

Metatranscriptomic Analysis of Larval Guts from Field-Collected and Laboratory-Reared *Spodoptera frugiperda* from the South American Subtropical Region

Christina B. McCarthy,^{a,b} Natalia A. Cabrera,^{a,b} Eduardo G. Virla^{c,d}

Centro Regional de Estudios Genómicos, Facultad de Ciencias Exactas, Universidad Nacional de La Plata, La Plata, Buenos Aires, Argentina^a; Departamento de Informática y Tecnología, Escuela de Ciencias Agrarias, Naturales y Ambientales, Universidad Nacional del Noroeste de la Provincia de Buenos Aires, Pergamino, Buenos Aires, Argentina^b; PROIMI-Biotecnología, División Control Biológico, San Miguel de Tucumán, Tucumán, Argentina^c; Instituto de Entomología, Fundación Miguel Lillo, San Miguel de Tucumán, Tucumán, Argentina^d

This is the first study to report a high-throughput approach integrating gene expression data from *Spodoptera frugiperda* guts and their associated metatranscriptomes. Our datasets provide information on the potential effects of environmental conditions on the expression profile of *S. frugiperda* larval guts, their associated metatranscriptome, and putative interactions between them.

Received 7 June 2015 Accepted 12 June 2015 Published 16 July 2015

Citation McCarthy CB, Cabrera NA, Virla EG. 2015. Metatranscriptomic analysis of larval guts from field-collected and laboratory-reared *Spodoptera frugiperda* from the South American subtropical region. *Genome Announc* 3(4):e00777-15. doi:10.1128/genomeA.00777-15.

Copyright © 2015 McCarthy et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Christina B. McCarthy, mccarthychristina@gmail.com.

Spodoptera frugiperda is a noctuid moth that devastates various crops including corn, rice, and cotton and is found in most of the American continent (1). Even though transgenic crops that produce insecticidal proteins from *Bacillus thuringiensis* are currently the most successful biotechnological pest management application (2), *S. frugiperda* has developed field-evolved resistance to Bt maize (3). In this sense, gut microbiota play important roles in the growth and development of insects, and metatranscriptomic analyses of insect hosts represent an invaluable tool which could lead to new targets for pest control (4, 5). Nevertheless, the assessment of metabolically active components in the gut microbiota (6) is surprisingly scarce. Furthermore, comprehensive analyses combining host gene expression data with information from associated microbiota (7), are even more rare. The purpose of this study was to integrate gene expression data from *S. frugiperda* guts and their associated metatranscriptomes, under natural and controlled conditions. For this, four *S. frugiperda* samples from the province of Tucumán (Argentina; subtropical region) were analyzed. Specimens were obtained from different environments, altitudes and food sources, namely, (1) a transgenic maize (*Zea mays*) field at 495 meters above sea level (MASL) where insecticides and fertilizers were applied (named Sf_MM; 26°49'50"S, 65°16'59.4"W), (2) *Sorghum halepense* at 495 MASL (Sf_MS; 26°49'50"S, 65°16'59.4"W), (3) a maize field at 2,283 MASL where no insecticides or fertilizers were used (Sf_TV; 26°55'40.75"S, 65°45'19.90"W), and (4) a colony established from larvae originally collected from the same transgenic maize field as Sf_MM, reared for 9 generations under controlled conditions on an artificial diet adapted from reference (8), without the addition of antibiotics (Sf_LR). For all samples, total RNA extracted from fifth instar larval guts (two digestive tracts per sample), was submitted to a one-step reverse transcription and PCR sequence-independent amplification procedure, modified from reference

(9), as described previously (7, 10). High-throughput pyrosequencing of the samples was performed using a Roche GS FLX (Macrogen, Inc., Republic of Korea), yielding ~1 Gb of metatranscriptomic reads with lengths of 50 to 1,600 base nucleotides (nt) (652 nt average).

Raw sequence reads were trimmed to remove nucleotides derived from the amplification primers using a custom application (7, 10). The nonredundant protein sequence NCBI database (DB:nr) was downloaded locally. Rapsearch2 (11) was used for the protein homology search against DB:nr, using the trimmed singlet reads, and the taxonomic and functional content of the datasets was analyzed with MEGAN (12, 13). An average of ~97% of the transcripts showed homology to eukaryota, as expected (less than 1% corresponded to nonlepidopteran transcripts, including fungi, kinetoplastida, and viridiplantae, among others), 0.28% to 0.07% to bacteria, 0.003% to 0.001% to archaea, and 0.03% to 0% to viruses. The highest number of bacterial transcripts was found in Sf_MS, whereas archaeal and viral transcripts were more abundant in Sf_LR. Statistical analysis (Fisher's exact test [14], $P < 0.05$) mostly indicated significant differences between samples.

This is the first study to report *S. frugiperda* metatranscriptomic datasets, which provide information on the potential effects of food source, insecticides, fertilizers, and environmental conditions on the expression profile of *S. frugiperda* larval guts, their associated metatranscriptome, and putative interactions between them.

Nucleotide sequence accession numbers. Nucleotide sequences were submitted to the NCBI Sequence Read Archive (SRA) under accession numbers [SRX1000994](https://www.ncbi.nlm.nih.gov/sra/SRX1000994), [SRX1000995](https://www.ncbi.nlm.nih.gov/sra/SRX1000995), and [SRX1000996](https://www.ncbi.nlm.nih.gov/sra/SRX1000996) (field-collected *S. frugiperda* larval guts) and [SRX684522](https://www.ncbi.nlm.nih.gov/sra/SRX684522) (laboratory-reared *S. frugiperda* larval guts).

ACKNOWLEDGMENTS

This research was supported by Agencia Nacional de Promoción Científica y Tecnológica (PICT PRH 112), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) (PIP 0294), and Universidad Nacional del Noroeste de la Provincia de Buenos Aires (UNNOBA) (Exp. 1388/2010 and Exp. 2581/2012) grants to C.B.M.

We gratefully acknowledge the Faculty of Agronomy and Zootechnics (Tucumán National University, Finca El Manantial, Lules Department, Tucumán) and Ing. Zoot. Carlos Marino (Azucarera Justiniano Frías S.A., Las Carreras, Valle de Tafí, Tucumán) for their support and for allowing us to collect *S. frugiperda* larvae from their fields.

C.B.M. and E.G.V. are members of the CONICET research career. N.A.C. is the recipient of an UNNOBA fellowship.

REFERENCES

1. Pogue M. 2002. A world revision of the genus *Spodoptera* (Guenée) *Lepidoptera: Noctuidae*. American Entomological Society, Philadelphia, PA.
2. Sanchis V. 2011. From microbial sprays to insect-resistant transgenic plants: history of the biopesticide *Bacillus thuringiensis*. A review. *Agron Sustain Dev* 31:217–231. <http://dx.doi.org/10.1051/agro/2010027>.
3. Storer NP, Babcock JM, Schlenz M, Meade T, Thompson GD, Bing JW, Huckaba RM. 2010. Discovery and characterization of field resistance to Bt maize: *Spodoptera frugiperda* (*Lepidoptera: Noctuidae*) in Puerto Rico. *J Econ Entomol* 103:1031–1038.
4. Morrison M, Pope PB, Denman SE, McSweeney CS. 2009. Plant biomass degradation by gut microbiomes: more of the same or something new? *Curr Opin Biotechnol* 20:358–363. <http://dx.doi.org/10.1016/j.copbio.2009.05.004>.
5. Dillon RJ, Dillon VM. 2004. The gut bacteria of insects: nonpathogenic interactions. *Annu Rev Entomol* 49:71–92. <http://dx.doi.org/10.1146/annurev.ento.49.061802.123416>.
6. Shao Y, Arias-Cordero E, Guo H, Bartram S, Boland W. 2014. *In vivo* pyro-SIP assessing active gut microbiota of the cotton leafworm, *Spodoptera littoralis*. *PLoS One* 9:e85948. <http://dx.doi.org/10.1371/journal.pone.0085948>.
7. McCarthy CB, Santini MS, Pimenta PF, Diambra LA. 2013. First comparative transcriptomic analysis of wild adult male and female *Lutzomyia longipalpis*, vector of visceral leishmaniasis. *PLoS One* 8:e58645. <http://dx.doi.org/10.1371/journal.pone.0058645>.
8. Murúa G, Virla EG. 2003. Evaluación de cuatro dietas artificiales para la cría de *Spodoptera frugiperda* (Lep.: Noctuidae) destinada a mantener poblaciones experimentales de himenópteros parasitoides. *Bol San Veg Plagas* 29:43–51.
9. Sambade A, Martín S, Olmos A, García ML, Cambra M, Grau O, Guerri J, Moreno P. 2000. A fast one-step reverse transcription and polymerase chain reaction (RT-PCR) amplification procedure providing highly specific complementary DNA from plant virus RNA. *J Virol Methods* 87: 25–28. [http://dx.doi.org/10.1016/S0166-0934\(00\)00145-2](http://dx.doi.org/10.1016/S0166-0934(00)00145-2).
10. McCarthy CB, Diambra LA, Rivera Pomar RV. 2011. Metagenomic analysis of taxa associated with *Lutzomyia longipalpis*, vector of visceral leishmaniasis, using an unbiased high-throughput approach. *PLOS Negl Trop Dis* 5:e1304. <http://dx.doi.org/10.1371/journal.pntd.0001304>.
11. Zhao Y, Tang H, Ye Y. 2012. RAPSearch2: a fast and memory-efficient protein similarity search tool for next-generation sequencing data. *Bioinformatics* 28:125–126. <http://dx.doi.org/10.1093/bioinformatics/btr595>.
12. Huson DH, Auch AF, Qi J, Schuster SC. 2007. MEGAN analysis of metagenomic data. *Genome Res* 17:377–386. <http://dx.doi.org/10.1101/gr.5969107>.
13. Huson DH, Mitra S, Ruscheweyh HJ, Weber N, Schuster SC. 2011. Integrative analysis of environmental sequences using MEGAN4. *Genome Res* 21:1552–1560. <http://dx.doi.org/10.1101/gr.120618.111>.
14. Fisher R. 1970. *Statistical methods for research workers*. 14th ed. Hafner, New York, NY.