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REVIEW ARTICLE

# Current status of cardiac MRI in small animals

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**Abstract** Cardiac magnetic resonance imaging (MRI) on small animals is possible but remains challenging and not well standardized. This publication aims to provide an overview of the current techniques, applications and challenges of cardiac MRI in small animals for researchers interested in moving into this field. Solutions have been developed to obtain a reliable cardiac trigger in both the rat and the mouse. Techniques to measure ventricular function and mass have been well validated and are used by several research groups. More advanced techniques like perfusion imaging, delayed enhancement or tag imaging are emerging. Regarding cardiac applications, not only coronary ischemic disease but several other pathologies or conditions including cardiopathies in transgenic animals have already benefited from these new developments. Therefore, cardiac MRI has a bright future for research in small animals.

**Keywords** Magnetic resonance imaging · Heart · Coronary ischemic disease · Rat

## Introduction

Magnetic resonance imaging (MRI) is a powerful technique used both in clinical practice and in research. In routine cardiac imaging, MRI is recognized as a robust and accurate method to measure cardiac function, perfusion and viability. The ease of transferring new methods developed on experimental models to clinical imaging is another stimulus to perform research on cardiac MRI. Research on small animals has numerous advantages such as realistic physiologic in vivo models, knockout animals as well as reduced team and cost to handle the experiments. Several imaging methods have been developed for the phenotypic analysis of small animals, amongst which MRI has been proposed [1]. Still, cardiac imaging on a small animal using MRI remains challenging and not well standardized. Therefore, this work aims to provide an overview of the current techniques, applications and challenges of cardiac MRI in small animals for researchers interested in moving into this field. First, the methodology will be discussed followed by the applications and results obtained so far.

## Cardiac MRI acquisition

### Optimal magnetic field strength

Usually, cardiac MRI research is performed on high-field magnets with a small bore specially designed for small animals. The advantage of using high fields relies on the increased signal that allows higher resolution to be achieved. Fields of 4.7 and 7 T are commonly used [2]. Experiences on 11 T and above have also been reported for applications like coronary angiography and phosphorus spectroscopy [2]. The position of the magnet (horizontal vs. vertical as encountered in high-field systems) does not seem to be relevant as prolonged upright body position exerts no significant changes in murine left ventricle hemodynamics [3]. However, high field is not mandatory as successful experiments have also be performed at 1.5 T on a clinical system [4, 5]. The loss of signal is compensated to some extent by dedicated coils that improve the signal reception.

### Trigger systems and animal monitoring

For structural studies like myocardial mass quantification, high-quality predominantly diastolic MR images may be obtained by signal averaging without any trigger (6). However, for all other techniques including a fast determination of the myocardial mass, an ECG trigger is absolutely required for cardiac MRI. Considering the high heart rate encountered in small animals (between 200 and 600 bpm) and the small voltage recorded for the ECG, the cardiac synchronization can be really challenging. Commercial trigger systems are available that can amplify the ECG signal recorded in small animals. However, the ECG may be corrupted during image acquisition by the imaging process, as for example in cardiac microscopy when short repetition time and high gradient-slew rate are used. Therefore other types of trigger devices have been developed [7]. The fiber-optic stethoscope system for example is inserted in the esophagus and optically detects pulsate compression of the esophageal wall [8]. As another type of device, the «lever-coil» is a coil mechanically coupled to the animal but not located within the resonator or gradient coil that is sensitive to both cardiac and respiratory motion [9]. Finally, the arterial pressure can also be used to trigger the MRI acquisition instead of the ECG [10].

Regarding the need for a respiratory gating, the position of the animal seems important as respiratory compensation is needed in vertical high-field magnets but not in horizontal magnets [11].

In addition to image triggering, cardiac and respiratory monitoring permits the display of vital signs. In conjunction with general anesthesia under isoflurane [12] and body temperature control [13], exam durations of around 2 h have been achieved.

### MR sequences and protocols

Various sequences are used for cardiac imaging in rats (10). There are no systematic studies comparing the different MR sequences. The choice of the MR sequences seems to depend more on both the investigators and the MR systems used. The most frequently used are T1 gradient echo cine sequences, yielding a typical in-plane resolution of 200–300 $\mu$m in less than a minute for a single slice on high-field magnets [14].

### Myocardial mass

Due to its ability to offer high spatial resolution, myocardial mass can be accurately quantified in vivo by MRI, as it has been demonstrated in the mouse [15–17]. The method relies on a complete coverage of the heart using cine images, followed by a delineation of the epicardial and endocardial contours of the left ventricle, as shown on Fig. 1. By considering the cardiac tissue density as constant, the segmented volume yields the mass.

This method has been validated by left-ventricle gravimetry [16]. Additional parameters like the volume of the cavities or the slice thickness can also be measured from the images.

On an isolated beating heart and high-field MR systems, the spatial resolution can be increased to observe the myocardial fiber structure [18–20]. Diffusion imaging has also been performed on isolated beating [19] and fixed hearts [21].

### Myocardial function

One of the strengths of cardiac MRI is the accuracy of measuring myocardial function in small animals. Several protocols have been proposed according to the type of measurements needed.

### Global and regional function

Global and regional functions are obtained using bright blood cine-imaging MR sequences [11, 22]. Parameters like ejection fraction, or wall thickness and wall thickening are reproducibly extracted from these data based on the high contrast between the blood and the myocardium. Stress cine-MRI of cardiac function have been performed using dobutamine in mice with an intravenous infusion at 4 or 40 $\mu$g/minute/kg [23], or an intra-peritoneal bolus injection of 1.5 $\mu$g/g body weight [24, 25].

### Strain and velocity imaging

Intra-myocardial wall motion, like radial and circumferential shortening or strains, torsion angle, can be measured

on strain imaging but not on cine imaging. The basic principle is to tag the myocardium physically using spatially selective saturation pulses and to track the displacement of the tagged myocardium. The feasibility of such an approach has already been demonstrated in the mouse using either SPAMM or DENSE imaging, as shown in Fig. 2 [26–30]. Another variant of these approaches uses phase-contrast imaging with bipolar gradients to encode the motion of the mouse myocardium [31]. The different merits and respective indications of these three techniques are yet to be determined in small animals.

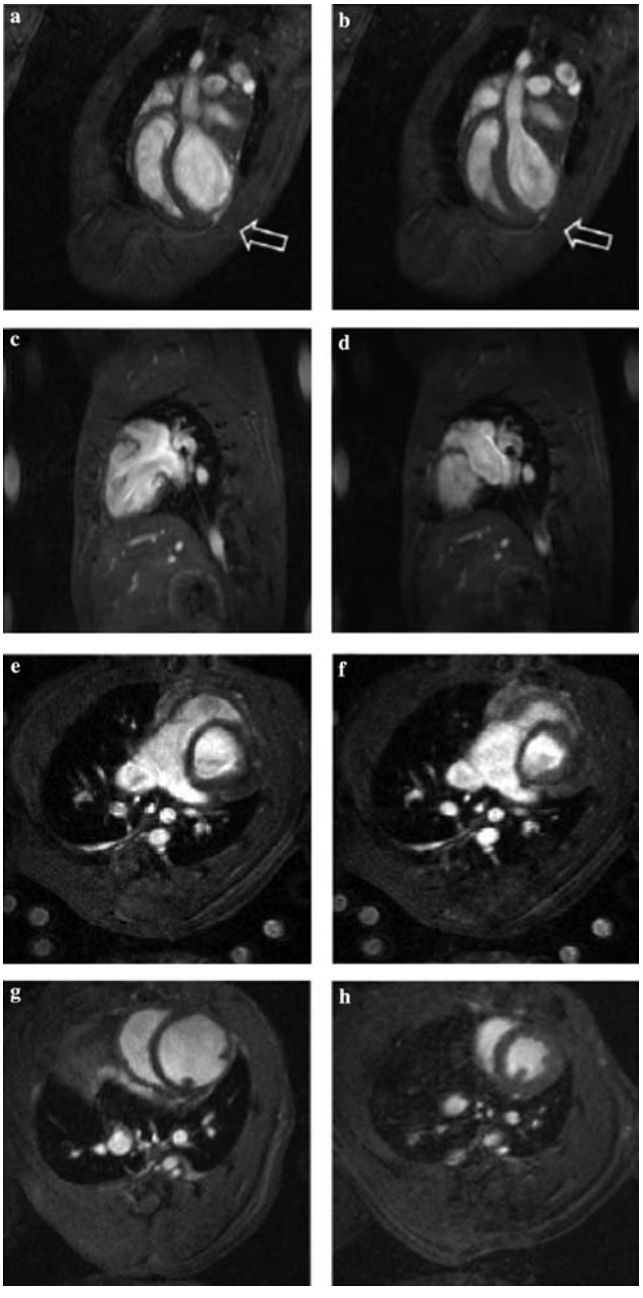


**Fig. 1** **a** and **b**: long-axis view of a rat heart 8 wk after large myocardial infarction (MI) in diastole (**a**) and systole (**b**). **c** and **d**: long-axis views of the RV 8 week after large MI in diastole (**c**) and systole (**d**). **e** and **f**: diastolic (**e**) and systolic (**f**) short-axis slices at the level of the tricuspid valve in a rat 8 wk after MI. **g** and **h**: diastolic short-axis slices at the level of the papillary muscles in a rat 8 wk after MI (**g**) and a sham-operated rat (**h**). Note the high spatial resolution that allows a clear delineation of the cardiac anatomy and to differential the normal and infarcted heart.(reprinted with permission form reference [65])

## Myocardial perfusion and blood volume

Quantification of coronary blood flow using phase-contrast techniques in small animals has only been achieved on isolated hearts [32]. Research has been more focused on measuring perfusion at the tissue level. Spin-labeling perfusion techniques [33] have been used in small animals [34–38] as an endogenous alternative to the myocardial perfusion measurement with exogenous contrast media used in humans. Indeed, in humans cardiac frequency is low enough to allow the acquisition of images with sufficient SNR and resolution within one heart beat to follow bolus. In small rodents, however, cardiac cycle duration is much shorter, of the order of 100–200 ms. This precludes acquisition of images with sufficient resolution and SNR within one heart beat. Therefore, MRI bolus tracking is less reliable in small animals. Also, reproducible bolus injections, required for perfusion quantification, are difficult to achieve in small animals [39].

The principle of spin-labeling perfusion quantification is based on inflowing, non-inverted spins into a selectively inverted slice. The mixing of non-inverted flowing and inverted spins in the imaging slice modifies the apparent relaxation time and thus creates flow-related contrast. Since T1 increases with higher magnetic fields, and higher T1 values allow longer inflow observation times, high magnetic fields are particularly useful for spin-labeling imaging.

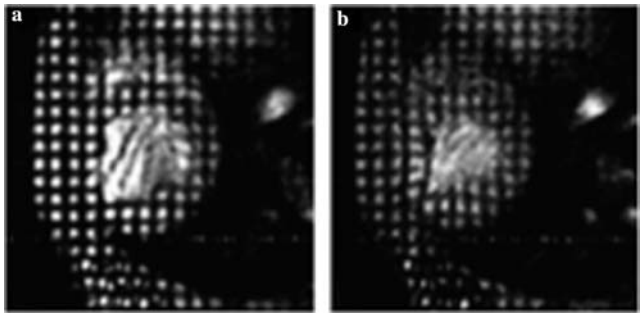


**Fig. 2** Typical images from an 11-phase (time points) tag MRI study in mouse are depicted shortly after end diastole (**a**) and at end systole (**b**). The analysis of the tag displacement is used to measure myocardial strains. (adapted with permission from reference [29])

Myocardial perfusion values at rest in rats from $3.5 \pm 0.1\,mg\,g^{-1}\,min^{-1}$ [35] to $5.5 \pm 0.7\,mg\,g^{-1}\,min^{-1}$ [34] have been reported in the literature and validated using a colored microspheres technique [35].

In addition to perfusion measurement by spin tagging, the injection of a T1 intravascular contrast media allows the determination of the myocardial blood volume [35, 40]. Other methods to measure regional myocardial blood volume is based on the accepted linear relationship between the regional blood volume and the transverse relaxation rate increase, $\Delta R2^*$, following iron oxide particle injection [41].

### Delayed enhancement MRI

Delayed imaging after contrast media injection is an established MR technique to reveal acute or chronic infarct in patients. This technique has been well validated in the mouse with an occlusion/reperfusion model and an extravascular contrast media (Gd-DTPA at a dose of 0.3–0.6 mmol/kg) by comparison to TTC staining [42]. In the rat with the occlusion/reperfusion model, however, the situation is less clear with more debated results. Both intravascular [43] and extravascular contrast media overestimated the infarct size whereas necrosis-avid gadolinium-based contrast media gave a correct measure [44]. In another rat study with a similar model, the enhanced region overestimated the infarct size immediately after the injection of Gd-DTPA, although it gradually decreased to match the size of the infarct [45]. Such a variation of the hyperenhanced region is not found in patients or other models and needs to be further studied [46].

Manganese-based contrast media have also been used to delineate myocardial infarct in rats as they accumulate inside normal myocytes but not in infarcted cells [47, 48]. However, the practical use of such contrast media that may also indicate the rate of calcium influx into the heart [25] remains to be defined.

### Metabolism and spectroscopy

The feasibility of 31 P MR spectroscopy has already been demonstrated in mice on isolated hearts [49, 50] and in vivo [51]. Using this technique, the myocardial phosphocreatine (PCr)-to-ATP (PCr/ATP) ratio remained unchanged in mice after dobutamine stress [49]. A report on sodium MR imaging in isolated rat hearts demonstrated an increase of the intracellular sodium during global ischemia [52]. However, these techniques remain difficult to master and are not widely used.

### Angiography and vessel wall imaging

Coronary MR angiography has been successfully performed on isolated rat hearts to demonstrate a coronary occlusion [2] but remains technically difficult, although possible, in mice in vivo [53]. Wall imaging of mice artery is a rapidly growing field due to the numerous knockout models that are available. With this technique, high MRI resolution of a limited vessel segment has been acquired in vivo in the aorta and carotid arteries of mice [54–59]. There are currently no reports on wall imaging of coronary arteries in small animals.

## Cardiac MRI applications

Using the previously described protocols on small animals, significant results have already been obtained in the cardiovascular field. These will be briefly reviewed to demonstrate the potential of cardiac MRI in small animals.

### Persistent coronary occlusion

This model is characterized by a permanent occlusion of a coronary artery, yielding an infarct of variable size depending on the site of the ligation (distal or proximal) [60]. The remodeling following myocardial infarct in the left ventricle has been well described using MRI [61]: the injured myocardium evolves toward scar tissue and the remote myocardium develops a hypertrophy of the wall as well as an increase of the intracapillary blood volume [62]. Ultimately, a cardiac failure may result [63]. MRI also demonstrated a right ventricle hypertrophy as in the left ventricle but no increase of the wall stress [64, 65].

The effect of transmyocardial laser revascularization [66], angiotensin converting enzyme inhibitors [67], testosterone [68] or statin [69] in this model of myocardial infarction have also been investigated.

### Occlusion/reperfusion model

This model is characterized by a transitory occlusion of a coronary artery, usually at its origin, followed by a release of the occlusion insuring a reperfusion of the injured myocardium. This results in an ischemia or an infarct of variable size depending of the duration of the occlusion. It allows also to study the no-reflow phenomenon characterized by obstructed intramyocardial micro-vessels despite reopened epicardial arteries [70].

MRI has been used to assess both infarct size and cardiac function in intact mice early after a large, reperfused myocardial infarction, revealing the existence of contractile dysfunction in non-infarcted regions of the heart [30, 70]. Nicorandil, a K-ATP channel opener with a nitrate-like effect attenuates left-ventricular dilatation and improves cardiac function in rats with reperfused myocardial infarction, as demonstrated by MRI [71–73]. In transgenic

mice with the occlusion/reperfusion model, MRI demonstrated the beneficial effect of over-expression of A1-adenosine receptors [74] and Angiotensin II type 2 receptors [75] on both systolic function and infarct size.

### Partial coronary occlusion (chronic ischemia)

In this model, a subtotal occlusion of the coronary artery results in impaired cardiac function and anatomy with multiple sites of injury including myocyte loss and hypertrophy and reparative fibrosis [76]. The effect of partial coronary occlusion on the residual myocardial blood flow at rest is controversial as it was found normal when measured by microspheres [77] but decreased in a recent study using MRI [78]. Perfusion, function and energy metabolism as assayed by 31 P spectroscopy in this model have been successfully obtained by MRI [2]. Although less often studied than the two previous models, the partial coronary occlusion technique has a strong potential as it can mimic the ischemic cardiopathy encountered in man.

### Other models

As the MRI technique to measure the cardiac function and mass is now validated for mice, numerous MR studies have been performed using transgenic animals such as myoglobin knockout mice [79], VEGF knockout mice [23], apolipoprotein E-deficient mice [80], caveoline-deficient mice [81] and transgenic mice expressing tumor necrosis factor alpha (TNF-alpha) [4].

Hyperthyroid and hypertensive cardiomyopathies have been studied by MRI. It was observed that the continuous blockade of calcium channels suppresses activation of calcineurin and the development of cardiac hypertrophy in spontaneously hypertensive rats [82, 83]. Mice with cardiomyocyte-specific disruption of the endothelin-1 gene were resistant to hyperthyroid cardiac hypertrophy [5].

MRI demonstrated the reduction of the myocardial mass and the amelioration of cardiac function induced by the angiotensin-converting enzyme (ACE) inhibitor captopril in streptozotocin (STZ)-diabetic male Wistar rats [84, 85].

MRI has also been used to study the cardiac graft rejection [86] with the accumulation of macrophages inside the transplant [87].

Finally, a series of experiments has been conducted to study the treatment and consequences of chagasic heart disease in mice [88–91, 91].

## Conclusions

This review illustrates the strong potential of MRI for cardiac imaging of small animals. Solutions have been found to obtain a reliable cardiac trigger in both the rat and the mouse. Techniques to measure ventricular function and mass have been well validated and are used by several research groups. More advanced techniques like perfusion imaging, delayed enhancement or tag imaging are emerging. However, an effort is still needed to standardize the acquisition protocols and data analysis. In this respect, better gold standards of cardiac parameters are required. For example, an atlas of normal and pathological cardiac strains as measured by tag MRI would be particularly useful. Regarding the cardiac application, not only coronary ischemic disease but several other pathologies including transgenic animals have already benefited from these new developments. Therefore, cardiac MRI has a bright future for research in small animals.

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