

# Ca<sup>2+</sup> signaling by T-type Ca<sup>2+</sup> channels in neurons

Lucius Cueni · Marco Canepari ·  
John P. Adelman · Anita Lüthi

Received: 14 August 2008 / Accepted: 15 August 2008 / Published online: 11 September 2008  
© Springer-Verlag 2008

**Abstract** Among the major families of voltage-gated Ca<sup>2+</sup> channels, the low-voltage-activated channels formed by the Ca<sub>v</sub>3 subunits, referred to as T-type Ca<sup>2+</sup> channels, have recently gained increased interest in terms of the intracellular Ca<sup>2+</sup> signals generated upon their activation. Here, we provide an overview of recent reports documenting that T-type Ca<sup>2+</sup> channels act as an important Ca<sup>2+</sup> source in a wide range of neuronal cell types. The work is focused on T-type Ca<sup>2+</sup> channels in neurons, but refers to non-neuronal cells in cases where exemplary functions for Ca<sup>2+</sup> entering through T-type Ca<sup>2+</sup> channels have been described. Notably, Ca<sup>2+</sup> influx through T-type Ca<sup>2+</sup> channels is the predominant Ca<sup>2+</sup> source in several neuronal cell types and carries out specific signaling roles. We also emphasize that Ca<sup>2+</sup> signaling through T-type Ca<sup>2+</sup> channels occurs often in select subcellular compartments, is mediated through strategically co-localized targets, and is exploited for unique physiological functions.

**Keywords** Ca<sub>v</sub>3 channel · Low-threshold burst · Oscillation · Thalamus · SK channel · SERCA

## Introduction

Voltage-gated Ca<sup>2+</sup> channels (VGCCs) transduce electrical excitability into intracellular Ca<sup>2+</sup> concentration increases and play an important role in a wide range of neuronal physiology. VGCCs are composed of three major subfamilies, which are encoded by the Ca<sub>v</sub>1, Ca<sub>v</sub>2, and Ca<sub>v</sub>3 genes that give rise to ten different  $\alpha_1$  channel subunits. A great deal is known about Ca<sup>2+</sup> influx through the high-voltage-activated (HVA) Ca<sup>2+</sup> channels encoded by the Ca<sub>v</sub>1 and Ca<sub>v</sub>2 channel genes and their contributions to neuronal signaling (for review, see, e.g., [19]). In contrast, the Ca<sub>v</sub>3 subfamily, generating the low-voltage-activated Ca<sup>2+</sup> currents, also referred to as T-type Ca<sup>2+</sup> currents [89], is poorly characterized in terms of the free Ca<sup>2+</sup> concentration ([Ca<sup>2+</sup>]<sub>i</sub>) increases resulting from channel activation and the signaling functions exerted by Ca<sup>2+</sup> ions entered through these channels. This deficit in our understanding is remarkable since T-type Ca<sup>2+</sup> currents have long been known to uniquely influence neuronal excitability [47, 59, 105]; they are widely expressed throughout the central nervous system (CNS), sometimes at very high densities [106]; and they are recruited during quite specific physiological circumstances [47, 88].

T-type Ca<sup>2+</sup> currents mediate the low-threshold burst discharges that occur during physiological and pathological forms of neuronal rhythmogenesis [47], such as the sleep-related slow electroencephalographic waves [27], rhythms related to motor coordination [6, 10], to learning [100], and hyper-synchronous oscillations during epilepsy [59, 96]. Additionally, T-type Ca<sup>2+</sup> currents are involved in sensory

---

Lucius Cueni and Marco Canepari contributed equally to this review.

L. Cueni · M. Canepari  
Division of Pharmacology and Neurobiology, Biozentrum,  
University of Basel,  
Klingelbergstrasse 70,  
CH-4056, Basel, Switzerland

J. P. Adelman  
Vollum Institute, OHSU,  
Portland, OR, 97239, USA

A. Lüthi (✉)  
Department of Cell Biology and Morphology,  
Faculty of Biology and Medicine, University of Lausanne,  
Rue du Bugnon 9,  
CH-1005, Lausanne, Switzerland  
e-mail: [anita.luthi@unil.ch](mailto:anita.luthi@unil.ch)

transmission [64], in dendritic integration [72, 76, 80], neurotransmitter release [18], neural development [68], mechanosensation [45], and pain [51]. The  $[Ca^{2+}]_i$  dynamics resulting from T-type  $Ca^{2+}$  channel activation are now being measured using advanced optical techniques and will shed light on  $Ca^{2+}$  signaling related to important physiological conditions. This review discusses recent insights into neuronal  $Ca^{2+}$  signaling mediated through T-type  $Ca^{2+}$  channels. It focuses on studies in which the corresponding  $[Ca^{2+}]_i$  dynamics have been optically resolved and associated with specific neuronal functions. The review will also discuss strong evidence for intracellular signaling events initiated through  $Ca^{2+}$  entry via T-type  $Ca^{2+}$  channels, as obtained from electrophysiological studies.

### Basic principles of T-type $Ca^{2+}$ channel function

T-type  $Ca^{2+}$  channels are distinct from other classes of VGCCs in terms of their gating characteristics and kinetics (Fig. 1a). T-type  $Ca^{2+}$  channels are activated by subthreshold membrane depolarizations between  $-65$  and  $-50$  mV. By virtue of their rapid and mostly complete inactivation at moderately depolarized membrane potentials, T-type  $Ca^{2+}$  channels can generate spike-like membrane depolarizations [47, 89, 105]. Depending on the specific  $Ca_v3$  channel subtype, the time courses of activation, inactivation, deactivation, and recovery from inactivation vary [63, 66, 111]. T-type  $Ca^{2+}$  channels require a preceding period of hyperpolarization to recover from inactivation and to become primed to open. Therefore, in many instances, T-type  $Ca^{2+}$  channel activation occurs as a rebound to inhibitory input. Accordingly, in neurons expressing T-type  $Ca^{2+}$  channels,  $[Ca^{2+}]_i$  increases will typically follow periods of hyperpolarization (Fig. 1b).

There are a number of reasons why our understanding of the  $[Ca^{2+}]_i$  signals generated by T-type  $Ca^{2+}$  channels lags behind that of the HVA  $Ca^{2+}$  channels. First, the contribution of T-type  $Ca^{2+}$  channels to  $[Ca^{2+}]_i$  is often masked by a more robust  $Ca^{2+}$  entry through HVA  $Ca^{2+}$  channels (see, e.g., [49, 75, 76, 113]). Second, the lack of potent and selective T-type  $Ca^{2+}$  channel blockers has hampered progress in dissecting their physiological contributions. Low concentrations of  $Ni^{2+}$  (50–100  $\mu$ M) or mibefradil (10–100  $\mu$ M) are commonly used to block T-type  $Ca^{2+}$  channels, but both of these affect other ion channels [84, 92]. T-type  $Ca^{2+}$  channels also have a comparatively low single-channel conductance, which, together with their rapid inactivation [47, 105], inherently limits  $Ca^{2+}$  influx. Third, in many neurons, T-type  $Ca^{2+}$  channels are predominantly expressed in the dendrites [26, 32, 57, 72], often in distal portions [26, 31, 73], which make resolving fluorescent signals arising from T-type  $Ca^{2+}$  channel activity difficult. Finally, in some cell types,  $Ca^{2+}$

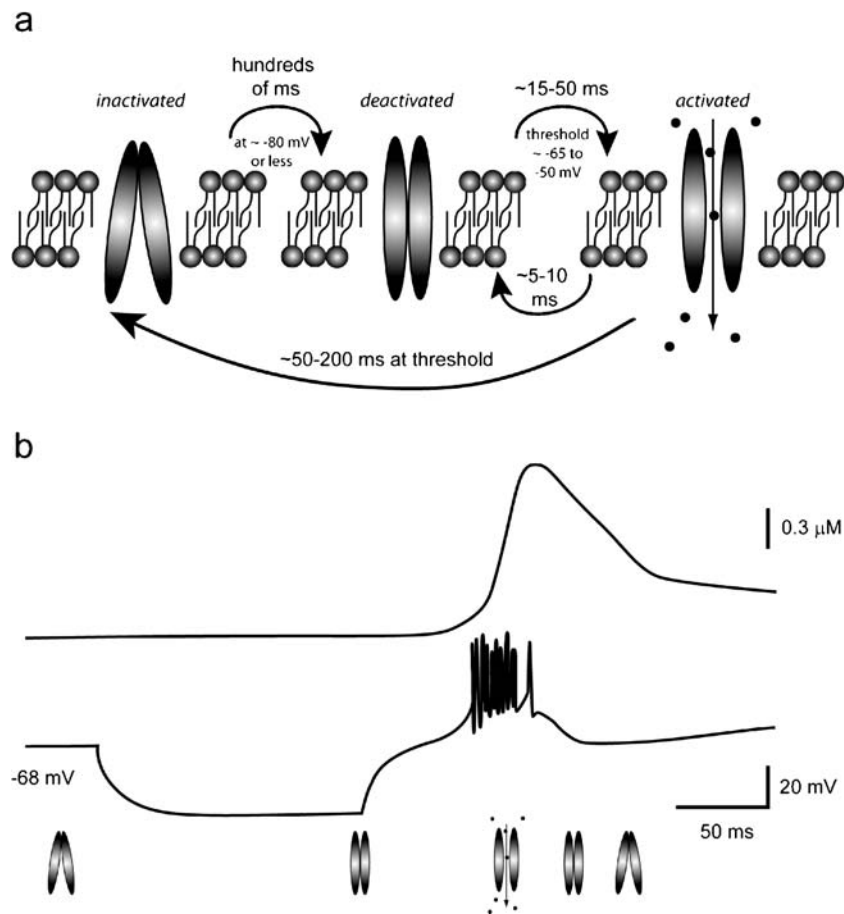
currents resistant to HVA channel blockers contain a large contribution from the  $Ca_v2$ -channels of the R-type, which resemble T-type  $Ca^{2+}$  currents in their comparatively hyperpolarized voltage range of activation and recovery from inactivation, and in overlapping pharmacological profiles, including mibefradil sensitivity [92]. Extended pharmacological and biophysical analyses, sometimes combined with genetic ablation techniques, have been applied recently to convincingly distinguish between the two  $Ca^{2+}$  channel types (see, e.g., [99, 108]).

Despite these obstacles, there is considerable evidence implicating T-type  $Ca^{2+}$  currents in important  $Ca^{2+}$  signaling roles (see also [88]). First, in several types of bursting neurons,  $Ca^{2+}$  entering during a burst discharge is important for the temporal patterning of oscillations [4, 6, 10, 14, 43, 70, 88, 104], although, in most cases, the  $Ca^{2+}$  current subtypes dominating  $Ca^{2+}$  entry during bursting have not yet been defined. Second, T-type  $Ca^{2+}$  channels expressed in non-bursting neurons contribute to  $Ca^{2+}$ -dependent ionic currents accompanying action potentials (APs) [77, 116]. Third, several forms of long-term synaptic plasticity depend on T-type  $Ca^{2+}$  currents [1, 12, 83, 85, 91]. Finally, pathological oscillations, such as those found during epileptic activity, are accompanied by enhanced T-type  $Ca^{2+}$  channel function [65, 103, 109, 117].

### $Ca^{2+}$ entry determined by the unique voltage-gating of T-type $Ca^{2+}$ channels

Three distinct voltage-gating characteristics endow  $Ca_v3$  channels with a unique identity as a  $Ca^{2+}$  source (Fig. 2). First, T-type  $Ca^{2+}$  currents are activated close to resting membrane potentials [47, 89, 105] and produce  $[Ca^{2+}]_i$  elevations at subthreshold potentials (Fig. 2a). For example, a  $Ni^{2+}$ -sensitive low-threshold  $Ca^{2+}$  signal accompanies subthreshold excitatory input in hippocampal CA1 and cortical layer V dendrites [72, 76]. In contrast, these channels typically make a minor contribution to  $Ca^{2+}$  entry resulting from a single AP [75, 77]. Through their low-threshold of activation, T-type  $Ca^{2+}$  channels can thus boost weak depolarizing inputs [41, 58, 110] and facilitate dendritic  $Ca^{2+}$  spikes [55, 117].

Second, T-type  $Ca^{2+}$  channels, while activating rapidly, deactivate comparatively slowly upon repolarization (Fig. 1a). During an AP lasting a few milliseconds, T-type  $Ca^{2+}$  channels may be activated but undergo little inactivation. Therefore, T-type  $Ca^{2+}$  channels remain open after the downstroke of the AP, permitting large  $Ca^{2+}$  influx due to the high driving force for  $Ca^{2+}$  ions (Fig. 2b) [77]. The dynamics of  $Ca^{2+}$  entry during AP repolarization, important for the timing of  $Ca^{2+}$ -dependent after-hyperpolarizing  $K^+$  currents [113], vary considerably



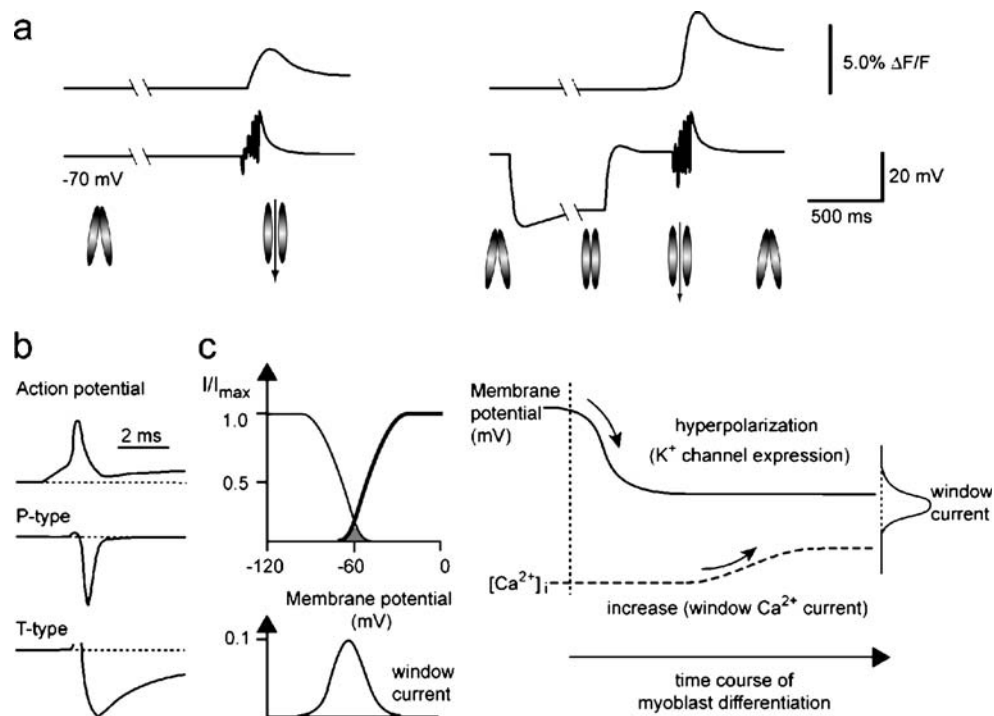
**Fig. 1** Schematic representation of T-type  $\text{Ca}^{2+}$  channel gating and the  $[\text{Ca}^{2+}]_i$  signals generated during a low-threshold burst. **a** A channel is depicted as two associated transmembrane proteins in a membrane bilayer in three different configurations (inactivated, deactivated, activated). Arrows point to the transitions between these three states of the channel. Next to the arrows, a range of time constants is indicated that is derived from work on heterologously expressed [63] and on native channels [47]. **b** Schematic traces presenting  $[\text{Ca}^{2+}]_i$

signals (*top trace*) obtained by a single low-threshold burst (*bottom trace*). This burst was evoked by a brief negative current injection to hyperpolarize membrane potential such that channels could recover from inactivation. Typical channel configurations accompanying these electrical events are schematically depicted below the traces. The  $[\text{Ca}^{2+}]_i$  signal is slightly delayed with respect to burst onset because it was derived from a recording with the low-affinity  $\text{Ca}^{2+}$  dye magfura-2 [29]

among the  $\text{Ca}_v3$  channel subtypes [63]. In cerebellar Purkinje cells, the time course of activation of  $\text{Ca}^{2+}$ -dependent afterhyperpolarizing  $\text{K}^+$  currents overlaps with that of T-type  $\text{Ca}^{2+}$  current deactivation [104], suggesting that the gating of the  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channels activated in between APs may be mediated via  $\text{Ca}^{2+}$  entering through T-type  $\text{Ca}^{2+}$  channels.

Third, although well-known for their prominent inactivation, all three T-type  $\text{Ca}^{2+}$  channel isoforms show a window current [66]. Window currents result from a small fraction of channels that remain open and lead to a stationary  $\text{Ca}^{2+}$  influx and a persistent enhancement of  $[\text{Ca}^{2+}]_i$  [28, 53] (Fig. 2c, left). The  $\text{Ca}_v3.3$  channel subtype gives rise to a comparatively large window current [66], which results in a steadily elevated basal  $[\text{Ca}^{2+}]_i$  and a propensity to generate oscillatory  $[\text{Ca}^{2+}]_i$  signals [25]. A

standing inward  $\text{Ca}^{2+}$  current in thalamocortical neurons underlies a membrane potential bi-stability [115] and the capability of switching abruptly between activated and silent states, such as those accompanying a slow ( $< 1$  Hz) sleep rhythm [46]. During the up-states of these rhythms, the window current additionally recruits  $\text{Ca}^{2+}$ -dependent cationic currents and permits a plateau-like up-state that may last for periods of seconds, whereas  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  channels initiate the down state [14]. Window currents ensure a steady, yet comparatively small,  $\text{Ca}^{2+}$  influx for prolonged periods of time around the resting membrane potentials of typical neurons and could thus promote cellular differentiation ([68], see also the “ $\text{Ca}^{2+}$  signaling through developmentally regulated expression of T-type  $\text{Ca}^{2+}$  channels” section), but also lead to abnormal cell growth and tumor proliferation [87].



**Fig. 2**  $[Ca^{2+}]_i$  signals generated through the unique voltage-gating characteristics of T-type  $Ca^{2+}$  channels. **a** *Left*, membrane potential showing a subthreshold response to repetitive stimulation (*bottom trace*) which is accompanied by a small  $[Ca^{2+}]_i$  increase (*top trace*). *Right*, this increase is boosted when synaptic stimulation is preceded by a short membrane hyperpolarization to allow for recovery from inactivation. Typical channel configurations introduced in Fig. 1a are shown *below the traces*. Traces adapted from [72]. **b** Time courses of the HVA P-type and T-type  $Ca^{2+}$  currents during an AP. Note the slow deactivation time course of the T-type  $Ca^{2+}$  current, leading to substantial  $Ca^{2+}$  influx during the postspike hyperpolarization. Traces

adapted from [104]. **c** *Left*, generation of a window  $Ca^{2+}$  current due to overlapping steady-state activation (*thick line*) and inactivation (*thin line*) curves. Voltage region covered by both curves is shaded. An expanded portion of this graph is shown below to highlight the amplitude and voltage dependence of the window current. *Right*, qualitative graphic representation of changes in membrane potential (*continuous line*) and  $[Ca^{2+}]_i$  (*dashed line*) during myoblast differentiation. Voltage range of window current is indicated to the right to illustrate that the hyperpolarization of membrane potential into this voltage range is a prerequisite for  $[Ca^{2+}]_i$  increases. Traces adapted from [11]

### $Ca^{2+}$ signaling due to spatially defined expression of T-type $Ca^{2+}$ channels

Electrophysiological recordings,  $Ca^{2+}$  imaging, and modeling studies support a nonuniform expression of T-type  $Ca^{2+}$  channels in diverse neuronal cell types.  $Ca^{2+}$  imaging of hippocampal dendrites of young adult rats revealed a graded expression profile for T-type  $Ca^{2+}$  channel-dependent signals, increasing with dendritic distance from the soma and reaching highest levels in distal dendrites [26, 56, 57, 73], in which T-type  $Ca^{2+}$  channel-mediated boosting of subthreshold potentials will facilitate dendrosomatic communication [71]. In thalamic neurons, T-type  $Ca^{2+}$  channels are heavily expressed in the dendrites, as shown by  $Ca^{2+}$  imaging [29, 81, 118], computational modeling [31], and cell-attached dendritic recordings [52]. In thalamocortical neurons of the paraventricular thalamic nucleus, T-type  $Ca^{2+}$  channels, predominantly formed by  $Ca_v3.1$  subunits, are concentrated in proximal dendrites and lead to numerous, closely spaced hotspots of increased  $[Ca^{2+}]_i$  [94], suggesting channel clustering, whereas in the

nucleus reticularis thalami (nRT), both  $Ca_v3.2$  and  $3.3$  subtypes are expressed and found along the somatodendritic axis [29, 52], but might be most strongly expressed in distal dendrites [31]. The dendritic localization of T-type  $Ca^{2+}$  channels underlies the robust bursting propensity of many neurons and enables the dendrites to be highly responsive to synaptic input [31]. T-type  $Ca^{2+}$  channels are also expressed on dendritic membranes of cerebellar Purkinje neurons [80] and contribute to the climbing fiber-evoked complex spikes observed in culture [20, 37]. Notably, two-photon imaging in Purkinje cell dendritic trees revealed expression of T-type  $Ca^{2+}$  channels at the level of single spines [49]. Therefore, these channels likely contribute to compartmentalized  $Ca^{2+}$  signaling in Purkinje cells in response to subthreshold input [36] and may influence the  $Ca^{2+}$  dependence of cerebellar synaptic plasticity [54].

The spatial compartmentalization of T-type  $Ca^{2+}$  channels has two important consequences for dendritic processing. First, the amplification of distal inputs through T-type  $Ca^{2+}$  channels boosts the sensitivity of peripheral sensory perception. In olfactory receptor neurons, odor-induced  $Ca^{2+}$

transients in the cilia-containing knobs are strongly boosted by  $\text{Ca}^{2+}$  entry through T-type  $\text{Ca}^{2+}$  channels. Blocking T-type  $\text{Ca}^{2+}$  currents with mibefradil reduces the knob  $[\text{Ca}^{2+}]_i$ , and transient and somatic signals are virtually absent [39]. Therefore, the local boosting of olfactory stimuli in the knob helps to propagate incoming olfactory signals to the soma, thereby ensuring AP generation [58]. Similarly, the high density of T-type  $\text{Ca}^{2+}$  channels in thalamic dendrites helps in the amplification of proximally impinging excitatory input and in securing robust burst discharge [29, 114]. Second, the non-uniform expression of T-type  $\text{Ca}^{2+}$  channels along the somatodendritic axis can lead to highly polarized  $[\text{Ca}^{2+}]_i$  increments. In cells of the vestibulocerebellum, burst firing induces a strong  $[\text{Ca}^{2+}]_i$  signal in the brush, which is predominantly carried through  $\text{Ca}_v3.1$  channels, whereas tonic AP firing produces a homogeneous  $[\text{Ca}^{2+}]_i$  increase throughout the cell [33]. Therefore, the bimodal activity pattern in these cells is accompanied by distinctly compartmentalized  $\text{Ca}^{2+}$  signaling.

### **$\text{Ca}^{2+}$ signaling through developmentally regulated expression of T-type $\text{Ca}^{2+}$ channels**

T-type  $\text{Ca}^{2+}$  channel expression undergoes marked developmental regulation and, in some cases, coincides with periods of neuronal differentiation (see also [68]). In the mouse peripheral vestibular system, large  $\text{Ca}_v3.2$ -type currents are recorded during embryonic development but current amplitude sharply decreases, reaching a minimum at postnatal day P4, after neurite outgrowth and synapse formation is terminated [2, 21]. Similarly, development of cochlear hair cells is accompanied by a marked reduction of  $\text{Ca}_v3.1$  channel mRNA and T-type  $\text{Ca}^{2+}$  currents [67]. In these cells, expression of T-type  $\text{Ca}^{2+}$  currents occurs during periods of elevated spontaneous AP generation, however, the signaling functions adopted by T-type  $\text{Ca}^{2+}$  channels during development remain to be specified. In *Xenopus* spinal neurons, the role of T-type  $\text{Ca}^{2+}$  currents is to facilitate rhythmic intracellular  $[\text{Ca}^{2+}]_i$  transients by initiating membrane depolarization that recruits HVA  $\text{Ca}^{2+}$  currents [43]. There is also evidence that the neuronal cell-adhesion molecules that promote differentiation regulate  $[\text{Ca}^{2+}]_i$  through mechanisms involving T-type  $\text{Ca}^{2+}$  channels [61].

Increased  $[\text{Ca}^{2+}]_i$  through window currents mediated by T-type  $\text{Ca}^{2+}$  currents have been implicated in skeletal muscle development. Skeletal muscle differentiation is a  $\text{Ca}^{2+}$ -dependent process that involves the transcription of a network of muscle-specific genes, and the fusion of myoblasts into multinucleate myotubes. At the onset of the fusion process, a subpopulation of myoblasts hyperpolarizes due to the expression of an inward rectifier  $\text{K}^+$  current [62], starts expressing  $\text{Ca}_v3.2$  T-type  $\text{Ca}^{2+}$  channels,

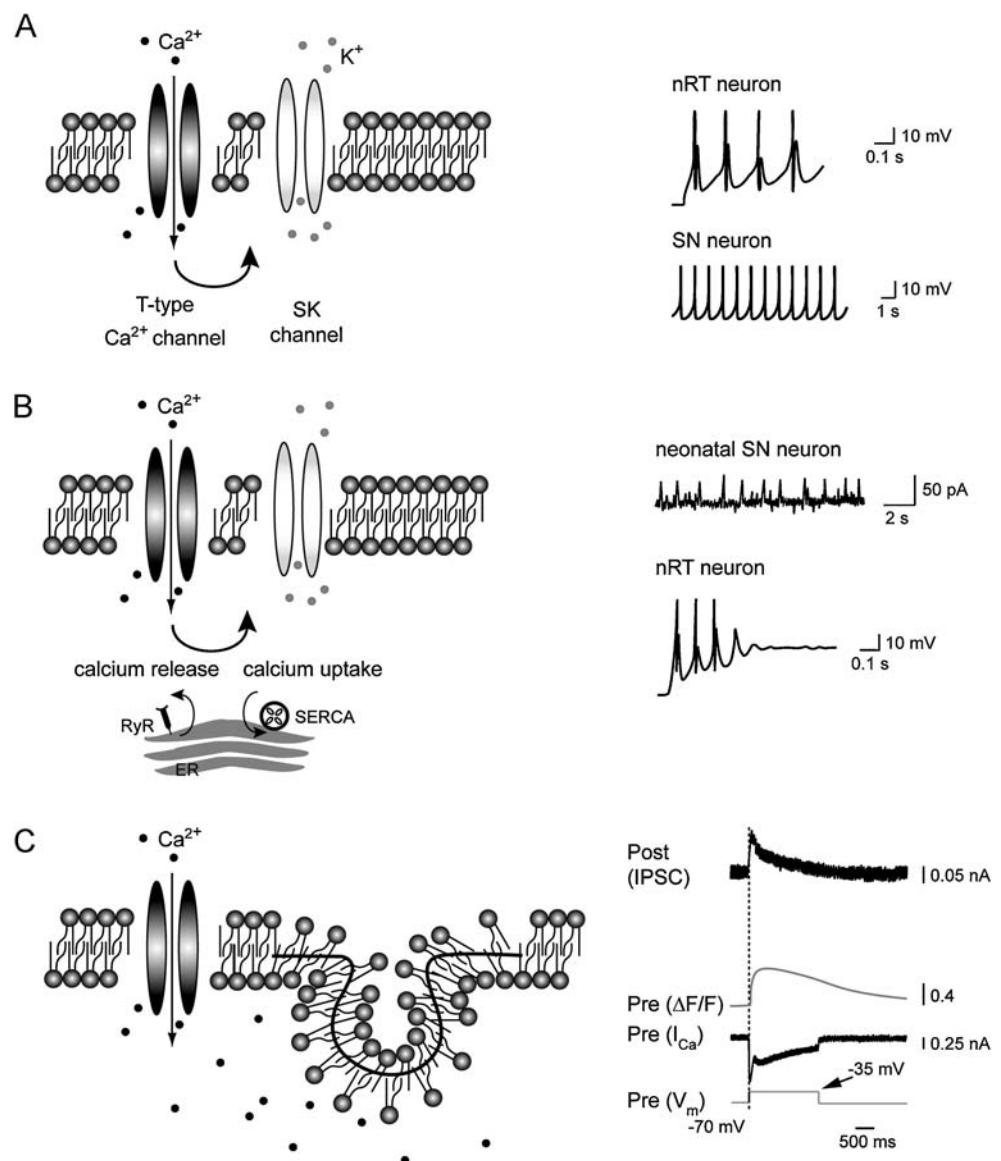
and shows elevated  $[\text{Ca}^{2+}]_i$  levels prior to fusion [11] (Fig. 2c, right). Pharmacological inhibition of T-type  $\text{Ca}^{2+}$  channels prevents myotube fusion [11]. Therefore, in skeletal muscle,  $\text{Ca}_v3.2$ -type  $\text{Ca}^{2+}$  channels appear responsible for the augmentation of basal  $[\text{Ca}^{2+}]_i$  in response to membrane hyperpolarization, consistent with the generation of a window current. Whether T-type  $\text{Ca}^{2+}$  window currents in neurons play similar roles in developmental processes remains an intriguing open question.

### **$\text{Ca}^{2+}$ signaling through co-localization of T-type $\text{Ca}^{2+}$ channels with $\text{Ca}^{2+}$ -dependent ion channels**

$\text{Ca}^{2+}$  entering through T-type  $\text{Ca}^{2+}$  channels can provide selective signaling through coupling with  $\text{Ca}^{2+}$ -dependent ion channels. In several cases, T-type  $\text{Ca}^{2+}$  channels are selectively coupled to  $\text{Ca}^{2+}$ -activated small conductance-type  $\text{K}^+$  (SK) channels [101] (Fig. 3a). In the nRT, the physiological relevance of T-type  $\text{Ca}^{2+}$  to SK channel coupling is apparent at the cellular as well as the large-scale network level. The nRT is a thin inhibitory shell of neurons that is important for sleep rhythm generation and for sensory information transfer [90]. During slow-wave sleep, nRT neurons generate complex oscillatory burst discharges [4, 14, 38]. These rhythmic bursting patterns are strongly dependent on the activation of SK2 channels in between low-threshold bursts [3, 4]. The activation of SK2 channels occurs via  $\text{Ca}^{2+}$  ions entering through T-type  $\text{Ca}^{2+}$  channels in the long and thin dendrites of nRT neurons [29]. The density of T-type  $\text{Ca}^{2+}$  channels is high enough such that a single low-threshold burst leads to an elevation of  $[\text{Ca}^{2+}]_i$  approaching 1  $\mu\text{M}$ , essentially saturating the nearby SK2 channels. The coupling is apparent as SK2 channel activity closely follows the time course of dendritically averaged  $[\text{Ca}^{2+}]_i$  signals and leads to a burst-induced afterhyperpolarization. This silent period allows for the next oscillatory burst, thereby generating the robust oscillatory behavior that is typical of nRT neurons. Accordingly, mice lacking SK2 channels show strongly suppressed electroencephalographic sleep waves and interrupted deep sleep behavior, suggesting an important role for  $\text{Ca}^{2+}$  entry through T-type  $\text{Ca}^{2+}$  channels in boosting sleep oscillations and in stabilizing sleep [29].

In dopaminergic midbrain neurons, T-type  $\text{Ca}^{2+}$  channels are coupled to SK3 channels [116]. Here, T-type to SK3 channel coupling results in an outward  $\text{K}^+$  current that underlies the hyperpolarization necessary for regular, pacemaker-like AP discharge. However, during neonatal development of dopaminergic neurons, T-type  $\text{Ca}^{2+}$  channels are dually coupled not only to SK3 channels, but also to ryanodine receptors (RyRs) [30] (Fig. 3b). Therefore, the  $[\text{Ca}^{2+}]_i$  transient that occurs via spontaneous openings of single T-type  $\text{Ca}^{2+}$  channels is boosted by  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$

**Fig. 3**  $\text{Ca}^{2+}$  signaling of T-type  $\text{Ca}^{2+}$  channels due to co-localization with  $\text{Ca}^{2+}$ -dependent effectors and with the intracellular  $\text{Ca}^{2+}$  handling machinery. **a** Schematic representation of co-localized T-type  $\text{Ca}^{2+}$  channels and SK channels in a membrane and representative oscillatory discharge patterns of nRT [29] and substantia nigra (SN) neurons [116]. **b** Co-localization of the T-type  $\text{Ca}^{2+}$ /SK channel pair with RyRs or SERCAs. In neonatal SN neurons, coupling to RyRs gives rise to spontaneous miniature outward currents [30]. In nRT neurons, coupling to SERCA induces oscillatory dampening [29]. ER Endoplasmic reticulum. **c**  $\text{Ca}^{2+}$ -induced synaptic transmission due to co-localization with the presynaptic release machinery (not shown). A single vesicle fused with the plasma membrane is depicted schematically. Representative pre- and postsynaptic recordings are shown to the right, as adapted from [50]. *IPSC* Inhibitory postsynaptic potential; *Pre* ( $\Delta F/F$ ) Presynaptic changes in fluorescent signal, *Pre* ( $I_{\text{Ca}}$ ) presynaptic  $\text{Ca}^{2+}$  current, *Pre* ( $V_m$ ) presynaptic voltage command



release (CICR, see also the “Coupling of T-type  $\text{Ca}^{2+}$  channels to intracellular  $\text{Ca}^{2+}$  release and sequestration” section) and becomes large enough to gate SK3 channels. The resulting random SK3 channel-dependent miniature outward currents make the spontaneous AP discharge irregular, which is a characteristic signature of neonatal dopamine neurons [30] (Fig. 3b).

In thalamic and habenular neurons, T-type  $\text{Ca}^{2+}$  currents may also be coupled to  $\text{Ca}^{2+}$ -dependent cationic currents, thereby leading to afterdepolarizations following burst discharges [4, 14, 22, 70]. In thalamocortical neurons, the coupling of T-type  $\text{Ca}^{2+}$  channels to cation channels is not due to direct  $\text{Ca}^{2+}$ -mediated activation of the effector channels. Rather, coupling occurs through recruitment of  $\text{Ca}^{2+}$ -dependent adenylyl cyclase and synthesis of cAMP which increases the activity of hyperpolarization-activated

cation-nonspecific (HCN) channels [69]. The resulting membrane depolarization feeds back to limit the recruitment of T-type  $\text{Ca}^{2+}$  channels and causes thalamocortical neurons to fail in the generation of low-threshold bursts [5]. Such cell-intrinsic mechanisms of regulating oscillatory activity are thought to be critical in the periodicity of thalamic network oscillations in vitro, which reproduce fundamental aspects of sleep spindle-related activities [5]. Similar to the co-localization of T-type  $\text{Ca}^{2+}$  and SK2 channels in nRT dendrites, thalamocortical dendrites co-express T-type  $\text{Ca}^{2+}$  channels and HCN channels [102]. The mechanisms of activation of  $\text{Ca}^{2+}$ -dependent cationic currents, such as those in nRT and habenular neurons, remain to be determined [4, 14, 22].

Apart from the CNS, coupling of T-type  $\text{Ca}^{2+}$  channels to large-conductance-type  $\text{Ca}^{2+}$ -activated  $\text{K}^+$  (BK) channels has

been suggested for the  $\text{Ca}_v3.2$  isoform expressed in the vascular smooth muscle of coronary arteries in heart. Mice lacking the  $\text{Ca}_v3.2$  isoform show constricted and malformed arteries, and impaired relaxation responses of arterial vessels [24]. This deficiency in arterial relaxation was proposed to be due to the loss of  $\text{Ca}^{2+}$  entering through  $\text{Ca}_v3.2$  channels to antagonize contraction via activating BK channels.  $\text{Ca}_v3.2$  channel and BK channel proteins from brain tissue co-immunoprecipitate, suggesting that they may co-localize in membranes through molecular interaction to translate  $\text{Ca}^{2+}$  entry rapidly into membrane hyperpolarization. In coronary arteries,  $\text{Ca}^{2+}$  transients due to T-type  $\text{Ca}^{2+}$  channels may mediate vasodilatation.

In addition, the coupling of T-type  $\text{Ca}^{2+}$  channels to BK and SK channels has been reported in a number of bursting neurons, such as cholinergic nucleus basalis neurons [113], intralaminar thalamic neurons [42], Purkinje neurons [104], and cartwheel cells of dorsal cochlear nucleus [60].

### Coupling of T-type $\text{Ca}^{2+}$ channels to intracellular $\text{Ca}^{2+}$ release and sequestration

An emerging aspect of  $\text{Ca}^{2+}$  signaling via T-type  $\text{Ca}^{2+}$  channels is the coupling to intracellular  $\text{Ca}^{2+}$  release and sequestration (Fig. 3b). In addition to immature dopaminergic neurons (see the “[Ca<sup>2+</sup> signaling through co-localization of T-type Ca<sup>2+</sup> channels with Ca<sup>2+</sup>-dependent ion channels](#)” section), T-type  $\text{Ca}^{2+}$  channel coupling to intracellular  $\text{Ca}^{2+}$  handling mechanisms has emerged principally from thalamic neurons (see also [88]). Thalamic nuclei are subdivided into first-order and higher-order nuclei depending on whether they receive their excitatory input from the periphery or from the cortex, respectively [95]. Accordingly, first-order nuclei are considered the classical relay stations of the thalamus, whereas higher-order nuclei are implicated in cortico-thalamo-cortical information transfer. These functionally defined thalamic subdivisions, are accompanied by a nucleus-specific relationship between  $\text{Ca}_v3.1$  channels and CICR. In the visual and somatosensory first-order nuclei, activation of a low-threshold  $\text{Ca}^{2+}$  spike induces rapid  $\text{Ca}^{2+}$  entry and a  $[\text{Ca}^{2+}]_i$  transient that decays on the time scales of the low-threshold current, without coupling to CICR [16, 93]. Tonic firing and HVA  $\text{Ca}^{2+}$  channel activation, however, leads to CICR in these cells [16]. Conversely, in higher-order neurons, a single low-threshold burst is sufficient to induce a tail in the corresponding intracellular  $[\text{Ca}^{2+}]_i$  transient that persists for seconds and that is, at least in part, due to CICR [93]. In nRT neurons,  $\text{Ca}^{2+}$  entry is mediated through  $\text{Ca}_v3.2$  and  $\text{Ca}_v3.3$  channels and, rather than being boosted through CICR, is rapidly sequestered selectively into the endoplasmic reticulum. Thus, the coupling between T-type  $\text{Ca}^{2+}$  and SK2 channels is controlled through the sarco/endoplasmic reticu-

lum  $\text{Ca}^{2+}$ -ATPases (SERCAs). SERCAs and SK2 channels compete for the same pool of  $\text{Ca}^{2+}$  ions entered through T-type  $\text{Ca}^{2+}$  channels, shaping and effectively limiting SK2 channel gating, thereby dampening nRT oscillations [29]. Notably, SERCAs specifically sequester  $\text{Ca}^{2+}$  ions entering through T-type but not through HVA  $\text{Ca}^{2+}$  channels, suggesting that the triad of T-type  $\text{Ca}^{2+}$  channels, SK2 channels, and SERCAs forms a signaling complex that is both self-triggering and self-limiting in rhythm generation.

T-type  $\text{Ca}^{2+}$  channel-induced CICR is also present in the heart. In addition to their major counterpart, the L-type  $\text{Ca}^{2+}$  channels, T-type  $\text{Ca}^{2+}$  channels act as  $\text{Ca}^{2+}$  sources and couple to the sarcoplasmic reticulum to induce CICR [98]. T-type  $\text{Ca}^{2+}$  channel density is highest in pacemaking structures, with a prominent expression of  $\text{Ca}_v3.1$ , whereas low levels are present in ventricular cells [112]. In pacemaking sinoatrial node and atrial cells, T-type  $\text{Ca}^{2+}$  channels contribute to the late phase of diastolic depolarization [44]. During this period, numerous and rapid (20–30 ms) localized  $[\text{Ca}^{2+}]_i$  sparks appear at the sarcolemmal surface, which sum to a pedestal increase in  $[\text{Ca}^{2+}]_i$  at subthreshold potentials that is further boosted by  $\text{Ca}^{2+}$  release from the sarcoplasmic reticulum [48]. Consistent with this finding, mice lacking the  $\text{Ca}_v3.1$   $\text{Ca}^{2+}$  channel subunit show no T-type  $\text{Ca}^{2+}$  currents in pacemaking cardiac tissue, a slowing of the late phase of diastolic depolarization, and bradycardia [74], whereas  $\text{Ca}_v3.2$ -deficient mice showed normal rhythms [24]. Thus, via the amplification of  $[\text{Ca}^{2+}]_i$  through sarcolemmal  $\text{Ca}^{2+}$  release,  $\text{Ca}_v3.1$  channels serve an important role in cardiac pacemaking.

### Involvement of T-type $\text{Ca}^{2+}$ channels in exocytosis

Presynaptic  $\text{Ca}^{2+}$  transients that underlie fast neurotransmitter release are among the largest and most highly localized  $[\text{Ca}^{2+}]_i$  elevations, reaching levels of up to tens of micromolar within <1 ms, on spatial scales of 10–100 nm. While it is well established that presynaptic HVA  $\text{Ca}^{2+}$  channels contribute to fast excitatory neurotransmission, in some neurons, T-type  $\text{Ca}^{2+}$  channels are also involved in neurotransmitter release, engendering a low-threshold component of fast exocytosis [18]. A remarkable case is the graded transmission between reciprocally connected oscillator heart interneurons of leech, which inhibit each other through burst discharges in alternating sequence [50]. T-type  $\text{Ca}^{2+}$  current-dependent burst discharges lead to robust  $\text{Ca}^{2+}$  increases at the presynaptic sites of contact, whereas tonic APs produce comparatively minor  $[\text{Ca}^{2+}]_i$  elevations. The amplitude of T-type  $\text{Ca}^{2+}$  currents correlates well with presynaptic  $[\text{Ca}^{2+}]_i$  dynamics and with graded synaptic transmission (Fig. 3c) [50]. A role for T-type  $\text{Ca}^{2+}$  currents in graded neurotransmission has

also been shown for retinal bipolar cells. These cells transform light-evoked synaptic input from photoreceptor cells into a sustained response and use graded neurotransmitter release to communicate with postsynaptic retinal amacrine and ganglion neurons.  $\text{Ca}^{2+}$  imaging and membrane capacitance measurements, coupled with mibefradil application, identified a role for T-type  $\text{Ca}^{2+}$  currents in triggering vesicle fusion at the giant terminals of bipolar cells [86]. Based on its  $\text{Ni}^{2+}$  and mibefradil sensitivity, it was also proposed that AP-independent excitatory miniature synaptic currents in dorsal spinal chord are triggered by T-type  $\text{Ca}^{2+}$  channels [7].

T-type  $\text{Ca}^{2+}$  currents contribute to different forms of  $\gamma$ -aminobutyric acid (GABA) release in granule cells of the olfactory bulb [34]. These axonless inhibitory neurons exert a lateral inhibition via dendro-dendritic synapses formed with mitral cells, the major output cells of the bulb. Dendro-dendritic interactions may occur in a local mode, in which single synapses between granule and mitral cells enter into a reciprocal communication. Under these conditions,  $\text{Ca}^{2+}$  influx through *N*-methyl-D-aspartate receptors (NMDARs), VGCCs, and release from internal stores all contribute to trigger GABA release from granule cell dendrites [35]. Alternatively, release from GABA can be elicited via backpropagating APs initiated in the granule cell somata [34] or via strong subthreshold synaptic stimulation [35]. These stimuli provoke a global  $[\text{Ca}^{2+}]_i$  increase in numerous distant spines, which is generated predominantly by T-type  $\text{Ca}^{2+}$  currents that evoke the release of GABA along large stretches of dendritic spines. These cells can thus release GABA in an AP-independent manner to exert global lateral inhibition of mitral cells.

Important forms of slow secretion, such as hormone release from neuroendocrine cells, are regulated by T-type  $\text{Ca}^{2+}$  channels [18]. An intriguing aspect is the recruitment of T-type  $\text{Ca}^{2+}$  channels for secretion in chromaffin cells. T-type  $\text{Ca}^{2+}$  channels are weakly expressed under control conditions, but the  $\text{Ca}_v3.2$  subtype is strongly upregulated by day-long exposure to cAMP-producing stimuli or by hypoxia and functionally couples to catecholamine release [17]. Secretion controlled by T-type  $\text{Ca}^{2+}$  channels in these cells is initiated at more hyperpolarized potentials, but couples with equal efficacy and velocity to the release apparatus [40], thus permitting catecholamine release in response to subthreshold stimuli and after stressful conditions.

### Involvement of T-type $\text{Ca}^{2+}$ channels in synaptic plasticity

$\text{Ca}^{2+}$  influx through T-type  $\text{Ca}^{2+}$  channels has been implicated in several forms of synaptic plasticity. In neonatal and young hippocampus, T-type  $\text{Ca}^{2+}$  channels

have been implicated in long-term depression (LTD) induced by metabotropic glutamate receptors (mGluRs) [9, 12, 83, 85]. At least in one of these cases, T-type  $\text{Ca}^{2+}$  channels may also play a role in retrograde signaling mediated by endocannabinoids [83]. Conversely, in mature CA1 pyramidal cells, a contribution of T-type  $\text{Ca}^{2+}$  channels to NMDAR-dependent long-term potentiation (LTP) was reported [107]. T-type  $\text{Ca}^{2+}$  currents could facilitate the generation of dendritic  $\text{Ca}^{2+}$  spikes, thereby promoting the  $\text{Mg}^{2+}$  unblock from the external vestibule of the NMDAR, as shown for spike-timing-dependent plasticity between pairs of layer V cortical neurons [55].

Subthreshold  $\text{Ca}^{2+}$  signaling has been previously suggested to play a role in cerebellar synaptic plasticity [36, 49], and roles for rebound burst discharges generated by T-type  $\text{Ca}^{2+}$  channels have been recently discovered. Low-frequency stimulation of inhibitory afferents onto neurons of the deep cerebellar nuclei, each evoking a rebound burst discharge, lead to LTP or LTD, depending on the number of APs generated during the rebound bursting [1]. Furthermore, when mossy fiber activation was paired with a postsynaptic rebound current, LTP was generated that strongly depended on the relative timing of synaptic stimulation and the rebound current [91]. Possibly,  $\text{Ca}^{2+}$  entry through T-type  $\text{Ca}^{2+}$  channels potentiates a plasticity-promoting  $\text{Ca}^{2+}$  signal generated by the synaptic stimuli at individual contact sites. Such a mechanism would point to an important role of non-synaptic  $[\text{Ca}^{2+}]_i$  signals generated by T-type  $\text{Ca}^{2+}$  channels in some forms of synaptic plasticity.

### Conclusions and outlook

Recent work clearly shows that T-type  $\text{Ca}^{2+}$  currents, beyond their biophysical role as burst generators, produce marked  $[\text{Ca}^{2+}]_i$  elevations that adopt intracellular signaling functions. To make use of this low-conductance, rapidly inactivating  $\text{Ca}^{2+}$  source active at subthreshold voltages, neurons have evolved specific strategies.

First, in a number of cases, T-type  $\text{Ca}^{2+}$  channels are highly expressed in subcellular compartments with comparatively weaker expression of other classes of VGCCs.  $[\text{Ca}^{2+}]_i$  changes are then dominated by T-type  $\text{Ca}^{2+}$  channels in a spatially confined, yet highly efficient manner. Prominent examples are the thin dendrites of nRT neurons [29], the knobs of olfactory sensory neurons [39], and the brushes of vestibulocerebellar unipolar brush cells [33]. Dendrites and spines of olfactory granule neurons also express high densities of T-type  $\text{Ca}^{2+}$  channels, allowing for global subthreshold  $[\text{Ca}^{2+}]_i$  increments in addition to local synaptic ones at single spines [35]. A relatively large T-type  $\text{Ca}^{2+}$  channel density is also present in the distal dendrites



of hippocampal [26, 57, 73] and cortical cells [76] and in dendritic spines of cerebellar Purkinje neurons [49].

Second, targets for  $\text{Ca}^{2+}$  ions entering through T-type  $\text{Ca}^{2+}$  channels are selectively co-expressed with T-type  $\text{Ca}^{2+}$  channels within these compartments. Particularly important are SK channels, BK channels, constituents of CICR, SERCAs, and dendritic vesicular release sites, but this list is likely to expand and may include additional types of ion channels, as well as proteins related to plasticity and development. The functional characteristics of their colocalization with T-type  $\text{Ca}^{2+}$  channels show remarkable cell type specificity. In the case of SK2 channels in nRT [29], the dendritic confinement of SK2 channels permits their selective activation upon a low-threshold  $\text{Ca}^{2+}$  burst. This activation is close to saturating, and SK2 currents are large and slow enough to allow for recovery of T-type  $\text{Ca}^{2+}$  channels. Conversely, in dopaminergic neurons, afterhyperpolarizations mediated by T-type–SK3 channel coupling are more rapid such that a full-fledged low-threshold burst is effectively prevented, and tonic discharge predominates [116]. This suggests that SK channel gating through T-type  $\text{Ca}^{2+}$  channels is tailored to control the deactivation/inactivation of  $\text{Ca}_v3$  channels according to the neuronal function close to specific system. Additional targeting of T-type  $\text{Ca}^{2+}$  currents is found for ER membrane proteins. SERCA-mediated  $\text{Ca}^{2+}$  sequestration in nRT dendrites is selective for  $\text{Ca}^{2+}$  entry through T-type  $\text{Ca}^{2+}$  channels [29]. A preferential localization of SERCAs on dendritic ER protrusions could underlie its selectivity for T-type  $\text{Ca}^{2+}$ , whereas SERCAs may be rare in subcellular compartments expressing HVA  $\text{Ca}^{2+}$  channels. Consistent with this hypothesis, there is evidence for a functional distinction between  $\text{Ca}^{2+}$ -sequestering and non-sequestering elements of the ER [13]. Furthermore, in the case of dopaminergic neurons, developmental maturation of AP discharge regularity is likely accompanied by a reorganization of CICR elements, followed by uncoupling of RyRs from the T-type  $\text{Ca}^{2+}$  channel–SK3 channel pair [30].

Additional aspects of T-type  $\text{Ca}^{2+}$  channel signaling described here are just beginning to be recognized and will progress rapidly with new optical techniques, the availability of genetically modified animals lacking T-type  $\text{Ca}^{2+}$  channel subunits (for review, see [97]) and the advance of small interfering RNA technologies [87]. The distinct properties of channel subtypes are well established, and these shape cell-specific characteristics of low-threshold bursting [15, 47, 79, 82] and of  $[\text{Ca}^{2+}]_i$  signals [63]. In keeping with this, the subcellular distribution of T-type  $\text{Ca}^{2+}$  channel subtypes [78], and subtype-specific  $\text{Ca}^{2+}$  signaling, including the generation of post-AP  $\text{Ca}^{2+}$  entry and window currents is an important aspect of future investigation. The prominent dendritic and spine expression of these channels in numerous cell types suggests that an assessment of their role in dendritic integration of synaptic inputs and in dendritic  $\text{Ca}^{2+}$  spikes

is likely to be important. Depending on expression density and channel subtype, recruitment of T-type  $\text{Ca}^{2+}$  currents can confer local or global dendritic  $[\text{Ca}^{2+}]_i$  signals and could hence contribute to various forms of synaptic plasticity. The rapidly expanding recognition of subtype-specific modulatory pathways on the short-term [23] and long-term ([8, 17]) provides an additional means for varying intracellular  $[\text{Ca}^{2+}]_i$  dynamics. Moreover, T-type  $\text{Ca}^{2+}$  currents can contribute to neurotransmitter release, particularly in neurons with graded or dendritic forms of synaptic release. Such coupling may be more widespread in cells with dendrodendritic coupling in the CNS (see [33, 35]) and could be linked to retrograde signaling through dendritic neuropeptide and endocannabinoid release.

T-type  $\text{Ca}^{2+}$  currents appear to provide crucial signaling functions during developmental periods in which neurons undergo profound changes in morphology, electrophysiology, and synapse formation. Whether these channels facilitate neuronal excitability by providing a subthreshold depolarizing or window current, or whether  $\text{Ca}^{2+}$  entry through these channels carries specific signals remains to be determined. In the mature thalamus, sleep-related rhythmic  $[\text{Ca}^{2+}]_i$  elevations are large and are handled in a cell-specific manner according to thalamocortical connectivity. It is thus tempting to speculate that these  $[\text{Ca}^{2+}]_i$  oscillations may represent an analogue of developmental oscillations that drive plasticity. These may contribute to some of the subcellular mechanisms by which sleep affects neural functions related to synaptic remodeling and plasticity.

Gain-of-function mutations in  $\text{Ca}_v3$  channel genes, or abnormal channel expression patterns, have been linked to states of chronic neuronal hyperexcitability and stress, such as chronic epilepsy, neuropathic pain, tumor formation, and hypoxia [8, 17, 59, 87], whereas genetic ablation confers resistance to seizures [96]. Abnormal  $[\text{Ca}^{2+}]_i$  load in dendrites and perturbed  $\text{Ca}^{2+}$  homeostasis is a central theme in the affected cells. Understanding the contributions of T-type  $\text{Ca}^{2+}$  channels to these pathological states, in particular, how they alter excitability and how they impose or alter neurotransmission, may be important for determining their signaling profile and for identifying potential therapeutic targets to prevent the development of these diseases (see [87]).

As we learn more about the roles for T-type  $\text{Ca}^{2+}$  channels in neurons, we will find that they are important for the cytoplasmic, biochemical, and genetic consequences of particular behavioral states such as sleep, reward actions, learning and development, and for pathological states related to neuronal hyperexcitability.

**Acknowledgements** We acknowledge F. Longordo, Dr. L. Mapelli and Prof. K. Vogt for useful discussions and E. Bernardi for his help with figure preparation. The work of J.P.A. is supported by NIH grants. A.L. is supported by the Swiss National Science Foundation (Nr. 3100A0-116006).

## References

- Aizenman CD, Manis PB, Linden DJ (1998) Polarity of long-term synaptic gain change is related to postsynaptic spike firing at a cerebellar inhibitory synapse. *Neuron* 21:827–835
- Autret L, Mechaly I, Scamps F, Valmier J, Lory P, Desmadryl G (2005) The involvement of  $\text{Ca}_v3.2/\alpha_{1H}$  T-type calcium channels in excitability of mouse embryonic primary vestibular neurones. *J Physiol (Lond)* 567:67–78
- Avanzini G, de Curtis M, Panzica F, Spreafico R (1989) Intrinsic properties of nucleus reticularis thalami neurones of the rat studied in vitro. *J Physiol (Lond)* 416:111–122
- Bal T, McCormick DA (1993) Mechanisms of oscillatory activity in guinea-pig nucleus reticularis thalami in vitro: a mammalian pacemaker. *J Physiol (Lond)* 468:669–691
- Bal T, McCormick DA (1996) What stops synchronized thalamocortical oscillations? *Neuron* 17:297–308
- Bal T, McCormick DA (1997) Synchronized oscillations in the inferior olive are controlled by the hyperpolarization-activated cation current  $I_h$ . *J Neurophysiol* 77:3145–3156
- Bao J, Li JJ, Perl ER (1998) Differences in  $\text{Ca}^{2+}$  channels governing generation of miniature and evoked excitatory synaptic currents in spinal laminae I and II. *J Neurosci* 18:8740–8750
- Beck H, Yaari Y (2008) Plasticity of intrinsic neuronal properties in CNS disorders. *Nat Rev Neurosci* 9:357–369
- Bender VA, Bender KJ, Brasier DJ, Feldman DE (2006) Two coincidence detectors for spike timing-dependent plasticity in somatosensory cortex. *J Neurosci* 26:4166–4177
- Beurrier C, Congar P, Bioulac B, Hammond C (1999) Subthalamic nucleus neurons switch from single-spike activity to burst-firing mode. *J Neurosci* 19:599–609
- Bijlenga P, Liu JH, Espinos E, Haenggeli CA, Fischer-Lougheed J, Bader CR, Bernheim L (2000) T-type  $\alpha_{1H}$   $\text{Ca}^{2+}$  channels are involved in  $\text{Ca}^{2+}$  signaling during terminal differentiation (fusion) of human myoblasts. *Proc Natl Acad Sci USA* 97:7627–7632
- Birtoli B, Ulrich D (2004) Firing mode-dependent synaptic plasticity in rat neocortical pyramidal neurons. *J Neurosci* 24:4935–4940
- Blaustein MP, Golovina VA (2001) Structural complexity and functional diversity of endoplasmic reticulum  $\text{Ca}^{2+}$  stores. *Trends Neurosci* 24:602–608
- Blethyn KL, Hughes SW, Toth TI, Cope DW, Crunelli V (2006) Neuronal basis of the slow (<1 Hz) oscillation in neurons of the nucleus reticularis thalami in vitro. *J Neurosci* 26:2474–2486
- Broicher T, Kanyshkova T, Landgraf P, Rankovic V, Meuth P, Meuth SG, Pape HC, Budde T (2007) Specific expression of low-voltage-activated calcium channel isoforms and splice variants in thalamic local circuit interneurons. *Mol Cell Neurosci* 36:132–145
- Budde T, Sieg F, Braunewell KH, Gundelfinger ED, Pape HC (2000)  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release supports the relay mode of activity in thalamocortical cells. *Neuron* 26:483–492
- Carabelli V, Marcantoni A, Comunanza V, de Luca A, Diaz J, Borges R, Carbone E (2007) Chronic hypoxia up-regulates  $\alpha_{1H}$  T-type channels and low-threshold catecholamine secretion in rat chromaffin cells. *J Physiol* 584:149–165
- Carbone E, Giacippoli A, Marcantoni A, Guido D, Carabelli V (2006) A new role for T-type channels in fast “low-threshold” exocytosis. *Cell Calcium* 40:147–154
- Catterall WA, Perez-Reyes E, Snutch TP, Striessnig J (2005) International Union of Pharmacology. XLVIII. Nomenclature and structure–function relationships of voltage-gated calcium channels. *Pharmacol Rev* 57:411–425
- Cavelier P, Lohof AM, Lonchamp E, Beekenkamp H, Mariani J, Bossu JL (2008) Participation of low-threshold  $\text{Ca}^{2+}$  spike in the Purkinje cells complex spike. *Neuroreport* 19:299–303
- Chambard JM, Chabbert C, Sans A, Desmadryl G (1999) Developmental changes in low and high voltage-activated calcium currents in acutely isolated mouse vestibular neurons. *J Physiol (Lond)* 518:141–149
- Chang S-Y, Kim U (2004) Ionic mechanism of long-lasting discharges of action potentials triggered by membrane hyperpolarization in the medial lateral habenula. *J Neurosci* 24:2172–2181
- Chemin J, Traboulsie A, Lory P (2006) Molecular pathways underlying the modulation of T-type calcium channels by neurotransmitters and hormones. *Cell Calcium* 40:121–134
- Chen CC, Lamping KG, Nuno DW, Barresi R, Prouty SJ, Lavoie JL, Cribbs LL, England SK, Sigmund CD, Weiss RM, Williamson RA, Hill JA, Campbell KP (2003) Abnormal coronary function in mice deficient in  $\alpha_{1H}$  T-type  $\text{Ca}^{2+}$  channels. *Science* 302:1416–1418
- Chevalier M, Lory P, Mironneau C, Macrez N, Quignard JF (2006) T-type  $\text{Ca}_v3.3$  calcium channels produce spontaneous low-threshold action potentials and intracellular calcium oscillations. *Eur J Neurosci* 23:2321–2329
- Christie BR, Eliot LS, Ito K, Miyakawa H, Johnston D (1995) Different  $\text{Ca}^{2+}$  channels in soma and dendrites of hippocampal pyramidal neurons mediate spike-induced  $\text{Ca}^{2+}$  influx. *J Neurophysiol* 73:2553–2557
- Crunelli V, Cope DW, Hughes SW (2006) Thalamic T-type  $\text{Ca}^{2+}$  channels and NREM sleep. *Cell Calcium* 40:175–190
- Crunelli V, Toth TI, Cope DW, Blethyn K, Hughes SW (2005) The ‘window’ T-type calcium current in brain dynamics of different behavioural states. *J Physiol (Lond)* 562:121–129
- Cueni L, Canepari M, Luján R, Emmenegger Y, Watanabe M, Bond CT, Franken P, Adelman JP, Lüthi A (2008) T-type  $\text{Ca}^{2+}$  channels, SK2 channels and SERCAs gate sleep-related oscillations in thalamic dendrites. *Nat Neurosci* 11:683–692
- Cui G, Okamoto T, Morikawa H (2004) Spontaneous opening of T-type  $\text{Ca}^{2+}$  channels contributes to the irregular firing of dopamine neurons in neonatal rats. *J Neurosci* 24:11079–11087
- Destexhe A, Contreras A, Steriade M, Sejnowski TJ, Huguenard JR (1996) In vivo, in vitro, and computational analysis of dendritic calcium currents in thalamic reticular neurons. *J Neurosci* 16:169–185
- Destexhe A, Neubig M, Ulrich D, Huguenard J (1998) Dendritic low-threshold calcium currents in thalamic relay cells. *J Neurosci* 18:3574–3588
- Diana MA, Otsu Y, Maton G, Collin T, Chat M, Dieudonne S (2007) T-type and L-type  $\text{Ca}^{2+}$  conductances define and encode the bimodal firing pattern of vestibulocerebellar unipolar brush cells. *J Neurosci* 27:3823–3838
- Egger V, Svoboda K, Mainen ZF (2003) Mechanisms of lateral inhibition in the olfactory bulb: efficiency and modulation of spike-evoked calcium influx into granule cells. *J Neurosci* 23:7551–7558
- Egger V, Svoboda K, Mainen ZF (2005) Dendrodendritic synaptic signals in olfactory bulb granule cells: local spine boost and global low-threshold spike. *J Neurosci* 25:3521–3530
- Eilers J, Augustine GJ, Konnerth A (1995) Subthreshold synaptic  $\text{Ca}^{2+}$  signalling in fine dendrites and spines of cerebellar Purkinje neurons. *Nature* 373:155–158
- Feltz A (2006) Low-threshold-activated Ca channels: from molecules to functions: over 25 years of progress. *Crit Rev Neurobiol* 18:169–178
- Fuentealba P, Timofeev I, Steriade M (2004) Prolonged hyperpolarizing potentials precede spindle oscillations in the thalamic reticular nucleus. *Proc Natl Acad Sci USA* 101:9816–9821

39. Gautam SH, Otsuguro KI, Ito S, Saito T, Habara Y (2007) T-type  $\text{Ca}^{2+}$  channels mediate propagation of odor-induced  $\text{Ca}^{2+}$  transients in rat olfactory receptor neurons. *Neuroscience* 144:702–713
40. Giacippoli A, Novara M, de Luca A, Baldelli P, Marcantoni A, Carbone E, Carabelli V (2006) Low-threshold exocytosis induced by cAMP-recruited  $\text{Ca}_v3.2$  ( $\alpha_{1H}$ ) channels in rat chromaffin cells. *Biophys J* 90:1830–1841
41. Gillissen T, Alzheimer C (1997) Amplification of EPSPs by low  $\text{Ni}^{2+}$ —and amiloride-sensitive  $\text{Ca}^{2+}$  channels in apical dendrites of rat CA1 pyramidal neurons. *J Neurophysiol* 77:1639–1643
42. Goillard J-M, Vincent P (2002) Serotonin suppresses the slow afterhyperpolarization in rat intralaminar and midline neurones by activating 5 HT-7 receptors. *J Physiol (Lond)* 541:453–465
43. Gu X, Spitzer NC (1993) Low-threshold  $\text{Ca}^{2+}$  current and its role in spontaneous elevations of intracellular  $\text{Ca}^{2+}$  in developing *Xenopus* neurons. *J Neurosci* 13:4936–4948
44. Hagiwara N, Irisawa H, Kameyama M (1988) Contribution of two types of calcium currents to the pacemaker potentials of rabbit sino-atrial node cells. *J Physiol (Lond)* 395:233–253
45. Heppenstall PA, Lewin GR (2006) A role for T-type  $\text{Ca}^{2+}$  channels in mechanosensation. *Cell Calcium* 40:165–174
46. Hughes SW, Cope DW, Blythyn KL, Crunelli V (2002) Cellular mechanisms of the slow (<1 Hz) oscillation in thalamocortical neurons in vitro. *Neuron* 33:947–958
47. Huguenard JR (1996) Low-threshold calcium currents in central nervous system neurons. *Annu Rev Physiol* 58:329–348
48. Hüser J, Blatter LA, Lipsius SL (2000) Intracellular  $\text{Ca}^{2+}$  release contributes to automaticity in cat atrial pacemaker cells. *J Physiol (Lond)* 524:415–422
49. Isope P, Murphy TH (2005) Low threshold calcium currents in rat cerebellar Purkinje cell dendritic spines are mediated by T-type calcium channels. *J Physiol (Lond)* 562:257–269
50. Ivanov AI, Calabrese RL (2000) Intracellular  $\text{Ca}^{2+}$  dynamics during spontaneous and evoked activity of leech heart interneurons: low-threshold Ca currents and graded synaptic transmission. *J Neurosci* 20:4930–4943
51. Jevtovic-Todorovic V, Todorovic SM (2006) The role of peripheral T-type calcium channels in pain transmission. *Cell Calcium* 40:197–203
52. Joksovic PM, Bayliss DA, Todorovic SM (2005) Different kinetic properties of two T-type  $\text{Ca}^{2+}$  currents of rat reticular thalamic neurones and their modulation by enflurane. *J Physiol (Lond)* 566:125–142
53. Jones SW (2003) Calcium channels: unanswered questions. *J Bioenerg Biomembr* 35:461–475
54. Jorntell H, Hansel C (2006) Synaptic memories upside down: bidirectional plasticity at cerebellar parallel fiber-Purkinje cell synapses. *Neuron* 52:227–238
55. Kampa BM, Letzkus JJ, Stuart GJ (2006) Requirement of dendritic calcium spikes for induction of spike-timing-dependent synaptic plasticity. *J Physiol (Lond)* 574:283–290
56. Karst H, Joels M, Wadman WJ (1993) Low-threshold calcium current in dendrites of the adult rat hippocampus. *Neurosci Lett* 164:154–158
57. Kavalali ET, Zhuo M, Bito H, Tsien RW (1997) Dendritic  $\text{Ca}^{2+}$  channels characterized by recordings from isolated hippocampal dendritic segments. *Neuron* 18:651–663
58. Kawai F, Miyachi E (2001) Enhancement by T-type  $\text{Ca}^{2+}$  currents of odor sensitivity in olfactory receptor cells. *J Neurosci* 21:RC144, (141–145)
59. Khosravani H, Zamponi GW (2006) Voltage-gated calcium channels and idiopathic generalized epilepsies. *Physiol Rev* 86:941–966
60. Kim Y, Trussell LO (2007) Ion channels generating complex spikes in cartwheel cells of the dorsal cochlear nucleus. *J Neurophysiol* 97:1705–1725
61. Kiryushko D, Korshunova I, Berezin V, Bock E (2006) Neural cell adhesion molecule induces intracellular signaling via multiple mechanisms of  $\text{Ca}^{2+}$  homeostasis. *Mol Biol Cell* 17:2278–2286
62. König S, Béguet A, Bader CR, Bernheim L (2006) The calcineurin pathway links hyperpolarization (Kir2.1)-induced  $\text{Ca}^{2+}$  signals to human myoblast differentiation and fusion. *Development* 133:3107–3114
63. Kozlov AS, McKenna F, Lee JH, Cribbs LL, Perez-Reyes E, Feltz A, Lambert RC (1999) Distinct kinetics of cloned T-type  $\text{Ca}^{2+}$  channels lead to differential  $\text{Ca}^{2+}$  entry and frequency-dependence during mock action potentials. *Eur J Neurosci* 11:4149–4158
64. Krahe R, Gabbiani F (2004) Burst firing in sensory systems. *Nature Rev Neurosci* 5:13–23
65. Kuisle M, Wanaverbecq N, Brewster AL, Frère SG, Pinault D, Baram TZ, Lüthi A (2006) Functional stabilization of weakened thalamic pacemaker channel regulation in rat absence epilepsy. *J Physiol (Lond)* 575:83–100
66. Lee JH, Daud AN, Cribbs LL, Lacerda AE, Pereverzev A, Klockner U, Schneider T, Perez-Reyes E (1999) Cloning and expression of a novel member of the low voltage-activated T-type calcium channel family. *J Neurosci* 19:1912–1921
67. Levic S, Nie L, Tuteja D, Harvey M, Sokolowski BH, Yamoah EN (2007) Development and regeneration of hair cells share common functional features. *Proc Natl Acad Sci USA* 104:19108–19113
68. Lory P, Bidaud I, Chemin J (2006) T-type calcium channels in differentiation and proliferation. *Cell Calcium* 40:135–146
69. Lüthi A, McCormick DA (1998) H-current: properties of a neuronal and network pacemaker. *Neuron* 21:9–12
70. Lüthi A, McCormick DA (1998) Periodicity of thalamic synchronized oscillations: the role of  $\text{Ca}^{2+}$ -mediated upregulation of  $I_h$ . *Neuron* 20:553–563
71. Magee JC (2000) Dendritic integration of excitatory synaptic input. *Nat Rev Neurosci* 1:181–190
72. Magee JC, Christofi G, Miyakawa H, Christie B, Lasser-Ross N, Johnston D (1995) Subthreshold synaptic activation of voltage-gated  $\text{Ca}^{2+}$  channels mediates a localized  $\text{Ca}^{2+}$  influx into the dendrites of hippocampal pyramidal neurons. *J Neurophysiol* 74:1335–13342
73. Magee JC, Johnston D (1995) Characterization of single voltage-gated  $\text{Na}^+$  and  $\text{Ca}^{2+}$  channels in apical dendrites of rat CA1 pyramidal neurons. *J Physiol (Lond)* 487:67–90
74. Mangoni ME, Traboulsie A, Leoni AL, Couette B, Marger L, Le Quang K, Kupfer E, Cohen-Solal A, Vilar J, Shin HS, Escande D, Charpentier F, Nargeot J, Lory P (2006) Bradycardia and slowing of the atrioventricular conduction in mice lacking  $\text{Ca}_v3.1/\alpha_{1G}$  T-type calcium channels. *Circ Res* 98:1422–1430
75. Markram H, Helm PJ, Sakmann B (1995) Dendritic calcium transients evoked by single back-propagating action potentials in rat neocortical pyramidal neurons. *J Physiol (Lond)* 485:1–20
76. Markram H, Sakmann B (1994) Calcium transients in dendrites of neocortical neurons evoked by single subthreshold excitatory postsynaptic potentials via low-voltage-activated calcium channels. *Proc Natl Acad Sci USA* 91:5207–5211
77. McCobb DP, Beam KG (1991) Action potential waveform voltage-clamp commands reveal striking differences in calcium entry via low and high voltage-activated calcium channels. *Neuron* 7:119–127
78. McKay BE, McRory JE, Molineux ML, Hamid J, Snutch TP, Zamponi GW, Turner RW (2006)  $\text{Ca}_v3$  T-type calcium channel isoforms differentially distribute to somatic and dendritic compartments in rat central neurons. *Eur J Neurosci* 24:2581–2594

79. Molineux ML, McRory JE, McKay BE, Hamid J, Mehaffey WH, Rehak R, Snutch TP, Zamponi GW, Turner RW (2006) Specific T-type calcium channel isoforms are associated with distinct burst phenotypes in deep cerebellar nuclear neurons. *Proc Natl Acad Sci USA* 103:5555–5560
80. Mougnot D, Bossu JL, Gähwiler BH (1997) Low-threshold  $\text{Ca}^{2+}$  currents in dendritic recordings from Purkinje cells in rat cerebellar slice cultures. *J Neurosci* 17:160–170
81. Munsch T, Budde T, Pape HC (1997) Voltage-activated intracellular calcium transients in thalamic relay cells and interneurons. *Neuroreport* 8:2411–2418
82. Murbartian J, Arias JM, Perez-Reyes E (2004) Functional impact of alternative splicing of human T-type  $\text{Ca}_v3.3$  calcium channels. *J Neurophysiol* 92:3399–3407
83. Nevian T, Sakmann B (2006) Spine  $\text{Ca}^{2+}$  signaling in spike-timing-dependent plasticity. *J Neurosci* 26:11001–11013
84. Nilius B, Talavera K, Verkhratsky A (2006) T-type calcium channels: the never ending story. *Cell Calcium* 40:81–88
85. Oliet SHR, Malenka RC, Nicoll RA (1997) Two distinct forms of long-term depression coexist in CA1 hippocampal pyramidal cells. *Neuron* 18:969–982
86. Pan ZH, Hu HJ, Perring P, Andrade R (2001) T-type  $\text{Ca}^{2+}$  channels mediate neurotransmitter release in retinal bipolar cells. *Neuron* 32:89–98
87. Panner A, Wurster RD (2006) T-type calcium channels and tumor proliferation. *Cell Calcium* 40:253–259
88. Pape HC, Munsch T, Budde T (2004) Novel vistas of calcium-mediated signalling in the thalamus. *Pflügers Arch* 448:131–138
89. Perez-Reyes E (2003) Molecular physiology of low-voltage-activated T-type calcium channels. *Physiol Rev* 83:117–161
90. Pinault D (2004) The thalamic reticular nucleus: structure, function and concept. *Brain Res Brain Res Rev* 46:1–31
91. Pugh JR, Raman IM (2006) Potentiation of mossy fiber EPSCs in the cerebellar nuclei by NMDA receptor activation followed by postinhibitory rebound current. *Neuron* 51:113–123
92. Randall AD, Tsien RW (1997) Contrasting biophysical and pharmacological properties of T-type and R-type calcium channels. *Neuropharmacology* 36:879–893
93. Richter TA, Kolaj M, Renaud LP (2005) Low voltage-activated  $\text{Ca}^{2+}$  channels are coupled to  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release in rat thalamic midline neurons. *J Neurosci* 25:8267–8271
94. Richter TA, Kolaj M, Renaud LP (2006) Heterogeneity in low voltage-activated  $\text{Ca}^{2+}$  channel-evoked  $\text{Ca}^{2+}$  responses within neurons of the thalamic paraventricular nucleus. *Eur J Neurosci* 24:1316–1324
95. Sherman SM (2007) The thalamus: more than just a relay. *Curr Opin Neurobiol* 17:417–422
96. Shin HS (2006) T-type  $\text{Ca}^{2+}$  channels and absence epilepsy. *Cell Calcium* 40:191–196
97. Shin HS, Cheong E-J, Choi S, Lee J, Na HS (2008) T-type  $\text{Ca}^{2+}$  channels as therapeutic targets in the nervous system. *Curr Opin Pharmacol* 8:33–41
98. Sipido KR, Cameliet E, Van de Werf F (1998) T-type  $\text{Ca}^{2+}$  current as a trigger for  $\text{Ca}^{2+}$  release from the sarcoplasmic reticulum in guinea-pig ventricular myocytes. *J Physiol (Lond)* 508:439–451
99. Sochivko D, Pereverzev A, Smyth N, Gissel C, Schneider T, Beck H (2002) The  $\text{Ca}_v2.3$   $\text{Ca}^{2+}$  channel subunit contributes to R-type  $\text{Ca}^{2+}$  currents in murine hippocampal and neocortical neurones. *J Physiol (Lond)* 542:699–710
100. Sotty F, Danik M, Manseau F, Laplante F, Quirion R, Williams S (2003) Distinct electrophysiological properties of glutamatergic, cholinergic and GABAergic rat septohippocampal neurons: novel implications for hippocampal rhythmicity. *J Physiol (Lond)* 551:927–943
101. Stocker M (2004)  $\text{Ca}^{2+}$ -activated  $\text{K}^{+}$  channels: molecular determinants and function of the SK family. *Nat Rev Neurosci* 5:758–770
102. Stuart GJ, Williams SR (2000) Co-localization of  $\text{I}_H$  and  $\text{I}_T$  channels in dendrites of thalamocortical neurons. *Soc Neurosci Abstr* 30:610–619
103. Su H, Sochivko D, Becker A, Chen J, Jiang Y, Yaari Y, Beck H (2002) Upregulation of a T-type  $\text{Ca}^{2+}$  channel causes a long-lasting modification of neuronal firing mode after status epilepticus. *J Neurosci* 22:3645–3655
104. Swensen AM, Bean BP (2003) Ionic mechanisms of burst firing in dissociated Purkinje neurons. *J Neurosci* 23:9650–9663
105. Talavera K, Nilius B (2006) Biophysics and structure-function relationship of T-type  $\text{Ca}^{2+}$  channels. *Cell Calcium* 40:97–114
106. Talley EM, Cribbs LL, Lee JH, Daud A, Perez-Reyes E, Bayliss DA (1999) Differential distribution of three members of a gene family encoding low voltage-activated (T-type) calcium channels. *J Neurosci* 19:1895–1911
107. Thomas MJ, Watabe AM, Moody TD, Makhinson M, O'Dell TJ (1998) Postsynaptic complex spike bursting enables the induction of LTP by theta frequency synaptic stimulation. *J Neurosci* 18:7118–7126
108. Tottene A, Volsen S, Pietrobon D (2000)  $\alpha_{1E}$  subunits form the pore of three cerebellar R-type calcium channels with different pharmacological and permeation properties. *J Neurosci* 20:171–178
109. Tsakiridou E, Bertolini L, de Curtis M, Avanzini G, Pape HC (1995) Selective increase in T-type calcium conductance of reticular thalamic neurons in a rat model of absence epilepsy. *J Neurosci* 15:3110–3117
110. Turner JP, Leresche N, Guyon A, Soltesz I, Crunelli V (1994) Sensory input and burst firing output of rat and cat thalamocortical cells: the role of NMDA and non-NMDA receptors. *J Physiol (Lond)* 480:281–295
111. Uebachs M, Schaub C, Perez-Reyes E, Beck H (2006) T-type  $\text{Ca}^{2+}$  channels encode prior neuronal activity as modulated recovery rates. *J Physiol (Lond)* 571:519–536
112. Vassort G, Talavera K, Alvarez JL (2006) Role of T-type  $\text{Ca}^{2+}$  channels in the heart. *Cell Calcium* 40:205–220
113. Williams S, Serafin M, Mühlenthaler M, Bernheim L (1997) Distinct contributions of high- and low-voltage-activated calcium currents to afterhyperpolarizations in cholinergic nucleus basalis neurons of the guinea pig. *J Neurosci* 17:7307–7315
114. Williams SR, Stuart GJ (2000) Action potential backpropagation and somato-dendritic distribution of ion channels in thalamocortical neurons. *J Neurosci* 20:1307–1317
115. Williams SR, Toth TI, Turner JP, Hughes SW, Crunelli V (1997) The 'window' component of the low-threshold  $\text{Ca}^{2+}$  current produces input signal amplification and bistability in cat and rat thalamocortical neurones. *J Physiol (Lond)* 505:689–705
116. Wolfart J, Roeper J (2002) Selective coupling of T-type calcium channels to SK potassium channels prevents intrinsic bursting in dopaminergic midbrain neurons. *J Neurosci* 22:3404–3413
117. Yaari Y, Yue C, Su H (2007) Recruitment of apical dendritic T-type  $\text{Ca}^{2+}$  channels by backpropagating spikes underlies de novo intrinsic bursting in hippocampal epileptogenesis. *J Physiol (Lond)* 580:435–450
118. Zhou Q, Godwin DW, O'Malley DM, Adams PR (1997) Visualization of calcium influx through channels that shape the burst and tonic firing modes of thalamic relay cells. *J Neurophysiol* 77:2816–2825