1	The pituitary TGF $\beta$ 1 system as a novel target for the treatment of resistant
2	prolactinomas.
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# 25 ABSTRACT

26 Prolactinomas are the most frequently observed pituitary adenomas and most 27 of them respond well to conventional treatment with dopamine agonists. However, a subset of prolactinomas fails to respond to such therapies and is considered as 28 29 dopamine agonist-resistant prolactinomas (DARPs). New therapeutic approaches are 30 necessary for these tumors. TGF<sup>β1</sup> is a known inhibitor of lactotroph cell proliferation 31 and prolactin secretion, and it partly mediates dopamine inhibitory action. TGF<sup>β</sup>1 is 32 secreted to the extracellular matrix as an inactive latent complex, and its bioavailability 33 is tightly regulated by different components of the TGFB1 system including latent binding proteins (LTBPs), local activators (Thrombospondin-1, matrix metalloproteases, 34 35 integrins, among others), and TGF $\beta$  receptors. Pituitary TGF $\beta$ 1 activity and the 36 expression of different components of the TGF $\beta$ 1 system, are regulated by dopamine and estradiol. Prolactinomas (animal models and humans) present reduced TGF<sup>β1</sup> 37 38 activity as well as reduced expression of several components of the TGFB1 system. 39 Therefore, restoration of TGF $\beta$ 1 inhibitory activity represents a novel therapeutic approach to bypass dopamine action in DARPs. The aim of this review is to summarize 40 41 the large literature supporting TGF<sup>β1</sup> important role as a local modulator of pituitary 42 lactotroph function; as well to provide recent evidence of the restoration of TGF<sup>β1</sup> 43 activity as an effective treatment in experimental prolactinomas.

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# 46 **Pituitary tumors**

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Pituitary tumors are commonly benign, slow growing adenomas and account for 10-15% of all intracranial neoplasms (Farrell 2006; Melmed 2015). The prevalence of these tumors is relatively high in the general population, with approximately 77 cases per 100.000 (Daly *et al.* 2009; Fernandez *et al.* 2010), and studies of autopsy specimens identified up to a 20% prevalence of clinically occult pituitary adenomas (Ezzat *et al.* 2004).

Despite their benign features pituitary tumors can cause considerably morbidity due to both hypersecretion of pituitary trophic hormones and excessive tumor growth that can affect surrounding tissue. Common symptoms of a pituitary tumor compressive 'mass effect' include visual impairment, headaches, neurological disorders and hypopituitarism caused by disruption of the hypothalamic-pituitary axis (Arafah & Nasrallah 2001; Melmed 2011). Based on their size, pituitary adenomas are classified as microadenomas (< 10mm), macroadenomas (>10mm) or giant adenomas (>40mm).

61 Pituitary tumors usually present with monoclonal growth and can also be 62 classified according to their cell type origin and hormone secretion. Thus, somatotropinomas secrete growth hormone (GH), prolactinomas secrete prolactin 63 (PRL), thyrotropinomas thyroid-stimulating 64 secrete hormone (TSH), and corticotropinomas secrete adrenocorticotropin hormone (ACTH). In contrast, the non-65 66 functioning pituitary adenomas do not produce any hormone and usually derive from gonadotropes (Kovacs et al. 2001; Syro et al. 2015). 67

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## 69 **Prolactinomas**

Among functioning pituitary tumors, prolactinomas are the most frequently observed in the clinic (40%) (Ciccarelli *et al.* 2005). Excessive PRL secretion by these tumors leads to hyperprolactinemia, which primarily affects gonadal/reproductive function, causing hypogonadism, galactorrhea, decreased libido and infertility both in

men and women. Large macroprolactinomas can also cause neurological symptoms
 due to compression of adjacent tissues.

Prolactinomas are usually benign, and although some tumors show invasion into the parasellar compartment and/or sphenoid sinuses, malignant transformation and metastatic spread are extremely rare. Macroprolactinomas tend to be more aggressive and resistant to therapies than microprolactinomas (Wong *et al.* 2015a).

Differences in prolactinoma incidence, tumor size and behavior have been 80 81 described among genders. The prevalence of prolactinomas is higher in women during 82 the fertile period (20-50 years), while the frequency is similar between sexes after the fifth decade of life (Colao et al. 2003; Gillam et al. 2006). Also, women usually present 83 84 with microprolactinomas whereas men more often present with macroprolactinomas ( 85 Delgrange et al. 1997; Nishioka et al. 2003). These differences have been associated to the earlier diagnosis in woman due to the readily detection of symptoms caused by 86 87 high prolactin (amenorrea/ galactorrea) (Delgrange et al. 1997;Colao et al. 2003; 88 Nishioka et al. 2003; Gillam et al. 2006). However, delayed diagnosis in men may not 89 be the only explanation for the differences in tumor size, since young men also present 90 with macroprolactinomas, and prolactinomas in men tend to be more aggressive, with 91 higher proliferative indexes and lower rates of surgical cure, suggesting a sex-specific 92 behavior for these tumors (Delgrange et al. 1997; Gillam MP & Molitch ME 2015).

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## Prolactinoma treatment

The major goals of treatment in patients with prolactinomas are to normalize serum PRL levels, to restore gonadal function, to reduce tumor size and to preserve or improve residual pituitary function. Prolactin secretion in the normal pituitary is tonically inhibited by hypothalamic dopamine through dopamine D2 receptors (Drd2) expressed on lactotroph cell membranes (Ben Jonathan & Hnasko 2001). The majority of prolactinomas retain an intact response to dopamine inhibition, therefore medical treatment with dopamine agonists (DAs), such as cabergolina and bromocriptine,

represents the first line therapy for this tumors, including microprolactinomas, macroprolactinomas and giant prolactinomas (Wong *et al.* 2015b). Dopamine agonists are highly effective in achieving therapeutic aims with a favorable benefit/risk balance compared with surgical treatment.

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# 107 Dopamine agonist-resistant prolactinomas (DARPs)

Despite the universal use of dopamine agonists and their high efficiency in reducing PRL levels and decreasing tumor size, there is a subset of prolactinomas (10-15%) that do not respond appropriately to the treatment, even at high doses of DA (Vroonen *et al.* 2012). These tumors represent a major challenge for clinical management. DARPs are more prevalent in men than woman, occur most frequently as macroprolactinomas and tend to be invasive, exhibiting extension to the cavernous sinuses.

115 The molecular mechanisms underlying the escape from dopaminergic 116 regulation in DARPS are not fully elucidated. The main candidate thought to be 117 responsible for resistance is the Drd2 itself. However, to date, no point mutation in the 118 Drd2 gene has been identified in DARPs (Friedman et al. 1994; Molitch 2003; Gillam et 119 al. 2006; Vroonen et al. 2012; Molitch 2014). Nevertheless, several mechanisms that 120 lead to reduced Drd2 sensitivity were described in resistant prolactinomas, including 121 evidence of decreased Drd2 mRNA expression, and differential expression of short and long Drd2 isoforms (Caccavelli et al. 1994; Vasilev et al. 2011; Shimazu et al. 2012); 122 reduced Drd2 density and reduced dopamine binding sites in plasma membranes of 123 124 DARP cells (Pellegrini et al. 1989). Alterations in dopamine signaling, such as 125 decreased expression of the inhibitory alpha G protein subunit (G<sub>ai2</sub>) have also been 126 described (Caccavelli et al. 1996); as well as decreased expression of the nerve growth factor receptor (NGFR), which indirectly modulates Drd2 expression (Passos et al. 127 128 2009). Histological studies on DARPs also revealed increased angiogenesis, cellular atypia (multinucleated cells, irregular nuclei) and increased proliferation index 129

measured by Ki67 staining, indicating an overall increase invasiveness (reviewed in
(Gurlek *et al.* 2007)).

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## 133 Alternative treatments for DARPs

At present, there is no alternative medical treatment for DARPs, and transsphenoidal surgery is indicated if the tumor is still resectable (Primeau *et al.* 2012; Smith *et al.* 2015). However, some aggressive prolactinomas recur post-operatively and show progressive growth, in which case, radiotherapy is the next therapeutic option, but with limited efficiency (Molitch 2014).

The chemotherapy agent temozolomide (TMZ) has been recently used as a last resort therapy and showed a moderately successful response in large aggressive DARPs (Whitelaw *et al.* 2012; Liu *et al.* 2015; McCormack *et al.* 2011). However the efficacy of TMZ therapy in aggressive pituitary adenomas remains controversial (Bruno *et al.* 2015) and clinical trials are now necessary to establish the indications, doses, and duration of TMZ administration to more accurately determine the efficacy of this agent.

New therapeutic approaches are necessary for those prolactinomas that are resistant to conventional treatments. Few reports of experimental treatments can be found in the literature and show variable effectiveness in pre-clinical and *in vitro* models. For instance, treatment with somatostatin receptor (SSTR) analogs failed to inhibit prolactin secretion by cultured cells derived from DARPs (Fusco *et al.* 2008) despite the expression of all subtypes of SSTR in human prolactinomas (Jaquet *et al.* 1999).

Based on the counteracting effects of estradiol on dopamine action in lactotrophs, targeting of the estrogen receptor with tamoxifen was evaluated in the precabergoline era in patients with bromocriptine-resistant prolactinomas, but only a moderated reduction in PRL levels was observed (Volker *et al.* 1982). A novel antiestrogen agent, fulvestrant, also reduced PRL secretion in pituitary cell lines, and

decreased tumor growth and serum PRL in estrogen-induced prolactinomas in rats

159 (Cao *et al.* 2014).

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## 161 In the search for new therapeutic targets for DARPs

Studies in animal models of prolactinomas with altered sensitivity to DA, such as the estrogen induced prolactinomas in rats and the Drd2 knock-out mice (Drd2<sup>-/-</sup>) have been very helpful to identify molecular pathways altered in these tumors and to test potential future therapies. Many of these studies suggest that the deregulation of local growth factors and extracellular matrix remodeling participate in the pathogenesis of prolactinomas by promoting cell proliferation, angiogenesis and invasiveness (Paez-Pereda *et al.* 2005; Cristina *et al.* 2005; Cristina *et al.* 2007; Recouvreux *et al.* 2013).

Transforming growth factor beta 1 (TGFβ1), a well-known inhibitor in lactotroph
 physiology, has been recently identified as a novel target for the development of new
 therapies in resistant prolactinomas.

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## 173 The complexity of the TGF $\beta$ system and biology

TGFβs are multifunctional cytokines known to play crucial regulatory roles in 174 cellular proliferation and differentiation, angiogenesis, extracellular matrix modification 175 176 and immunomodulation (Yoshinaga et al. 2008), and have powerful effects on embryogenesis, development and tissue homeostasis (Heldin et al. 2009; Galvin-177 Burgess et al. 2013; Itoh et al. 2014). The TGFβ family comprises more than 30 highly 178 179 pleiotropic molecules including activins, inhibins, nodal, bone morphogenetic proteins 180 (BMPs), the anti-muellerian hormone (AMH), and several growth and differentiation 181 factors (GDFs) among others (Derynck & Akhurst 2007). Three isoforms of TGFβ have 182 been identified (TGF $\beta$ 1, 2, and 3).

The importance of TGFβ1 is clearly demonstrated by the fact that TGFβ1 null mutation causes excessive inflammatory response and early death (Kulkarni *et al.* 1993). On the contrary, an excess of TGFβ1 activity is associated to connective tissue

diseases, fibrosis and inflammation, cirrhosis, arthritis and sclerosis, cardiovascular diseases and cancer, making TGF $\beta$  an interesting target for therapeutic research (Pohlers *et al.* 2009; Doyle *et al.* 2012; Akhurst & Hata 2012).

Nearly all cell types are sensitive to TGF $\beta$ 1, but TGF $\beta$  action is highly dependent on cell type, developmental stage, physiological- pathological conditions, interaction with components of the extracellular matrix and, once bond to its receptor, interaction with other signaling pathways.

193 The potent biological activity of TGF $\beta$ 1 is tightly regulated at different levels, 194 including its synthesis, secretion, storage and activation. The three TGFβ isoforms are synthesized as homodimeric precursor molecules that contain a pro-peptide sequence, 195 196 called latency associated peptide (LAP), and the functional mature TGF<sup>β</sup> sequence 197 (Figure 1. 1). After proteolytic processing by furin within the trans-golgi, LAP remains 198 associated with the mature TGFB by non-covalent interactions in a small latent 199 complex (Figure 1. 2). While in the endoplasmic reticulum, LAP is linked, by disulfide 200 bonds, with a latent TGF $\beta$  binding protein (LTBP) (Figure 1. 3). LTBPs belong to a 201 family of large secretory extracellular matrix (ECM) glycoproteins. Although LTBPs are 202 not required for maintenance of TGF $\beta$  latency, they facilitate the secretion, storage, 203 and activation of the TGF $\beta$ -LAP complex (Rifkin 2005).

204 TGF $\beta$  is secreted as part of this large latent TGF $\beta$  complex (LLC), (Figure 1. 3), 205 and is incorporated as component of the ECM, which acts as a cytokine reservoir 206 (Figure 1. 4). Trapped in the ECM, TGF $\beta$  remains latent because of persistent binding 207 of LAP and must undergo a highly regulated activation process by which mature cytokine is released from its latent complex to enable the active form to bind and signal 208 209 through its receptor (Figure 1. 5). Latent TGF $\beta$  activation is a crucial event in governing the cytokine biological function and availability in the ECM (Annes et al. 2003; Annes et 210 211 al. 2004; Rifkin 2005).

Several latent TGF $\beta$ 1 activators have been described, including proteases 212 (such as plasmin, matrix metalloproteinase 2 (MMP2), matrix metalloproteinase 9 213 (MMP9), BMP-1, thrombospondin-1 (TSP-1), kallikrein 1 (KLK1), integrins  $\alpha\nu\beta6$  and 214 215  $\alpha\nu\beta$ 8, and reactive oxygen species (ROS) or pH changes in the local environment, among others. However, their individual biological importance in releasing TGF $\beta$ 1 from 216 217 its latent complex and their local regulation in different tissues are not fully understood 218 (Annes et al. 2003; Annes et al. 2004; Yoshinaga et al. 2008). Since all these factors 219 are related to ECM perturbations, the latent TGF $\beta$  complex has been postulated as a "sensor" of environment disturbances (Annes et al. 2003). 220

221 Once TGF $\beta$ 1 is released from the ECM, the active cytokine binds to its transmembrane receptor, the type II TGF $\beta$  receptor (T $\beta$ RII), a constitutively active 222 kinase that recruits and phosphorylates T $\beta$ RI (type I TGF $\beta$  receptor) forming a 223 224 heterotetrameric complex of serine/threonine kinase receptors containing two type I and two type II subunits (Figure 1, 6). Next, T $\beta$ RI phosphorylates the downstream 225 receptor-associated Smads (R-Smads: Smad2/Smad3), which form a heteromeric 226 complex with Smad4, and translocate to the nucleus to regulate the transcription of 227 target genes (Figure 1. 7). Additionally, an inhibitory Smad, Smad7, competes with the 228 229 Smad2/3 for binding to the activated T $\beta$ RI, thereby exerting a negative effect on 230 TGFβ/Smad signaling (Shi & Massague 2003; Han *et al.* 2015).

TGFβ can also signal through smad-independent pathways, including the mitogenactivated protein kinases (ERK1/2, JNK, p38), small GTP-binding proteins (Ras, RhoA, Rac1, CDC42, mTOR), the NF-κB pathway and Wnt/β-catenin pathway, the AKT/PKB pathway and phosphatidylinositol-4,5-bisphosphate 3-kinase (PI3K) (Attisano & Wrana 2002; Derynck & Zhang 2003; Moustakas & Heldin 2005).

As a multifunctional cytokine with powerful effects on cell proliferation, cellular
 migration and inflammation, TGFβ signaling has been targeted for drug development

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and numerous strategies have proceeded through preclinical to clinical trials (reviewed
 in (Akhurst & Hata 2012)).

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## **TGF**β1 in the pituitary: a brief history

243 The earliest publications on TGF $\beta$ 1 action in the pituitary date from the late 80s and early 90s (Ying et al. 1986; Mueller & Kudlow 1991). Dr. Sarkar and col were the 244 245 first to demonstrate local TGF $\beta$ 1 mRNA and protein expression in the pituitary gland, and the inhibitory action of TGF $\beta$ 1 on prolactin secretion and lactotrophic growth in 246 1992 (Sarkar et al. 1992). Although these first evidences were found in animal models 247 248 (rat), TGF<sup>β1</sup> and T<sup>β</sup>RII expression were promptly found to be expressed in human 249 pituitaries (Halper et al. 1992; Fujiwara et al. 1995), as well as in human pituitary adenomas (Fujiwara et al. 1995; Jin et al. 1997). 250

251 The main physiological modulators of lactotroph function are dopamine and estradiol, which exert inhibitory and stimulatory actions, respectively (Ben Jonathan & 252 Hnasko 2001). The pro-mitotic effect of estradiol (pharmacological doses) and its role 253 254 in prolactinoma induction is very well described in the literature (Heaney et al. 2002). 255 However, estrogens also participate in the lactotroph cell turnover in normal pituitary 256 glands, sensitizing lactotroph cells to apoptotic stimuli. Therefore the effect of estradiol 257 on lactotroph function depends on the dose and normal/tumoral condition of the cells 258 (Pisera et al. 2004; Zaldivar et al. 2009; Jaita et al. 2015). Interestingly, dopamine and 259 estradiol also regulate the expression of both TGFβ1 and its receptor, but in opposite 260 ways. Thus, while estrogen stimulation increases serum PRL levels and lactotroph proliferation, it decreases the expression of TGF $\beta$ 1 in the anterior pituitary. On the 261 262 contrary, dopamine, acting through the Drd2, up-regulates TGF<sup>β1</sup> expression and secretion in vivo and in vitro, with a concomitant reduction in the proliferation rate of 263 lactotrophs. Moreover, it has been proposed that TGF<sup>β1</sup> partially mediates the 264 inhibitory effect of dopamine on lactotroph proliferation (Sarkar et al. 2005). Our group 265 has recently described that the amount of pituitary active TGF $\beta$ 1 is also locally 266

regulated by dopamine and estradiol treatment in mice, and moreover we found an inverse correlation between active TGF $\beta$ 1 levels and serum PRL (Recouvreux *et al.* 2011). It is worth noting that less than 8% of total pituitary TGF $\beta$ 1 was found in the active form, similar to what has been described in other tissues (Yoshinaga *et al.* 2008). This underscores the tightly regulation of the latent TGF $\beta$  activation process.

Other important factor regulating lactotroph homeostasis is PRL itself, acting through the PRL receptor (PRLR). It has been shown that endogenous PRL exerts paracrine/autocrine antiproliferative and proapoptotic effects on lactotrophs, and moreover, knock-out mice lacking PRLR develop prolactinomas, further demonstrating the important role of PRL in the negative feedback on lactotroph function (Schuff *et al.* 2002; Ferraris *et al.* 2012; Ferraris *et al.* 2014).

Whether PRL can as well regulate TGFβ1 expression or function in the pituitary gland
 is an open question that has not yet been addressed.

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Alterations in the components of the TGFβ1 system during prolactinoma
 development

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Evidences of TGFβ1 alterations in estradiol-induced prolactinomas in rats 284 The estrogen-treated rat is a well-known model for prolactinoma development 285 with increased pituitary weight, hyperprolactinemia, lactotroph hyperplasia and reduced 286 dopaminergic action at the pituitary level (Heaney et al. 1999; Heaney et al. 2002b). 287 Furthermore, estradiol treatment decreases pituitary TGFB1 and TBRII mRNA and 288 protein, together with an increase in PRL levels (Sarkar et al. 1992; Pastorcic et al. 289 1995; De et al. 1996) (Figure 2). Therefore, the inhibition of TGF<sup>β1</sup> and T<sup>β</sup>RII might 290 291 cooperate in the development of prolactinomas induced by estradiol (Hentges & Sarkar 292 2001). In agreement with this idea, pituitary tumorigenesis induced by estrogen

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293 treatment is greatly accelerated in TβRII heterozygous knockout mice (TβRII<sup>+/-</sup>) where

the expression of T $\beta$ RII is markedly reduced (Shida *et al.* 1998).

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# **TGF**β1 alterations in the prolactinoma development in Drd2<sup>-/-</sup> mice

Another well characterized model to study prolactinoma development are the 297 transgenic knock out mice lacking functional Drd2 (Drd2<sup>-/-</sup>). This model represents an 298 299 excellent model to mimick dopamine agonist-resistance. Because of the absence of inhibitory dopaminergic control, these mice display chronic hyperprolactinemia and 300 301 lactotroph hyperplasia (Kelly et al. 1997; Diaz-Torga et al. 2002; Cristina et al. 2006), 302 but the loss of dopamine inhibition has deeper effect on pituitary function in female than 303 in male mice (Saiardi et al. 1997; Diaz-Torga et al. 2002). In females the increase in 304 serum prolactin levels is much more pronounced than in males, and females develop 305 lactotroph hyperplasia from 6 month onwards, while age-matched Drd2-deficient males 306 develop pituitary lactotroph adenomas at 17 to 20 months of age (Asa et al. 1999).

Interestingly, active and total TGF<sup>β1</sup> levels, as well as T<sup>β</sup>RII and LTBP1 307 expression are reduced in Drd2<sup>-/-</sup> pituitaries compared to controls (wild type or wt). 308 309 highlighting the stimulatory role of dopamine on pituitary TGF $\beta$ 1 system (Recouvreux et 310 al. 2011) (Figure 2). On the other hand, the impact of the chronic loss of dopaminergic tone on the TGF $\beta$ 1 system was also stronger in females, evidenced by the 311 downregulation of several putative TGF $\beta$ 1 activators (MMP2, MMP9, MT1-MMP, 312 313 thrombospondin-1 and kallikrein) as well as the decreased expression of TGF $\beta$ 1 target genes observed only in females (Drd2<sup>-/-</sup> vs. their wt counterpart). In this model we 314 315 found sex differences in the regulation of the TGFβ1 system: males express higher levels of several components of the TGF $\beta$ 1 system, and it could be due to the lower 316 serum estradiol levels present in males, as estradiol negatively controls most of the 317 components of the system (Recouvreux et al. 2013). We suggest that stronger pituitary 318 TGFβ1 system could protect males from excessive lactotroph proliferation and 319

prolactinoma development. Then, sex differences found in the regulation of the TGFβ1
 system could explain sex differences found in the incidence of prolactinoma
 development in this model.

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# **324** - **TGF**β1 alterations in human pituitary tumors

In humans the expression of several components of the TGF<sup>β</sup> signaling 325 326 pathway was recently compared in five normal human anterior pituitaries, 29 invasive 327 nonfunctioning pituitary adenomas (NFPAs) and 21 noninvasive NFPAs (Zhenye et al. 2014). This report demonstrated that TGFβ1 mRNA expression and p-Smad3 protein 328 329 levels gradually decreased, while Smad7 mRNA levels gradually increased from 330 normal anterior pituitaries to noninvasive NFPAs and invasive NFPAs. The authors concluded that the activity of TGF $\beta$  signaling would be limited during tumor 331 332 development.

Recent work also described a significant down-regulation of the TGF $\beta$ 1/Smad signaling cascade in 12 cases of DARPs compared to normal human anterior pituitaries. The authors showed that TGF $\beta$ 1 mRNA levels, and Smad2 and Smad3 mRNA and protein expression were significantly decreased in human prolactinomas (Li *et al.* 2015) (Figure 2).

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Overall, decreased TGF $\beta$ 1 activity and decreased expression of different components of the TGF $\beta$ 1 system have been described in animal models of prolactinomas as well as in human prolactinomas. Taking into account that TGF $\beta$ 1 inhibits lactotroph proliferation and PRL synthesis and secretion, we speculate that recovering local TGF $\beta$ 1 activity could contribute to revert the adenoma development and to normalize prolactinemia.

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Recovery of local TGFβ1 activity. Successful treatment in an experimental
 model of prolactinoma

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Thrombospondin-1 (TSP1) is one of the main physiologic latent TGFβ1 activators *in vitro* and *in vivo* (Schultz-Cherry *et al.* 1994). TSP1 is a large multifunctional matrix glycoprotein involved in cell growth, adhesion, migration (Lawler 2002). TSP1 also functions as an endogenous antiangiogenic factor, inhibiting the proliferation and migration of endothelial cells by interaction with its cell surface receptor CD36 and by antagonizing VEGF activity (Lawler & Lawler 2012).

Based on the CD36-binding peptide sequence from TSP-1, small-molecules were 355 356 developed to mimic TSP-1 antiangiogenic properties (Haviv et al. 2005). Several of these new drugs were able to slow tumor growth in preclinical models (Anderson et al. 357 358 2007; Yang et al. 2007; Garside et al. 2010). Among them, ABT-510 and ABT-898 359 (Abbott Laboratories), two of such TSP1 analogs, were assayed in several solid tumors (Haviv et al. 2005). ABT-510 was evaluated in phase II clinical trials for the treatment of 360 361 head and neck cancer, non-small cell lung cancer, lymphoma, and renal cell carcinoma 362 (Haviv et al. 2005; Ebbinghaus et al. 2007; Markovic et al. 2007; Yang et al. 2007; Gordon et al. 2008; Nabors et al. 2010). The second-generation TSP-1 synthetic 363 364 analogue, ABT-898, was found to have greater potency than ABT-510 and is expected to have greater efficacy than the other available TSP-1-mimetic peptides (Garside et al. 365 2010; Campbell et al. 2011) due to its lower clearance rate. 366

Immunoreactive TSP-1 is present in the anterior pituitary, particularly in endothelial cells (Burns & Sarkar 1993), and TSP-1 levels and its antiangiogenic effect are reduced in prolactinomas induced by estradiol in rats (Sarkar *et al.* 2007) and in the hyperplastic pituitaries of  $Drd2^{-/-}$  mice (Recouvreux *et al.* 2013). TSP-1 expression was also found down-regulated in invasive *vs.* non-invasive prolactinomas in humans (Jiang *et al.* 2012).

Given that: 1) TSP1 is an antiangiogenic factor, 2) TSP1 expression is reduced
 during prolactinoma development, 3) TSP1 is a known TGFβ1 activator, 4) TGFβ1
 activity is also reduced during the development of prolactinomas, and 5) TGFβ1 is an

inhibitory factor of lactotroph proliferation and synthesis; we speculated that treatments that improve pituitary TSP-1 and/or TGF $\beta$ 1 activities could reduce the progression of prolactinomas.

379 We first evaluated whether the TSP1 analogs were able to activate TGF<sup>β</sup>1 in 380 the pituitary. In fact, an *in vivo* short term treatment (100 mg/kg ABT-510, ip, 30 min) enhanced the biological activity of pituitary TGF $\beta$ 1, with a concomitant reduction in 381 382 serum prolactin levels (Recouvreux et al. 2012). Notably, same effect was observed 383 after short term treatment with ABT-510 in female rats carrying prolactinoma induced by chronic estradiol treatment. We next evaluated whether an *in vivo* treatment for two 384 385 weeks with the TSP1 analogs could counteract the development of estradiol-induced prolactinomas in rats. ABT-510 and ABT-898 treatment (100mg/kg ip, thrice a week for 386 two weeks) significantly decreased pituitary tumor size, reduced tissue angiogenesis 387 388 and pituitary proliferation markers, as well as serum prolactin levels, in female rats with 389 prolactinomas induced by chronic treatment with estradiol (Recouvreux et al. 2012). 390 Furthermore, ABT-510 and ABT-898 treatment markedly increased active TGF<sup>β1</sup> 391 content, measured by ELISA within the tumors. The increase in cytokine activation was 392 also reflected in the recovery of intrapituitary pSMAD2/3 expression (Figure 3). Besides 393 from the well-known antiangiogenic effect of these TSP-1 mimetic peptides, the 394 improvement of the local TGF $\beta$ 1 biological activity, most likely contributed to the 395 reduction in serum prolactin and in the inhibition of prolactinoma growth.

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# Overall conclusions and Perspectives

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Prolactinomas are the most frequent pituitary tumors in adults accounting for 60% of all functioning pituitary tumors (Ciccarelli *et al.* 2005). Even though prolactinomas are usually benign and in most cases respond well to treatment with dopaminergic agents, 15% of these tumors are resistant to classical therapy, become

403 invasive and aggressive, and require extirpation. The mechanisms underlying the escape from dopaminergic regulation in DARPS are not fully understood, and the main 404 405 candidate to be responsible for resistance is the Drd2 itself. Since TGF $\beta$ 1 mediates, at 406 least partially, the inhibitory action exerted by dopamine on lactotrophs, and reduced 407 TGF<sub>β1</sub> activity is a common feature of prolactinoma development, treatments that improve pituitary TGF $\beta$ 1 activity represent a rational approach to develop alternative 408 409 therapies for DARPs. Supporting this, we provide evidence of the effectiveness of a 410 treatment with the small TSP1-analog peptides ABT-510 and ABT-898 to restore 411 TGF $\beta$ 1 activity and to counteract prolactinoma development in rats.

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Taken together the data summarized here, the recovery of TGFβ1 activity
emerges as a novel therapeutic target for treatment of dopamine agonist resistant
prolactinomas.

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## FIGURE LEGENDS

## Figure 1: The biology of TGFβ system

TGF $\beta$  is synthesized as homodimeric precursor containing a pro-peptide sequence LAP, (1), and then processed by furin. LAP remains associated with the mature TGF $\beta$  by non-covalent interactions in a small latent complex (2), which in turn is linked by disulfide bonds to one of the latent TGF $\beta$  binding proteins (LTBP1-4) (3). TGF $\beta$  is secreted as part of this large latent TGF $\beta$  complex (LLC) (3), and it is incorporated as component of the extracellular matrix (ECM), which acts as a reservoir of the cytokine (4). TGF $\beta$  must undergo a highly regulated activation process by which mature cytokine is released (5) to enable binding to its receptor complex (T $\beta$ RI and T $\beta$ RII) (6) and signal through Smad2/Smad3 pathway (7). Known TGF $\beta$  activators are listed in the upper left.

# Figure 2: Alterations of TGFβ1 system in prolactinomas

Decreased expression of different components of pituitary TGFβ1 system in prolactinoma models are represented with down-pointed arrows. Gray arrows indicate findings in human prolactinoma specimens; black arrows indicate findings in Drd2<sup>-/-</sup> mice; white arrows indicate findings in estradiol induced prolactinomas in rats.

# **Figure 3:** Recovery of TGFβ1 activity emerges as a novel therapeutic target for treatment of dopamine agonist resistant prolactinomas. Treatment with Thrombospondin-1 analogs (ABT-510, ABT-898) recover pituitary TGFβ1 activity, reduce tumor size, tumor angiogenesis and proliferation markers, as well as serum prolactin in estradiol-induced prolactinomas in female rats.



Figure 1: The biology of TGFβ system. TGFβ is synthesized as homodimeric precursor containing a propeptide sequence LAP, (1), and then processed by furin. LAP remains associated with the mature TGFβ by non-covalent interactions in a small latent complex (2), which in turn is linked by disulfide bonds to one of the latent TGFβ binding proteins (LTBP1-4) (3). TGFβ is secreted as part of this large latent TGFβ complex (LLC) (3), and it is incorporated as component of the extracellular matrix (ECM), which acts as a reservoir of the cytokine (4). TGFβ must undergo a highly regulated activation process by which mature cytokine is released (5) to enable binding to its receptor complex (TβRI and TβRII) (6) and signal through Smad2/Smad3 pathway (7). Known TGFβ activators are listed in the upper left. 254x190mm (96 x 96 DPI)



# Fig.2 Decreased expression of the TGF $\beta$ 1 components in prolactinomas

Figure 2: Alterations of TGFβ1 system in prolactinomas. Decreased expression of different components of pituitary TGFβ1 system in prolactinoma models are represented with down-pointed arrows. Gray arrows indicate findings in human prolactinoma specimens; black arrows indicate findings in Drd2-/- mice; white arrows indicate findings in estradiol induced prolactinomas in rats. 254x190mm (96 x 96 DPI)



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