miRNAs, namely, hsa-miR-223-3p and hsa-miR-126-3p, are interesting because of their biological role and proven abundance in platelets [44–46]. miR-223-3p is highly expressed in platelets and megakaryocytes. Expression of hsa-miR-223-3p is strongly correlated with platelet response to antiplatelet therapy, and high miR-223 levels predict efficacious platelet inhibition with clopidogrel [44, 47]. In the present study, qPCR screening did not detect significant abundance of hsa-miR-223-3p in the *Ect*-enriched plasma fraction, and in patients with T2DM or healthy controls. However, some studies have shown that platelet-derived microvesicles deliver miR-223 as an effector complex along with miRNA machinery protein AGO2 and affect the expression of FBXW7 and EFNA1 mRNA transcripts [46]. In our previous studies, we have confirmed the presence of AGO proteins in larger platelet vesicles corresponding to *Ects*, obtained from activated platelets (unpublished data).

In the present study, mature miRNAs generated from the miR-126 precursor locus, i.e., hsa-miR-126-3p and hsa-miR-126-5p, were not significantly regulated in the *Ect*-enriched plasma fraction obtained from patients with T2DM. Our data differ from those of a previous study by Jansen *et al.*, who showed decreased expression of circulating hsa-miR-126-3p in patients with DM [42]. This discrepancy might be because of heterogeneity between the present study and the previous study with respect to the inclusion of patients with different DM types and absence of

information on treatment regimen and duration. In contrast, another study showed that circulating miR-126 levels are elevated in patients with T2DM and are positively correlated with glycemic control measure (glucose AUC) [48], which is consistent with the present study. miR-126 is expressed in platelets and endothelial cells, and circulating miR-126 is carried by both platelet- and endothelial cell-derived microvesicles. However, results of sorting experiments showed that most miR-126 is transferred by endothelial cell-derived *Ects* [27, 49]. These data indicate that miR-126 expression can be upregulated and downregulated in cardiovascular diseases and suggest that both platelet activity and endothelial cell dysfunction affect miR-126 availability [45]. Similarly, the level of another miRNA (hsa-miR-495-3p), which is suggested to be associated with DM complications, was not increased in the *Ect*-enriched plasma fraction in the present study.

Moreover, we observed that hsa-miR-193b-3p and hsa-miR-95-3p expression was significantly upregulated in the *Ect*-enriched plasma fraction obtained from patients with T2DM. To the best of our knowledge, no study has reported the involvement of miR-193 or miR-95 in angiogenesis regulation and vascular function, to date. Increased expression of miR-95 has been observed in cancer cells, and its activity is associated with radioresistance (or radiosensitivity) of non-small cell lung cancer [50]. miR-95 functions as an oncogene by directly targeting sorting nexin 1 and other proteins involved in cell proliferation [51]. In contrast to hsa-miR-95-3p, which stimulates cell proliferation, hsa-miR-193b exerts an anti-proliferative effect [52]. Moreover, accumulating evidence suggests that miR-193b inhibits the migration and invasion of cancer cells [53]. In the present study, we found that these two miRNAs were highly expressed in *Ects* obtained from patients with T2DM, suggesting that pro- and anti-proliferation processes are dysregulated in T2DM and that *Ects* contribute to the regulation of these processes.

Interestingly, hsa-miR-409-3p expression was downregulated in the *Ect*-enriched plasma fraction obtained from patients with T2DM. Previous studies indicate that cancer-associated stromal-like cells release miR-409 through EVs and promote epithelial-to-mesenchymal transition (EMT) both *in vitro* and *in vivo* [54]. EMT is a transdifferentiation process responsible for several clinical complications in T2DM. In experimental hyperglycemia, TGF- β signaling pathway mediates endothelial-to-mesenchymal transition (EndMT), which induces endothelial damage and increases mesenchymal biomarker expression [55]. The results of the present study suggest that hsa-miR-409-3p regulates EndMT

in T2DM.

In addition, we compared the distribution of circulating miRNAs between *Ect*-enriched and *Ect*-depleted plasma compartments in healthy controls and patients with T2DM. Our results showed specific distribution patterns of miRNAs between both these compartments in healthy controls and patients with T2DM. miRNA enrichment patterns in *Ects* isolated from patients with T2DM compared with those present in corresponding supernatants indicated the presence of pro- and anti-angiogenic miRNAs, which were different from that observed in healthy controls. Specific exosomal miRNA transfer in the context of angiogenesis has already been described in the communication between cardiomyocytes and endothelial cells mediated by miR-320 in rat models of DM [56].

Our results suggest an imbalance in the distribution of miRNAs in patients with T2DM, which may be associated with the physiological role of these miRNAs. This unique molecular signature may vary in different DM types showing different progression and associated complications [23, 42, 48], suggesting the role of genetics in *Ect* formation. However, one should consider environmental factors and baseline characteristics of patients (disease duration, age, weight, and gender). Moreover, relative tissue specificity, expression, and stability of different miRNAs significantly affect miRNA expression.

Role of *Ect*-enriched miRNAs in the regulation of angiogenesis

We found that both pro- and anti-angiogenic miRNAs were upregulated in *Ects* and that the miRNA expression pattern was associated with various angiogenic genes. As proven, *Ect*-enriched miRNAs in patients with T2DM included miRNAs that exerted antagonistic effects on angiogenesis and vasculature development (Table 3). miR-26 family miRNAs and miR-221-3p exert anti-angiogenic effects, whereas miR-30 family miRNAs and miR-199a-3p exert pro-angiogenic effect. Jansen *et al.* showed that DM is associated with reduced endothelial miR-126 and miR-26a expression in circulating microvesicles; however, the authors analyzed patients with different DM types and for varied durations [42]. These two miRNAs exert both pro- and anti-angiogenic effects by promoting vascular regeneration, inhibiting apoptosis, and reducing insulin sensitivity in endothelial cells [57-59]. In the present study, we found that hsa-miR-26b expression was upregulated in the *Ect*-enriched plasma fraction obtained from patients with DM. These results suggest the presence of a delicate equilibrium in miRNA cargo that may be

associated with the functional regulatory properties of *Ects* in patients with DM.

Experimental evidence supports this hypothesis in other fields. For example, monocyte-derived *Ects* carrying pro-angiogenic miRNAs (miR-150) exert strong pro-angiogenic effects, and this pro-angiogenic activity of these *Ects* is attenuated after depletion of the pro-angiogenic miRNAs [60]. In stem cells, relative proportions of exosomal miRNAs with antagonistic regulatory functions are important for exerting anti-proliferative effect on cancer cells [61]. Moreover, variability of miRNAs in exosomes derived from different sarcomas is important for inducing the formation of a colonization niche around the primary tumor by these miRNAs [62].

We previously reported the pro-angiogenic potential of *Ects* with *Ects* obtained from patients with T2DM expressing cytokines and angiogenic factors, including those regulated once activated, normal T-cell expressed and secreted (RANTES), and angiotensin-2 [30]. Thus, *Ects* can be considered specific miRNA carriers that may promote angiogenesis in endothelial cells by targeting different angiogenic pathways, including active pSTAT3, nuclear factor (NF)- κ B, Wnt, PPAR γ , and ERK5/MAPK7 signaling pathways [23, 63]. Moreover, *Ect*-induced endothelial dysfunction should be considered by comparing data from specific endothelial biomarkers in T2DM [34].

Technical and clinical limitations associated with *Ect* study

The high diversity of EVs may hamper compatible measurement of *Ects* by using different analytical methods. To prevent these complications, different biophysical and biochemical properties of *Ects*, including size, mass density, shape, and surface antigen expression, can be used for their isolation [64]. In the present study, we used high-speed centrifugation for performing plasma fractionation and *Ect* fractionation as described elsewhere [29, 30]. This strategy cannot fully evade contamination with HDL and LDL proteins, which can bind and transfer miRNAs within a circulation system [65], as well as AGO proteins. To eliminate these effects, we used the normalization method to reduce the potential effect on miRNA analysis.

EV number depends on detection techniques and different pre-analytical variables such as blood collection and handling, plasma preparation, and storage conditions. Pre-analytical variables, namely collection, handling and storage of samples, have been extensively studied in previous work and showed insignificant effects over EVs when performed in identical conditions [66]. Therefore,

optimization and standardization of detection techniques are important to precisely define microvesicles and to prevent false-high or false-low quantification. Three ISTH Scientific and Standardization Subcommittees (SSC Vascular Biology, DIC, and Haemostasis & Malignancy) have initiated a project for standardizing the enumeration of cellular MPs by performing flow cytometry. Two collaborative workshops were implemented to establish the resolution and threshold levels of flow cytometers currently used in laboratories. Results of these workshops showed the potential and laboratory limitations of microvesicle evaluation methods [67]. Thus, *Ect* data should be considered in pre-analytical and analytical phases as well as with patient characteristics, especially for clinical purposes. In the present study, we used different imaging methods to visualize the presence of EV, such as TEM, cryo-TEM, and AFM. The accompanying wide-field images (Figure S1) and wide-field cryo-TEM images (Figure 4A-B, E-F) show potential contaminants, the presence of which should be accounted for in data analysis and normalization methods.

The clinical limitation of the present study was associated with patient characteristics. Body mass index (BMI) was significantly different between patients with T2DM and healthy controls. Patients with T2DM included in the present study were mostly overweight (mean BMI, \sim 30 kg/m²), which strongly increases the risk of T2DM. In our logistic regression model, BMI (as well as age and arterial hypertension) was analyzed as a potential confounding factor of miRNA levels in DM incidence, without any significance. However, the effect of gender bias should be considered because our study groups were not analyzed with due respect to this risk factor. Similarly, while smoking behavior was not the subject of this study, the role of cigarette smoking on miRNA levels cannot be overlooked.

Subcutaneous adipose tissues, including those present in the abdominal area, possess adipose mesenchymal stem cells that produce EVs, and these cells contribute to the diversity of miRNA profiles in *Ects* [61]. Therefore, excessive fat accumulation should be considered as an additional factor that impairs angiogenesis in patients with T2DM. Another important issue is glucose regulation in patients with T2DM and healthy controls. The median HbA1C level in patients included in the present study was 7.5% (7.1%–7.8%), which was similar to that reported in other studies [11, 42], and indicated imbalanced glucose regulation in patients with T2DM included in the present study. Moreover, two healthy subjects included in the present study showed incidentally elevated glucose levels, which affected the mean

glucose level. However, healthy controls included in the present study were not considerably different from those included in previous studies. Moreover, they were carefully selected with respect to age, gender, smoking behavior, and cardiovascular diseases (except hypertension). Healthy controls differed from T2DM patients with respect to biochemical characteristics, having higher total and LDL cholesterol levels and lower triglycerides levels than T2DM patients, which may affect *Ect* formation. These differences were probably associated with hyperlipidemia treatment in patients with DM that were included in the present study. Diabetes management guidelines recommend the use of statins to normalize the lipid profile, and to reduce cardiovascular risk in patients with DM; the use of antiplatelet and hypotensive drug therapies is also recommended [28].

Conclusions

The present study is the first systematic study to determine the signatures of *Ect*-specific miRNAs in patients with T2DM, and to analyze their pro- and anti-angiogenic potential by using bioinformatics tools. Our results clearly show the distribution of pro- and anti-angiogenic miRNAs in patients with T2DM, as indicated by the upregulation or downregulation of miRNAs in the *Ect*-enriched plasma fraction. This finding may highlight a possible mechanism underlying the occurrence of vascular complications due to impaired angiogenesis in patients with T2DM.

Abbreviations

ACS: acute coronary syndrome; AFM: atomic force microscopy; BMI: body mass index; CB: cacodylic buffer; CHOL HDL: cholesterol high-density lipoprotein fraction; CHOL LDL: cholesterol low-density lipoprotein fraction; CAD: coronary artery disease; Cryo-TEM: cryogenic transmission electron microscopy; DM: diabetes mellitus; Ects: ectosomes; EVs: extracellular vesicles; HbA1C: glycated haemoglobin A1C fraction; hs-CRP: high-sensitivity C-reactive protein; IS: ischemic stroke; LoD: limit of detection; MPs: microparticles; miRNAs: microRNAs; NTA: nanoparticle tracking analysis; NGSP: national glycohemoglobin standardization program; OGTT: oral glucose tolerance test; PS: phosphatidylserine; PPP: platelet-poor plasma; SD: standard deviation; TC: total cholesterol; TG: triglycerides; TEM: transmission electron microscopy; T1DM: type 1 diabetes mellitus; T2DM: type 2 diabetes mellitus.

Supplementary Material

Supplementary figure and tables.

<http://www.thno.org/v08p3874s1.pdf>

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Competing Interests

The authors have declared that no competing interest exists.

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