

1 **Interaction between contrasting rice genotypes and soil physical conditions induced by**  
2 **hydraulic stresses typical of alternate wetting and drying irrigation of soil**

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26 **Abstract**

27 *Background and aims* Alternate wetting and drying (AWD) saves water in paddy rice production  
28 but could influence soil physical conditions and root growth. This study investigated the  
29 interaction between contrasting rice genotypes, soil structure and mechanical impedance  
30 influenced by hydraulic stresses typical of AWD.

31 *Methods* Contrasting rice genotypes, IR64 and deeper-rooting Black Gora were grown in various  
32 soil conditions for two weeks. For the AWD treatments the soil was either maintained in a  
33 puddled state, equilibrated to -5 kPa (WET), or dried to -50 kPa and then rewetted at the water  
34 potential of -5 kPa (DRY-WET). There was an additional manipulated macropore structure  
35 treatment, i.e. the soil was broken into aggregates, packed into cores and equilibrated to -5 kPa  
36 (REPACKED). A flooded treatment (puddled soil remained flooded until harvest) was set as a  
37 control (FLOODED). Soil bulk density, penetration resistance and X-ray Computed  
38 Tomography (CT) derived macropore structure were measured. Total root length, root surface  
39 area, root volume, average diameter, and tip number were determined by WinRhizo.

40 *Results* AWD induced formation of macropores and slightly increased soil mechanical  
41 impedance. The total root length of the AWD and REPACKED treatments were 1.7-2.2 and 3.5-  
42 4.2 times greater than that of the FLOODED treatment. There was no significant difference  
43 between WET and DRY-WET treatments. The differences between genotypes were minimal.

44 *Conclusions* AWD influenced soil physical properties and some root characteristics of rice  
45 seedlings, but drying soil initially to -50 kPa versus -5 kPa had no impact. Macropores formed  
46 intentionally from repacking caused a large change in root characteristics.

47 **Keywords** rice roots; genotype; macropores; mechanical impedance; soil structure; X-Ray CT

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## 50 **Introduction**

51 Rice (*Oryza sativa* L.) is the staple food for over half of the world's population (Chen et al.  
52 2014). About 75% of total rice productivity comes from irrigated lowland rice systems (Bouman  
53 and Tuong 2001) that consume an estimated 24%-30% of the world's developed freshwater  
54 resources (Bouman et al. 2007). This is a major sustainability challenge (Bouman and Tuong  
55 2001) as water scarcity threatens the productivity of irrigated rice systems (Bouman et al. 2005).  
56 As a solution, alternate wetting and drying (AWD) is gaining adoption to decrease water  
57 demands, with large-scale international projects by the International Rice Research Institute  
58 (IRRI) and others promoting this technology (Lampayan et al. 2015).

59 AWD is likely to produce different soil physical conditions for rice growth than a flooded  
60 system, potentially influencing cultivar choice to maximise plant performance. Drying and  
61 wetting cycles from AWD have been shown to affect paddy soil structure compared to flooded  
62 systems (Zhang et al. 2003) and it has been demonstrated that AWD irreversibly increases soil  
63 strength at least in the top 12 cm of the soil (Norton et al. 2017). Puddled and flooded rice soils  
64 have little strength and much of the soil structure has been broken apart by mechanical action  
65 (Liu et al. 2005, Ringrose-Voase et al. 2000). Drying by AWD consolidates and shrinks the soil.  
66 Yoshida and Hallett (2008) found drying of paddy soils to -50 kPa water potential increased  
67 mechanical strength considerably, and that this strength did not decrease with subsequent  
68 wetting. Macropores may form as cracks and pre-existing pores that extend (Bottinelli et al.  
69 2016), creating connected pore systems favourable to rapid root growth. Under AWD, roots may  
70 therefore experience greater mechanical impedance from the soil matrix, but take greater  
71 advantage of newly formed pore networks. Root elongation of cereal crops is strongly influenced  
72 by physical properties (Bengough et al. 2011; Valentine et al. 2012; White and Kirkegaard 2010).  
73 Cairns et al. (2004) found that the increase of penetration resistance induced by drying potentially  
74 limits the growth of new rice nodal roots. However, the presence of macropores may offset the  
75 effect of mechanical impedance. In an arable, upland farming system, Lampurlanés and Cantero-  
76 Martínez (2003) found that greater soil strength under no-tillage does not greatly affect root  
77 growth in well-structured soils.

78 Zhang et al. (2009) concluded moderate AWD (re-watered when soil water potential  
79 reached -15 kPa) can enhance rice root growth and improve grain yield, while a severe AWD  
80 (re-watered when soil water potential reached -30 kPa) limits rice root growth and decreases  
81 grain yield. These results were also reflected in a recent meta-analysis of 56 studies on the

82 impacts of AWD on yield, it was observed that mild AWD ( $\geq -20$  kPa) did not cause a significant  
83 decrease in yield, however under AWD when water potential was less than -20 kPa a significant  
84 decrease in yield was observed (Carrijo et al. 2017). Perhaps -20 kPa drying produced highly  
85 restrictive root growth conditions. Monshausen and Gilroy (2009) found that mechanical  
86 stimulation of roots (i.e. transient bending) could elicit lateral root formation, possibly  
87 contributing to the positive impact of AWD at -15 kPa drying found by Zhang et al. (2009). The  
88 response of rice roots to AWD of paddy soil is still poorly understood and merits greater research  
89 interest.

90 Many reports have shown that root system architecture is influenced by both the soil  
91 environment and genotype (Acuña et al. 2007; Rogers et al. 2016). Rice genotypes with deeper  
92 roots are selected to improve resource capture under water saving irrigation strategies, such as  
93 AWD (Fang et al. 2013; Trachsel et al. 2011, Venuprasad et al. 2011), whereas shallow roots  
94 capture phosphorous more effectively (Clark et al. 2011). Breeding for root system architectures  
95 to maximize soil exploration and plant fitness (McCully 1995) offers considerable potential to  
96 improve yields, but too little thought has been given to root system response to soil physical  
97 properties (McKenzie et al. 2009). Roots may also induce changes to soil pore structure. By  
98 penetrating the soil, roots form macropores and create weak zones that are easy to fragment  
99 (Angers and Caron 1998). In AWD, surface cracks can be evident and the soil pore structure  
100 changes with drying (Ringrose-Voase et al. 2000).

101 The aim of this study was to explore the response of two contrasting rice genotypes with  
102 contrasting root architecture to soil physical conditions induced by hydraulic stress history or a  
103 manipulated macropore structure. To our knowledge, this is the first study to explore the  
104 interaction between soil strength, pore structure and rice genotype on rice root growth, with the  
105 results relevant to crop selection and soil management in AWD systems. For the AWD  
106 treatments the soil was either maintained in a puddled state, equilibrated to a constant water  
107 potential (-5 kPa water potential), or dried (-50 kPa water potential) and then rewetted (-5 kPa  
108 water potential). Our AWD simulates an initial drying cycle, with young plants studied so that  
109 cores of a suitable size for X-ray Computed Tomography (CT) could be used with minimal  
110 confinement of root growth. In AWD a water potential of -5 kPa will typically occur before water  
111 reaches a 15 to 20 cm depth where subsequent flooding is recommended (Yang et al. 2017). A  
112 water potential of -50 kPa is more extreme, above the thresholds of -10 to -20 kPa water potential  
113 where adverse effects on plant growth stages can occur, but often achieved in the field during  
114 periods of high plant transpiration and hydraulic gradients from the soil surface to the water

115 table. Soil physical conditions were characterised from core measurements of bulk density and  
116 penetration resistance, and a detailed X-ray CT analysis of macropore structure. We compared  
117 two rice genotypes, shallow rooting IR64 and deeper rooting Black Gora. Our hypothesis was  
118 that soil mechanical impedance to root growth in a paddy system would be dictated by the  
119 greatest drying stress, with differing impacts on the root morphology of rice seedlings between  
120 deep- and shallow-rooting genotypes. Some of these differences would be due to the creation of  
121 macropores, which would provide preferential channels for root growth that would overcome  
122 limitations from soil strength. The research has relevance to developing screening approaches  
123 of rice genotypes specifically for AWD systems. Moreover, it provides new information on how  
124 AWD may influence soil physical conditions.

## 125 **Material and methods**

### 126 **Rice cultivars and soil properties**

127 Contrasting rice genotypes were used in the study: IR 64 (an indica type with a shallow root  
128 system from the Oryza SNP set (McNally et al. 2009)) and Black Gora (an aus type with deep  
129 root system from the Rice Diversity Panel 1 (Zhao et al. 2011)). The soil used in the experiment  
130 was sampled from a paddy field maintained as permanent rice by the CREA Unità di Ricerca per  
131 la Riscicoltura in Vercelli, Italy and located at 45°19'25'' N and 8°22'25'' E. In the top 20 cm  
132 the soil texture consisted of 61% sand, 26% silt and 13% clay determined by the combination of  
133 wet sieving and hydrometer methods. It has 2.5 % organic carbon measured with a CNS  
134 elemental analyser (CE Instruments, Wigan, UK) and pH of 6.7 measured in a 1:5 soil to CaCl<sub>2</sub>  
135 using a pH meter (Hanna Instruments, Leighton Buzzard, UK).

### 136 **Experimental design and growth conditions**

137 We used PVC soil cores that were 5 cm in diameter and 8 cm high chosen to obtain an X-ray CT  
138 resolution < 40 µm so that macropores could be resolved. The cores were filled with soil that  
139 had been puddled by stirring soil and water thoroughly to mimic a paddy field. Four treatments  
140 were established: (1) FLOODED: puddled soil which remained flooded until harvest; (2) WET:  
141 puddled soil which was dried and maintained at -5 kPa to simulate the wet end of AWD; (3)  
142 DRY-WET: Puddled soil first equilibrated to -50 kPa followed by flooding and drying to -5 kPa  
143 to simulate a more drastic AWD cycle; and (4) REPACKED: Puddled soil that was first  
144 equilibrated to -50 kPa then broken into aggregates smaller than 4 mm before packing into cores  
145 to a similar bulk density to FLOODED, then flooded and dried to -5 kPa. A suction plate with a

146 bubbling pressure of -75 kPa was used to equilibrate water potential (Ecotech, Bonn, Germany).  
147 Each treatment had four replicates.

148 Each core was planted with one rice seedling. The seeds were germinated on wet filter  
149 paper at 30 °C for 48 h before being planted 3 mm below the soil surface. Plants were grown in  
150 a heated greenhouse with day/night temperatures of 28/26 °C and an 11 h photoperiod. The WET,  
151 DRY-WET and REPACKED treatments were grown on a large sand table that maintained the  
152 water potential. Over the first three days, the water potential was kept at -0.5 kPa to decrease  
153 stress on young seedlings, and then changed to -5 kPa until harvest. The FLOODED treatment  
154 cores were placed in a plastic tray filled with water to keep the cores flooded during the whole  
155 growth period. Plastic beads were put on the surface of cores to reduce evaporation. Rice  
156 cultivars were grown for 14 days. Each pot was irrigated daily by adding 10 ml of water to the  
157 top to compensate for the evaporative losses. This was quickly drained by the sand table for the  
158 WET, DRY-WET and REPACKED treatments to the prescribed water potential, but the  
159 hydraulic gradient in the core from evaporation may have induced a more negative water  
160 potential at the soil surface before watering.

#### 161 **Penetration resistance and bulk density measurements**

162 Before planting the rice, penetration resistance was measured by a Z005 mechanical test frame  
163 fitted with a 5 N load cell accurate to 0.05 mN (Zwick/Roell AG, Ulm, Germany). A 1 mm  
164 diameter, 30° full opening angle miniature cone penetrometer was inserted into the cores to a  
165 depth of 4 mm at a speed of 2 mm min<sup>-1</sup>. Three points were measured for every core. We defined  
166 soil penetration resistance as the plateau in the penetration stress measured during penetration.  
167 At the same time the bulk density was calculated from the mass of dry soil and volume.

#### 168 **X-ray Computed Tomography and image processing**

169 Soil cores were scanned using a XT H 225 ST CT scanner (Nikon Metrology, Tring, UK) with  
170 settings of 180 kV, 285 μA, 0.12° steps with 500 ms exposure time, 0.5 mm Cu filter and pixel  
171 size at 43.09 μm. Shortly before scanning, all the shoots were cut to slow root growth. All the  
172 cores were then drained to -50 kPa on the suction table to improve the quality of the CT images  
173 (Zappala et al. 2013). Drainage likely induces shrinkage in the FLOODED and WET treatments,  
174 which will be considered when interpreting the results. Drainage of pore water and storage after  
175 scanning was undertaken at 4 °C to avoid decomposition of roots until root-washing.

176 Three dimensional reconstruction was performed on the original images using the software  
177 CT Pro 3D (Nikon Metrology, version XT 4.3.1). The digital image processing and analysis were  
178 conducted with ImageJ (Version 1.50e). The 3D image stack of each soil core column was  
179 cropped to a region of interest of  $600 \times 600$  pixels ( $25.85 \times 25.85$  mm) and a depth of 800  
180 continuous slices (34.47 mm). Cropping the images and reducing the stacks was necessary to  
181 avoid ring artefacts caused by edge effects and beam hardening (Deurer et al. 2009; Mooney et  
182 al. 2006). Images were segmented using a 'Default' thresholding method, a variation on the  
183 'IsoData' method where the average of the object and background image are used to compute  
184 the threshold (Ridler and Calvard, 1978). Porosity and pore size distribution were computed  
185 using the 'thickness' plugin of 'BoneJ'. This approach fits the largest sphere inside the 3D pore  
186 space that touches the bordering soil matrix and then measures the sphere diameter.

187 Unfortunately the moisture content of the soil created considerable overlap between the  
188 greyscale values for the roots and the adjacent water filled pore space. This, combined with the  
189 small diameter of the rice roots ( $<0.1$  mm), meant that it was not possible to accurately segment  
190 the roots in the CT images in this study.

### 191 **Root traits**

192 After CT scanning, roots were carefully washed from the soil. Roots with soil were placed on a  
193 sieve (0.5 mm) and carefully washed with tap water to remove all soil particles. Root samples  
194 were placed in a plexiglas tray (100 by 200 mm) with a 4 to 6 mm deep layer of water, and spread  
195 out with tweezers to minimize overlapping. Grayscale images (800 DPI) of roots were obtained  
196 using an Expression 10000XL scanner (Epson, Suwa, Japan). Total root length, root surface area,  
197 root volume, average diameter, and tip number were determined by root analysis software,  
198 WinRhizo (Version 2013e) (Regent Instrument Canada Inc.). If not scanned immediately, the  
199 roots were immersed in a 50% ethanol solution in plastic containers with lids, and stored at 4 °C.  
200 On a subset of cores, manual counts of root tips were performed to check the accuracy of  
201 WinRhizo.

### 202 **Statistical analysis**

203 Data were checked for normality with probability plots. One-way ANOVA and post hoc analysis  
204 were conducted by the Fisher's protected least significant difference (LSD) procedure with SPSS  
205 24.0 to evaluate for significant differences between treatments ( $P \leq 0.01$ ). Significant statistical

206 differences of pore size distribution between rice cultivars were established by the Students t-  
207 test.

## 208 **Results**

### 209 **Soil physical conditions**

210 There was no significant difference between rice cultivars for the initial soil physical properties  
211 and growing conditions. The soil of the FLOODED treatment was the wettest and weakest, and  
212 its penetration resistance was 69.2%-77.3% less than the other treatments (Fig. 1). Penetration  
213 resistance of the DRY-WET treatment was 35.7% greater than that of the WET treatment (Fig.  
214 1). This was in agreement with our hypotheses that soils dried to -50 kPa and then rewet to -5  
215 kPa would be stronger than soils maintained at -5 kPa. For the REPACKED treatment, the  
216 penetration resistance was ranked between the WET treatment and the DRY-WET treatment  
217 (Fig. 1).

218 A small 2.4% increase in bulk density was caused by shrinkage of the WET and DRY-WET  
219 treatments, compared to FLOODED and REPACKED soils that had similar bulk densities ( $P <$   
220 0.05) (Table 1). This was reflected in the calculated total porosity, but when separated into air-  
221 filled porosity at -5 kPa, the equivalent of 60  $\mu\text{m}$  macropores, the REPACKED cores were very  
222 different to the FLOODED ones. In the DRY-WET treatment, visible cracks were created when  
223 dried to -50 kPa, as detected by the greater air-filled porosity compared to the WET treatment.  
224 Although the water potentials during the growing period were the same for the treatments, except  
225 for the FLOODED treatment, the water contents were different because their different soil  
226 structures affected their water holding capacity (Table 1).

### 227 **Macroporosity structure from X-ray CT images**

228 Cross-sections of the cores before plant growth are shown in Figure 2. After plant growth, harvest  
229 and drying to -50 kPa, the total cumulative macroporosity ( $>43 \mu\text{m}$ ) and pore size distribution  
230 obtained by CT showed differences between treatments (Fig. 3). For the REPACKED treatment,  
231 the macroporosity of both IR64 and Black Gora was much greater than other treatments ( $P <$   
232 0.01), with no difference between cultivars (Fig. 2). For the WET treatment, the macroporosity  
233 of IR64 was 47.0% more than Black Gora and their pore size distribution also showed significant  
234 differences ( $P < 0.01$ ). Visual examination showed this was caused by pores  $>500 \mu\text{m}$ , with IR64  
235 having 128% greater pore volume in this size range than Black Gora (Fig. 3). The macroporosity  
236 of FLOODED and DRY-WET treatments was less than the other two treatments and porosity of



237 pores in each size class (43-4900  $\mu\text{m}$ ) was also less than the other two treatments. Pore size  
238 distribution of IR64 and Black Gora were not statistically different for either the FLOODED and  
239 DRY-WET treatments based on t-tests at a range of pores sizes (Fig. 3).

## 240 **Root traits**

241 Images of the different root architectures between treatments are shown in Figure 4. For all the  
242 root parameters, only the average diameter of the root system was significantly different between  
243 genotypes. The diameter of Black Gora was 20.6% greater than that of IR64 in the FLOODED  
244 treatment and was 10.8% less than that of IR64 in WET treatment (Table 2). When comparing  
245 the differences between soil treatments for the same genotype, they followed the same trend. The  
246 diameter of the REPACKED treatment was 16.1%-22.1% less than other treatments for IR64  
247 and 14.4%-35.3% less than other treatments for Black Gora. The diameter of other three  
248 treatments were not significantly different (Table 2).

249 For both IR64 and Black Gora, total root length, surface area, root volume and number of  
250 tips of the REPACKED treatment were significantly greater than other treatments ( $P < 0.01$ )  
251 (Table 2). Specifically, the total root length of the REPACKED treatment was 3.47 times greater  
252 than that of the FLOODED treatment and c. 2 times greater than the WET and DRY-WET  
253 treatments for IR64. For Black Gora, the total root length of the REPACKED treatment was  
254 4.26 times greater than that of FLOODED treatment and c. 2 times greater than that of WET and  
255 DRY-WET treatments (Table 2). Surface area, root volume and the number of root tips followed  
256 the same trend with total root length, i.e. REPACKED > WET and DRY-WET > FLOODED.  
257 The difference between treatments of number of tips was greater than that of other root  
258 parameters. For IR64, the number of tips of the REPACKED treatment was 4.37 times greater  
259 than that of the FLOODED treatment and 1.99-2.31 times greater than that of the WET and  
260 DRY-WET treatment. For Black Gora, the number of tips of the REPACKED treatment was  
261 5.33 times greater than that of the FLOODED treatment and 1.88-1.97 times greater than the  
262 WET and DRY-WET treatment (Table 2). The root mass of different soil treatments did not  
263 show significant differences for IR64, while for Black Gora, the root mass of FLOODED  
264 treatment was 27.3%-33.3% less than other treatments (Table 2). The shoot mass of the WET  
265 treatment was 26.9% greater than the FLOODED treatment for IR64, while for Black Gora, it  
266 was not affected by AWD or soil packing. In addition, the shoot mass of Black Gora was 43.3%  
267 greater than that of IR64 for the REPACKED treatment (Table 2).

## 268 **Discussion**

269 The hypothesis that root morphology would vary due to the severity of AWD was not confirmed  
270 in this study. In rice production systems, AWD usually re-floods rice when it is wetter than -50  
271 kPa water potential, the driest water potential that we used (Belder et al. 2004; Bouman et al.  
272 2007; Norton et al. 2017). However, the hydraulic gradient to the evaporating surface and spatial  
273 heterogeneity of soil physical properties in the field (Becel et al. 2012; Valentine et al. 2012)  
274 could impart even greater hydraulic stresses on the soil. We found that soil drying caused an  
275 irreversible change to penetration resistance upon rewetting, albeit with mechanical impedance  
276 levels not limiting to root growth (Bengough et al. 2011). Root growth and AWD did induce the  
277 formation of macropores, and in comparison to the FLOODED treatment, this may be one cause  
278 of differences in root architecture. When macroporosity was intentionally manipulated in the  
279 REPACKED treatment, there were huge differences in root morphology. This treatment, with a  
280 prominence of interaggregate macropores, promoted root elongation and branching (Table 2).

281 Despite observing large differences in soil physical condition between the FLOODED,  
282 WET, DRY-WET and REPACKED treatments, the contrasting deep rooting Black Gora and  
283 shallow rooting IR64 genotypes generally did not differ markedly in either root structure or their  
284 impact on soil macroporosity. Between these genotypes, the only plant phenotypic difference  
285 was slightly greater average root diameter (12%) for IR64. To enable X-ray CT imaging we  
286 limited the study to small cores and young plants, but the root traits of seedlings may not be  
287 indicative of older plants (Atkinson et al. 2014), so follow-on work with larger cores is necessary.  
288 Moreover, in an unsuccessful attempt to resolve rice roots in X-Ray CT imaging, all soils were  
289 dried to -50 kPa before final scanning to improve segmentation (Zappala et al. 2013). This will  
290 inevitably induce shrinkage and crack formation (Yoshida and Hallett 2008), particularly in the  
291 WET and FLOODED treatments that never experienced -50 kPa during plant growth. With  
292 drying to -50 kPa the combination of the presence of roots and the shrinkage stress could  
293 dissipate macropore formation to a greater number of smaller pores. This was particularly  
294 evident for the WET treatment. In the DRY-WET treatment, shrinkage to -50 kPa before plant  
295 growth likely consolidated the soil, with only a few large shrinkage cracks forming near the  
296 sample edge (Fig. 2) that were outside the analysed volume.

297 An interesting finding for AWD systems was the formation of macropores and their  
298 potential to influence root morphology. Macropores could provide rapid root growth pathways  
299 in soil, and on re-wetting AWD systems they could improve water permeability to soil in the  
300 rooting zone above confining plough pans that are common in paddy rice systems. Passioura  
301 (1991) hypothesised that roots are not evenly distributed throughout the soil matrix and are

302 possibly trapped in large pores. The hypothesis has been verified by many subsequent studies.  
303 Colombi et al. (2017) created artificial, vertical macropores in the soil and observed via X-ray  
304 imaging that roots of wheat, soybean and maize can grow preferentially towards these  
305 macropores, although they may choose to cross through them rather than penetrate through them.  
306 Pfeifer et al. (2014) also found from X-ray imaging that barley roots tended to grow towards  
307 macropores in compacted soils. White and Kirkegaard (2010) observed that wheat roots  
308 preferred to grow in pores and structural cracks in dense, structured soil below 0.6 m.

309 Root elongation remains relatively unimpeded as long as the root tip remains “trapped” in  
310 the macropore (Pierret et al. 2007). Pierret et al. (1999) grew wheat plants in undisturbed soil  
311 cores from the field and in repacked soil cores filled at the same bulk density and found plants  
312 grew better in repacked cores than in undisturbed cores with 80% of roots located in macropores.  
313 They also found no difference between macropore sheath and bulk soil for both bacterial  
314 population and elements concentrations in repacked soil. This suggests that our REPACKED  
315 treatment might provide a good physical environment for roots, rather than biochemistry driving  
316 differences.

317 Mechanical impedance is one of the major limitations for root system growth and  
318 development (Bengough et al. 2011). Although the penetration resistance between our different  
319 soil treatments was far below the critical threshold of 2 MPa (Ringrose-Voase et al. 2000). A  
320 negative relationship between root elongation rate and penetration resistance has been observed  
321 for weaker soils (Bengough et al. 2011; Thangaraj et al. 1990). Whitmore and Whalley (2009)  
322 proposed root elongation rate decreases almost linearly with the increase of penetration  
323 resistance up to critical levels where elongation may cease. Our study did not find a negative  
324 relationship between root length (i.e. elongation) and penetration resistance. Whilst the  
325 penetration resistance of the AWD and REPACKED treatments was much greater than that of  
326 FLOODED treatment, the root length of the FLOODED treatment was much less than that of  
327 other treatments (Fig. 1 & Table 2). Others have observed a poor correlation between soil  
328 strength and rice root elongation in weaker soils (Rogers et al. 2016).

329 The greatest impact on root growth that we observed was the influence of macropores in  
330 the REPACKED treatment. Despite the REPACKED treatment having a penetration resistance  
331 that was closer to the WET and DRY-WET soil treatments than the flooded treatment, the  
332 REPACKED treatment had the largest root mass and length of all. In structured soils this  
333 suggests that mechanical impedance measurements obtained by rigid penetrometers could be of  
334 limited use (Bengough and Mullins 1990). Although a penetrometer is a direct and easy way to

335 measure penetration resistance, measurements need to be interpreted with associated information  
336 on pore structure to assess restrictions to root growth with greater rigour. This is supported by a  
337 study on spring wheat by Gaiser et al. (2013) who found that soil penetration resistance became  
338 much less significant for spring wheat root growth above biopore volumes between 0.015 and  
339  $0.030 \text{ m}^3 \text{ m}^{-3}$ .

340 Soil structure affects a range of physical limitations to root growth, including water, air and  
341 mechanical impedance (Whitmore and Whalley 2009). Mechanical impedance has been found  
342 to have a larger impact than water stress during drought (volumetric water content was 17%-  
343 24%) on rice root growth (Cairns et al. 2004). In a broad field survey of physical limitation to  
344 barley root growth, Valentine et al. (2012) found that the volume of pores between  $60 \mu\text{m}$  and  
345  $300 \mu\text{m}$  equivalent diameter (estimated from water-release characteristics) accounted for 65.7%  
346 of the variation in root elongation rates. We observed that greater drying by AWD increased both  
347 mechanical impedance and macropore development, with recovery not found with subsequent  
348 flooding. Consequently, root morphology was also altered, but not following expected trends for  
349 penetration or bulk density differences. Bulk density is a widely used parameter to quantify soil  
350 compaction, but it is poor at describing soil functions like the rate of root growth (Colombi et  
351 al. 2017). In our study, the bulk density of the four soil treatments was almost the same, but  
352 closer examination of soil macropores found large differences that may explain observed root  
353 morphology differences. Simple measurements like penetration resistance and bulk density  
354 provide an incomplete description of physical stresses experienced by growing roots, suggesting  
355 that macropores should not be neglected. There is great potential with non-invasive imaging to  
356 study these processes in much greater detail, including in naturally structured soils.

## 357 **Conclusions**

358 In comparison to the puddled state of paddy rice systems, the hydraulic stress induced by drying  
359 similar to the first cycle of AWD increased many root traits that are important to plant  
360 productivity. Imparting mild drying stresses of  $-5 \text{ kPa}$  or  $-50 \text{ kPa}$  increased penetration resistance  
361 by more than 400% compared to puddled soil, with subsequent rewetting having minimal impact  
362 on soil strength. The increased root tips after a hydraulic stress was imposed may be due to  
363 branching induced by mechanical impedance or the development of macropores that serve as  
364 preferential root growth pathways. Further investigations with REPACKED cores containing a  
365 large volume of macropores found an even greater impact to root traits than AWD. A comparison  
366 between contrasting deep-rooting (Black Gora) and shallow-rooting (IR64) rice genotypes found

367 little cultivar specific impact of the soil physical properties to root traits, or of the roots to soil  
368 physical properties. Further research should explore more mature plants and tracking the  
369 interaction between soil strength, pore structure development with AWD and rice genotypes.  
370 There may be potential in rice cultivation systems to manipulate soil structure through either  
371 tillage, cycles of wetting and drying or structure forming amendments like organic residues to  
372 enhance root structure. Simple measurements of soil physical properties such as bulk density or  
373 penetration resistance, as to their effects on root growth alone, may provide an incomplete  
374 assessment. A greater emphasis on the properties of macropores that provide easier growth  
375 pathways for roots is needed. Future research should also explore root phenotypic traits that may  
376 improve root:soil interactions in mechanically constrained soils where macropores provide  
377 important growth pathways.

378

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