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Signature sequences in diverse proteins provide evidence for the late divergence of the Order *Aquificales*

Summary. The Aquificales species are presently believed to be the earliest branching lineage within Bacteria. However, the branching order of this group in different phylogenetic trees is highly variable and not resolved. In the present work, the phylogenetic placement of Aquificales was examined by means of a cladistic approach based on the shared presence or absence of definite signature sequences (consisting of conserved inserts or deletions) in many highly conserved and important proteins, e.g. RNA polymerase β (RpoB), RNA polymerase β' (RpoC), alanyl-tRNA synthetase (AlaRS), CTP synthase, inorganic pyrophosphatase (PPase), Hsp70 and Hsp60. For this purpose, fragments of the above genes that contained the signature regions were cloned from different Aquificales species (Calderobacterium hydrogenophilum, Hydrogenobacter marinus, and Thermocrinis ruber) and the sequence data were compared with those available from all other species. The presence in Aquificales species of distinctive inserts in Hsp70 and Hsp60 that are not found in any Firmicutes, Actinobacteria, or Thermotoga-Clostridium species excluded them from these groups of Bacteria. The shared presence of prominent indels in the RpoB (>100 amino acids), RpoC (>100 amino acids) and AlaRS (4 amino acids) proteins, which are only found in the various Aquificales species, the Chlamydiae, the CFBG (Cytophaga-Flavobacteria-Bacteroides-green sulfur bacteria) group, and Proteobacteria, strongly suggests their placement within these groups of Bacteria. A specific relationship between Proteobacteria and Aquificales is suggested by the presence in inorganic pyrophosphatase of a 2-amino-acid insert that is uniquely found in these phyla. However, the Aquificales species lacked a number of other protein signatures (e.g. indels in CTP synthase and Hsp70) that are characteristic of Proteobacteria, indicating that they constitute a distinct phylum related to Proteobacteria. These results provide strong and consistent evidence that the Aquificales diverged after the branching of Firmicutes, Actinobacteria, Thermotoga, Deinococcus-Thermus, green nonsulfur bacteria, Cyanobacteria, Spirochetes, Chlamydiae, and CFBG group, but before the emergence of the Proteobacteria. [Int Microbiol 2004; 7(1):41-52]

Key words: Aquifex · bacterial phylogeny · branching order

Introduction

The phylogenetic trees based on 16S rRNA provide the presently accepted framework for understanding the evolutionary relationships among *Bacteria* [29,33]. Based on these

trees, a number of main groups or phyla within the *Bacteria* have been recognized. These include *Thermotoga*, green nonsulfur bacteria (GNS), *Deinococcus-Thermus*, Cyanobacteria, low G + C gram-positive (Firmicutes), high G + C gram-positive (Actinobacteria), Chlamydiae, CFBG (*Cytophaga-*Flavobacteria-*Bacteroides*-Green sulfur bacteria), *Planctomy*- *ces*, and relatives, and the Proteobacteria [29,41]. In addition, a number of other phyla consisting of only a limited number of species are also recognized [29]. Although the branching orders of different phyla in rRNA or other phylogenetic trees have not been resolved, the *Aquificales* species (represented by the genus *Aquifex*) are thought to be the earliest branching lineage within *Bacteria* [1,4,6,8,14,29,30,33,35,36,40]. However, the deep branching of *Aquifex* is not supported by many protein phylogenies, in which it exhibits a closer relationship to the δ_{ϵ} -Proteobacteria and the Chlamydiae groups [5,7,10,18–20,27,31,36,39].

Despite the lack of a consistent picture concerning its phylogenetic placement, the deep branching of Aquifex has become a central aspect of the current view of bacterial phylogeny [8,11,29,33,36]. In this context, it is important to further investigate the branching position of this phylum relative to other groups of Bacteria using different approaches. In the present work, we used a cladistic approach based on shared conserved indels or signature sequences in various proteins to deduce the branching order of bacterial groups [19,23]. This approach has provided evidence that the major groups within Bacteria have branched off in the following order: low G + C gram-positives \rightarrow high G + C gram-positives \rightarrow Deinococcus/Thermus-GNS-Cyanobacteria-Spirochetes \rightarrow Chlamydiae, CFBG, Aquifex $\rightarrow \delta_{,\varepsilon}$ -Proteobacteria $\rightarrow \alpha$ -Proteobacteria $\rightarrow \beta$ -Proteobacteria $\rightarrow \gamma$ -Proteobacteria [20,21,23]. By means of this approach, the genus Aquifex was found to branch at a position similar to that of the Chlamydiae and CFBG groups [20,21].

The present report describes work on a number of new as well as previously described protein signatures (i.e. Hsp70, Hsp60, CTP synthase, PPase, AlaRS, RpoB, and RpoC) that are helpful in understanding the phylogenetic placement of *Aquificales*. The Order *Aquificales* comprises four genera: *Aquifex, Calderobacterium, Hydrogenobacter* and *Thermocrinis* [36]. Of these, sequence information is mainly available for *Aquifex* species. We have cloned and sequenced gene fragments of the above-mentioned proteins, containing the signature regions from species belonging to various *Aquificales* genera. Results of these studies provide consistent evidence that this group should be placed between the $\delta_{,\varepsilon}$ -Proteobacteria and the Chlamydiae and CFBG groups and it constitutes a late-branching phylum within *Bacteria*.

Materials and methods

DNA. Purified *C. hydrogenophilum* (type strain; Z-829) DNA was generously provided by Dr. Karel Mikulik (Academy of Sciences, Czech Republic) [32]. The DNA for *H. marinus* (DSM 12046T) was kindly provided by Dr. Micheal Thomm (Institut für Allgemeine Mikrobiologie, Kiel, Germany) [38], and the DNA from *T. ruber* (DSM 12173) was a generous gift of Dr. K.O. Stetter (University of Regensburg, Berlin, Germany) [24]. The complete genome of *Aquifex aeolicus* have been sequenced [11] and sequence information for several *Aquifex pyrophilus* genes is available in the NCBI database.

PCR amplification and sequencing. Degenerate oligonucleotide primers, in opposite orientations, were designed for highly conserved regions that flanked the identified signatures in sequence alignments. The sequences of various PCR primers used in these studies are detailed in Table 1. Because these primers are based on highly conserved regions, they may also prove effective in amplifying these genes from other species. Ten-µl PCR reactions (approximately 0.2 mg DNA per reaction) were optimized for Mg²⁺ concentration (1.5-4 mM) for each set of primers. PCR was carried out using a Techne Progene thermocycler, over 30 cycles (15 s at 94°C, 15 s at 55°C, 1 min at 72°C) with an initial 1-min hot start at 94°C, and a final extension step (15 s at 94°C, 15 s at 55°C, 7 min at 72°C). The DNA fragments of the expected size were purified from 0.8 % (w/v) agarose gels and subcloned into the plasmid pDRIVE using a UA cloning kit (Qiagen). Due to DNA limitation, some gene fragments from T. ruber were not amplified. After transforming E. coli JM109 cells with the plasmids, inserts from a number of positive clones were sequenced. The sequence data for various Aquificales species have been deposited in the GenBank and the accession numbers for these sequences are included in the alignment figures.

Results

Determining the branching Order of *Aquificales* **based on signatures sequences.** Signature sequences provide a powerful means to deduce the relative branching order and interrelationships among different groups. By making use of conserved and defined indels that are commonly shared by different species, it is possible to group different species or taxa into distinct clades, which show specific relationships to each other [3,19,21,26,37]. The application of this approach for determining the phylogenetic placement of *Aquificales* is described below.

The Hsp70 (DnaK) family of proteins contain a 21-23amino-acid (aa) insert in the N-terminal quadrant that distinguishes various gram-negative (or diderm) bacteria from gram-positive (or monoderm) bacteria (Fig. 1) [19]. The Hsp70 homologs are found in all Bacteria and the identified insert is present in all diderm (i.e., primarily gram-negative) bacteria, but not in Thermotoga or any Firmicutes and Actinobacteria. The insert in Hsp70 is not found in any archaeal homolog where this protein is found, which suggests that this indel constitutes an insert, and that the species lacking it are ancestral (Fig. 1) [19]. Both Aquifex aeolicus and Aquifex pyrophilus, whose sequences are available [11,17], contained this insert. The presence of this insert in C. hydrogenophilum, H. marinus, and T. ruber was examined by PCR amplifying the *dnaK* gene fragments covering the indel region. Sequences of the resulting fragments showed that this insert is present in all of these species (Fig. 1), indicat-

Gene	Primer	Primer sequence*	Fragment size
Hsp70	Forward	5'-GGNATHGAYYTNGGNACNAC-3'	1.1 kb
	Reverse	5'- GCNACNGCYTCRTCNGGRTT- 3'	
Hsp60	Forward	5'-AAGCTTTCNCCRAANCCNGGNGCY TTNACNGC-3'	0.5 kb
	Reverse	5'-GGYGAYGGYACYACHACWGC-3'	
Alanine tRNA synthetase	Forward	5'-TTYACNAAYGCNGGNATG-3'	170 bp
	Reverse	5'-CATYTCRAARAANGTRTG-3'	
CTP synthase (Proteobacterial insert)	Forward	5'-GGNATTHTGYYTNGGNATGCA-3'	420 bp
	Reverse	5'-AAYTCNGGRTGRAAYTG-3'	
Inorganic pyrophosphatase	Forward	5'-AARTAYGARMTIGAYAARGA-3'	333 bp
	Reverse	5'-TYYTTRTAIKKYTCRAARAA-3'	
RNA polymerase subunit	Forward		
	Calderobacterium	5'-TTYYTWGAGCACRAYGAYGCDAA YMGNGCNYTNATGGG-3'	830 bp
	Hydrogenothermus	5'-GGNTAYAAYTWYGARGAYGC-3'	1.2 kb
	Reverse	5'-ACCYTTRTTWCCRTGHCKTCCHGC CATYTTRTCDCC- 3'	
RNA polymerase β subunit	Forward	5'-ATHGGNGARCCNGGNACNCA-3'	642 bp
	Reverse	5'-GGNARNCCNCCNGTDATRTC-3'	
CTP synthase (<i>Aquifex</i> -Proteob. insert)	Forward	5'-GGNCAYTAYGARMGNTT-3'	357 bp
	Reverse	5'-TGYTGNGTNGGYTTNGTYTT-3'	-

Table 1. PCR primers for am	plifying different	gene sequences
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* Where N=A,T,C or G; H=A,C or T; Y=C or T; R=A or G; W=A or T; M=A or C; K=G or T; D=G,A or T; I=inositol.

ing that this group diverged after the branching of gram-positive bacteria (Actinobacteria and Firmicutes) and *Thermotoga* (Fig. 1).

Hsp60, which is present in all bacteria except a few mycoplasmas, contains a conserved 1-aa insert that is common to various Proteobacteria, Chlamydiae, CFBG group, Spirochetes, and Cyanobacteria, but absent from various Firmicutes, Actinobacteria, Thermotoga, Deinococcus /Thermus group, and GNS bacteria (Fig. 2) [19,20]. The sequence information for Hsp60 is available from >400 species and no exception to this pattern has been observed. A 0.5-kb fragment of groEL covering this region was amplified from the three Aquificales species, and all of them were found to contain this insert in the appropriate position (Fig. 2). This insert is also present in the published sequence of A. aeolicus [11]. The shared presence of this insert in different Aquificales as well as various Cyanobacteria, Spirochetes, Chlamydiae, CFBG and Proteobacteria provides evidence that it should be placed within these groups. The bacterial groups lacking this indel (Firmicutes, Actinobacteria, Thermotoga, Deinococcus/ Thermus, and GNS bacteria) are indicated to have branched off prior to the insertion of this indel.

A prominent signature was identified in the β '-subunit of RNA polymerase (RpoC) that consists of a large insert of *ca*. 200 aa in various Proteobacteria, Chlamydiae, CFBG group, and Spirochetes. In this position, the cyanobacteria contain a much larger insert (~600 aa), which could be of independent origin or may have changed subsequently. However, no insert

is present in this position in various Archaea, Firmicutes, Actinobacteria, *Thermotoga, Deinococcus/Thermus* group, and GNS bacteria, indicating that these groups diverged prior to the introduction of this indel (Fig. 3). A 642-bp fragment of *rpoC* covering this region was amplified from *H. marinus* and *C. hydrogenophilum*, and both these species contained a 188-aa insert in this region. Similar inserts of 188 aa are also present in the published sequences of *A. aeolicus* and *A. pyrophilus* (Fig. 3) [11,27]. The shared presence of this large (~200 aa) insert in the Spirochetes, Chlamydiae, CFBG group, and Proteobacteria again suggests that *Aquificales* should be placed within these groups of *Bacteria*.

Alanyl-tRNA synthetase contains a highly conserved 4-aa insert that is common to various Proteobacteria, Chlamydiae, CFBG groups, and *A. aeolicus* and *A. pyrophilus*, but it is not found in any other group of prokaryotes, including Archaea (Fig. 4). A 170-bp fragment of AlaRS covering this region was amplified and sequenced from *C. hydrogenophilum*, *H. marinus*, and *T. ruber*. The results of these studies (Fig. 4) show that this insert is a common characteristic of all *Aquificales* species, thereby strongly supporting their placement within the Proteobacteria, Chlamydiae, and CFBG group and indicating that these phyla diverged after branching of groups lacking this indel (Fig. 4).

Another prominent signature showing a relationship similar to that of AlaRS was identified in the β -subunit of RNA polymerase (RpoB). This protein contains a large insert of ~120 aa that is present in Chlamydiae, the CFBG group,

			61					123
r.	E. coli	SP/P04475	NPONTLFAIKRLIG	RRF	OD	EEVQRDVSIMPFKIIAAD	NGDAWVEVKGOK	MAPPOISAEVLLKM
	X. fastidiosa	AAF85139	KFY-V		G-	AK-LDLV-YTOH-	ATADAK	LOEKE
	Pse. aeruginosa	AAG08147	Y-V		EE	NVK-IOMV-YS-VK		K
	Pas. multocida	15602601	K			E-V	E-	
	Vibrio cholerae	2623605			E-	IKYVK	A	AV
	Ral. solanacearum	16429657	RY-V		EE	KK-IGLYT-SK	RDK-	R
	Nei. meningitidis	H81185	-AKIY-A		E-	KIESEK-N	KAO-KE	LSR
	Bru. melitensis	17988285			D-	PM-TK-KDLV-YVKG-	H-K-	YS-SMI-O
Proteobacteria	Mesorhizobium loti	13473981	EIV		D-	PVTEK-KKLV-YVKG-	AG-K-	0S-SMI-0
Toteobacteria			RIY-V		I-	PM-RK-QG-V-YN-VK	ADNN-	YS-SFI-Q-
	Ri. conorii	15892156	TVV		T-	AEK-KKLV-YN-VDGG	R-E-	FS-A-VVI-0
	Rh. capsulatus	1373328		Y				
	A. tumefaciens	SP/P20442	TV			PT-EK-KALVE-VKG-	KAQDKN	YS-SMI-Q
	Ca. crescentus	GB/M95799	T	-TA		PV-EK-KGMV-YRSSR-R		YS-QEVFI-Q-
	Myx. xanthus	1805284	EVA		DS	P-GKKAIGVSVASSP	IR-KG	YSEVIM
	Geo. metallireducens	ZP_00080430	E	1010	DT	RK-ISVK	AR-KM	YSA-EMQ
		ZP_00130430	ER-VVM-		DA	PG-WKEHS-YR-V-GA	Q-RP	YSA-EMI-G-I
	Camp. jejuni	CAB73024	EK-IYSIM-	LMI		DAAKEAKNRL-YH -TER	ACAI-IA-KI	YT-QEKM-1
1	Hel. pylori	2072520	EK-IYSIM-		NE	DKAKEAEKRL-Y-VDRN	GA-CAI-IS-KI	YT-QEKI-M-
1	Chl. trachomatis	SP/P17821	EKASTF	-K-		SESEIKTV-Y-VAPNS	KVFD-EQKL	YT-EE-G-QI-M-
	Chl. muridarum	AAF39496	EKASTF	-K-		SESEIKTV-Y-VAPNS	KVFENKL	YT-EE-G-QI-M
Oblamudias	Chlam. pneumoniae	AAF38114	EKGSF	-KY		SASEIQTV-YTVTSGS	KVFD-KQ	YT-EE-G-QI-N-
Chlamydiae,	Cyt. aquatilis	AF013110	TK-IASFM-	HT-		A-TTNESKRVSY-VVKGV	QQYST-DID-RL	YTAQELMT-Q
CFBG Group	Fl. ferrugineum	AF013111	ITSVFM-	-G-		NTEEI-HWSY-VAOG-	-NTVRIDID-RL	YT-QEMQ
	Fib. succinogenes	AY017382	EK-IYSFM-	-TA		G-CSAAEKNY-LVGTG	SDPVR-OIDDKO	FEAOT-
	Por. gingivalis	Q9ZAD3	TK-IYSFM-	ETY		DO-S-E-ERVVVRG-	-NTPR-DID-RL	YT-QEMI-Q-I
1	Cb. tepidum	AF130447	KI-SFM-	-KY		DPNEKKLASYDVVN-E		YS-QEMI-Q
	C. hydrogenophilum	AY188750	D-EIYESF	-KS		KEEAKRVSY-VVPDE		LVR-EEVG-HI-K-1
	Tc. ruber	AY254212	D-EIYESF	1.		-D-KEEAIRVSYMVV-DK		
	Hydro. marinus	AY188440	DIYESF	-K-		DKEEIKYV-Y-VV-D-		IVR-EEVG-QK-I
Aquificale		CAA06703	D-EVYESF	-K-		KEEAKRVSY-VVPDE		LVR-EEVG-HR-1
	Aqu. pyrophilus	2983493	D-EVYESF	-K-		NKEAKRVSY-VVPDE		VR-EEVG-HR-I
	Aqu. aeolicus			S			-D-VRE-KL	
0.1	Tre. pallidum	3322484	EH-IYSF			N-LTGEAKKV-Y-VPQG		YSTQEFI-Q
Spirochete	Bor. burgdorferi	SP/P28608	EIYSFM-	1.1.1.1.1.1.1.1		ASEIKMV-Y-EKGL	The Context of the South Street Control of the South Street Street Street Street Street Street Street Street St	Q-SEAT-T
	Lep. interrogans	AAN50903	-AVIRSAF	L		N-CESEMKHVSY-V-RSG	-EGVKF-TSAGE	FT-QERM-
-	Synechococcus PCC7942		RFANF			D-LTDESKRV-YTVRRDP		EFEEVA-MI-R-
Cyanobacteri	Synechocystis PCC6803		GFYSVF	-K-		D-ITNEATEVAYSVVKDG		QFEEQR-
-	Anabaena variabilis	2073390	FVY	Y		N-LSPESKRV-YT-RKD-	V-NIK-ARLNKE	F-AEEMK-
1	Cfx. aurantiacus	AF130446	EYSVF	-S-				EYQEMQ-
Deinococcus-	D. proteolyticus	1813672		W		DKDEAARSTVKEGP		YE-VR-1
Thermus, GNS	D. radiodurans	A75557	AAEVF	W		DKEEAARSTVKEGP	S-SVRIN-KD	LE-VR-1
	Thermus aquaticus	GB/Y07826	EG-I-EF			EEAKRV-Y-VVPGP	D-GVRKL	YT-EEMI-R-
Thermotoga	T. maritima	3882431	ER-IKSKM-				T-YK-RIDDKE	YT-QEFI-K-
Incinicitogu	Clo. acetobutylicum	118726	DK-IISKM-				TAEK-AIDDKN	YT-QEMI-Q-
	Clo. perfringens	SP/P26823	DK-IMSHM-				T-YK-NID-KD	LS-QEMI-Q-
	Myc. leprae	SP/P19993	-VDR-IRSVHM-				S-WSI-ID-K-	YTAQERM-
	Str. coelicolor	GB/014499	-VDR-IRSVHM-				T-WK-NLD-KD	FN-QFQ-
Gram-positive	Bif. longum	AAN24348	-VDR-ISSVHM-				S-WT-DID-K-	WT-QEAQILMK
	Cor. glutamicum	O8NLY6	-VDR-IRSVH				T-WS-AIDDKN	YTSQERT-M-
Bacteria	Bac. subtilis	GB/U39711	IMS HM-				T-YKIE-KD	YT-QEVII-QH
	M. pneumoniae	1674090	IVSM-				TSNK-TNPD	GSTKELQI-SY
	M. genitalium	1361531	IVSM-				TSNK-K-TTKE	LS-E-VQI-SY
		2145133	E-ILSSKM-				TSEK-SANAKE	YT-QEMI-QY
	Strep. mutans						TSEK-SAN-KE	YT-QEMI-QN
	T lootin							
	L. lactis Sta. aureus	GB/M98865 GB/X75428	E-IISSKM- VOSHM-				T-YK-DIE-KS	YT-QEMI-QN

Fig. 1. Partial alignment of Hsp70 sequences showing a signature sequence consisting of an insert of 21–23-amino-acid (aa) (outer box), which is specific for diderm (characterized by the presence of an outer membrane) bacteria which comprise mainly gram-negative phyla. The smaller 2-aa insert (inner box) within the large insert is distinctive of the proteobacterial group. *Dashes* in the alignment indicate sequence identity with the amino acids found in *Escherichia coli* sequence shown on the top line. The accession numbers of various sequences are shown in the second column. Only representative sequences from different *Bacteria* are shown. The absence of the large insert in *Aquificales* species supports their placement within the diderm (gram-negative) bacteria. Abbreviations of species names are: *A., Agrobacterium; Aqu., Aquifex; Bac., Bacillus; Bif., Bifidobacterium; Bor., Borrelia; Bru., Brucella; C., Calderobacterium; Ca., Caulobacter; Camp., Campylobacter; Cb., Chlorobium; Chl., Chlamydia; Chlam., Chlamydophila; Cfx., Chloroflexus; Clo., Clostridium; Cor., Corynebacterium; Cyt., Cytophaga; D., Deinococcus; Des., Desulfovibrio; E., Escherichia; Fib., Fibrobacter; Fl., Flavobacteria; Geo., Geobacter; Hel., Helicobacter; Hydro., Hydrogenothermus; L., Lactococcus; Lep., Leptospira; M., Mycoplasma; Myc., Mycobacterium; Myx., Myxococcus; Nei., Neisseria; Pas., Pasteurella; Por., Porphyromonas; Pse., Pseudomonas; Ral., Ralstonia; Ri., Rickettsia; Rh., Rhodobacter; Sta., Staphylococcus; Str., Streptomyces; Strep., Streptococcus; T., Thermotoga; Tc., Thermocrinis; Tre., Treponema; X., Xyllela. GNS in bacterial groups name refers to green nonsulfur bacteria (<i>Cf: aurantiacus*).

Proteobacteria, and *A. pyrophilus* and *A. aeolicus*, but is absent from all other *Bacteria* (Fig. 5). This insert is not found in the RpoB homologs from *Archaea*, providing evidence that the bacterial groups lacking this indel are ancestral. The shared presence of this insert in Proteobacteria and *A. pyrophilus* was first reported by Klenk et al. [27], but due

to limited sequence information, its evolutionary significance was not clear. We have now amplified a 0.83-kb fragment of *rpoB* from *C. hydrogenophilum* and a 1.2-kb fragment of this gene from *H. marinus* containing the signature region. Sequencing of these fragments revealed that this insert is present in both species (Fig. 5). The shared presence of this

			144	178
	E.coli	P06139	IAOVGTISA	
	Buch. aphidicola	Q59177	T	AKAEND
	X. fastidiosa	AE003849	A	SI-NIKI-
	Pse. aeruginosa	P30718		SI-OIE
	Pas. multocida	Q59687	-E	
	Nei. meningitidis	P42385	S	QAIE
	Ral. solanacearum	17545361	A	SI-AR
	Ri. Prowazekii	3861167	S -	G-KEI-EKKEE
Proteobacteria	Ca. crescentus	P48211		G-KEEMKN
	A. tumefaciens	P30779	V	GERQI-LDQRN
	Bru. abortus	P25967	V	AEAEIMQN
	Rh. capsulatus	P95678	V	GEAQI-RFD-SQN
	Geo. sulfurreducens	ZP_00082424		N-K-I-DIQE
	Des. vulgaris	ZP_00129431		S-I-NIS
	Camp. jejuni	e1287421	A	KI-NDED
	Hel. pylori	P42383	-TA	HNIDED
	C. hydrogenophilum	AY188749	-EA	N-TNIT-RD-
Aquificales	Hydro. marinus	AY188441	-E-IA	N-PEII-RSR-END
Aquincules	Tc. ruber	AY254214	-EA	N-P-IIP-EAG-R
	Aqu. aeolicus Chl. muridarum	2984379 AAF39243	-EA	N-PEIIDEED
	Chl. pneumoniae	14916541	A	N-AEI-NEN-S N-SEI-NEN-S
	Cb. tepidum	AAM71772	A	N-PEI-ED
Oblematics	Fib. succinogenes	AY017380	A	N-PEI-E-L-N-ENDI-
Chlamydiae,	Cyt. aquatilis	AF335327	-K-IAS	NVI-ET-FA
CFBG group	Bact. forsythus	CAB43992	-EH-AK	GGIQK
	Rhodo. marinus	AAD37976	A	G-KAI-QD-FED
	Por. gingivalis	D17398	-EH-AK	GNI-SRK
	Fl. ferrugineum	AF335328	-QA	NIFA
	Bor. burgodferi	P27575	AS	N-SYI-EKD
Spirochetes	Tre. pallidum	AAC65026	V-H-ASV	N-KEI-RIL-S-IENDD-D
	Lep. interrogans	L14682	N-AS	N-N-I-NDD
	Synechocystis PCC6803	2506274	A G	NEQMNQSL-
A	Synechococcus PCC6301	48022	A G	N-FEQMDKSL-
Cyanobacteria	Nostoc punctiforme	17132795	A G	NEQMQSL-
	Thermo. elongatus	11131866	AA G	NERMDSL-
Delasses	Cfx. aurantiacus	AAD33468	H-A-N	A-SEI-EV-AD
Deinococcus-	Thermus aquaticus	P45746	-EE-A	N-PEDEI
Thermus, GNS	D. proteolyticus	AF130448	-KK-AG	N-PQEESI-
and the second	D.radiodurans	G75499	-KK-AG	NQESI-
Thermotoga	T. maritima	H72367	H-AA	NSAEI-EED
J	Myc. leprae	P09239	ATAA	G-QSI-DNN
	Myc. tuberculosis	P06806	ATAA	G-QSI-DNN
	Cor. glutamicum	19553910	N-A-V-S	RVEIV-AEDV
	Bif. longum Str. coelicolor	24850323 8248795	A-A	A-PEEKLQDV A-TOI-E
· · · · · ·	Clo. perfringens	P26821	STAS R-AA	AKIDEN
Cram negitive	L. lactis	X71132	A-V-S	RS-KEY-SDERSDI-
Gram-positive	Sta. aureus	15927604	A	AEI-RY-SENDI-
Bacteria	Bac. halodurans	15613125	AAS	A-DEIERNDI-
	Bac. subtilis	16077670	AA	AESERNDI-
	Strep. pneumoniae	2545870	AAV-S	RS-KEY-SEDI-
	Strep. pyogene	21911301	AAV-S	RS-KEY-SERNDI-
	M. genitalium	12045254	-EAAS	GSKEIQALNTD
	M. pneumoniae	P78012	-EAAS	GSKEIQALNTD
				Int. Microbiol.

Fig. 2. Excerpts from Hsp60 sequence alignment showing a 1-aa insert that is commonly shared by Spirochetes, Cyanobacteria, Chlamydiae, CFBG group and Proteobacteria, but not present in *Deinococcus-Thermus*, green nonsulfur bacteria, *Thermotoga*, Actinobacteria and Firmicutes. Sequences from only representative species are shown. The presence of this insert in *Aquificales* species provides evidence for their placement within the former groups of *Bacteria*. For abbreviations of species names, see the legend to Fig. 1. Additional abbreviations: *Bact., Bacteroides; Buch., Buchnera; Rhodo., Rhodothermus; Thermo., Thermosynechococcus.*

prominent insert in the *Aquificales*, Proteobacteria, Chlamydiae and CFBG group of species provides evidence that these groups are related and that they diverged subsequent to other groups lacking the indel.

Another useful signature providing further clarification of the relationships among the Proteobacteria, Chlamydiae, CFBG group and *Aquificales* has now been identified in the enzyme inorganic pyrophosphatase (PPase), which catalyzes the hydrolysis of pyrophosphate. This protein contains a 2-aa insert in a conserved region that is shared by various Proteobacteria as well as *A. aeolicus*, but not present in Chlamydiae and CFBG group nor in any other *Bacteria* (Fig. 6). Approximately 300–335 bp fragments of *PPase* from *C. hydrogenophilum, T. ruber*, and *H. marinus* were PCR amplified and sequenced. The results (Fig. 6) show that PPases from *Aquificales* species contain this insert, which indicates that it is a common characteristic of the group. The shared presence of this indel in only the Proteobacteria and

			926		1142
	E. coli	223476		AASR(182aa)QES	GGTKDITGGLPRVA
	Pas. multocida	15603601		A(182aa)	V
	Pse. aeruginosa	15599465		(182aa) T	SK-R
	Vibrio cholerae	15640356		T 182aa -K-	N
	X. fastidiosa	15839221		T-LS 182aaA	SK-R
	Ral. solanaceum	17547752		(183aa) T	OK-R
Proteobacteria	Nei. meningitidis	15676061		(184aa) -A-	SK-R
FIOLEODACLEIIA	Ri. prowazekii	15604017		TK(180aa)K	TT
	Ca. crescentus	16124758		T-OV(177aa)T-G	AK-R
	A. tumefaciens	15889249		T-NV(179aa)L	AK
	Rho. rubrum	22966128			SK-R
	Des. desulfuricans	23483807		T 177aa R	SKV
	Hel. pylori	15645812		T (326aa) KAT	VKSRS
	Camp. jejuni	15791843	S		AKSS
	Agu. aeolicus	15606950			AKVRVE
Aquificales	Aqu. pyrophilus	7531199			AKVRVE
Aquincules	Hydro. marinus	AY188438		(181aa)	AK-RF-
	C. hydrogenophilum	AY188439		(182aa)	AK-R
Chlamydiae,	Chl. trachomatis	15605035		I-AT (184aa) RGA	IK
	Chlam. pneumoniae	16752963		I-AT(184aa)RGA	IK
CFBG Group	Cyt. hutchinsonii	23137570		TN(173aa)RSV	К-ВТ
	Cb. tepidum	21672997		OG(151aa)NLD	RVGGAK-E
	Lep. interrogans	AAN50617		A(217aa) TVA	EK-RVD
Spirochetes		15594733		V-QA(186aa)KG-	EK-QN
ophochetes	Bor. burgdorferi	15639234		TS(186aa)KAI	QK-SS
	Tre. pallidum			VFTG(677aa)F-R	AK-GIOIE
Cyanobacteria	Nostoc sp. 7120	BAB77962			AK-GVOIE
	Synechocystis 6803	BAA17364		VFTG(629aa)F-R	
Deinococcus-	Thermo. elongatus	BAC08191		VFTG 629aa F-R	AK-GIQIE
	Cfx. aurantiacus	ZP_00018295	T		ASADQQ
Thermus, GNS	Thermus aquaticus	20139788	· · · · · · · · · · · · · · · · · · ·		AVGTQI
Thematers	D. radiodurans	AAF10485	T		A-GG
Thermotoga	T. maritima	15643225	V		M-ASVS-TTE
	Cor. glutamicum	BAB97882	Q		VGGQ VGE0
	Myc. leprae	419676	Q		VAAASOT
	Bif. longum	ZP_00120580	L-SS		
Gram-positive	Fuso. nucleatum	19705326	T T		VAGVQE
	Ocean. iheyensis	BAC12069			VAGDQIQ
Bacteria	M. pneumoniae	13508254	TV		STENNLAQ-FE-LK
	Bac. subtilis	16077176	T		VAGDQIQ
	Lis. innocua	7514789	T		VAGDQIQ
	Clo. perfringens	18311394	T		VAGAQE
	Sta. aureus	7470964	T		VAGSQIQ
	Strep. pneumoniae	15903818	T		ASNTQQ
Archese	L. lactis	15673781	T		ASSSQQ
Archaea	Halo. sp. NCR-1	NP_281211	YA-		VAEM-V-QLI
	Meth. barkeri	ZP_00079002	YA-		VAEINV-LLI
	Sulf. solfataricus	AAK40565	FA-		IRELNV-LLI
					Int. Microbiol.

Fig. 3. The shared presence of a large insert in RNA polymerase β' in Spirochetes, Chlamydiae, CFBG group, Proteobacteria, and various *Aquificales* species. The cyanobacteria contain a much larger insert (>600 aa) in this position. The observed indel is not present in *Archaea* or other bacterial groups. For abbreviations of species names, see previous figure legends. Additional abbreviations: *Fuso.*, *Fusobacterium*; *Halo.*, *Halobacterium*; *Lis.*, *Listeria*; *Meth.*, *Methanosarcina*; *Ocean.*, *Oceanobacillus*; *Rho.*, *Rhodospirillum*; *Sulf.*, *Sulfolobus*.

Aquificales species provides evidence that these two groups are specifically related and that they diverged after branching of the other groups.

We have previously described many signatures that are unique to Proteobacteria and which provide clear distinction among the species belonging to the α -, β -, γ - and the δ , ε -subdivisions [20,22]. Two of these signatures, one consisting of a 10-aa insert in CTP synthase (Fig. 7) and the other of a 2aa insert in Hsp70 (see Fig. 1), are distinctive of the entire proteobacterial phylum and are not found in any other *Bacteria*. The proteobacterial signature in Hsp70 (2-aa insert) is present within the large insert in this protein, whose sequence for various *Aquificales* species is shown in Fig. 1. The 2-aa insert common to various Proteobacteria is not found in any of the *Aquificales* species, which indicates that the two are distinct from each other. We cloned and sequenced a 420-bp fragment of the CTP synthase gene from *C. hydrogenophilum*, *T. ruber*, and *H. marinus*. The CTP synthase from all these species, as well as from *A. aeolicus*, did not contain the 10-aa insert common to various Proteobacteria (Fig. 7). In addition to these signatures, the published sequences of *A. aeolicus* do not contain any of the other signatures distinctive of the α , β and γ -Proteobacteria [23]. These results provide strong evidence that the *Aquificales* phylum is distinct from Proteobacteria and that it branched off prior to the latter phylum.

			57	9
1	E. coli	145220	Contraction of the second s	GGKH NDLENVGYTARHHTFFE
	Pas. multocida	AAK03371	-PA	
	Pse. aeruginosa	AAG04292	-A-TVK	
	Vibrio cholerae	2500962	-A-TA	F F
	X. fastidiosa	AAF82937	-SEV-VADV-C-L	DS
	Ral. solanaceum	CAD14499	-P-VASVL	
Proteobacteria	Nei. meningitidis	AAF41948	-PA-K	
roteobacteria	Rho. rubrum	ZP 00013842	-P-HK	D
	Ri. prowazekii	CAA15280	-S-NK-VKSL	H
	Ca. crescentus	AAK24500	HPASK	D
	A. tumefaciens	AAK87639	-PT-ASA-K	D
	Des. desulfuricans	ZP 00131262	-AK-L-V	RR
	Hel. pylori	2314404	PSIPASL-M	L
	Camp. jejuni	CAB75143	P-PP-K-SC-T-I	D
	Tc. ruber	AY254210	-P-TVSC-K-L-V	SS
	Agu. pyrophilus	4587716	-P-KSC-K-L-V	ss
Aquificales	Aqu. aeolicus	2983727	-P-KSC-K-L-V	SS
Aquincales	Hydro. marinus	AY188439	이 사람이 많아? 이 집안 같아요? 아파가 가지 않는 것이 많이 많이 했다.	S PP
	C. hydrogenophilum	AY188751	-P-TVSC-K-L-V	S SS
	Cyt. aquatilis	AF130452	PKSP-IADT-K-L-V	S E IDTY
Chlamydiae,	Fl. ferrugineum	AF130451	PASA-VADT-K-L-V	S EVDTY
	Cb. tepidum	AAM71414	Constant Constant Constant Constant	S D RDTY
CFBG group	Chl. trachomatis	6758113		H-SL
	Chlam. pneumoniae	AAD19030	SK-I	DH-SL
	Lep. interrogans	AAN50605	-TSC-K-L-T	TVK-EC
a : 1 /	Bor. burgdorferi	2688110	PSGDMLVNV-K-L-T	G-IDE DLS-L
Spirochetes	Tre. pallidum	AE001269	PAGT-LVNA-K-L-T	G-IDADNS -L
	Nostoc sp. PCC 7120	BAB74117	FKK-I-T	IR-KO
Cyanobacteria	Synechocystis 6803	1653611	AEFPK-I-T	IR
ojunozaciona	Thermo. elongatus	BAC09655	PKVPA-K-L-T	IR
Deinococcus-	The. aquaticus	1565288	-EWR-VC-E-L-V	G-IR-SN-Y
	The. thermophilus	2500960	-EWR-VC-E-L-V	G-IR-SN-Y-
Thermus	D. radiodurans	AAF11848	OPSK-VA-KV	G-IR-RLSL-
Thermotoga	T. maritima	4981959	PV-T-VA-C-K-L-T	V-IK-P
mermotoga	Cor. glutamicum	BAB99025	FENGTSI-KT	L-I-EI-TN
	Myc. leprae	3136021	PP-ATSI-K-I-T	P-IDDI-TN
	Str. ceolicolor	CAB93381	PPFDSV-KT	P-I-EK-TG
	Bif. longum	AAN24695	PPKR-MASN-KT	L-IDEK-TG
	Therm. tengcongensis		TPPS-VCI-T	P-I-RKA
1.1	Ocean. iheyensis	BAC13965	PDNP-IVNA-KSIRT	T
Gram-positive	Bac. subtilis	2635186	PENP-IVNA-KAI-T	IK
Bacteria	Lis. innocua	CAC96770	PDNP-MANA-KSI-T	IK
Dacteria	Clo. perfringens	BAB81486	PPKR-IC-K-I-T	G-IK-SG
	Sta. aureus	BAB95433	PKKP-IVNKAI-T	IF
	M. genitalium	1351145	PPSK-LVNA-I-L-V	IF-SO-L-
	Strep. pneumoniae	AAL00044	PENP-I-NA-KAI-T	IKM-
	L. lactis	AAK05822	PENP-L-NA-KAI-T	IKM-
Archaea	Halo. sp. NCR-1	AAG20397	PPANPLVVP-I-M	Q-IDK-GTMA-
Archaea	Meth. barkeri	ZP 00077929	PPANPL-IP-I-L	DSRSGL-T
	Pyr. aerophilum	AAL65007	PPANPLVIPSI-L	T-VDKRSGL-G
	TIT. GETOPHITTUN	111000001	TTUREDAT LOT-D	1 ADV - VOQ - T-Q

Fig. 4. A 4-aa insert in alanyl-tRNA synthetase that is a unique characteristic of Proteobacteria, Chlamydiae, CFBG group, and *Aquificales* homologs. This insert is not found in other groups of *Bacteria* or Archaea, indicating a specific relationship of *Aquificales* to the Proteobacteria, Chlamydiae and the CFBG group. For abbreviations of species names, see previous figure legends. Additional abbreviations: *Pyr., Pyrobaculum; The., Thermus; Therm., Thermoanaerobacter*.

Discussion

In this work, we have used a cladistic approach involving the shared presence of conserved indels in widely distributed proteins to clarify the phylogenetic placement of *Aquificales* [29,33]. Unlike the phylogenetic trees in which the deduced relationships are dependent upon a large number of variables and often not resolved, the relationships inferred by this method are based on minimal assumptions and are unambiguous [3,19,23,37]. The phylum *Aquificales* is made up of four genera, *Aquifex, Hydrogenobacter, Calderobacterium*, and *Thermocrinis* [36]. The first two genera contain two and

three species, respectively, whereas the last two genera are made up of only a single species [36]. Except for *A. aeolicus*, whose genome has been sequenced [11], very limited sequence information is available for other *Aquificales* species. This work describes a large number of signatures, many of them for the first time, that are relevant to understanding the phylogenetic placement of this phylum. The sequence information for various genes that were studied here was obtained from most of the *Aquificales* genera in order to ensure the general applicability of the derived inference to the entire phylum.

A summary diagram of the results obtained from different signatures is presented in Fig. 8. Based on their observed dis-

			919		1058
	E. coli	42818	RVPNGVSGTVIDVQVFTRD	GVE(91aa) KRR	IT OGDD LAPGVLKIVKVYLAVK
	Pas. multocida	15603602	S	(91aa) Q-G-	
	Pse. aeruginosa	15599466	T-TK	(103aa) -K-	
	Vibrio cholerae	15640355	A	(92aa)	
	X. fastidiosa	15839222	P-MD	-I-(104aa)S	MF
	Ral. solanaceum	17547753	S-ME	T(104aa)K	-LE PI-M
	Nei. meningitidis	15676060	-М-Т-МЕ	-IQ(104aa)-KK-	
Proteobacteria	Rho. rubrum	ZP 00013726	-I-PTVE-RN-R	(104aa) -KV	KL -RG-E-PMFV
	Ri. prowazekii	6652726	HSVE-RI-S-R	(104aa) -VE-	LO SPO-AVFI-T-F
	Bru. melitensis	17987032	-M-P-TYVE-RN-H	(104aa) -VE-	VQ RE MPM-MFV
	Ca. crescentus	16124757	-L-PAIVRN-H		Q RE -PM-MVFV
	A. tumefaciens	15889250	-M-P-TFIVE-RN-H		VO RE MPM-MFV
	Des. desulfuricans	ZP 00129103	KV-P-IEKL-N-R		
	Hel. pylori	15645812	YC-PSLEKKK		
	Camp. jejuni	15791842	YATASLE-V-VKIKK		
	Aqu. aeolicus	15606949	-C-PE-IK		
	Agu. pyrophilus	4753643	-C-PE-IK		
Aquificales	Hydro. marinus	AY188442	TE-I-VA-K		EVE K-AKNELI-O
	C. hydrogenophilum	AY188443		3.7 Properties Networks, Contraction	G KRSE -PAIALI-O
	Cyt. hutchinsonii	ZP-00119276			TL EVG-E-PA-IVQLAI-K
	Cb. tepidum	AAM71403			N VE -PIEELAI-Q
Chlamydiae,		10637868			TDA TIG-E-PN-IIO-ALI-K
CFBG Group	Chl. trachomatis	15605036			I-K EAD-DHIRQV-S
CFBG Gloup	Chlam. pneumoniae	16753076			H-R EAD-DHIRQV-S
1	Lep. interrogans	AAN50618	-MFEIKR-S-E	DITE (11200/1140	NO-E -PAEEMFV-R
Out the test	Bor. burgdorferi	15594734	KH-TERI-KE		DVGN -SEE-LV-K
Spirochetes	Tre. pallidum	15639233	HEBLR-S		ENSEVLI-T
1	Nostoc sp. PCC 7120	17229086	EK-R-VRLE		E -PANMV-RV-0
Cyanobacteria	Synechocystis 6803	16329957	EK-R-VRE		KE -PANMV-RI-V-Q
Cyanobacteria	Thermo. elongatus	BAC08193	EK-R-VRE		E -PANMV-R1-V-Q
Deinococcus-		00017399	R-KKS-S		E-AE -PVNOT-R-L-CO
	Thermus aquaticus	20139789	P-EG-I-VGRLRLR-G		DPGVE-KREV-R-FV-O
Thermus, GNS	D. radiodurans	15805937	QS-QG-I-VKTVR-R-G		DEGVD-KREM-RV-Q
Thermotoga	T. maritima	15643224	-L-HE-RR-D-YDON		DIAE -GAL-R-V-SR
1	Cor. glutamicum	19551731	KH-ET-KG-RH-S-E		DDNEMIRI-V-O
	Myc. leprae	418765	KH-EKGIRSHE		DD-E -PANEL-RV-O
	Str. ceolicolor	21223036	KH-EI-KG-RD-E		EE -PNOL-RV-O
	Therm. tengcongensis		TM-H-SK-V-V-ILELS-E		NE -KAN-SIR-LV-E
	Ocean. iheyensis	BAC12068	H-GG-I-LKI-N-E		DE -PNQLVRA-IVO
	Bac. subtilis	CAB11883	H-GG-IIHKN-E		DE -PNOL-RIVO
Gram -positive	Lis. innocua	16799362	H-GG-I-LKIE		AE -PNQL-RIVQ
	Clo. perfringens	18311395	H-EA-IIVKE		NSNEL-RC-I-0
Bacteria	Sta. aureus	15923532	H-AG-I-LKN-E		
	M. genitalium	12045200	K-SH-GD-I-SA-KR-SIA		ET-SNQL-RIVQ
	Strep. pneumoniae	15901784	H-AD-V-RKIV		NE -NDIEMIVVQ
1	L. lactis	NP 267957	H-GG-I-HRE		NE -QSNML-RI-Q
	Halo. sp. NCR-1	AAG20693	TMRS-ED-VVDT-TLMEG-		NE -PSN-L-R-FI-Q
Archaea	Meth. barkeri	ZP 00079000	TMRSNET-I-DT-ILTESI		DGSK-AK-SVRDE-
Alonaod	Pyr. aerophilum	AAL62934	A-RR-EK-I-DK-IITESP		NGTRLAKVRDE- EGN-LR-REL-

Fig. 5. Partial alignment of RNA polymerase β -subunit sequences showing a large insert (>100 aa) that is specific for the Proteobacteria, Chlamydiae, CFBG group, and *Aquificales* homologs. This insert is not found in other bacterial or archaeal homologs.

tribution in different bacterial phyla, the diagram depicts the inferred evolutionary stages where these signatures were introduced in these genes during the course of bacterial evolution. In this diagram, all of the marked signatures are present in the various bacterial groups above the indicated insertion points, but they are not found in any of the groups that lie below. The large inserts in Hsp70, RpoB, RpoC, and AlaRS, which are absent from various monoderm bacteria (Firmicutes, Actinobacteria and *Thermotoga*) are also not found in any archaeal homologs, providing evidence that within *Bacteria*, the gram-positive (monoderm) bacteria (Firmicutes, Actinobacteria) and *Thermotoga* constitute early branching lineages [28]. The presence in *Aquificales* of the large insert in Hsp70, which is a distinctive characteristic of diderm bacteria, supports their placement within this group.

The signature in Hsp60 further excludes *Aquificales* from the *Deinococcus/Thermus* and GNS phyla. The large insert in RpoC, which in addition to its presence in *Aquificales* is found only in Cyanobacteria, Spirochetes, Chlamydiae, CFBG group, and Proteobacteria, places *Aquificales* within these groups. The shared conserved indels in AlaRS and RpoB further refine the placement of *Aquificales* to Chlamydiae, the CFBG group, and Proteobacterial groups. The signature in PPase, which is present only in Proteobacteria and *Aquificales* species, points to a specific relationship between these groups to the exclusion of all others. However, the absence of various signatures that are distinctive of the Proteobacteria in *Aquificales* homologs indicates that *Aquificales* constitutes a distinct phylum that diverged prior to the Proteobacteria. The only arrangement of the different bacte-

			88	142
1	E. coli	BAB38627	RPVGVLKMTDEAGEDAKLVAVPHS	KL SKEYDHIKDVNDLPELLKAQIAHFFEHYK
	X. fastidiosa	AAF84970		-I FSG-SE-I-QVSAHWLER-G
	Pse. aeruginosa	AAG07418	BGID	VL-KDV-EYTALEKN
	Vibrio cholerae	AAF95686	T	-IR
	Pas. multocida	AAK03275	SV	TAOS
	Ral. solanacearum	CAD16056	-TL-MS-OVN	PATA-MT-LS-IGON-LDKO
	Nei. meningitidis	NP 273684	-VIM-FV-DGEV-D-I-CAD	DR NNGNAY -TLSQO-IKEFH-N
Proteobacteria	Ca. crescentus	AAK22035	I-T-M-VS-E-ILVD	HPF-TGVTSWRTI-TEQ
	Brucella melitensis	17986360	IV-E-NS-K-E-IISP	
	Ri. prowazekii	CAA15034	-AIM-ES-L-E-IIT-	DITFELDC-MKR-V
	Rho. rubrum	AAF21981	Y-ER-E-ILG	YPYH-NVNNFGRPIELRR-EA
	A. tumefaciens	AAK88383	IMI-E-DG-K-E-ILAP	TQRKVHNYTITLKE
	Hel. pylori	AAD07684	-LN-ES-M-EI-L-ID	-I DPTHSYVIDSKHTLDK-KT
1	Camp. jejuni	CAB75274	-LII-ES-M-ELN-	-I DARNTYTQATLNK-KNT
1	Aqu. aeolicus	AAC07463	I-M-E-RI-T-VIE	DPS-SNT-DNIVREK-K
Aquificales	C. hydrogenophilum	AY254208	I-G-E-RE-I-TL	-V DPTS-DKA-LDK-K
Aquincales	Hydro. marinus	AY254205	-CR-I-L-ES-VVIV-	TPLT-QSLD-F-K-ILE-VKP
1	Tc. ruber	AT254213	I-M-E-RE-V-T-VIT	DPTNV-TLDPVVLDR-K
1	Cb. tepidum	AAM72059	-VIMR-I-HGEN-D-IIAAD	DMSVSD-H-IGGKHF-MELQE
Oblematics	Cyt. hutchinsonii	23135798	IMR-L-GGEK-D-IIAAD	DISVK-YN-ISPHTILEVQRD
Chlamydiae,	Chlam. pneumoniae	BAA99126	I-G-RII-SGEA-D-IILED	DLVFAE-E-IS-C-GTVLDM-Q-Y-LT
CFBG Group	Chl. muridarum	AAF39030	I-GIRIL-SGEA-D-IILED	DLV-GAME-IS-C-GSVLDM-Q-Y-LT
	Chl. trachomatis	AAC68367	I-GIRIL-SEEA-D-IILED	DLV-GN-E-ISEC-GTVLDM-Q-Y-LT
Spirochetes	Lept. interrogans	24198094	IMR-L-SGED-IIAAN	DMSVNN-ISEPHFTLELKD
	Synechocystis 6803	BAA18431	I-M-E-I-GGDR-E-ILCAK	DPR-TYV-SIAGHRLDEERS
Cyanobactoria	Nostoc sp. PCC 7120	BAB75269	I-F-E-I-GGDR-E-ILDK	DPR-A-V-SLVAPHRLDEERS
Cyanobacteria	Pro. marinus pastoris	23122353	-IIM-FD-GGEV-D-VILSD	D-RMHTFEG-HWLNETKYYW
	Thermo. elongatus	22295609	I-M-E-I-SGDR-E-ILCVD	DPR-AEV-SLK-IAPHRLEEERT
	Dehalo. ethenogenes	TIGR 61435	KLFYK-PLCIG	DPHFNF-R-LS-V-PH-LKE-FNI
Thermus, GNS	Thermus aquaticus	585324	-VL-L-EK-GVIG-VAE	DQRLQ-IG-VGV-QE-QT
	Myc. leprae	CAC29718	MFR-VH-G-D-VLCVN	DHRWHGII-V-TFELDA-KV
	Str. coelicolor	CAB42762	-AI-MFRG-DLCST	DPRVE-LR-IHHVS-FDRLE-QV
	Thermobif. fusca ZP	00058806	-AI-MFR-RG-D-VLCAT	DPRME-LR-IHHVN-FERLE-QTV-
	Cor. glutamicum	25029102	-IIFG-DLS-LD	DPRQ-IS-VSDFDE-EV
Cram nanitiva	Bif. longum	23335625	-ALYH-VG-D-VLCA	DVRF-DID-VN-YHE-KQ
Gram-positive	M. pneumoniae	AAB95962	-IA-E-V-DGEL-TLG-IDC	DPR-KE-NSKHRVDE-IG-LKT
Bacteria	M. genitalium	H64238	-IA-E-I-DGEI-TLG-IDC	DPR-KE-NQISKHRIEE-LI-LKT
	M. pulmonis	CAC13643	-IAMIGET-TIHDD	DYRLENALEDSKWLEE-KYSN
	U. urealyticum	B82908	-VL-AMI-GGET-TLIDV	DPR-KNNFK-I-LHWL-EVQDN
	Bac. stearothermophilus	2059312	-VI-Y-N-V-SGEIGVE	DPRF-EVRSIEQHKLKER
	Bac. halodurans	BAB05323	KVL-F-N-I-GGEQITE	DPRFKDVNSLKHKLDEKT
				Int. Microbiol.

Fig. 6. Excerpts from sequence alignment of inorganic pyrophosphatase showing a 2-aa insert (boxed), which is found only in *Aquificales* and proteobacterial homologs, indicating a specific relationship between them. For abbreviations of species names, see previous figure legends. Additional abbreviations: *Dehalo., Dehalococcoides; Pro., Prochlorococcus; Thermobif., Thermobifida; U., Ureaplasma.*

rial groups that is compatible with the various signatures places *Aquificales* in a position between Chlamydiae and CFBG groups and Proteobacteria. All of the other main groups within *Bacteria* appear to have branched off at earlier stages.

The question can now be asked whether these results can be explained by any other reasonable means. In this context, note that most of the proteins in which the various signatures are found (e.g., RpoB, RpoC, AlaRS, Hsp60, Hsp70, CTP synthase) are single-copy essential genes present in virtually all *Bacteria*. For most of these proteins, >50 sequences are available from the bacterial groups containing the inserts (e.g., Proteobacteria, *Aquificales*, Chlamydiae, and CFBG group) and an equally large number from the bacterial groups lacking them (e.g., Spirochetes, Cyanobacteria, *Deinococcus -Thermus*, *Thermotoga*, gram-positives). If these genes (or indels) were subjects of frequent lateral gene transfers (LGTs) [9,13,25], then one would expect that some of the species from the former groups would be lacking the signatures and at the same time several species from the latter group would possess these indels, resulting in a more random arrangement. However, the fact that all species from the former groups contain these signatures, and none of these signatures are found in species from the latter groups, provides strong evidence that they were introduced only once in a common ancestor of the first group of species. (A second homolog of Hsp70 lacking the large indel has been found in *Borrelia burgdorferi*. This homolog is likely derived by means of LGT and it is readily distinguished from the normal Hsp70 homolog [21].) The χ^2 probability (assuming two degrees of freedom) that the observed distribution of these indels is due to random occurrence is virtually nil (<10⁻¹⁰).

These results challenge and do not support the presently held view that the *Aquificales* group constitutes the deepest branching lineages within *Bacteria*. The deep branching of *Aquificales* in phylogenetic trees could result from a variety of factors, including the long branch-length effect and LGT [15,34]. The *Aquifex* genome appears to be rapidly evolving

			375		442
1	E. coli	147478	ANSTEFVPDCKYPVVALITE	WRDENGNVEV	RSEKSDLGGTMRLG
	Pas. multocida	AAK03956	TRT-DYG		A
	Pse. aeruginosa	AAG07025	TSSGHG	T-AI	EA
	Vibrio cholerae	AAF95590	G		
	X. fastidiosa	AAF84097	NDROSPHI	TTT-ER	-D
	Ral. solanaceum	CAD14828	NLETEH	-O-RD-RK	AD
	Nei. meningitidis	AAF41908	TLK-AAD-	T-D-SV-T	-DESA
Proteobacteria	Nit. europaea	3493582	-H-TP-TPYLG	-R-RC-RV-K	-SAOT
	Ca. crescentus	AAK23696	-SG PTERGIM	- IKGNE-VO	-RAND
	A. tumefaciens	AAK87397	-SG-TAEG-M	-VKG-EL -K	T-G
	Rho, rubrum	22968980	-S-SGVPEV -L-G-M	-VRG-ER-A	-GSGD
	Des. desulfuricans	23475278	EDEOTPDK-IY-M	이 같이 많은 것이 같이 많이 많이 많이 없다.	-DSG-EK
	Hel. pylori	3183132	TNQRYYGD		YN-P
	Camp. jejuni	CAB72520	VNEK-ONY-D-		AKTP
1	Aqu. aeolicus	2983754	TP-TPFIDIME-	the the blogs	OKKVDK
	Tc. ruber	AY254211	TPYTPYIDIMEG		OKKVDK
Aquificales	Hydro. marinus	AY254204	TPETPYID-MP-		ORNINNK
riganiouroo	C. hydrogenophilum	AY188752	-H-APNTPH-IID-LP-		OK-IE
i	Cyt. hutchinsonii	ZP 0011859	MNKTANINMMES		OKNVTMK
	Cb. tepidum	NP 661048	TN-RTRFID-MEH		OKKVKEK
Chlamydiae,	· · · · · · · · · · · · · · · · · · ·	AY017383	TENTTHID-MD-		OKNVTEK
	Fib. succinogenes Chl. trachomatis	2494759	M-MNPETPDCMMEG		ODSVVK
CFBG Group					
	Chlam. pneumoniae	4376510	L-M-PNTPH-I-YVMEG		QDPLVAT
Spirochetes	Lep. interrogans	AAN49608	IRTEHIS-LE-		QNDIEQM
op. constate	Tre. pallidum	AE001210	-H-RAV-TPHD-LPG		CV- TPTSL
Cuanahastaria	Nostoc sp. PCC 7120	BAB76699	ADSTTIN-LP-		QQDVV
Cyanobacteria	Thermo. elongatus	BAC08319	ADANTPHIH-LP-		QQDIV
	Synechococcus 7942	1399854	AD-ETPHILP-		QQDVV
Deinococcus,	Synechocystic 6803	2494761	AETETPNIN-LP-		QQ-VV
	Cfx. aurantiacus	22970289	N-QTAHID-MPD		QRDITEK
GNS bacteria	D. radiodurans	AE002001	AEYAKNK-ID-MP-		QLEVAGM
Thermotoga	T. maritima	AE001748	TPNTPYD-ME-		QKRILK
	Cor. glutamicum	BAB98810	-S-TPAATI-TME-		VSG-A
11.10.0	Myc. leprae	1709961	AEPATPDISTMAD		VAG-A-F
High G + C	Str. ceolicolor	5738492	D-ATAHSTMA-		VAGEG-M
Gram-positives	Thermobif fusca	23017870	LDKA-HSTMAD		VAG-R-M
orani poorarioo	Bif. longum	23326100	SEANI-TME-		VAG-G-M-H
1	Bac. subtilis	131733	-H-A-I-PSTQY-IID-LP-		QK-VEL
1	Ocean. iheyensis	BAC14963	A-IN-HTPHID-LP-		QKDIE
Low G + C	Lis. innocua	CAC97930	-H-A-IE-ET-HNIID-LP-		QKNIENML
	Clo. perfringens	BAB81914	-HI-PQTKYID-MPD		QK-ID-K
Gram-positives	Sta. aureus	BAB95916	-H-A-L-PATPY-IID-LP-		QK-IEL
	M. genitalium	629288	-D-TN-NTTH-IFDY-K		GI-RDNIL
	Strep. pneumoniae	AAK99242	A-LAPETKY-IIDIMRD		QI-IE-ML
	L. lactis	3757664	-H-FAL-PETKYIDIMRD		OV-VE-ML

Fig. 7. Partial alignment of CTP synthase sequences showing a 10-aa insert that is distinctive of various Proteobacteria, but not found in *Aquifica les* species or other groups of *Bacteria*. A smaller 4-aa insert specific for *Actinobacteria* is also present in this region. For abbreviations of species names, see previous figure legends. *Nit.*, *Nitrosomas*.

[11,36], and thus many Aquifex genes are subjects of long branch-length effects leading to their abnormal branching in phylogenetic trees. LGT is another important factor that can lead to abnormal branching in phylogenetic trees. About 10% of the A. aeolicus genes exhibit extensive sequence identity to homologs from various Archaea, implicating massive LGT between these groups [2]. If this is the case, then in phylogenetic trees constructed from homologs of the transferred genes, Aquificales species will branch near the root of the tree, as their sequences would closely resemble those of the Archaea. Note in this regard that, in contrast to the A. aeolicus genome, which has a G + C content of 43.4%, the G + Ccontent of 16S-23S-5S operons in this species is 65% [11], which suggests that either the rRNA genes in this species selectively evolved at a very rapid rate, or that they have been acquired from a high G + C species by means of LGT. However, the differences in evolutionary rates are not expected to have a significant effect on the placement of species into different clades based upon conserved indels in widely distributed proteins [19].

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The late divergence of the *Aquificales*, as suggested by the present work, has important implications for bacterial /prokaryotic phylogeny. The clustering of the *Aquificales* and other hyperthermophiles at the base of the prokaryotic tree has provided the main argument for a hot origin of life [8,12]. However, a later divergence of *Aquificales* suggests that thermal adaptation within *Bacteria* probably occurred in many different lineages independently [16].

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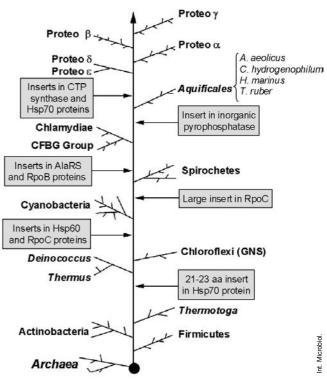


Fig. 8. A summary diagram indicating the evolutionary branch points where the signature sequences studied in the present work may have been introduced. All of the marked signatures are present in various groups above the indicated insertion points (i.e. those diverging later), but not in the groups lying below, which presumably diverged earlier. The insert in the RpoC protein, due to its different length, also distinguishes cyanobacteria from other *Bacteria*. The inserts in Hsp70, AlaRS, RpoB and RpoC are also not found in any archaeal homologs, supporting the inference that the groups indicated at the base of the tree are ancestral. The relative branching of other groups not studied here is based on signatures described in earlier work [21,23].

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Las secuencias signatura de diversas proteínas demuestran la divergencia tardía del Orden *Aquificales*

Resumen. Actualmente se cree que las especies de Aquificales son las que primero se separaron dentro del dominio Bacteria. No obstante, el orden de ramificación de este grupo no está resuelto y en los diferentes árboles filogenéticos es altamente variable. En este trabajo hemos examinado la posición filogenética de Aquificales mediante un enfoque cladístico basado en la presencia o ausencia de secuencias signatura definidas (consistentes en adiciones o deleciones conservadas) en muchas proteínas importantes y muy conservadas, como son la RNA polimerasa β (RpoB), la RNA polimerasa β' (RpoC), la alanil-tRNA sintetasa (AlaRS), la CTP sintasa, la pirofosfatasa inorgánica (PPasa), Hsp70 y Hsp60. Con este objeto, se clonaron fragmentos de los genes de las proteínas enumeradas que contenían las regiones signatura provenientes de diferentes especies de Aquificales (Calderobacterium hydrogenophilum, Hydrogenobacter marinus y Thermocrinis ruber) y se compararon las secuencias con las disponibles del resto de las especies. La presencia de insertos distintivos en las proteínas Hsp70 y Hsp60 de las especies de Aquificales, no presentes en ninguna especie de Firmicutes, Actinobacteria o Thermotoga-Clostridium, las excluyen de estos grupos del dominio Bacteria. La presencia compartida de importantes inserciones-deleciones en las proteínas RpoB (>100 aa), RpoC (>100 aa) y AlaRS (4 aa) que sólo se encuentran en varias especies de Aquificales, así como de Clamidias, el grupo CFBG (Cytophaga-Flavobacterias-Bacteroides-Bacterias verdes del azufre) y las Proteobacterias indica su pertenencia a estos grupos de Bacteria. Un inserto de 2 aa en la pirofosfatasa inorgánica, únicamente presente en los genes homólogos de Aquificales y Proteobacterias, parece indicar una relación específica entre estos dos filums. No obstante, las especies de Aquificales carecen de algunas otras signaturas de proteínas (por ejemplo, los indeles en CTP sintasa y Hsp70) características de las Proteobacterias, lo cual indica que constituyen un filum separado pero relacionado con las Proteobacterias. Estos resultados prueban intensa y consistentemente que las Aquificales se separaron después de la ramificación de los grupos Firmicutes, Actinobacterias, Thermotoga, Deinococcus-Thermus, Bacterias verdes del azufre, Cianobacterias, Espiroquetas y Clamidias-CFBG, pero antes de la emergencia de las Proteobacterias. [Int Microbiol 2004; 7(1):41-52]

Palabras clave: Aquifex · filogenia bacteriana · orden de ramificación

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As seqüências assinaturas de diversas proteínas demonstram a divergência tardia da Ordem *Aquificales*

Resumo. Atualmente acredita-se que as espécies de Aquificales são as que primeiro se separaram dentro do dominio Bacteria. No entanto, a ordem de ramificação deste grupo não está resolvida e é altamente variável nas diferentes árvores filogenéticas. Neste trabalho foi examinada a posição filogenética de Aquificales mediante uma aproximação cladística baseada na presença ou ausência compartilhada de seqüências com assinaturas definidas (representando adições ou deleções conservadas), encontradas em muitas proteinas importantes e altamente conservadas como são a RNA polimerase β (RpoB), a RNA polimerase β ' (RpoC), a alanil-tRNA sintetase (AlaRS), a CTP sintase, a pirosfosfatase inorgânica (PPase), Hsp 70 e Hsp60. Com esse obtetivo foram clonados fragmentos dos genes das proteínas enumeradas e que continham as regiões assinatura provenientes de diferentes espécies de Aquificales (Calderobacterium hydrogenophilum, Hydrogenobacter marinus e Thermocrinis ruber) e comparadas as seqüências com as disponíveis para as demais espécies. A presença de inserções distintas nas proteínas Hsp70 e Hsp60 das espécies de Aquificales as quais não foram encontrados em nenhuma espécie de Firmicutes, Actinobactérias ou Thermotoga-Clostridium, exclue estes grupos do domínio Bacteria. A presença compartilhada de importantes inserções-deleções nas proteínas RpoB (>100 aa), RpoC (>100 aa) e AlaRS (4 aa) em várias espécies de Aquificales assim como de Clamídeas, o grupo CFBG (Cytophaga-Flavobactérias-Bacteroides-Bactérias verdes do enxofre) e as Proteobactérias indica fortemente a inserção dessas espécies nos grupos de Bacteria. Uma adição de 2aa na pirofosfatase inorgânica, presente unicamente nos genes homólogos de Aquificales e Proteobactérias, parece indicar uma relação específica entre estes dois filos. Entretanto, as espécies de Aquificales não possuem assinatura em algumas proteínas (por exemplo, os indels em CTP sintase e Hsp70) que são características das Proteobactérias indicando que elas constituem um filo distinto, embora relacionado com as Proteobactérias. Estes resultados representam uma forte e consistente evidência de que os Aquificales se divergiram depois da ramificação dos Firmicutes, Actinobactérias, Thermotoga, Deinococcus-Thermus, Bactérias verdes do enxofre, Cianobactérias, Espioquetas e os grupos Clamídeas-CFBG, porém antes do surgimento das Proteobactérias. [Int Microbiol 2004; 7(1):41-52]

Palavras chave: Aquifex · filogenia bacteriana · ordem de ramificação