

EXPERT REVIEW

From gut dysbiosis to altered brain function and mental illness: mechanisms and pathways

GB Rogers¹, DJ Keating², RL Young³, M-L Wong⁴, J Licinio⁴ and S Wesselingh¹

The human body hosts an enormous abundance and diversity of microbes, which perform a range of essential and beneficial functions. Our appreciation of the importance of these microbial communities to many aspects of human physiology has grown dramatically in recent years. We know, for example, that animals raised in a germ-free environment exhibit substantially altered immune and metabolic function, while the disruption of commensal microbiota in humans is associated with the development of a growing number of diseases. Evidence is now emerging that, through interactions with the gut–brain axis, the bidirectional communication system between the central nervous system and the gastrointestinal tract, the gut microbiome can also influence neural development, cognition and behaviour, with recent evidence that changes in behaviour alter gut microbiota composition, while modifications of the microbiome can induce depressive-like behaviours. Although an association between enteropathy and certain psychiatric conditions has long been recognized, it now appears that gut microbes represent direct mediators of psychopathology. Here, we examine roles of gut microbiome in shaping brain development and neurological function, and the mechanisms by which it can contribute to mental illness. Further, we discuss how the insight provided by this new and exciting field of research can inform care and provide a basis for the design of novel, microbiota-targeted, therapies.

Molecular Psychiatry (2016) **21**, 738–748; doi:10.1038/mp.2016.50; published online 19 April 2016

INTRODUCTION

The disruption of the microbes that are resident in our gastrointestinal tract has long been implicated in the development or exacerbation of mental disorders. There is, for example, a long history of anecdotal reports of psychiatric side-effects of antibiotics, even in those without a premorbid psychiatric history.¹ There have also been attempts to influence the composition of the gut microbiota to achieve clinical benefit. For example, in the first decades of the twentieth century, probiotic preparations containing *Lactobacillus* strains were marketed widely as a means to improve mental health or treat psychiatric disorders.² These approaches fell from favour in the 1920s because of a lack of mechanistic understanding and their link to the increasingly unfashionable ‘auto-intoxication’ model. However, the interest in the role of gut microbes in mental health, and our ability to improve psychiatric wellbeing through their manipulation, is resurgent.^{2,3}

In this review, we consider the potential of dysbiosis to contribute to psychopathology and the evidence linking disruption of gut microbiota with specific psychiatric disorders. We examine the role of the microbiome in neurological development and regulation, and consider its contribution to aging-related morbidity. Finally, we discuss the potential for modification of the gut microbiome to provide clinical benefit in the context of altered brain function.

REGULATION OF NEUROLOGICAL FUNCTION BY THE GUT MICROBIOME

The potential contribution of bidirectional communication between the gut and central nervous system (CNS) is suggested

by high rates of comorbidity between gastrointestinal and psychiatric illnesses.^{4,5} For example, mood disorders affect more than half of all patients with irritable bowel syndrome,⁶ with antidepressants being one of the most common pharmaceutical interventions for irritable bowel syndrome.⁴ The gut–brain axis consists of a bidirectional communication network that monitors and integrates gut functions and link them to cognitive and emotional centres of the brain. It encompasses the central, autonomic and enteric nervous systems, as well as the neuro-endocrine, enteroendocrine and neuroimmune systems.^{7,8} It mediates the effects of both genetic and environmental factors on brain development and function, and has been implicated in the aetiology of a number of psychiatric disorders.^{9–12}

In recent years, we have increasingly understood the contribution made by the gut microbiome not only in the regulation of host physiology, particularly metabolism and immunity,^{13–17} but also the CNS and brain function.^{11,18,19} Given mounting evidence that the microbiome has a key role in influencing the development and function of the nervous system through its interaction with the gut–brain axis, it has been suggested that a ‘microbiome–gut–brain axis’ may be a more appropriate model.^{19–22}

The delicate balance between the human microbiome and the development of psychopathologies is particularly interesting given the ease with which the microbiome can be altered by external factors, such as diet,²³ exposure to antimicrobials^{24,25} or disrupted sleep patterns.²⁶ For example, a link between antibiotic exposure and altered brain function is well evidenced by the psychiatric side-effects of antibiotics, which range from anxiety

¹South Australian Health and Medical Research Institute, Infection and Immunity Theme, School of Medicine, Flinders University, Adelaide, SA, Australia; ²South Australian Health and Medical Research Institute, Centre for Neuroscience and Department of Human Physiology, Flinders University, Adelaide, SA, Australia; ³South Australian Health and Medical Research Institute, Department of Medicine, University of Adelaide, Adelaide, SA, Australia and ⁴South Australian Health and Medical Research Institute, Mind and Brain Theme, and Flinders University, Adelaide, SA, Australia. Correspondence: Professor GB Rogers, Microbiology and Infectious Diseases, 5D332, Flinders Medical Centre, Flinders Drive, Bedford Park, Adelaide 5042, SA, Australia.

E-mail: geraint.rogers@sahmri.com

Received 15 February 2016; revised 22 February 2016; accepted 25 February 2016; published online 19 April 2016

and panic to major depression, psychosis and delirium.¹ A recent large population study reported that treatment with a single antibiotic course was associated with an increased risk for depression and anxiety, rising with multiple exposures.²⁷ Bercik *et al.*²⁸ showed that oral administration of non-absorbable antimicrobials transiently altered the composition of the gut microbiota in adult mice and increased exploratory behaviour and hippocampal expression of brain-derived neurotrophic factor (BDNF), while intraperitoneal administration had no effect on behaviour. Alteration of brain function may therefore add to the many reasons that inappropriate antibiotic use should be avoided. It should be noted though that unchecked bacterial infection also represents an acute stressor, and has been shown to be associated with memory dysfunction in mice.²⁹

Diet is another important determinant of gut microbiota composition and function that is strongly linked with psychopathological outcomes. For example, consumption of high fat diet (HFD) is associated with altered microbial diversity and reduced synaptic plasticity,^{30–31} with increased vulnerability to anxiety-like behaviour in mice,³² while altered microbial diversity upon consumption of a diet high in sucrose results in significantly impaired development of a spatial bias for long-term memory, short-term memory and reversal training.³³ In contrast, adolescent rats fed a low-calorie diet show augmented neurogenesis and BDNF levels, and improved cognition in adulthood,³⁴ and a diet that increases microbiota diversity is associated with improved cognitive ability.³⁵ Although human data have shown reduced microbial diversity in individuals is linked with increased adiposity, insulin resistance, dyslipidaemia and more pronounced inflammatory phenotype,^{36,37} strong evidence of a direct microbiome effect comes from studies using conventionally housed mice subjected to a microbiome depletion and/or transplantation paradigm using microbiota isolated from donors on either an HFD or control diet. Following re-colonization, mice given the HFD exposed microbiota showed significant and selective disruptions in exploratory, cognitive, and stereotypical behaviour.³⁸ Although it is not possible to exclude the direct effect of host metabolism on brain function, such findings do suggest that diet-induced changes in the intestinal microbiome substantially influence brain function.

Diet and antibiotic exposure are only two factors that potentially influence brain function through shaping the gut microbiome (Figure 1). An array of common variables may be equally important. For example, alcohol consumption,^{39,40} smoking habits⁴¹ and disruption of diurnal rhythm,²⁶ have all been shown to substantially affect microbiota composition. As such, how wider influences on the microbiome contribute to dysregulation of brain function is an area of growing interest.

THE MICROBIOME IN SPECIFIC PSYCHIATRIC CONDITIONS

While the links between the microbiome and specific psychiatric conditions have been reviewed elsewhere,^{18,42–45} a brief examination of the contribution of inter-kingdom interactions to two particularly distressing neuropsychiatric disorders provides a useful illustration.

Major depressive disorder (MDD) is typified by markers known to be influenced by the microbiome. For example, depression-associated changes seen in the hypothalamic-pituitary-adrenal (HPA) stress response,⁴⁶ and altered levels of depression-associated monoamines (or their receptors) in corticolimbic regions of the brain, have both been demonstrated in germ-free (GF) mice.^{28,47–50} The increased concentrations of pro-inflammatory cytokines seen in MDD⁴⁶ may also result from interactions with gut microbes. Levels of serum antibodies against lipopolysaccharide from gram-negative enterobacteria are higher in patients with MDD than in controls,⁵¹ and cause stress-associated with increased gut permeability and bacterial

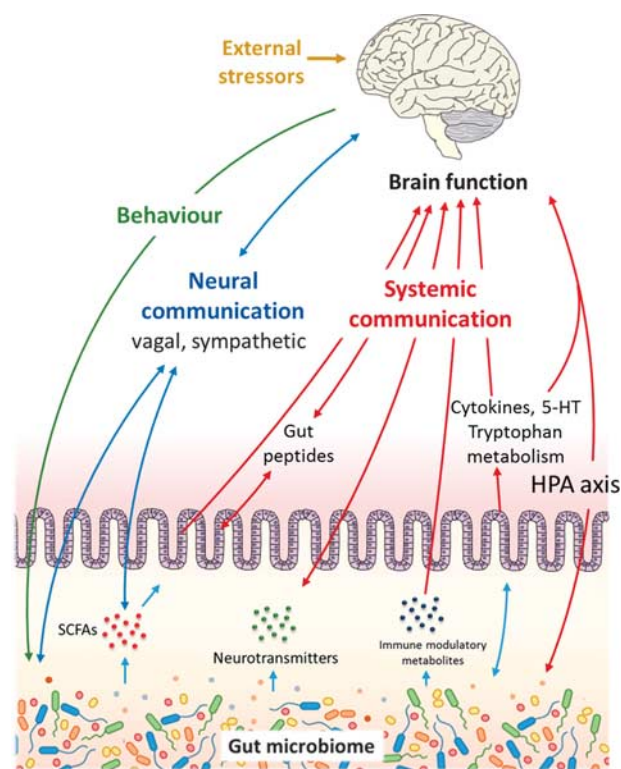


Figure 1. Communication pathways linking the gut microbiome with brain function.

translocation in animal models.^{22,52} Evidence also exists that depression alters the gut microbiota, as demonstrated in mice in which chronic depression- and anxiety-like behaviours has been induced by olfactory bulbectomy,⁵³ suggesting a feedback loop between depressive states and dysbiosis. A reflection of the importance of this circular relationship may be the existence of host mechanisms that regulate microbiota composition.^{54,55}

Similar parallels between dysbiosis and psychopathogenesis exist in schizophrenia (reviewed by Dinan *et al.*⁵⁶ and Nemani *et al.*⁴⁴). Many of the strongest associations identified between genetic risk and schizophrenia relate to genes involved in immunity,^{57,58} paralleling clinical studies that report an upregulated immune and inflammatory status in schizophrenia patients.^{59–68} Serological markers of bacterial translocation are also substantially elevated in schizophrenia subjects and significantly correlated with systemic inflammatory markers.⁶⁹ In turn, cytokine levels are correlated with the severity of clinical symptoms,^{59,70,71} and it has been suggested that the resulting neuroinflammation is involved directly in schizophrenia pathogenesis.^{72–74}

As described later, the microbiota also modulate a range of neurotrophins and proteins involved in brain development and plasticity.^{48,49,75} There is evidence that such alterations are central to the pathophysiology of schizophrenia. For example, BDNF expression is believed to have a role in the molecular mechanism underlying altered cognition,⁷⁶ and through its influence on brain plasticity, may contribute to the *N*-methyl-D-aspartate receptor dysfunction seen in schizophrenia.⁷⁷

TREATMENT INTERACTIONS WITH THE MICROBIOME IN MENTAL ILLNESS

In addition to influencing psychopathogenesis directly, the gut microbiome makes an important contribution to drug metabolism, and potentially explains much of the inter-individual

variability in treatment efficacy and side-effects.^{78,79} For example, the gut microbiota has been implicated in the reductive metabolism of psychotropic medications, including benzodiazepine clonazepam,⁸⁰ risperidone⁸¹ and levodopa.⁸² In addition, the gut microbiome is also able to influence the gene expression of hepatic enzymes that aid in the metabolism and detoxification of drugs outside of the gut.^{83,84}

A reciprocal interaction also exists, with drugs used to target psychiatric or neurological disorders having the potential to affect the composition and function of the gut microbiome. For example, the atypical antipsychotic olanzapine has been shown to affect microbiota composition in rats, as well as triggering inflammatory effects and weight gain,^{85,86} with the co-administration of antibiotics shown to attenuate these physiological effects.⁸⁷ The impact of atypical antipsychotics on the gut microbiota may therefore explain to some extent the increased levels of cardiac and metabolic disease in patients receiving these medications.^{88–90}

The clinical implications of these pathways remain poorly understood, but suggest the utility of a precision approach to therapy, as has been advocated in psychiatry^{91,92} and other disease contexts.⁹³

THE ROLE OF THE MICROBIOME IN BRAIN DEVELOPMENT

Prenatal neurodevelopment

Brain development spans the prenatal period to post adolescence and involves the interplay of genetic and environmental factors.⁹⁴ Disruption of these interactions can alter normal developmental trajectories and contribute substantially to neuropsychiatric outcomes in later in life.^{95,96}

Neural development begins early in embryonic life with a number of important stages occurring before birth.⁹⁴ Areas of the brain undergoing these events exhibit greater fragility⁹⁷ and the significant impact of insults that occur during gestation is increasingly recognized.⁹⁸ During this period, maternal immunity and metabolism represents a link between neurodevelopment in the womb and the external environment. Challenges to maternal homeostasis, such as infection, poor nutrition or prenatal stress (PNS), are associated with neurodevelopmental disorders, including anxiety, autism, attention deficit hyperactivity disorder, depression and schizophrenia.^{99–109} Disruption of the maternal microbiome, or 'dysbiosis', appears to act as a link between external stressors and fetal development, either by altering normal developmental cues, or through the presentation of inappropriate developmental stimuli.

The precise nature of relationships between maternal microbiome interactions, altered neurodevelopment and subsequent psychopathologies, remain poorly defined. To a large extent, this is due to the challenge of determining the relative contribution of parallel and overlapping pathways that link multiple interacting systems. Even in animal models, it is extremely difficult to identify the relative contribution of pathways by which a single factor can lead to an array of behavioural disorders. As an illustration, the consumption of a HFD during pregnancy is associated with subsequent behavioural disorders.^{110,111} However, HFD has been shown to influence multiple regulatory pathways in the immune,^{112,113} metabolic¹¹⁴ and neuroendocrine¹¹⁰ systems, through both microbiome dependent and independent mechanisms, as well as resulting in the vertical transmission of the associated dysbiosis.²⁵ Further, the impact of an insult such as HFD consumption depends on the developmental stage at which it occurs, with similar adverse events during early or late periods associated with different outcomes.^{105,115,116}

One important contributor to aberrant neurodevelopment appears to be the disruption of the immuno-regulatory role of the gut microbiome, resulting in a pro-inflammatory maternal

state. Increased levels of circulating cytokines during pregnancy have been shown to negatively affect neural development¹¹⁰ and could act by altering the fetal immune milieu (reviewed in detail elsewhere^{94,117–119}).

Immune-dysregulation could result from factors that ablate the normal microbiota, such as antibiotics, thereby suppressing microbial interactions with toll-like receptors and Treg cells in the gut^{120–122} or the production of immuno-regulatory metabolites, such as short-chain fatty acids (SCFAs).^{122–124} Alternatively, factors that trigger dysbiosis, such as high fat consumption, could act by promoting the production of pro-inflammatory bacterial metabolites.¹²⁵ In addition, the dysbiotic changes in the gut microbiota could influence inflammation and CNS function through changes in activation of vagal and/or spinal nerve pathways.^{22,108,126,127} The contribution of such a microbiota-immune interaction to stress-associated pathologies is supported by the observation that exposure to repeated stress affects the gut microbiota in a manner that correlates with changes in levels of pro-inflammatory cytokines.¹²⁸

The maternal HPA axis is likely to represent another important link between prenatal insults and developmental abnormalities. The HPA axis is affected by factors such as PNS^{129,130} and infection,¹³¹ which are risk factors for a wide range of neurodevelopmental disorders.^{132–136} In animal models of early-life postnatal stress, hyper-responsiveness of the HPA axis is coupled with altered visceral pain sensitivity and impaired intestinal barrier function,^{137,138} while aberrant dietary protein:carbohydrate ratios during gestation have moderate long-term effects on the function of the HPA and sympatho-adrenomedullary axes in offspring.¹³⁹ It is useful to note direct responses to *in utero* stressors such as hypoxia also involve the adrenal system^{140,141} and are essential to fetal survival and neurodevelopment.¹⁴² Whether the maternal microbiome can influence these pathways remains unknown.

The manner in which a hyperactive maternal HPA stress response influences fetal development is unclear; however, an emerging hypothesis involves maternal cortisol crossing the placenta in a quantity sufficient to affect gene expression in fetal brain cells.¹⁴³ This model is supported by *in vitro* analysis of human fetal brain aggregates¹⁴⁴ and the observation that the effects of PNS on offspring can be partially mimicked by giving pregnant animals a synthetic glucocorticoid or adrenocorticotropic hormone.^{130,145} However, the interaction of the HPA axis with the maternal microbiome is likely to be complex. In addition to affecting fetal neurodevelopment directly, stress-induced alterations to the HPA axis trigger maternal gut dysbiosis.¹⁴⁶ These changes in the gut microbiota could further influence HPA axis dysfunction through altered tryptophan metabolism, as well as contributing to other dysbiosis-associated dysregulatory pathways.⁹⁴ In addition, there is evidence that the gut microbiome influences the function of the placenta via the HPA axis, thereby altering fetal exposure to specific compounds in maternal circulation.^{147–150}

The maternal gut microbiota could also affect fetal neurodevelopment by influencing levels of circulating 5-hydroxytryptamine (5-HT). The gut microbiome regulates 5-HT biosynthesis by enterochromaffin (EC) cells in the gut.¹³ In turn, 5-HT regulates fetal neuronal cell division, differentiation and synaptogenesis¹⁵¹ and its depletion results in altered brain development.¹⁵² Furthermore, maternal plasma serotonin is required for proper neuronal morphogenesis during developmental stages that precede the appearance of serotonergic neurons, with embryos depending more on maternal plasma serotonin than their own during *in utero* development.¹⁵³ Maternal gut dysbiosis is also likely to influence blood-brain barrier (BBB) formation, a critical component in CNS development, ensuring an optimal micro-environment for neuronal growth and specification.¹⁵⁴ This is suggested by analysis of the embryos of GF mice, where the BBB has been shown to be substantially compromised.¹⁵⁵

Postnatal neurodevelopment

Neurodevelopment continues outside the womb with the neonatal period characterized by substantial neurological development, including morphological changes, cell differentiation and acquisition of function.^{156,157} Synaptogenesis begins shortly after birth and reaches maximum levels by around 2 years of age, before a process of synaptic refinement and elimination reduces the number of synapses in the postnatal brain to adult levels by mid-adolescence.¹⁵⁸ Remodelling continues well into the third decade of life,¹⁵⁹ providing a lengthy window of vulnerability to external perturbations. This critical period of neurodevelopment parallels the establishment and maturation of the microbiome, a process now known to be essential for the establishment of normal immune function,^{160–164} the neuroendocrine system¹⁶⁵ and metabolic regulation.^{166,167} Disruption of the microbiome in early life therefore has the potential to influence neurodevelopment and long-term mental health outcomes, particularly through its interaction with the immune system and the gut–brain axis.

Gnotobiotic animal models have been important in demonstrating the contribution of the developing microbiome to early-life neurodevelopment and the establishment of appropriate stress responses. For example, GF mice have an exaggerated hypothalamic-pituitary response to mild restraint stress, with elevated plasma adrenocorticotropic hormone and corticosterone and reduced BDNF expression levels in the cortex and hippocampus.⁴⁹ Furthermore, mice that develop in the absence of microbes exhibit increased motor activity and reduced anxiety, associated with differential expression of synaptophysin and PSD-95, proteins that are specifically involved in synaptogenesis pathways.⁴⁸ Microbial colonization is also required for programming and presentation of normal social behaviours, and is important for the regulation of repetitive behaviours,¹⁶⁸ the development of non-spatial memory,²⁹ and the development of pain signalling from the body.¹⁶⁹ It is important to note that the absence of appropriate microbial developmental cues in early-life can result in aberrant mental development that is not corrected by later microbial exposure (Neufeld *et al.*)¹⁷⁰

It is clear from these and other GF animal studies that the absence of a commensal microbiota during early-life substantially affects both neurophysiology and the risk of abnormal behaviour development. However, while a useful tool for highlighting mechanistic pathways, the GF animal poorly reflects the types of microbiome disruption that may occur in humans. As such, other investigations have attempted to recreate real-world early-life insults in the controlled context of animal models. For example, while associations between caesarean-section delivery, altered early life microbial colonization^{171,172} and the incidence of behavioural disorders and abnormal cognitive development in humans^{173–175} have been known for some time, the extent to which a direct causal relationship exists is difficult to discern, given the number of other potentially contributing variables. However, when vaginally delivered mouse pups are compared with those delivered via caesarean section they show an altered gut microbiome and increased anxiety, social deficits and repetitive behaviours reminiscent of autism spectrum disorder-like behaviours in humans.¹⁷⁶

Even in animal models though, the line between pre- and post-delivery periods is blurred by factors such as the vertical transmission of microbiota, the influence of the maternal microbiome of milk composition,¹⁷⁷ and the continuation of stressors in the external environment. An example of this complexity is the impact of PNS on neurodevelopment. PNS has been shown to alter the composition of the gut¹⁷⁸ and maternal vaginal microbiota in mice,^{98,179} thereby altering the pool of microbes that can be passed to the neonate (an analogous situation has been described in humans, where PNS has been shown to affect the composition of the human infant gut microbiota over the first

110 days after birth¹⁸⁰). As above although PNS also alters prenatal development, and therefore the nature of interactions between the neonate and microbes in early life. Determining the relative contribution and timing of contributory pathways to long-term psychopathological outcomes is therefore challenging.

The lasting impact of antibiotic exposure on the microbiome, whether during pregnancy,^{181,182} intrapartum¹⁸³ or in the neonatal period^{24,184} is an example of a further complex factor. There is clear potential for antibiotic dysbiosis to contribute to maternally mediated antenatal neurodevelopment, while antibiotic dysbiosis is also heritable.²⁵ Early-life exposure to antibiotics has been shown to result in long-term immune dysregulation¹⁸⁵ and visceral hypersensitivity.¹⁸⁶ Further, the developmental impact of antibiotic dysbiosis is not limited to the neonatal period, with adolescent rats exhibiting an altered tryptophan metabolic pathway, reduced anxiety and cognitive defects.¹⁸⁷

Diet-induced maternal dysbiosis may also affect early-life neurodevelopment through milk composition. For example, the offspring of mice fed an HFD during lactation show developmental and neurobehavioral changes that suggest possible disruption of physical and sensory-motor maturation, and increased susceptibility to depressive and aggressive-like behaviour.¹⁸⁸ These observations suggests further work in relation to dietary inputs will be important in understanding brain function determinants in humans.

MECHANISMS OF INTERACTION

Activation of inflammatory pathways appears to be a particularly important link between the microbiome and neonatal neurodevelopment. The gut microbiota can affect the immune system directly via activation of the vagus nerve,^{22,126,189–191} in turn triggering bidirectional communication with the CNS.¹⁹² In addition, indirect effects of the gut microbiota on the innate immune system can result in alterations in the circulating levels of pro- and anti-inflammatory cytokines that directly affect brain function.

Bacterial metabolites from the gut have a substantial influence on the regulation of the gut–brain axis and local and systemic immunity. SCFAs, produced by the bacterial fermentation of dietary carbohydrates, have immunomodulatory properties^{121,123,124,193} and can interact with nerve cells by stimulating the sympathetic and autonomic nervous system via G-protein-coupled (GPR) receptor 41 (GPR41)¹⁹⁴ and GPR43.¹⁹⁵ In addition, they can cross the BBB, modulate brain development and behaviour^{196–198} and have been implicated in the development of autism.¹⁹⁹ Further, gut microbiota derived SCFAs have been shown to regulate microglia homeostasis,²⁰⁰ necessary for proper brain development and brain tissue homeostasis.^{201–203} GF mice display global defects in microglia with altered cell proportions and an immature phenotype, leading to impaired innate immune responses in the CNS.²⁰⁰ SCFAs also regulate the release of gut peptides from enteroendocrine cells,²⁰⁴ which in turn affect gut–brain hormonal communication.^{205,206} SCFAs have recently been shown to regulate the synthesis of gut-derived 5-HT from EC cells.¹³ The gut provides ~95% of total body 5-HT,²⁰⁷ most of which exists in plasma. Although this source of 5-HT has intrinsic roles within the gut^{208,209} and peripherally in metabolic control,²¹⁰ EC cell 5-HT can activate afferent nerve endings to signal to the CNS.²¹¹ Furthermore, this source of 5-HT has significant links to psychiatric disorders with the most commonly used antidepressant, fluoxetine, blocking the transport of gut 5-HT into plasma, while elevated plasma serotonin is observed in 25–50% of children with autism^{212–215} and an inverse correlation between high plasma serotonin and low serotonergic neurotransmission has been demonstrated in young male adults with autism spectrum disorder.²¹⁶ In addition to SCFAs, gut bacteria are also capable of producing an array of other neuroactive and

immunomodulatory compounds, including dopamine,²¹⁷ γ -aminobutyric acid,²¹⁸ histamine²¹⁹ and acetylcholine,²²⁰ while the gut microbiome is an important regulator of bile acid pool size and composition,²²¹ and, in turn, BBB integrity and HPA function.²²²

The gut microbiota could also contribute to the regulation of brain function by influencing tryptophan metabolism (reviewed by O'Mahony and colleagues⁹⁵). Tryptophan is an essential, diet-derived, amino acid,²²³ required for serotonin synthesis in the CNS.²²⁴ Once absorbed from the gut, tryptophan can cross the BBB and participate in serotonin synthesis.²²⁴ However, there are many other pathways through which tryptophan can be metabolized,²²⁴ including the largely hepatic kynurenine pathway²²⁵ and the major serotonin synthesis pathway in gut EC cells.^{226–228}

The availability of tryptophan is heavily influenced by the gut microbiota. GF mice have been shown to have increased plasma tryptophan concentrations,^{47,48} which can be normalized following post-weaning colonization.⁴⁷ Resident gut bacteria can utilize tryptophan for growth²²⁹ and in some cases, production of indole,^{230,231} or serotonin (reviewed by O'Mahony and colleagues⁹⁵), while the microbiota might also affect tryptophan availability by influencing host enzymes responsible for its degradation.⁴⁷ By limiting the availability of tryptophan for serotonin production in the CNS (EC-derived serotonin does not cross the BBB), the gut microbiota could influence serotonergic neurotransmission.⁹⁵ In vulnerable populations, reducing the circulating concentrations of tryptophan has been shown to affect mood, and to reinstate depressive symptoms in patients who have successfully responded to selective serotonin reuptake inhibitors.^{232,233} The gut microbiota could also influence the production of both neuroprotective and neurotoxic components of the kynurenine pathway.²²⁴

Other pathways by which the gut microbiota could influence the development and activity of brain tissue include regulation of the release of gut peptides from enteroendocrine cells,²⁰⁴ which in turn affect gut–brain hormonal communication,^{205,206} and, as described above, the regulation of microglia homeostasis.

Two recent, related papers by Wong *et al.* and Zheng *et al.* indicate that the microbiota–gut–brain axis functions in a bidirectional manner in the regulation of depressive-like behaviours. Data in the paper by Wong *et al.*²³⁴ demonstrate that changes in behaviour caused by increased stress levels, knockout of caspase 1 leading to decreased inflammasome function, or pharmacological treatments result in changes in the gut microbiome. The paper by Zheng *et al.* shows three key findings: (i) the absence of gut microbiota in GF mice resulted in decreased immobility time in the forced swimming test relative to conventionally-raised healthy control mice. (ii) From clinical sampling, the gut microbiotic compositions of MDD patients and healthy controls were significantly different from that of MDD patients. (iii) Faecal microbiota transplantation of GF mice with 'depression microbiota' derived from MDD patients resulted in depression-like behaviours compared with colonization with 'healthy microbiota' derived from healthy control individuals. Moreover, the concerned authors showed that mice harbouring 'depression microbiota' primarily exhibited disturbances of microbial genes and host metabolites involved in carbohydrate and amino acid metabolism, indicating that the development of depressive-like behaviours is mediated through the host's metabolism.²³⁵ The combined findings of these two papers suggest that the microbiota–gut–brain axis is fully bidirectional, functioning in a manner through which changes in microbiota affect behaviour, while conversely, changes in behaviour brought about by chronic stress, genetic manipulation, or pharmacological intervention, result in alterations in microbiota composition. Novel approaches to target this bidirectional interface of gut microbiota and depressive-like behaviour may offer novel approaches for the treatment of major depression.

THE ROLE OF THE MICROBIOME IN AGE-RELATED COGNITIVE DECLINE

Despite fluctuating in response to external influences, the gut microbiota is thought to remain relatively stable during adulthood.²³⁶ However, just as the microbiome has a critical role in the development of the nervous system in the neonate, it also appears to have a substantial influence on CNS degeneration in old age. Aging affects the brain on both cellular and functional levels, and is associated with decline in sensory, motor and higher cognitive functions.^{237–239} This period of life is also associated with marked changes in the microbiome.^{240,241} In keeping with dysbiosis arising from a range of insults, age-related changes in gut microbiota composition appear to involve a reduction in microbial diversity, with an increased relative abundance of Proteobacteria and a reduction in bifidobacteria species, and reduced SCFA production.²³⁹

It has been suggested that the processes of age-related dysbiosis and neurological decline are linked through the former mediating chronic low-grade inflammation as a common basis for a broad spectrum of age-related pathologies, or so-called 'inflamm-aging'.²⁴² Inflammation has a substantial role in cognitive decline, not only in the context of normal aging but also in neurological disorders and sporadic Alzheimer's disease.²⁴³ There are a number of ways in which gut dysbiosis could contribute to this process, including direct inflammatory stimulation, the production of pro-inflammatory metabolites, and the loss of immune-regulatory function. In addition, the gut microbiome is essential to the bioavailability of polyphenols, unsaturated fats and antioxidants, all of which may help protect against neuronal and cell aging role under normal circumstances (reviewed by Caracciolo *et al.*²³⁹). Notably, dysbiosis-associated inflammation is also strongly implicated in obesity and diabetes, both of which have been shown to exacerbate normal cognitive decline.^{244–247}

Age-related changes in the brain are most pronounced in the amygdala, hippocampus and frontal cortex,²⁴⁸ whose function is heavily dependent on serotonergic neurotransmission,²⁴⁹ potentially implicating microbiome-influenced changes in tryptophan metabolism. Further, altered serotonin systems could represent a common link with changes in sleep, sexual behaviour and mood in the elderly, as well as disorders such as diabetes, faecal incontinence and cardiovascular diseases.^{94,250}

An association between loss of microbiome function, specifically genes that encode SCFAs, and increased levels of circulating pro-inflammatory cytokines, has been shown in healthy elderly people.²⁵¹ Further, markers of microbiome change are significantly correlated with diet, and with indices of frailty and poor health among long-term institutionalized people,²⁵¹ while feeding cognitively healthy elderly individuals a diet low in meat and meat products is associated with subsequent increases in brain volume and cognitive function.²⁵² Interestingly, in mice, the same HFD predisposes to physiological and anxiety-like effects in adults, while aged mice display deficits in spatial cognition,²⁵³ suggesting the effect of stressors changes during the aging process.

With a growing appreciation of the healthcare implications of an aging global population^{254–256} obtaining a better understanding of how the bidirectional interaction between the microbiome and gut–brain axis that influences age-related changes in brain function, must be a priority.

MODIFICATION OF THE GUT MICROBIOTA TO AFFECT THERAPEUTIC CHANGE

As described above, studies in mice have shown that alteration of the microbial composition of the gut can induce changes in behaviour, raising the possibility of therapeutic manipulation of the microbiome. What approach might be appropriate depends on the specific role of the microbiome in pathogenesis.

In instances where the absence of particular bacterial species is linked to altered brain function, the addition of discrete microbes may be clinically effective. For example, in rats deprived of maternal contact at an early age, treatment with *Bifidobacterium infantis* results in normalization of the immune response, reversal of behavioural deficits, and restoration of basal noradrenaline concentrations in the brainstem,²⁵⁷ while in a mouse model of gastrointestinal inflammation and infection, exposure to *B. longum* normalizes anxiety-like behaviour.^{258,259} The effects of psychosocial stress are also reversed in mice following probiotic treatments.^{260,261} Such effects are not limited to rodent models; in healthy women, a probiotic cocktail alters activity of brain regions that control central processing of emotion and sensation.²⁶² Broadly, such probiotic effects appear to mediate behavioural changes through stimulation of the vagus nerve^{22,191,258} or through modulation of cytokine production.²⁶³

Probiotic therapies have limitations, including a poor ability to establish a stable population within the recipient. Further, in many instances, pathogenesis may be contributed to by broad functions conserved across many different species, such as the ability to produce metabolites that are immunomodulatory, or that directly influence brain activity.^{264,265} Here, it may be the absence of suitable drivers of beneficial behaviour that is limiting, rather than the absence of microbes capable of exhibiting them. In such instances, the broad-scale alteration of the microbiome using selective dietary microbial growth substrates, or prebiotics, may be more appropriate and result in longer lasting change. For example, consumption of fructooligosaccharides or a non-digestible galactooligosaccharide formulation (BGOS) elevates BDNF levels and NMDAR subunit expression in rats,²⁶⁶ BGOS consumption also reduces anxiety in mice injected with lipopolysaccharide to induce sickness behaviour, an effect that appears to be related to the modulation of cortical interleukin-1 β and 5-HT_{2A} receptor expression.²⁶⁷ In humans, daily consumption of BGOS for 3 weeks results in a significantly lower salivary cortisol awakening response compared with placebo and a decreased attentional vigilance to negative versus positive information.²⁶⁸ Pusceddu *et al.*²⁶⁹ showed that long-term supplementation with n-3 polyunsaturated fatty acids corrected dysbiosis seen in maternally separated female rats, and was associated with an attenuation of the corticosterone response to acute stress. Interestingly, while the supporting evidence for the efficacy or such approaches is only now emerging, the consumption of wholegrain and high fibre foods, essentially prebiotics, is already recommended to patients.²⁷⁰

Demonstrations of the transmissibility of behavioural traits between animals by faecal microbiota transfer are also intriguing. Faecal microbiota transfer is employed increasingly widely in the treatment of conditions such as recurrent *Clostridium difficile* infection.²⁷¹ Its ability to influence behaviour suggests that it might also have a role in the treatment of psychopathology (reviewed by Collins *et al.*²⁷²). It is important to note, however, that these observations also raise important questions about current approaches to donor screening for therapeutic faecal microbiota transfer.

FUTURE DIRECTIONS

The advances in our understanding of the role of the microbiome in neurodevelopment and mental health, particularly in the past 5 years, have been remarkable. The implications of this new insight are only beginning to become apparent; however, the potential value of microbiome analyses in revealing mechanisms that underpin altered brain development and mental illness is hugely exciting. There is now a need to close the gap between practice, including the increasing use of pro- and prebiotics, and the supporting science. The importance of achieving this is reflected in the substantial investments made to 'microbiome-

European axis' research by both the US government and the European Union.²⁷³

Achieving a better understanding of the contribution of the microbiome to mental health will require further development of analytical approaches. Studies based on reductive animal models, particularly those involving GF animals, have been important in identifying underlying mechanisms; however, they exclude the complexity of real-world interactions. The rapidly falling costs of 'omics' approaches to microbiome analysis now allow them to be applied to large human cohorts within life-course studies, with data generated assessed in the context of detailed genetic, epigenetic, demographic and clinical assessments. Exploiting these opportunities will result in substantial improvement in our understanding of altered brain function and mental illness in the relative near-term.

In addition to changing analytical strategies, the conceptual framework within which these data are assessed must also continue to develop. A 'three-hit' model of vulnerability and resilience to mental health issues, based on genetic predisposition, the prenatal environment, and later life experiences, has been proposed.²⁷⁴ However, just as the gut-brain axis might be extended to include the microbiome, such developmental pathways must also take into consideration points of interaction with our resident microbiota. Refining these models based on empirical data now represents a key challenge in understanding the processes behind altered brain function and mental illness.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

- 1 Sternbach H, State R. Antibiotics: neuropsychiatric effects and psychotropic interactions. *Harv Rev Psychiatry* 1997; **5**: 214–226.
- 2 Bsted AC, Logan AC, Selhub EM. Intestinal microbiota, probiotics and mental health: from Metchnikoff to modern advances: part I - autointoxication revisited. *Gut Pathog* 2013; **5**: 5.
- 3 Bsted AC, Logan AC, Selhub EM. Intestinal microbiota, probiotics and mental health: from Metchnikoff to modern advances: part II - contemporary contextual research. *Gut Pathog* 2013; **5**: 3.
- 4 Neufeld KA, Foster JA. Effects of gut microbiota on the brain: implications for psychiatry. *J Psychiatry Neurosci* 2009; **34**: 230–231.
- 5 Kennedy PJ, Cryan JF, Dinan TG, Clarke G. Irritable bowel syndrome: a microbiome-gut-brain axis disorder? *World J Gastroenterol* 2014; **20**: 14105–14125.
- 6 Whitehead WE, Palsson O, Jones KR. Systematic review of the comorbidity of irritable bowel syndrome with other disorders: what are the causes and implications? *Gastroenterology* 2002; **122**: 1140–1156.
- 7 Grenham S, Clarke G, Cryan JF, Dinan TG. Brain-gut-microbe communication in health and disease. *Front Physiol* 2011; **2**: 94.
- 8 Mayer EA, Tillisch K. The brain-gut axis in abdominal pain syndromes. *Annu Rev Med* 2011; **62**: 381–396.
- 9 Cryan JF, Dinan TG. Mind-altering microorganisms: the impact of the gut microbiota on brain and behaviour. *Nat Rev Neurosci* 2012; **13**: 701–712.
- 10 Collins SM, Surette M, Bercik P. The interplay between the intestinal microbiota and the brain. *Nat Rev Microbiol* 2012; **10**: 735–742.
- 11 Foster JA, McVey Neufeld KA. Gut-brain axis: how the microbiome influences anxiety and depression. *Trends Neurosci* 2013; **36**: 305–312.
- 12 Aziz Q, Doré J, Emmanuel A, Guarner F, Quigley EM. Gut microbiota and gastrointestinal health: current concepts and future directions. *Neurogastroenterol Motil* 2013; **25**: 4–15.
- 13 Yano JM, Yu K, Donaldson GP, Shastri GG, Ann P, Ma L *et al.* Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis. *Cell* 2015; **161**: 264–276.
- 14 Crane JD, Palanivel R, Mottillo EP, Bujak AL, Wang H, Ford RJ *et al.* Inhibiting peripheral serotonin synthesis reduces obesity and metabolic dysfunction by promoting brown adipose tissue thermogenesis. *Nat Med* 2015; **21**: 166–172.
- 15 Fu J, Bonder MJ, Cenit MC, Tigchelaar EF, Maatman A, Dekens JA *et al.* The gut microbiome contributes to a substantial proportion of the variation in blood lipids. *Circ Res* 2015; **117**: 817–824.

- 16 Valentini M, Piermattei A, Di Sante G, Migliara G, Delogu G, Ria F. Immunomodulation by gut microbiota: role of Toll-like receptor expressed by T cells. *J Immunol Res* 2014; **2014**: 586939.
- 17 Sorini C, Falcone M. Shaping the (auto)immune response in the gut: the role of intestinal immune regulation in the prevention of type 1 diabetes. *Am J Clin Exp Immunol* 2013; **2**: 156–171.
- 18 Dinan TG, Cryan JF. Melancholic microbes: a link between gut microbiota and depression? *Neurogastroenterol Motil* 2013; **25**: 713–719.
- 19 Cryan JF, O'Mahony SM. The microbiome-gut-brain axis: from bowel to behavior. *Neurogastroenterol Motil* 2011; **23**: 187–192.
- 20 Collins SM, Bercik P. The relationship between intestinal microbiota and the central nervous system in normal gastrointestinal function and disease. *Gastroenterology* 2009; **136**: 2003–2014.
- 21 Rhee SH, Pothoulakis C, Mayer EA. Principles and clinical implications of the brain-gut-enteric microbiota axis. *Nat Rev Gastroenterol Hepatol* 2009; **6**: 306–314.
- 22 Bravo JA, Julio-Pieper M, Forsythe P, Kunze W, Dinan TG, Bienenstock J et al. Communication between gastrointestinal bacteria and the nervous system. *Curr Opin Pharmacol* 2012; **12**: 667–672.
- 23 Gohir W, Whelan FJ, Surette MG, Moore C, Schertzer JD, Sloboda DM. Pregnancy-related changes in the maternal gut microbiota are dependent upon the mother's periconceptual diet. *Gut Microbes* 2015; **6**: 310–320.
- 24 Russell SL, Gold MJ, Hartmann M, Willing BP, Thorson L, Wlodarska M et al. Early life antibiotic-driven changes in microbiota enhance susceptibility to allergic asthma. *EMBO Rep* 2012; **13**: 440–447.
- 25 Ma J, Prince AL, Bader D, Hu M, Ganu R, Baquero K et al. High-fat maternal diet during pregnancy persistently alters the offspring microbiome in a primate model. *Nat Commun* 2014; **5**: 3889.
- 26 Thaiss CA, Zeevi D, Levy M, Zilberman-Schapira G, Suez J, Tengeler AC et al. Transkingdom control of microbiota diurnal oscillations promotes metabolic homeostasis. *Cell* 2014; **159**: 514–529.
- 27 Lurie I, Yang YX, Haynes K, Mamtani R, Boursi B. Antibiotic exposure and the risk for depression, anxiety, or psychosis: a nested case-control study. *J Clin Psychiatry* 2015; **76**: 1522–1528.
- 28 Bercik P, Denou E, Collins J, Jackson W, Lu J, Jury J et al. The intestinal microbiota affect central levels of brain-derived neurotrophic factor and behavior in mice. *Gastroenterology* 2011; **141**: 599–609.e1–3.
- 29 Gareau MG, Wine E, Rodrigues DM, Cho JH, Whary MT, Philpott DJ et al. Bacterial infection causes stress-induced memory dysfunction in mice. *Gut* 2011; **60**: 307–317.
- 30 Liu Z, Patil IY, Jiang T, Sancheti H, Walsh JP, Stiles BL et al. High-fat diet induces hepatic insulin resistance and impairment of synaptic plasticity. *PLoS One* 2015; **10**: e0128274.
- 31 Daniel H, Moghaddas Gholami A, Berry D, Desmarchelier C, Hahne H, Loh G et al. High-fat diet alters gut microbiota physiology in mice. *ISME J* 2014; **8**: 295–308.
- 32 Sharma S, Fernandes MF, Fulton S. Adaptations in brain reward circuitry underlie palatable food cravings and anxiety induced by high-fat diet withdrawal. *Int J Obes (Lond)* 2013; **37**: 1183–1191.
- 33 Magnusson KR, Hauck L, Jeffrey BM, Elias V, Humphrey A, Nath R et al. Relationships between diet-related changes in the gut microbiome and cognitive flexibility. *Neuroscience* 2015; **300**: 128–140.
- 34 Kaptan Z, Akgün-Dar K, Kapucu A, Dedeakayoğulları H, Batu Ş, Üzümlü G. Long term consequences on spatial learning-memory of low-calorie diet during adolescence in female rats; hippocampal and prefrontal cortex BDNF level, expression of NeuN and cell proliferation in dentate gyrus. *Brain Res* 2015; **1618**: 194–204.
- 35 Li W, Dowd SE, Scurlock B, Acosta-Martinez V, Lyte M. Memory and learning behavior in mice is temporally associated with diet-induced alterations in gut bacteria. *Physiol Behav* 2009; **96**: 557–567.
- 36 Le Chatelier E, Nielsen T, Qin J, Prifti E, Hildebrand F, Falony G et al. Richness of human gut microbiome correlates with metabolic markers. *Nature* 2013; **500**: 541–546.
- 37 Cottillard A, Kennedy SP, Kong LC, Prifti E, Pons N, Le Chatelier E et al. Dietary intervention impact on gut microbial gene richness. *Nature* 2013; **500**: 585–588.
- 38 Bruce-Keller AJ, Salbaum JM, Luo M, Blanchard E 4th, Taylor CM, Welsh DA et al. Obese-type gut microbiota induce neurobehavioral changes in the absence of obesity. *Biol Psychiatry* 2015; **77**: 607–615.
- 39 Engen PA, Green SJ, Voigt RM, Forsyth CB, Keshavarzian A. The gastrointestinal microbiome: alcohol effects on the composition of intestinal microbiota. *Alcohol Res* 2015; **37**: 223–236.
- 40 Bull-Otterson L, Feng W, Kirpich I, Wang Y, Qin X, Liu Y et al. Metagenomic analyses of alcohol induced pathogenic alterations in the intestinal microbiome and the effect of *Lactobacillus rhamnosus* GG treatment. *PLoS One* 2013; **8**: e53028.
- 41 Biedermann L, Zeitz J, Mwinji J, Sutter-Minder E, Rehman A, Ott SJ et al. Smoking cessation induces profound changes in the composition of the intestinal microbiota in humans. *PLoS One* 2013; **8**: e59260.
- 42 Fond G, Boukouaci W, Chevalier G, Regnault A, Eberl G, Hamdani N et al. The "psychomicrobiotic": targeting microbiota in major psychiatric disorders: a systematic review. *Pathol Biol (Paris)* 2015; **63**: 35–42.
- 43 Castro-Nallar E, Bendall ML, Pérez-Losada M, Sabuncyan S, Severance EG, Dickerson FB et al. Composition, taxonomy and functional diversity of the oropharynx microbiome in individuals with schizophrenia and controls. *PeerJ* 2015; **3**: e1140.
- 44 Nemani K, Hosseini Ghomi R, McCormick B, Fan X. Schizophrenia and the gut-brain axis. *Prog Neuropsychopharmacol Biol Psychiatry* 2015; **56**: 155–160.
- 45 Evrensel A, Ceylan ME. The gut-brain axis: the missing link in depression. *Clin Psychopharmacol Neurosci* 2015; **13**: 239–244.
- 46 O'Brien SM, Scott LV, Dinan TG. Cytokines: abnormalities in major depression and implications for pharmacological treatment. *Hum Psychopharmacol* 2004; **19**: 397–403.
- 47 Clarke G, Grenham S, Scully P, Fitzgerald P, Moloney RD, Shanahan F et al. The microbiome-gut-brain axis during early life regulates the hippocampal serotonergic system in a sex-dependent manner. *Mol Psychiatry* 2013; **18**: 666–673.
- 48 Diaz Heijtz R, Wang S, Anuar F, Qian Y, Björkholm B, Samuelsson A et al. Normal gut microbiota modulates brain development and behavior. *Proc Natl Acad Sci USA* 2011; **108**: 3047–3052.
- 49 Sudo N, Chida Y, Aiba Y, Sonoda J, Oyama N, Yu XN et al. Postnatal microbial colonization programs the hypothalamic-pituitary-adrenal system for stress response in mice. *J Physiol* 2004; **558**: 263–275.
- 50 Neufeld KM, Kang N, Bienenstock J, Foster JA. Reduced anxiety-like behavior and central neurochemical change in germ-free mice. *Neurogastroenterol Motil* 2011; **23**: 255–264, e119.
- 51 Maes M, Kubera M, Leunis JC. The gut-brain barrier in major depression: intestinal mucosal dysfunction with an increased translocation of LPS from gram negative enterobacteria (leaky gut) plays a role in the inflammatory pathophysiology of depression. *Neuro Endocrinol Lett* 2008; **29**: 117–124.
- 52 Ait-Belgnaoui A, Bradesi S, Fioramonti J, Theodorou V, Bueno L. Acute stress-induced hypersensitivity to colonic distension depends upon increase in paracellular permeability: role of myosin light chain kinase. *Pain* 2005; **113**: 141–147.
- 53 Park AJ, Collins J, Blennerhasset PA, Ghia JE, Verdu EF, Bercik P et al. Altered colonic function and microbiota profile in a mouse model of chronic depression. *Neurogastroenterol Motil* 2013; **25**: 733–e575.
- 54 Hubbard TD, Murray IA, Bisson WH, Lahoti TS, Gowda K, Amin SG et al. Adaptation of the human aryl hydrocarbon receptor to sense microbiota-derived indoles. *Sci Rep* 2015; **5**: 12689.
- 55 Marcobal A, Kashyap PC, Nelson TA, Aronov PA, Donia MS, Spormann A et al. A metabolomic view of how the human gut microbiota impacts the host metabolome using humanized and gnotobiotic mice. *ISME J* 2013; **7**: 1933–1943.
- 56 Dinan TG, Borre YE, Cryan JF. Genomics of schizophrenia: time to consider the gut microbiome? *Mol Psychiatry* 2014; **19**: 1252–1257.
- 57 Stefansson H, Ophoff RA, Steinberg S, Andreassen OA, Cichon S, Rujescu D et al. Common variants conferring risk of schizophrenia. *Nature* 2009; **460**: 744–747.
- 58 Sekar A, Bialas AR, de Rivera H, Davis A, Hammond TR, Kamitaki N et al. Schizophrenia risk from complex variation of complement component 4. *Nature* 2016; **530**: 177–183.
- 59 Fan X, Goff DC, Henderson DC. Inflammation and schizophrenia. *Expert Rev Neurother* 2007; **7**: 789–796.
- 60 Song X, Fan X, Song X, Zhang J, Zhang W, Li X et al. Elevated levels of adiponectin and other cytokines in drug naive, first episode schizophrenia patients with normal weight. *Schizophr Res* 2013; **150**: 269–273.
- 61 Miller BJ, Buckley P, Seabolt W, Mellor A, Kirkpatrick B. Meta-analysis of cytokine alterations in schizophrenia: clinical status and antipsychotic effects. *Biol Psychiatry* 2011; **70**: 663–671.
- 62 Meyer U, Feldon J. Prenatal exposure to infection: a primary mechanism for abnormal dopaminergic development in schizophrenia. *Psychopharmacology (Berl)* 2009; **206**: 587–602.
- 63 Müller N, Myint AM, Schwarz MJ. Inflammation in schizophrenia. *Adv Protein Chem Struct Biol* 2012; **88**: 49–68.
- 64 Potvin S, Stip E, Sepelhy AA, Gendron A, Bah R, Kouassi E. Inflammatory cytokine alterations in schizophrenia: a systematic quantitative review. *Biol Psychiatry* 2008; **63**: 801–808.
- 65 Drexhage RC, Hoogenboezem TA, Cohen D, Versnel MA, Nolen WA, van Beveren NJ et al. An activated set point of T-cell and monocyte inflammatory networks in recent-onset schizophrenia patients involves both pro- and anti-inflammatory forces. *Int J Neuropsychopharmacol* 2011; **14**: 746–755.
- 66 Francesconi LP, Ceresér KM, Mascarenhas R, Stertz L, Gama CS, Belmonte-de-Abreu P. Increased annexin-V and decreased TNF- α serum levels in chronic-medicated patients with schizophrenia. *Neurosci Lett* 2011; **502**: 143–146.

- 67 Kunz M, Ceresér KM, Goi PD, Fries GR, Teixeira AL, Fernandes BS et al. Serum levels of IL-6, IL-10 and TNF- α in patients with bipolar disorder and schizophrenia: differences in pro- and anti-inflammatory balance. *Rev Bras Psiquiatr* 2011; **33**: 268–274.
- 68 Pedrini M, Massuda R, Fries GR, de Bittencourt Pasquali MA, Schnorr CE, Moreira JC et al. Similarities in serum oxidative stress markers and inflammatory cytokines in patients with overt schizophrenia at early and late stages of chronicity. *J Psychiatr Res* 2012; **46**: 819–824.
- 69 Severance EG, Gressitt KL, Stallings CR, Origoni AE, Khushalani S, Leweke FM et al. Discordant patterns of bacterial translocation markers and implications for innate immune imbalances in schizophrenia. *Schizophr Res* 2013; **148**: 130–137.
- 70 Fan X, Liu EY, Freudenreich O, Park JH, Liu D, Wang J et al. Higher white blood cell counts are associated with an increased risk for metabolic syndrome and more severe psychopathology in non-diabetic patients with schizophrenia. *Schizophr Res* 2010; **118**: 211–217.
- 71 Hope S, Ueland T, Steen NE, Dieset I, Lorentzen S, Berg AO et al. Interleukin 1 receptor antagonist and soluble tumor necrosis factor receptor 1 are associated with general severity and psychotic symptoms in schizophrenia and bipolar disorder. *Schizophr Res* 2013; **145**: 36–42.
- 72 Na KS, Jung HY, Kim YK. The role of pro-inflammatory cytokines in the neuroinflammation and neurogenesis of schizophrenia. *Prog Neuropsychopharmacol Biol Psychiatry* 2014; **48**: 277–286.
- 73 Ribeiro-Santos A, Lucio Teixeira A, Salgado JV. Evidence for an immune role on cognition in schizophrenia: a systematic review. *Curr Neuropharmacol* 2014; **12**: 273–280.
- 74 Severance EG, Alaedini A, Yang S, Halling M, Gressitt KL, Stallings CR et al. Gastrointestinal inflammation and associated immune activation in schizophrenia. *Schizophr Res* 2012; **138**: 48–53.
- 75 Douglas-Escobar M, Elliott E, Neu J. Effect of intestinal microbial ecology on the developing brain. *JAMA Pediatr* 2013; **167**: 374–379.
- 76 Nieto R, Kukuljan M, Silva H. BDNF and schizophrenia: from neurodevelopment to neuronal plasticity, learning, and memory. *Front Psychiatry* 2013; **4**: 45.
- 77 Coyle JT. NMDA receptor and schizophrenia: a brief history. *Schizophr Bull* 2012; **38**: 920–926.
- 78 Nierenberg AA. Predictors of response to antidepressants: general principles and clinical implications. *Psychiatr Clin North Am* 2003; **26**: 345–352, viii.
- 79 Zhang JP, Malhotra AK. Pharmacogenetics and antipsychotics: therapeutic efficacy and side effects prediction. *Expert Opin Drug Metab Toxicol* 2011; **7**: 9–37.
- 80 Elmer GW, Rummel RP. Role of the intestinal microflora in clonazepam metabolism in the rat. *Xenobiotica* 1984; **14**: 829–840.
- 81 Taylor K, Elliott S. An unusual case of risperidone instability in a fatality presenting an analytical and interpretative challenge. *Drug Test Anal* 2013; **5**: 748–752.
- 82 Fiddian-Green RG. *Helicobacter pylori* eradication and L-dopa absorption in patients with PD and motor fluctuations. *Neurology* 2007; **68**: 1085.
- 83 Meini W, Sczesny S, Brigelius-Flohé R, Blaut M, Glatt H. Impact of gut microbiota on intestinal and hepatic levels of phase 2 xenobiotic-metabolizing enzymes in the rat. *Drug Metab Dispos* 2009; **37**: 1179–1186.
- 84 Claus SP, Ellero SL, Berger B, Krause L, Bruttin A, Molina J et al. Colonization-induced host-gut microbial metabolic interaction. *MBio* 2011; **2**: e00271–10.
- 85 Davey KJ, O'Mahony SM, Schellekens H, O'Sullivan O, Bienenstock J, Cotter PD et al. Gender-dependent consequences of chronic olanzapine in the rat: effects on body weight, inflammatory, metabolic and microbiota parameters. *Psychopharmacology (Berl)* 2012; **221**: 155–169.
- 86 Ley RE, Turnbaugh PJ, Klein S, Gordon JI. Microbial ecology: human gut microbes associated with obesity. *Nature* 2006; **444**: 1022–1023.
- 87 Davey KJ, Cotter PD, O'Sullivan O, Crispie F, Dinan TG, Cryan JF et al. Antipsychotics and the gut microbiome: olanzapine-induced metabolic dysfunction is attenuated by antibiotic administration in the rat. *Transl Psychiatry* 2013; **3**: e309.
- 88 McEvoy JP, Meyer JM, Goff DC, Nasrallah HA, Davis SM, Sullivan L et al. Prevalence of the metabolic syndrome in patients with schizophrenia: baseline results from the Clinical Antipsychotic Trials of Intervention Effectiveness (CATIE) schizophrenia trial and comparison with national estimates from NHANES III. *Schizophr Res* 2005; **80**: 19–32.
- 89 Osby U, Correia N, Brandt L, Ekblom A, Sparén P. Time trends in schizophrenia mortality in Stockholm county, Sweden: cohort study. *BMJ* 2000; **321**: 483–484.
- 90 Emul M, Kalelioglu T. Etiology of cardiovascular disease in patients with schizophrenia: current perspectives. *Neuropsychiatr Dis Treat* 2015; **11**: 2493–2503.
- 91 Wong EH, Fox JC, Ng MY, Lee CM. Toward personalized medicine in the neuropsychiatric field. *Int Rev Neurobiol* 2011; **101**: 329–349.
- 92 Alhajji L, Nemeroff CB. Personalized medicine and mood disorders. *Psychiatr Clin North Am* 2015; **38**: 395–403.
- 93 Rogers GB, Wesselingh S. Precision respiratory medicine and the microbiome. *Lancet Respir Med* 2016; **4**: 73–82.
- 94 O'Mahony SM, Clarke G, Borre YE, Dinan TG, Cryan JF. Serotonin, tryptophan metabolism and the brain-gut-microbiome axis. *Behav Brain Res* 2015; **277**: 32–48.
- 95 Ben-Ari Y. Neuropaediatric and neuroarchaeology: understanding development to correct brain disorders. *Acta Paediatr* 2013; **102**: 331–334.
- 96 Rapoport JL, Giedd JN, Gogtay N. Neurodevelopmental model of schizophrenia: update 2012. *Mol Psychiatry* 2012; **17**: 1228–1238.
- 97 Guerri C. Neuroanatomical and neurophysiological mechanisms involved in central nervous system dysfunctions induced by prenatal alcohol exposure. *Alcohol Clin Exp Res* 1998; **22**: 304–312.
- 98 Jašarević E, Rodgers AB, Bale TL. A novel role for maternal stress and microbial transmission in early life programming and neurodevelopment. *Neurobiol Stress* 2015; **1**: 81–88.
- 99 Brown AS, Susser ES. Prenatal nutritional deficiency and risk of adult schizophrenia. *Schizophr Bull* 2008; **34**: 1054–1063.
- 100 Van Lieshout RJ, Taylor VH, Boyle MH. Pre-pregnancy and pregnancy obesity and neurodevelopmental outcomes in offspring: a systematic review. *Obes Rev* 2011; **12**: e548–e559.
- 101 Wischhof L, Irrsack E, Osorio C, Koch M. Prenatal LPS-exposure—a neurodevelopmental rat model of schizophrenia—differentially affects cognitive functions, myelination and parvalbumin expression in male and female offspring. *Prog Neuropsychopharmacol Biol Psychiatry* 2015; **57**: 17–30.
- 102 Meyer U, Feldon J, Yee BK. A review of the fetal brain cytokine imbalance hypothesis of schizophrenia. *Schizophr Bull* 2009; **35**: 959–972.
- 103 Finegold SM. State of the art; microbiology in health and disease. Intestinal bacterial flora in autism. *Anaerobe* 2011; **17**: 367–368.
- 104 Glynn LM, Wadhwa PD, Dunkel-Schetter C, Chicz-Demet A, Sandman CA. When stress happens matters: effects of earthquake timing on stress responsivity in pregnancy. *Am J Obstet Gynecol* 2001; **184**: 637–642.
- 105 Brown PL, Shepard PD, Elmer GI, Stockman S, McFarland R, Mayo CL et al. Altered spatial learning, cortical plasticity and hippocampal anatomy in a neurodevelopmental model of schizophrenia-related endophenotypes. *Eur J Neurosci* 2012; **36**: 2773–2781.
- 106 Howerton CL, Bale TL. Prenatal programming: at the intersection of maternal stress and immune activation. *Horm Behav* 2012; **62**: 237–242.
- 107 Marques AH, O'Connor TG, Roth C, Susser E, Børke-Monsen AL. The influence of maternal prenatal and early childhood nutrition and maternal prenatal stress on offspring immune system development and neurodevelopmental disorders. *Front Neurosci* 2013; **7**: 120.
- 108 Goehler LE, Park SM, Opitz N, Lyte M, Gaykema RP. *Campylobacter jejuni* infection increases anxiety-like behavior in the holeboard: possible anatomical substrates for viscerosensory modulation of exploratory behavior. *Brain Behav Immun* 2008; **22**: 354–366.
- 109 Bilbo SD, Levkoff LH, Mahoney JH, Watkins LR, Rudy JW, Maier SF. Neonatal infection induces memory impairments following an immune challenge in adulthood. *Behav Neurosci* 2005; **119**: 293–301.
- 110 Sullivan EL, Riper KM, Lockard R, Valleeau JC. Maternal high-fat diet programming of the neuroendocrine system and behavior. *Horm Behav* 2015; **76**: 153–161.
- 111 Sasaki A, de Vega W, Sivanathan S, St-Cyr S, McGowan PO. Maternal high-fat diet alters anxiety behavior and glucocorticoid signaling in adolescent offspring. *Neuroscience* 2014; **272**: 92–101.
- 112 Strandberg L, Verdrengh M, Enge M, Andersson N, Amu S, Onnheim K et al. Mice chronically fed high-fat diet have increased mortality and disturbed immune response in sepsis. *PLoS One* 2009; **4**: e7605.
- 113 Verwaerde C, Delanoye A, Macia L, Tailleux A, Wolowczuk I. Influence of high-fat feeding on both naive and antigen-experienced T-cell immune response in DO10.11 mice. *Scand J Immunol* 2006; **64**: 457–466.
- 114 Choi MS, Kim YJ, Kwon EY, Ryoo JY, Kim SR, Jung UJ. High-fat diet decreases energy expenditure and expression of genes controlling lipid metabolism, mitochondrial function and skeletal system development in the adipose tissue, along with increased expression of extracellular matrix remodelling- and inflammation-related genes. *Br J Nutr* 2015; **113**: 867–877.
- 115 Selevan SG, Kimmel CA, Mendola P. Identifying critical windows of exposure for children's health. *Environ Health Perspect* 2000; **108**: 451–455.
- 116 Khandaker GM, Zimbron J, Lewis G, Jones PB. Prenatal maternal infection, neurodevelopment and adult schizophrenia: a systematic review of population-based studies. *Psychol Med* 2013; **43**: 239–257.
- 117 Hsu P, Nanan R. Foetal immune programming: hormones, cytokines, microbes and regulatory T cells. *J Reprod Immunol* 2014; **104-105**: 2–7.
- 118 Kim DR, Bale TL, Epperson CN. Prenatal programming of mental illness: current understanding of relationship and mechanisms. *Curr Psychiatry Rep* 2015; **17**: 5.
- 119 Glover V. Prenatal stress and its effects on the fetus and the child: possible underlying biological mechanisms. *Adv Neurobiol* 2015; **10**: 269–283.

- 120 Atarashi K, Tanoue T, Oshima K, Suda W, Nagano Y, Nishikawa H et al. Treg induction by a rationally selected mixture of Clostridia strains from the human microbiota. *Nature* 2013; **500**: 232–236.
- 121 Furusawa Y, Obata Y, Fukuda S, Endo TA, Nakato G, Takahashi D et al. Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* 2013; **504**: 446–450.
- 122 Smith PM, Howitt MR, Panikov N, Michaud M, Gallini CA, Bohlooly-Y M et al. The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science* 2013; **341**: 569–573.
- 123 Park BO, Kim SH, Kong GY, Kim DH, Kwon MS, Lee SU et al. Selective novel inverse agonists for human GPR43 augment GLP-1 secretion. *Eur J Pharmacol* 2015; **771**: 1–9.
- 124 Donohoe DR, Garge N, Zhang X, Sun W, O'Connell TM, Bunger MK et al. The microbiome and butyrate regulate energy metabolism and autophagy in the mammalian colon. *Cell Metab* 2011; **13**: 517–526.
- 125 Schaap LA, Koster A, Visser M. Adiposity, muscle mass, and muscle strength in relation to functional decline in older persons. *Epidemiol Rev* 2013; **35**: 51–65.
- 126 Wang H, Yu M, Ochani M, Amella CA, Tanovic M, Susarla S et al. Nicotinic acetylcholine receptor alpha7 subunit is an essential regulator of inflammation. *Nature* 2003; **421**: 384–388.
- 127 de Lartigue G, de La Serre CB, Raybould HE. Vagal afferent neurons in high fat diet-induced obesity; intestinal microflora, gut inflammation and cholecystokinin. *Physiol Behav* 2011; **105**: 100–105.
- 128 Bailey MT, Dowd SE, Galley JD, Hufnagle AR, Allen RG, Lyte M. Exposure to a social stressor alters the structure of the intestinal microbiota: implications for stressor-induced immunomodulation. *Brain Behav Immun* 2011; **25**: 397–407.
- 129 Glover V, O'Connor TG, O'Donnell K. Prenatal stress and the programming of the HPA axis. *Neurosci Biobehav Rev* 2010; **35**: 17–22.
- 130 Weinstock M. The long-term behavioural consequences of prenatal stress. *Neurosci Biobehav Rev* 2008; **32**: 1073–1086.
- 131 McCann SM, Antunes-Rodrigues J, Franci CR, Anselmo-Franci JA, Karanth S, Rettori V. Role of the hypothalamic pituitary adrenal axis in the control of the response to stress and infection. *Braz J Med Biol Res* 2000; **33**: 1121–1131.
- 132 Boersma GJ, Moghadam AA, Cordner ZA, Tamashiro KL. Prenatal stress and stress coping style interact to predict metabolic risk in male rats. *Endocrinology* 2014; **155**: 1302–1312.
- 133 Ronald A, Pennell CE, Whitehouse AJ. Prenatal maternal stress associated with ADHD and autistic traits in early childhood. *Front Psychol* 2011; **1**: 223.
- 134 Class QA, Abel KM, Khshan AS, Rickert ME, Dalman C, Larsson H et al. Offspring psychopathology following preconception, prenatal and postnatal maternal bereavement stress. *Psychol Med* 2014; **44**: 71–84.
- 135 O'Connor TG, Heron J, Golding J, Beveridge M, Glover V. Maternal antenatal anxiety and children's behavioural/emotional problems at 4 years. Report from the Avon Longitudinal Study of Parents and Children. *Br J Psychiatry* 2002; **180**: 502–508.
- 136 Huizink AC, Robles de Medina PG, Mulder EJ, Visser GH, Buitelaar JK. Stress during pregnancy is associated with developmental outcome in infancy. *J Child Psychol Psychiatry* 2003; **44**: 810–818.
- 137 Dumont TM, Rughani AI, Penar PL, Horgan MA, Tranmer BI, Jewell RP. Increased rate of complications on a neurological surgery service after implementation of the Accreditation Council for Graduate Medical Education work-hour restriction. *J Neurosurg* 2012; **116**: 483–486.
- 138 Söderholm JD, Yates DA, Gareau MG, Yang PC, MacQueen G, Perdue MH. Neonatal maternal separation predisposes adult rats to colonic barrier dysfunction in response to mild stress. *Am J Physiol Gastrointest Liver Physiol* 2002; **283**: G1257–G1263.
- 139 Otten W, Kanitz E, Tuchscherer M, Gräbner M, Nürnberg G, Bellmann O et al. Effects of low and high protein:carbohydrate ratios in the diet of pregnant gilts on maternal cortisol concentrations and the adrenocortical and sympathoadrenal reactivity in their offspring. 2013. *J Anim Sci* 2013; **91**: 2680–2692.
- 140 Keating DJ, Rychkov GY, Adams MB, Holgert H, McMillen IC, Roberts ML. Opioid receptor stimulation suppresses the adrenal medulla hypoxic response in sheep by actions on Ca(2+) and K(+) channels. *J Physiol* 2004; **555**: 489–502.
- 141 Keating DJ, Rychkov GY, Roberts ML. Oxygen sensitivity in the sheep adrenal medulla: role of SK channels. *Am J Physiol Cell Physiol* 2001; **281**: C1434–C1441.
- 142 Slotkin TA, Orband-Miller L, Queen KL. Do catecholamines contribute to the effects of neonatal hypoxia on development of brain and heart? Influence of concurrent alpha-adrenergic blockade on ornithine decarboxylase activity. *Int J Dev Neurosci* 1987; **5**: 135–143.
- 143 Glover V. Prenatal stress and child outcomes. In: Antonelli MC (ed). *Advances in Neurobiology Perinatal Programming of Neurodevelopment*. Springer: New York, 2015, pp 269–283.
- 144 Salaria S, Chana G, Caldara F, Feltrin E, Altieri M, Faggioni F et al. Microarray analysis of cultured human brain aggregates following cortisol exposure: implications for cellular functions relevant to mood disorders. *Neurobiol Dis* 2006; **23**: 630–636.
- 145 Crudo A, Suderman M, Moisiadis VG, Petropoulos S, Kostaki A, Hallett M et al. Glucocorticoid programming of the fetal male hippocampal epigenome. *Endocrinology* 2013; **154**: 1168–1180.
- 146 Golubeva AV, Crampton S, Desbonnet L, Edge D, O'Sullivan O, Lomasney KW et al. Prenatal stress-induced alterations in major physiological systems correlate with gut microbiota composition in adulthood. *Psychoneuroendocrinology* 2015; **60**: 58–74.
- 147 Jansson T, Powell TL. Role of the placenta in fetal programming: underlying mechanisms and potential interventional approaches. *Clin Sci (Lond)* 2007; **113**: 1–13.
- 148 Mairesse J, Lesage J, Breton C, Bréant B, Hahn T, Darnaudéry M et al. Maternal stress alters endocrine function of the feto-placental unit in rats. *Am J Physiol Endocrinol Metab* 2007; **292**: E1526–E1533.
- 149 Welberg LA, Thirivikraman KV, Plotsky PM. Chronic maternal stress inhibits the capacity to up-regulate placental 11beta-hydroxysteroid dehydrogenase type 2 activity. *J Endocrinol* 2005; **186**: R7–R12.
- 150 Glover V, Bergman K, Sarkar P, O'Connor TG. Association between maternal and amniotic fluid cortisol is moderated by maternal anxiety. *Psychoneuroendocrinology* 2009; **34**: 430–435.
- 151 Gaspar P, Cases O, Maroteaux L. The developmental role of serotonin: news from mouse molecular genetics. *Nat Rev Neurosci* 2003; **4**: 1002–1012.
- 152 Mazer C, Muneyirci J, Taheny K, Raio N, Borella A, Whitaker-Azmitia P. Serotonin depletion during synaptogenesis leads to decreased synaptic density and learning deficits in the adult rat: a possible model of neurodevelopmental disorders with cognitive deficits. *Brain Res* 1997; **760**: 68–73.
- 153 Côté F, Fligny C, Bayard E, Launay JM, Gershon MD, Mallet J et al. Maternal serotonin is crucial for murine embryonic development. *Proc Natl Acad Sci USA* 2007; **104**: 329–334.
- 154 Engelhardt B. Development of the blood-brain barrier. *Cell Tissue Res* 2003; **314**: 119–129.
- 155 Braniste V, Al-Asmakh M, Kowal C, Anuar F, Abbaspour A, Tóth M et al. The gut microbiota influences blood-brain barrier permeability in mice. *Sci Transl Med* 2014; **6**: 263ra158.
- 156 Anderson GM. Genetics of childhood disorders: XLV. Autism, part 4: serotonin in autism. *J Am Acad Child Adolesc Psychiatry* 2002; **41**: 1513–1516.
- 157 Borre YE, O'Keefe GW, Clarke G, Stanton C, Dinan TG, Cryan JF. Microbiota and neurodevelopmental windows: implications for brain disorders. *Trends Mol Med* 2014; **20**: 509–518.
- 158 Herschkowitz N, Kagan J, Zilles K. Neurobiological bases of behavioral development in the first year. *Neuropediatrics* 1997; **28**: 296–306.
- 159 Petanjek Z, Judaš M, Šimic G, Rasin MR, Uylings HB, Rakic P et al. Extraordinary neoteny of synaptic spines in the human prefrontal cortex. *Proc Natl Acad Sci USA* 2011; **108**: 13281–13286.
- 160 Karmarkar D, Rock KL. Microbiota signalling through MyD88 is necessary for a systemic neutrophilic inflammatory response. *Immunology* 2013; **140**: 483–492.
- 161 Abt MC, Osborne LC, Monticelli LA, Doering TA, Alenghat T, Sonnenberg GF et al. Commensal bacteria calibrate the activation threshold of innate antiviral immunity. *Immunity* 2012; **37**: 158–170.
- 162 Arpaia N, Campbell C, Fan X, Dikiy S, van der Veeke J, deRoos P et al. Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature* 2013; **504**: 451–455.
- 163 Artis D. Epithelial-cell recognition of commensal bacteria and maintenance of immune homeostasis in the gut. *Nat Rev Immunol* 2008; **8**: 411–420.
- 164 Kaplan JL, Shi HN, Walker WA. The role of microbes in developmental immunologic programming. *Pediatr Res* 2011; **69**: 465–472.
- 165 Clarke G, O'Mahony SM, Dinan TG, Cryan JF. Priming for health: gut microbiota acquired in early life regulates physiology, brain and behaviour. *Acta Paediatr* 2014; **103**: 812–819.
- 166 Goulet O. Potential role of the intestinal microbiota in programming health and disease. *Nutr Rev* 2015; **73**: 32–40.
- 167 Arrieta MC, Stiemsma LT, Amenyogbe N, Brown EM, Finlay B. The intestinal microbiome in early life: health and disease. *Front Immunol* 2014; **5**: 427.
- 168 Desbonnet L, Clarke G, Shanahan F, Dinan TG, Cryan JF. Microbiota is essential for social development in the mouse. *Mol Psychiatry* 2014; **19**: 146–148.
- 169 Amaral FA, Sachs D, Costa VV, Fagundes CT, Cisalpino D, Cunha TM et al. Commensal microbiota is fundamental for the development of inflammatory pain. *Proc Natl Acad Sci USA* 2008; **105**: 2193–2197.
- 170 Neufeld KA, Kang N, Bienenstock J, Foster JA. Effects of intestinal microbiota on anxiety-like behavior. *Commun Integr Biol* 2011; **4**: 492–494.
- 171 Dominguez-Bello MG, Costello EK, Contreras M, Magris M, Hidalgo G, Fierer N et al. Delivery mode shapes the acquisition and structure of the initial microbiota across multiple body habitats in newborns. *Proc Natl Acad Sci USA* 2010; **107**: 11971–11975.
- 172 Biasucci G, Rubini M, Riboni S, Morelli L, Bessi E, Retetangos C. Mode of delivery affects the bacterial community in the newborn gut. *Early Hum Dev* 2010; **86**: 13–15.

- 173 Glasson EJ, Bower C, Petterson B, de Klerk N, Chaney G, Hallmayer JF. Perinatal factors and the development of autism: a population study. *Arch Gen Psychiatry* 2004; **61**: 618–627.
- 174 Li HT, Ye R, Achenbach TM, Ren A, Pei L, Zheng X *et al*. Caesarean delivery on maternal request and childhood psychopathology: a retrospective cohort study in China. *BJOG* 2011; **118**: 42–48.
- 175 Al Khalaf SY, O'Neill SM, O'Keefe LM, Henriksen TB, Kenny LC, Cryan JF *et al*. The impact of obstetric mode of delivery on childhood behavior. *Soc Psychiatr Epidemiol* 2015; **50**: 1557–1567.
- 176 Borre YE, Moloney RD, Clarke G, Dinan TG, Cryan JF. The impact of microbiota on brain and behavior: mechanisms & therapeutic potential. *Adv Exp Med Biol* 2014; **817**: 373–403.
- 177 Thum C, Cookson AL, Otter DE, McNabb WC, Hodgkinson AJ, Dyer J *et al*. Can nutritional modulation of maternal intestinal microbiota influence the development of the infant gastrointestinal tract? *J Nutr* 2012; **142**: 1921–1928.
- 178 Tannock GW, Savage DC. Influences of dietary and environmental stress on microbial populations in the murine gastrointestinal tract. *Infect Immun* 1974; **9**: 591–598.
- 179 Jašarević E, Howerton CL, Howard CD, Bale TL. Alterations in the vaginal microbiome by maternal stress are associated with metabolic reprogramming of the offspring gut and brain. *Endocrinology* 2015; **156**: 3265–3276.
- 180 Zijlman MA, Korpela K, Riksen-Walraven JM, de Vos WM, de Weerth C. Maternal prenatal stress is associated with the infant intestinal microbiota. *Psychoneuroendocrinology* 2015; **53**: 233–245.
- 181 Fåk F, Ahrné S, Molin G, Jeppsson B, Weström B. Microbial manipulation of the rat dam changes bacterial colonization and alters properties of the gut in her offspring. *Am J Physiol Gastrointest Liver Physiol* 2008; **294**: G148–G154.
- 182 Tormo-Badia N, Håkansson Å, Vasudevan K, Molin G, Ahrné S, Cilio CM. Antibiotic treatment of pregnant non-obese diabetic mice leads to altered gut microbiota and intestinal immunological changes in the offspring. *Scand J Immunol* 2014; **80**: 250–260.
- 183 Azad MB, Konya T, Persaud RR, Guttman DS, Chari RS, Field CJ *et al*. Impact of maternal intrapartum antibiotics, method of birth and breastfeeding on gut microbiota during the first year of life: a prospective cohort study. *BJOG*; doi: 10.1111/1471-0528.13601; e-pub ahead of print.
- 184 Tanaka S, Kobayashi T, Songjinda P, Tateyama A, Tsubouchi M, Kiyohara C *et al*. Influence of antibiotic exposure in the early postnatal period on the development of intestinal microbiota. *FEMS Immunol Med Microbiol* 2009; **56**: 80–87.
- 185 Deshmukh HS, Liu Y, Menkiti OR, Mei J, Dai N, O'Leary CE *et al*. The microbiota regulates neutrophil homeostasis and host resistance to *Escherichia coli* K1 sepsis in neonatal mice. *Nat Med* 2014; **20**: 524–530.
- 186 O'Mahony SM, Felice VD, Nally K, Savignac HM, Claesson MJ, Scully P *et al*. Disturbance of the gut microbiota in early-life selectively affects visceral pain in adulthood without impacting cognitive or anxiety-related behaviors in male rats. *Neuroscience* 2014; **277**: 885–901.
- 187 Desbonnet L, Clarke G, Traplin A, O'Sullivan O, Crispie F, Moloney RD *et al*. Gut microbiota depletion from early adolescence in mice: implications for brain and behaviour. *Brain Behav Immun* 2015; **48**: 165–173.
- 188 Giriko C, Andreoli CA, Mennitti LV, Hosoume LF, Souto Tdos S, Silva AV *et al*. Delayed physical and neurobehavioral development and increased aggressive and depression-like behaviors in the rat offspring of dams fed a high-fat diet. *Int J Dev Neurosci* 2013; **31**: 731–739.
- 189 Borovikova LV, Ivanova S, Zhang M, Yang H, Botchkina GI, Watkins LR *et al*. Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. *Nature* 2000; **405**: 458–462.
- 190 Lyte M, Li W, Opitz N, Gaykema RP, Goehler LE. Induction of anxiety-like behavior in mice during the initial stages of infection with the agent of murine colonic hyperplasia *Citrobacter rodentium*. *Physiol Behav* 2006; **89**: 350–357.
- 191 Perez-Burgos A, Wang B, Mao YK, Mistry B, McVey Neufeld KA, Bienenstock J *et al*. Psychoactive bacteria *Lactobacillus rhamnosus* (JB-1) elicits rapid frequency facilitation in vagal afferents. *Am J Physiol Gastrointest Liver Physiol* 2013; **304**: G211–G220.
- 192 Dantzer R. Cytokine, sickness behavior, and depression. *Immunol Allergy Clin North Am* 2009; **29**: 247–264.
- 193 Macfarlane S, Macfarlane GT. Regulation of short-chain fatty acid production. *Proc Nutr Soc* 2003; **62**: 67–72.
- 194 Kimura I, Inoue D, Maeda T, Hara T, Ichimura A, Miyauchi S *et al*. Short-chain fatty acids and ketones directly regulate sympathetic nervous system via G protein-coupled receptor 41 (GPR41). *Proc Natl Acad Sci USA* 2011; **108**: 8030–8035.
- 195 Nøhr MK, Pedersen MH, Gille A, Egerod KL, Engelstoft MS, Husted AS *et al*. GPR41/FFAR3 and GPR43/FFAR2 as cosensors for short-chain fatty acids in enteroendocrine cells vs FFAR3 in enteric neurons and FFAR2 in enteric leukocytes. *Endocrinology* 2013; **154**: 3552–3564.
- 196 MacFabe DF, Cain DP, Rodriguez-Capote K, Franklin AE, Hoffman JE, Boon F *et al*. Neurobiological effects of intraventricular propionic acid in rats: possible role of short chain fatty acids on the pathogenesis and characteristics of autism spectrum disorders. *Behav Brain Res* 2007; **176**: 149–169.
- 197 MacFabe DF, Cain NE, Boon F, Ossenkopp KP, Cain DP. Effects of the enteric bacterial metabolic product propionic acid on object-directed behavior, social behavior, cognition, and neuroinflammation in adolescent rats: relevance to autism spectrum disorder. *Behav Brain Res* 2011; **217**: 47–54.
- 198 Macfabe DF. Short-chain fatty acid fermentation products of the gut microbiome: implications in autism spectrum disorders. *Microb Ecol Health Dis* 2012; **23**; doi: 10.3402/mehd.v23i0.19260.
- 199 de Theije CG, Wopereis H, Ramadan M, van Eijndthoven T, Lambert J, Knol J *et al*. Altered gut microbiota and activity in a murine model of autism spectrum disorders. *Brain Behav Immun* 2014; **37**: 197–206.
- 200 Ery D, Hrabě de Angelis AL, Jaitin D, Wieghofer P, Staszewski O, David E *et al*. Host microbiota constantly control maturation and function of microglia in the CNS. *Nat Neurosci* 2015; **18**: 965–977.
- 201 Ransohoff RM, Perry VH. Microglial physiology: unique stimuli, specialized responses. *Annu Rev Immunol* 2009; **27**: 119–145.
- 202 Kettenmann H, Hanisch UK, Noda M, Verkhratsky A. Physiology of microglia. *Physiol Rev* 2011; **91**: 461–553.
- 203 Prinz M, Priller J. Microglia and brain macrophages in the molecular age: from origin to neuropsychiatric disease. *Nat Rev Neurosci* 2014; **15**: 300–312.
- 204 Schéle E, Grahnmemo L, Anesten F, Hallén A, Bäckhed F, Jansson JO. The gut microbiota reduces leptin sensitivity and the expression of the obesity-suppressing neuropeptides proglucagon (Gcg) and brain-derived neurotrophic factor (Bdnf) in the central nervous system. *Endocrinology* 2013; **154**: 3643–3651.
- 205 Cameron J, Doucet E. Getting to the bottom of feeding behaviour: who's on top? *Appl Physiol Nutr Metab* 2007; **32**: 177–189.
- 206 Wren AM, Bloom SR. Gut hormones and appetite control. *Gastroenterology* 2007; **132**: 2116–2130.
- 207 Erspamer V. Pharmacology of indole-alkylamines. *Pharmacol Rev* 1954; **6**: 425–487.
- 208 Keating DJ, Spencer NJ. Release of 5-hydroxytryptamine from the mucosa is not required for the generation or propagation of colonic migrating motor complexes. *Gastroenterology* 2010; **138**: 659–670.e1–2.
- 209 Spencer NJ, Nicholas SJ, Robinson L, Kyloh N, Flack N, Brookes SJ *et al*. Mechanisms underlying distension-evoked peristalsis in guinea pig distal colon: is there a role for enterochromaffin cells? *Am J Physiol Gastrointest Liver Physiol* 2011; **301**: G519–G527.
- 210 Young RL, Lumsden AL, Keating DJ. Gut serotonin is a regulator of obesity and metabolism. *Gastroenterology* 2015; **149**: 253–255.
- 211 Bertrand PP, Kunze WA, Bornstein JC, Furness JB, Smith ML. Analysis of the responses of myenteric neurons in the small intestine to chemical stimulation of the mucosa. *Am J Physiol* 1997; **273**: G422–G435.
- 212 Cook EH Jr, Leventhal BL, Heller W, Metz J, Wainwright M, Freedman DX. Autistic children and their first-degree relatives: relationships between serotonin and norepinephrine levels and intelligence. *J Neuropsychiatry Clin Neurosci* 1990; **2**: 268–274.
- 213 Leboyer M, Philippe A, Bouvard M, Guillaud-Bataille M, Bondoux D, Tabuteau F *et al*. Whole blood serotonin and plasma beta-endorphin in autistic probands and their first-degree relatives. *Biol Psychiatry* 1999; **45**: 158–163.
- 214 Hranilovic D, Bujas-Petkovic Z, Vragovic R, Vuk T, Hock K, Jernej B. Hyperserotonemia in adults with autistic disorder. *J Autism Dev Disord* 2007; **37**: 1934–1940.
- 215 Mulder EJ, Anderson GM, Kema IP, de Bildt A, van Lang ND, den Boer JA *et al*. Platelet serotonin levels in pervasive developmental disorders and mental retardation: diagnostic group differences, within-group distribution, and behavioral correlates. *J Am Acad Child Adolesc Psychiatry* 2004; **43**: 491–499.
- 216 McBride PA, Anderson GM, Hertzig ME, Sweeney JA, Kream J, Cohen DJ *et al*. Serotonergic responsivity in male young adults with autistic disorder. Results of a pilot study. *Arch Gen Psychiatry* 1989; **46**: 213–221.
- 217 Tsavkelova EA, Botvinko IV, Kudrin VS, Oleskin AV. Detection of neurotransmitter amines in microorganisms with the use of high-performance liquid chromatography. *Dokl Biochem* 2000; **372**: 115–117.
- 218 Barrett E, Ross RP, O'Toole PW, Fitzgerald GF, Stanton C. γ -Aminobutyric acid production by culturable bacteria from the human intestine. *J Appl Microbiol* 2012; **113**: 411–417.
- 219 Thomas CM, Hong T, van Pijkeren JP, Hemarajata P, Trinh DV, Hu W *et al*. Histamine derived from probiotic *Lactobacillus reuteri* suppresses TNF via modulation of PKA and ERK signaling. *PLoS One* 2012; **7**: e31951.
- 220 Stephenson M, Rowatt E. The production of acetylcholine by a strain of *Lactobacillus plantarum*. *J Gen Microbiol* 1947; **1**: 279–298.
- 221 Ridlon JM, Kang DJ, Hylemon PB, Bajaj JS. Bile acids and the gut microbiome. *Curr Opin Gastroenterol* 2014; **30**: 332–338.
- 222 McMillin M, Frampton G, Quinn M, Divan A, Grant S, Patel N *et al*. Suppression of the HPA axis during cholestasis can be attributed to hypothalamic bile acid signaling. *Mol Endocrinol* 2015; **29**: 1720–1730.
- 223 Le Floch N, Otten W, Merlot E. Tryptophan metabolism, from nutrition to potential therapeutic applications. *Amino Acids* 2011; **41**: 1195–1205.

- 224 Ruddick JP, Evans AK, Nutt DJ, Lightman SL, Rook GA, Lowry CA. Tryptophan metabolism in the central nervous system: medical implications. *Expert Rev Mol Med* 2006; **8**: 1–27.
- 225 Schwarcz R, Bruno JP, Muchowski PJ, Wu HQ. Kynurenines in the mammalian brain: when physiology meets pathology. *Nat Rev Neurosci* 2012; **13**: 465–477.
- 226 Mawe GM, Hoffman JM. Serotonin signalling in the gut—functions, dysfunctions and therapeutic targets. *Nat Rev Gastroenterol Hepatol* 2013; **10**: 473–486.
- 227 Spiller R. Serotonin and GI clinical disorders. *Neuropharmacology* 2008; **55**: 1072–1080.
- 228 Gershon MD. 5-Hydroxytryptamine (serotonin) in the gastrointestinal tract. *Curr Opin Endocrinol Diabetes Obes* 2013; **20**: 14–21.
- 229 Milligan TW, Doran TI, Straus DC, Mattingly SJ. Growth and amino acid requirements of various strains of group B streptococci. *J Clin Microbiol* 1978; **7**: 28–33.
- 230 Li G, Young KD. Indole production by the tryptophanase TnaA in *Escherichia coli* is determined by the amount of exogenous tryptophan. *Microbiology* 2013; **159**: 402–410.
- 231 Lee Y, Yeom J, Kim J, Jung J, Jeon CO, Park W. Phenotypic and physiological alterations by heterologous acylhomoserine lactone synthase expression in *Pseudomonas putida*. *Microbiology* 2010; **156**: 3762–3772.
- 232 Delgado PL, Charney DS, Price LH, Aghajanian GK, Landis H, Heninger GR. Serotonin function and the mechanism of antidepressant action. Reversal of antidepressant-induced remission by rapid depletion of plasma tryptophan. *Arch Gen Psychiatry* 1990; **47**: 411–418.
- 233 Hood SD, Bell CJ, Nutt DJ. Acute tryptophan depletion. Part I: rationale and methodology. *Aust N Z J Psychiatry* 2005; **39**: 558–564.
- 234 Wong M-L, Inserra A, Lewis MD, Mastronardi CA, Kentish S, Leong L et al. Inflammasome signaling affects anxiety- and depressive-like behavior and gut microbiome composition. *Mol Psychiatry* 2016 (in press).
- 235 Zheng P, Zeng B, Zhou C, Liu M, Xu X, Zeng L et al. Microbiome remodeling induces depression-like behaviors in a pathway that is mediated through the host's metabolism. *Mol Psychiatry* 2016; doi: 10.1038/mp.2016.44 (in press).
- 236 Dominguez-Bello MG, Blaser MJ, Ley RE, Knight R. Development of the human gastrointestinal microbiota and insights from high-throughput sequencing. *Gastroenterology* 2011; **140**: 1713–1719.
- 237 Salthouse TA. Decomposing age correlations on neuropsychological and cognitive variables. *J Int Neuropsychol Soc* 2009; **15**: 650–661.
- 238 Schaffer S, Asseburg H, Kuntz S, Muller WE, Eckert GP. Effects of polyphenols on brain ageing and Alzheimer's disease: focus on mitochondria. *Mol Neurobiol* 2012; **46**: 161–178.
- 239 Caracciolo B, Xu W, Collins S, Fratiglioni L. Cognitive decline, dietary factors and gut-brain interactions. *Mech Ageing Dev* 2014; **136–137**: 59–69.
- 240 Biagi E, Nylund L, Candela M, Ostan R, Bucci L, Pini E et al. Through ageing, and beyond: gut microbiota and inflammatory status in seniors and centenarians. *PLoS One* 2010; **5**: e10667.
- 241 Claesson MJ, Jeffery IB, Conde S, Power SE, O'Connor EM, Cusack S et al. Composition, variability, and temporal stability of the intestinal microbiota of the elderly. *Proc Natl Acad Sci USA* 2011; **108**: 4586–4591.
- 242 Franceschi C, Bonafè M, Valensin S, Olivieri F, De Luca M, Ottaviani E et al. Inflamm-aging. An evolutionary perspective on immunosenescence. *Ann N Y Acad Sci* 2000; **908**: 244–254.
- 243 Griffin WS. Neuroinflammatory cytokine signaling and Alzheimer's disease. *N Engl J Med* 2013; **368**: 770–771.
- 244 Hassing LB, Dahl AK, Thorvaldsson V, Berg S, Gatz M, Pedersen NL et al. Overweight in midlife and risk of dementia: a 40-year follow-up study. *Int J Obes (Lond)* 2009; **33**: 893–898.
- 245 Kivipelto M, Ngandu T, Fratiglioni L, Viitanen M, Kåreholt I, Winblad B et al. Obesity and vascular risk factors at midlife and the risk of dementia and Alzheimer disease. *Arch Neurol* 2005; **62**: 1556–1560.
- 246 Cheng CM, Chiu MJ, Wang JH, Liu HC, Shyu YI, Huang GH et al. Cognitive stimulation during hospitalization improves global cognition of older Taiwanese undergoing elective total knee and hip replacement surgery. *J Adv Nurs* 2012; **68**: 1322–1329.
- 247 Exalto LG, Whitmer RA, Kappelle LJ, Biessels GJ. An update on type 2 diabetes, vascular dementia and Alzheimer's disease. *Exp Gerontol* 2012; **47**: 858–864.
- 248 Lupien SJ, McEwen BS, Gunnar MR, Heim C. Effects of stress throughout the lifespan on the brain, behaviour and cognition. *Nat Rev Neurosci* 2009; **10**: 434–445.
- 249 Berger M, Gray JA, Roth BL. The expanded biology of serotonin. *Annu Rev Med* 2009; **60**: 355–366.
- 250 Fidalgo AR. Experimental insights into age-exacerbated cognitive dysfunction after peripheral surgery. *Ageing Cell* 2013; **12**: 523–524.
- 251 Claesson MJ, Jeffery IB, Conde S, Power SE, O'Connor EM, Cusack S et al. Gut microbiota composition correlates with diet and health in the elderly. *Nature* 2012; **488**: 178–184.
- 252 Titova OE, Ax E, Brooks SJ, Sjögren P, Cederholm T, Kilander L et al. Mediterranean diet habits in older individuals: associations with cognitive functioning and brain volumes. *Exp Gerontol* 2013; **48**: 1443–1448.
- 253 Kesby JP, Kim JJ, Scadeng M, Woods G, Kado DM, Olefsky JM et al. Spatial cognition in adult and aged mice exposed to high-fat diet. *PLoS One* 2015; **10**: e0140034.
- 254 Jin K, Simpkins JW, Ji X, Leis M, Stambler I. The critical need to promote research of aging and aging-related diseases to improve health and longevity of the elderly population. *Ageing Dis* 2014; **6**: 1–5.
- 255 Suzman R, Beard JR, Boerma T, Chatterji S. Health in an ageing world—what do we know? *Lancet* 2015; **385**: 484–486.
- 256 Chatterji S, Byles J, Cutler D, Seeman T, Verdes E. Health, functioning, and disability in older adults—present status and future implications. *Lancet* 2015; **385**: 563–575.
- 257 Desbonnet L, Garrett L, Clarke G, Kiely B, Cryan JF, Dinan TG. Effects of the probiotic *Bifidobacterium infantis* in the maternal separation model of depression. *Neuroscience* 2010; **170**: 1179–1188.
- 258 Bercik P, Verdu EF, Foster JA, Macri J, Potter M, Huang X et al. Chronic gastrointestinal inflammation induces anxiety-like behavior and alters central nervous system biochemistry in mice. *Gastroenterology* 2010; **139**: 2102–2112.e1.
- 259 Bercik P, Park AJ, Sinclair D, Khoshdel A, Lu J, Huang X et al. The anxiolytic effect of *Bifidobacterium longum* NCC3001 involves vagal pathways for gut-brain communication. *Neurogastroenterol Motil* 2011; **23**: 1132–1139.
- 260 Zareie M, Johnson-Henry K, Jury J, Yang PC, Ngan BY, McKay DM et al. Probiotics prevent bacterial translocation and improve intestinal barrier function in rats following chronic psychological stress. *Gut* 2006; **55**: 1553–1560.
- 261 Ait-Belgnaoui A, Durand H, Cartier C, Chaumaz G, Eutamene H, Ferrier L et al. Prevention of gut leakiness by a probiotic treatment leads to attenuated HPA response to an acute psychological stress in rats. *Psychoneuroendocrinology* 2012; **37**: 1885–1895.
- 262 Tillisch K, Labus J, Kilpatrick L, Jiang Z, Stains J, Ebrat B et al. Consumption of fermented milk product with probiotic modulates brain activity. *Gastroenterology* 2013; **144**: 1394–401.e1–4.
- 263 Desbonnet L, Garrett L, Clarke G, Bienenstock J, Dinan TG. The probiotic *Bifidobacteria infantis*: an assessment of potential antidepressant properties in the rat. *J Psychiatr Res* 2008; **43**: 164–174.
- 264 Wikoff WR, Anfora AT, Liu J, Schultz PG, Lesley SA, Peters EC et al. Metabolomics analysis reveals large effects of gut microflora on mammalian blood metabolites. *Proc Natl Acad Sci USA* 2009; **106**: 3698–3703.
- 265 Hanstock TL, Mallet PE, Clayton EH. Increased plasma d-lactic acid associated with impaired memory in rats. *Physiol Behav* 2010; **101**: 653–659.
- 266 Savignac HM, Corona G, Mills H, Chen L, Spencer JP, Tzortzis G et al. Prebiotic feeding elevates central brain derived neurotrophic factor, N-methyl-D-aspartate receptor subunits and D-serine. *Neurochem Int* 2013; **63**: 756–764.
- 267 Savignac HM, Couch Y, Stratford M, Bannerman DM, Tzortzis G, Anthony DC et al. Prebiotic administration normalizes lipopolysaccharide (LPS)-induced anxiety and cortical 5-HT_{2A} receptor and IL-1 β levels in male mice. *Brain Behav Immun* 2015; **52**: 120–131.
- 268 Schmidt K, Cowen PJ, Harmer CJ, Tzortzis G, Errington S, Burnet PW. Prebiotic intake reduces the waking cortisol response and alters emotional bias in healthy volunteers. *Psychopharmacology (Berl)* 2015; **232**: 1793–1801.
- 269 Pusceddu MM, El Aidy S, Crispie F, O'Sullivan O, Cotter P, Stanton C et al. N-3 polyunsaturated fatty acids (PUFAs) reverse the impact of early-life stress on the gut microbiota. *PLoS One* 2015; **10**: e0139721.
- 270 Mental Health Foundation, 2007. Available at: <http://www.mentalhealth.org.uk/help-information/mental-health-a-z/D/diet/> (accessed on January 2016).
- 271 Rogers GB, Bruce KD. Challenges and opportunities for faecal microbiota transplantation therapy. *Epidemiol Infect* 2013; **141**: 2235–2242.
- 272 Collins SM, Kassam Z, Bercik P. The adoptive transfer of behavioral phenotype via the intestinal microbiota: experimental evidence and clinical implications. *Curr Opin Microbiol* 2013; **16**: 240–245.
- 273 Smith PA. The tantalizing links between gut microbes and the brain. *Nature* 2015; **526**: 312–314.
- 274 Daskalakis NP, Bagot RC, Parker KJ, Vinkers CH, de Kloet ER. The three-hit concept of vulnerability and resilience: toward understanding adaptation to early-life adversity outcome. *Psychoneuroendocrinology* 2013; **38**: 1858–1873.



This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by-nc-nd/4.0/>